SUPPLEMENTARY INFORMATION

Spatiotemporal resolution of the Ntla transcriptome in axial mesoderm development

Ilya A. Shestopalov¹, Cameron L. W. Pitt¹, and James K. Chen¹

¹Department of Chemical and Systems Biology, Stanford University School of Medicine, Stanford, CA 94305, USA

SUPPLEMENTARY RESULTS

Supplementary Table 1	
Supplementary Table 2	7
Supplementary Table 3	
Supplementary Table 4	
Supplementary Table 5	
Supplementary Table 6	
Supplementary Table 7	
Supplementary Table 8	
Supplementary Table 9	
Supplementary Table 10	
Supplementary Table 11	
Supplementary Table 12	
Supplementary Table 13	
Supplementary Figure 1	
Supplementary Figure 2	
Supplementary Figure 3	
Supplementary Figure 4	
Supplementary Figure 5	
Supplementary Figure 6	
Supplementary Figure 7	
Supplementary Figure 8	
Supplementary Figure 9	

Supplementary Table 1. cFD vs. *ntla* cMO + cFD comparison at 9 hpf: 87 microarray hits ranked by fold change.

Ensembl ID and chromosome locations correspond to zebrafish genome assembly Zv8, gene build 59 (May 2010).

Entry	Gene symbol	Gene description	Fold change	Zfin ID	Ensembl gene ID	Nimblegen SEQ_ID	Chr	Start	End
1	si:ch211-215a10.5-001 (hpcal4)	novel protein similar to vertebrate hippocalcin like 4	-6.543	si:ch211- 215a10.5-001	NP_001108159.1	OTTDART00000018441	19	29460115	29462786
2	LOC563543 (inpp5b)	novel protein similar to vertebrate inositol polyphosphate-5-phosphatase	-6.093	none	LOC563543	ZV700S00001662	16	2599817	2639066
3	lhx1a	LIM-homeobox 1a	-6.092	lhx1a	lhx1a	AW077429	15	28135534	28143145
4	snai2	snail homolog 2	-5.700	snai2	snai2	OTTDART00000027124	24	33995692	34001705
5	notch1b	notch homolog 1b	-5.524	notch1b	notch1b	OTTDART00000023258	5	67392870	67496053
6	si:dkey-49o11.4 (tbx3l)	novel protein similar to vertebrate t-box 3	-5.072	si:dkey-49o11.4	LOC568479	OTTDART00000019838	5	24885216	24887665
7	wnt8a	wingless-type MMTV integration site family, member 8a	-4.652	wnt8a	wnt8a	OTTDART00000025914	14	36040558	36047793
8	LOC566628 (pskh2)	novel protein similar to vertebrate protein serine kinase H2	-4.616	none	LOC566628	ENSDART00000037703	12	40283700	40307825
9	Irrc8da	novel protein similar to vertebrate leucine rich repeat containing 8 family, member D alpha	-4.602	Irrc8da	A2BIH6_DANRE	OTTDART00000023505	2	22633903	22636380
10	atp1a1b	ATPase, Na+/K+ transporting, alpha 1b polypeptide	-4.371	atp1a1b	atp1a1b	OTTDART00000026276	9	36416946	36437549
11	myod1	myogenic determination factor 1	-4.290	myod1	myod1	OTTDART00000026919	25	32781654	32783858
12	si:dkey-146l2.1-001 (ankrd15)	novel protein similar to vertebrate ankyrin repeat domain 15	-4.281	si:dkey-146l2.1- 001	A2CEC3_DANRE	TC243640	5	43852500	43944459
13	zgc:91787 (b3gnt7)	novel protein similar to vertebrate beta-3- galactosyltransferase	-4.215	zgc:91787	zgc:91787	OTTDART00000020258	8	22683762	22685318
14	col8a1a	collagen, type VIII, alpha 1	-4.113	col8a1a	col8a1a	AI397462	9	30487274	30499313
15	plp2	proteolipid protein 2	-4.036	plp2	plp2	TC260151	8	50452420	50499168
16	plod1a	lysyl hydroxylase 1	-3.848	plod1a	plod1a	ZV700S00006183	8	49269611	49320332
17	ribc1	RIB43A domain with coiled-coils 1	-3.614	ribc1	zgc:158280	ZV700S00001481	23	28138233	28145545
18	pip5k1c	phosphatidylinositol-4-phosphate 5-kinase, type I, gamma	-3.592	pip5k1c	pip5k1c	ENSDART00000105443	22	17671179	17691603
19	pde4ba	cAMP-specific 3',5'-cyclic phosphodiesterase 4B alpha	-3.547	pde4ba	LOC565706	ZV700S00006639	6	32670067	32747756
20	rbm38	RNA binding motif protein 38	-3.483	rbm38	rbm38	OTTDART00000026624	23	4620570	4666179
21	lamp1	lysosomal-associated membrane protein 1	-3.426	lamp1	none	ZV700S00006436	9	34697159	34701323
22	cpn1	carboxypeptidase N, polypeptide 1	-3.379	cpn1	cpn1	BC066689.1	12	30496591	30523965
23	abcc6b	ATP-binding cassette, sub-family C, member 6b	-3.344	abcc6b	abcc6	ENSDART00000065433	6	11984572	12024328
24	dopey2	dopey family member 2	-3.341	dopey2	dopey2	ENSDART00000078256	9	33581620	33595018

25	kirrel3l	kin of IRRE like 3 like	-3.334	kirrel3l	kirrel3l	ZV700S00005154	9	26637783	26712362
26	aldh1a2	aldehyde dehydrogenase 1 family, member A2	-3.255	aldh1a2	aldh1a2	OTTDART00000025116	7	30625839	30666282
27	mnx1	motor neuron and pancreas homeobox 1	-3.252	mnx1	mnx1	OTTDART00000013585	7	41141692	41144486
28	caskin1	CASK-interacting protein 1	-3.206	none	ENSDARG0000004 6107	ENSDART00000040925	3	32954587	32996156
29	hmbox1b	similar to homeobox containing 1	-3.133	hmbox1b	LOC564816	ENSDART0000008808	Zv8_NA 11417	178371	190934
30	pak1	P21/Cdc42/Rac1-activated kinase 1	-3.114	pak1	pak1	ZV700S00006164	18	2188979	2204559
31	ppargc1al	peroxisome proliferator activated receptor gamma coactivator 1 alpha - like	-3.026	ppargc1al	ppargc1al	ENSDART00000077731	Zv8_NA 6114	4577	28421
32	unc45b	homolog of <i>C. elegans</i> unc-45B	-3.013	unc45b	unc45b	OTTDART00000024993	8	10979015	11004475
33	zgc:66052 (tagln3b)	novel protein similar to vertebrate transgelin 3	-2.939	zgc:66052	zgc:66052	OTTDART00000028979	24	21471592	21485670
34	tbx6	T-box gene 6	-2.919	tbx6	tbx6	OTTDART00000022473	5	41292606	41309337
35	myf5	myogenic factor 5	-2.788	myf5	myf5	OTTDART0000009544	4	21534374	21538246
36	hdr	hematopoietic death receptor	-2.773	hdr	hdr	OTTDART00000019823	5	25528873	25539033
37	rgmb	RGM domain family, member B	-2.721	rgmb	rgmb	OTTDART00000022855	5	48756554	48774922
38	trpv4	transient receptor potential cation channel, subfamily V, member 4	-2.712	trpv4	trpv4	OTTDART00000019252	5	17734760	17772071
39	tacc2	transforming, acidic coiled-coil containing protein 2	-2.693	tacc2	tacc2	ZV700S00006103	13	33046650	33082675
40	adora2ab	adenosine receptor A2a.2	-2.677	adora2ab	NP_001035125.1	NM_001040036	21	10544202	10563719
41	hspa12a	heat shock 70 kDa protein 12A	-2.660	hspa12a	zgc:153612	ZV700S00001957	17	18786426	18805408
42	zgc:153027 (esm1)	novel protein similar to vertebrate endothelial cell- specific molecule 1	-2.637	zgc:153027	zgc:153027	TC263451	21	8194010	8198200
43	kidins220a	kinase D-interacting substance of 220 kDa	-2.603	kidins220a	si:dkey-177f17.1	OTTDART0000007862	17	33031518	33087170
44	si:dkey-195c14.1 (sall3l)	novel protein similar to vertebrate sal-like 3	-2.552	si:dkey-195c14.1	si:dkey-195c14.1	OTTDART00000017492	19	20764504	20773732
45	smyd1b	SET and MYND domain containing 1b	-2.532	smyd1b	smyd1b	NM_001039636	8	1232945	1278872
46	her1	hairy-related 1	-2.529	her1	her1	OTTDART00000023081	5	70458828	70465219
47	p4ha1	procollagen-proline, 2-oxoglutarate 4-dioxygenase (proline 4-hydroxylase), alpha polypeptide I	-2.512	p4ha1	p4ha1	OTTDART00000025381	17	16492035	16511758
48	hfe2	hemochromatosis type 2	-2.485	hfe2	hfe2	NM_213643	16	42382086	42527246
49	zgc:163023 (tspan4)	novel protein similar to vertebrate tetraspanin 4	-2.460	zgc:163023	zgc:163023	 NM_001082807	3	29844005	29853390
50	Irch2	leucine-rich repeats and calponin homology (CH) domain containing 2	-2.431	Irch2	Irch2	OTTDART00000030270	5	34818824	34844320
51	lipg	lipase, endothelial	-2.429	lipg	lipg	OTTDART00000028618	8	33133042	33142110
52	ttna	titin a	-2.415	ttna	ttna	OTTDART00000029928	9	43263480	43328198
53	itgb5	integrin, beta 5	-2.400	itgb5	itgb5	OTTDART00000014003	9	22031250	22101761
54	actn2	actinin, alpha 2	-2.384	actn2	actn2	OTTDART0000029626	17	16515238	16537141
55	zgc:92198 (camk2n2)	novel protein similar to vertebrate Ca2+/calmodulin- dependent protein kinase II inhibitor 2	-2.367	zgc:92198	zgc:92198	NM_001002642	24	25360048	25362300

56	fadd	fas (tnfrsf6)-associated via death domain	-2.341	fadd	fadd	TC242683	7	55882294	55888612
57	wnt3l	wingless-type MMTV integration site family, member 3 like	-2.338	wnt3l	wnt3l	NM_001007185	2	2309950	2311326
58	mesp2	mesoderm posterior 2 homolog	-2.333	mesp2	A3KPB3_DANRE	ENSDART00000102928	25	9383009	9384267
59	twist2	twist 2	-2.333	twist2	twist2	NM_001005956	9	46391351	46391842
60	ttnb	titin b	-2.327	ttnb	ttnb	OTTDART00000026898	9	42989675	43130539
61	st3gal2l	ST3 beta-galactoside alpha-2,3-sialyltransferase 2, like	-2.321	st3gal2l	st3gal2l	TC249907	11	26392769	26414674
62	sort1b	sortilin 1, like	-2.284	sort1b	sort1b	OTTDART00000013761	8	25809473	25842079
63	hmcn1	hemicentin 1	-2.272	hmcn1	hmcn1	OTTDART0000006813	20	34056841	34169854
64	irx3a	iroquois homeobox protein 3a	-2.242	irx3a	irx3a	ZV700S00006366	7	36391615	36394934
65	sall1a	sal-like 1a	-2.206	sall1a	sall1a	ZV700S00001341	7	37784931	37793542
66	tnfaip6	tumor necrosis factor, alpha-induced protein 6	-2.200	tnfaip6	tnfaip6	TC245382	9	22117982	22133555
67	zgc:66433 (KIAA1211)	novel protein similar to vertebrate uncharacterized protein KIAA1211	-2.200	zgc:66433	zgc:66433	OTTDART00000028901	14	22214800	22296654
68	inka1a	induced in neural crest by AP2, 1a	-2.170	inka1a	inka1a	ZV700S0000040	11	35101810	35108146
69	meis4.1a	myeloid ecotropic viral integration site 4.1a	-2.155	meis4.1a	meis4.1a	OTTDART00000023940	1	51288174	51319427
70	dlc	deltaC	-2.146	dlc	dlc	OTTDART00000025961	15	20634176	20639029
71	kitlga	kit ligand a	-2.145	kitlga	kitlga	ENSDART00000104502	25	18638925	18658676
72	prickle2	prickle-like 2	-2.135	prickle2	prickle2	AY278987.1	11	18849431	18895050
73	wwp2	WW domain containing E3 ubiquitin protein ligase 2	-2.121	wwp2	wwp2	TC261671	25	34924572	34978790
74	ENSDARG00000063075 (c20orf117)	novel protein similar to vertebrate uncharacterized protein c20orf117	-2.120	none	ENSDARG0000006 3075	ENSDART00000091852	23	8047246	8130572
75	ube2e3	ubiquitin-conjugating enzyme E2E 3	-2.120	ube2e3	ube2e3	OTTDART00000026433	9	44146061	44228245
76	kif1b	kinesin family member 1B	-2.085	kif1b	kif1b	ZV700S00001465	23	28894800	28957280
77	notch2	notch homolog 2	-2.072	notch2	notch2	ZV700S00005786	8	19943482	19983080
78	si:ch211-285f17.1 (KIAA1217)	novel protein similar to vertebrate sickle tail protein homolog	-2.030	si:ch211-285f17.1	ENSDARG0000003 1658	TC253143	24	9246424	9333914
79	id:ibd2573 (creb3l1)	novel protein similar to vertebrate cAMP responsive element binding protein 3-like 1	-2.021	id:ibd2573	LOC566720	ENSDART00000074532	7	39330933	39358257
80	fgf24	fibroblast growth factor 24	-2.020	fgf24	fgf24	OTTDART0000002076	14	6156422	6194919
81	tnfrsfa	tumor necrosis factor receptor superfamily, member a	-2.013	tnfrsfa	tnfrsfa	ZV700S0000235	21	16565637	16612107
82	zgc:165655	novel pleckstrin domain protein	-2.009	zgc:165655	zgc:165655	NM_001098779	16	34734527	34792568
	82/87 (94%) downregulate	ed							
83	туот3	myomesin family, member 3	+2.002	туот3	туот3	ENSDART00000101501	Zv8_NA 513	18129	78285
84	nmnat2	nicotinamide nucleotide adenylyltransferase 2	+2.027	nmnat2	nmnat2	OTTDART00000020501	2	34068428	34082906
85	hbp1	HMG box-containing protein 1	+2.037	hbp1	zgc:112297	AW282104	25	2564423	2584269

86	rbms3	Single-stranded RNA interacting protein	+2.095	rbms3	rbms3	TC267504	16	38456043	38671893
87	pdgfrb2	platelet-derived growth factor receptor beta 1.2 fragment	+2.442	pdgfrb2	pdgfrb2	ENSDART00000028255	14	4208337	4211507
	5/87 (6%) upregulated								

Supplementary Table 2. cFD vs. *ntla* cMO + cFD comparison at 16 hpf: 12 microarray hits ranked by fold change.

Ensembl ID and chromosome locations correspond to zebrafish genome assembly Zv8, gene build 59 (May 2010).

Entry	Gene symbol	Gene description	Fold change	Zfin ID	Ensembl gene ID	Nimblegen SEQ_ID	Chr	Start	End
1	LOC795255 (slc38a8)	novel protein similar to vertebrate soluble carrier family 38 protein	-4.917	none	LOC795255	BQ132767	7	29413983	29422236
2	cav3	caveolin 3	-2.717	cav3	cav3	AY124574.1	6	45637427	45645163
3	znf385b	zinc finger protein 385b	-2.357	znf385b	znf385b	OTTDART0000001965	9	43499966	43886239
4	tspan7b	tetraspanin 7b	-2.318	tspan7b	tspan7b	ZV700S00005092	22	13702361	13754893
5	si:dkey-205o12.2 (prk1)	novel protein similar to vertebrate pim oncogene related kinase 1	-2.073	si:dkey-205o12.2	si:dkey-205o12.2	ENSDART00000100940	8	13504646	13505939
6	si:dkeyp-122e7.1 (znf804a)	novel protein similar to vertebrate zinc finger 804a	-2.051	si:dkeyp-122e7.1	ENSDARG00000027 079	ZV700S0000778	9	11484108	11611524
7	ptrfb	polymerase I and transcript release factor b	-2.046	ptrfb	ptrfb	ZV700S00003617	24	7593206	7623842
8	zgc:174919	novel protein containing zinc finger domain	-2.039	zgc:174919	zgc:174919	ENSDART00000106259	22	3102571	3109743
9	zgc:110418 (kcnk6)	novel protein similar to vertebrate potassium inwardly-rectifying channel, subfamily K, member 6	-2.030	zgc:110418	zgc:110418	TC241224	18	45049012	45058866
	9/12 (75%) downregulate	d							
10	wu:fd47f06 (guca2b)	novel protein similar to vertebrate guanylate cyclase activator 2b	+2.095	wu:fd47f06	none	ENSDART00000028699	11	27970374	27971060
11	ntla	no tail-a	+2.225	ntla	ntla	ZV700S00001498	19	20033640	20038792
12	frzb	frizzled-related protein	+3.320	frzb	frzb	ZV700S00006289	9	47742998	47749719
	3/12 (25%) upregulated								

Supplementary Table 3. Microarray hits that overlap with previously reported Ntla targets.

*Mesodermal expression domains in gastrula-stage embryos (6-9 hpf) **Mesodermal expression domains in bud-stage embryos (10 hpf)

Entry	Gene symbol	Gene description	Expression domain in zebrafish	Previously reported direct target? ¹	Previously reported transcriptional target?
1	dic	deltaC	Ventrolateral margin* Paraxial**	6 probes	No
2	wnt8a	wingless-type MMTV integration site family, member 8a	Ventrolateral margin* Tailbud**	5 probes, T-box clusters	Yes ²
3	aldh1a2	aldehyde dehydrogenase 1 family, member A2	Ventrolateral margin* Paraxial**	4 probes, T-box clusters	1.5-fold decrease in expression upon loss of Ntla ³
4	her1	hairy-related 1	Ventrolateral margin* Tailbud, Paraxial**	4 probes	1.5-fold decrease in expression upon loss of Ntla ³
5	kirrel3l	kin of IRRE like 3 like	Axial, Margin* Axial**	3 probes, T-box clusters	No
6	rbm38	RNA binding motif protein 38	Tailbud, Paraxial**	3 probes	2.1-fold decrease in expression with loss of Ntla ³
7	irx3a	iroquois homeobox protein 3a	Axial* Axial**	3 probes	No
8	tbx6	T-box gene 6	Ventrolateral margin* Tailbud, Paraxial**	2 probes, T-box clusters	1.7-fold decrease in expression with loss of Ntla ³
9	fgf24	fibroblast growth factor 24	Margin* Tailbud, Paraxial**	2 probes, T-box clusters	No
10	ube2e3	ubiquitin-conjugating enzyme E2E 3	Not reported	2 probes	No
11	myod1	myogenic determination factor 1	Paraxial margin* Paraxial**	1 probe	No
12	tnfaip6	tumor necrosis factor, alpha-induced protein 6	Not reported	1 probe, T-box clusters	No
13	zgc:66052 (tagln3b)	novel protein similar to vertebrate transgelin 3	Axial, Other* Axial, Other**	T-box clusters	5.0-fold decrease in expression with loss of Ntla ³
14	zgc:163023 (tspan4)	novel protein similar to vertebrate tetraspanin 4	Axial, Margin*	No	6.9-fold decrease in expression with loss of Ntla ³
15	hspa12a	heat shock 70 kDa protein 12A	Not reported	No	4.4-fold decrease in expression with loss of Ntla ³

16	tacc2	transforming, acidic coiled-coil containing protein 2	Axial, Margin* Axial**	No	2.7-fold decrease in expression with loss of Ntla ³
17	plp2	proteolipid protein 2	Paraxial**	No	2.3-fold decrease in expression with loss of Ntla ³

Supplementary Table 4. cFD vs. *ntla* cMO + cFD comparison at 9 hpf: 55 microarray hits with known expression patterns, grouped by their expression domain at 10 hpf and ranked by fold change.

*Expression domains highlighted in bold font were confirmed to be Ntla-dependent by *in situ* hybridization.

Entry	Gene symbol	Gene description	Expression domain at 10 hpf*	Fold change
1	si:ch211-215a10.5- 001 (hpcal4)	novel protein similar to vertebrate hippocalcin like 4	Axial	-6.543
2	LOC563543 (inpp5b)	novel protein similar to vertebrate inositol polyphosphate-5-phosphatase	Axial	-6.093
3	snai2	snail homolog 2	Axial	-5.700
4	LOC566628 (pskh2)	novel protein similar to vertebrate protein serine kinase H2	Axial	-4.616
5	lrrc8da	novel protein similar to vertebrate leucine rich repeat containing 8 family, member D alpha	Axial	-4.602
6	zgc:91787 (b3gnt7)	novel protein similar to vertebrate beta-3-galactosyltransferase	Axial	-4.215
7	col8a1a	collagen, type VIII, alpha 1	Axial	-4.113
8	plod1a	lysyl hydroxylase 1	Axial	-3.848
9	mnx1	motor neuron and pancreas homeobox 1	Axial	-3.252
10	pak1	P21/Cdc42/Rac1-activated kinase 1	Axial	-3.114
11	hdr	hematopoietic death receptor	Axial	-2.773
12	rgmb	RGM domain family, member B	Axial	-2.721
13	trpv4	transient receptor potential cation channel, subfamily V, member 4	Axial	-2.712
14	tacc2	transforming, acidic coiled-coil containing protein 2	Axial	-2.693
15	p4ha1	procollagen-proline, 2-oxoglutarate 4-dioxygenase (proline 4-hydroxylase), alpha polypeptide I	Axial	-2.512
16	hfe2	hemochromatosis type 2	Axial	-2.485
17	Irch2	leucine-rich repeats and calponin homology (CH) domain containing 2	Axial	-2.431
18	lipg	lipase, endothelial	Axial	-2.429
19	zgc:92198 (camk2n2)	novel protein similar to vertebrate Ca2+/calmodulin-dependent protein kinase II inhibitor 2	Axial	-2.367
20	fadd	fas (tnfrsf6)-associated via death domain	Axial	-2.341
21	twist2	twist 2	Axial	-2.333
22	hmcn1	hemicentin 1	Axial	-2.272
23	irx3a	iroquois homeobox protein 3a	Axial	-2.242
24	sall1a	sal-like 1a	Axial	-2.206
25	inka1a	induced in neural crest by AP2, 1a	Axial	-2.17
26	prickle2	prickle-like 2	Axial	-2.135
27	wwp2	WW domain containing E3 ubiquitin protein ligase 2	Axial	-2.121

28	id:ibd2573 (creb3l1)	novel protein similar to vertebrate cAMP responsive element binding protein 3-like 1	Axial	-2.021
29	tnfrsfa	tumor necrosis factor receptor superfamily, member a	Axial	-2.013
30	lhx1a	LIM-homeobox 1a	Axial, Other	-6.092
31	notch1b	notch homolog 1b	Axial, Other	-5.524
32	kirrel3l	kin of IRRE like 3 like	Axial, Other	-3.334
33	zgc:66052 (tagln3b)	novel protein similar to vertebrate transgelin 3	Axial, Other	-2.939
34	zgc:66433 (KIAA1211)	novel protein similar to vertebrate uncharacterized protein KIAA1211	Axial, Other	-2.200
	34/55 (62%) with axia	I expression		
35	myod1	myogenic determination factor 1	Paraxial	-4.290
36	plp2	proteolipid protein 2	Paraxial	-4.036
37	cpn1	carboxypeptidase N, polypeptide 1	Paraxial	-3.379
38	aldh1a2	aldehyde dehydrogenase 1 family, member A2	Paraxial	-3.255
39	ppargc1al	peroxisome proliferator activated receptor gamma coactivator 1 alpha - like	Paraxial	-3.026
40	unc45b	homolog of <i>C. elegans</i> unc-45B	Paraxial	-3.013
41	myf5	myogenic factor 5	Paraxial	-2.788
42	smyd1b	SET and MYND domain containing 1b	Paraxial	-2.532
43	ttna	titin a	Paraxial	-2.415
44	ttnb	titin b	Paraxial	-2.327
45	meis4.1a	myeloid ecotropic viral integration site 4.1a	Paraxial	-2.155
46	dlc	deltaC	Paraxial	-2.146
47	notch2	notch homolog 2	Paraxial	-2.072
	13/55 (24%) with para	axial expression		
48	wnt8a	wingless-type MMTV integration site family, member 8a	Tailbud, Other	-4.652
49	rbm38	RNA binding motif protein 38	Tailbud, Paraxial	-3.483
50	adora2ab	adenosine receptor A2a.2	Tailbud, Other	-2.677
51	her1	hairy-related 1	Tailbud, Paraxial	-2.529
52	tbx6	T-box gene 6	Tailbud, Paraxial	-2.919
53	wnt3l	wingless-type MMTV integration site family, member 3 like	Tailbud ⁴ , Paraxial	-2.338
54	st3gal2l	ST3 beta-galactoside alpha-2,3-sialyltransferase 2, like	Tailbud, Paraxial	-2.321
55	fgf24	fibroblast growth factor 24	Tailbud, Paraxial	-2.020
	8/55 (14%) with tailbu			

Supplementary Table 5. cFD vs. *ntla* cMO + cFD comparison at 16 hpf: 9 microarray hits with known expression patterns, grouped by their expression domain at 16 hpf and ranked by fold change.

*Expression domains in highlighted in bold font were confirmed to be Ntla-dependent *in situ* hybridization.

Entry	Gene symbol	Gene description	Expression domain at 16 hpf*	Fold change
1	LOC795255 (slc38a8)	novel protein similar to vertebrate soluble carrier family 38 protein	Axial	-4.917
2	znf385b	zinc finger protein 385b	Axial	-2.357
3	tspan7b	tetraspanin 7b	Axial	-2.318
4	ptrfb	polymerase I and transcript release factor b	Axial	-2.046
5	zgc:110418 (kcnk6)	novel protein similar to vertebrate potassium inwardly-rectifying channel, subfamily K, member 6	Axial	-2.030
6	frzb	frizzled-related protein	Axial	+3.320
7	ntla	no tail-a	Axial	+2.225
8	cav3	caveolin 3	Axial, Paraxial	-2.717
9	wu:fd47f06 (guca2b)	novel protein similar to vertebrate guanylate cyclase activator 2b	Ectoderm	+2.095
	8/9 (89 %) with axial exp	ression		·

Supplementary Table 6. cFD vs. *ntla* cMO comparison at 9 hpf: 87 microarray hits grouped by cellular function and ranked by fold change.

Entry	Gene symbol	Gene description	Cellular function	Fold change
1	hdr	hematopoietic death receptor	Apoptosis	-2.773
2	fadd	fas (tnfrsf6)-associated via death domain	Apoptosis	-2.341
3	tnfaip6	tumor necrosis factor, alpha-induced protein 6	Apoptosis	-2.200
4	tnfrsfa	tumor necrosis factor receptor superfamily, member a	Apoptosis	-2.013
	4/87 (5%) apoptosis			
5	Irrc8da	novel protein similar to vertebrate leucine rich repeat containing 8 family, member D alpha	Cell adhesion/migration	-4.602
6	zgc:91787 (b3gnt7)	novel protein similar to vertebrate beta-3-galactosyltransferase	Cell adhesion/migration	-4.215
7	kirrel3l	kin of IRRE like 3 like	Cell adhesion/migration	-3.334
8	pak1	P21/Cdc42/Rac1-activated kinase 1	Cell adhesion/migration	-3.114
9	zgc:66052 (tagln3b)	novel protein similar to vertebrate transgelin 3	Cell adhesion/migration	-2.939
10	zgc:163023 (tspan4)	novel protein similar to vertebrate tetraspanin 4	Cell adhesion/migration	-2.460
11	itgb5	integrin, beta 5	Cell adhesion/migration	-2.400
12	st3gal2l	ST3 beta-galactoside alpha-2,3-sialyltransferase 2, like	Cell adhesion/migration	-2.321
13	hmcn1	hemicentin 1	Cell adhesion/migration	-2.272
14	inka1a	induced in neural crest by AP2, 1a	Cell adhesion/migration	-2.170
	10/87 (12%) cell adhesio	n/migration		
15	si:ch211-215a10.5-001 (hpcal4)	novel protein similar to vertebrate hippocalcin like 4	Cell signaling	-6.543
16	LOC563543 (inpp5b)	novel protein similar to vertebrate inositol polyphosphate-5-phosphatase	Cell signaling	-6.093
17	notch1b	notch homolog 1b	Cell signaling	-5.524
18	wnt8a	wingless-type MMTV integration site family, member 8a	Cell signaling	-4.652
19	LOC566628 (pskh2)	novel protein similar to vertebrate protein serine kinase H2	Cell signaling	-4.616
20	pip5k1c	phosphatidylinositol-4-phosphate 5-kinase, type I, gamma	Cell signaling	-3.592
21	pde4ba	cAMP-specific 3',5'-cyclic phosphodiesterase 4B alpha	Cell signaling	-3.547
22	aldh1a2	aldehyde dehydrogenase 1 family, member A2	Cell signaling	-3.255
23	rgmb	RGM domain family, member B	Cell signaling	-2.721
24	trpv4	transient receptor potential cation channel, subfamily V, member 4	Cell signaling	-2.712
25	adora2ab	adenosine receptor A2a.2	Cell signaling	-2.677
26	zgc:153027 (esm1)	novel protein similar to vertebrate endothelial cell-specific molecule 1	Cell signaling	-2.637
27	kidins220a	kinase D-interacting substance of 220 kDa	Cell signaling	-2.603
28	zgc:92198 (camk2n2)	novel protein similar to vertebrate Ca2+/calmodulin-dependent protein kinase II inhibitor 2	Cell signaling	-2.367
29	wnt3l	wingless-type MMTV integration site family, member 3 like	Cell signaling	-2.338
30	dlc	deltaC	Cell signaling	-2.146

31	kitlga	kit ligand a	Cell signaling	-2.145
32	kif1b	kinesin family member 1B	Cell signaling	-2.085
33	notch2	notch homolog 2	Cell signaling	-2.072
34	fgf24	fibroblast growth factor 24	Cell signaling	-2.020
35	zgc:165655	novel pleckstrin domain protein	Cell signaling	-2.009
36	pdgfrb2	platelet-derived growth factor receptor beta 1.2 fragment	Cell signaling	+2.442
	22/87 (25%) cell signaling	g		
37	plp2	proteolipid protein 2	Endocytosis	-4.036
38	sort1b	sortilin 1, like	Endocytosis	-2.284
	2/87 (2%) endocytosis			
39	col8a1a	collagen, type VIII, alpha 1	Extracellular matrix	-4.113
40	plod1a	lysyl hydroxylase 1	Extracellular matrix	-3.848
41	cpn1	carboxypeptidase N, polypeptide 1	Extracellular matrix	-3.379
42	p4ha1	procollagen-proline, 2-oxoglutarate 4-dioxygenase (proline 4-hydroxylase), alpha polypeptide I	Extracellular matrix	-2.512
	4/87 (5%) extracellular m	atrix		
43	lhx1a	LIM-homeobox 1a	Gene expression	-6.092
44	snai2	snail homolog 2	Gene expression	-5.700
45	si:dkey-49o11.4 (tbx3l)	novel protein similar to vertebrate t-box 3	Gene expression	-5.072
46	myod1	myogenic determination factor 1	Gene expression	-4.290
47	rbm38	RNA binding motif protein 38	Gene expression	-3.483
48	mnx1	motor neuron and pancreas homeobox 1	Gene expression	-3.252
49	hmbox1b	similar to homeobox containing 1	Gene expression	-3.133
50	tbx6	T-box gene 6	Gene expression	-2.919
51	myf5	myogenic factor 5	Gene expression	-2.788
52	si:dkey-195c14.1 (sall3l)	novel protein similar to vertebrate sal-like 3	Gene expression	-2.552
53	smyd1b	SET and MYND domain containing 1b	Gene expression	-2.532
54	her1	hairy-related 1	Gene expression	-2.529
55	mesp2	mesoderm posterior 2 homolog	Gene expression	-2.333
56	twist2	twist 2	Gene expression	-2.333
57	irx3a	iroquois homeobox protein 3a	Gene expression	-2.242
58	sall1a	sal-like 1a	Gene expression	-2.206
59	meis4.1a	myeloid ecotropic viral integration site 4.1a	Gene expression	-2.155
60	prickle2	prickle-like 2	Gene expression	-2.135
61	si:ch211-285f17.1 (KIAA1217)	novel protein similar to vertebrate sickle tail protein homolog	Gene expression	-2.0300
62	id:ibd2573 (creb3l1)	novel protein similar to vertebrate cAMP responsive element binding protein 3- like 1	Gene expression	-2.021

63	hbp1	HMG box-containing protein 1	Gene expression	+2.037
64	rbms3	RNA binding motif, single stranded interacting protein	Gene expression	+2.095
	22/87 (25%) gene expres	sion		
65	hfe2	hemochromatosis type 2	Other (homeostasis)	-2.485
66	lipg	lipase, endothelial	Other (homeostasis)	-2.429
67	nmnat2	nicotinamide nucleotide adenylyltransferase 2	Other (homeostasis)	+2.027
68	atp1a1b	ATPase, Na+/K+ transporting, alpha 1b polypeptide	Other (membrane transporter)	-4.371
69	abcc6b	ATP-binding cassette, sub-family C, member 6b	Other (membrane transporter)	-3.344
70	lamp1	lysosomal-associated membrane protein 1	Other (protein homeostasis)	-3.426
71	hspa12a	heat shock 70 kDa protein 12A	Other (protein homeostasis)	-2.660
72	wwp2	WW domain containing E3 ubiquitin protein ligase 2	Other (protein homeostasis)	-2.121
73	ube2e3	ubiquitin-conjugating enzyme E2E 3	Other (protein homeostasis)	-2.120
	9/87 (10%) other			
74	ppargc1al	peroxisome proliferator activated receptor gamma coactivator 1 alpha - like	Sarcomere assembly	-3.026
75	unc45b	homolog of <i>C. elegans</i> unc-45B	Sarcomere assembly	-3.013
76	ttna	titin a	Sarcomere assembly	-2.415
77	actn2	actinin, alpha 2	Sarcomere assembly	-2.384
78	ttnb	titin b	Sarcomere assembly	-2.327
79	myom3	myomesin family, member 3	Sarcomere assembly	+2.002
	6/87 (7%) sarcomere ass	embly		
80	si:dkey-146l2.1-001 (ankrd15)	novel protein similar to vertebrate ankyrin repeat domain 15	Unknown	-4.281
81	ribc1	RIB43A domain with coiled-coils 1	Unknown	-3.614
82	dopey2	dopey family member 2	Unknown	-3.341
83	caskin1	CASK-interacting protein 1	Unknown	-3.206
84	tacc2	transforming, acidic coiled-coil containing protein 2	Unknown	-2.693
85	lrch2	leucine-rich repeats and calponin homology (CH) domain containing 2	Unknown	-2.431
86	zgc:66433 (KIAA1211)	novel protein similar to vertebrate uncharacterized protein KIAA1211	Unknown	-2.200
87	ENSDARG0000006307 5 (c20orf117)	novel protein similar to vertebrate uncharacterized protein c20orf117	Unknown	-2.1200
	8/87 (9%) unknown			1

Supplementary Table 7. cFD vs. *ntla* cMO + cFD comparison at 16 hpf: 12 microarray hits grouped by cellular function and ranked by fold change.

Entry	Gene symbol	Gene description	Cellular function	Fold change
1	tspan7b	tetraspanin 7b	Cell adhesion/migration	-2.318
	1/12 (8%) cell adhesion/r	nigration		
2	si:dkey-205o12.2 (prk1)	novel protein similar to vertebrate pim oncogene related kinase 1	Cell signaling	-2.073
3	frzb	frizzled-related protein	Cell signaling	+3.320
4	wu:fd47f06 (guca2b)	novel protein similar to vertebrate guanylate cyclase activator 2b	Cell signaling	+2.095
	3/12 (25%) cell signaling			
5	LOC795255 (slc38a8)	novel protein similar to vertebrate soluble carrier family 38 protein	Endocytosis	-4.917
6	cav3	caveolin 3	Endocytosis	-2.717
7	ptrfb	polymerase I and transcript release factor b	Endocytosis	-2.046
	3/12 (25%) endocytosis			
8	znf385b	zinc finger protein 385b	Gene expression	-2.357
9	si:dkeyp-122e7.1 (znf804a)	novel protein similar to vertebrate zinc finger 804a	Gene expression	-2.051
10	zgc:174919	novel protein containing zinc finger domain	Gene expression	-2.039
11	ntla	no tail-a	Gene expression	+2.225
	4/12 (33 %) gene express	sion		
12	zgc:110418 (kcnk6)	novel protein similar to vertebrate potassium inwardly-rectifying channel, subfamily K, member 6	Membrane transporter	-2.030
	1/12 (8%) other			

Supplementary Table 8. cFD vs. *ntla* cMO + cFD comparison at 9 hpf: 34 microarray hits with reported loss-of-function studies, grouped by their expression domain at 10 hpf and ranked by phenotype and fold change.

*Expression domains highlight in bold font were confirmed to be Ntla-dependent by *in situ* hybridization.

**Loss-of-function methods highlighted in bold font were performed in this study.

***Phenotypes related to mesodermal patterning are indicated. Those highlighted in bold font were investigated in this study.

Entry	Gene symbol	Gene description	Expression domain at 10 hpf*	Fold Change	Loss-of-function method**	Mesodermal defects***
1	pak1	P21/Cdc42/Rac1-activated kinase 1	Axial	-3.114	Morpholino	Gastrulation ⁵
2	irx3a	iroquois homeobox protein 3a	Axial	-2.242	Morpholino	Gastrulation ⁶
3	prickle2	prickle-like 2	Axial	-2.135	Morpholino	Gastrulation ⁷
4	col8a1a	collagen, type VIII, alpha 1	Axial	-4.113	Mutant, Morpholino	Notochord ⁸
5	mnx1	motor neuron and pancreas homeobox 1	Axial	-3.252	Morpholino	Notochord, Muscle
6	si:ch211- 215a10.5-001 (hpcal4)	novel protein similar to vertebrate hippocalcin like 4	Axial	-6.543	Morpholino	None
7	LOC563543 (inpp5b)	novel protein similar to vertebrate inositol polyphosphate-5-phosphatase	Axial	-6.093	Morpholino	None
8	snai2	snail homolog 2	Axial	-5.700	Morpholino	None
9	hdr	hematopoietic death receptor	Axial	-2.773	Morpholino	None ⁹
10	trpv4	transient receptor potential cation channel, subfamily V, member 4	Axial	-2.712	Mutant, Morpholino	None ¹⁰
11	tacc2	transforming, acidic coiled-coil containing protein 2	Axial	-2.693	Morpholino	None
12	fadd	fas (tnfrsf6)-associated via death domain	Axial	-2.341	Morpholino	None ⁹
13	hmcn1	hemicentin 1	Axial	-2.272	Mutant	None ^{11,12}
14	sall1a	sal-like 1a	Axial	-2.206	Morpholino	None ¹³
15	inka1a	induced in neural crest by AP2, 1a	Axial	-2.170	Mutant, Morpholino	None 14
16	tnfrsfa	tumor necrosis factor receptor superfamily, member a	Axial	-2.013	Morpholino	None ⁹
17	kirrel3l	kin of IRRE like 3 like	Axial, Other	-3.334	Morpholino	Muscle 15
18	lhx1a	LIM-homeobox 1a	Axial, Other	-6.092	Morpholino	None
19	notch1b	notch homolog 1b	Axial, Other	-5.524	Morpholino	None 16-18
	2/19 (10%) with axi	al expression had notochord defects				
20	myod1	myogenic determination factor 1	Paraxial	-4.290	Mutant, Morpholino	Muscle ¹⁹
21	unc45b	homolog of <i>C. elegans</i> unc-45B	Paraxial	-3.013	Mutant, Morpholino	Muscle 20

22	myf5	myogenic factor 5	Paraxial	-2.788	Mutant, Morpholino	Muscle ²¹
23	smyd1b	SET and MYND domain containing 1b	Paraxial	-2.532	Morpholino	Muscle ²²
24	ttna	titin a	Paraxial	-2.415	Mutant, Morpholino	Muscle ²³
25	ttnb	titin b	Paraxial	-2.327	Morpholino	Muscle ²³
26	dlc	deltaC	Paraxial	-2.146	Mutant, Morpholino	Muscle ²⁴
27	aldh1a2	aldehyde dehydrogenase 1 family, member A2	Paraxial	-3.255	Mutant, Morpholino	No ^{25,26}
28	meis4.1a	myeloid ecotropic viral integration site 4.1a	Paraxial	-2.155	Morpholino	None ²⁷
29	notch2	notch homolog 2	Paraxial	-2.072	Morpholino	None 16
	7/10 (70%) with pa	raxial expression had muscle defects				
30	wnt8a	wingless-type MMTV integration site family, member 8a	Tailbud, Other	-4.652	Mutant, Morpholino	Notochord, Mesoderm ²⁸⁻³⁰
31	wnt3l	wingless-type MMTV integration site family, member 3 like	Tailbud, Paraxial	-2.338	Morpholino	Mesoderm 29,30
32	fgf24	fibroblast growth factor 24	Tailbud, Paraxial	-2.020	Mutant, Morpholino	Mesoderm ³¹
33	ppargc1al	peroxisome proliferator activated receptor gamma coactivator 1 alpha - like	Paraxial	-3.026	Morpholino	Muscle ³²
34	her1	hairy-related 1	Tailbud, Paraxial	-2.529	Mutant, Morpholino	Muscle ³³
	3/5 (60%) with tailbud expression had mesoderm defects					

Supplementary Table 9. cFD vs. *ntla* cMO + cFD comparison at 16 hpf: 4 microarray hits with reported loss-of-function studies, grouped by their expression domain at 16 hpf and ranked by fold change.

*Expression domains highlighted in bold font were confirmed to be Ntla-dependent by *in situ* hybridization. **Phenotypes highlighted in bold font were investigated in this study.

Entry	Gene symbol	Gene description	Expression domain at 16 hpf*	Fold Change	Loss-of-function method	Notochord defects**
1	LOC795255 (slc38a8)	novel protein similar to vertebrate soluble carrier family 38 protein	Chordamesoderm	-4.917	Morpholino	Yes
2	znf385b	zinc finger protein 385b	Chordamesoderm	-2.357	Morpholino	Yes
3	ptrfb	polymerase I and transcript release factor b	Chordamesoderm	-2.046	Morpholino	Yes ³⁴
4	cav3	caveolin 3	Chordamesoderm, Paraxial	-2.717	Morpholino	Yes ³⁵

Supplementary Table 10. MO sequences and doses.

*Bases complementary to start codons are underlined, where appropriate. **Intron/exon assignments are based upon the zebrafish genome assembly Zv8, gene build 59 (May 2010).

МО	Sequence*	Targeted region**	Dose (per embryo)	Functional validation
cav3 MO	CGTTAGTGTTGTACTGGTCCGC <u>CAT</u>	translational start site	3 ng + 4.5 ng <i>tp53</i> MO	previously reported morphant ³⁵
col8a1a MO	CCGTAGGAGAAGATAATCTCAAGGA	translational start site	6 ng	mutant phenocopy ⁸
<i>lhx1a</i> MO1	CCCGCACAGTGGAC <u>CAT</u> CGTCTTTG	translational start site	18 ng	
lhx1a MO2	GAC <u>CAT</u> CGTCTTTGGATGTGCTCCC	translational start site	8 ng	
LOC563543 (inpp5b)	CTGGAAACTACCGGAGGAAAACACA	splice – i9e10	3 ng + 4.5 ng <i>tp53</i> MO	missplicing – Supp. Fig. 6C
LOC795255 (slc38a8) MO1	TGGCATAGCCTCTAACACACCTGGT	splice - e3i3	6 ng	missplicing – Supp. Fig. 8G
LOC795255 (slc38a8) MO2	AGTTCCTC <u>CAT</u> CCTCAGCGCCTCAA	translational start site	1.5 ng + 2.2 ng <i>tp53</i> MO	phenocopy of MO1
mnx1 MO1	GCCTGGCCTGCATGGAGAATTTTAA	splice - i1e2	9 ng	missplicing – Supp. Fig. 5D
mnx1 MO2	AAAAGAAAGGTTTACCTGCGTCTCT	splice - e2i2	6 ng	missplicing – Supp. Fig. 5D
<i>ntla</i> cMO	GACTTGAGGCAGA <u>CAT</u> ATTTCCGAT- linker-GCCTCAAGTC	сМО	1.5 ng	mutant phenocopy 36,37
ntla MO	GACTTGAGGCAGA <u>CAT</u> ATTTCCGAT	translational start site	1 ng	mutant phenocopy 36,37
ptrfb MO	GACGGCTGTCTTCAATCACCTC <u>CAT</u>	translational start site	6 ng	previously reported morphant ³⁴
si:ch211-215a10.5-001 (hpcal4) MO1	GTTTTCC <u>CAT</u> CTCTACACCTCACTG	translational start site	6 ng + 9 ng <i>tp53</i> MO	
si:ch211-215a10.5-001 (hpcal4) MO2	GCTTGCTGTTGTGTTTTCC <u>CAT</u> CTC	translational start site	2 ng + 3 ng <i>tp53</i> MO	
snai2	ATACATGTCATTTTCTCACCCGTGT	splice - e2i2	3 ng	missplicing – Supp. Fig. 6E
tacc2	TCCATCTTTGTCCTCTTACCATCA	splice – e7i7	1 ng + 2 ng <i>tp5</i> 3 MO	missplicing – Supp. Fig. 6F
<i>tp53</i> MO	GCGC <u>CAT</u> TGCTTTGCAAGAATTG	translational start site	_	previously reported morphant ³⁸
znf385b MO1	TTCTGGAGGTCTTACCTTGACTGCT	splice - e2i2	3 ng + 4.5 ng <i>tp5</i> 3 MO	missplicing – Supp. Fig. 8K
znf385b MO2	TGGCTCTACAAAGGAGAACAAATGA	splice - i4e5	3 ng + 4.5 ng <i>tp53</i> MO	missplicing – Supp. Fig. 8K

Supplementary Table 11. Phenotype statistics for morpholino experiments in order of presentation.

Figure	Panel	Phenotypic criteria	Total number of embryos	Number with phenotype	% Penetrance
1	c, top	Fluorescent notochord and medial floor plate	20	20	100
1	c, bottom	Fluorescent medial floor plate alone	26	23	88
1	d, top	Fluorescent notochord and neural tube	29	29	100
1	d, bottom	Fluorescent, incompletely vacuolated notochord and neural tube	24	24	100
2	b	Axial and ventral mesoderm expression of <i>lhx1a</i>	21	21	100
2	С	Loss of axial expression of <i>lhx1a</i>	20	20	100
2	d	Axial expression of mnx1	18	18	100
2	е	Loss of axial expression of mnx1	21	21	100
2	f	Tailbud expression of <i>rbm</i> 38	22	22	100
2	g	Loss of axial tailbud expression of <i>rbm38</i>	21	21	100
2	h	Adaxial expression of unc45b	17	17	100
2	i	Reduced adaxial expression of unc45b	23	20	87
2	j	Chordamesodermal and somitic expression of cav3	10	10	100
2	k	Reduced chordamesodermal expression of cav3	10	8	80
2		Chordamesodermal expression of <i>tspan7b</i>	12	12	100
2	m	Reduced chordamesodermal expression of <i>tspan7b</i>	12	12	100
2	n	Chordamesodermal expression of <i>slc38a8</i>	12	12	100
2	0	Reduced chordamesodermal expression of <i>slc38a8</i>	10	10	100
2	р	Chordamesodermal expression of <i>znf385b</i>	16	15	94
2	q	Reduced chordamesodermal expression of <i>znf385b</i>	10	8	80
3	C	Undulating notochord	37	37	100
3	d	Wildtype-like notochord	26	26	100
3	f	Somite patterning defects	24	24	100
3	g	Somite patterning defects	26	26	100
3	h	Axial expression of shha	20	20	100
3	i	Axial expression of <i>shha</i>	17	17	100
3	i	Axial expression of <i>ihhb</i>	20	20	100
3	k	Reduced axial expression of <i>ihhb</i>	20	20	100
3		Adaxial expression of <i>ptc1</i>	21	18	86
3	m	Adaxial expression of <i>ptc1</i>	18	16	89
3	0	Incomplete notochord vacuolization	16	12	75
3	p	Incomplete notochord vacuolization	18	9	50
S1	6 hpf	Ntla expression in the germ ring at wildtype level	8	8	100
S1	7 hpf	Ntla expression in the germ ring and axial mesoderm at wildtype level	8	6	75

S1	8 hpf	Ntla expression in axial mesoderm is reduced where co-localized with FD	8	8	100
S2	12 hpf	Ntla expression in the chordamesoderm at wildtype level	8	8	100
S2	14 hpf	Ntla expression in the chordamesoderm at nearly wildtype level	6	5	100
S2	16 hpf	Ntla expression in the chordamesoderm is reduced where co-localized with FD	7	7	100
S3	aldh1a2, WT	Paraxial expression	18	18	100
S3	aldh1a2, MO	Reduced paraxial expression	21	16	76
S3	b3gnt7, WT	Axial expression	34	34	100
S3	b3gnt7, MO	Loss of axial expression	33	33	100
S3	col8a1a, WT	Axial expression	22	21	95
S3	col8a1a, MO	Loss of axial expression	25	22	88
S3	her1, WT	Tailbud and paraxial expression	23	18	78
S3	her1, MO	Reduced tailbud expression	22	22	100
S3	hpcal4, WT	Axial expression	18	18	100
S3	hpcal4, MO	Loss of axial expression	16	16	100
S3	inpp5b, WT	Axial expression	18	18	100
S3	inpp5b, MO	Loss of axial expression	22	22	100
S3	kirrel3l, WT	Axial and anterior expression	22	22	100
S3	kirrel3l, MO	Loss of axial expression	37	37	100
S3	lhx1a, WT	Axial and lateral expression	18	14	78
S3	lhx1a, MO	Loss of axial expression	20	20	100
S3	Irch2, WT	Axial expression	22	20	91
S3	Irch2, MO	Loss of axial expression	20	20	100
S3	<i>Irrc8da</i> , WT	Axial expression	20	18	90
S3	<i>Irrc8da</i> , MO	Loss of axial expression	21	20	95
S3	mnx1, WT	Axial expression	18	17	94
S3	mnx1, MO	Loss of axial expression	21	21	100
S3	myod1, WT	Adaxial expression	20	20	100
S3	myod1, MO	Reduced adaxial expression	28	28	100
S3	notch1b, WT	Axial, adaxial, tailbud, and anterior expression	19	19	100
S3	notch1b, MO	Loss of axial expression	14	14	100
S3	<i>p4ha1</i> , WT	Axial expression	21	20	95
S3	p4ha1, MO	Loss of axial expression	22	22	100
S3	pak1, WT	Axial expression	21	20	95
S3	pak1, MO	Loss of axial expression	23	23	100
S3	plod1a, WT	Axial expression	19	19	100
S3	plod1a, MO	Loss of axial expression	33	33	100
S3	prickle2, WT	Axial expression	17	17	100
S3	prickle2, MO	Loss of axial expression	35	33	94
S3	pskh2, WT	Axial expression	24	24	100

S3	pskh2, MO	Loss of axial expression	24	22	92
S3	<i>rbm</i> 38, WT	Tailbud expression	22	22	100
S3	<i>rbm</i> 38, MO	Reduced tailbud expression	21	21	100
S3	snai2, WT	Axial expression	17	17	100
S3	snai2, MO	Loss of axial expression	19	18	95
S3	tacc2, WT	Axial expression	18	17	94
S3	tacc2, MO	Loss of axial expression	17	17	100
S3	tbx6, WT	Paraxial expression	19	19	100
S3	tbx6, MO	Reduced paraxial expression	21	21	100
S3	unc45b, WT	Adaxial expression	17	17	100
S3	unc45b, MO	Reduced adaxial expression	23	20	87
S3	wnt8a, WT	Tailbud expression	19	19	100
S3	wnt8a, MO	Reduced tailbud expression	13	11	85
S4	cav3, WT	Chordamesodermal and somitic expression	9	9	100
S4	cav3, no UV	Chordamesodermal and somitic expression	10	10	100
S4	cav3, UV	Reduced chordamesodermal expression	10	8	80
S4	frzb, WT	No expression	11	11	100
S4	frzb, no UV	No expression	9	6	67
S4	frzb, UV	Chordamesodermal expression	9	9	100
S4	guca2b, WT	Ectodermal expression	11	11	100
S4	guca2b, no UV	Ectodermal expression	13	13	100
S4	guca2b, UV	Ectodermal expression	10	10	100
S4	kcnk6, WT	Chordamesodermal expression	18	18	100
S4	<i>kcnk</i> 6, no UV	Chordamesodermal expression	12	12	100
S4	kcnk6, UV	Reduced chordamesodermal expression	12	12	100
S4	<i>ntla</i> , WT	Chordamesodermal expression	6	6	100
S4	<i>ntla</i> , no UV	Chordamesodermal expression	13	13	100
S4	<i>ntla</i> , UV	Increased chordamesodermal expression	6	6	100
S4	<i>ptrfb</i> , WT	Chordamesodermal expression	10	10	100
S4	<i>ptrfb</i> , no UV	Chordamesodermal expression	12	11	92
S4	<i>ptrfb</i> , UV	Reduced chordamesodermal expression	8	8	100
S4	<i>slc38a8</i> , WT	Chordamesodermal expression	9	9	100
S4	<i>slc38a8</i> , no UV	Chordamesodermal expression	12	12	100
S4	<i>slc38a8</i> , UV	Reduced chordamesodermal expression	10	10	100
S4	<i>tspan7b</i> , WT	Chordamesodermal expression	21	21	100
S4	<i>tspan7b</i> , no UV	Chordamesodermal expression	12	12	100
S4	tspan7b, UV	Reduced chordamesodermal expression	12	12	100
S4	<i>zn</i> f385b, WT	Chordamesodermal expression	22	22	100
S4	<i>zn</i> f385b, no UV	Chordamesodermal expression	16	15	94

S5 S5 S5 S5 S5 S6 S6	b c e f b, MO1	Somite patterning defects Somite patterning defects U-shaped somites and ventral body curvature Somite patterning defects	13 21 17	11 19	85 90
S5 S5 S6 S6	e f b, MO1	U-shaped somites and ventral body curvature			90
S5 S6 S6	f b, MO1		17	17	
S6 S6	b, MO1	Somite patterning defects		17	100
S6	,		24	24	100
	1 1/00	Ventral body curvature	24	19	79
	b, MO2	Ventral body curvature	19	11	58
S6	С	Mild necrosis throughout embryo	15	15	100
S6	d, MO1	Wildtype-like morphology	45	45	100
S6	d, MO2	Ventral body curvature	51	42	82
S6	е	Wildtype-like morphology	17	17	100
S6	f	Wildtype-like morphology	15	15	100
S7	<i>mnx1</i> , F59	Shortened slow muscle fibers	18	18	100
S7	<i>mnx1</i> , F310	Wildtype-like fast muscle fibers	21	21	100
S7	<i>ntla</i> , F59	Disorganized, shortened slow muscle fibers	23	23	100
S7	<i>ntla</i> , F310	Shortened fast muscle fibers	20	20	100
S7	Cyc, F59	Disorganized and fewer slow muscle fibers	26	26	100
S7	Cyc, F310	Wildtype-like fast muscle fibers	29	29	100
S8	b	Incomplete notochord vacuolization	35	34	97
S8	С	Reduced cell-cell contacts within the notochord	27	25	93
S8	d	Incomplete notochord vacuolization	16	12	75
S8	е	Incomplete notochord vacuolization	20	14	70
S8	f	Incomplete notochord vacuolization	21	14	67
S8	h	Incomplete notochord vacuolization	18	9	50
S8	i	Incomplete notochord vacuolization	21	8	38
S8	j	Incomplete notochord vacuolization	27	18	67
S9	a, 6-hpf UV	Wildtype-like <i>flh</i> expression	12	12	100
S9	b, 6-hpf UV	Wildtype-like <i>flh</i> expression	12	12	100

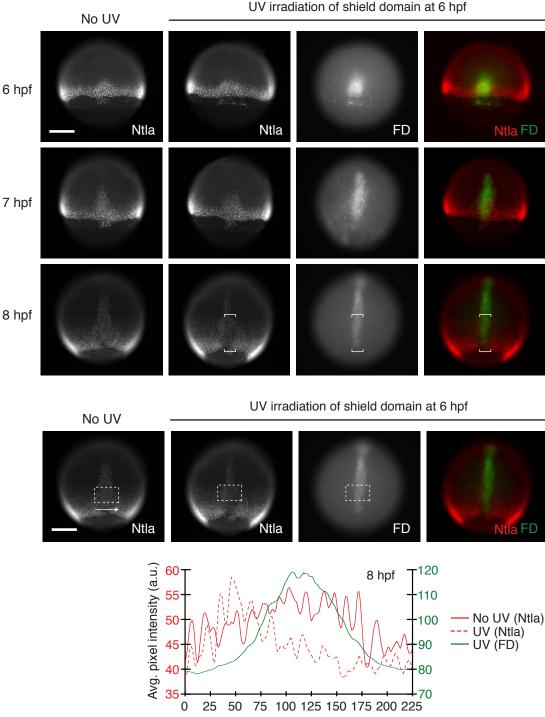
Supplementary Table 12. Primer sequences used to verify MO-induced RNA missplicing.

Transcript	Forward	Reverse
LOC563543 (inpp5b) MO	TTCGGCCTGCGGGATAACCTC	GAACTGGAAGCGGATGGAAACTGC
LOC795255 (slc38a8) MO1	AAGGCTGGCGGCGTGAACA	GAAGCCCATGAGCCCAATCCT
mnx1 MO1 or MO2	CTCCCTGCCCACCTCATCCAT	GGCCTTTTTACTGCGCTTCCATTTC
snai2 MO	GGAAGATGAGGCTCTGCTGGAACG	CGATCTGCGAATGCACGACTGC
tacc2 MO	ATGGCTCAATCAGTCCCTTCAAATCA	GCCCTTCGAAACGCTGCTGTC
znf385 MO1	TGCAACACAGAGTGACGCCAAGTCT	ACCGCGGGCGCAACAGG
znf385 MO2	CCGGTCGGCCTGTTCCCTAAT	GGTTTGCCTGCCACTCTGTCCTT

Supplementary Table 13. Primer sequences used to generate *in situ* probes.

Transcript	Forward	Reverse	
aldh1a2	AGACGCGATGACCTCCAGTGAAGT	CCGAGGTAGAGGCAGGTGAGAGG	
cav3	CGCCGCAGCTGTCTCTACCATT	AGGCCACCCGAACACTGCTAAAG	
col8a1a	CCCGGGCCAAAAGGAGAAGTT	AACCCGGAGAAGGAGGAGTGGA	
frzb	CTGGCCTTCGCATGTCTCCTG	TATCTGGCCCCTCCGCTTTGA	
her1	CCAAAACCTCCGCCTCTGCTC	CCTGGGACGACCGGTAATGAAGT	
kirrel3l	CTGCTGCCGCCACAATGCTAA	TCCCCGTCCGATTCCACATTC	
lhx1a	CGGAGGGCACGGAGCAAAGT	GGGCCCGAAAACATCTGAGGAC	
LOC563543 (inpp5b)	ACTCGCATCTCGCCGCACAC	GAAGCAGCTGGGCAGGTAGGAG	
LOC566628 (pskh2)	GGGCTGGCCTGTTGGGATAAGT	CCGGCGCTGGAGGTTTCTG	
LOC795255 (slc38a8)	CTCATTTGGGGCCATTTTCATCAT	AGTCGCAGCCAGAGTTCCCATTAC	
Irch2	TGAACTGTCCGATCTGCCATTGA	TCAGCCCGTGCCGTTTCTG	
Irrc8da	TGACCATTTGCGATGCCTACACA	TCTGGGATGCTGGTGATGTTGTTATG	
mnx1	CTCCCTGCCCACCTCATCCAT	GGCCTTTTTACTGCGCTTCCATT	
notch1b	ACGGGCGCACAGGTCTGTTG	ATCCCGGTTCGCAGGCACAC	
p4ha1	AGCCGCCTGTTTTGCCGTTAC	ACTGCTGCTCCTACATCCGTGAAG	
pak1	CCGCCCATGAGGAACACCAG	CCCCTGGCCGATCTTCTCAAA	
plod1a	CGCCCGCTCAGAGGATTATGTG	TGAAAGTGGAGGCGTCGTGATG	
prickle2	CTGCGGGGAACACATCGGTATT	GTGGCCCAACATCCTTTTCCTTC	
ptrfb	GGAAGGGGAAGCTGAGGGTGAA	AGCGGGCGTAGATGGTGTGGT	
rbm38	GGATAAAAGTGCAGCGGAAAGAGC	GGGTGGAGATGGCGGAGGAT	
si:ch211-215a10.5-001 (hpcal4)	CAGCAAGCTGGCTCCTGAGGTG	TTTGGCCGCTTCTTTGAACTCC	
snai2	AGCGCTGGAAGATGAGGCTCTG	GCAGGTGGGCTCGGAGGTTC	
tacc2	CCAATCCCCAAGGCATCATACAAC	CGGTCGGCTGTGGGAAATCTT	
tbx6	GGAACGGCCTGTGCTGGACTTA	ATGTGTAGACGGGGCTGGTATTTGTG	
tspan7b	AGCGGAGGGAGGAAAGGCACTA	CAAAGGCAACAAAACCACCGATAAC	
unc45b	GGCAGGTCGGCAGGGTGTAAT	GCAGGCGACCGCATACAGGAT	
wnt8a	ATCGGAAAAATGGGTGGTCGTG	GCCGCCTGCAGCTTCTTCTCT	
wu:fb47f06 (guca2b)	CCGTCGCTTTCCTCGTCGTG	TGCAAGCCGCGAAAGCACA	
zgc:110418 (kcnk6)	CGCCGGTAAAGCCTTCTCCATC	CCGTCCAGCCGTAAACATCAGC	
zgc:91787 (b3gnt7)	GCGGCGGCGTGGATGTT	GACTTTGCGGGCCACCTGAGA	
znf385b	GAGGCACGAAATGGAGCAGGTC	GAGCGGCACGGAGAAACGAG	

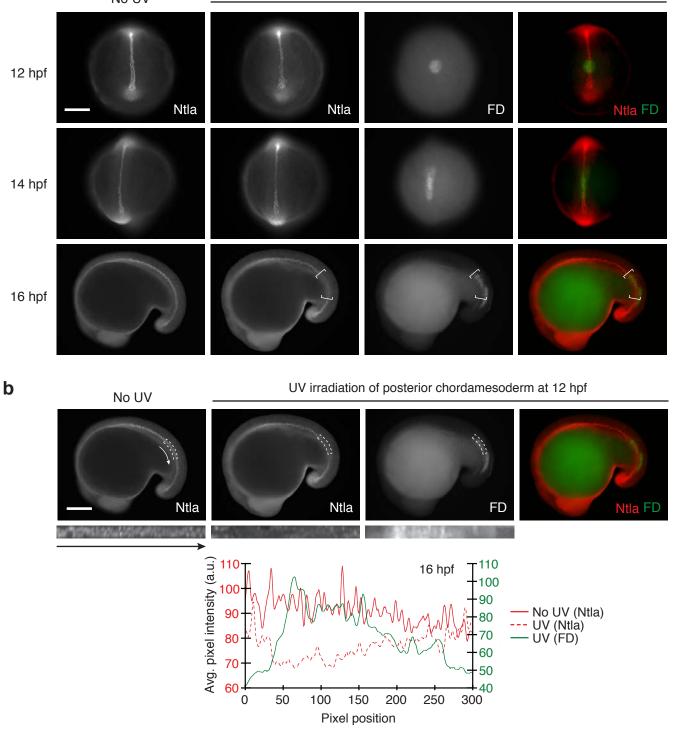
b



Pixel position

Supplementary Figure 1. Ntla protein depletion after *ntla* **cMO photoactivation within the embryonic shield. a**, Zebrafish zygotes were injected with a *ntla* caged morpholino (cMO)/caged fluorescein-conjugated dextran (cFD) mixture and then irradiated within a 100 m-diameter region centered on the shield domain at 6 hours post fertilization (hpf). The irradiated embryos fixed at various developmental time points were assessed by whole-mount immunostaining for Ntla and uncaged fluorescein-conjugated dextran (FD) levels. Non-irradiated embryos were processed and analyzed in an equivalent manner to provide a comparison control, and regions of Ntla protein depletion are indicated by the brackets. b, Quantification of Ntla protein levels in 8-hpf embryos after *ntla* cMO photoactivation. Regions demarcated by the dashed white lines were selected for quantification, and average pixel intensities for each position along the horizontal axis (arrow) were determined using ImageJ software. Pixel intensities for Ntla immunostaining in non-irradiated (red line) and locally irradiated (dashed red line) embryos are shown, as well as those for FD immunostaining in locally irradiated embryos (green line). Embryo orientations: dorsal view and anterior up. Scale bars: 200 m.

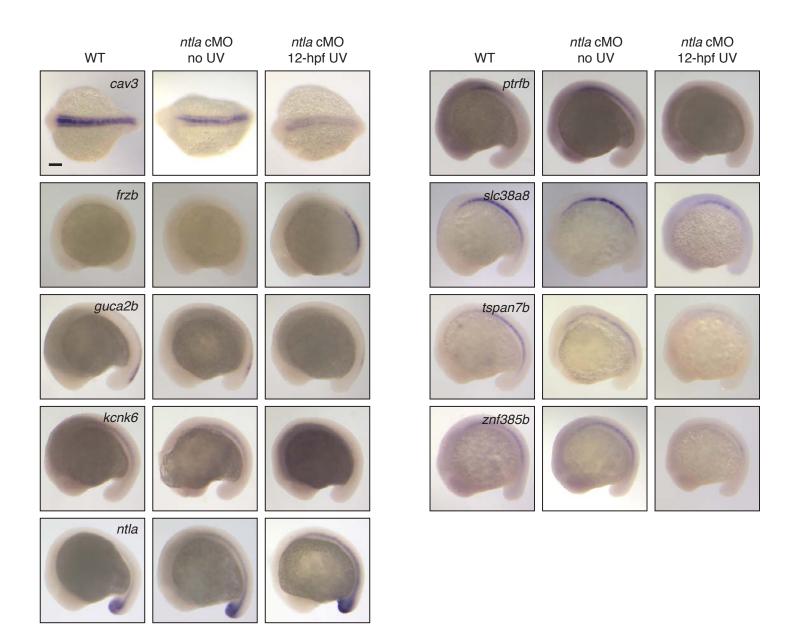
No UV



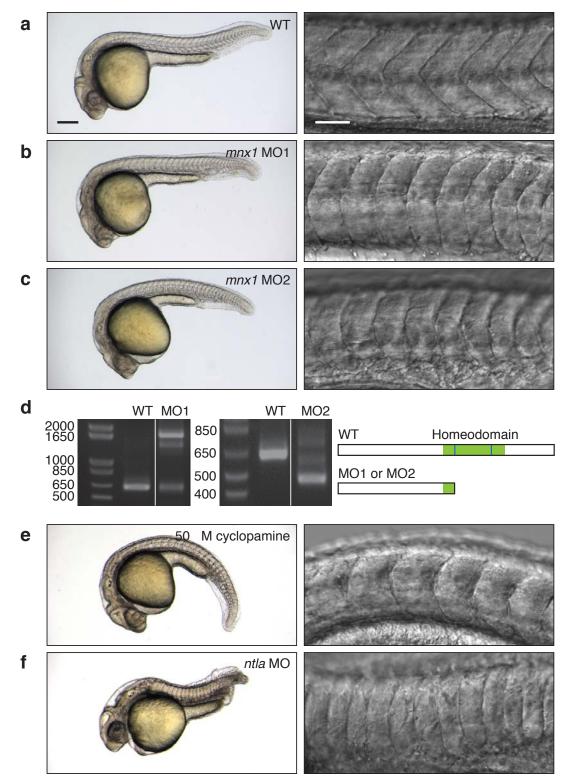
Supplementary Figure 2. Ntla protein depletion after *ntla* cMO photoactivation within posterior chordamesoderm. a, Zebrafish zygotes were injected with a *ntla* cMO/cFD mixture and then were irradiated within a 100 m-diameter region within the posterior chordamesoderm at 12 hpf. The irradiated embryos were fixed at various time points, and Ntla and FD were detected by immunofluorescence. Non-irradiated embryos were processed and analyzed in an equivalent manner to provide a comparison control, and regions of Ntla protein depletion are indicated by the brackets. b, Quantification of Ntla protein levels in 16-hpf embryos after *ntla* cMO photoactivation. Regions demarcated by the dashed white lines were selected for quantification, and image data straightened by ImageJ software is shown below each embryo micrograph. Average pixel intensities for each position along the anterior-posterior axis (arrows) were determined using ImageJ software. Pixel intensities for Ntla immunostaining in non-irradiated (red line) and locally irradiated (dashed red line) embryos are shown, as well as those for FD immunostaining in locally irradiated embryos (green line). Embryo orientations: 12 hpf, dorsal view and anterior up; 14 hpf, posterior dorsal view and dorsal up; 16 hpf, lateral view and anterior left. Scale bars: 200 m.

WT	ntla MO	WT	ntla MO	WT	ntla MO
aldh1a2		b3gnt7 pax2a	0	col8a1a pax2a	
her1	0	hpcal4 pax2a	0	inpp5b pax2a	
kirrel3l	0	lhx1a pax2a	0	Irch2 pax2a	0
Irrc8da pax2a		mnx1 pax2a	0	myod1	
notch1b		p4ha1 pax2a		pak1 pax2a	0
plod1a pax2a		prickle2 pax2a		pskh2 pax2a	
rbm38		snai2 pax2a	0	tacc2 pax2a	\bigcirc
tbx6	0	unc45b	10	wnt8a	0

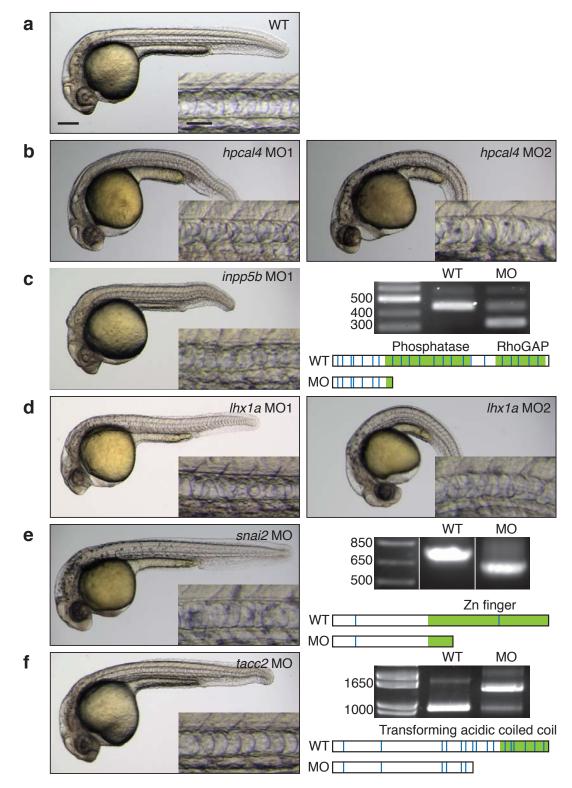
Supplementary Figure 3. Confirmation of selected microarray hits associated with notochord cell fate commitment by *in situ* hybridization. 10-hpf wildtype and *ntla* MO-injected embryos stained for candidate Ntla targets expressed during gastrulation are shown, with co-labeling of *pax2a* transcripts in some cases to determine embryo orientation. All 24 genes tested were confirmed to be transcribed in a Ntla-dependent manner. *kirrel3, lhx1a,* and *notch1b* transcript levels were specifically reduced in the axial mesoderm of *ntla* morphants, whereas their other expression domains were unaffected. Similarly, expression of *her1, rbm38, tbx6,* and *wnt8a* in *ntla* morphants was specifically reduced in the tailbud. In addition to these genes, another hit in our microarray data (**Supplementary Table 1**), *wnt3l,* has previously been shown to be expressed in a Ntla-dependent manner (Ref. 4). Embryo orientations: *her1, rbm38, tbx6, wnt8a,* posterior dorsal view and anterior up; all others, dorsal view and anterior up. Scale bar: 100 m.



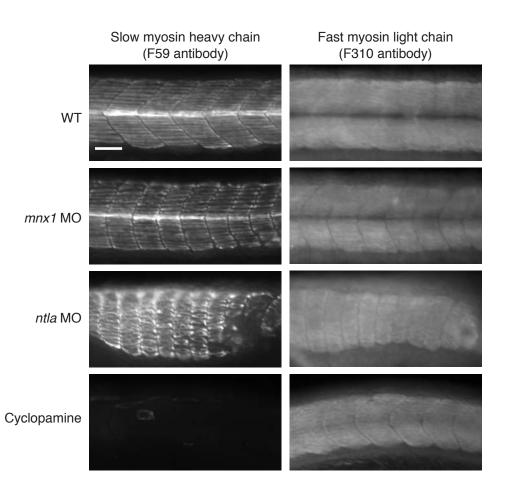
Supplementary Figure 4. Confirmation of selected microarray hits associated with notochord maturation by *in situ* **hybridization.** 16-hpf *ntla* cMO-injected embryos that were either cultured in the dark or globally UV irradiated at 12 hpf are shown. Of the nine genes tested, all but *guca2b* were confirmed to be transcribed within the axial mesoderm in a Ntla-dependent manner. *cav3* transcript levels were specifically reduced in the chordamesoderm of irradiated embryos, whereas somitic expression was unaffected. The increase in *ntla* transcript levels upon *ntla* cMO photoactivation is potentially a consequence of *ntla* mRNA stabilization by the uncaged MO. Embryo orientations: *cav3*, dorsal view and anterior left; all others, lateral view and anterior left. Scale bar: 100 m.



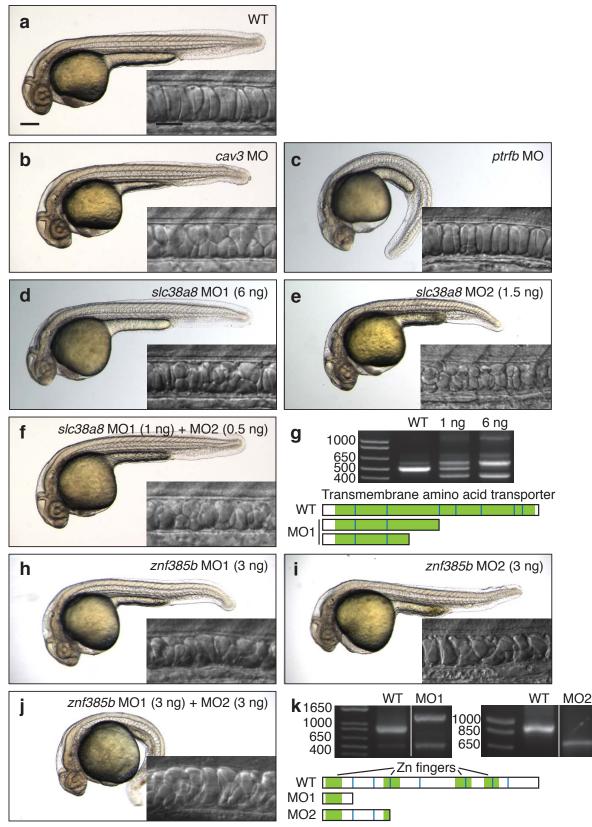
Supplementary Figure 5. Knockdown of *mnx1* expression induces somite morphology defects that differ from Hedgehog pathway loss-of-function phenotypes. a-c, Somite patterning in wildtype embryos and those previously injected with non-overlapping, splice-blocking *mnx1* MOs. Brightfield and DIC micrographs of 1-dpf embryos are shown. d, Confirmation of *mnx1* MO-dependent RNA missplicing by RT-PCR. Mnx1 proteins encoded by the observed transcripts are shown as bar diagrams, with sites corresponding to exon-intron boundaries depicted as blue lines and known protein domains colored green. Exon-intron assignments are based on the Zv8 zebrafish genome assembly Zv8, gene build 59. e, Cyclopamine treatment of embryos starting at 5 hpf resulted in ventral body curvature and U-shaped somites that are shortened along the dorsoventral axis, consistent with loss of Hedgehog pathway-dependent muscle fates. f, *ntla* morphants also exhibited curved somites, but they were morphologically more similar to *mnx1* morphants then to cyclopamine-embryos. Embryo orientations: lateral view and anterior left. Scale bars: whole-embryo brightfield micrograph, 200 μ m; DIC micrograph, 50 m.



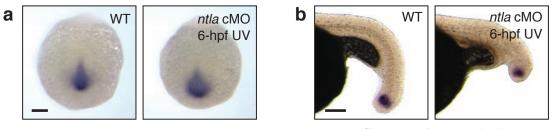
Supplementary Figure 6. Representative morphants of Ntla-dependent genes expressed during gastrulation that do not exhibit clear notochord defects. Brightfield micrographs of selected morphants at 1 day post fertilization (dpf) are shown. In cases where a splice-blocking MO was used, RNA missplicing was confirmed by RT-PCR. Proteins encoded by these transcripts are shown as bar diagrams as described in **Supplementary Figure 5**. **a**, Wildtype control. **b**, Embryos injected with two different translation-blocking MOs targeting *hpcal4* at their maximum non-toxic doses exhibited ventral body curvature but wildtype-like notochord cells. **c**, Embryos injected with a splice-blocking MO targeting *inpp5b* had mild notochord necrosis despite co-injection with a *tp53* MO. **d**, Embryos injected with two different translation-blocking MOs targeting *hx1a* at their maximum non-toxic doses had morphologically normal notochords, although one MO induced ventral curvature and shortening along the anterior-posterior axis. **e-f**, Embryos injected with splice-blocking MOs against *snai2* or *tacc2* had wildtype-like notochords even though the targeted RNA was misspliced as expected. Embryo orientations: lateral view, dorsal up and anterior left. Scale bars: whole-embryo micrograph, 200 m; inset, 50 m.



Supplementary Figure 7. Knockdown of *mnx1* expression induces muscle differentiation defects that differ from Hedgehog pathway loss-of-function phenotypes. Slow myosin heavy chain and fast myosin light chain expression in wildtype zebrafish and embryos treated with an *mnx1* MO, a *ntla* MO, or 50 μ M cyclopamine. 1-dpf embryos are shown. Embryo orientations: lateral view and anterior left. Scale bar: 50 m.



Supplementary Figure 8. Morphants of several Ntla-dependent genes expressed during somitogenesis exhibit aberrant notochord cell morphology. Brightfield and DIC (inset) micrographs of selected morphants at 1.5 dpf are shown. In cases where a splice-blocking MO was used, RNA missplicing was confirmed by RT-PCR. Proteins encoded by these transcripts are shown as bar diagrams as described in **Supplementary Figure 5**. **a**, Wildtype control. **b**, Embryos injected with a *cav3* MO had disorganized, incompletely vacuolated notochord cells. **c**, Embryos injected with a *ptrfb* MO exhibited reduced notochord cell-cell contacts and ventral body curvature. **d-e**, Embryos injected with either splice-blocking or translation-blocking *slc38a8* MOs (MO1 and MO2, respectively) had incompletely vacuolated notochord cells. **f**, Co-injection of both *slc38a8* MOs at sub-phenotypic doses synergistically induced similar notochord defects. **g**, Confirmation of *slc38a8* MO-dependent RNA missplicing by RT-PCR. **h-i**, Embryos injected with splice-blocking *znf385b* MOs (MO1 or MO2) exhibited partially vacuolated notochord cells. **j**, Co-injection of both *znf385b* MOs led to a stronger notochord defect. **k**, Confirmation of *znf385b* MO-dependent RNA missplicing by RT-PCR. Embryo orientations: lateral view and anterior left. Scale bars: whole-embryo brightfield micrograph, 200 m; DIC inset, 50 m.



flh expression at 12 hpf

flh expression at 20 hpf

Supplementary Figure 9. Temporal analysis of Ntla-dependent *flh* **transcription. a-b**, Expression of *flh* at 12 hpf and 20 hpf in wildtype zebrafish, as well as in embryos injected with the ntla cMO and irradiated globally at 6 hpf. Embryo orientations: **a**, dorsal posterior view and dorsal up; **b**, lateral view and anterior left. Scale bars: **a**, 100 m; **b**, 200 m.

SUPPLEMENTARY METHODS

Caged fluorescein dextran	
Ntla polyclonal antibody	
Imaging of zebrafish embryos	
Cyclopamine treatment of zebrafish embryos	

Caged fluorescein dextran (cFD) preparation

Aminodextran (3.5-5 mg of 10-kDa polymer; Invitrogen, D1860) dissolved in 500 μ L of 0.1 M Na₂B₄O₇ buffer (pH 8.5) was added to carboxymethylnitrobenzyl (CMNB)-caged fluorescein N-hydroxysuccinimide ester (1 mg; Invitrogen, C20050) in the manufacturer-supplied tinted tube, and the reaction mixture was vortexed overnight. The resulting cFD was separated from unreacted caged fluorescein using a Zeba Desalt spin column (Thermo Fisher Scientific, 89889) according to the manufacturer's instructions. The yellow-colored eluent was lyophilized to dryness, weighed, dissolved in water to make a 1% (w/v) stock solution, and stored at -20 °C as 2- μ L aliquots. Spectroscopic analysis indicated an average loading of 2.5 caged fluoresceins per dextran molecule.

Ntla polyclonal antibody

Full-length *ntla* cDNA flanked by 5' EcoRI and 3' BamHI sites was generated by PCR using cDNA derived from 10-hpf zebrafish embryos as a template and the following primers: 5'-CGAATTCATGTCTGCCTCAAGTCCCGACCA-3' and 5'-CGGATCCTCAGTAGCTCTGAGCCA-CAGGCG-3'. The *ntla* cDNA was then cloned into a pCR-Blunt II-TOPO vector (Invitrogen), digested with EcoRI and BamHI, and ligated into similarly cut pMALc2x vector (NEB) to generate a bacterial expression construct encoding an Ntla-maltose binding protein (MBP) fusion (pMALc2x-Ntla). The pMALc2X-Ntla vector was transformed into BL21 competent cells and a single colony was cultured in 80 mL LB-ampicillin broth overnight. This starter culture was then used to innoculate 1 L of 2X YT broth (16 g tryptone, 10 g yeast extract, 5 g NaCl, 2 g glucose, 100 mg ampicillin, 1 L water) and cultured for 2 hours at 37 °C, followed by 1 hour at 30 °C to achieve an OD₆₀₀ of 0.8-1.0. IPTG (0.5 mM) was added to the culture, which was grown for another 6 hours at 30 °C. Cells were then harvested by centrifugation (4,000 x *g*, 20 minutes, 4 °C), resuspended in column buffer (50 mL; 200 mM NaCl, 1 mM EDTA, 0.07% (v/v) 2-mercaptoethanol, 20 mM Tris-HCl, pH 7.4), chilled to -72 °C in a dry ice/ethanol bath,

37

thawed in ice water, and sonicated 8 x 15 seconds in ice water. Triton X-100 was added to a final concentration of 1% (v/v), and the lysate was mixed gently at 4 °C for 30 minutes solubilize the Ntla-MBP fusion. The lysate was next centrifuged (12,000 x g, 30 minutes, 4 °C) and the supernatant was passed over amylose resin (5 mL; NEB) that was previously packed into a gravity flow column and equilibrated with the column buffer (40 mL). The loaded amylose resin was washed with column buffer (60 mL) and eluted in 1-mL fractions with column buffer containing 20 mM maltose. Fractions containing the 89 kDa Ntla-MBP fusion protein were pooled and concentrated to 1 mg/mL using an Amicon Centricon centrifugal filter device.

The purified Ntla-MBP fusion protein (4 mg) was cleaved with Factor Xa (40 µg) and Ntla was separated from MBP by DEAE-Sepharose ion exchange chromatography using a pMAL Fusion & Purification System kit (NEB) according to the manufacturer's instructions. Rabbit innoculation and serum collection were conducted by Rockland Immunochemicals using their standard procedures. To affinity purify the polyclonal anti-Ntla antibody, an affinity column was prepared by expressing and purifying the Ntla-MBP fusion protein (4 mg) as described above, dialyzing the protein in 1 L 1X PBS for 12 hours, and immobilizing it onto 1 mL of AminoLink resin (Pierce) according to the manufacturer's instructions. The polyclonal anti-Ntla antibody serum (1 mL) was then passed over the gravity-flow affinity column, eluted with 0.1 M glycine-HCl buffer, pH 2.5-3.0. Each 200-µL fraction containing the purified antibody was then neutralized with 10 µL of 1 M Tris-HCl, pH 9.

Imaging of zebrafish embryos

For live imaging of 24- or 36-hpf zebrafish, the embryos were manually dechorinated and immobilized in E3 medium containing 0.5% (w/v) low-melt agarose and 0.05% (w/v) Tricaine mesylate. Brightfield images were acquired using a Leica M205FA fluorescence stereoscope equipped with a Leica DFC500 digital camera. Differential interference contrast (DIC) micrographs were obtained with a Leica DM4500B epifluorescence microscope equipped

38

with a 20x/0.5 NA water-immersion objective and a QImaging Retiga-SRV digital camera. Confocal fluorescence and DIC images were obtained with a Zeiss Axio Imager Z1 upright microscope equipped with a 20x/0.5 NA water-immersion objective, fluorescence and transmission photomultiplier tube detectors, and an LSM700 laser-scanning confocal head. Fixed embryos stained by *in situ* hybridization were mounted in 1X PBS containing 2% (w/v) methylcellulose and imaged with the Leica M205FA/DFC500 system using LED illumination arrays and the Leica LAS montage imaging module. Fixed immunostained embryos were mounted in 1X PBS containing 0.5% (w/v) low-melt agarose, and fluorescence images were obtained with a DM4500B/Retiga-SRV system equipped with GFP and TXR filter sets and a 10x/0.30 NA objective.

Cyclopamine treatment of zebrafish embryos

5-hpf embryos in 10 mL of E3 medium (5 mM NaCl, 0.17 mM KCl, 0.33 mM CaCl₂, and 0.33 mM MgSO₄) were treated with an ethanolic solution of cyclopamine (5 μ L of a 100 mM stock; 50 μ M final concentration) and cultured at 28.5 °C.

REFERENCES

- 1. Morley, R.H. et al. A gene regulatory network directed by zebrafish No tail accounts for its roles in mesoderm formation. *Proc. Natl. Acad. Sci. U. S. A.* **106**, 3829-34 (2009).
- 2. Goering, L.M. et al. An interacting network of T-box genes directs gene expression and fate in the zebrafish mesoderm. *Proc Natl Acad Sci U S A* **100**, 9410-5 (2003).
- Garnett, A.T. et al. Identification of direct T-box target genes in the developing zebrafish mesoderm. *Development* 136, 749-60 (2009).
- 4. Martin, B.L. & Kimelman, D. Regulation of canonical Wnt signaling by Brachyury is essential for posterior mesoderm formation. *Dev. Cell* **15**, 121-33 (2008).
- 5. Lightcap, C.M. et al. Interaction with LC8 is required for Pak1 nuclear import and is indispensable for zebrafish development. *PLoS One* **4**, e6025 (2009).
- Lewis, K.E., Bates, J. & Eisen, J.S. Regulation of iro3 expression in the zebrafish spinal cord. *Dev. Dyn.* 232, 140-8 (2005).
- Veeman, M.T., Slusarski, D.C., Kaykas, A., Louie, S.H. & Moon, R.T. Zebrafish prickle, a modulator of noncanonical Wnt/Fz signaling, regulates gastrulation movements. *Curr. Biol.* 13, 680-5 (2003).
- Gansner, J.M. & Gitlin, J.D. Essential role for the alpha 1 chain of type VIII collagen in zebrafish notochord formation. *Dev. Dyn.* 237, 3715-26 (2008).
- Eimon, P.M. et al. Delineation of the cell-extrinsic apoptosis pathway in the zebrafish. *Cell Death Differ.* 13, 1619-30 (2006).
- Kottgen, M. et al. TRPP2 and TRPV4 form a polymodal sensory channel complex. *J. Cell Biol.* 182, 437-47 (2008).
- Carney, T.J. et al. Genetic analysis of fin development in zebrafish identifies furin and hemicentin1 as potential novel fraser syndrome disease genes. *PLoS Genet.* 6, e1000907 (2010).

- 12. van Eeden, F.J. et al. Genetic analysis of fin formation in the zebrafish, Danio rerio. *Development* **123**, 255-62 (1996).
- Harvey, S.A. & Logan, M.P. sall4 acts downstream of tbx5 and is required for pectoral fin outgrowth. *Development* 133, 1165-73 (2006).
- 14. Luo, T. et al. Inca: a novel p21-activated kinase-associated protein required for cranial neural crest development. *Development* **134**, 1279-89 (2007).
- Srinivas, B.P., Woo, J., Leong, W.Y. & Roy, S. A conserved molecular pathway mediates myoblast fusion in insects and vertebrates. *Nat. Genet.* **39**, 781-6 (2007).
- Lorent, K. et al. Inhibition of Jagged-mediated Notch signaling disrupts zebrafish biliary development and generates multi-organ defects compatible with an Alagille syndrome phenocopy. *Development* **131**, 5753-66 (2004).
- Milan, D.J., Giokas, A.C., Serluca, F.C., Peterson, R.T. & MacRae, C.A. Notch1b and neuregulin are required for specification of central cardiac conduction tissue. *Development* 133, 1125-32 (2006).
- Yeo, S.Y., Kim, M., Kim, H.S., Huh, T.L. & Chitnis, A.B. Fluorescent protein expression driven by her4 regulatory elements reveals the spatiotemporal pattern of Notch signaling in the nervous system of zebrafish embryos. *Dev. Biol.* **301**, 555-67 (2007).
- 19. Maves, L. et al. Pbx homeodomain proteins direct Myod activity to promote fast-muscle differentiation. *Development* **134**, 3371-82 (2007).
- 20. Etard, C. et al. The UCS factor Steif/Unc-45b interacts with the heat shock protein Hsp90a during myofibrillogenesis. *Dev. Biol.* **308**, 133-43 (2007).
- 21. Chen, Y.H. & Tsai, H.J. Treatment with Myf5-morpholino results in somite patterning and brain formation defects in zebrafish. *Differentiation* **70**, 447-56 (2002).
- Tan, X., Rotllant, J., Li, H., De Deyne, P. & Du, S.J. SmyD1, a histone methyltransferase, is required for myofibril organization and muscle contraction in zebrafish embryos. *Proc. Natl. Acad. Sci. U. S. A.* **103**, 2713-8 (2006).

- 23. Seeley, M. et al. Depletion of zebrafish titin reduces cardiac contractility by disrupting the assembly of Z-discs and A-bands. *Circ. Res.* **100**, 238-45 (2007).
- van Eeden, F.J. et al. Mutations affecting somite formation and patterning in the zebrafish, Danio rerio. *Development* **123**, 153-64 (1996).
- 25. Wingert, R.A. et al. The cdx genes and retinoic acid control the positioning and segmentation of the zebrafish pronephros. *PLoS Genet.* **3**, 1922-38 (2007).
- 26. Alexa, K. et al. Maternal and zygotic aldh1a2 activity is required for pancreas development in zebrafish. *PLoS One* **4**, e8261 (2009).
- Bessa, J. et al. meis1 regulates cyclin D1 and c-myc expression, and controls the proliferation of the multipotent cells in the early developing zebrafish eye. *Development* 135, 799-803 (2008).
- Lekven, A.C., Thorpe, C.J., Waxman, J.S. & Moon, R.T. Zebrafish wnt8 encodes two wnt8 proteins on a bicistronic transcript and is required for mesoderm and neurectoderm patterning. *Dev. Cell* 1, 103-14 (2001).
- Thorpe, C.J., Weidinger, G. & Moon, R.T. Wnt/beta-catenin regulation of the Sp1-related transcription factor sp5l promotes tail development in zebrafish. *Development* **132**, 1763-72 (2005).
- Shimizu, T., Bae, Y.K., Muraoka, O. & Hibi, M. Interaction of Wnt and caudal-related genes in zebrafish posterior body formation. *Dev. Biol.* 279, 125-41 (2005).
- Draper, B.W., Stock, D.W. & Kimmel, C.B. Zebrafish fgf24 functions with fgf8 to promote posterior mesodermal development. *Development* 130, 4639-54 (2003).
- 32. Hanai, J. et al. The muscle-specific ubiquitin ligase atrogin-1/MAFbx mediates statininduced muscle toxicity. *J. Clin. Invest.* **117**, 3940-51 (2007).
- 33. Oates, A.C. & Ho, R.K. Hairy/E(spl)-related (Her) genes are central components of the segmentation oscillator and display redundancy with the Delta/Notch signaling pathway in

the formation of anterior segmental boundaries in the zebrafish. *Development* **129**, 2929-46 (2002).

- 34. Hill, M.M. et al. PTRF-Cavin, a conserved cytoplasmic protein required for caveola formation and function. *Cell* **132**, 113-24 (2008).
- Nixon, S.J. et al. Zebrafish as a model for caveolin-associated muscle disease; caveolin-3 is required for myofibril organization and muscle cell patterning. *Hum. Mol. Genet.* 14, 1727-43 (2005).
- Ouyang, X. et al. Versatile synthesis and rational design of caged morpholinos. J. Am. Chem. Soc. 131, 13255-69 (2009).
- 37. Shestopalov, I.A., Sinha, S. & Chen, J.K. Light-controlled gene silencing in zebrafish embryos. *Nat. Chem. Biol.* **3**, 650-1 (2007).
- 38. Robu, M.E. et al. p53 activation by knockdown technologies. PLoS Genet. 3, e78 (2007).