Disorders of Bile Acid Metabolism in Cholesterol Gallstone Disease

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Abstract

The aim of the study was to evaluate the metabolism of individual bile acids in patients with cholesterol gallstone disease. Therefore, we determined pool size and turnover of deoxycholic (DCA), cholic (CA), and chenodeoxycholic acid (CDCA) in 23 female gallstone patients classified according to their gallbladder function and in 15 healthy female controls. Gallstone patients had normal hepatic bile acid synthesis, but, depending on gallbladder function, differed with respect to turnover and size of the bile acid pools: Patients with well-emptying gallbladder (group A, n = 9) had enhanced turnover and reduced pools of CA (-46%; P < 0.01 vs. controls) and CDCA (-24%; P < 0.05), but normal input and size of the DCA pool. With reduced gallbladder emptying (< 50% of volume; group B, n = 6), turnover and pools of CA, CDCA, and DCA were similar as in controls. Patients with loss of gallbladder reservoir (group C, n = 8) had increased input (+100%; P < 0.01) and pool size of DCA (+45%; P = 0.07) caused by rapid conversion of CA to DCA, while the pools of CA (-71%; P < 0.001 vs. controls) and CDCA (-36%; P < 0.05) were reduced by enhanced turnover. Thus, in patients with cholesterol gallstones, the pools of primary bile acids are diminished, unless gallbladder emptying is reduced. Furthermore, in a subgroup of gallstone patients, who had completely lost gallbladder function, the CA pool is largely replaced by DCA owing to rapid transfer of CA to the DCA pool. This probably contributes to supersaturation of bile with cholesterol. (J. Clin. Invest. 1992. 90:859-868.) Key words: bile acid metabolism • cholesterol gallstone disease • deoxycholic acid • gallbladder emptying • cholesterol saturation of bile Introduction

In most nonobese patients with cholesterol gallstones, the pools of cholic acid $(CA)^1$ and chenodeoxycholic (CDCA) are reduced (1-6), and deoxycholic acid (DCA) is often increased in bile (7). Both changes could contribute to supersaturation of bile with cholesterol (1, 7, 8): the diminution of the bile acid pools by reducing the mass of bile acids in the enterohepatic

J. Clin. Invest. © The American Society for Clinical Investigation, Inc. 0021-9738/92/09/0859/10 \$2.00 Volume 90, September 1992, 859-868 circulation (9), the increase of DCA by raising biliary secretion of cholesterol (10).

The small pools of CA and CDCA may be caused by a small gallbladder reservoir (11), by enhanced turnover (2, 5), or by inhibition of bile acid synthesis by an oversensitive feedback mechanism (12) or by elevated levels of DCA (13, 14). An increased fraction of DCA in bile could be caused by high input and large pool size of DCA. CA is nearly completely 7α -dehydroxylated to DCA by anaerobic bacteria in the colon (7, 15, 16), but only 30–40% of this DCA is absorbed from the intestine (17). The DCA pool could be expanded by increased input of DCA owing to increased synthesis of the precursor CA or to an increased fraction of CA transferred to the DCA pool.

It is still unclear which of the above factors account(s) for the reduction in CA and CDCA pool size and which for an increase in biliary DCA. Therefore, we studied the size and the turnover of the pools of CA, CDCA, and DCA in gallstone patients classified according to reservoir volume and emptying of the gallbladder.

Methods

Study subjects (Table I). The study comprised 15 female controls with normal gallbladder function and 23 female patients with radiolucent gallbladder stones. All were Caucasians. The mean age and the mean body mass index of the control subjects were 34 yr and 23.09 kg/m², respectively, and of the gallstone patients 44 yr and 24.35 kg/m². None was grossly obese (> 30 kg/m²) or had evidence of hepatic, intestinal, or renal disease, cholangitis, diabetes mellitus, or thyroid dysfunction. None had taken antibiotics, lipid-lowering drugs, or hormones within the last four weeks. Control subjects were 13 healthy, paid volunteers recruited among employees at the University Hospital (Munich) and two otherwise healthy patients, one with a gallbladder polyp and the other with polycystic liver disease.

All participants were classified according to the reservoir function and the emptying of the gallbladder. Gallstone patients with reservoir function (n = 15) showed on ultrasound normal wall echoes of the gallbladder, which was filled to < 30% (vol/vol) with stones; 13 of them had an opacified gallbladder on oral cholecystography, 2 had no cholecystography, but contracted the gallbladder in response to cholecystokinin (CCK). Emptying of the gallbladder was classified as normal, when on ultrasound $(18) \ge 50\%$ of the volume had emptied in response CCK (19, 20). Nine of these gallstone patients had normal emptying of the gallbladder (group A, n = 9) and six had reduced emptying (group B, n = 6). The gallstone patients without reservoir function of the gallbladder (group C, n = 8) showed no opacification on oral cholecystography. Occlusion of the cystic duct (n = 5) or a shrunken gallbladder (n = 3) were confirmed by endoscopic retrograde cholangiography and/or subsequent cholecystectomy. Controls showed on ultrasound a sludge-free lumen, normal wall echoes, and normal emptying of the gallbladder in response to CCK.

Study protocol. The protocol had been approved by the Ethics Committee of the Hospitals of the University of Munich. All participants gave their written informed consent. The protocol (21) consisted of (a) stable isotope kinetics of CA, CDCA, and DCA (22), (b) serial sonographic estimation of gallbladder volume (18) during an intrave-

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^{1.} *Abbreviations used in this paper:* CA, cholic acid; CCK, cholecystokinin; CDCA, chenodeoxycholic acid; DCA, deoxycholic acid; FTR, fractional turnover rate.

					Body	Dietary cholesterol	Serum		
Subject No.	Diagnosis	Treatment*	Age	Body weight	mass index	cholesterol intake	Triglycerides	Cholesterol	Stool frequency
			(yr)	(kg)	(kg/m²)	(mg/d)	(mg/dl)	(mg/dl)	(d ⁻¹)
Gallston	e patients								
		Group A:	with prese	rved reservo	ir and empty	ing of the gallbl	adder		
1	GBS	CCX	39	71	27.73	627	151	227	0.6
2	GBS	CCX	44	67	24.61	267	56	160	1.0
3	GBS [‡]	CCX	38	55	23.81	349	130	189	1.0
4	GBS	CCX	35	76	26.30	207	90	156	1.0
5	GBS	ESWL	35	66	25.78	624	44	151	0.4
6	GBS	ESWL	42	60	21.26	286	109	189	1.0
7	GBS	none	27	54	20.83	320	87	164	0.7
8	GBS	CCX	34	53	23.56	524	154	192	1.0
9	GBS	ESWL	41	67	24.61	328	118	209	1.0
\bar{X}	(n = 9)		37.2	63.2	24.28	392	104	182	0.86
SD	. ,		5.1	8.1	2.24	158	39	26	0.23
		Group B: with	preserved	reservoir an	d reduced er	nptying of the g	allbladder		
10	CBS	CCX.	27	50	77 57	450	02	104	1.0
10	GBS		37	50 70	25.55	450	83 184	194	1.0
12	GBS		30 11	70	23.71	430	104	233	0.0
12	CBS	ESWI	44	/ I 5 A	23.72	373	129	239	1.0
13	CBS	ESWL	49	50	21.09	300	184	208	1.2
14	GBS	ESWL	44 38	39 71	25.05	404	8 <i>3</i> 54	187	0.5
			41.7	(2.0	20.10			107	1.0
X SD	(n=6)		41.7 48	63.8 77	23.71	436 64	114	213	0.88
50			4.0		1.04	04	-10	21	0.27
		Gro	oup C: with	out reservoir	r function of	the gallbladder			
16	OCD, GBS	CCX	40	66	23.95	363	174	114	1.00
17	SGB, GBS	CCX	45	53	21.50	404	69	232	1.00
18	SGB, GBS	CCX	48	78	26.99	440	107	245	0.60
19	OCD, GBS	none	80	63	23.71	400	78	206	0.70
20	OCD, GBS	CCX	68	63	24.11	340	320	197	0.80
21	OCD, GBS	CCX	30	90	29.79	—	183	241	0.80
22	SGB, GBS	CCX	45	73	25.86	570	215	229	1.00
23	SGB, OCD, GBS	CCX	69	62	23.34	415	108	246	2.00
\bar{X}	(n=8)		53.1	68.5	24.91	419	157	214	0.99
SD			17.2	11.5	2.57	74	84	44	0.44
				Groups A	, B, and C				
\bar{v}	(n - 22)		42.0	65 7	24.25	415	127	201	0.01
SD	(n - 23)		43.9 12.6	9.2	24.33	413	64	35	0.31
Control	nubients								
Control	Subjects								
1	GBpolyp	CCX	44	52	21.10	196	71	218	1.0
2	Healthy	none	47	69	24.74	566	62	165	0.8
3	Healthy	none	33	55	20.96	585	162	195	0.8
4	Healthy	none	26	72	25.21	234	74	176	1.0
5	Healthy	none	23	62	20.24	259	68	186	1.2
6	Healthy	none	26	64	24.69	543	52	. 214	0.6
7	Healthy	none	47	64	25.64	325	90	231	1.0
8	Healthy	none	23	52	18.21	272	67	183	1.0
9	Healthy	none	22	59	22.21	602	92	179	1.2
10	Healthy	none	45	71	26.40	334	88	213	0.8

Table I. Characteristics of Gallstone Patients and Control Subjects

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					Body	Dietary	Serum lipids		
Subject No.	Diagnosis	Treatment*	Age	Body weight	mass index	intake	Triglycerides	Cholesterol	frequency
			(yr)	(kg)	(kg/m²)	(mg/d)	(mg/dl)	(mg/dl)	(d ⁻¹)
Control	subjects (continued)								
11	Healthy	none	36	58	20.80	218	96	238	1.0
12	Healthy	none	23	67	24.61	239	155	195	0.6
13	PLD	CCX	48	66	24.84	253	110	232	0.8
14	Healthy	none	36	50	19.53	435	82	204	1.0
15	Healthy	none	33	72	27.13	188	115	245	1.2
\bar{X}	(n = 15)		34.1	62.2	23.09	350	92	205	0.93
SD			10.0	7.5	2.78	153	32	25	0.20

Abbreviations: CCX, cholecystectomy; ESWL, extracorporeal shock wave lithotripsy; GB, gallbladder; GBS, gallbladder stones; OCD, occluded cystic duct; PLD, polycystic liver disease; SGB, shrunken GB.

(4)

* All studies were concluded before treatment.

[‡] Subject had an aortic bioprosthesis for 1 yr.

nous infusion of CCK (19, 20) with (c) sampling of unstimulated and CCK-stimulated duodenal bile and (d) records of dietary intake and bowel habits kept for 7 d to estimate daily intake of cholesterol (23) and average stool frequency.

Bile acid kinetics. Pool sizes and turnover rates of CA, CDCA, and DCA were simultaneously determined from postprandial serum samples obtained before and for 4 d after oral intake of a single dose of marker bile acids (50 mg each) which were labeled with stable isotopes (¹³C or ²H) (22). In the first three participants (subjects 1 and 2 in group A and subject 1 in controls), only kinetics of CDCA and DCA were determined using ¹³C-labeled marker bile acids; their CA pool size was calculated as the product of the CDCA pool size times the CA/ CDCA molar ratio of intraoperative gallbladder bile (24). Marker bile acids (24-13C-CA, 90% 13C; 24-13C-CDCA, 91.9% 13C; 24-13C-DCA, 91.5% ¹³C; 2,2,4,4-²H₄-DCA, 99.4% ²H₄) were purchased from Merck Sharp & Dohme, Montreal, Canada. Materials, sample preparation, and measurement of isotope ratios of individual bile acids using combined capillary gas-liquid chromatography-mass spectrometry-selected ion monitoring have previously been described (22). From the isotopic enrichment R and the natural abundance R_0 (before intake of label) of ¹³C or ²H₄ of the respective bile acid, atom percent excess (APE) was calculated (22):

$$APE = R - R_0 / 1 + (R - R_0) \times 100(\%)$$
⁽¹⁾

The APE vs. time curve showed monoexponential first-order decay with excellent fit (r > 0.90). Pool size and synthesis rate were calculated:

Pool size = dose
$$\times b \times 100/e^a$$
 - dose (2)

Synthesis rate = pool size
$$\times$$
 FTR (3)

where *a* is the *y*-intercept and fractional turnover rate (*FTR*) the slope of the first-order decay curve of APE, and *b* the degree of labeling of the marker bile acid. Hepatic synthesis rate of bile acids was calculated as the sum of CA and CDCA synthesis rates and total bile acid pool size as the sum of the pools of CA, CDCA, and DCA. The transfer of ¹³C label from CA to the DCA pool was estimated from the amount of excess [¹³C]DCA (21):

Excess^{[13}C]DCA_t =
$$APE_{1^{13}C|DCA,t} \times \text{pool size}_{DCA}$$

imes body weight/100

where APE_t is the APE in the sample drawn at time t after intake of 50 mg of [¹³C]CA. The pool size of DCA was estimated from the dose and the decay of [²H₄]DCA.

Gallbladder reservoir and emptying (Table II). The fasting volume was calculated according to the sum of cylinders method (18) from sonographic images of the gallbladder in the maximum longitudinal and anterior-posterior diameters. The recordings (Sonolayer B, Toshiba Corp., Tokyo) were taken in triplicate on the morning after a 12-h fast. The average coefficient of variation for triplicate determination (n = 30) was $6.0\% \pm 3.9\%$ ($\bar{X} \pm SD$). Serial volumes (every 5 min) and the first-order rate constant of emptying were determined (18) during a subsequent intravenous infusion of CCK (0.02 U kg⁻¹min⁻¹; Kabi-Vitrum, Stockholm) (20).

The fasting volume of the gallbladder was 24.3 ± 8.8 ml in gallstone patients with gallbladder reservoir (groups A and B) and 21.6 ± 8.8 ml in controls (NS). The residual volume (10.6 ± 7.9 vs. 2.3 ± 1.6 ml; P< 0.01), even when corrected for stone volume (Table II; P < 0.05), was larger in gallstone patients than in controls and the fraction of volume emptied (F_e) was diminished ($66\pm27\%$ vs. $89\pm6\%$; P < 0.02). The gallstone patients with reduced emptying of the gallbladder (group B) differed from gallstone patients with normal emptying (group A) with regard to the fraction of gallbladder volume emptied ($38\pm10\%$ vs. $86\pm12\%$), residual volume (14.5 ± 6.7 vs. 2.7 ± 2.8 ml, corrected for stone volume; P < 0.01), and rate of emptying (0.020 ± 0.007 vs. 0.069 ± 0.035 min⁻¹; P < 0.01) (Fig. 1).

Analysis of bile and gallstones. Duodenal bile was sampled through a thin Teflon duodenal tube between 8 and 10 a.m. after an overnight (12 h) fast. Bile was collected during the CCK infusion from 10 to 20 min and from 20 to 30 min, light-shielded on ice, and the darkest fraction sampled (3 ml). The samples were kept in chloroform/methanol (1:2 vol/vol; 0.5 ml of bile per 7.5 ml of solvent) at -20° C and analyzed for cholesterol (25) and lipid-soluble phosphorus (26) by colorimetric tests and for bile acids by gas-liquid chromatography (27). Cholesterol saturation was calculated (28) assuming a total lipid concentration of 10 g/dl in undiluted bile (29). Gallstones obtained at cholecystectomy were assessed for volume (by water displacement) and cholesterol content (21). In seven patients gallstone volume (V) was calculated from sonographic estimates of the maximum diameter $(d_1 = 2a)$ and the maximal perpendicular diameter $(d_2 = 2b)$ assuming a three-dimensional ellipsoid $(V = 4/3 a b^2)$ (Table II).

Statistical analysis. Data are given as $\bar{X}\pm$ SD. The statistical significance of differences between controls and patient groups A-C was tested by one way ANOVA using the program Solo 101 of BMDP

	.	Fasting	Residual					Gallstones	
	volume	volume (corr.)*	(corr.)*	F _e	k,	t _{1/2}	n	Volume	Cholestero
		(<i>ml</i>)		(%)	(min ⁻¹)	(min)		(ml)	(%dry wt)
Gallstone pati	ents								
		(Group A: with re	eservoir and e	mptying of the	gallbladder			
1	19.0	16.0	8.0	50.0	0.058	12.0	2	3.0	79
· 2	32.0	24.5	8.5	65.3	0.059	11.8	2	7.5	65
3	18.8	18.3	1.0	94.5	0.100	6.9	4	0.5	73
4	15.5	14.4	0.4	97.2	0.090	7.7	40	1.1	70
5	20.5	17.4	1.6	90.8	0.050	13.9	1	3.1	ND
6	36.5	34.3	1.0	97.1	0.140	5.0	2	2.2	ND
7	10.3	8.6	1.3	84.9	0.045	15.4	3	1.7	ND
8	23.1	20.3	0.6	97.0	0.039	17.8	4	2.8	78
9	34.8	25.8	5.3	79.5	0.037	18.7	1	9.0	ND
$\bar{X}(n=9)$	23.4	20.0	2.7	86.1	0.069	12.1	7	3.4	73
SD	9.1	7.5	2.8	12.4	0.035	4.8	13	2.9	6
		C D	· · · · · · · · · · · · · · · · · · ·				1.1		
		Group B:	with preserved i	eservoir and	reduced empty	ing of the gail	Diadder		
10	19.5	17.3	11.3	34.7	0.023	30.1	76	2.2	75
11	20.0	16.5	9.5	42.4	0.013	53.3	8	3.5	83
12	39.5	38.0	22.0	42.1	0.024	28.9	2	1.5	78
13	15.4	14.0	8.2	41.4	0.012	57.8	1	1.4	ND
14	28.2	22.2	12.0	45.9	0.029	23.9	2	6.0	73
15	31.5	29.7	24.0	19.2	0.020	34.7	1	1.8	ND
X(n=6)	25.7	23.0	14.5	37.6	0.020	38.1	15	2.7	77
SD	9.0	9.2	6.7	9.7	0.007	14.0	30	1.8	4
			C	Groups A and	B (<i>n</i> = 15)				
Ā	24.3	21.2	7.7	65.5	0.049	22.5	10	3.2	75
SD	8.8	8.0	7.5	27.3	0.036	16.0	21	2.5	5
			Group C: with	out reservoir f	unction of the	gallbladder			
16		08	010 up 01				0	0.5	70
10	12.5	0.	0.	0	0		8	8.3	/8
10	12.5	3.5	3.5	0	0		9	9.2	83
18	2.0	1.0	1.0	0.	0			1.0	//
19		0	0					11.0*	ND
20		0	0				3	7.5	ND
21		0	0				32	8.6	67
22 23	12.5	2.0	2.0	0	0		9	10.5	71 80
2J		0	0.0	0	0		1	0.0	50
X SD	9.0 5.7	0.8	0.8	0	0		10	8.0 3.1	6
50	5.7	1.5	1.5				10	5.1	Ū
Controls with	gallbladder rese	ervoir and emp	tying						
1	13.0		1.5	88.5	0.100	6.9	0		
2	18.5		1.5	91.5	0.059	11.7	0		
3	45.5		2.0	95.6	0.067	10.3	0		
4	12.5		1.5	88.0	0.086	8.1	0		
5	17.0		1.2	92.9	0.081	8.6	0		
6	22.0		4.4	80.0	0.033	21.0	0		
7	17.7		2.2	87.4	0.062	11.2	0		
8	29.9		7.4	75.3	0.034	20.4	0		
9	24.0		2.0	91.7	0.056	12.4	0		

Table II. Gallbladder Motility and Gallstone Characteristics

Tab	le II.	(Continued)	

		Fasting		Residual			Gallstones			
	Fasting volume	volume (corr.)*	volume (corr.)*	F _e	k,	t _{1/2}	n	Volume	Cholesterol	
		(ml)		(%)	(min ⁻¹)	(min)		(ml)	(% dry wt)	
Controls with a	gallbladder rese	rvoir and empt	ying (<i>continued</i>)						
10	28.0		2.5	91.7	0.059	11.7	0			
11	24.0		1.2	95.0	0.106	6.5	0			
12	14.5		2.2	84.8	0.080	8.7	0			
13	15.0		1.5	90.0	0.084	8.3	0			
14	28.5		1.4	95.1	0.082	8.5	0			
15	14.2		1.4	90.1	0.060	11.6	0			
$\bar{X}(n=15)$	21.6		2.3	89.2	0.070	11.1	0			
SD	8.8		1.6	5.6	0.021	4.3	0			

 F_e , fraction (%) of gallbladder fasting volume emptied (18); k_e , fractional rate constant of emptying (18) during CCK infusion (20); ND, not determined.

* Corrected by subtraction of total volume of stones.

[‡] Several stones with estimated total volume of 11 ml.

[§] Occluded cystic duct of the gallbladder. Therefore the corrected fasting and residual volumes were arbitrarily set to zero.

Statistics Corp., Sepulveda, CA. Differences were confirmed by unpaired t test (for equal variance) or by Satterthwaite's t test (for unequal variance) at the level of P < 0.05 (30). Associations between variables were searched with scatter plots and confirmed by linear regression analysis (31).

Results

Bile acid pools (Table III). The total group of gallstone patients had a slightly smaller (-26%; P < 0.01) total bile acid pool (52.4 ± 14.9 mol kg⁻¹; n = 23) than controls (70.7 ± 20.5 mol kg⁻¹; n = 15), reduced pools of CA (-42%, P < 0.01) and CDCA (-29%, P < 0.02) and a normal pool of DCA (+8%). The pools of CA, CDCA, and DCA turned over at higher fractional rates (+46%, P = 0.07; +46%, +39%, P < 0.02). Sub-



Figure 1. Time course of emptying of the gallbladder during intravenous infusion of CCK (0.02 Ivy dog units kg⁻¹min⁻¹). $\bar{X}\pm$ SEM; gallbladder volume not corrected for stone volume. (\odot) Controls (n = 15); (\blacksquare) gallstone patients with well-emptying gallbladder (group A; n = 9); (\blacklozenge) gallstone patients with reduced emptying (< 50) of the gallbladder (group B; n = 6); (\blacktriangle) gallstone patients without gallbladder reservoir (group C; n = 8). *Significantly different (P < 0.05).

groups (A-C) of gallstone patients classified according to gallbladder function showed three different bile acid pool patterns (Fig. 2): (a) Patients with normal emptying (group A) had reduced pools of total bile acids (-33%, P < 0.05), CA (-46%, PP < 0.01), and CDCA (-24%, P < 0.05 vs. controls) with enhanced fractional turnover (CA +54%, P < 0.05; CDCA +38%, NS) and normal size and FTR of the DCA pool. (b) Patients with defective emptying of the gallbladder (group B) had normal-sized pools of total bile acids, CA, and DCA. The CDCA pool was reduced by 23%, but this was not statistically significant. The fractional turnover rates of CA, CDCA, and DCA were normal. (c) The patients without gallbladder reservoir (group C), however, had an expanded DCA pool (+45%, P = 0.07) combined with reduced pools of CA (-71%, P < 0.001) and CDCA (-37%, P < 0.05). Their fractional turnover rates of CA (+97%, P < 0.05) and CDCA (+83%, P < 0.01 vs. control) were enhanced (Table III).

Hepatic synthesis of bile acids was determined in 16 gallstone patients and 13 controls. Gallstone patients and controls had a normal rate of hepatic synthesis of bile acids (15.6 ± 4.6 vs. 16.1 ± 8.5 mol kg⁻¹ d⁻¹), both of CA and CDCA (Table III, groups A-C vs. controls). In one control (subject 10) and five gallstone patients (subject 9 in group A and subjects 16, 18, 20, and 22 in group C), CA synthesis was not measurable, in that the [13 C]CA marker was converted to [13 C]DCA within 24 h (see below). The input rate of DCA in these subjects (9.4 ± 3.2 mol kg⁻¹ d⁻¹) was comparable to the synthesis rate of the precursor CA in controls (10.2 ± 5.6 mol kg⁻¹ d⁻¹); the sum of their CDCA synthesis and DCA input rates (19.3 ± 3.7 mol kg⁻¹ d⁻¹; n = 5) matched (+20%, NS) hepatic synthesis rate of bile acids in controls.

Input of DCA was 38% higher in the total group of gallstone patients $(5.8\pm3.1 \ \mu\text{mol} \ \text{kg}^{-1}\text{d}^{-1})$ than in controls $(4.2\pm3.0 \ \mu\text{mol} \ \text{kg}^{-1}\text{d}^{-1})$, but this difference did not reach statistical significance. Only subgroup C showed a significant twofold increase of DCA input (P < 0.01 vs. controls and group A&B). Except for one woman (subject 17), all in group C had the same disorder of bile acid kinetics characterized by enhanced

Subject		CDCA			CA			DCA		Transfer of
No.	Pool	Synthesis	FTR	Pool	Synthesis	FTR	Pool	Input	FTR	Ca to DCA*
	(µmol kg ⁻¹ body wt)	(µmol kg⁻¹ d⁻¹)	(d⁻¹)	(µmol kg ⁻¹ body wt)	(µmol kg ⁻¹ d ⁻¹)	(d ⁻¹)	(µmol kg ⁻¹ body wt)	(µmol kg⁻¹ d⁻¹)	(d ⁻¹)	(%)
Gallstone _J	patients									
		C	Group /	A: with reservoir an	nd emptying of	the gal	lbladder			
1	19.4	5.2	0.27	17.7 [‡]	ND	ND	13.3	2.6	0.20	ND
2	14.8	2.5	0.17	20.7 [‡]	ND	ND	9.2	1.8	0.28	ND
3	14.5	7.5	0.52	12.9	14.3	1.18	14.4	5.7	0.40	40
4	13.6	5.4	0.40	17.9	8.1	0.45	16.2	5.9	0.37	73
5	12.9	4.8	0.37	14.4	5.2	0.36	11.4	3.9	0.34	75
6	42.4	11.4	0.27	17.1	10.6	0.62	3.9	1.9	0.49	18
7	16.9	5.1	0.30	22.0	10.8	0.49	7.7	5.0	0.64	46
8	25.7	5.4	0.21	14.7	5.0	0.34	2.5	1.0	0.39	19
9	23.0	11.3	0.49	0			24.1	7.9	0.33	
$\bar{X}(n=9)$	20.4	6.5	0.33	15.3	9.0	0.57	11.4	4.0	0.38	45.2
SD	9.4	3.0	0.12	6.4	3.6	0.31	6.6	2.3	0.13	25.0
		Group B:	with pr	eserved reservoir a	nd reduced em	ptying	of the gallbladder			
10	24.9	7.0	0.28	26.8	13.9	0.52	25.8	5.2	0.20	37
11	14.3	3.0	0.22	31.3	16.0	0.51	14.6	4.5	0.31	28
12	18.1	3.8	0.21	31.5	7.2	0.23	15.9	3.5	0.22	49
13	19.4	7.8	0.40	30.1	11.1	0.37	12.4	5.3	0.43	48
14	19.6	5.9	0.30	13.4	5.0	0.37	10.6	3.8	0.36	76
15	27.8	4.7	0.17	28.5	7.4	0.26	32.0	5.1	0.16	69
$\bar{X}(n=6)$	20.7	5.4	0.26	26.9	10.1	0.38	18.6	4.6	0.28	51.1
SD	4.9	1.9	0.08	6.9	4.3	0.12	8.5	0.8	0.10	18.2
				Groups A	and B $(n = 15)$					
\bar{X}	20.5	61	0 31	20.0	9.6	0 48	14.3	4 2	0 34	48 1
SD	7.7	2.6	0.11	9.4	3.8	0.25	8.0	1.8	0.12	21.1
			Group	C: without reserve	oir function of t	the gall	bladder			
16	12.8	5 5	0 / 3	0			30.6	12.7	0 32	
10	23.0	5.5	0.75	247	15.3	0.62	31.2	53	0.52	35
17	13.3	0.J 5.6	0.27	24.7	15.5	0.02	21.0	5.5	0.17	35
10	80	3.5	0.42	11.2	86	0.77	17.0	9.0	0.40	103
20	0.9 20 A	20.6	0.33	0	0.0	0.77	13.7	0.0	0.32	105
20	10.9	3.8	0.70	90	9.5	0 97	27 4	9.6	0.35	101
21	10.9	5.8	0.55	9.0	9.5	0.97	27.4	9.0 11.9	0.57	101
22	26.1	0.0 7.0	0.00	15.1	8.2	0.55	16.9	8.6	0.55	105
$\vec{\mathbf{v}}$ (0)	16.0	7 4	0.44	76	10.4	0.72	22.5	0.0	0.40	96.0
X(n=8)	10.9	/.4	0.44	/.0	10.4	0.73	23.5	8.9	0.40	80.U 34.0
SD	8.2	5.5	0.16	9.2	3.3	0.19	8./	2.8	0.13	34.0
				Groups	A, B, and C					
\bar{X}	19.2	6.5	0.35	15.3	9.8	0.54	17.5	5.8	0.36	57.6
SD	7.8	3.8	0.14	11.0	3.6	0.26	9.2	3.1	0.13	29.1
Control su	bjects									
1	37.6	5.6	0.15	44.5 [‡]	ND	ND	4.7	1.4	0.24	ND
2	52.5	4.7	0.09	34.6	8.0	0.23	0.0	0.0	0.37	0
3	26.5	14.6	0.55	20.7	24.2	1.17	24.9	8.5	0.34	35
4	26.8	6.4	0.24	40.1	17.6	0.44	26.9	9.5	0.35	54
5	29.2	5.5	0.19	16.4	4.6	0.28	13.9	3.0	0.22	65

Table III.	Kinetics of	Bile Acids in	Control Subjects and	d Gallstone Patients
			·	

	c	CDCA			CA			DCA		
Subject No.	Pool	Synthesis	FTR	Pool	Synthesis	FTR	Pool	Input	FTR	Transfer of Ca to DCA*
	(µmol kg ⁻¹ body wt)	(µmol kg ⁻¹ d ⁻¹)	(d⁻¹)	(µmol kg ^{−1} body wt)	(µmol kg ⁻¹ d ⁻¹)	(d ^{−1})	(µmol kg ⁻¹ body wt)	(µmol kg⁻¹ d⁻¹)	(d ^{−1})	(%)
Control sul	bjects (continued)									
6	20.9	3.3	0.16	15.2	5.4	0.36	9.2	2.1	0.23	39
7	20.9	7.7	0.37	38.4	13.8	0.36	14.7	6.3	0.43	46
8	35.6	5.0	0.14	38.7	7.0	0.18	19.3	4.4	0.23	63
9	26.6	5.3	0.20	44.7	10.7	0.24	20.8	4.2	0.20	39
10	16.0	4.8	0.30	0			28.5	8.0	0.28	
11	20.1	6.2	0.31	25.4	8.1	0.32	14.7	3.7	0.25	46
12	23.7	8.0	0.34	22.2	8.7	0.39	31.5	6.3	0.20	72
13	16.8	3.0	0.18	20.3	5.9	0.09	10.8	1.1	0.10	19
14	39.0	9.0	0.23	33.3	11.3	0.34	18.8	4.1	0.22	36
15	13.2	2.6	0.20	18.0	6.7	0.37	4.0	0.7	0.17	10
X(n = 15)	27.0	6.1	0.24	27.5	10.2	0.37	16.2	4.2	0.26	40.3
SD	10.5	3.0	0.12	12.9	5.6	0.26	9.4	3.0	0.09	21.3

* Transfer of CA to the DCA pool = (DCA input/CA synthesis) \times 100.

* This CA pool was calculated (24) from the kinetically determined CDCA pool size times the CA/CDCA molar ratio of intraoperative gallbladder bile.

transfer of CA to the DCA pool. This could be directly demonstrated by measurement of the ¹³C label in the DCA pool (Fig. 3): after intake of [¹³C]CA, 35–80% of the ¹³C label peaked in the DCA pool within 24 h. By contrast, controls (except subject 10) and gallstone patients of groups A and B (except subject 9 in group A) transferred less than half of that amount of ¹³C label and reached peak enrichment of the DCA pool with ¹³C 3–4 d after intake.



Figure 2. Patterns ($\bar{X}\pm$ SEM) of bile acid pools and CA to DCA transfer in healthy controls (n = 15) and three groups of gallstone patients classified according to fraction of emptying of the gallbladder: A (n = 9) with emptying gallbladder; B (n = 6) with reduced emptying (< 50%) of the gallbladder; C (n = 8) without gallbladder reservoir. Symbols and abbreviations: (**u**) bile acid (BA); (**u**) CA pool; (**u**) Percent transfer (T%) of CA to the DCA pool (=DCA input/CA synthesis ×100). *Significantly different from controls; **significantly different from all other groups.

Linear correlations with bile acid kinetics. The hepatic synthesis rates of CA and CDCA were directly correlated (r = 0.608, P < 0.001). Input of DCA increased with the synthesis rate of its precursor CA (r = 0.511, P < 0.01) and with the fractional transfer of CA to the DCA pool (r = 0.664, P < 0.001). The size of the CDCA pool was mainly related to CDCA synthesis rate (r = 0.559, P < 0.005) and gallbladder fasting volume (r = 0.489, P < 0.01). The size of the CA pool



Figure 3. Time course of the amount of excess ¹³C-labeled DCA $(\bar{X}\pm SEM)$ in the DCA pool after oral intake of 110 μ mol [¹³C]CA in controls (\odot) (n = 14), and three groups of gallstone patients: (**n**) group A (n = 7); (\bullet) group B (n = 6); (\blacktriangle) group C (n = 8) (compare Fig. 2). Note enhanced formation and increased input of [¹³C]DCA in group C of gallstone patients. *Significantly different (C vs. other groups). ¹³C transfer was not determined in patients 1 and 2 in group A and subject 1 in controls.

Table IV. Lipid Composition of CCK-stimulated Duodenal Bile

Group	No.	Total lipids	Cholesterol saturation [‡]
		(g/dl)	
Gallstone patients			
Group A	8	4.76 ± 2.41	1.27±0.07 [§]
Group B	6	5.23±3.03 ^{II}	1.40±0.23 [§]
Group C	7	2.78±1.24 [§]	1.53±0.35 [§]
Controls	15	5.26±1.73	0.89±0.17

* In three subjects (10–12 in group B) intraoperative gallbladder bile was analyzed instead of CCK bile.

[‡] Saturation index according to Carey's Critical Tables (29).

[§] Significantly different from controls (P < 0.02).

^{II} Significantly different (P < 0.05, group C vs. B).

correlated poorly with CA synthesis rate (r = +0.34; P = 0.07) and inversely with the fractional transfer of CA to the DCA pool (r = -0.38; P = 0.03). The size of the DCA pool correlated (r = 0.80; P < 0.001) with the input rate of DCA. Neither pool size nor input rate of DCA were related to age. The fraction of DCA in the total bile acid pool was correlated with the fraction of DCA (r = 0.71; P < 0.001) and the cholesterol saturation index (r = 0.42; P = 0.01) of CCK-stimulated bile, and with the total volume of the gallstones (r = 0.53; P < 0.01).

Composition of biliary lipids (Table IV). CCK-stimulated duodenal bile was supersaturated (29) in all gallstone patients (P < 0.01 vs. controls) and in 4 of 15 controls (subjects 4, 10, 12, and 15), who either had a very small bile acid pool (subject 15) or a large DCA pool (subjects 4, 10, and 12). Gallstone patients showed an increased cholesterol/phospholipid ratio (+41%, +58%, +54% in groups A, B, and C; P < 0.01 vs. controls), but only group C showed increased ratios of phospholipids/bile acids (+41%; P < 0.02). The degree of cholesterol saturation was directly correlated with the fraction of DCA in biliary bile acids (r = 0.552; P = 0.001) (Fig. 4).

Discussion

This study substantiates the concept that the reduction of bile acid pool size in patients with cholesterol gallstones is caused by an increased turnover of the bile acid pools, not by a reduction of bile acid synthesis. The major new finding is a disorder of CA-DCA metabolism observed in a subgroup of gallstone patients, whose disease had progressed to loss of the reservoir function of the gallbladder.

Reduction of the bile acid pool is a common finding in non-obese cholesterol gallstone patients (1, 2, 4-6), which has been explained by reduced synthesis of bile acids (12) or enhanced enterohepatic cycling of the bile acid pool (3, 5, 9). Only in a group of Italian gallstone patients the reduction of the CA pool was caused by a combination of reduced synthesis and enhanced turnover (32). In our study and in studies of American whites (1, 3, 4) and American Indians (33) gallstone patients had normal hepatic synthesis of bile acids. Also Swedish gallstone patients had bile acid synthesis not significantly decreased (5). Contrary to the hypothesis on feedback inhibition



Figure 4. Cholesterol saturation (29) is correlated (r = 0.552; P = 0.001) with the percentage of DCA in bile acids of CCK-stimulated duodenal bile from healthy controls (\circ) and from gallstone patients with normal emptying (\blacksquare) (group A, n = 8) or reduced emptying of the gallbladder (\blacklozenge) (group B, n = 6) and gallstone patients without reservoir function of the gallbladder (\blacklozenge) (group C, n = 7).

of bile acid synthesis by DCA (13, 14), even the patients of group C with twofold increased input of DCA had normal bile acid synthesis. This strongly suggests that hepatic synthesis of bile acids is not deranged in gallstone disease. The reduction of the bile acid pool in gallstone disease is explained by increased fractional turnover of the bile acid pools (1, 2, 4-6, 33) (Table III, groups A and C).

The increased turnover of the bile acid pools in gallstone patients, as observed in groups A and C (Table III), is thought to result from increased enterohepatic cycling of bile acids (3, 9, 12). This could be caused by enhanced emptying of the gallbladder, or by hastened transit (34) or decreased fractional absorption of bile acids in the small intestine. Emptying of the gallbladder is normal or reduced, but not enhanced, in gallstone patients (35, 36); reduced emptying of the gallbladder is associated with slower fractional turnover and larger pool size of bile acids in healthy subjects (34) as well as in gallstone patients (37) (Fig. 2, group B). Hastening the small intestinal transit enhances the turnover and reduces the size of the bile acid pool in healthy subjects to the extent seen in gallstone patients (34). We did not determine small intestinal transit time. However, mouth to cecum transit time had not been shortened in similar gallstone patients with functioning gallbladder (38). Alternatively, the increase (+54%) in fractional turnover and the reduction (-44%) of the CA pool in the gallstone patients of group A could be caused by a reduction in fractional absorption of CA by only 5% (from 95.4% [3] to 90.5%), provided the pool cycles at a normal rate of seven times per day (9). Such a small decrease in absorption of CA and CDCA cannot be excluded by the available studies (3, 12). Thus, the cause for the enhanced bile acid turnover is still unknown in gallstone disease.

Expansion of the DCA pool was related to increased input of DCA (r = 0.80; P < 0.001). DCA was conserved in the enterohepatic circulation as efficiently as CDCA judged from similar fractional turnover rates. In this study, high input of DCA was not caused by old age or constipation, two factors reported to increase input of DCA (17, 39). Highly significant direct associations suggested that the input rate of DCA depended on the synthesis rate of its precursor CA and even more on the fractional transfer of CA to the DCA pool. This fractional transfer, which is usually less than 40% in healthy subjects (17), ranged up to 100%.

Simultaneous turnover studies of CA and DCA revealed a disorder of CA-DCA metabolism in a subgroup of gallstone patients. This disorder can be described as an increased DCA input (> 7 μ mol kg⁻¹d⁻¹) or transfer (> 75%) of CA to DCA. It is associated with a reduced pool of CA and an expanded DCA pool (DCA/CA pool ratio > 1.5) (Fig. 2, group C). Between 35% and 80% of the ¹³C label given as [¹³C]CA by mouth appeared within 24 h as [¹³C]DCA in the bile acid pool, whereas this transfer was < 25% in controls (Fig. 3). The accelerated and increased transfer of ¹³C label implies faster degradation of CA to DCA as well as increased absorption of that DCA.

Which mechanism is responsible for the enhanced transfer of CA to DCA? In theory, two factors should be considered: accelerated loss of CA into the colon, where it is almost completely (> 95%) degraded to DCA (15), and increased bacterial conversion of CA to DCA in the small intestine. Both could be operative in patients without gallbladder reservoir (group C), because loss of the gallbladder reservoir (a) enhances daily enterohepatic cycling (40) and loss of CA into the colon and (b) prolongs the time of daily exposure of CA to bacteria in the small intestine (41). Cholecystectomy, however, led only to a moderate increase (from 46% to 66%) of the fraction of CA transferred to the DCA pool (21). On the other hand, the disorder occurred also in two of the women with normal gallbladder reservoir and emptying (subject 9 in group A and subject 10 in controls). Therefore, we speculate that other mechanisms in addition to loss of the gallbladder reservoir are involved. If ileal absorption of CA were impaired, rapid loss of CA into the colon would reduce the pool of CA and accelerate the conversion of CA to DCA and the input of that DCA. An alternative mechanism would be colonization of the distal small bowel with 7α -dehydroxylating anaerobic bacteria. CA could then be more rapidly converted to DCA and that DCA could be well absorbed via the ileal bile acid transport system (42).

In patients with type III pattern of the bile acid pools the molar ratios of DCA/CA in fasting bile exceeded 1.0 (data not shown). Such high ratios have been reported for gallstone patients with nonvisualizing gallbladder in one (5) but not another study (43), for cholecystectomized gallstone patients in some (41, 44) but not other studies (21, 45), and for some conditions with a high risk of gallstone formation such as partial resection of the ileum (46) or hyperlipidemia type IIb or IV (47).

This study showed reasonable positive correlations between the fraction of DCA in the total bile acid pool and the DCA fraction (r = 0.65; P < 0.005) and cholesterol saturation index (r = 0.44; P < 0.05) of CCK-stimulated duodenal bile. The degree of cholesterol saturation of bile was also correlated with the fraction of DCA in biliary bile acids (Fig. 4). These associations support the concept that an expansion of the DCA fraction in the bile acid pool contributes to supersaturation of bile in gallstone disease.

Interestingly, the study revealed that bile was supersaturated in gallstone disease also in presence of a normal sized bile acid pool in subgroup B. In this group of patients, the reduced emptying of the gallbladder may have decreased the cycling of the pools and the secretion rate of bile acids. Low hepatic output of bile acids could increase cholesterol saturation by raising the secretory ratio of cholesterol to phospholipids (48).

Generally, loss of the gallbladder reservoir enhances enterohepatic cycling of bile acids (40) and lowers cholesterol saturation of bile (4, 21, 43). Nevertheless, the patients in group C had supersaturated bile in spite of loss of the gallbladder reservoir, presumably because other factors had raised cholesterol content of bile. We speculate that supersaturation of bile in group C may have been caused by DCA-induced hypersecretion of cholesterol (10), in that these patients had a high fraction of DCA which exceeded the CA fraction in bile.

In summary, two different disturbances of bile acid metabolism, which are likely to contribute to supersaturation of bile with cholesterol, were observed in cholesterol disease: (a) reduction of the bile acid pool and (b) enhanced conversion of CA to DCA with replacement of the CA pool by an expanded DCA pool. The first disorder may be caused by more rapid loss of primary bile acids from the small intestine into the colon, the second disturbance may result from enhanced 7α -dehydroxylation of CA (possibly in the ileum) and increased absorption of newly formed DCA.

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