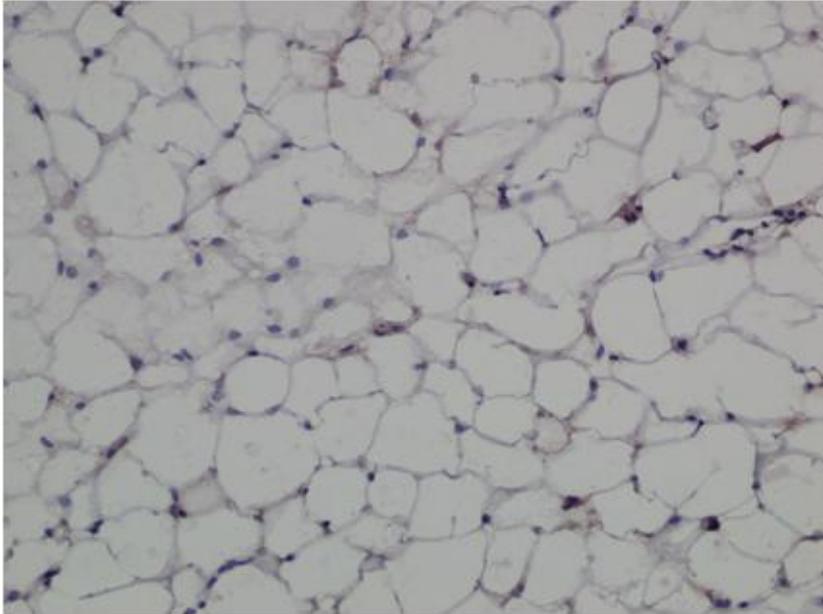
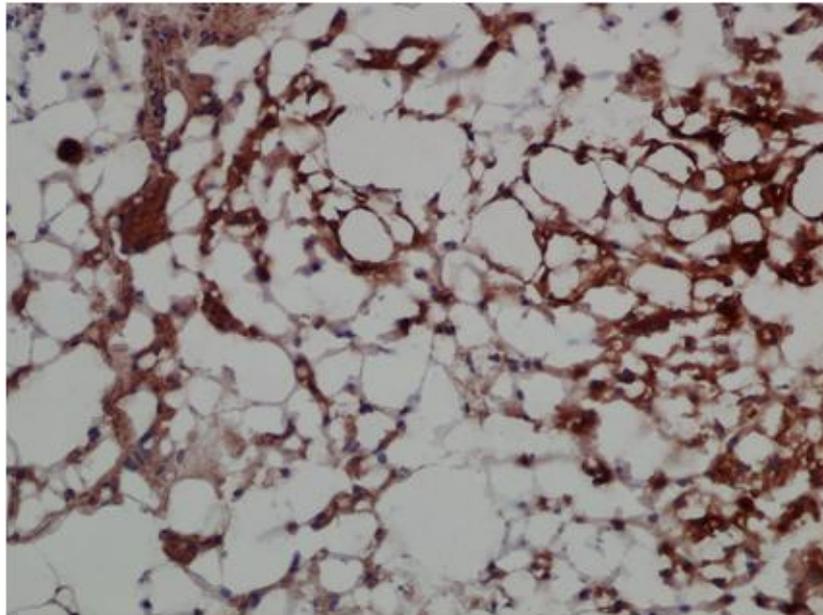


**Figure S1. ANP and  $\beta$ -agonist increase lipolysis in human adipocytes.** Lipolysis and second messenger levels in culture human subcutaneous adipocytes (A, B, C) and hMADS adipocytes (D, E, F). Cells were treated with Iso (○), or ANP (●) and compared with untreated control cells (■). Glycerol, cAMP and cGMP concentrations were all determined as described in Methods.

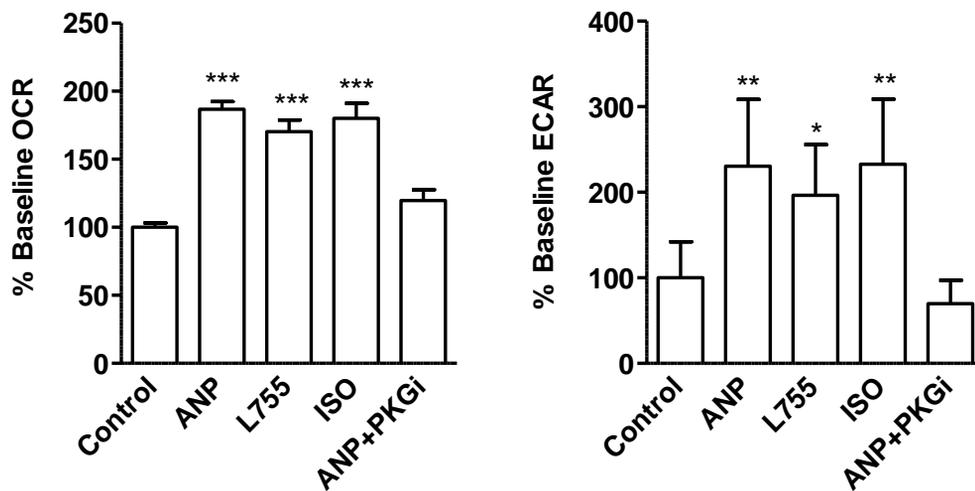
*Nprc*<sup>+/+</sup>



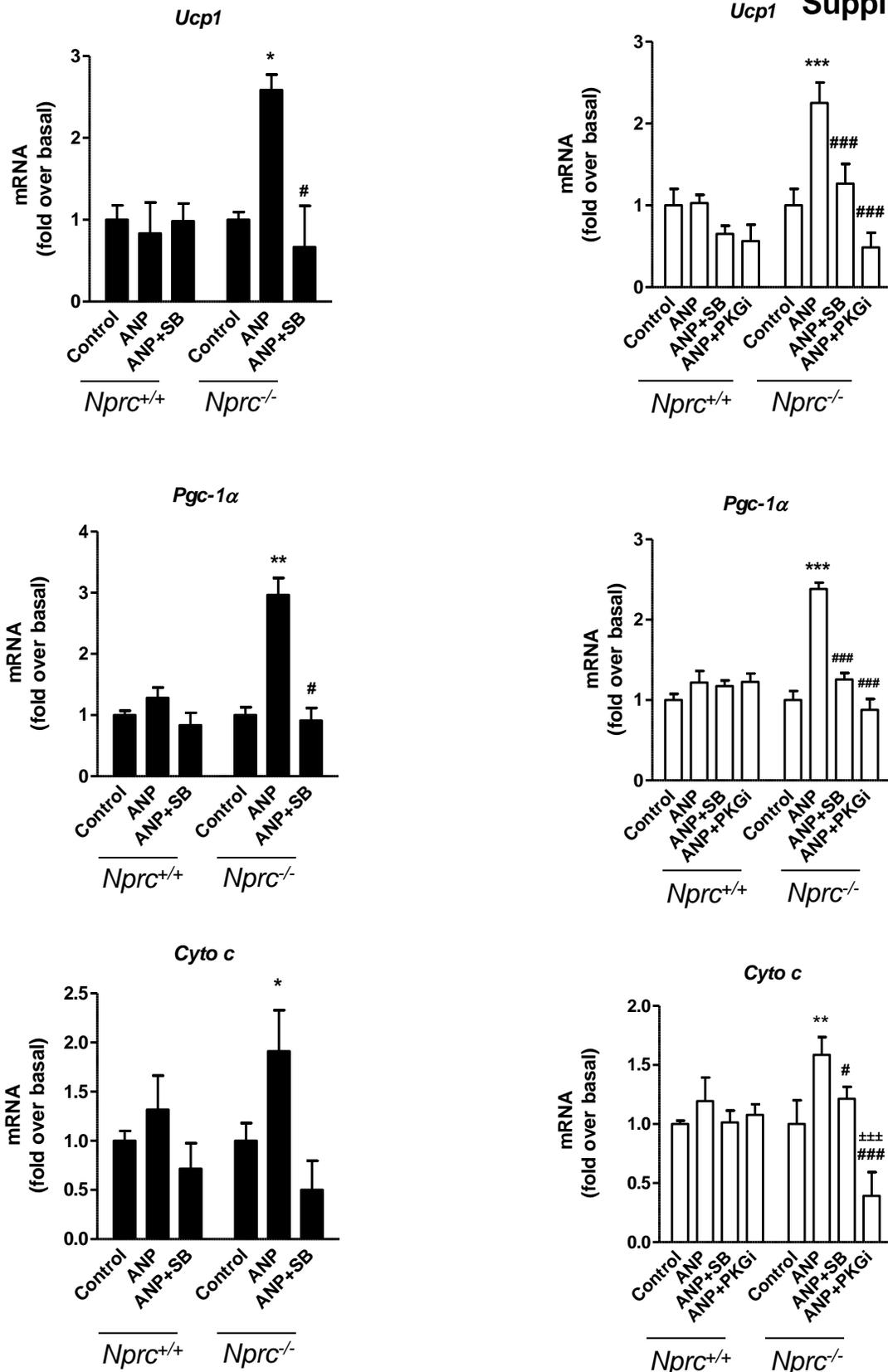
*Nprc*<sup>-/-</sup>



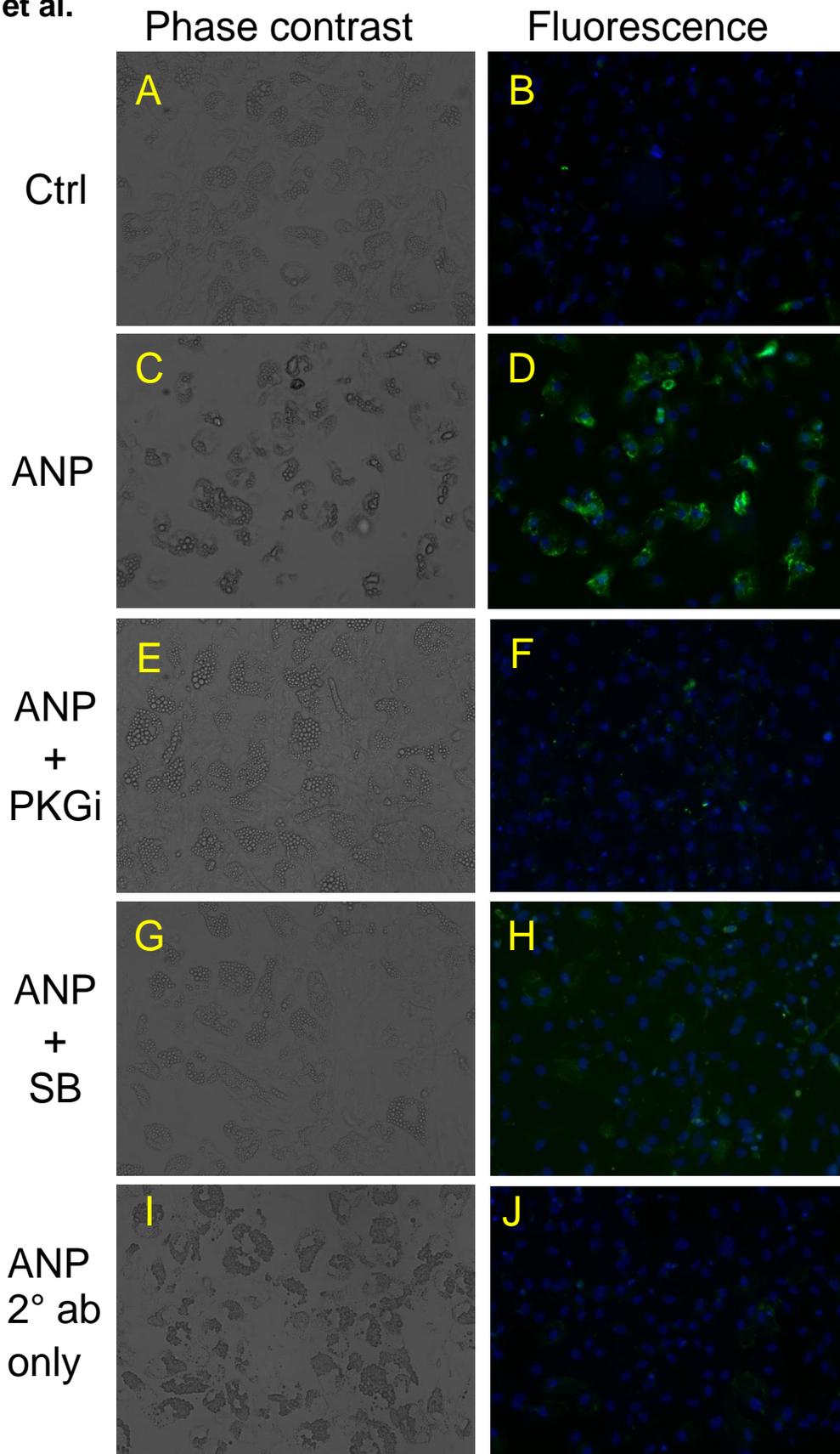
**Figure S2. UCP1 immunohistochemical analysis of epididymal WAT.** *Nprc*<sup>+/+</sup> and *Nprc*<sup>-/-</sup> epididymal WAT were processed to detect UCP1 immunoreactivity.



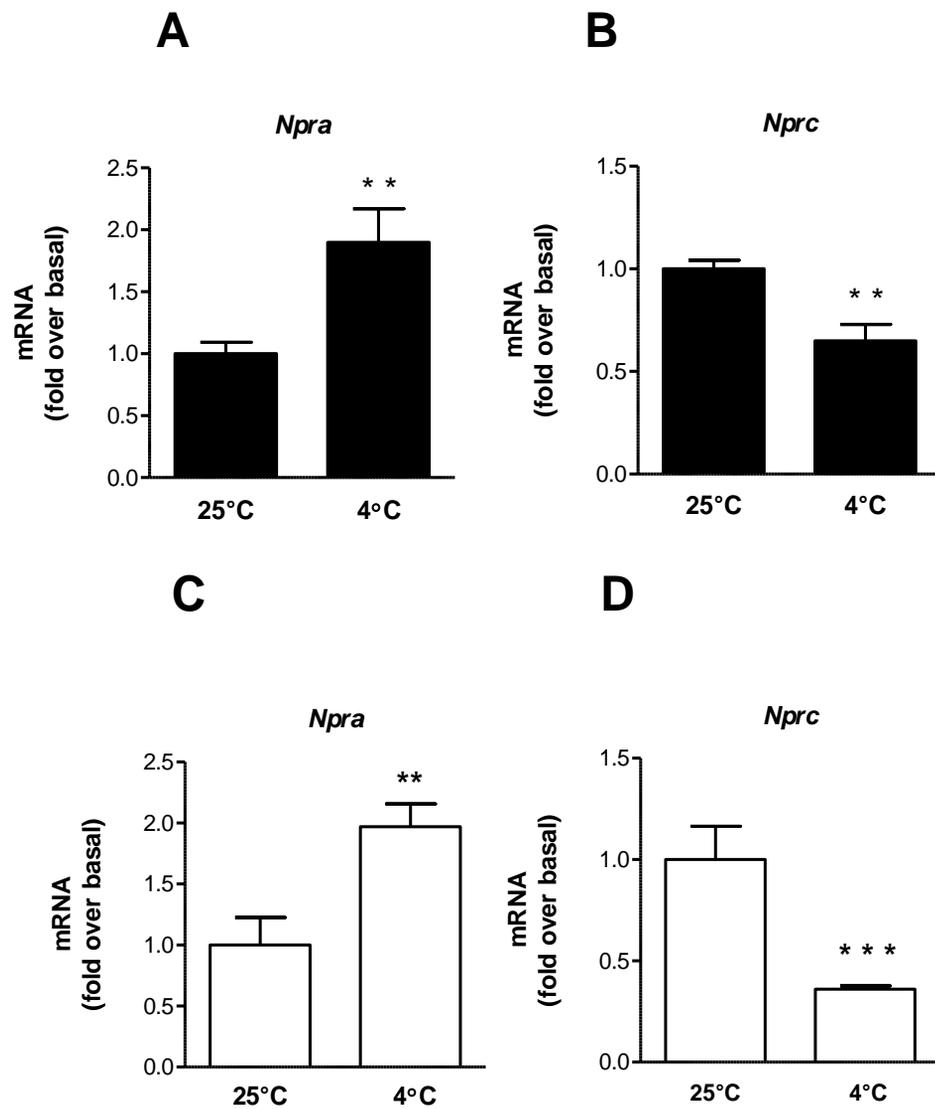
**Figure S3. Histograms summarizing the average maximal percent increase of OCR or extracellular acidification rates (ECAR) over their baseline rates.** Results are from 6-8 independent experiments. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$  for treated vs. untreated cells.



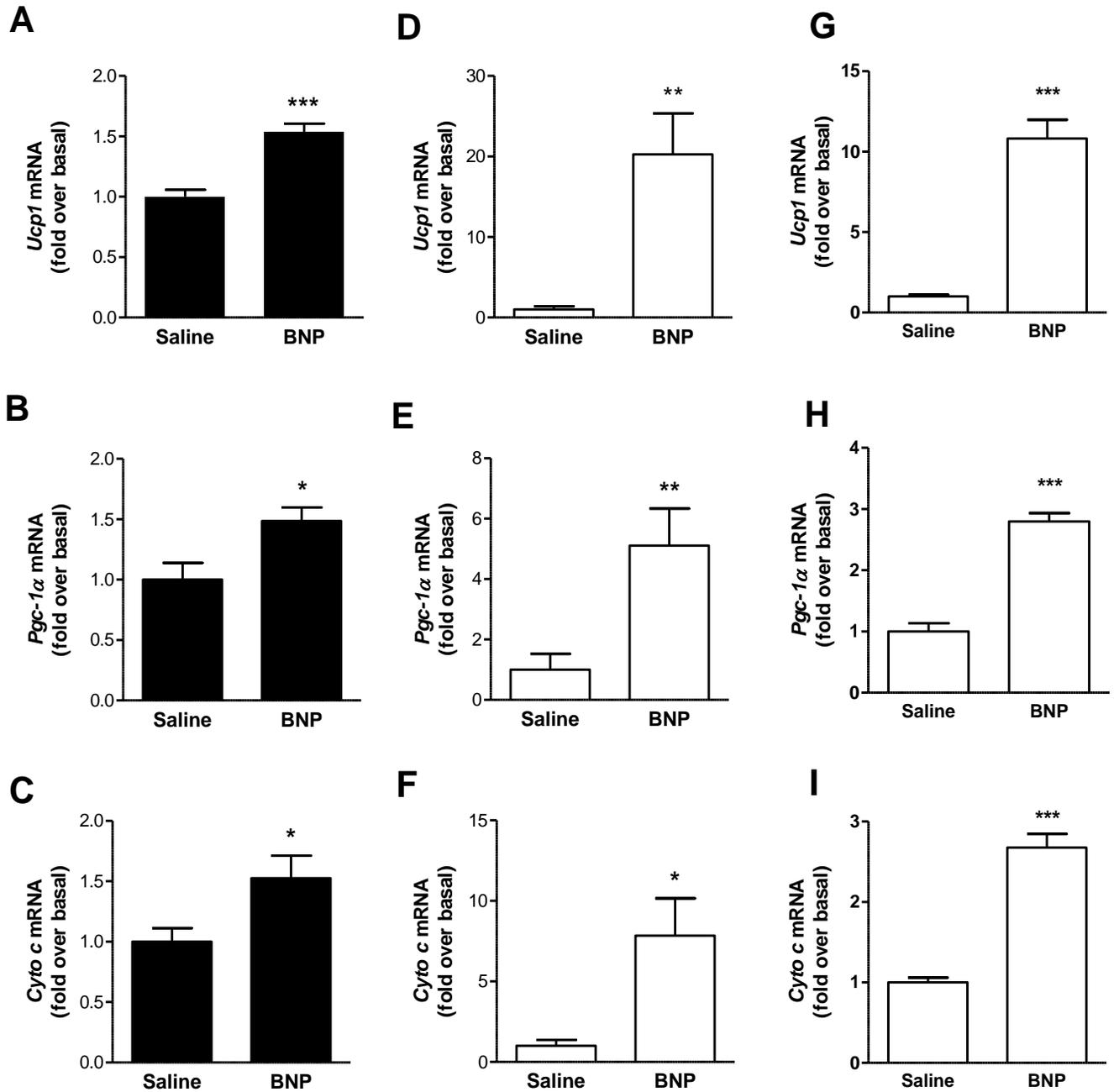
**Figure S4.** *Ucp1*, *Pgc-1α*, and *Cyto c* gene expression levels were evaluated in adipocytes obtained from iBAT (black bars) and epididymal WAT (white bars) of *Nprc*<sup>+/+</sup> vs *Nprc*<sup>-/-</sup> mice after treatment or not with ANP and pretreatment or not with PKGi or SB. mRNA levels were measured by quantitative real-time PCR, normalized to 36B4 and expressed relative to untreated control cells as described in Methods. Results are the means ± SD of at least 3 independent experiments.



**Figure S5. Images of hMADS immunostained for UCP1 and visualized under phase-contrast or fluorescence microscopy.** Cells were differentiated on the slide and either untreated (**A, B**), or treated with ANP (**C, D**) alone or after a 30 min pre-treatment period with PKGi (**E, F**), or SB (**G, H**). Specificity of UCP1 immunostaining was verified by absence of fluorescence with secondary antibody alone (**I, J**). In all slides nuclei are shown by the blue DAPI staining.



**Figure S6. Gene expression analysis of natriuretic peptide receptors in C57BL6 after 6 hrs of cold exposure (4°C; n=9) compared to control mice maintained at room temperature (25°C; n=8). **A** and **B**) *Npra* and *Nprc* mRNA levels in BAT, respectively; **C** and **D**) *Npra* and *Nprc* mRNA levels in epididymal WAT, respectively.**



**Figure S7. Adipose tissues from C57BL/6 mice treated with BNP (2ng/kg/hr) express higher levels of brown adipocyte marker genes.** Gene expression analysis of *Ucp1*, *Pgc-1α* and *Cyto c* in iBAT (panel A to C, respectively), inguinal WAT (panel D to F, respectively) and epididymal WAT (panel G to I, respectively) from C57BL/6 mice treated with saline or BNP by Mini-pump. \* P < 0.05 vs. Saline \*\* P < 0.01 vs. Saline. \*\*\* P < 0.001 vs. Saline.

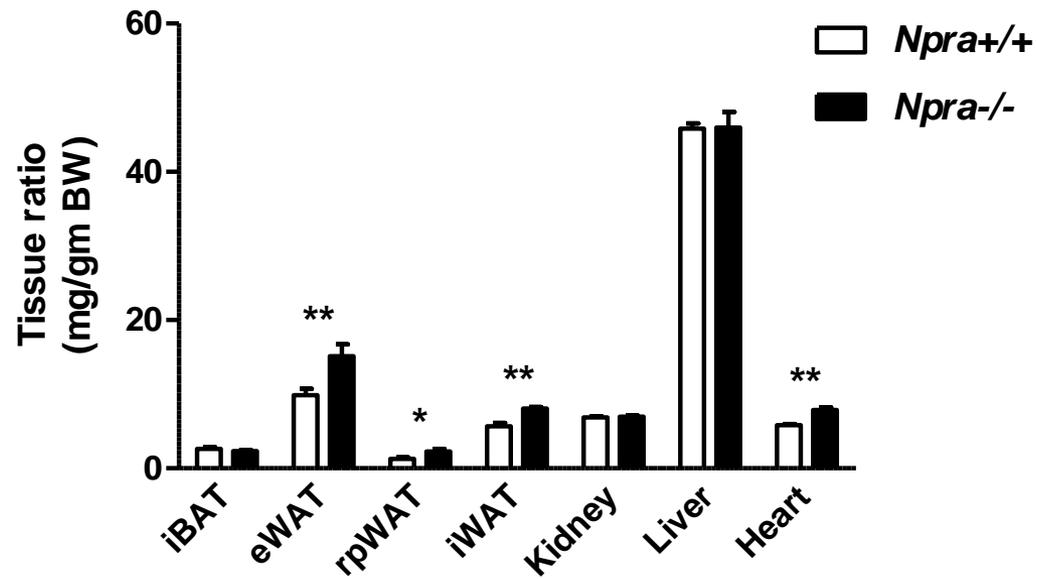


Figure S8. Ratio between weight of isolated tissues and total body weight of *Npra*<sup>+/+</sup> vs *Npra*<sup>-/-</sup> mice.

TABLE S1

Effect of ANP on glycerol release from cultured wild-type and *Nprc*<sup>-/-</sup> adipocytes

<i>Treatment</i>	<b><i>Brown adipocytes</i></b>				<b><i>White adipocytes</i></b>			
	<i>Nprc</i> <sup>+/+</sup>		<i>Nprc</i> <sup>-/-</sup>		<i>Nprc</i> <sup>+/+</sup>		<i>Nprc</i> <sup>-/-</sup>	
	x	sd	x	sd	x	sd	x	sd
Ctrl	61.30 ± 9.45	(3)	64.00 ± 4.86	(3)	65.66 ± 11.71	(3)	53.64 ± 4.39	(3)
ANP	71.20 ± 4.12	(3)	90.04 ± 7.75	(3)**	68.16 ± 7.47	(3)	72.95 ± 0.84	(3)**

Cultured brown and white adipocytes from *Nprc*<sup>+/+</sup> and *Nprc*<sup>-/-</sup> were treated with ANP (100 nM), and aliquots of culture medium were collected and assayed for glycerol release. Concentrations were determined from a standard curve. \*\*P < 0.01 vs. untreated cells.

**Table S2****Human primers**

<b><i>UCP1</i></b>	FWD primer RV primer	5'- ACGACACGGTCCAGGAGTTC -3' 5'- CTGAGCCCGCAGTGTGG -3'
<b><i>CYTO C</i></b>	FWD primer RV primer	5'- TGGCCCCTCCCATCTACAC -3' 5'- ATCCTTGGCTATCTGGGACATG -3'
<b><i>PGC-1<math>\alpha</math></i></b>	FWD primer RV primer	5'- CACCAAACCCACAGAGAACA -3' 5'- GGGTCATTTGGTGACTCTGG -3'
<b><i>PRDM16</i></b>	FWD primer RV primer	5'- CAGCACGGTGAAGCCATTC -3' 5'- GCGTGCATCCGCTTGTG -3'
<b><i>hCPT1B</i></b>	FWD primer RV primer	5'- CTGGAAGAAACGCCTGATCC -3' 5'- GCATCTCTGGATGCAACTGA -3'
<b><i>hMFN1</i></b>	FWD primer RV primer	5'- CTGTTGCCGGGTGATAGTTG -3' 5'- AGGAGTCAGGGCCAGAGC -3'
<b><i>hMFN2</i></b>	FWD primer RV primer	5'- TCCCTGCTAGGAGTTGCTGTAC -3' 5'- CACCTCAGCCCATGTGTCTCTT -3'
<b><i>hCOX10</i></b>	FWD primer RV primer	5'- CACACTCTCTCCTCACGCCTC -3' 5'- TTCTTTCAAGATACCAGACAGAGC -3'
<b><i>SIRT3</i></b>	FWD primer RV primer	5'- CTGTGTCAGCGGAAACT -3' 5'- TCCTATGTTACCATTTATTGTGTGC -3'
<b><i>GAPDH</i></b>	FWD primer RV primer	5'- TGGTCTCCTCTGACTTCAAC -3' 5'- GTGAGGGTCTCTCTCCTCCT -3'

**Mouse primers**

<b><i>Ucp1</i></b>	FWD primer RV primer	5'- TAAGCCGGCTGAGATCTTGT -3' 5'- GGCCTCTACGACTCAGTCCA -3'
<b><i>Pgc-1<math>\alpha</math></i></b>	FWD primer RV primer	5'- CGGAAATCATATCCAACCAG -3' 5'- TGAGAACCGCTAGCAAGTTTG -3'
<b><i>Cyto c</i></b>	FWD primer RV primer	5'- TGGCCCCTCCCATCTACAC -3' 5'- ATCCTTGGCTATCTGGGACATG -3'
<b><i>Npra</i></b>	FWD primer RV primer	5'- CGAAGACAAGTGCATCCTGAG -3' 5'- TGGAGACACAGTCAACACAGC -3'
<b><i>Nprc</i></b>	FWD primer RV primer	5'- AGCTGGCTACAGCAAGAAGG -3' 5'- CGGCGATACCTTCAAATGTC -3'
<b><i>36B4</i></b>	FWD primer RV primer	5'- GATGCCCAGGGAAGACAG -3' 5'- ACAATGAAGCATTTTGGATAATCA -3'

**Primers for quantification of mtDNA**

<b>NADH dehydrogenase Subunit 1 (mitochondrial)</b>	FWD primer RV primer	5'- CCCTAAAACCCGCCACATCT -3' 5'- GAGCGATGGTGAGAGCTAAGGT -3'
<b>Lipoprotein Lipase (genomic)</b>	FWD primer RV primer	5'- CGAGTCGTCTTTCTCCTGATGAT -3' 5'- TTCTGGATTCCAATGCCTTCGA -3'

**Primers for ChIP assay**

<b>UCP1 enhancer</b>	FWD primer RV primer	5'- GTGCAGCGATTTCTGATTGA -3' 5'- AGGGTCACAGGAGAAGCTGA -3'
<b>UCP1 exon 2</b>	FWD primer RV primer	5'- ACGACACGGTCCAGGAGTTC -3' 5'- GTTTGGGGAAGGGAGAGTTC -3'

**Suppl Table 1.** Primer sequences used for gene expression analysis, chromatin immune-precipitation (ChIP) assay and quantification of mitochondria DNA content (mtDNA).