SUPPLEMENTARY INFORMATION

Spatiotemporal resolution of the Ntla transcriptome in axial mesoderm development

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SUPPLEMENTARY RESULTS

Supplementary Table 1. cFD vs. *ntla* **cMO + cFD comparison at 9 hpf: 87 microarray hits ranked by fold change.**

Ensembl ID and chromosome locations correspond to zebrafish genome assembly Zv8, gene build 59 (May 2010).

Supplementary Table 2. cFD vs. *ntla* **cMO + cFD comparison at 16 hpf: 12 microarray hits ranked by fold change.**

Ensembl ID and chromosome locations correspond to zebrafish genome assembly Zv8, gene build 59 (May 2010).

Supplementary Table 3. Microarray hits that overlap with previously reported Ntla targets.

*Mesodermal expression domains in gastrula-stage embryos (6-9 hpf) **Mesodermal expression domains in bud-stage embryos (10 hpf)

Supplementary Table 4. cFD vs. *ntla* **cMO + cFD comparison at 9 hpf: 55 microarray hits with known expression patterns, grouped by their expression domain at 10 hpf and ranked by fold change.**

*Expression domains highlighted in bold font were confirmed to be Ntla-dependent by *in situ* hybridization.

Supplementary Table 5. cFD vs. *ntla* **cMO + cFD comparison at 16 hpf: 9 microarray hits with known expression patterns, grouped by their expression domain at 16 hpf and ranked by fold change.**

*Expression domains in highlighted in bold font were confirmed to be Ntla-dependent *in situ* hybridization.

Supplementary Table 6. cFD vs. *ntla* **cMO comparison at 9 hpf: 87 microarray hits grouped by cellular function and ranked by fold change.**

Supplementary Table 7. cFD vs. *ntla* **cMO + cFD comparison at 16 hpf: 12 microarray hits grouped by cellular function and ranked by fold change.**

Supplementary Table 8. cFD vs. *ntla* **cMO + cFD comparison at 9 hpf: 34 microarray hits with reported loss-of-function studies, grouped by their expression domain at 10 hpf and ranked by phenotype and fold change.**

*Expression domains highlight in bold font were confirmed to be Ntla-dependent by *in situ* hybridization.

**Loss-of-function methods highlighted in bold font were performed in this study.

***Phenotypes related to mesodermal patterning are indicated. Those highlighted in bold font were investigated in this study.

Supplementary Table 9. cFD vs. *ntla* **cMO + cFD comparison at 16 hpf: 4 microarray hits with reported loss-of-function studies, grouped by their expression domain at 16 hpf and ranked by fold change.**

*Expression domains highlighted in bold font were confirmed to be Ntla-dependent by *in situ* hybridization. **Phenotypes highlighted in bold font were investigated in this study.

Supplementary Table 10. MO sequences and doses.

*Bases complementary to start codons are underlined, where appropriate. **Intron/exon assignments are based upon the zebrafish genome assembly Zv8, gene build 59 (May 2010).

Supplementary Table 11. Phenotype statistics for morpholino experiments in order of presentation.

Supplementary Table 12. Primer sequences used to verify MO-induced RNA missplicing.

Supplementary Table 13. Primer sequences used to generate *in situ* **probes.**

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Supplementary Figure 1. Ntla protein depletion after *ntla* **cMO photoactivation within the embryonic shield. a**, Zebrafish zygotes were injected with a *ntla* caged morpholino (cMO)/caged fluorescein-conjugated dextran (cFD) mixture and then irradiated within a 100 m-diameter region centered on the shield domain at 6 hours post fertilization (hpf). The irradiated embryos fixed at various developmental time points were assessed by whole-mount immunostaining for Ntla and uncaged fluorescein-conjugated dextran (FD) levels. Non-irradiated embryos were processed and analyzed in an equivalent manner to provide a comparison control, and regions of Ntla protein depletion are indicated by the brackets. **b**, Quantification of Ntla protein levels in 8-hpf embryos after *ntla* cMO photoactivation. Regions demarcated by the dashed white lines were selected for quantification, and average pixel intensities for each position along the horizontal axis (arrow) were determined using ImageJ software. Pixel intensities for Ntla immunostaining in non-irradiated (red line) and locally irradiated (dashed red line) embryos are shown, as well as those for FD immunostaining in locally irradiated embryos (green line). Embryo orientations: dorsal view and anterior up. Scale bars: 200 m.

Supplementary Figure 2. Ntla protein depletion after *ntla* **cMO photoactivation within posterior chordamesoderm. a**, Zebrafish zygotes were injected with a *ntla* cMO/cFD mixture and then were irradiated within a 100 m-diameter region within the posterior chordamesoderm at 12 hpf. The irradiated embryos were fixed at various time points, and Ntla and FD were detected by immunofluorescence. Non-irradiated embryos were processed and analyzed in an equivalent manner to provide a comparison control, and regions of Ntla protein depletion are indicated by the brackets. **b**, Quantification of Ntla protein levels in 16-hpf embryos after *ntla* cMO photoactivation. Regions demarcated by the dashed white lines were selected for quantification, and image data straightened by ImageJ software is shown below each embryo micrograph. Average pixel intensities for each position along the anterior-posterior axis (arrows) were determined using ImageJ software. Pixel intensities for Ntla immunostaining in non-irradiated (red line) and locally irradiated (dashed red line) embryos are shown, as well as those for FD immunostaining in locally irradiated embryos (green line). Embryo orientations: 12 hpf, dorsal view and anterior up; 14 hpf, posterior dorsal view and dorsal up; 16 hpf, lateral view and anterior left. Scale bars: 200 m.

Supplementary Figure 3. Confirmation of selected microarray hits associated with notochord cell fate commitment by *in situ* **hybridization.** 10-hpf wildtype and *ntla* MO-injected embryos stained for candidate Ntla targets expressed during gastrulation are shown, with co-labeling of *pax2a* transcripts in some cases to determine embryo orientation. All 24 genes tested were confirmed to be transcribed in a Ntla-dependent manner. *kirrel3*, *lhx1a*, and *notch1b* transcript levels were specifically reduced in the axial mesoderm of *ntla* morphants, whereas their other expression domains were unaffected. Similarly, expression of *her1*, *rbm38*, *tbx6*, and *wnt8a* in *ntla* morphants was specifically reduced in the tailbud. In addition to these genes, another hit in our microarray data (**Supplementary Table 1**), *wnt3l*, has previously been shown to be expressed in a Ntla-dependent manner (Ref. 4). Embryo orientations: *her1*, *rbm38*, *tbx6*, *wnt8a*, posterior dorsal view and anterior up; all others, dorsal view and anterior up. Scale bar: 100 m.

Supplementary Figure 4. Confirmation of selected microarray hits associated with notochord maturation by *in situ* **hybridization.** 16-hpf *ntla* cMO-injected embryos that were either cultured in the dark or globally UV irradiated at 12 hpf are shown. Of the nine genes tested, all but *guca2b* were confirmed to be transcribed within the axial mesoderm in a Ntladependent manner. *cav3* transcript levels were specifically reduced in the chordamesoderm of irradiated embryos, whereas somitic expression was unaffected. The increase in *ntla* transcript levels upon *ntla* cMO photoactivation is potentially a consequence of *ntla* mRNA stabilization by the uncaged MO. Embryo orientations: *cav3*, dorsal view and anterior left; all others, lateral view and anterior left. Scale bar: 100 m.

Supplementary Figure 5. Knockdown of *mnx1* **expression induces somite morphology defects that differ from Hedgehog pathway loss-of-function phenotypes. a-c**, Somite patterning in wildtype embryos and those previously injected with non-overlapping, splice-blocking *mnx1* MOs. Brightfield and DIC micrographs of 1-dpf embryos are shown. **d**, Confirmation of *mnx1* MO-dependent RNA missplicing by RT-PCR. Mnx1 proteins encoded by the observed transcripts are shown as bar diagrams, with sites corresponding to exon-intron boundaries depicted as blue lines and known protein domains colored green. Exon-intron assignments are based on the Zv8 zebrafish genome assembly Zv8, gene build 59. **e**, Cyclopamine treatment of embryos starting at 5 hpf resulted in ventral body curvature and U-shaped somites that are shortened along the dorsoventral axis, consistent with loss of Hedgehog pathway-dependent muscle fates. **f**, *ntla* morphants also exhibited curved somites, but they were morphologically more similar to *mnx1* morphants then to cyclopamineembryos. Embryo orientations: lateral view and anterior left. Scale bars: whole-embryo brightfield micrograph, 200 μ m; DIC micrograph, 50 m.

Supplementary Figure 6. Representative morphants of Ntla-dependent genes expressed during gastrulation that do not exhibit clear notochord defects. Brightfield micrographs of selected morphants at 1 day post fertilization (dpf) are shown. In cases where a splice-blocking MO was used, RNA missplicing was confirmed by RT-PCR. Proteins encoded by these transcripts are shown as bar diagrams as described in **Supplementary Figure 5**. **a**, Wildtype control. **b**, Embryos injected with two different translation-blocking MOs targeting *hpcal4* at their maximum non-toxic doses exhibited ventral body curvature but wildtype-like notochord cells. **c**, Embryos injected with a splice-blocking MO targeting *inpp5b* had mild notochord necrosis despite co-injection with a *tp53* MO. **d**, Embryos injected with two different translation-blocking MOs targeting *lhx1a* at their maximum non-toxic doses had morphologically normal notochords, although one MO induced ventral curvature and shortening along the anterior-posterior axis. **e-f**, Embryos injected with splice-blocking MOs against *snai2* or *tacc2* had wildtype-like notochords even though the targeted RNA was misspliced as expected. Embryo orientations: lateral view, dorsal up and anterior left. Scale bars: whole-embryo micrograph, 200 m; inset, 50 m.

Supplementary Figure 7. Knockdown of *mnx1* **expression induces muscle differentiation defects that differ from Hedgehog pathway loss-of-function phenotypes.** Slow myosin heavy chain and fast myosin light chain expression in wildtype zebrafish and embryos treated with an *mnx1* MO, a ntla MO, or 50 **µ**M cyclopamine. 1-dpf embryos are shown. Embryo orientations: lateral view and anterior left. Scale bar: 50 m.

Supplementary Figure 8. Morphants of several Ntla-dependent genes expressed during somitogenesis exhibit aberrant notochord cell morphology. Brightfield and DIC (inset) micrographs of selected morphants at 1.5 dpf are shown. In cases where a splice-blocking MO was used, RNA missplicing was confirmed by RT-PCR. Proteins encoded by these transcripts are shown as bar diagrams as described in **Supplementary Figure 5**. **a**, Wildtype control. **b**, Embryos injected with a *cav3* MO had disorganized, incompletely vacuolated notochord cells. **c**, Embryos injected with a *ptrfb* MO exhibited reduced notochord cell-cell contacts and ventral body curvature. **d-e**, Embryos injected with either splice-blocking or translation-blocking *slc38a8* MOs (MO1 and MO2, respectively) had incompletely vacuolated notochord cells. **f**, Co-injection of both *slc38a8* MOs at sub-phenotypic doses synergistically induced similar notochord defects. **g**, Confirmation of *slc38a8* MO-dependent RNA missplicing by RT-PCR. **h-i**, Embryos injected with splice-blocking *znf385b* MOs (MO1 or MO2) exhibited partially vacuolated notochord cells. **j**, Co-injection of both *znf385b* MOs led to a stronger notochord defect. **k**, Confirmation of *znf385b* MO-dependent RNA missplicing by RT-PCR. Embryo orientations: lateral view and anterior left. Scale bars: whole-embryo brightfield micrograph, 200 m; DIC inset, 50 m.

flh expression at 12 hpf

flh expression at 20 hpf

Supplementary Figure 9. Temporal analysis of Ntla-dependent *flh* **transcription. a-b**, Expression of *flh* at 12 hpf and 20 hpf in wildtype zebrafish, as well as in embryos injected with the ntla cMO and irradiated globally at 6 hpf. Embryo orientations: **a**, dorsal posterior view and dorsal up; **b**, lateral view and anterior left. Scale bars: **a**, 100 m; **b**, 200 m.

SUPPLEMENTARY METHODS

Caged fluorescein dextran (cFD) preparation

Aminodextran (3.5-5 mg of 10-kDa polymer; Invitrogen, D1860) dissolved in 500 µL of 0.1 M $Na₂B₄O₇$ buffer (pH 8.5) was added to carboxymethylnitrobenzyl (CMNB)-caged fluorescein N-hydroxysuccinimide ester (1 mg; Invitrogen, C20050) in the manufacturer-supplied tinted tube, and the reaction mixture was vortexed overnight. The resulting cFD was separated from unreacted caged fluorescein using a Zeba Desalt spin column (Thermo Fisher Scientific, 89889) according to the manufacturer's instructions. The yellow-colored eluent was lyophilized to dryness, weighed, dissolved in water to make a 1% (w/v) stock solution, and stored at -20 °C as 2-µL aliquots. Spectroscopic analysis indicated an average loading of 2.5 caged fluoresceins per dextran molecule.

Ntla polyclonal antibody

Full-length *ntla* cDNA flanked by 5' EcoRI and 3' BamHI sites was generated by PCR using cDNA derived from 10-hpf zebrafish embryos as a template and the following primers: 5'- CGAATTCATGTCTGCCTCAAGTCCCGACCA-3' and 5'-CGGATCCTCAGTAGCTCTGAGCCA-CAGGCG-3'. The *ntla* cDNA was then cloned into a pCR-Blunt II-TOPO vector (Invitrogen), digested with EcoRI and BamHI, and ligated into similarly cut pMALc2x vector (NEB) to generate a bacterial expression construct encoding an Ntla-maltose binding protein (MBP) fusion (pMALc2x-Ntla). The pMALc2X-Ntla vector was transformed into BL21 competent cells and a single colony was cultured in 80 mL LB-ampicillin broth overnight. This starter culture was then used to innoculate 1 L of 2X YT broth (16 g tryptone, 10 g yeast extract, 5 g NaCl, 2 g glucose, 100 mg ampicillin, 1 L water) and cultured for 2 hours at 37 °C, followed by 1 hour at 30 °C to achieve an OD_{600} of 0.8-1.0. IPTG (0.5 mM) was added to the culture, which was grown for another 6 hours at 30 °C. Cells were then harvested by centrifugation (4,000 x *g,* 20 minutes, 4 °C), resuspended in column buffer (50 mL; 200 mM NaCl, 1 mM EDTA, 0.07% (v/v) 2-mercaptoethanol, 20 mM Tris-HCl, pH 7.4), chilled to -72 $^{\circ}$ C in a dry ice/ethanol bath,

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thawed in ice water, and sonicated 8 x 15 seconds in ice water. Triton X-100 was added to a final concentration of 1% (v/v), and the lysate was mixed gently at 4 °C for 30 minutes solubilize the Ntla-MBP fusion. The lysate was next centrifuged (12,000 x *g*, 30 minutes, 4 °C) and the supernatant was passed over amylose resin (5 mL; NEB) that was previously packed into a gravity flow column and equilibrated with the column buffer (40 mL). The loaded amylose resin was washed with column buffer (60 mL) and eluted in 1-mL fractions with column buffer containing 20 mM maltose. Fractions containing the 89 kDa Ntla-MBP fusion protein were pooled and concentrated to 1 mg/mL using an Amicon Centricon centrifugal filter device.

The purified Ntla-MBP fusion protein (4 mg) was cleaved with Factor Xa (40 µg) and Ntla was separated from MBP by DEAE-Sepharose ion exchange chromatography using a pMAL Fusion & Purification System kit (NEB) according to the manufacturer's instructions. Rabbit innoculation and serum collection were conducted by Rockland Immunochemicals using their standard procedures. To affinity purify the polyclonal anti-Ntla antibody, an affinity column was prepared by expressing and purifying the Ntla-MBP fusion protein (4 mg) as described above, dialyzing the protein in 1 L 1X PBS for 12 hours, and immobilizing it onto 1 mL of AminoLink resin (Pierce) according to the manufacturer's instructions. The polyclonal anti-Ntla antibody serum (1 mL) was then passed over the gravity-flow affinity column, eluted with 0.1 M glycine-HCl buffer, pH 2.5-3.0. Each 200-µL fraction containing the purified antibody was then neutralized with 10 µL of 1 M Tris-HCl, pH 9.

Imaging of zebrafish embryos

For live imaging of 24- or 36-hpf zebrafish, the embryos were manually dechorinated and immobilized in E3 medium containing 0.5% (w/v) low-melt agarose and 0.05% (w/v) Tricaine mesylate. Brightfield images were acquired using a Leica M205FA fluorescence stereoscope equipped with a Leica DFC500 digital camera. Differential interference contrast (DIC) micrographs were obtained with a Leica DM4500B epifluorescence microscope equipped

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with a 20x/0.5 NA water-immersion objective and a QImaging Retiga-SRV digital camera. Confocal fluorescence and DIC images were obtained with a Zeiss Axio Imager Z1 upright microscope equipped with a 20x/0.5 NA water-immersion objective, fluorescence and transmission photomultiplier tube detectors, and an LSM700 laser-scanning confocal head. Fixed embryos stained by *in situ* hybridization were mounted in 1X PBS containing 2% (w/v) methylcellulose and imaged with the Leica M205FA/DFC500 system using LED illumination arrays and the Leica LAS montage imaging module. Fixed immunostained embryos were mounted in 1X PBS containing 0.5% (w/v) low-melt agarose, and fluorescence images were obtained with a DM4500B/Retiga-SRV system equipped with GFP and TXR filter sets and a 10x/0.30 NA objective.

Cyclopamine treatment of zebrafish embryos

5-hpf embryos in 10 mL of E3 medium (5 mM NaCl, 0.17 mM KCl, 0.33 mM CaCl₂, and 0.33 mM MgSO₄) were treated with an ethanolic solution of cyclopamine (5 μ L of a 100 mM stock; 50 µM final concentration) and cultured at 28.5 °C.

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