

Supporting Information

Brønsted Acid Mediated Cyclization of Enaminones. Rapid and Efficient Access to the Tetracyclic Framework of the *Strychnos* Alkaloids

Rahul V. Edwankar,[†] Chitra R. Edwankar,[†] Ojas A. Namjoshi,[†] Jeffrey R. Deschamps,[‡]
and James M. Cook^{*†}

[†]Department of Chemistry & Biochemistry, University of Wisconsin-Milwaukee, Milwaukee,
WI-53201, United States; [‡]Centre for Bimolecular Science Engineering, Naval Research
Laboratory, Code 6930, Washington, D.C. 20375, United States

*To whom correspondence should be addressed. Tel: 414-229-5856; Fax: 414-229-5530;

E-mail: capncook@uwm.edu

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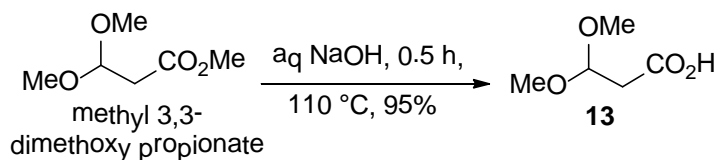
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General Experimental Considerations:

All reactions were carried out under an argon atmosphere with dry solvents using anhydrous conditions unless otherwise stated. Tetrahydrofuran (THF) and diethyl ether were freshly distilled from Na/benzophenone ketyl prior to use. Dichloromethane and acetonitrile were distilled from calcium hydride prior to use. Methanol was distilled over magnesium sulfate prior to use. Benzene and toluene were distilled over Na prior to use. Reagents were purchased at the highest commercial quality and used without further purification unless otherwise stated. Thin layer chromatography (TLC) was performed using Dynamic Adsorbents Inc. UV active silica gel, 200 μm , plastic backed; Dynamic Adsorbents Inc. TLC plates were visualized by exposure to short wavelength UV light (254 nm). Indoles were visualized with a saturated solution of ceric ammonium sulfate in 50% sulfuric acid.¹ Elemental analyses were performed on a Carlo Erba model EA-1110 carbon, hydrogen, and nitrogen analyzer. All samples submitted for CHN analyses were first dried under high vacuum for a minimum of six hours using a drying pistol with isopropyl alcohol or benzene as the solvent with potassium hydroxide pellets in the drying bulb. Melting points were taken on a Stuart melting point apparatus SMP3 manufactured by Barloworld Scientific US Ltd. Proton (^1H NMR) and carbon high resolution nuclear magnetic resonance spectra (^{13}C NMR) were obtained on a Bruker 300-MHz or a GE 500-MHz NMR spectrometer. ^1H NMR data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, ddd = doublet of doublet of doublet, td = triplet of doublet, m = multiplet), integration, and coupling constants (Hz). ^{13}C NMR data are reported in parts per million (ppm) on the δ scale. The low resolution mass spectra (LRMS) were obtained as electron

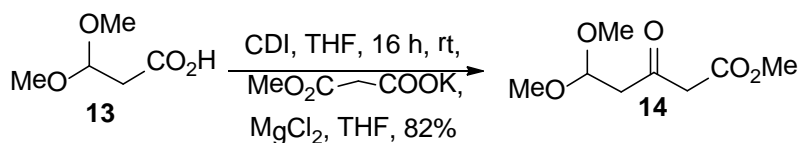
impact (EI, 70eV) mass spectrometer, which were recorded on a Hewlett-Packard 5985B gas chromatography-mass spectrometer, while high resolution mass spectra (HRMS) were recorded on a VG Autospec (Manchester England) mass spectrometer. HRMS recorded by electrospray ionization (ESI) methods were performed at the Laboratory for Biological Mass Spectrometry at Texas A&M University on a API QStar Pulsar model, manufactured by MDS Sciex. Optical rotations were measured on a JASCO Model DIP-370 digital polarimeter. Infra-red spectra were recorded on a Thermo Nicolet Nexus 870 FT-IR or a Perkin Elmer 1600 series FT-IR spectrometer. Flash and gravity chromatography was performed using silica gel P60A, 40-63 μm purchased from Silicycle. All reactions were carried out under an argon atmosphere with dry solvents using anhydrous conditions unless otherwise stated.

Experimental Procedures and Analytical Data:



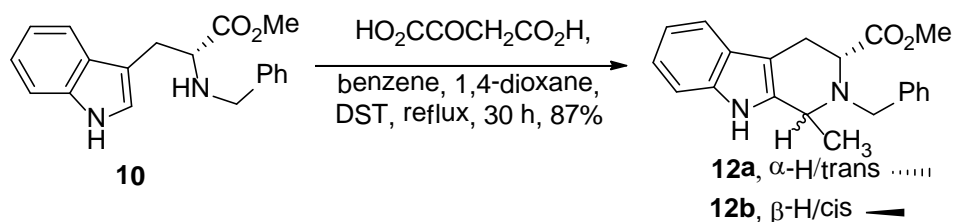
Hydrolysis of Methyl 3,3-dimethoxypropionate to 3,3-dimethoxypropionic Acid (13).² A mixture of methyl 3,3-dimethoxypropionate (14.2 g, 95.8 mmol) and 3.5 M aqueous NaOH (4.6 g of NaOH dissolved in 33 mL of H₂O) was stirred at 110 °C for 0.5 h. After 0.5 h, analysis by TLC on (silica gel, EtOAc/hexanes = 1:1) indicated the disappearance of starting material and a new spot at lower R_f had appeared. The clear solution which resulted was allowed to come to rt, cooled and neutralized by careful addition of cold aqueous 12 M HCl. After neutralization the solution was acidified to a

pH of 3 - 4 by addition of more aqueous 12 M HCl at rt. The acid **13** was extracted with EtOAc and CH₂Cl₂ (after each extraction the aqueous phase was acidified with 12 M HCl), and the combined extracts were dried (MgSO₄) and concentrated under reduced pressure to yield 12.2 g (95%) of the title acid **13** as a colorless oil. IR (NaCl) ν_{\max} 3090, 2942, 1716, 1076 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 4.85 (1H, t, *J* = 5.8 Hz), 3.40 (6H, s), 2.72 (2H, d, *J* = 5.8 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 175.2, 100.9, 53.5, 38.6. The data for this compound matched in all respects with that reported in the literature.³



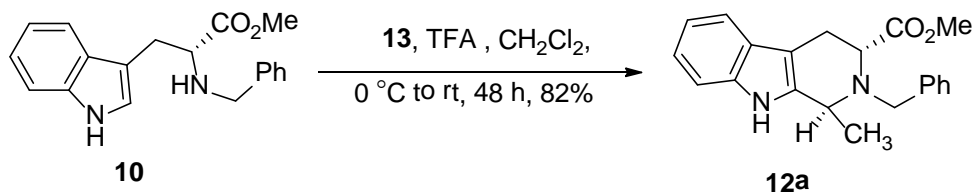
Condensation of carboxylic acid 13 to give Methyl 5,5-dimethoxy-3-oxo-4-pentanoate (14). To the carboxylic acid **13** (8.1 g, 60.2 mmol) in THF (323 mL) was added 10.8 g (66.3 mmol) of 1,1'-carbonyldiimidazole and the solution was stirred overnight at rt under argon. In a second flask potassium methyl malonate (18.7 g, 120 mmol), powdered anhydrous magnesium chloride (11.5 g, 120 mmol) and a catalytic amount of 4-(dimethylamino) pyridine (660 mg, 5.4 mmol) were vigorously stirred in a mixture of THF (323 mL) and acetonitrile (176 mL). After stirring for 2 h, the above amide in THF was added dropwise to the malonate solution. The solution which resulted was allowed to stir at rt with the help of an overhead stirrer for 12 h. The suspension which resulted was concentrated under reduced pressure. The residue was dissolved in cold aqueous 1 N HCl (500 mL) and the aqueous layer was extracted with ether (500 mL x 3). The combined organic extracts were washed with a saturated aqueous NaHCO₃ solution (500 mL), brine (250 mL) and dried (MgSO₄). The solvent was removed under

reduced pressure. The residue was subjected to flash silica gel chromatography to furnish **14** (9.7 g, 82%) as a yellow oil. IR (NaCl) ν_{\max} 2955, 1749, 1720, 1438, 1121, 1069 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ 4.78 (1H, t, $J = 5.5$ Hz), 3.75 (3H, s), 3.53 (2H, s), 3.38 (6H, s), 2.86 (2H, d, $J = 5.5$ Hz) (Proton NMR contains 8.5% of enol); ^{13}C NMR (75 MHz, CDCl_3) δ 199.6, 167.3, 101.3, 53.9, 52.3, 49.8, 46.6; HRESIMS m/z 197.0994 [$\text{M} + \text{Li}$] $^+$ (calcd for $\text{C}_8\text{H}_{14}\text{O}_5\text{Li}$, 197.1001).



Indoles **12a** & **12b**: Thermal Conditions

To a solution of optically active N_6 -benzyl-D-tryptophan methyl ester **10** (500 mg, 1.6 mmol) in dry benzene (7 mL) was added a solution of oxalacetic acid (257 mg, 2.0 mmol) in dry 1,4-dioxane (6 mL). After refluxing the above solution for 24 h with the continuous removal of water via a Dean-Stark trap, the solvent was removed under reduced pressure to provide an orange residue, which was purified by flash silica gel column chromatography to provide **12a** & **12b** (470 mg, 87% yield). The spectral data for these two β -carbolines **12a** and **12b** was identical to the published values.⁴

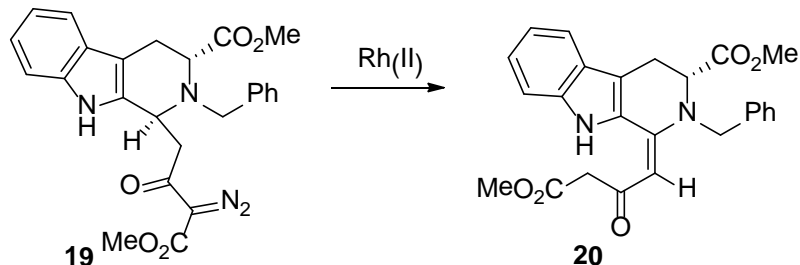


Indole **12a**: Protic Conditions

Condensation of **10 to provide (1*S*,3*R*)-Methyl-2-benzyl-1-methyl-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole-3-carboxylate (**12a**)**

To a round bottom flask which contained a solution of optically active *N*_b-benzyl-D-tryptophan methyl ester **10** (200 mg, 0.7 mmol) in dry CH₂Cl₂ (5 mL) was added 3,3-dimethoxy propionic acid **13** (174 mg, 1.3 mmol) and TFA (96 μL, 1.3 mmol) at 0 °C with stirring. The reaction mixture which resulted was stirred at rt for 48 h and then cooled in an ice bath and brought to pH 8 with a cold aqueous solution of saturated NaHCO₃. The aqueous layer was separated and extracted with CH₂Cl₂ (3 x 50 mL). The combined organic layers were washed with brine (2 x 50 mL) and dried (Na₂SO₄). Removal of the solvent under reduced pressure afforded an oil which was purified by flash silica gel column chromatography (EtOAc : hexanes) to give **12a** (177 mg, 82% yield). *R*_f 0.72 (silica gel, EtOAc/hexanes, 3 : 5); ¹H NMR (300 MHz, CDCl₃) δ 7.68 (1H, s), 7.53 (1H, d, *J* = 8.7 Hz), 7.44 (2H, d, *J* = 8.1 Hz), 7.38-7.26 (4H, m), 7.19 (1H, td, *J* = 7.1, 1.4 Hz), 7.14 (1H, td, *J* = 7.1, 1.2 Hz), 4.23 (1H, q, *J* = 6.6 Hz), 4.04 (1H, t, *J* = 6.2 Hz), 3.90 (2H, dd, *J* = 23.0, 14.4 Hz), 3.73 (3H, s), 3.10 (2H, ddd, *J* = 16.0, 10.9, 1.1 Hz), 1.48 (3H, d, *J* = 6.7 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 173.5 (C), 139.9 (C), 136.1 (2 x C), 128.3 (2 x CH), 128.2 (2 x CH), 127.0 (C), 126.9 (CH), 121.5 (CH), 119.4 (CH), 118.0 (CH), 110.6 (CH), 106.4 (C), 56.9 (CH), 53.9 (CH₂), 51.6 (CH₃), 50.9 (CH), 22.3 (CH₂), 21.1 (CH₃); EIMS *m/z* 334 [M⁺] (56), 319 (53), 275 (52), 243 (61), 183 (27), 170 (44), 157 (54), 130 (17), 115 (13), 91 (100); HRESIMS *m/z* 335.1759 (M + H)⁺ (calcd for C₂₁H₂₃N₂O₂, 335.1761). The spectral data for this compound matched in all respects with that reported in the literature.⁴

Table 1



(*R,E*)-Methyl-2-benzyl-1-(4-methoxy-2,4-dioxobutylidene)-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole-3-carboxylate (20**)**

Procedure described for Entry 1: To a solution of **19** (207 mg, 0.45 mmol) in dry benzene (6 mL) under nitrogen, rhodium(II) acetate (20 mg, 0.045 mmol) was added. The mixture was allowed to stir at rt for 4.0 h. The solution was then filtered through a pad of celite and washed with ether. The filtrate was concentrated under reduced pressure and the residue was subjected to flash silica gel column chromatography (EtOAc : hexanes) to furnish **20** (130 mg, 67% yield) as a light yellow oil. ¹H NMR (500 MHz, CDCl₃) δ 13.8 (1H, s), 7.58 (1H, d, *J* = 8.1 Hz), 7.50 (1H, d, *J* = 8.4 Hz), 7.43-7.40 (2H, m), 7.37-7.33 (3H, m), 7.31 (1H, ddd, *J* = 8.3, 7.0, 1.0 Hz), 7.14 (1H, ddd, 1H, *J* = 7.8, 7.3, 0.5 Hz), 5.55 (1H, s), 5.13 (1H, d, *J* = 16.3 Hz), 4.41-4.38 (2H, m), 3.68 (3H, s), 3.65-3.62 (4H, m), 3.50 (2H, dd, *J* = 16.2, 14.4 Hz), 3.39 (1H, dd, *J* = 16.4, 6.7 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 187.6 (C), 172.0 (C), 169.6 (C), 152.9 (C), 137.1 (C), 136.3 (C), 129.6 (CH), 129.5 (CH), 128.4 (CH), 128.2 (C), 127.4 (2 x CH), 125.5 (CH), 125.2 (C), 120.5 (CH), 119.8 (CH), 113.6 (CH), 112.6 (C), 95.2 (CH), 63.0 (CH), 57.3 (CH₂), 53.4 (CH₃), 52.6 (CH₃), 51.1 (CH₂), 24.34 (CH₂).

Entry 2: Procedure same as entry 1 except the reaction was performed at 40 °C for 1.5 h. (130 mg, 67% yield).

Entry 3: Procedure same as entry 1 except the reaction was performed at 75 °C for 2.0 h. (145 mg, 75% yield).

Entry 4: Procedure same as entry 1 except CuOTf was used as the catalyst and the reaction performed in CH₂Cl₂ as the solvent at rt for 1 h (100 mg, 51% yield).

Entry 5: Procedure same as entry 1 except CuOTf was used as the catalyst and the reaction performed in CH₂Cl₂ as the solvent, under reflux for 1 h (92 mg, 47% yield).

Entry 6: Procedure same as entry 1 except CuOTf was used as the catalyst and the reaction performed in benzene as the solvent, at rt for 2 h (100 mg, 51% yield).

Entry 7: To a solution of diazo compound **19** (50 mg, 0.11 mmol) in dry CH₂Cl₂ (4 mL) was added dropwise tetrafluoroboric acid diethylether complex (70 mg, 0.44 mmol) under an atmosphere of argon at 0 °C (ice bath). The reaction mixture was stirred at rt for more than 24 h. During this period regular analysis by TLC indicated presence of starting material **19** and an increase in baseline formation. After 33 h TLC analysis indicated presence of only baseline impurities. The reaction mixture was neutralized with an ice cold aqueous solution of saturated NaHCO₃. The two layers were separated, the organic layer was washed with brine, dried (Na₂SO₄) and the solvent concentrated under reduced pressure to give a brown residue. The same result was observed with 8 eq of HBF₄•OEt₂. Reaction was stopped after 24 h.

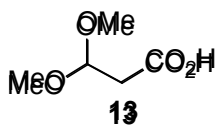
Entry 8: Procedure same as entry 7, except the reaction was performed under refluxing condition for 4 h. Only baseline impurities were observed.

Entry 9: Under a balloon of argon, diazo compound **19** (50 mg, 0.11 mmol) was dissolved in CCl₄ (2 mL) and the vial was placed in between two UV lamps. The mixture was irradiated with UV light (shortwave $\lambda \geq 254$ nm) for 12 h at rt. After 12 h analysis by TLC (silica gel, EtOAc/hexanes, 2 : 3) indicated presence of starting material **19** and baseline impurities.

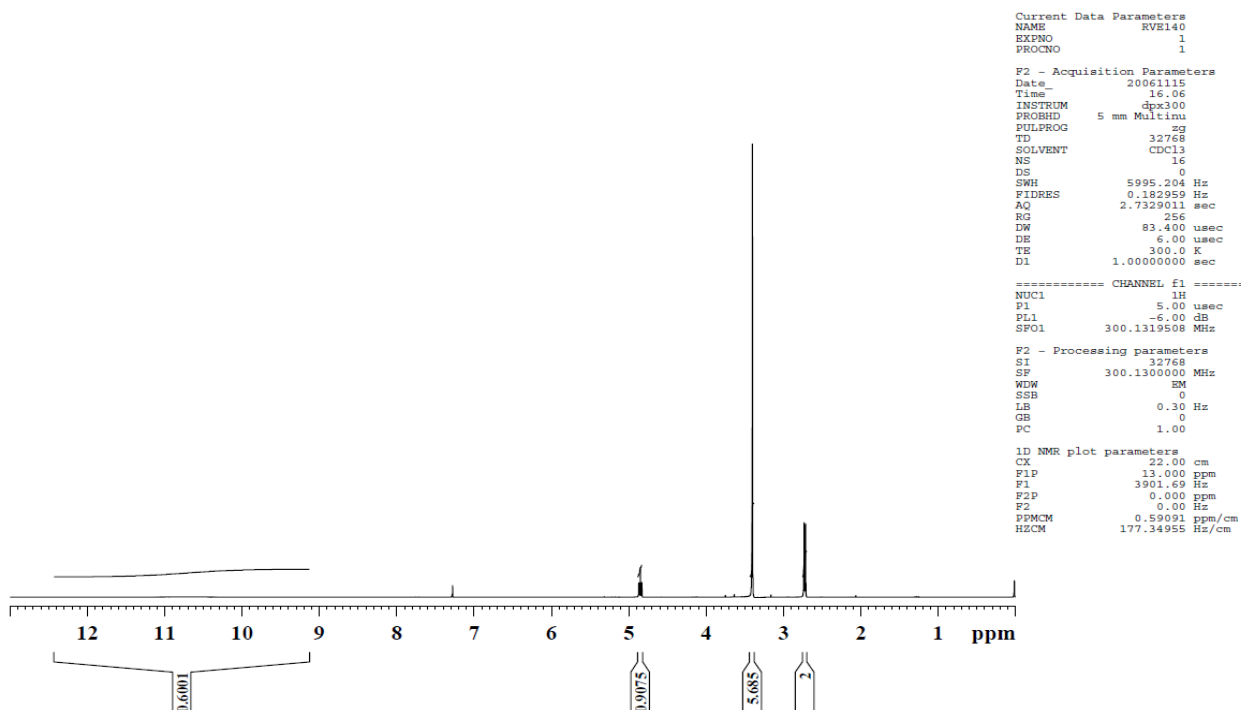
References:

- 1) Li, J. J.; Limberakis, C.; Pflum, D. A. *Modern Organic Synthesis in the Laboratory*, Oxford University Press, Inc: New York, NY, 2007, Chapter 1, pp 17-20.
- 2) Compound **13** was synthesized following a procedure by: Zaragoza, F.; Stephensen, H.; Peschke, B.; Rinvall, K. *J. Med. Chem.* **2005**, *48*, 306-311.
- 3) Valade, A.-G.; Dugat, D.; Jeminet, G.; Royer, J.; Husson, H.-P. *Eur. J. Org. Chem.* **2001**, 2041-2053.
- 4) Sandrin, J.; Hollinshead, S. P.; Cook, J. M. *J. Org. Chem.* **1989**, *54*, 5636-5640.

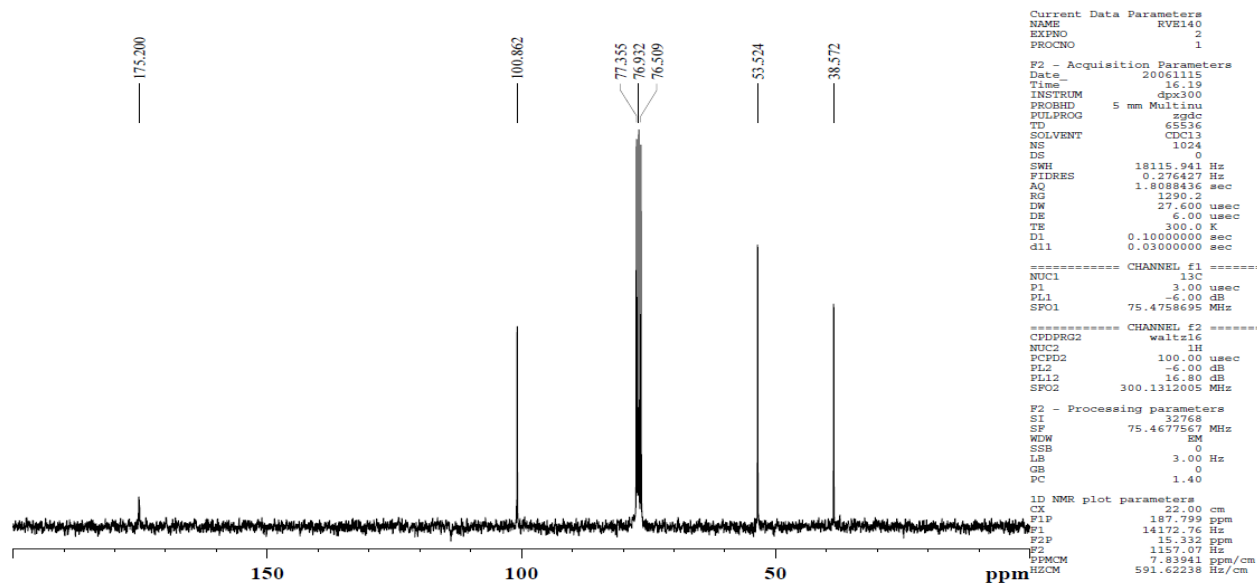
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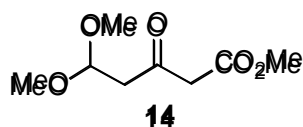


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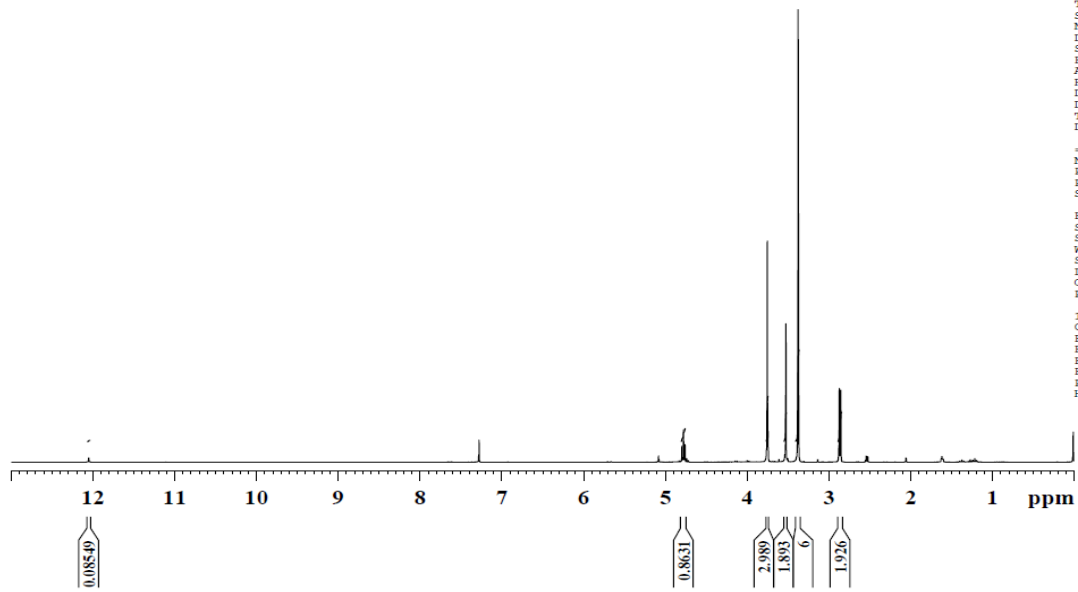


Carbon Spectrum





1D NMR Spectrum: Methyl-2,2-dimethoxy-3-oxopentanoate



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PROCNO        1

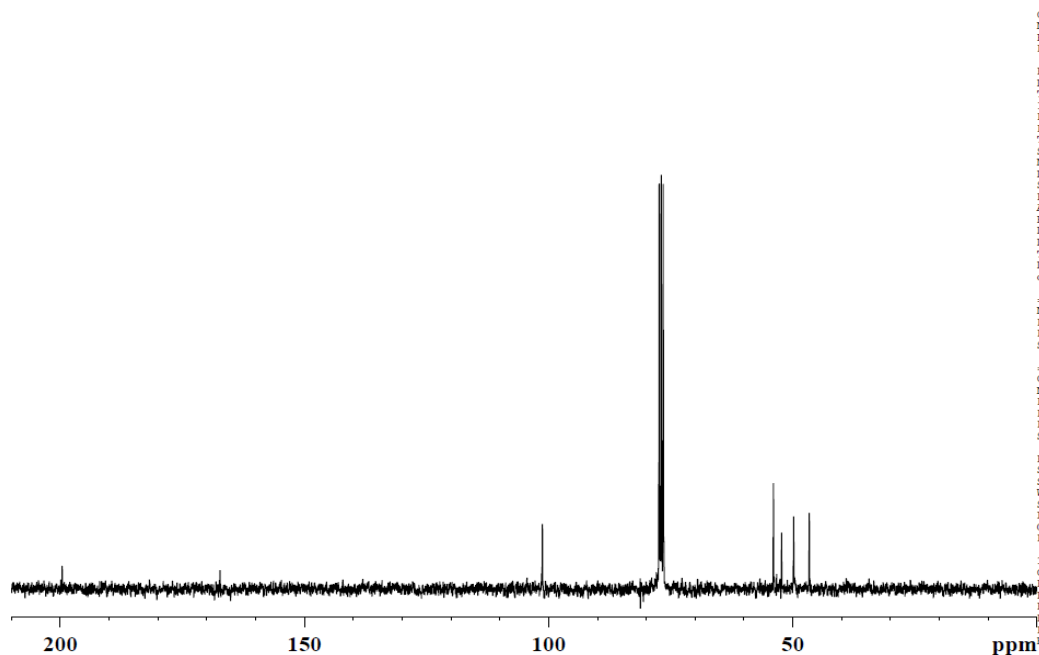
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Carbon Spectrum



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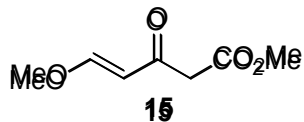
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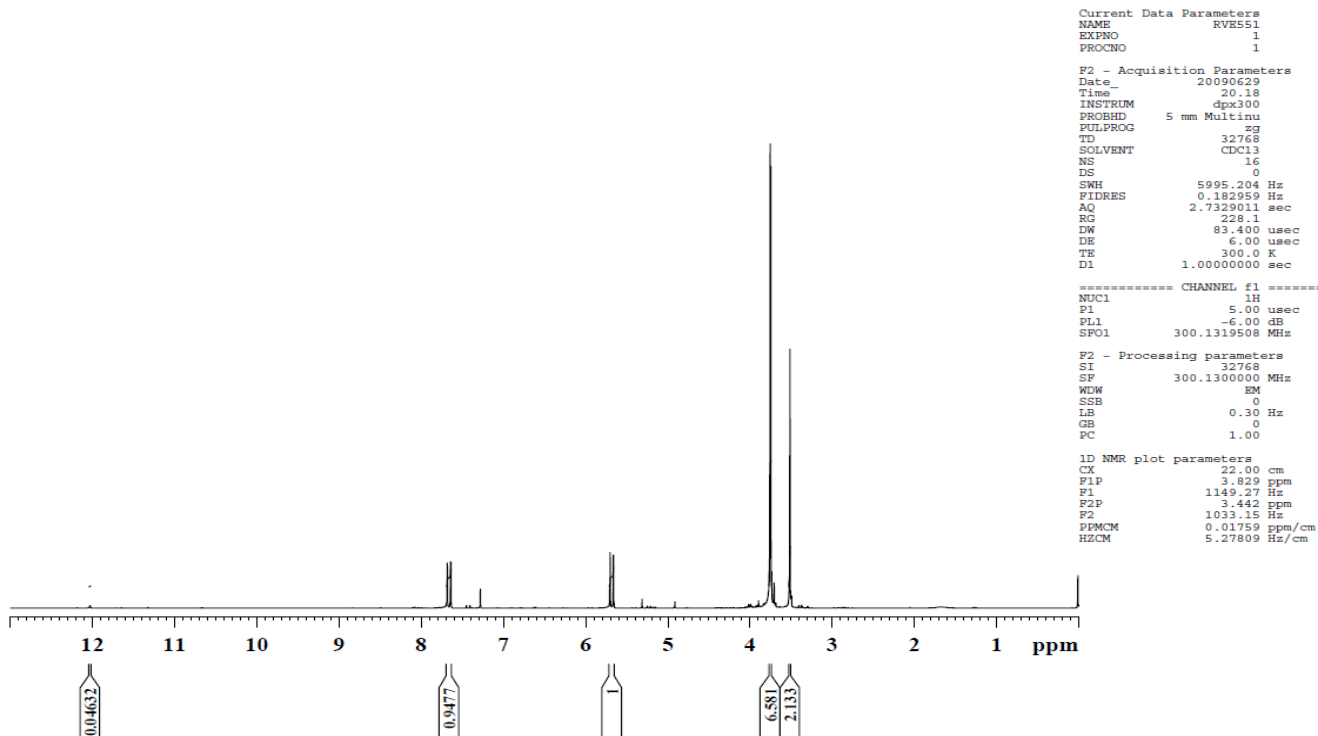
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Proton Spectrum



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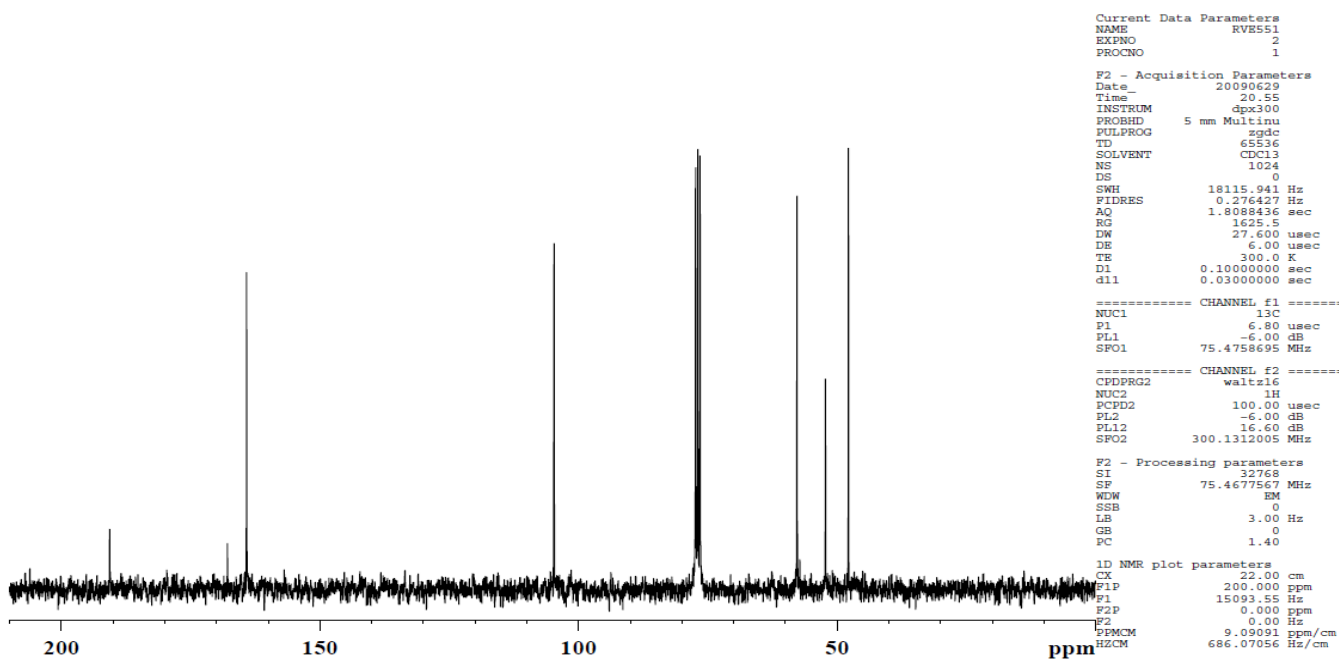
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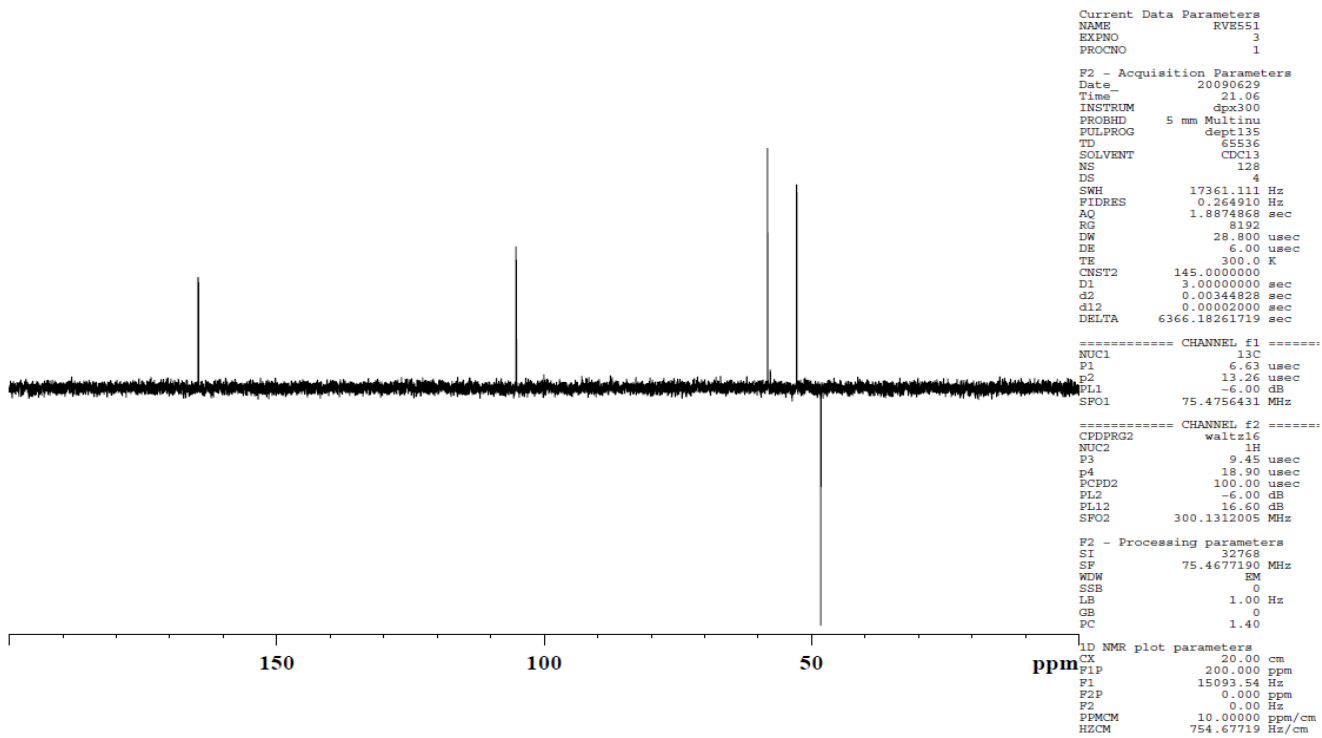
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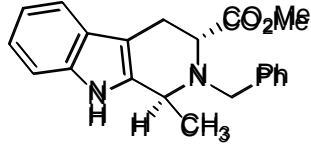
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Carbon Spectrum



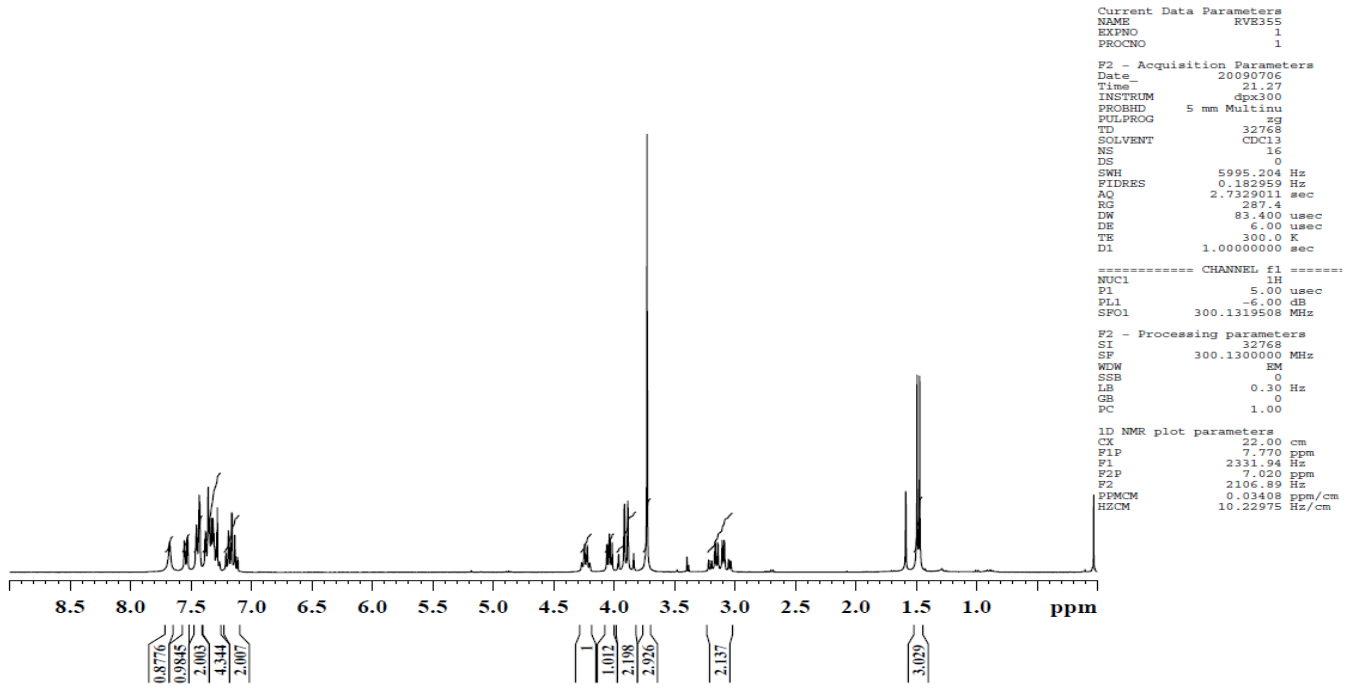
C-13 DEPT 135



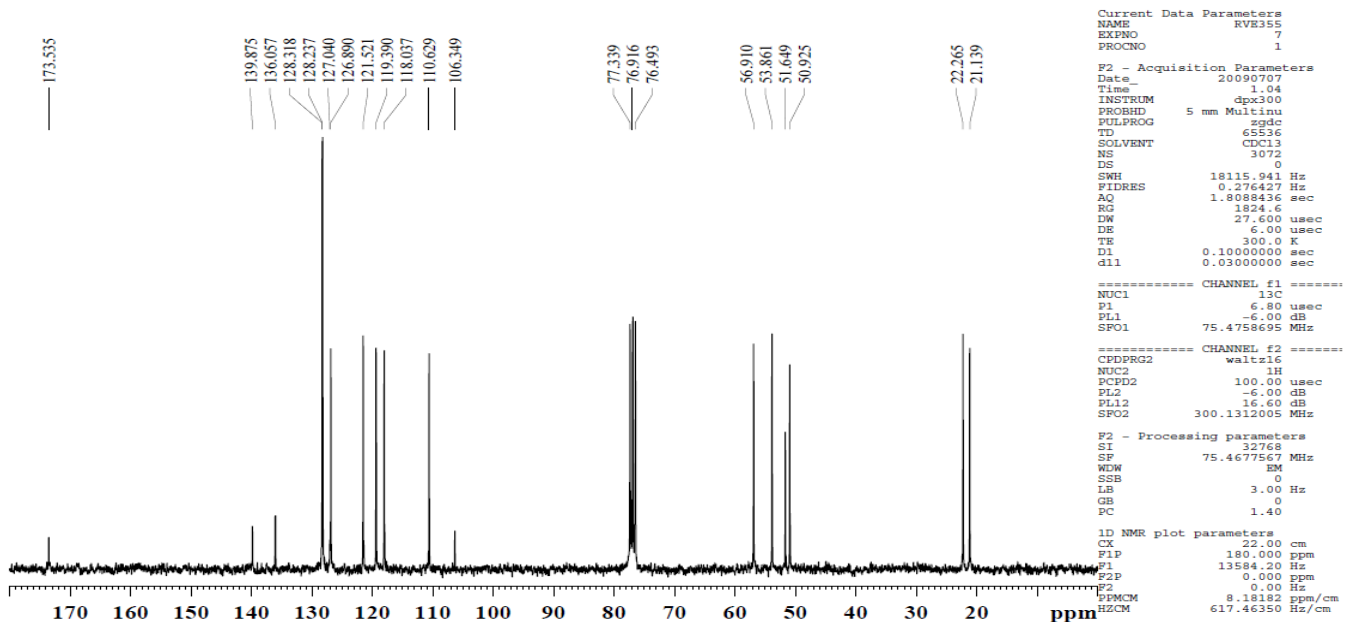


12a

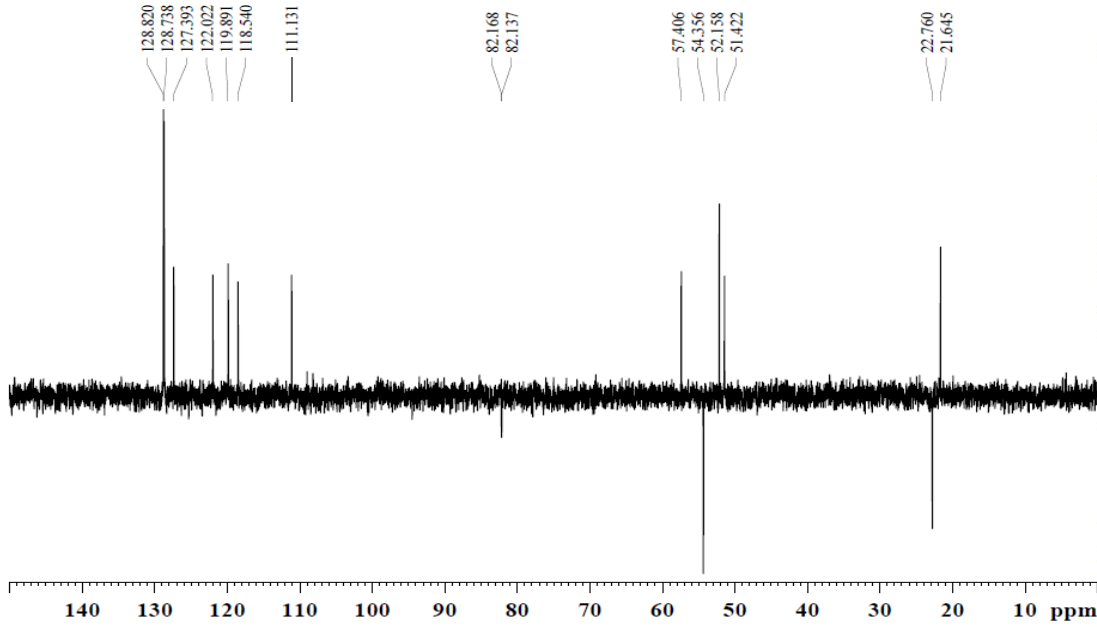
Proton Spectrum



Carbon Spectrum



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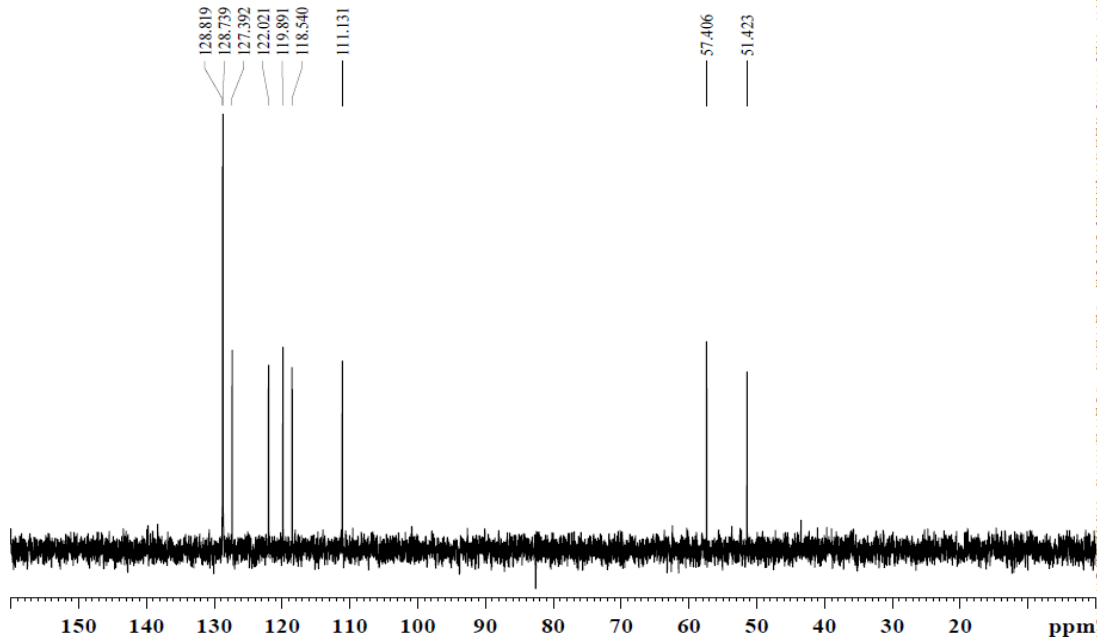
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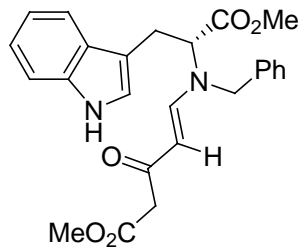
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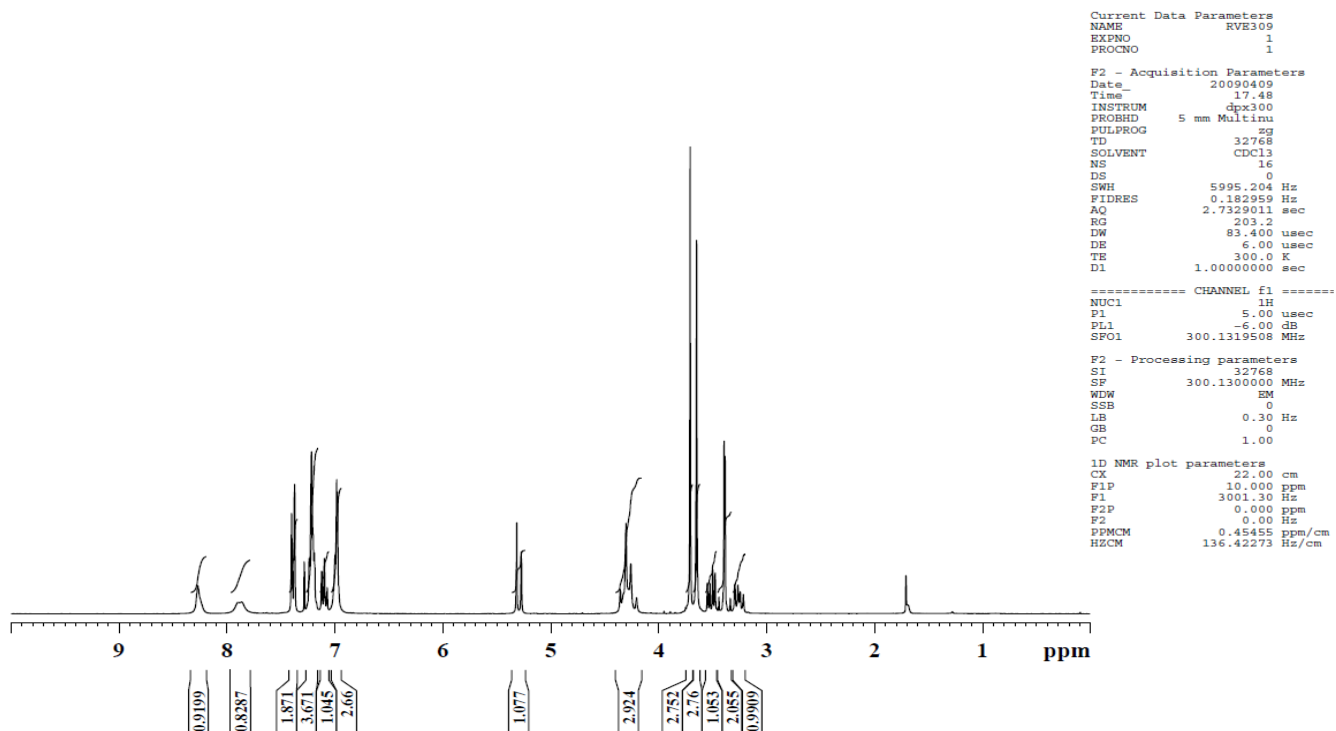
F2 - Processing parameters
SI         32768
SF         75.4677190 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

ID NMR plot parameters
CX         20.00 cm
F1P        150.000 ppm
F1         11320.16 Hz
F2P        0.000 ppm
F2         0.00 Hz
PPMCM      7.50000 ppm/cm
HZCM       566.00761 Hz/cm
    
```

16

Proton Spectrum



```

Current Data Parameters
NAME           RVE309
EXPNO          1
PROCNO         1

F2 - Acquisition Parameters
Date_          20090409
Time           17.48
INSTRUM        dpx300
PROBHD         5 mm Multinu
PULPROG        zg
TD             32768
SOLVENT        CDCl3
NS            16
DS            0
SWH           5995.204 Hz
FIDRES        0.182959 Hz
AQ            2.7329011 sec
RG            203.2
DW            83.400 usec
DE            6.00 usec
TE            300.0 K
D1            1.0000000 sec

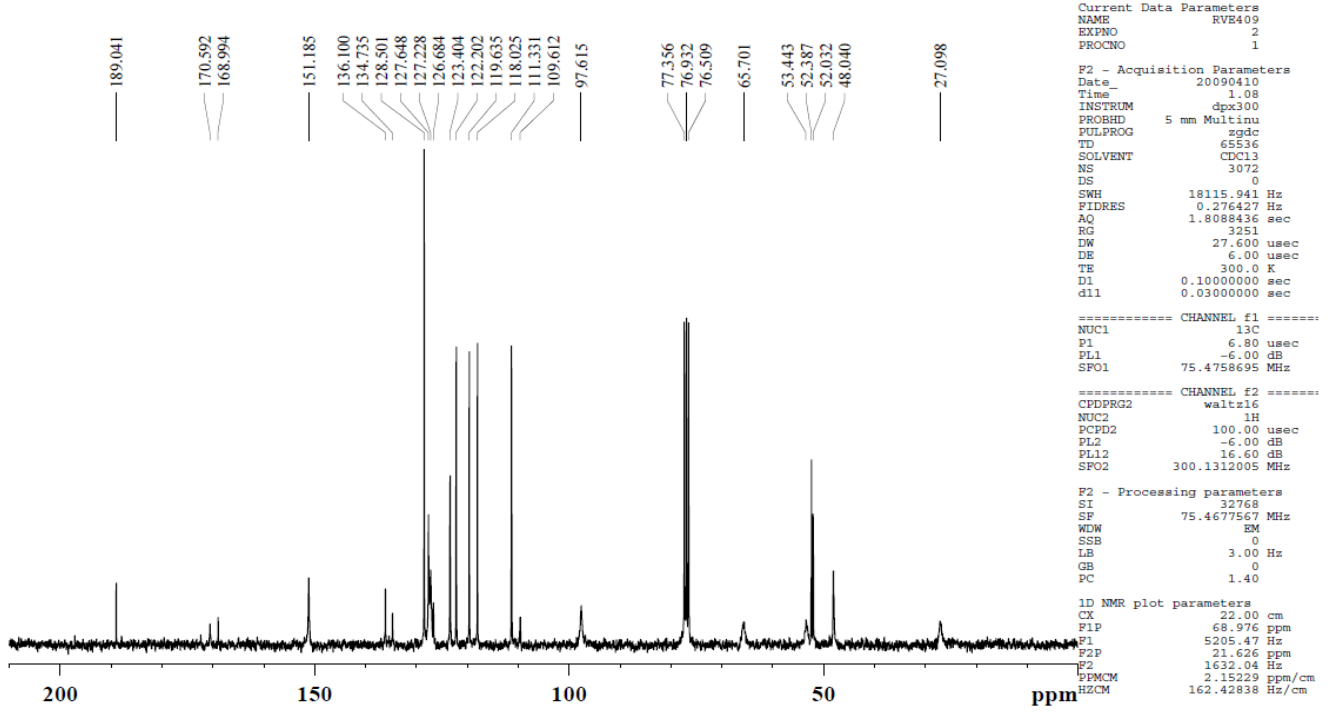
===== CHANNEL f1 =====
NUC1           1H
P1            5.00 usec
PL1           -6.00 dB
SFO1          300.1319508 MHz

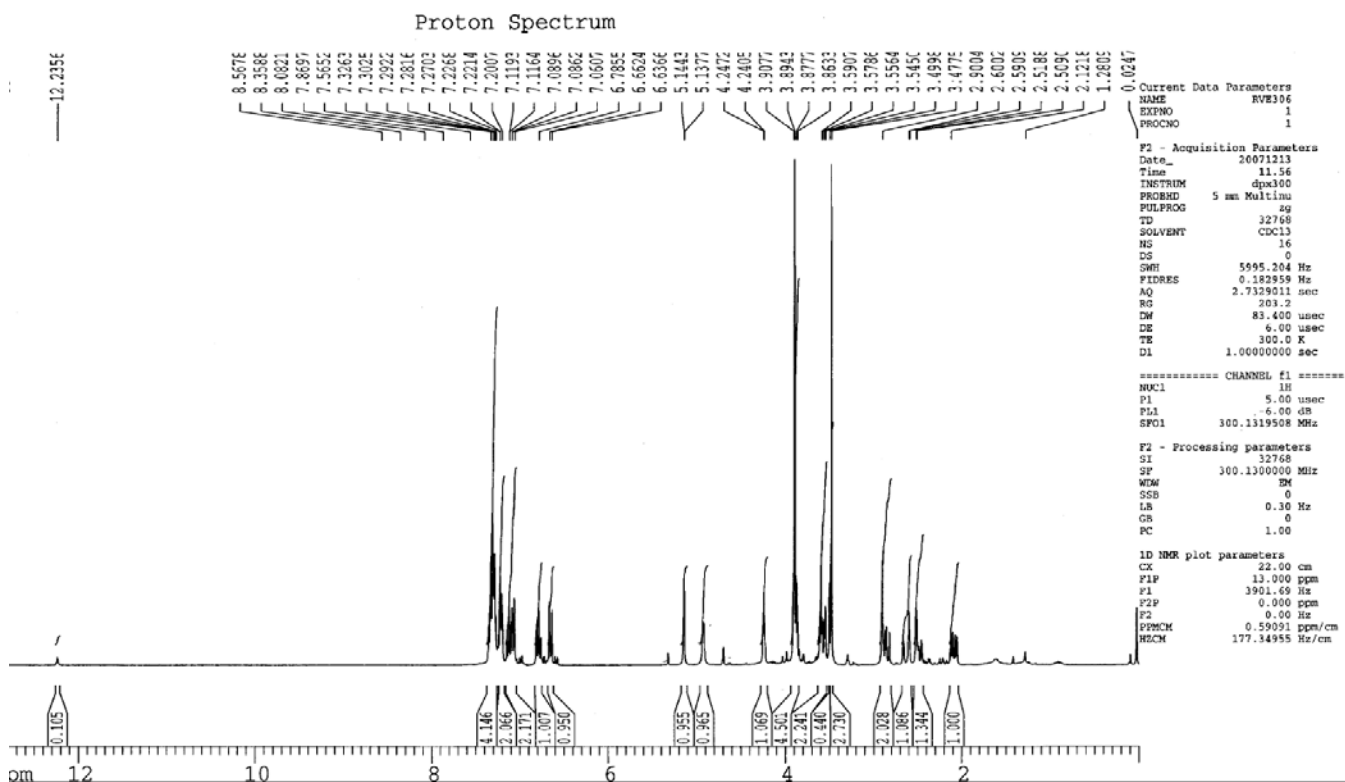
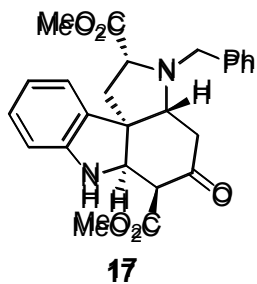
F2 - Processing parameters
SI            32768
SF           300.1300000 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00

1D NMR plot parameters
CX            22.00 cm
FIP           10.000 ppm
F1            3001.30 Hz
F2            0.000 ppm
F2            0.00 Hz
PPMCM         0.45455 ppm/cm
HZCM          136.42273 Hz/cm

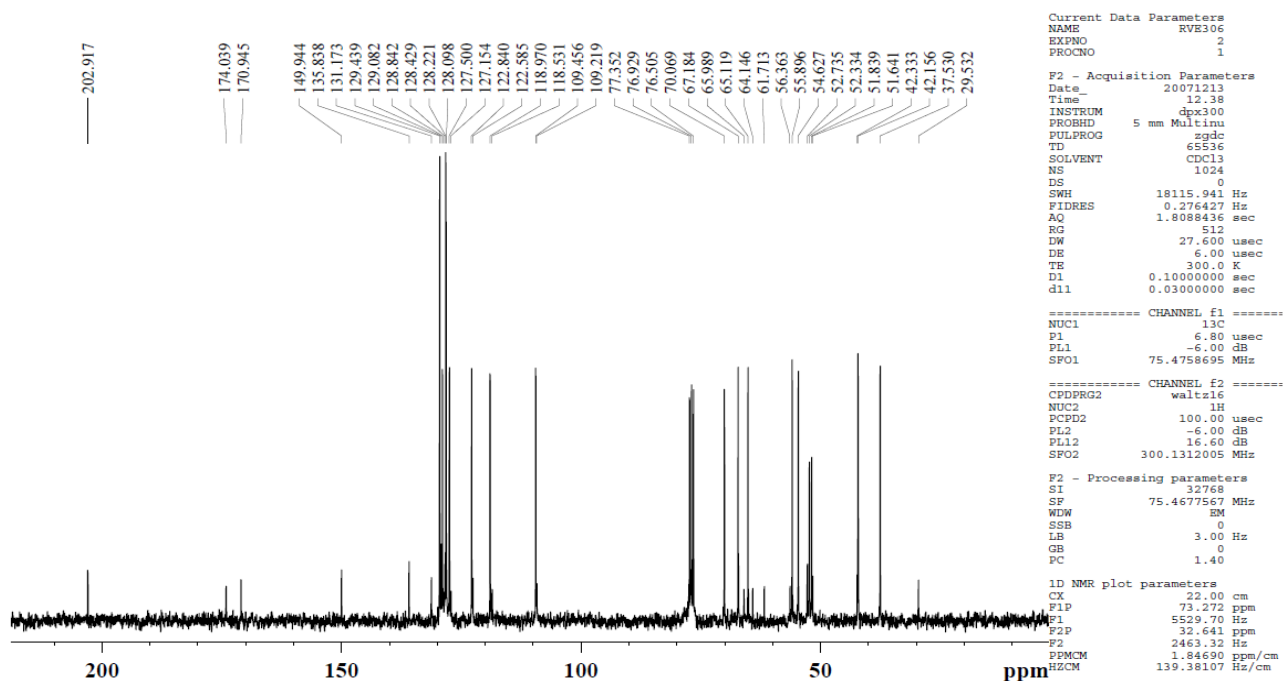
```

Carbon Spectrum

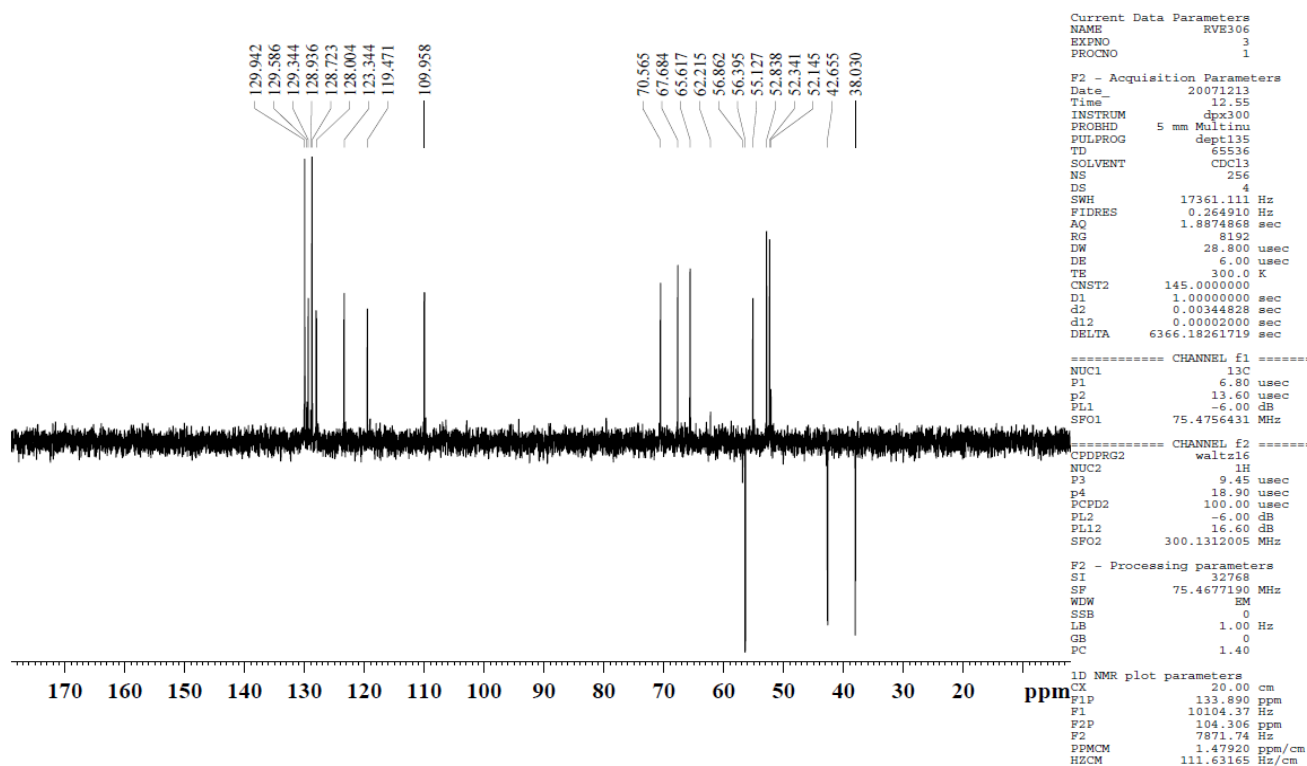




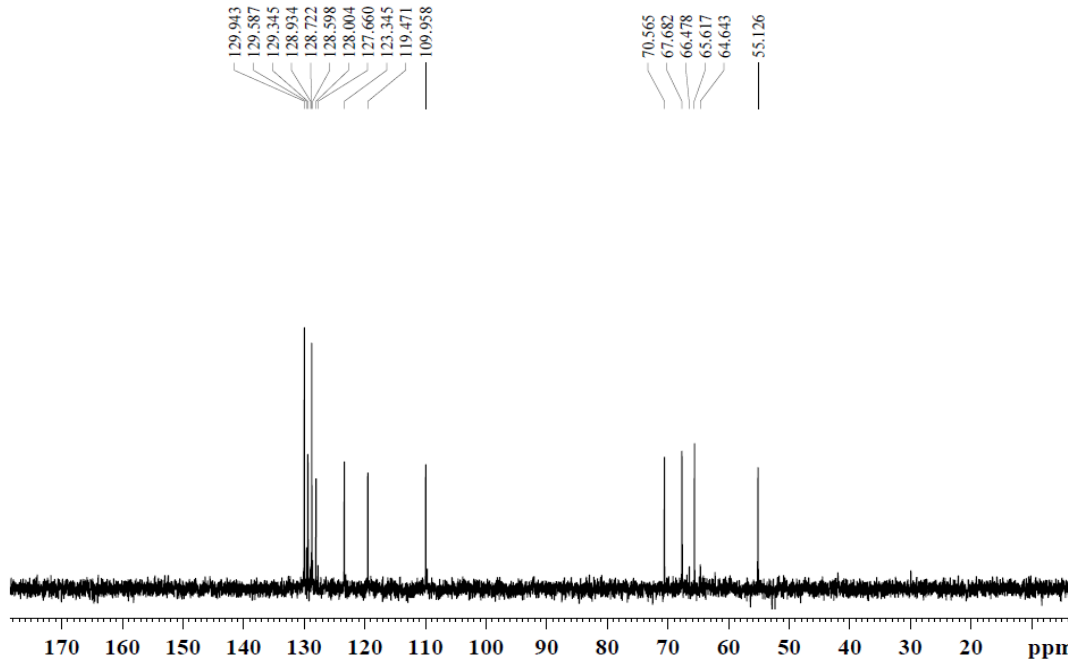
Carbon Spectrum



C-13 DEPT 135



C13 DEPT 90



```

Current Data Parameters
NAME          RVE306
EXPNO         4
PROCNO        1

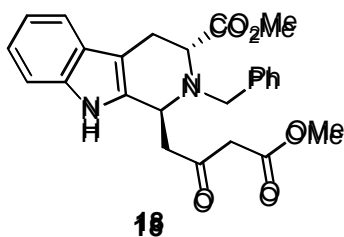
F2 - Acquisition Parameters
Date_         20071213
Time          13.09
INSTRUM       dpx300
PROBHD        5 mm Multinu
PULPROG       dept90
TD            65536
SOLVENT       CDCl3
NS            256
DS            4
SWH           23809.523 Hz
FIDRES        0.363304 Hz
AQ            1.3763061 sec
RG            7298.2
DW            21.000 usec
DE            6.00 usec
TE            300.0 K
CNST2         145.0000000
D1            1.0000000 sec
d2            0.00344828 sec
d12           0.00002000 sec
DELTA         6366.18261719 sec

===== CHANNEL f1 =====
NUC1          13C
P1            6.80 usec
P2            13.60 usec
PL1           -6.00 dB
SFO1          75.4772501 MHz

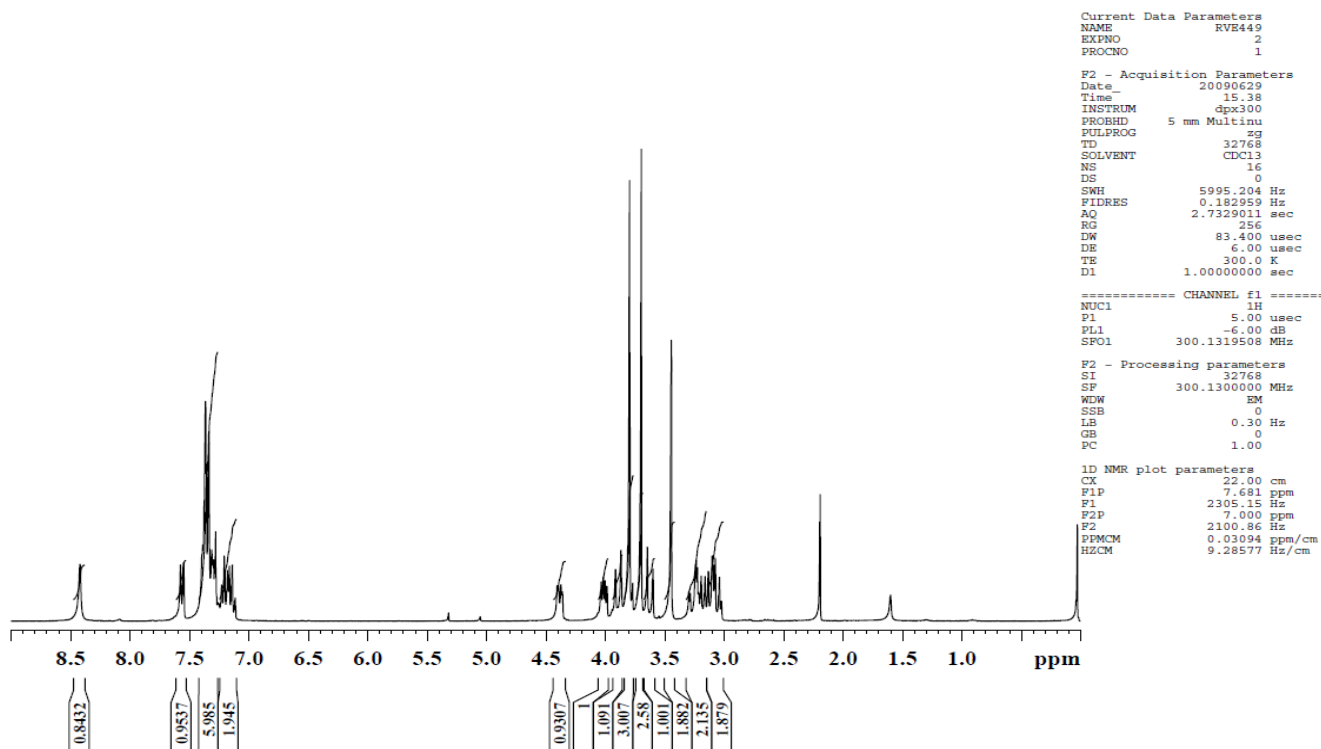
===== CHANNEL f2 =====
CDDPRG2       waltz16
NUC2          1H
P3            9.45 usec
P4            18.90 usec
PCPD2         100.00 usec
PL2           -6.00 dB
PL12          16.60 dB
SFO2          300.1312005 MHz

F2 - Processing parameters
SI            32768
SF            75.4677190 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            1.40

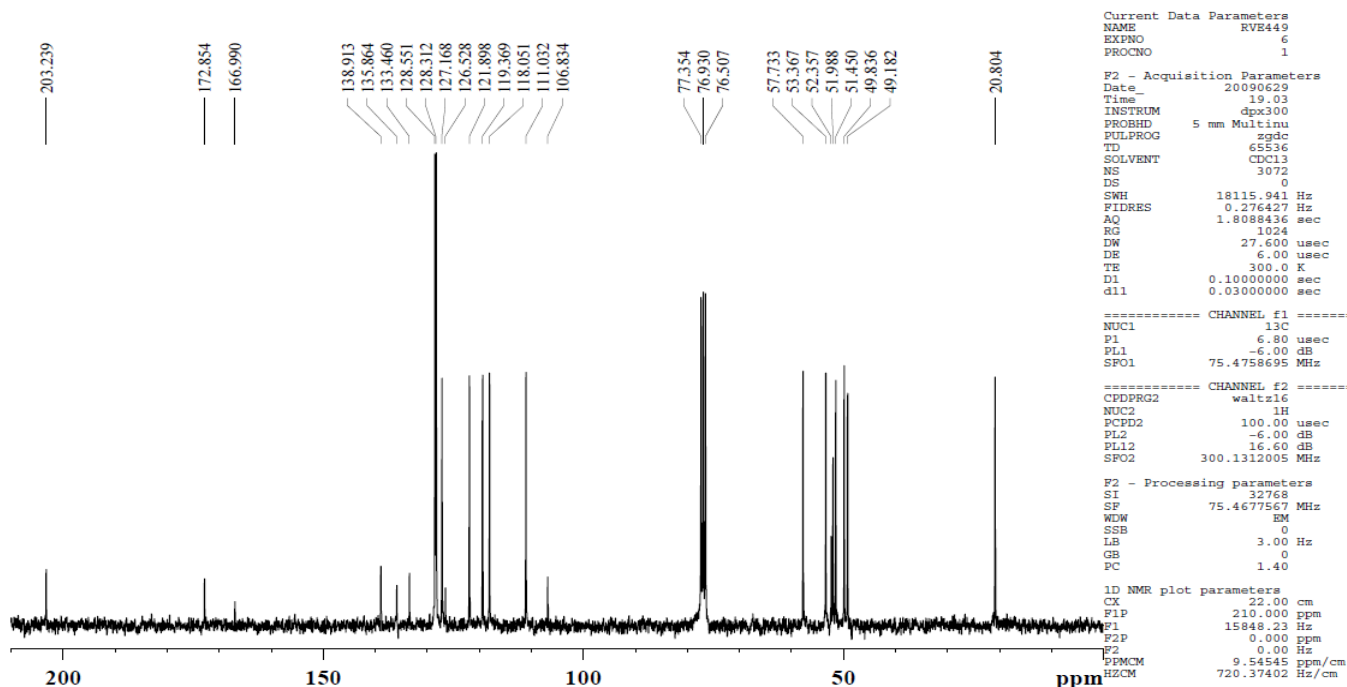
1D NMR plot parameters
CX            20.00 cm
F1P           133.480 ppm
F1            10073.43 Hz
F2P           105.816 ppm
F2            7985.71 Hz
PFMCM         1.38319 ppm/cm
HZCM          104.38629 Hz/cm
    
```



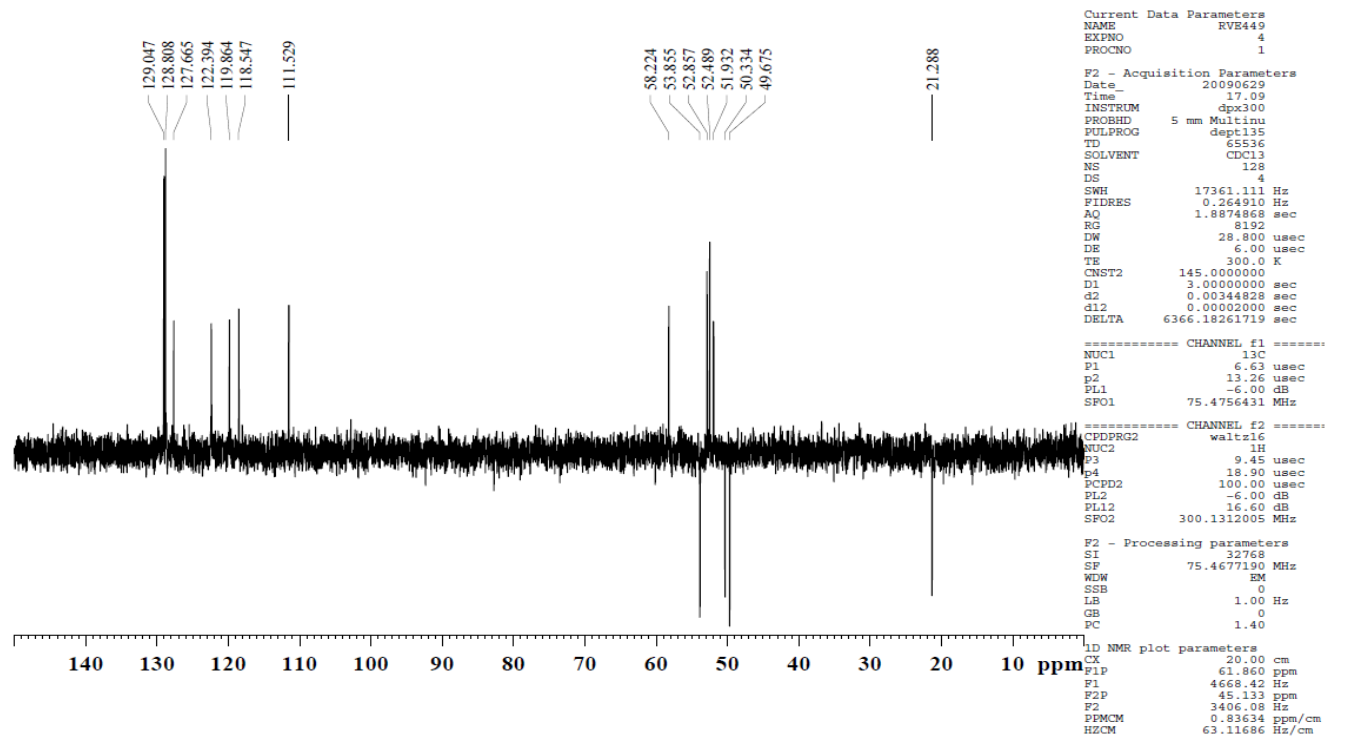
Proton Spectrum



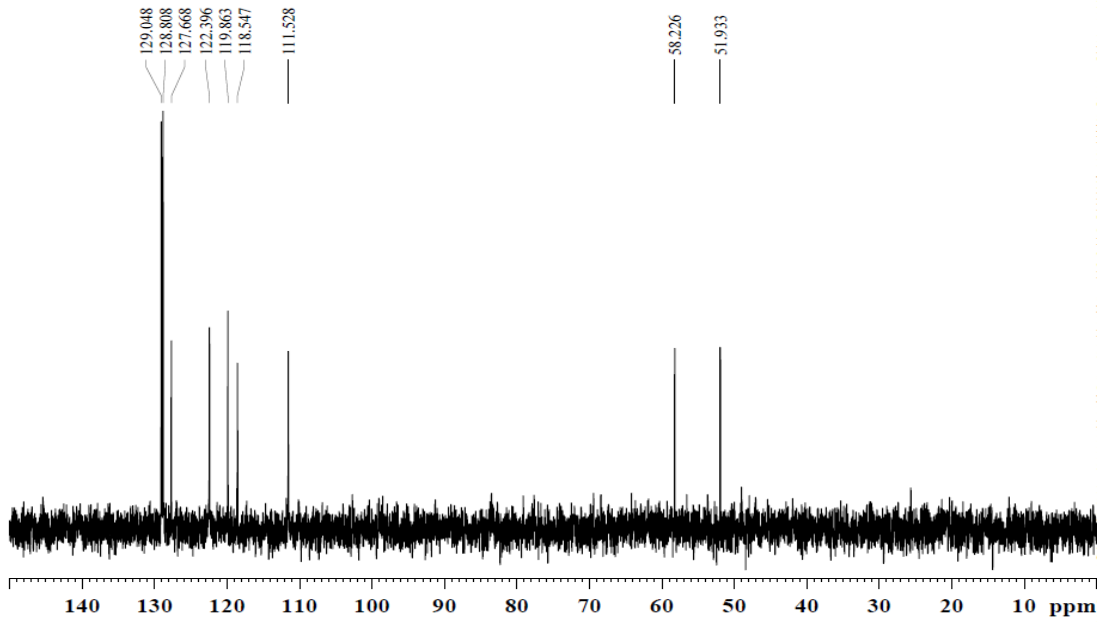
Carbon Spectrum



C-13 DEPT 135



C13 DEPT 90



```

Current Data Parameters
NAME          RV5449
EXPNO         5
PROCNO        1

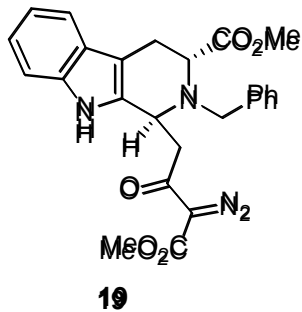
F2 - Acquisition Parameters
Date_         20090629
Time          17.18
INSTRUM       dpx300
PROBHD        5 mm Multinu
PULPROG       dept90
TD             65536
SOLVENT       CDCl3
NS             128
DS             4
SWH            23809.523 Hz
FIDRES         0.363304 Hz
AQ             1.3763061 sec
RG             6502
DW             21.000 usec
DE             6.00 usec
TE             300.0 K
CNST2         145.0000000
D1             3.0000000 sec
d12            0.00344828 sec
d12            0.0002000 sec
DELTA         6366.18261719 sec

===== CHANNEL f1 =====
NUC1           13C
P1             6.63 usec
P2            131.26 usec
PL1            -6.00 dB
SFO1           75.4772501 MHz

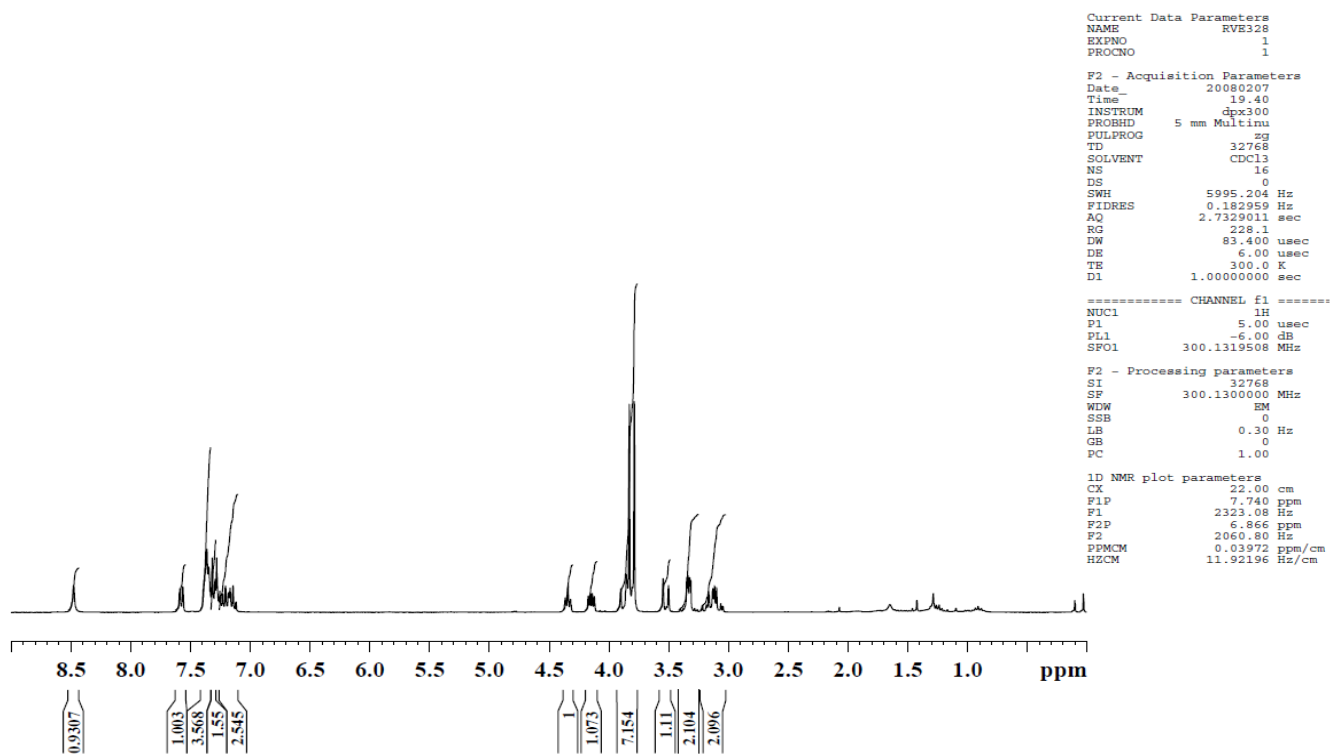
===== CHANNEL f2 =====
CFDPRG2       waltz16
NUC2           1H
P3             9.45 usec
P4            18.90 usec
PCPD2         100.00 usec
PL2            -6.00 dB
PL12           16.60 dB
SFO2           300.1312005 MHz

F2 - Processing parameters
SI             32768
SF             75.4677190 MHz
RGW            EM
SSB            0
LB             1.00 Hz
GB             0
PC             1.40

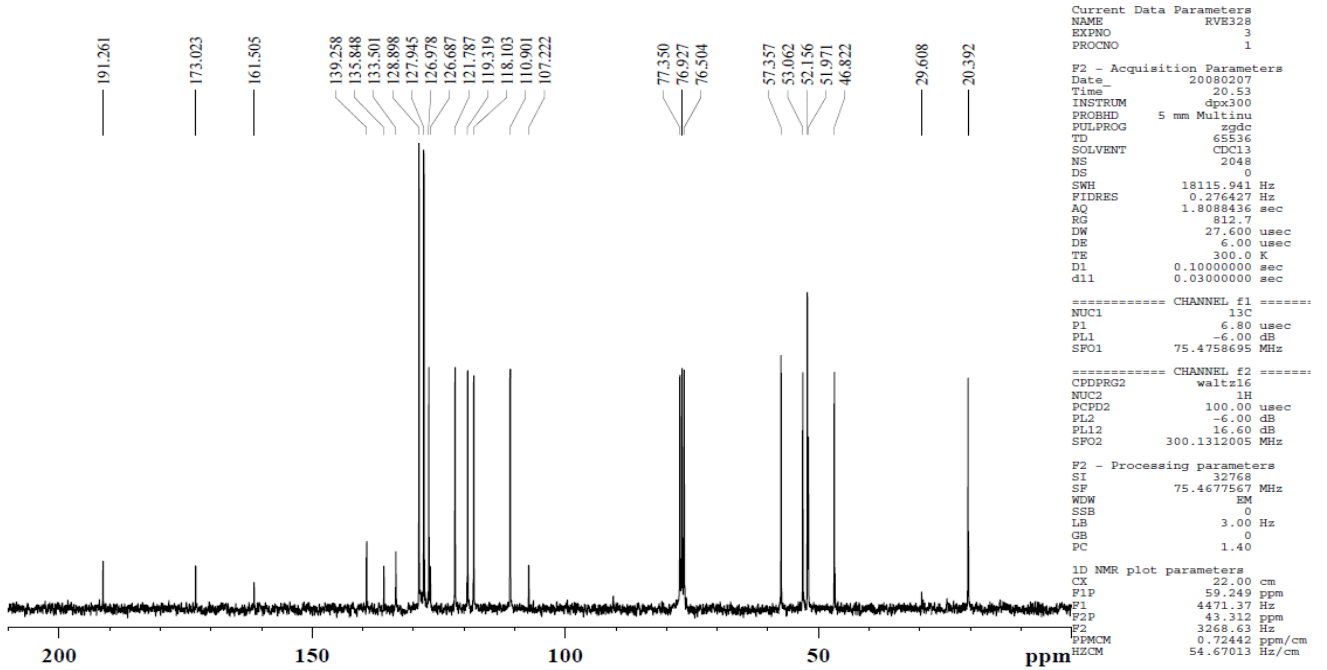
ID NMR plot parameters
CX             20.00 cm
F1P            132.665 ppm
F1             10011.89 Hz
F2P            106.899 ppm
F2             8067.42 Hz
PPMCM         1.28828 ppm/cm
HZCM          97.22371 Hz/cm
    
```

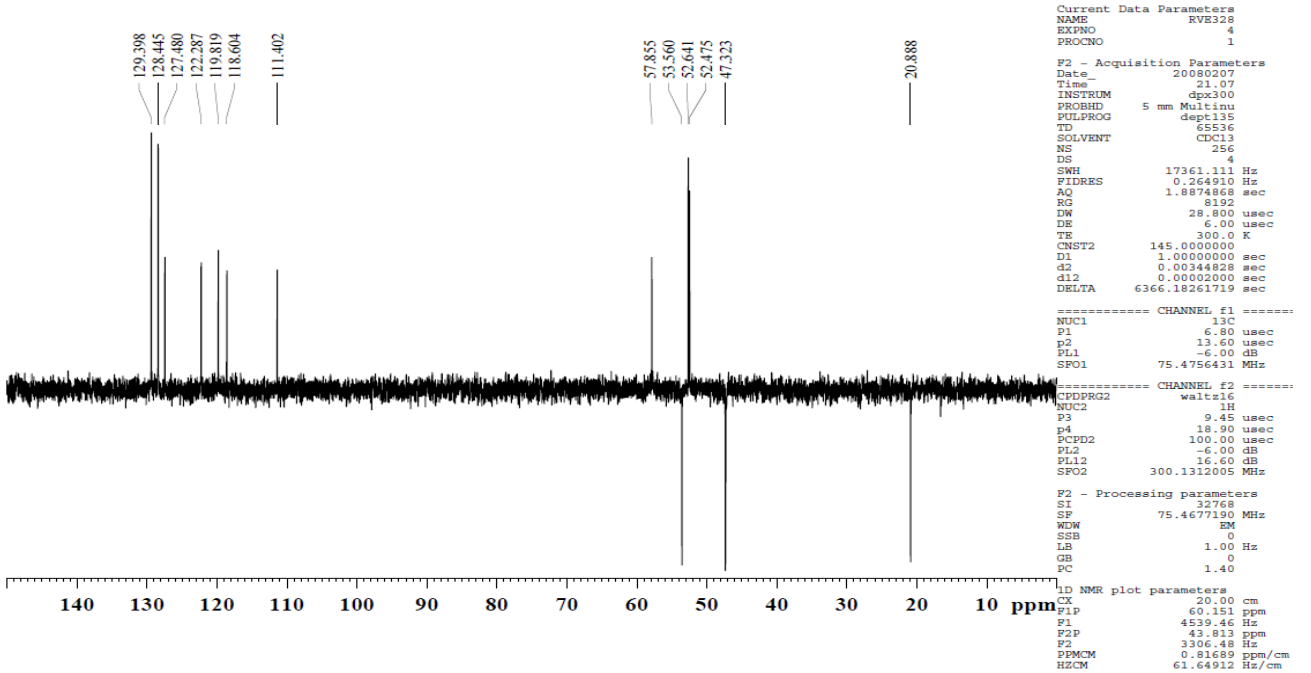
Proton Spectrum



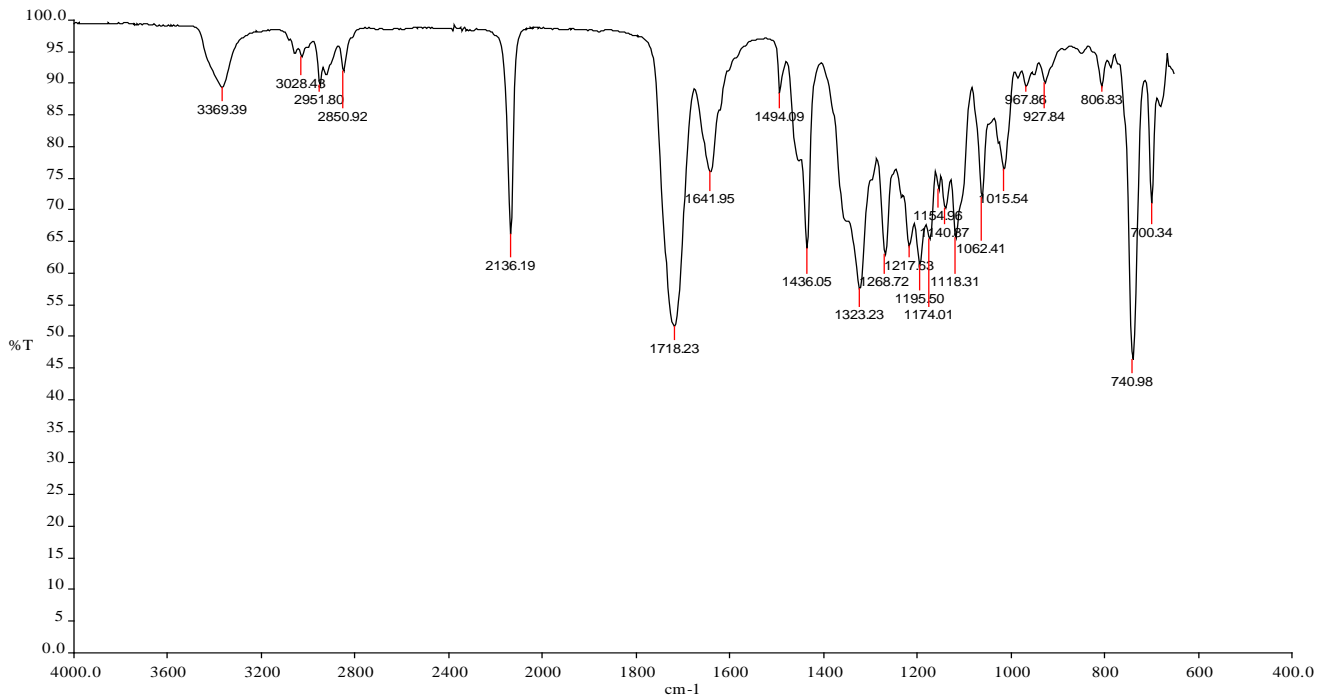
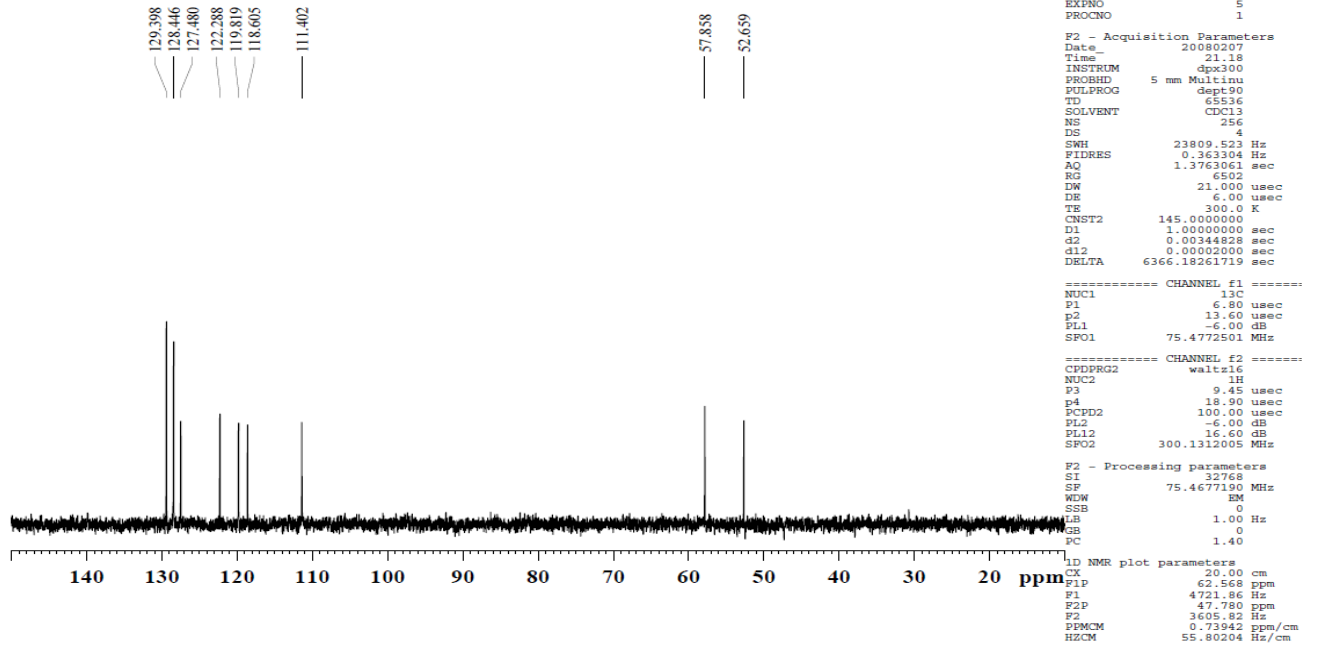
Carbon Spectrum

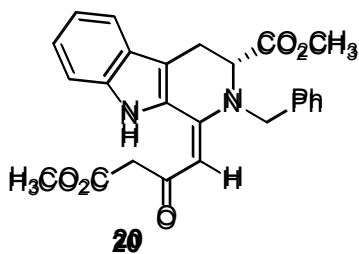


C-13 DEPT 135



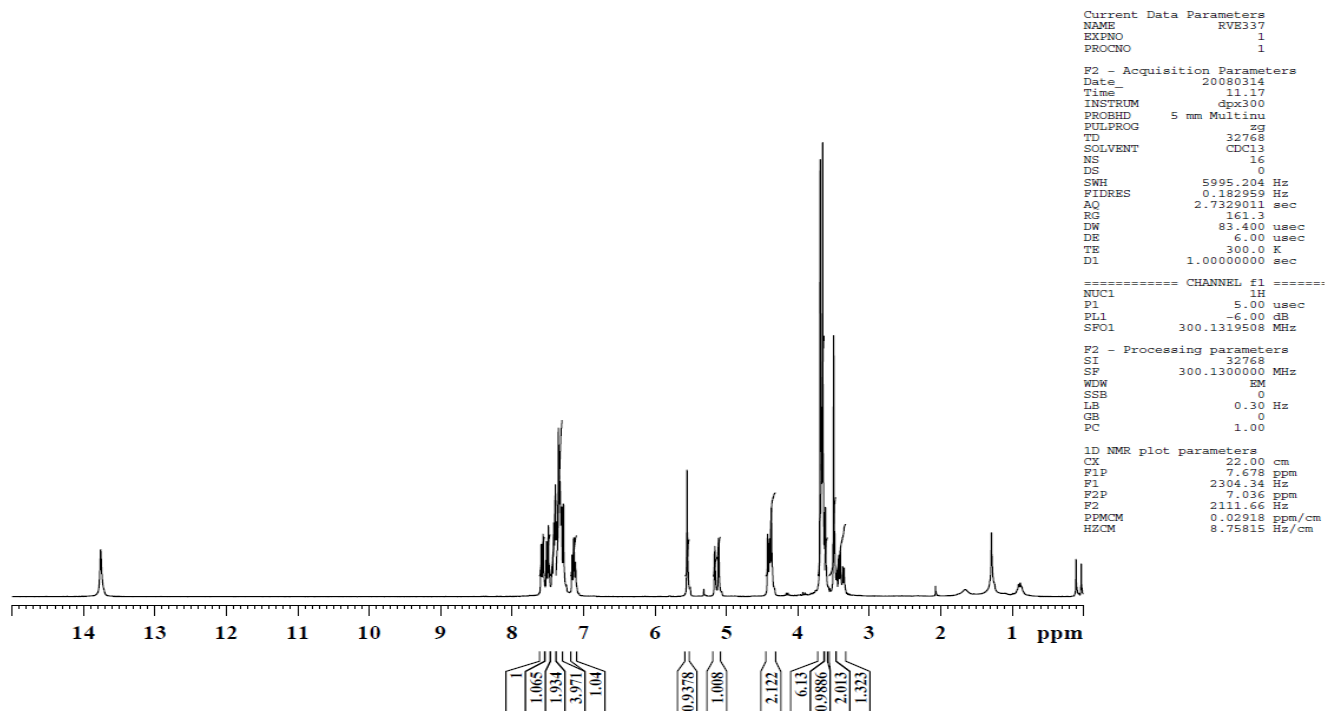
C13 DEPT 90



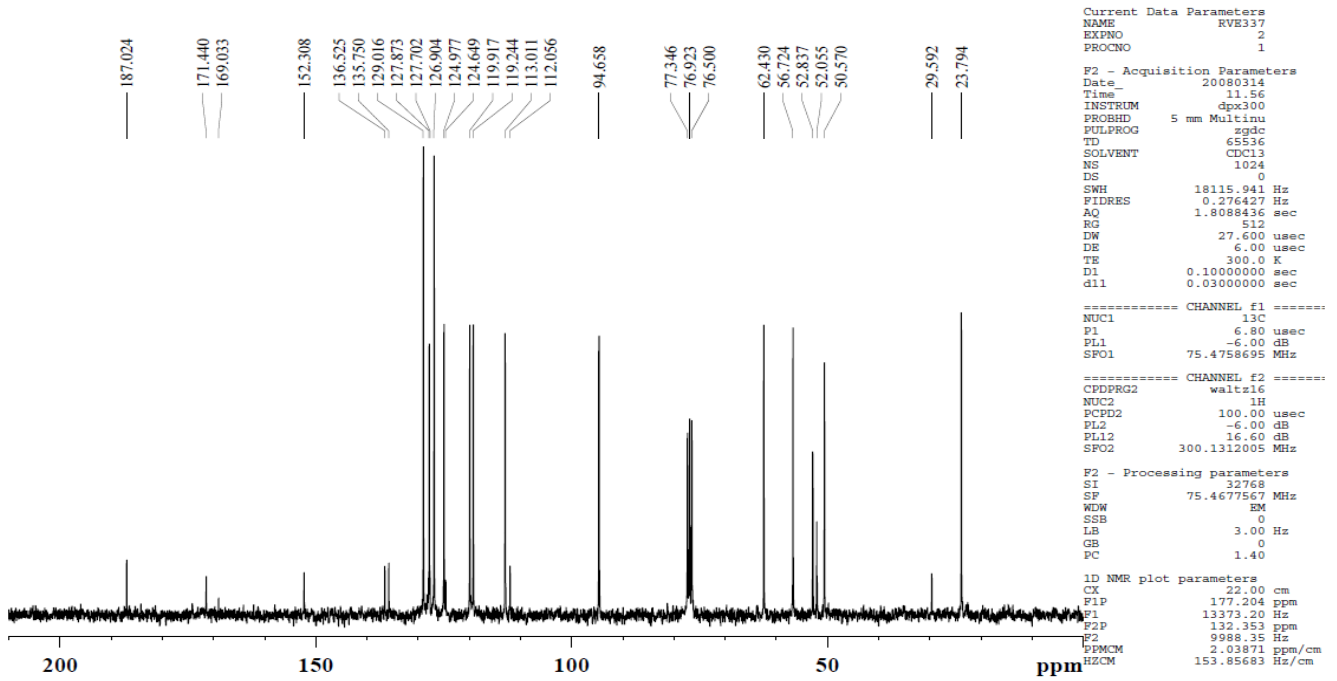


Compound 20 (300 MHz)

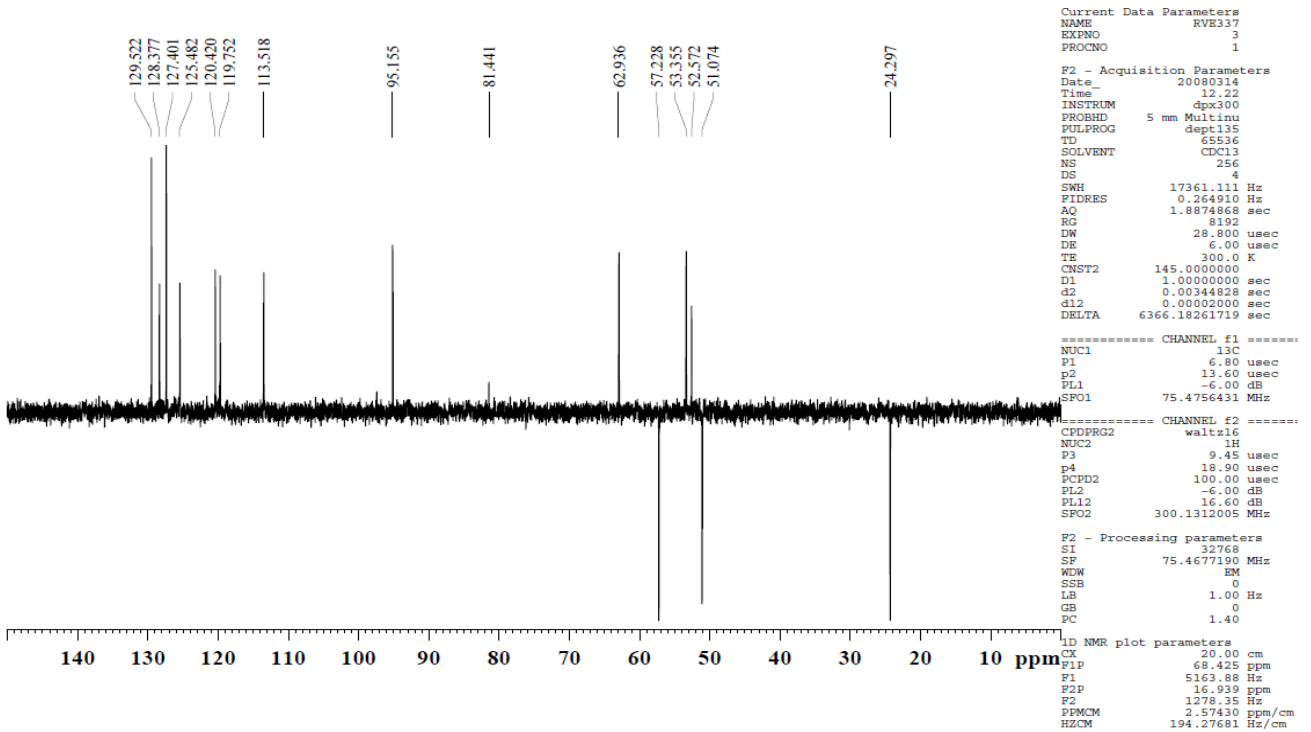
Proton Spectrum



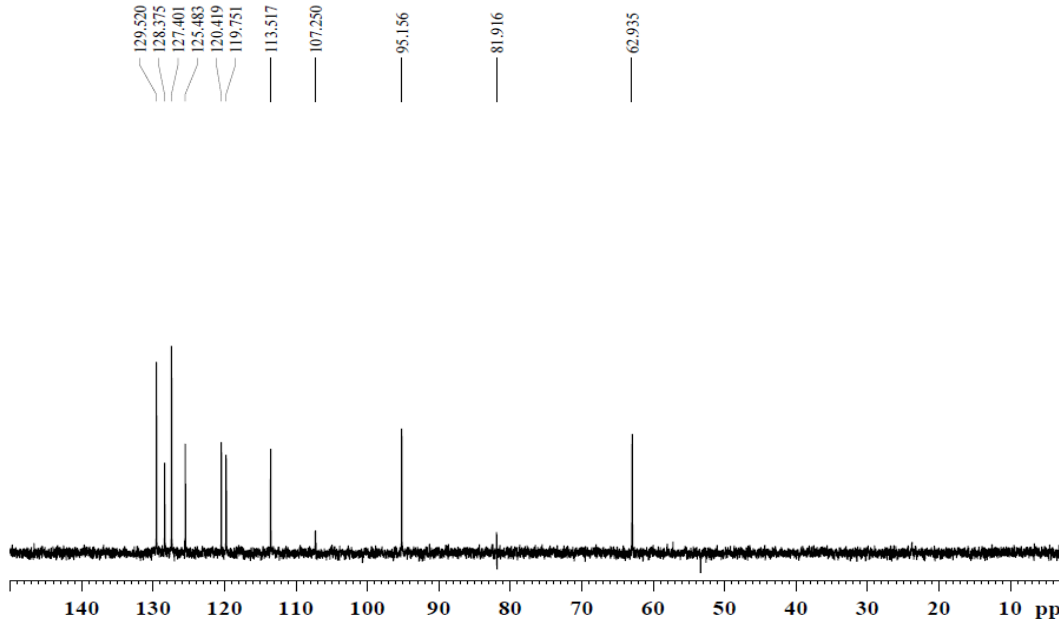
Carbon Spectrum



C-13 DEPT 135



C13 DEPT 90



```

Current Data Parameters
NAME      RVE337
EXPNO     4
PROCNO    1

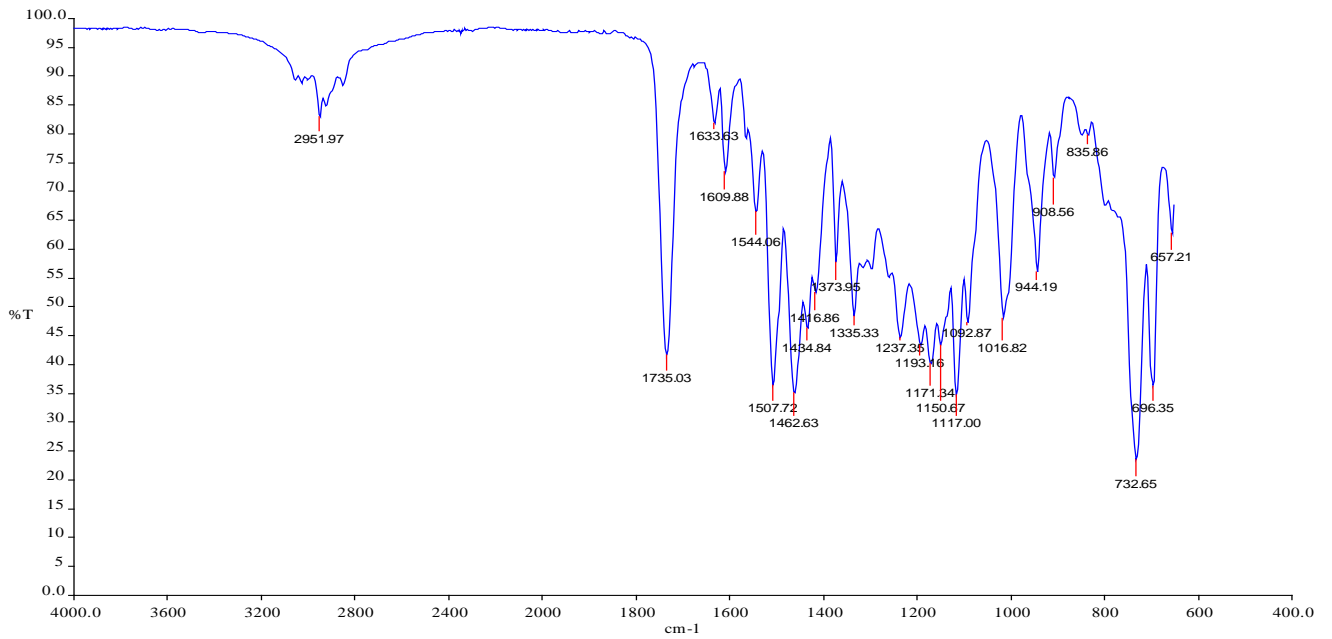
F2 - Acquisition Parameters
Date_     20080314
Time      12.33
INSTRUM   dpx300
PROBHD    5 mm Multinu
PULPROG   dept90
TD         65536
SOLVENT   CDC13
NS         256
DS         4
SWH        23809.523 Hz
FIDRES     0.363304 Hz
AQ         1.3763061 sec
RG         6502
DW         21.000 usec
DE         6.00 usec
TE         300.0 K
CNST2     145.0000000
D1         1.000000000 sec
d2         0.00344828 sec
d12        0.000020000 sec
DELTA     6366.18261719 sec

===== CHANNEL f1 =====
NUC1      13C
P1        6.80 usec
P2        13.60 usec
PL1       -6.00 dB
SFO1      75.4772501 MHz

===== CHANNEL f2 =====
NUC2      1H
P3        9.45 usec
P4        18.90 usec
PCPD2     100.00 usec
PL2       -6.00 dB
PL12      16.60 dB
SFO2      300.1312005 MHz

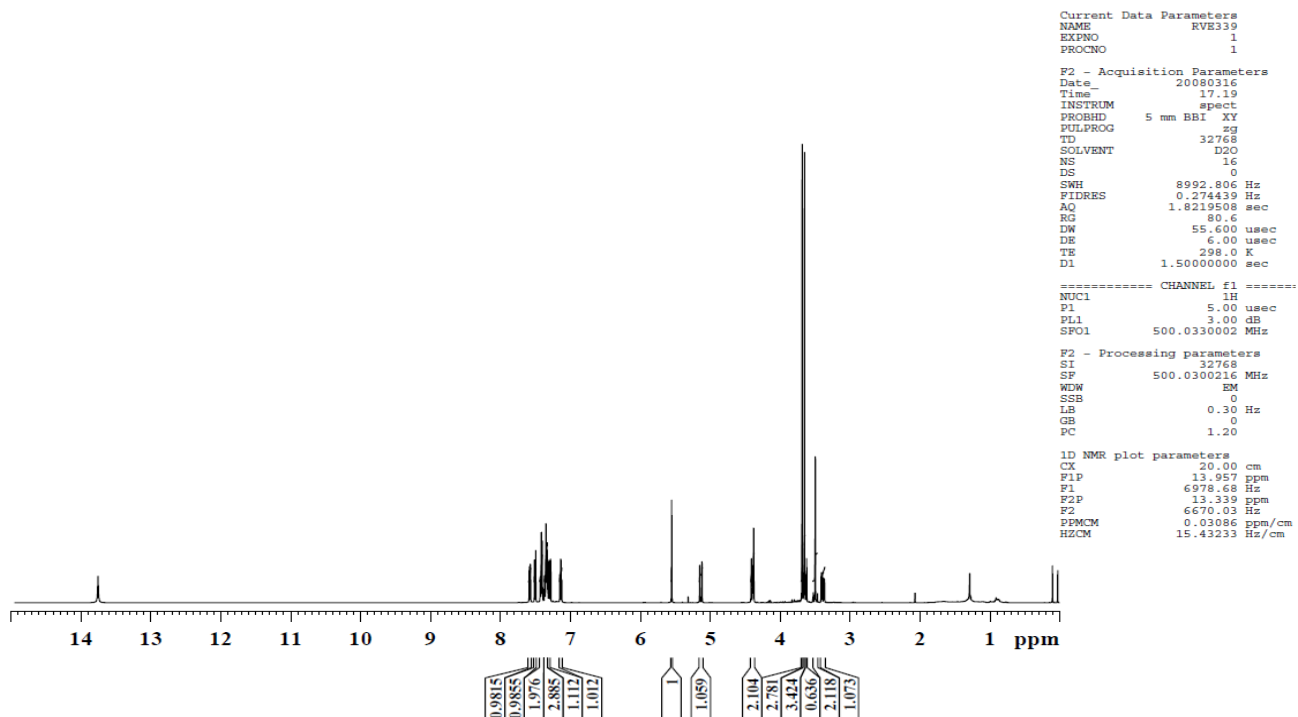
F2 - Processing parameters
SI         32768
SF         75.4677190 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
SI         20.00 cm
CX         110.254 ppm
F1P        8320.62 Hz
F2P        46.895 ppm
F2         3539.04 Hz
PPMCM      3.16797 ppm/cm
HZCM       239.07912 Hz/cm
    
```

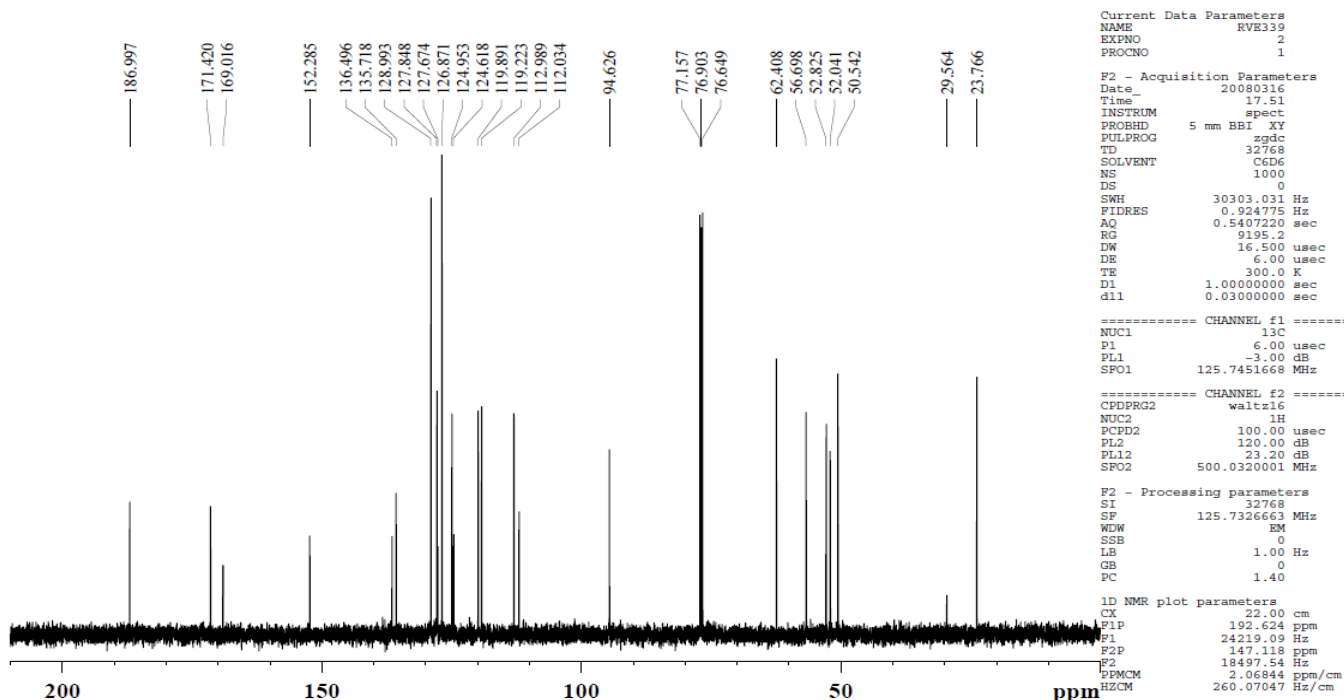


Compound 20 (500 MHz)

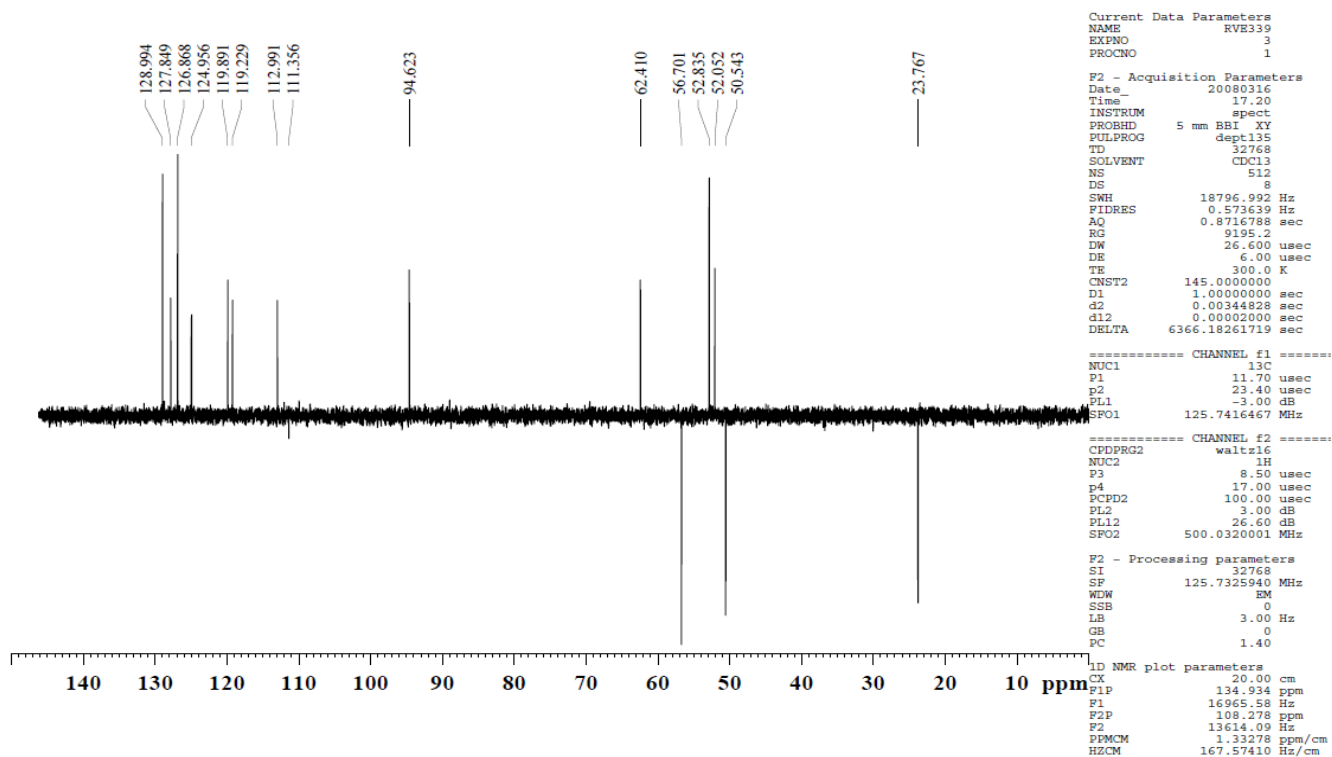
proton spectrum



carbon spectrum



C-13 DEPT-135



X-ray Structural Analysis and Methods for 17 and 18

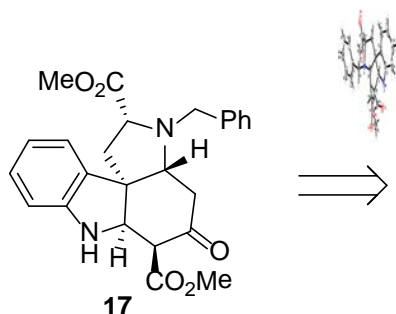


Figure S1: Displacement ellipsoid plot of (2*R*,3*aR*,6*S*,6*aR*,11*bR*)-Dimethyl 3-benzyl-5-oxo-2,3,3*a*,4,5,6,6*a*,7-octahydro-1*H*-pyrrolo[2,3-*d*]carbazole-2,6-dicarboxylate (**17**).

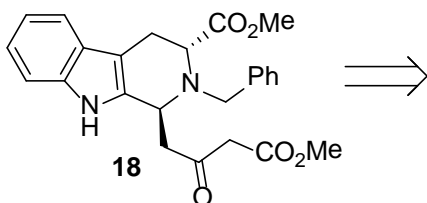


Figure S2: Displacement ellipsoid plot of (1*S*,3*R*)-Methyl-2-benzyl-1-(4-(methoxy-2,4-dioxobutyl)-2,3,4,9-tetrahydro-1*H*-pyrido[3,4-*b*]indole-3-carboxylate (**18**).

X-ray Crystal methods for 17 and 18

Single-crystal X-ray diffraction data on compounds **17** and **18** were collected at room temperature (approximately 23 °C) using MoK α radiation and a Bruker APEX 2 CCD area detector. Crystals were prepared for data collection by coating with high viscosity microscope oil (Paratone-N, Hampton Research). The oil-coated crystal was mounted on a Micro Mesh mount (MiteGen Inc.) and transferred immediately to the diffractometer. Corrections were applied for Lorentz, polarization, and absorption effects. All structures were solved by direct methods and refined by

full-matrix least squares on F^2 values using the programs found in the SHELXTL suite (Bruker, SHELXTL v6.10, 2000, Bruker AXS Inc., Madison, WI). Parameters refined included atomic coordinates and anisotropic thermal parameters for all non-hydrogen atoms. Hydrogen atoms on carbons were included using a riding model [coordinate shifts of C applied to H atoms] with C-H distance set at 0.96 Å.

The crystal of **17**, was monoclinic in space group $P2_1$ with unit cell dimensions $a = 10.9243(19)$ Å, $b = 7.6452(13)$ Å, $c = 13.624(2)$ Å, and $\beta = 92.005(3)^\circ$. Data were 97.9% complete to 25.00° (approximately 0.82 Å) with an average redundancy of 3.62.

The crystal of **18**, was monoclinic in space group $P2_1$ with unit cell dimensions $a = 7.9956(8)$ Å, $b = 12.2939(13)$ Å, $c = 11.3263(11)$ Å, and $\beta = 91.511(2)^\circ$. Data were 100% complete to 28.28° (approximately 0.75 Å) with an average redundancy of 4.07.