

Supporting Information for:

**Endoperoxide Synthesis by Photocatalytic Aerobic [2+2+2] Cycloadditions**

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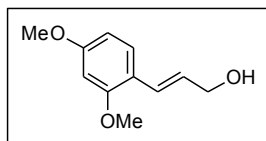
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**I. General information**

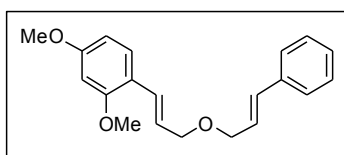
THF and CH<sub>2</sub>Cl<sub>2</sub> were purified by elution through alumina as described by Grubbs.<sup>1</sup> MeNO<sub>2</sub> was purified by distillation from MgSO<sub>4</sub> prior to use. A 200 W (3980 lumens) GE Crystal Clear light bulb was used for all photochemical reactions depicted in Tables 1 and 2. Flash column chromatography was performed with Silicycle 40-63 Å silica (230-400 mesh). 9,10-Dicyanoanthracene (DCA), 5,10,15,20-tetraphenyl-21H,23H-porphine (TPP) and 2,4,6-triphenylpyrylium tetrafluoroborate (TPT) were purchased from Aldrich and used without further purification. Ru(bpz)<sub>3</sub>(PF<sub>6</sub>)<sub>2</sub><sup>2</sup> and Ru(bpy)<sub>3</sub>(PF<sub>6</sub>)<sub>2</sub><sup>3</sup> were prepared as previously described. Diastereomer ratios for all compounds were determined by <sup>1</sup>H NMR analysis of the unpurified reaction mixture. <sup>1</sup>H and <sup>13</sup>C NMR data for all previously uncharacterized compounds were obtained using Varian Inova-500 and Varian Unity-500 spectrometers and are referenced to TMS (0.0 ppm), CDCl<sub>3</sub> (77.0 ppm), and CD<sub>3</sub>CN (118.6 ppm), respectively. IR spectral data were obtained using a Bruker Vector 22 spectrometer (thin film on NaCl). Melting points were obtained using a Mel-Temp II (Laboratory Devices, Inc., USA) melting point apparatus. Mass spectrometry was performed with a Waters (Micromass) AutoSpec®. These facilities are funded by the NSF (CHE-9974839, CHE-9304546) and the University of Wisconsin.

**II. Synthesis of cyclization substrates**

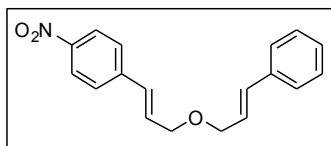
The substrates for entries 1–4, 7, 8, 10, 11, 13, and 15 were prepared as previously described.<sup>3</sup> The substrate for entry 6 was prepared as described by Nair.<sup>4</sup>



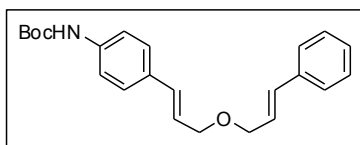
***o,p*-Dimethoxycinnamyl alcohol.** A solution of ethyl 2,4-dimethoxycinnamate (4.39 g, 18.5 mmol) in 37 mL CH<sub>2</sub>Cl<sub>2</sub> was placed in a 250 mL round-bottomed flask. The solution was cooled to -78 °C, and a 1.0 M solution of DIBAL-H in hexanes (46.3 mL, 46.3 mmol) was added over 5 minutes. The reaction was then stirred for 2 h at -78 °C. After warming to 0 °C, the reaction was quenched with a saturated aqueous solution of Rochelle's salt. The mixture was stirred and allowed to warm to room temperature overnight. The layers were separated, and the aqueous layer was extracted with three portions of Et<sub>2</sub>O. The combined organic fractions were dried over MgSO<sub>4</sub> and concentrated by rotary evaporation. The residue was purified by chromatography on SiO<sub>2</sub> (8:1 to 2:1 hexanes:EtOAc) to give 3.39 g (17.4 mmol, 94% yield) of a yellow oil. IR (neat) 3477, 2939, 1110 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ 7.35 (d, J = 8.4 Hz, 1H), 6.83 (d, J = 15.9 Hz, 1H), 6.47 (dd, J = 8.4, 2.5 Hz, 1H), 6.44 (d, J = 2.5 Hz, 1H), 6.28 (dt, J = 15.9 Hz, 6.6, 1H), 4.29 (m, 2H), 3.83 (s, 3H), 3.81 (s, 3H), 1.46 (m, 1H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>) δ 160.5, 157.9, 127.8, 127.0, 126.2, 118.7, 104.8, 98.4, 64.4, 55.4, 55.3. HRMS (EI) calculated for [C<sub>11</sub>H<sub>14</sub>O<sub>3</sub>]<sup>+</sup> requires m/z 194.0938, found m/z 194.0939.



***(E)*-*o,p*-Dimethoxycinnamyl *(E)*-cinnamyl ether** (Table 2, Entry 5). A 50 mL round-bottomed flask was charged with 60% NaH (575 mg, 14.4 mmol) and 5 mL THF. A solution of 2,4-dimethoxycinnamyl alcohol (2.00 g, 10.3 mmol) in 7.5 mL THF was added slowly. The reaction was stirred at room temperature for 30 min and then cooled to 0 °C. A solution of cinnamyl bromide (2.42 g, 12.3 mmol) in 8 mL of THF was added, and the reaction was warmed to room temperature. After 24 h, the reaction was quenched with water and the aqueous layer was extracted with two portions of Et<sub>2</sub>O. The combined organic fractions were dried over MgSO<sub>4</sub> and concentrated via rotary evaporation. Purification by chromatography on SiO<sub>2</sub> (10:1 to 4:1 hexanes:EtOAc) afforded 2.91 g (9.40 mmol, 91% yield) of a yellow oil. IR (neat) 3003, 2938, 1209 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ 7.39–7.20 (m, 6H), 6.86 (d, J = 16.0 Hz, 1H), 6.62 (d, J = 16.0 Hz, 1H), 6.46 (dd, J = 8.6, 2.5 Hz, 1H), 6.42 (d, J = 2.5 Hz, 1H), 6.32 (dt, J = 16.0, 6.2 Hz, 1H), 6.23 (dt, J = 16.0, 6.2 Hz, 1H), 4.18 (d, J = 6.2 Hz, 4H), 3.8 (s, 3H), 3.78 (s, 3H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>) δ 160.4, 157.8, 136.7, 132.3, 128.4, 127.7, 127.6, 127.5, 126.4, 126.2, 124.2, 118.7, 104.7, 98.3, 71.5, 70.4, 55.3, 55.2. HRMS (EI) calculated for [C<sub>20</sub>H<sub>22</sub>O<sub>3</sub>]<sup>+</sup> requires m/z 310.1564, found m/z 310.1549.

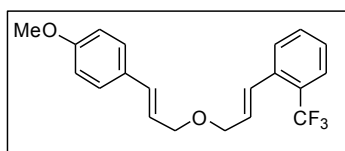


***(E)*-*p*-Nitrocinnamyl *(E)*-cinnamyl ether.** A solution of 4-nitrocinnamyl alcohol (1.20 g, 6.70 mmol) and cinnamyl bromide (1.98 g, 10.0 mmol) in 27 mL DMF was placed in a 100 mL round-bottomed flask. The solution was cooled to 0 °C and 60% NaH (402 mg, 10.0 mmol) was added slowly. The mixture was allowed to warm to room temperature and stirred for 7 h. The reaction was quenched by slowly adding a saturated aqueous solution of NH<sub>4</sub>Cl and the aqueous layer was extracted with two portions of Et<sub>2</sub>O. The combined organic fractions were dried over MgSO<sub>4</sub> and concentrated via rotary evaporation. Purification by chromatography on SiO<sub>2</sub> (20:1 to 4:1 hexanes:EtOAc) gave 885 mg (3.00 mmol, 45% yield) of a red oil. IR (neat) 3080, 1343, 1110 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ 8.11 (d, J = 9.3 Hz, 2H), 7.44 (d, J = 9.3 Hz, 2H), 7.40–7.20 (m, 5H), 6.68 (d, J = 16.0 Hz, 1H), 6.64 (d, J = 16.0 Hz, 1H), 6.45 (dt, J = 16.0, 5.5 Hz, 1H), 6.31 (dt, J = 16.0, 5.5 Hz, 1H), 4.22–4.19 (m, 4H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>) δ 146.6, 143.0, 136.4, 132.5, 131.1, 129.2, 128.4, 127.6, 126.7, 126.3, 125.5, 123.7, 71.0, 69.8. HRMS (EI) calculated for [C<sub>18</sub>H<sub>17</sub>NO<sub>3</sub>]<sup>+</sup> requires m/z 195.1208, found m/z 195.1215.



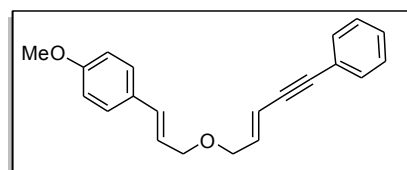
***(E)*-*p*-*N*-Boc-Aminocinnamyl *(E)*-cinnamyl ether** (Table 2, Entry 9). A solution of Cu(acac)<sub>2</sub> (213 mg, 0.81 mmol) in 10 mL EtOH was placed in a 100 mL round-bottomed flask. NaBH<sub>4</sub> (300 mg, 7.93 mmol) was added, and the mixture was stirred for 10 min. The flask was then placed

in a water bath and vigorously stirred while a suspension of (*E*)-*p*-nitrocinnamyl (*E*)-cinnamyl ether (1.20 g, 4.06 mmol) in 15 mL of EtOH was added dropwise. After 1 h, additional NaBH<sub>4</sub> (161 mg, 4.25 mmol) was added, and the mixture was stirred for 12 h before being quenched with water. The aqueous phase was extracted with two portions of EtOAc, and the combined organic fractions were dried over MgSO<sub>4</sub> and concentrated via rotary evaporation. Purification by flash column chromatography (20:1 to 1:1 hexanes:EtOAc) afforded the free amine. This material was dissolved in 20 mL 1,4-dioxane in a 100 mL round-bottomed flask. Triethylamine (0.54 mL, 3.90 mmol) and Boc<sub>2</sub>O (851 mg, 3.90 mmol) were added, and the reaction was stirred at room temperature for 18 h. The solution was concentrated via rotary evaporation, and the residue was eluted through a short plug of SiO<sub>2</sub> with Et<sub>2</sub>O. The eluent was concentrated by rotary evaporation, and the residue was purified by chromatography on SiO<sub>2</sub> (20:1 to 4:1 hexanes:EtOAc) to afford 325 mg (0.912 mmol, 22% yield) of a yellow solid (mp = 82–84 °C). IR (neat) 3329, 1727, 1522 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ 7.40–7.22 (m, 9H), 6.64 (d, J = 15.9 Hz, 1H), 6.57 (d, J = 15.9 Hz, 1H), 6.51 (s, 1H), 6.32 (dt, J = 15.9, 5.9 Hz, 1H), 6.23 (dt, J = 15.9, 5.9 Hz, 1H), 4.21–4.18 (m, 4H), 1.52 (s, 9H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>) δ 152.6, 137.8, 136.7, 132.5, 132.2, 131.6, 128.5, 127.6, 127.1, 126.5, 126.0, 124.5, 118.4, 80.6, 70.8, 70.7, 28.3. HRMS (EI) calculated for [C<sub>23</sub>H<sub>27</sub>O<sub>3</sub>N+Na]<sup>+</sup> requires m/z 388.1884, found m/z 388.1880.



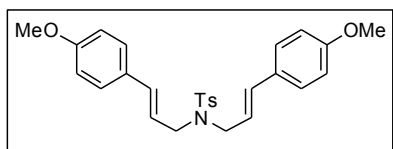
**(*E*)-*p*-Methoxycinnamyl (*E*)-*o*-trifluoromethylcinnamyl ether** (Table 2, Entry 12). A solution of 2-trifluoromethyl cinnamyl alcohol<sup>5</sup> (1.69 g, 8.36 mmol) in 20 mL THF was placed in a 250 mL round-bottomed flask. 60% NaH (402 mg, 10.1 mmol) was added, and the reaction was stirred for 1 h. The flask was then cooled to 0 °C, and a solution of *p*-methoxycinnamyl

bromide (2.85 g, 12.5 mmol) in 15 mL of THF was added slowly. The solution was allowed to warm to room temperature and stirred for 48 h. The reaction was quenched with water and the aqueous layer was extracted with two portions of Et<sub>2</sub>O. The combined organic fractions were washed with brine, dried over MgSO<sub>4</sub>, and concentrated via rotary evaporation. Purification by chromatography on SiO<sub>2</sub> (20:1 to 10:1 hexanes:EtOAc) afforded 1.45 g (4.2 mmol, 50% yield) of a colorless oil. IR (neat) 3004, 1514, 1315 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ 7.62 (m, 2H), 7.47 (t, J = 7.5 Hz, 1H), 7.34–7.20 (m, 3H), 7.02 (d, J = 15.7 Hz, 1H), 6.85 (d, J = 8.6 Hz, 2H), 6.58 (d, J = 15.7 Hz, 1H), 6.28 (dt, J = 15.7, 5.9 Hz, 1H), 6.18 (dt, J = 15.7, 5.9 Hz, 1H), 4.22 (d, J = 5.9 Hz, 2H), 4.18 (d, J = 5.9 Hz, 2H), 3.78 (s, 3H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>) δ 159.3, 135.9, 132.5, 131.8, 130.6, 129.4, 128.2, 127.7, 127.5, 127.3, 127.0, 125.7 (q, J = 5.8 Hz), 124.3 (q, J = 273.9 Hz), 123.5, 114.0, 70.9, 70.2, 55.2. HRMS (EI) calculated for [C<sub>20</sub>H<sub>19</sub>O<sub>2</sub>F<sub>3</sub>]<sup>+</sup> requires m/z 348.1332, found m/z 348.1346.

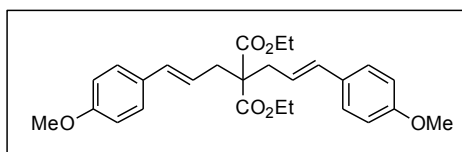


**(*E*)-*p*-Methoxycinnamyl (*E*)-5-phenylpent-2-en-4-yn-1-yl ether** (Table 2, Entry 14). A suspension of 60% NaH (290 mg, 7.20 mmol) in 20 mL THF was placed in a 100 mL round-bottomed flask. The flask was cooled to 0 °C, and (*E*)-5-phenylpent-2-en-4-yn-1-ol<sup>6</sup> (1.14 g, 7.20 mmol) was added. The mixture was stirred for 30 min

before adding *p*-methoxycinnamyl bromide (1.63 g, 7.20 mmol). The reaction was allowed to warm to room temperature. After stirring for 12 h, the reaction was cooled to 0 °C and quenched by adding a saturated aqueous solution of NH<sub>4</sub>Cl. The aqueous layer was extracted with two portions of EtOAc. The combined organic fractions were washed with brine, dried over MgSO<sub>4</sub>, and concentrated via rotary evaporation. Purification by chromatography on SiO<sub>2</sub> (20:1 to 8:1 hexanes:EtOAc) afforded 1.65 g (5.42 mmol, 75% yield) of the title product as a yellow solid (mp = 49–51 °C). IR (neat) 2935, 2198, 1513 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ 7.45–7.30 (m, 7H), 6.86 (d, J = 9.0 Hz, 2H), 6.57 (d, J = 16.1 Hz, 1H), 6.3 (dt, J = 16.01, 5.6 Hz, 1H), 6.16 (dt, J = 16.0, 6.0 Hz, 1H), 5.98 (dt, J = 16.0, 1.7 Hz, 1H), 4.16 (dd, J = 6.0, 1.3 Hz, 2H), 4.13 (dd, J = 5.6, 1.7 Hz, 2H), 3.81 (s, 3H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>) δ 159.3, 139.5, 132.4, 131.5, 129.4, 128.3, 128.2, 127.7, 123.4, 123.2, 114.0, 111.7, 90.0, 87.4, 71.1, 69.7, 55.3. HRMS (EI) calculated for [C<sub>21</sub>H<sub>20</sub>O<sub>2</sub>]<sup>+</sup> requires m/z 304.1458, found m/z 304.1469.

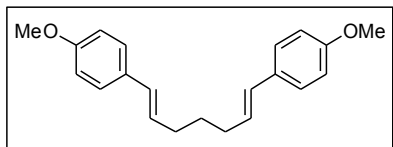


**(*E,E*)-*N,N*-Bis-(*p*-methoxycinnamyl) toluenesulfonamide** (Table 2, Entry 16). A suspension of 60% NaH (568 mg, 14.2 mmol) in 30 mL DMF was placed in a 250 mL round-bottomed flask. The mixture was cooled to 0 °C and a solution of *N-p*-methoxycinnamyl *p*-toluenesulfonamide<sup>7</sup> (1.5 g, 4.73 mmol) in 30 mL of DMF was added dropwise over 10 min. The reaction was warmed to room temperature and stirred for 30 min before a solution of *p*-methoxycinnamyl bromide (2.69 g, 11.8 mmol) in 35 mL of DMF was added. After 24 h, the reaction was carefully quenched with a saturated aqueous solution of NH<sub>4</sub>Cl and the aqueous layer was extracted with two portions of Et<sub>2</sub>O. The combined organic fractions were dried over MgSO<sub>4</sub> and concentrated via rotary evaporation. Purification by flash column chromatography on SiO<sub>2</sub> (20:1 to 2:1 hexanes:EtOAc) afforded 1.29 g (2.8 mmol, 59% yield) of the title product as a yellow solid (mp = 92–94 °C). IR (neat) 3033, 1512, 1251 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ 7.75 (d, J = 7.8 Hz, 2H), 7.30 (d, J = 7.8 Hz, 2H), 7.18 (d, J = 8.9 Hz, 4H), 6.82 (d, J = 8.9 Hz, 4H), 6.36 (d, J = 16.0 Hz, 2H), 5.82 (dt, J = 16.0, 6.7 Hz, 2H), 3.98 (d, J = 6.7 Hz, 4H), 3.80 (s, 6H), 2.43 (s, 3H). <sup>13</sup>C NMR: 159.4, 143.2, 137.6, 133.5, 129.7, 129.0, 127.6, 127.3, 121.6, 114.0, 55.3, 49.0, 21.5. HRMS (EI) calculated for [C<sub>27</sub>H<sub>29</sub>NO<sub>4</sub>S+Na]<sup>+</sup> requires m/z 486.1710, found m/z 486.1700.



**(*E,E*)-Diethyl 2,2-bis(*p*-methoxycinnamyl)malonate** (Table 2, Entry 17). Diethyl malonate (1.00 g, 6.24 mmol) was added to a suspension of 60% NaH (929 mg, 23.2 mmol) in 16 mL THF. The mixture was stirred for 1 h then cooled to 0 °C. A solution of *p*-methoxycinnamyl bromide (3.54 g, 15.6 mmol) in 15 mL of

DMF was added. After 12 h, the reaction was quenched with water and the aqueous layer was extracted with two portions of Et<sub>2</sub>O. The combined organic fractions were dried over MgSO<sub>4</sub> and concentrated via rotary evaporation. Purification by chromatography on SiO<sub>2</sub> (15:1 to 10:1 hexanes:EtOAc) afforded 2.45 g (5.41 mmol, 87% yield) of a white solid (mp = 74–75 °C). IR (neat) 1729, 1512, 1249 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ 7.32 (d, J = 9.1 Hz, 4H), 6.83 (d, J = 9.1 Hz, 4H), 6.39 (d, J = 15.7 Hz, 2H), 5.93 (dt, J = 15.9, 7.6 Hz, 2H), 4.20 (q, J = 7.2 Hz, 4H), 3.80 (s, 6H), 2.81 (dd, J = 7.6, 1 Hz, 4H), 1.24 (t, J = 7.2 Hz, 6H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>) δ 170.9, 159.1, 133.4, 130.0, 127.3, 121.7, 113.9, 61.3, 58.1, 55.3, 36.5, 14.2. HRMS (EI) calculated for [C<sub>27</sub>H<sub>32</sub>O<sub>6</sub>+H]<sup>+</sup> requires m/z 453.2272, found m/z 453.2276.



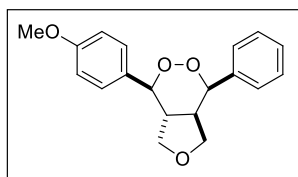
**(*E,E*)-1,7-bis(4-methoxyphenyl)hepta-1,6-diene** (Table 2, Entry 18). An aqueous solution of glutaraldehyde (50% w/w, 1.00 g, 4.99 mmol) and a solution of 4-methoxybenzyl triphenylphosphonium bromide (5.80 g, 12.5 mmol) in 20 mL THF were placed in a 100 mL round-bottomed flask and cooled to 0 °C. KO<sup>*t*</sup>-Bu (1.68 g, 15.0 mmol) was

added slowly over 5 min. The resulting orange suspension was stirred at room temperature for 18 h before being diluted with water and extracted with three portions of Et<sub>2</sub>O. The combined organic fractions were dried over MgSO<sub>4</sub> and concentrated via rotary evaporation. Purification by chromatography on SiO<sub>2</sub> (20:1 to 10:1 hexanes:EtOAc) afforded a mixture of isomeric bis(styrene)s. This material was dissolved in 50 mL of benzene in 100 mL round-bottomed flask, to which was added thiophenol (0.022 mL, 0.211 mmol) and AIBN (34.7 mg, 0.211 mmol). The solution was heated at reflux for 24 h. Upon cooling to room temperature, the solution was diluted with hexanes and washed with two portions of saturated aqueous NaHCO<sub>3</sub> and one portion of brine. The organic layer was then dried over MgSO<sub>4</sub> and concentrated by rotary evaporation. Purification by chromatography on SiO<sub>2</sub> (gradient, 25:1 to 20:1 hexanes/EtOAc) gave 207 mg (0.670 mmol, 13% yield) of the title product as a white solid (mp = 91–93 °C). IR (neat) 3004, 2916, 1250 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz, CDCl<sub>3</sub>) δ 7.24 (d, J = 8.9 Hz, 4H), 6.80 (d, J = 8.9 Hz, 4H), 6.32 (d, J = 15.9 Hz, 2H), 6.06 (dt, J = 15.9, 7 Hz, 2H), 3.72 (s, 6H), 2.21 (q, J = 7 Hz, 4H), 1.60 (p, J = 7.3 Hz,

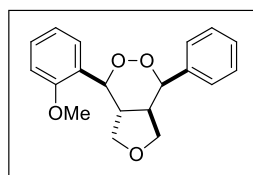
2H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  158.6, 130.7, 129.2, 128.8, 127.0, 113.9, 55.2, 32.9, 29.0. HRMS (EI) calculated for  $[\text{C}_{21}\text{H}_{24}\text{O}_2]^+$  requires  $m/z$  308.1672, found  $m/z$  308.1668.

### III. Photocycloadditions

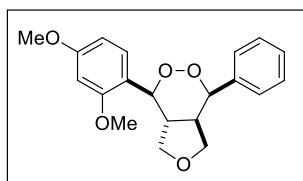
**General Procedure:** An oven-dried 135 mL glass pressure vessel containing a magnetic stirbar was charged with bis(alkene) substrate,  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$  (0.005 or 0.02 equiv), and  $\text{MeNO}_2$ . The vessel was fitted with a regulator and pressurized to 60 psi with  $\text{O}_2$  gas. The reaction was cooled in a  $5^\circ\text{C}$  water bath and irradiated with a 200 W incandescent bulb. After consumption of the substrate, the reaction mixture was eluted through a short pad of  $\text{SiO}_2$  using  $\text{EtOAc}$  and  $\text{CH}_2\text{Cl}_2$ . The solvent was removed by rotary evaporation, and purification of the residue by chromatography on  $\text{SiO}_2$  (8:1 to 2:1 hexanes: $\text{EtOAc}$ ) afforded the [2+2+2] cycloadduct.



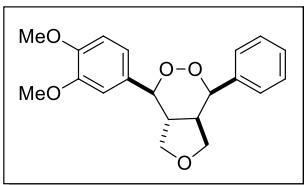
(Table 2, entry 1). Prepared according to the General Procedure using 100 mg (0.357 mmol) of bis(alkene), 1.5 mg (1.8  $\mu\text{mol}$ ) of  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$ , 17.8 mL of  $\text{MeNO}_2$  and an irradiation time of 30 min. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 100.6 mg (0.322 mmol, 90% yield, dr = 6:1), Experiment 2: 104.0 mg (0.333 mmol, 94% yield, dr = 6:1). IR (neat) 1253, 732, 702  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR: (500 MHz)  $\delta$  7.50 (d,  $J$  = 7.8 Hz, 2H), 7.39-7.35 (m, 2H), 7.33-7.30 (m, 3H), 6.91 (d,  $J$  = 8.8 Hz, 2H), 5.56 (d,  $J$  = 5.8 Hz, 1H), 5.04 (d,  $J$  = 10 Hz, 1H), 4.17 (t,  $J$  = 7.4 Hz, 1H), 3.82-3.77 (m, 4H), 3.43 (dd,  $J$  = 10.8, 8.0 Hz, 1H), 3.25 (dd,  $J$  = 11.4, 7.6 Hz, 1H), 2.88-2.79 (m, 1H), 2.66-2.57 (m, 1H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  161.8, 139.1, 130.72, 130.66, 129.5, 128.9, 128.7, 115.4, 88.5, 84.4, 69.8, 68.7, 56.4, 47.7, 42.3. HRMS (EI) calculated for  $[\text{C}_{19}\text{H}_{20}\text{O}_4]^+$  requires  $m/z$  312.1456, found  $m/z$  312.1457.



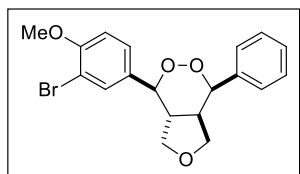
(Table 2, entry 3). Prepared according to the General Procedure using 100 mg (0.357 mmol) of bis(alkene), 6.2 mg (7.0  $\mu\text{mol}$ ) of  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$ , 17.8 mL of  $\text{MeNO}_2$  and an irradiation time of 24 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 52.0 mg (0.167 mmol, 47% yield, dr = 10:1), Experiment 2: 47.1 mg (0.151 mmol, 42% yield, dr = 10:1). IR (neat) 756, 735, 702  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR: (500 MHz)  $\delta$  7.50-7.28 (m, 7H), 7.00 (t,  $J$  = 7.9 Hz, 1H), 6.90 (d,  $J$  = 8.3 Hz, 1H), 5.62 (d,  $J$  = 6.3 Hz, 1H), 5.54 (d,  $J$  = 9.8 Hz, 1H), 4.10 (t,  $J$  = 7.3 Hz, 1H), 3.85-3.76 (m, 4H), 3.56 (dd,  $J$  = 10.9, 8.1 Hz, 1H), 3.13 (dd,  $J$  = 11.2, 7.7 Hz, 1H), 2.92 (m, 1H), 2.64 (m, 1H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  156.7, 136.7, 129.9, 128.3, 127.8, 127.3, 127.0, 120.8, 120.8, 110.5, 83.1, 81.7, 68.72, 68.69, 55.3, 46.0, 41.8. HRMS (EI) calculated for  $[\text{C}_{19}\text{H}_{20}\text{O}_4]^+$  requires  $m/z$  312.1456, found  $m/z$  312.1437.



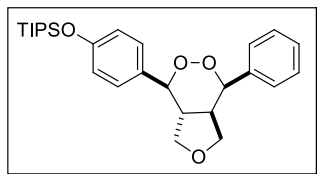
(Table 2, entry 5). Prepared according to the General Procedure using 100 mg (0.322 mmol) of bis(alkene), 1.4 mg (1.6  $\mu\text{mol}$ ) of  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$ , 16 mL of  $\text{MeNO}_2$  and an irradiation time of 2 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 75.4 mg (0.220 mmol, 68% yield, dr = 4:1), Experiment 2: 71.7 mg (0.209 mmol, 65% yield, dr = 4:1). IR (neat) 1210, 735, 702  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR: (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.49 (d,  $J$  = 7.2 Hz, 2H), 7.40-7.28 (m, 4H), 6.51 (dd,  $J$  = 8.5, 2.4 Hz, 1H), 6.46 (d,  $J$  = 2.4 Hz, 1H), 5.58 (d,  $J$  = 6 Hz, 1H), 5.48 (d,  $J$  = 9.7 Hz, 1H), 4.14 (t,  $J$  = 7.4 Hz, 1H), 3.86-3.81 (m, 7H), 3.48 (dd,  $J$  = 10.8, 8.1 Hz, 1H), 3.20 (dd,  $J$  = 11.3, 7.6 Hz, 1H), 2.92-2.83 (m, 1H), 2.71-2.61 (m, 1H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  161.5, 158.5, 137.0, 128.6, 128.1, 127.6, 127.1, 118.2, 104.4, 98.5, 83.1, 81.4, 68.8, 68.4, 55.43, 55.35, 46.4, 40.9. HRMS (EI) calculated for  $[\text{C}_{20}\text{H}_{22}\text{O}_5]^+$  requires  $m/z$  342.1462, found  $m/z$  342.1444.



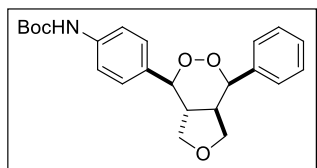
(Table 2, entry 6). Prepared according to the General Procedure using 100 mg (0.322 mmol) of bis(alkene), 1.4 mg (1.6  $\mu\text{mol}$ ) of  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$ , 16 mL of  $\text{MeNO}_2$  and an irradiation time of 1 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 100.9 mg (0.295 mmol, 92% yield, dr = 7:1), Experiment 2: 99.5 mg (0.291 mmol, 90% yield, dr = 7:1). IR (neat) 1261, 731, 702  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR: (500 MHz)  $\delta$  7.50 (d,  $J$  = 7.5 Hz, 2H), 7.39–7.29 (m, 3H), 6.96–6.91 (m, 2H), 6.87 (d,  $J$  = 8.2 Hz, 1H), 5.58 (d,  $J$  = 5.8 Hz, 1H), 5.05 (d,  $J$  = 9.9 Hz, 1H), 4.18 (t,  $J$  = 7.3 Hz, 1H), 3.90 (s, 3H), 3.88 (s, 3H), 3.83 (t,  $J$  = 7.8 Hz, 1H), 3.46 (dd,  $J$  = 10.8, 7.9 Hz, 1H), 3.26 (dd,  $J$  = 11.3, 7.6 Hz, 1H), 2.85 (m, 1H),  $\delta$  2.63 (m, 1H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  149.9, 149.2, 137.0, 129.3, 128.2, 127.7, 127.1, 120.0, 111.1, 111.0, 87.6, 83.1, 68.9, 68.2, 55.91, 55.89, 46.4, 41.1. HRMS (EI) calculated for  $[\text{C}_{20}\text{H}_{22}\text{O}_5]^+$  requires  $m/z$  342.1462, found  $m/z$  342.1472.



(Table 2, entry 7). Prepared according to the General Procedure using 100 mg (0.279 mmol) of bis(alkene), 4.8 mg (5.6  $\mu\text{mol}$ ) of  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$ , 14 mL of  $\text{MeNO}_2$  and an irradiation time of 12 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 70.5 mg (0.196 mmol, 65% yield, dr = 8:1), Experiment 2: 69.5 mg (0.194 mmol, 64% yield, dr = 8:1). IR (neat) 1262, 732, 702  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR: (500 MHz)  $\delta$  7.57 (d,  $J$  = 2.2 Hz, 1H), 7.50 (d,  $J$  = 7.5 Hz, 2H), 7.39 (t,  $J$  = 7.6 Hz, 2H), 7.32 (m, 2H), 6.91 (d,  $J$  = 8.47, 1H), 5.59 (d,  $J$  = 5.9 Hz, 1H), 5.01 (d,  $J$  = 10.1 Hz, 1H), 4.17 (t,  $J$  = 7.3 Hz, 1H), 3.91 (s, 3H), 3.82 (t,  $J$  = 7.7 Hz, 1H), 3.46 (dd,  $J$  = 10.7, 7.9 Hz, 1H), 3.24 (dd,  $J$  = 11.4, 7.7 Hz, 1H), 2.85 (m, 1H), 2.59 (m, 1H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  156.6, 136.7, 132.5, 130.6, 128.3, 127.84, 127.80, 127.0, 112.0, 111.9, 86.6, 83.2, 68.9, 68.0, 56.3, 46.3, 41.4. HRMS (EI) calculated for  $[\text{C}_{19}\text{H}_{19}\text{O}_4\text{Br}]^+$  requires  $m/z$  390.0462, found  $m/z$  390.0461.

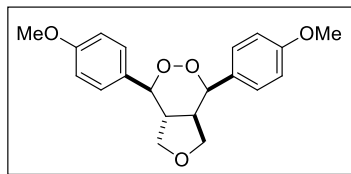


(Table 2, entry 8). Prepared according to the General Procedure using 100 mg (0.237 mmol) of bis(alkene), 1.0 mg (1.2  $\mu\text{mol}$ ) of  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$ , 11.8 mL of  $\text{MeNO}_2$  and an irradiation time of 2 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 77.1 mg (0.170 mmol, 72% yield, dr = 10:1), Experiment 2: 82.8 mg (0.182 mmol, 77% yield, dr = 10:1). IR (neat) 737, 701, 684  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR: (500 MHz)  $\delta$  7.50 (d,  $J$  = 7.5 Hz, 2H), 7.39–7.28 (m, 3H), 7.23 (d,  $J$  = 8.6 Hz, 2H), 6.88 (d,  $J$  = 8.6 Hz, 2H), 5.55 (d,  $J$  = 6.0 Hz, 1H), 5.02 (d,  $J$  = 9.8 Hz, 1H), 4.16 (t,  $J$  = 7.3 Hz, 1H), 3.76 (t,  $J$  = 7.8 Hz, 1H), 3.43 (dd,  $J$  = 10.8, 8.0 Hz, 1H), 3.24 (dd,  $J$  = 11.3, 7.6 Hz, 1H), 2.82 (m, 1H), 2.59 (m, 1H), 1.26 (septet,  $J$  = 7.5 Hz, 3H), 1.1 (d,  $J$  = 7.5 Hz, 18H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  157.0, 137.1, 129.4, 128.9, 128.2, 127.6, 127.1, 120.1, 87.5, 83.2, 68.9, 68.1, 46.5, 41.4, 17.9, 12.6. HRMS (EI) calculated for  $[\text{C}_{27}\text{H}_{38}\text{O}_4\text{Si}]^+$  requires  $m/z$  454.2534, found  $m/z$  454.2549.



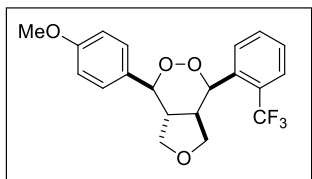
(Table 2, entry 9). Prepared according to the General Procedure using 100 mg (0.281 mmol) of bis(alkene), 1.2 mg (1.4  $\mu\text{mol}$ ) of  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$ , 14 mL of  $\text{MeNO}_2$  and an irradiation time of 30 min. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 103.7 mg (0.267 mmol, 95% yield, dr = 6:1), Experiment 2: 104.6 mg (0.269 mmol, 96% yield, dr = 6:1). IR (neat) 3333, 2876, 2885  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR: (500 MHz)  $\delta$  7.48 (d,  $J$  = 7.5 Hz, 2H), 7.40–7.35 (m, 4H), 7.39–7.28 (m, 3H), 6.57 (s, 1H), 5.58 (d,  $J$  = 5.9 Hz, 1H), 5.04 (d,  $J$  = 9.8 Hz, 1H), 4.17 (t,  $J$  = 7.3 Hz, 1H), 3.79 (t,  $J$  = 7.9 Hz, 1H), 3.45 (dd,  $J$  = 10.7, 8.0 Hz, 1H), 3.24 (dd,  $J$  = 11.4, 7.7 Hz, 1H), 2.89–2.80 (m, 1H), 2.66–2.56 (m, 1H), 1.51 (s, 9H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  152.5, 139.4, 136.9, 131.4, 128.33, 128.25, 127.7, 127.1, 118.6, 87.3, 83.2,

80.8, 68.9, 68.1, 46.4, 41.3, 28.3. HRMS (EI) calculated for  $[C_{23}H_{27}O_5N+Na]^+$  requires  $m/z$  420.1782, found  $m/z$  420.1790.



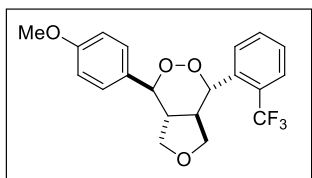
(Table 2, entry 11). Prepared according to the General Procedure using 100 mg (0.322 mmol) of bis(alkene), 1.4 mg (1.6  $\mu$ mol) of  $Ru(bpz)_3(PF_6)_2$ , 16.1 mL of  $MeNO_2$  and an irradiation time of 30 min. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 83.0 mg (0.242 mmol, 75% yield, dr >10:1), Experiment 2: 82.1 mg (0.239 mmol, 74% yield, dr >10:1). IR (neat)

1159, 732, 702  $cm^{-1}$ .  $^1H$  NMR: (500 MHz)  $\delta$  7.42 (d,  $J$  = 8.6 Hz, 2H), 7.32 (d,  $J$  = 8.6 Hz, 2H), 6.91 (d,  $J$  = 8.6 Hz, 2H), 6.90 (d,  $J$  = 8.6 Hz, 2H), 5.52 (d,  $J$  = 5.7 Hz, 1H), 5.04 (d,  $J$  = 9.9 Hz, 1H), 4.14 (t,  $J$  = 7.3 Hz, 1H), 3.83–3.80 (m, 7H), 3.44 (dd,  $J$  = 10.8, 8 Hz, 1H), 3.23 (dd,  $J$  = 11.2, 7.5 Hz, 1H), 2.86–2.77 (m, 1H), 2.69–2.60 (m, 1H).  $^{13}C$  NMR: (125 MHz,  $CDCl_3$ )  $\delta$  160.3, 159.0, 129.1, 129.03, 128.95, 128.4, 114.2, 113.6, 87.3, 82.9, 68.9, 68.1, 55.3, 55.2, 46.7, 41.1. HRMS (EI) calculated for  $[C_{20}H_{22}O_5]^+$  requires  $m/z$  342.1462, found  $m/z$  342.1453.



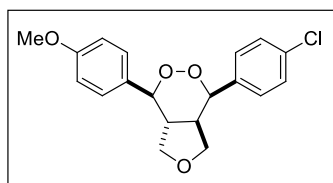
(Table 2, entry 12). Prepared according to the General Procedure using 100 mg (0.287 mmol) of bis(alkene), 1.2 mg (1.4  $\mu$ mol)  $Ru(bpz)_3(PF_6)_2$ , 14.4 mL of  $MeNO_2$  and an irradiation time of 2 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 90.3 mg (0.237 mmol, 83% yield, dr = 4:1), Experiment 2: 88.4 mg (0.232 mmol, 81% yield, dr = 4:1). IR (neat) 1312, 1120, 773  $cm^{-1}$ .  $^1H$  NMR: (500 MHz)  $\delta$

8.22 (d,  $J$  = 7.5 Hz, 1H), 7.68 (d,  $J$  = 7.8 Hz, 1H), 7.59 (t,  $J$  = 7.5 Hz, 1H), 7.45 (t,  $J$  = 7.5 Hz, 1H), 7.40 (d,  $J$  = 8.6 Hz, 2H), 6.97 (2,  $J$  = 8.8 Hz, 2H), 5.99 (d,  $J$  = 7.2 Hz, 1H), 5.07 (d,  $J$  = 9.7 Hz, 1H), 4.02 (td,  $J$  = 7.7, 1.7 Hz, 1H), 3.93 (t,  $J$  = 7.7 Hz, 1H), 3.84 (s, 3H), 3.53 (dd,  $J$  = 10.6, 7.9 Hz, 1H), 3.00 (m, 1H), 2.91–2.79 (m, 2H).  $^{13}C$  NMR: (125 MHz,  $CDCl_3$ )  $\delta$  160.2, 135.3, 132.1, 130.3, 128.8, 128.4, 128.2, 127.4 (q,  $J$  = 30.2 Hz), 126.2 (q,  $J$  = 5.8 Hz), 124.1 (t,  $J$  = 274.5 Hz), 114.3, 86.8, 78.8 (q,  $J$  = 2.7 Hz), 68.9 (q,  $J$  = 3.9 Hz), 68.7, 55.3, 45.5, 41.3. HRMS (EI) calculated for  $[C_{20}H_{19}O_4F_3]^+$  requires  $m/z$  380.1230, found  $m/z$  380.1237.



(Table 2, entry 12, minor diastereomer). IR (neat) 1315, 1253, 1037  $cm^{-1}$ .  $^1H$  NMR: (500 MHz)  $\delta$  7.73 (t,  $J$  = 7.6 Hz, 2H), 7.64 (t,  $J$  = 7.6 Hz, 1H), 7.50 (t,  $J$  = 7.6 Hz, 1H), 7.38 (d,  $J$  = 8.6 Hz, 2H), 6.94 (d,  $J$  = 8.6 Hz, 2H), 5.59 (d,  $J$  = 9.8 Hz, 1H), 5.18 (d,  $J$  = 10.0 Hz, 1H), 3.96 (t,  $J$  = 7.6 Hz, 1H), 3.84 (s, 3H), 3.77 (t,  $J$  = 7.5 Hz, 1H), 3.60 (dd,  $J$  = 11.0, 8.1 Hz, 1H), 3.55 (dd,  $J$  = 11.1, 8.0 Hz, 1H), 2.75 (m, 1H), 2.62 (m, 1H).  $^{13}C$  NMR: (125 MHz,  $CDCl_3$ )

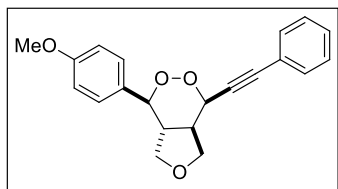
$\delta$  160.6, 135.1, 132.4, 129.3, 129.2, 128.54, 128.51, 127.8, 126.0 (q,  $J$  = 5.5 Hz), 123.9 (q,  $J$  = 274.5 Hz), 114.2, 87.1, 81.9, 68.2, 68.0, 55.3, 50.6, 48.5. HRMS (EI) calculated for  $[C_{20}H_{19}O_4F_3]^+$  requires  $m/z$  380.1230, found  $m/z$  380.1212.



(Table 2, entry 13). Prepared according to the General Procedure using 100 mg (0.318 mmol) of bis(alkene), 1.4 mg (1.6  $\mu$ mol) of  $Ru(bpz)_3(PF_6)_2$ , 15.9 mL of  $MeNO_2$  and an irradiation time of 2 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 94.0 mg (0.271 mmol, 85% yield, dr = 5:1), Experiment 2: 95.4 mg (0.275 mmol, 87% yield, dr = 5:1). IR (neat) 1252, 1062, 848  $cm^{-1}$ .  $^1H$  NMR:

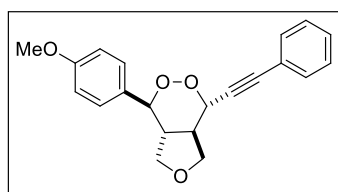
(500 MHz)  $\delta$  7.44 (d,  $J$  = 8.6 Hz, 2H), 7.35 (2,  $J$  = 8.6 Hz, 2H), 7.29 (d,  $J$  = 8.6 Hz, 2H), 6.91 (d,  $J$  = 8.6 Hz, 2H), 5.53 (d,  $J$  = 5.9 Hz, 1H), 5.04 (d,  $J$  = 10.0 Hz, 1H), 4.18 (t,  $J$  = 7.3 Hz, 1H), 3.83–3.77 (m, 4H), 3.43 (dd,  $J$  = 10.9, 8.1 Hz, 1H), 3.24 (dd,  $J$  = 11.4, 7.6 Hz, 1H), 2.82 (m, 1H), 2.55 (m, 1H).  $^{13}C$  NMR:

(125 MHz, CD<sub>3</sub>CN)  $\delta$  161.9, 138.0, 134.2, 130.8, 130.45, 130.39, 129.5, 115.4, 88.6, 83.8, 69.7, 68.6, 56.4, 47.6, 42.2;. HRMS (EI) calculated for [C<sub>19</sub>H<sub>19</sub>O<sub>4</sub>Cl]<sup>+</sup> requires m/z 346.0967, found m/z 346.0985.



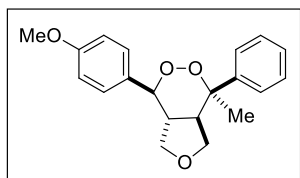
(Table 2, entry 14). Prepared according to the General Procedure using 100 mg (0.329 mmol) of bis(alkene), 1.4 mg (1.6  $\mu$ mol) of Ru(bpz)<sub>3</sub>(PF<sub>6</sub>)<sub>2</sub>, 16.6 mL of MeNO<sub>2</sub> and an irradiation time of 2 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 88 mg (0.262 mmol, 80% yield, dr = 2:1), Experiment 2: 92.1 mg (0.274 mmol, 83% yield, dr = 2:1). IR (neat) 1253,

758, 691 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz)  $\delta$  7.51 (m, 2H), 7.38–7.30 (m, 5H), 6.89 (d, J = 8.7 Hz, 2H), 5.34 (d, J = 4.7 Hz, 1H), 5.02 (d, J = 10.0 Hz, 1H), 4.17 (t, J = 7.4 Hz, 1H), 3.89 (t, J = 7.7 Hz, 1H), 3.80 (s, 3H), 3.74 (dd, J = 11.0, 7.1 Hz, 1H), 3.46 (dd, J = 11.4, 7.9 Hz, 1H), 2.96 (m, 1H), 2.65 (m, 1H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>)  $\delta$  160.5, 132.0, 129.2, 128.9, 128.4, 128.3, 122.0, 114.1, 89.0, 87.2, 82.8, 73.3, 68.8, 68.0, 55.3, 46.6, 43.5. HRMS (EI) calculated for [C<sub>21</sub>H<sub>20</sub>O<sub>4</sub>]<sup>+</sup> requires m/z 336.1456, found m/z 336.1464.



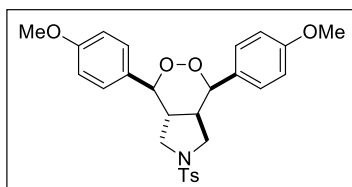
(Table 2, entry 14, minor diastereomer). IR (neat) 1252, 759, 691 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz)  $\delta$  7.46 (m, 2H), 7.38–7.28 (m, 5H), 6.90 (d, J = 8.7 Hz, 2H), 5.05 (d, J = 9.5 Hz, 2H), 4.23 (t, J = 7.3 Hz, 1H), 3.92 (t, J = 7.3 Hz, 1H), 3.81 (s, 3H), 3.69 (dd, J = 10.7, 7.8 Hz, 1H), 3.51 (dd, J = 10.7, 7.9 Hz, 1H), 2.55 (m, 2H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>)  $\delta$  160.6, 132.0, 129.21, 129.17, 128.4, 127.7, 121.5, 114.2, 88.8, 86.8, 82.7, 75.1, 68.4,

68.3, 55.3, 49.8, 48.1. HRMS (EI) calculated for [C<sub>21</sub>H<sub>20</sub>O<sub>4</sub>]<sup>+</sup> requires m/z 336.1456, found m/z 336.1461.



(Table 2, entry 15). Prepared according to the General Procedure using 100 mg (0.340 mmol) of bis(alkene), 5.9 mg (6.8  $\mu$ mol) of Ru(bpz)<sub>3</sub>(PF<sub>6</sub>)<sub>2</sub>, 17 mL of MeNO<sub>2</sub> and an irradiation time of 2 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 88.1 mg (0.270 mmol, 79% yield, dr >10:1), Experiment 2: 90.5 mg (0.277 mmol, 82% yield, dr >10:1). IR (neat) 1251, 738, 704 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz)  $\delta$

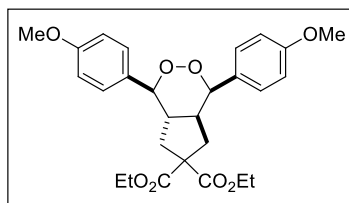
7.49 (m, 2H), 7.37 (m, 2H), 7.29 (m, 1H), 7.21 (d, J = 8.6 Hz, 2H), 6.86 (d, J = 8.6 Hz, 2H), 4.97 (d, J = 9.9 Hz, 1H), 4.28 (t, J = 7.0 Hz, 1H), 3.78 (s, 3H), 3.72 (t, J = 8.0 Hz, 1H), 3.66 (dd, J = 11.4, 7.4 Hz, 1H), 3.42 (dd, J = 10.6, 8.1 Hz, 1H), 2.51 (m, 1H), 2.30 (m, 1H), 1.61 (s, 3H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>)  $\delta$  160.3, 140.6, 129.1, 128.6, 127.9, 127.1, 126.9, 114.1, 87.3, 86.1, 68.8, 68.2, 55.2, 53.4, 43.0, 28.5. HRMS (EI) calculated for [C<sub>20</sub>H<sub>22</sub>O<sub>4</sub>]<sup>+</sup> requires m/z 326.1613, found m/z 326.1616.



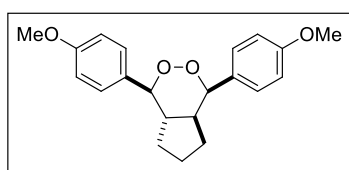
(Table 2, entry 16). Prepared according to the General Procedure using 100 mg (0.230 mmol) of bis(alkene), 1.0 mg (1.1  $\mu$ mol) of Ru(bpz)<sub>3</sub>(PF<sub>6</sub>)<sub>2</sub>, 11.5 mL of MeNO<sub>2</sub> and an irradiation time of 2 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 80.0 mg (0.171 mmol, 75% yield, dr >10:1), Experiment 2: 76.2 mg (0.163 mmol, 71% yield, dr >10:1). IR (neat)

1163, 733, 665 cm<sup>-1</sup>. <sup>1</sup>H NMR: (500 MHz)  $\delta$  7.57 (d, J = 8.2 Hz, 2H), 7.35 (d, J = 8.6 Hz, 2H), 7.25 (d, J = 8.2 Hz, 2H), 7.20 (d, J = 8.6 Hz, 2H), 6.91–6.86 (m, 4H), 5.32 (d, J = 5.7 Hz, 1H), 4.86 (d, J = 9.9 Hz, 1H), 3.83 (s, 3H), 3.82 (s, 3H), 3.66 (dd, J = 8.9, 7.5 Hz, 1H), 3.34 (dd, J = 9.8, 7.2 Hz, 1H), 2.82 (m, 2H), 2.47 (m, 1H), 2.43 (s, 3H), 2.28 (m, 1H). <sup>13</sup>C NMR: (125 MHz, CDCl<sub>3</sub>)  $\delta$  160.5, 159.2, 143.5, 134.2, 129.7, 129.05, 128.96, 128.5, 128.4, 127.1, 114.2, 113.7, 87.0, 82.7, 55.3, 55.2, 49.5, 48.9, 45.1, 39.7, 21.4;. HRMS (EI) calculated for [C<sub>27</sub>H<sub>29</sub>NO<sub>6</sub>S+Na]<sup>+</sup> requires m/z 518.1608, found m/z 518.1614.



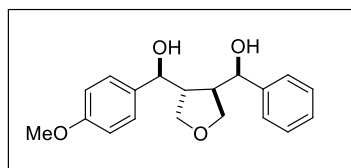


(Table 2, entry 17). Prepared according to the General Procedure using 100 mg (0.220 mmol) of bis(alkene), 1.0 mg (1.1  $\mu\text{mol}$ ) of  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$ , 11 mL of  $\text{MeNO}_2$  and an irradiation time of 2 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 97.4 mg (0.201 mmol, 91% yield, dr >10:1), Experiment 2: 98.8 mg (0.204 mmol, 92% yield, dr >10:1). IR (neat) 1253, 1034, 733  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR: (500 MHz)  $\delta$  7.49 (d,  $J$  = 8.6 Hz, 2H), 7.32 (d,  $J$  = 8.6 Hz, 2H), 6.93–6.87 (m, 4H), 5.37 (d,  $J$  = 5.7 Hz, 1H), 4.93 (d,  $J$  = 9.8 Hz, 1H), 4.18 (m, 2H), 4.06 (m, 2H), 3.82 (s, 3H), 3.81 (s, 3H), 2.64 (dd,  $J$  = 12.8, 6.1 Hz, 1H), 2.49 (m, 1H), 2.33 (dd,  $J$  = 13.1, 7.1 Hz, 1H), 2.26 (m, 1H), 1.74 (dd,  $J$  = 13.0, 11.6 Hz, 1H), 1.53 (t,  $J$  = 13.0 Hz, 1H), 1.23 (t,  $J$  = 7.2 Hz, 3H), 1.13 (t,  $J$  = 7.2 Hz, 3H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  172.2, 171.9, 160.2, 158.9, 129.4, 129.3, 128.9, 128.8, 114.1, 113.5, 89.5, 84.8, 61.6, 61.5, 57.4, 55.25, 55.18, 46.2, 40.8, 36.5, 35.1, 14.0, 13.8. HRMS (EI) calculated for  $[\text{C}_{27}\text{H}_{32}\text{O}_8+\text{Na}]^+$  requires  $m/z$  507.1990, found  $m/z$  507.2000.

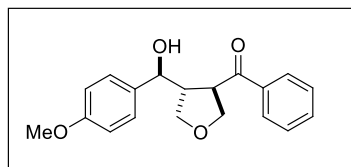


(Table 2, entry 18). Prepared according to the General Procedure using 100 mg (0.325 mmol) of bis(alkene), 1.4 mg (1.6  $\mu\text{mol}$ ) of  $\text{Ru}(\text{bpz})_3(\text{PF}_6)_2$ , 16 mL of  $\text{MeNO}_2$  and an irradiation time of 2 h. Purification by flash column chromatography afforded the cycloadduct as an oil. Experiment 1: 88.0 mg (0.259 mmol, 80% yield, >10:1dr), Experiment 2: 86.9 mg (0.256 mmol, 79% yield, >10:1dr). IR (neat) 1250, 1032, 743  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR: (500 MHz)  $\delta$  7.53 (d,  $J$  = 9.0 Hz, 2H), 7.32 (d,  $J$  = 9.0 Hz, 2H), 6.90 (d,  $J$  = 9.0 Hz, 4H), 5.39 (d,  $J$  = 5.8 Hz, 1H), 4.91 (d,  $J$  = 10 Hz, 1H), 3.82 (s, 3H), 3.81 (s, 3H), 2.27 (m, 1H), 2.03 (m, 1H), 1.87 (m, 1H), 1.66 (m, 1H), 1.55 (m, 2H), 1.19 (m, 1H), 1.06 (m, 1H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  160.0, 158.7, 130.5, 129.8, 129.3, 129.0, 113.9, 113.2, 90.7, 85.9, 55.24, 55.18, 47.4, 41.4, 27.3, 25.8, 21.2. HRMS (EI) calculated for  $[\text{C}_{21}\text{H}_{24}\text{O}_4]^+$  requires  $m/z$  340.1670, found  $m/z$  340.1657.

#### IV. Endoperoxide opening reactions



**Compound 6.** A 2 dram vial equipped with a stirbar was charged with 139 mg (0.440 mmol) of **5**. AcOH (2.2 mL) and zinc dust (144 mg, 2.2 mmol) were added, the vial was sealed with a Teflon cap, and the reaction was stirred at room temperature. After 2 h, the reaction mixture was concentrated by rotary evaporation and then eluted through a short plug of silica using EtOAc. The eluent was concentrated by rotary evaporation, and the residue was purified by chromatography on  $\text{SiO}_2$  (2:1 to 1:1 hexanes/EtOAc) to afford 126 mg (0.40 mmol, 91% yield) of the desired product as a colorless solid (mp = 150–151  $^\circ\text{C}$ ).  $^1\text{H}$  NMR: (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.34 (m, 4H), 7.28 (m, 1H), 7.19 (d,  $J$  = 8.6 Hz, 2H), 6.85 (d,  $J$  = 8.6 Hz, 2H), 4.79 (s, 1H), 4.48 (d,  $J$  = 9.0 Hz, 1H), 3.88 (dd,  $J$  = 9.0, 7.7 Hz, 1H), 3.80 (m, 4H), 3.79 (dd,  $J$  = 9.1, 6.6 Hz, 1H), 3.49 (dd,  $J$  = 9.1, 7.8 Hz, 1H), 3.39 (dd,  $J$  = 9.3, 6.1 Hz, 1H), 3.03 (s, 1H), 2.77 (m, 1H), 2.53 (m, 1H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  159.4, 142.4, 134.9, 128.4, 127.7, 127.6, 126.7, 114.0, 76.3, 74.4, 71.0, 70.2, 55.3, 49.8, 48.1. HRMS (EI) calculated for  $[\text{C}_{19}\text{H}_{22}\text{O}_4+\text{Na}]^+$  requires  $m/z$  337.1411, found  $m/z$  337.1409.



**Compound 7.** A 2 dram vial equipped with a stirbar was charged with 74.0 mg (0.240 mmol) of endoperoxide **5**. Dry  $\text{CH}_2\text{Cl}_2$  (1.2 mL) and  $\text{NEt}_3$  (10  $\mu\text{L}$ , 0.07 mmol) were added sequentially, the vial was sealed with a Teflon cap, and the reaction was stirred at room temperature. After 13 h, the reaction mixture was concentrated by rotary evaporation and the crude reaction mixture was purified by chromatography on  $\text{SiO}_2$  (2:1 to 1:1 hexanes/EtOAc) to afford 69.0 mg (0.22 mmol, 92% yield) of a colorless oil.  $^1\text{H}$  NMR: (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.95 (d,  $J$  = 7.8 Hz, 2H), 7.56 (t,  $J$  = 7.2 Hz, 1H), 7.44 (t,  $J$  = 7.5 Hz, 2H), 7.27 (d,  $J$  = 8.7 Hz, 2H), 6.85 (d,  $J$  = 8.7 Hz, 2H), 4.61 (d,  $J$  = 9.0 Hz, 1H), 4.26 (m, 2H), 3.89 (m, 1H), 3.78 (s, 3H), 3.74 (dd,

$J = 9.0, 7.2$  Hz, 1H), 3.60 (dd,  $J = 9.0, 4.6$  Hz, 1H), 3.16 (m, 1H), 2.33 (s, 1H).  $^{13}\text{C}$  NMR: (125 MHz,  $\text{CDCl}_3$ )  $\delta$  200.0, 159.4, 136.4, 134.9, 133.2, 128.62, 128.60, 127.6, 114.0, 75.7, 71.3, 70.8, 55.2, 50.6, 49.7. HRMS (E1) calculated for  $[\text{C}_{19}\text{H}_{20}\text{O}_4+\text{Na}]^+$  requires  $m/z$  335.1254, found  $m/z$  335.1256.

## V. Representative NOE data

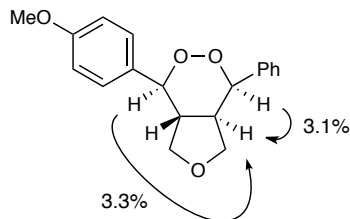


Table 2, entry 1

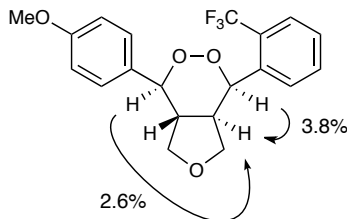


Table 2, entry 12

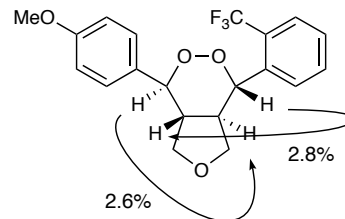


Table 2, entry 12  
minor diastereomer

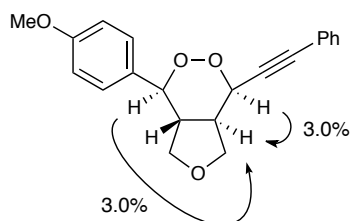


Table 2, entry 14

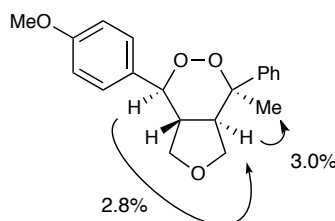


Table 2, entry 15

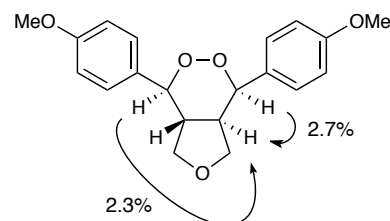


Table 2, entry 17

## VI. Cytotoxicity screening data

**Table S-1.** Tabulated cytotoxicity data for representative endoperoxides from Table 2.

Compound	IC <sub>50</sub> (μM)	SE
Table 2, Entry 1	>100	NA
Table 2, Entry 3	27.64	1.27
Table 2, Entry 5	>100	NA
Table 2, Entry 6	77.72	1.82
Table 2, Entry 7	38.71	2.66
Table 2, Entry 11	>100	NA
Table 2, Entry 12	13.18	0.63
Table 2, Entry 13	44.30	1.89
Table 2, Entry 14	4.56	0.19
Table 2, Entry 16	17.00	0.46

As an initial demonstration of their potential biological activity, a selection of the compounds reported in Table 2 were assayed for cytotoxicity in human prostate cancer cells (Du145s). Cells were treated for 72 hours with each test compound and then the amount of live cells was quantified using an ATP determining assay reagent, Cell Titer Glo (Promega). The IC<sub>50</sub> was determined using curve fitting software, xfit 5.0 (IDBS), to determine the concentration at which point 50% cytotoxicity was observed. Working stocks were prepared at a final concentration of 100X in anhydrous DMSO. Serial dilutions were made in DMSO in 96-well polypropylene plates using the Precision XS liquid handler (BioTek, Inc.). Compounds were divided equally into the corresponding wells of a 384-well plate in all 4 quadrants using a Biomek FX liquid handler with 96 channel pipetting head (Beckman-Coulter, Inc.). Compounds

were stored at  $-20\text{ }^{\circ}\text{C}$  in 100% DMSO until the day of the assay(s). Freeze-thaw cycles are minimized to a maximum of 10 per plate.

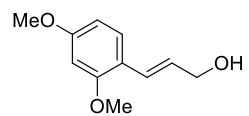
All cell lines were maintained as previously reported.<sup>8</sup> Cells were harvested by trypsinization using 0.25% trypsin and 0.1% EDTA and then counted in a Cellometer Auto T4 cell counter (Nexcelom, inc), before dilution for assay plating. Cell plating, compound handling and assay set up were performed as previously reported (Langenhan, et al) except that the cells were plated in 50  $\mu\text{L}$  volumes in 384-well clear bottom, tissue culture plates (Corning-Costar, Inc). Compounds were added from the 384-well compound stock plates at a 1:100 dilution using a Biomek FX liquid handler equipped with a 384-channel head (Beckman Coulter, Inc.). 15  $\mu\text{L}$  of cell titer-glo reagent (Promega Corporation, Inc.) was added and incubated for 10 min at room temperature with gentle agitation to lyse the cells.

$\text{IC}_{50}$ s for cytotoxicity assays were determined cell-titer glo using xfit 5.0 as previously reported in Langenhan, et al. The  $\text{IC}_{50}$  determined using the best fit curve for dosage response is reported as the final  $\text{IC}_{50}$ . The best fit curve is defined as the curve that has the lowest standard error. If the standard error exceeds 10% of the  $\text{IC}_{50}$ , the assay is deemed a failure and repeated.

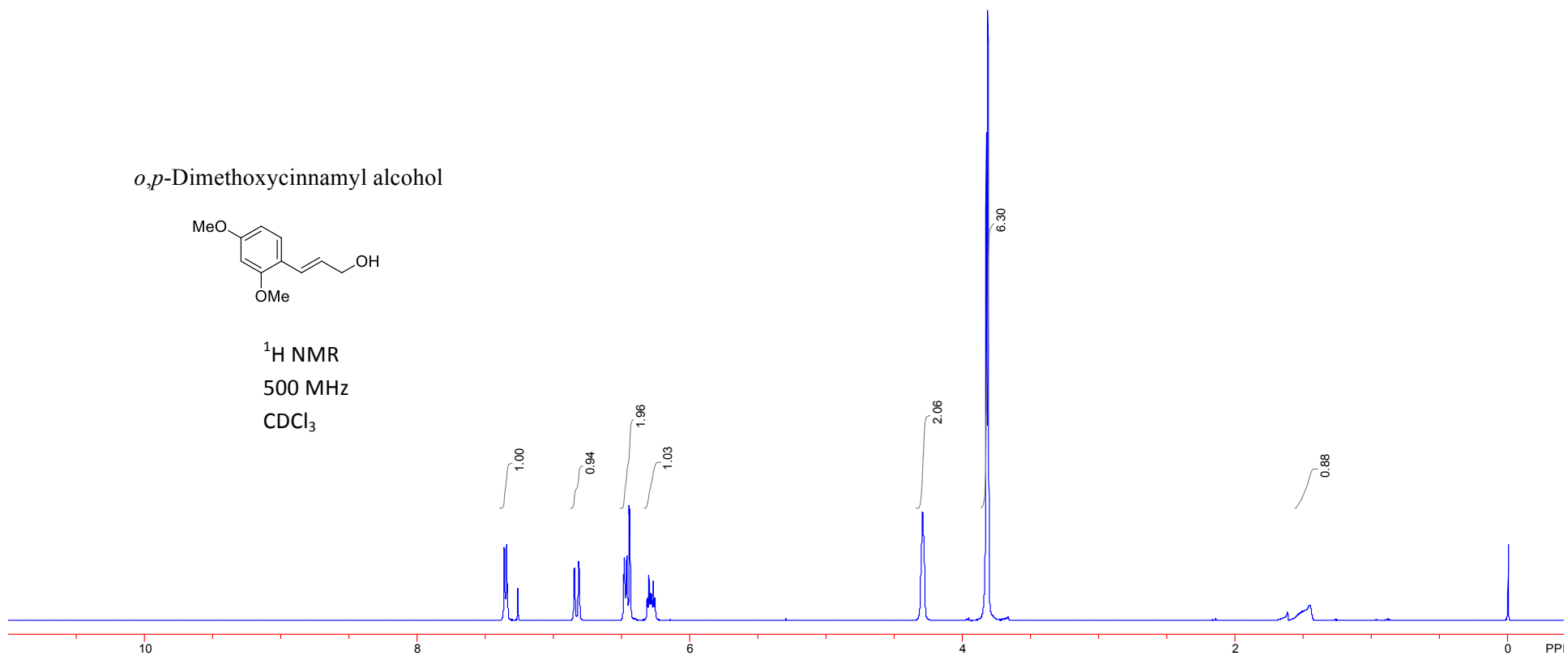
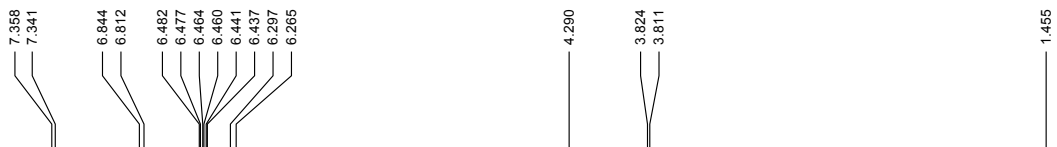
## **VII. References**

- <sup>1</sup>Pangborn, A. B.; Giardello, M. A.; Grubbs, R. H.; Rosen, R. K.; Timmers, F. T. *Organometallics* **1996**, *15*, 1518–1520.
- <sup>2</sup>Lin, S.; Ischay, M. A.; Fry, C. G.; Yoon, T. P. *J. Am. Chem. Soc.* **2011**, *133*, 19350–19353.
- <sup>3</sup>Ischay, M. A.; Lu, Z.; Yoon, T. P. *J. Am. Chem. Soc.* **2010**, *132*, 8572–8574.
- <sup>4</sup>Nair, V.; Balagopal, L.; Menon, R. S.; Ros, S.; Srinivas, R. *ARKIVOC* **2003**, *8*, 199–210.
- <sup>5</sup>McConnell, O.; He, Y.; Nogle, L.; Sarkahian, A. *Chirality* **2007**, *9*, 716–730.
- <sup>6</sup>Takeuchi, R.; Tanabe, K.; Tanaka, S. *J. Org. Chem.* **2000**, *65*, 1558–1561.
- <sup>7</sup>Ghorai, M. K.; Kumar, A.; Das, K. *Org. Lett.* **2007**, *9*, 5441–5444.
- <sup>8</sup>Langenhan, J. M.; Peters, N. R.; Guzei, I. A.; Hoffmann, F. M.; Thorson, J. S. *Proc. Nat. Acad. Sci.* **2005**, *102*, 12305–12310.

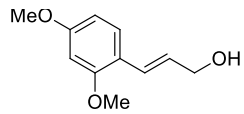
*o,p*-Dimethoxycinnamyl alcohol



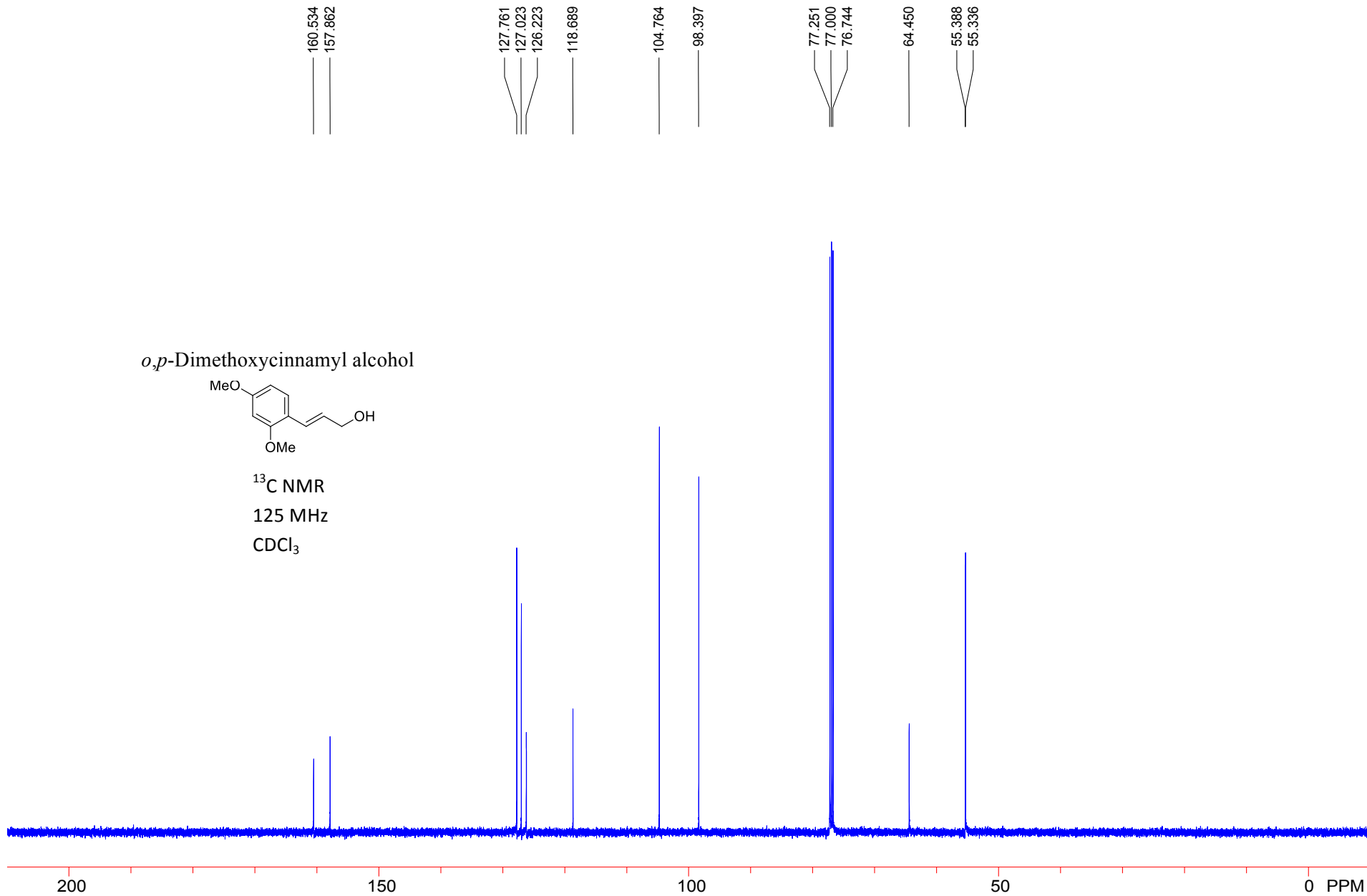
<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>



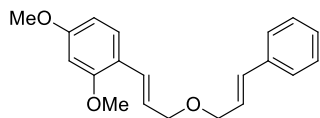
*o,p*-Dimethoxycinnamyl alcohol



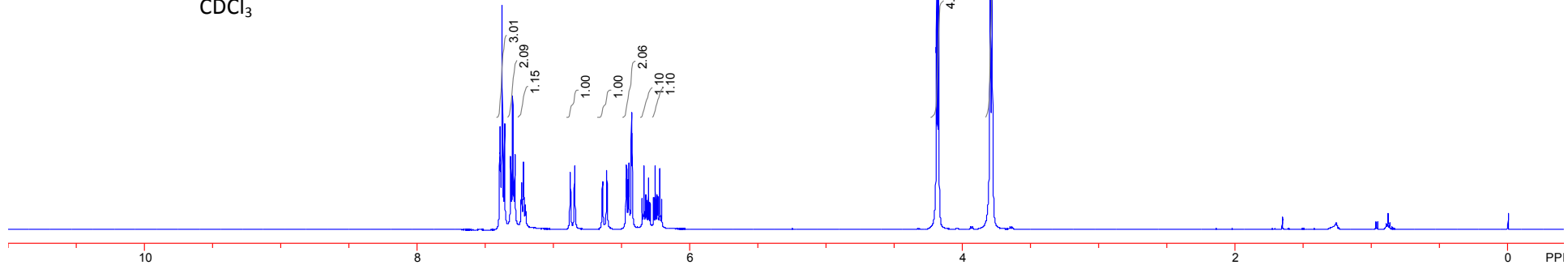
<sup>13</sup>C NMR  
125 MHz  
CDCl<sub>3</sub>



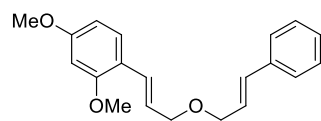
Substrate for Table 2, entry 5



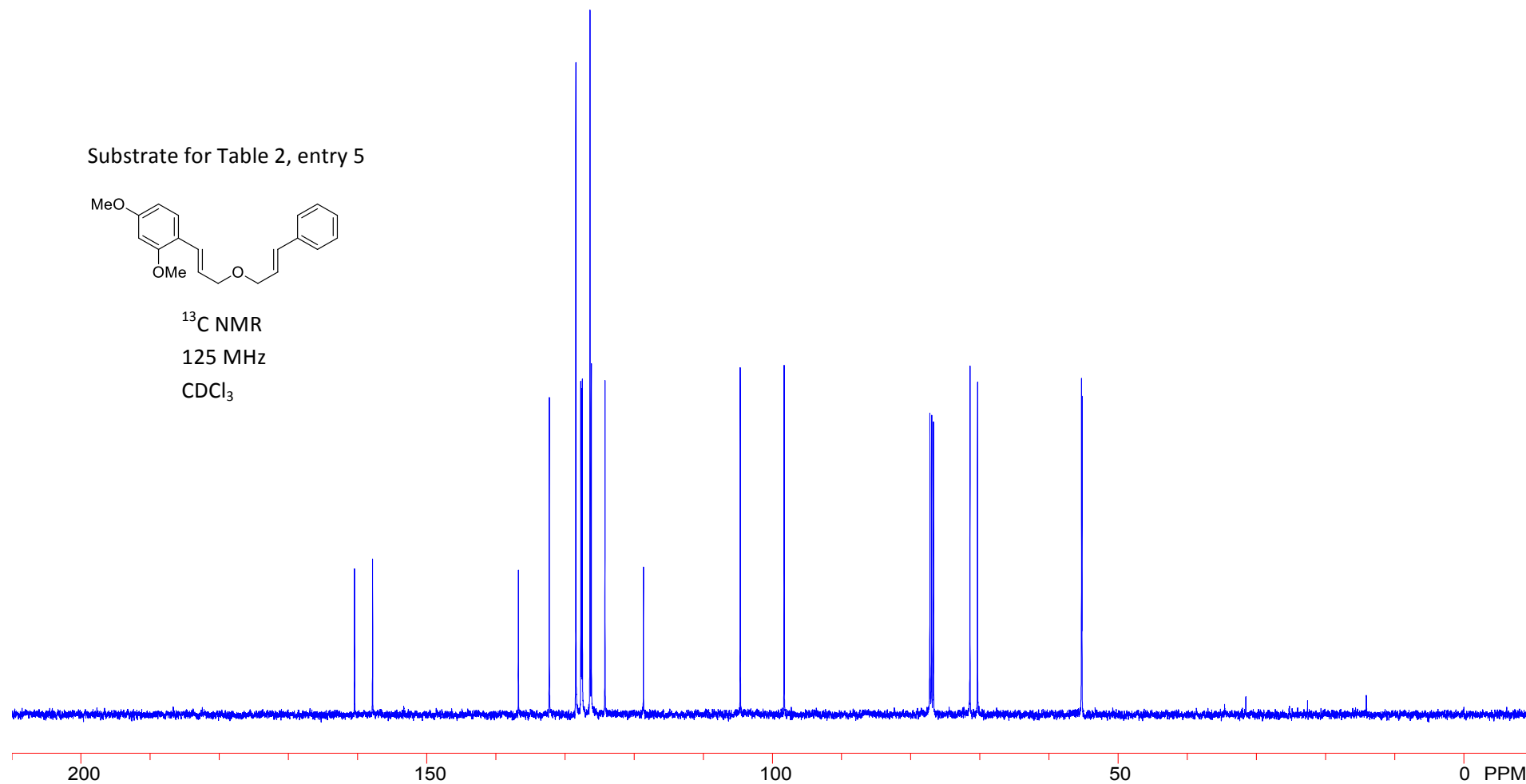
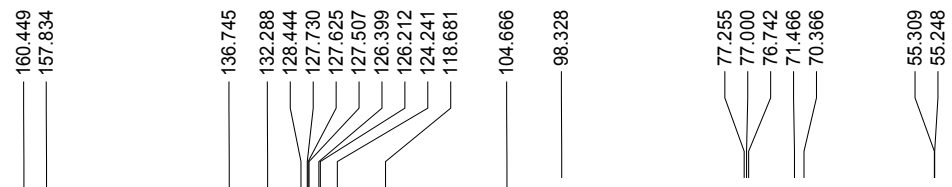
$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$

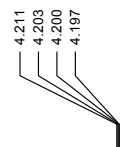
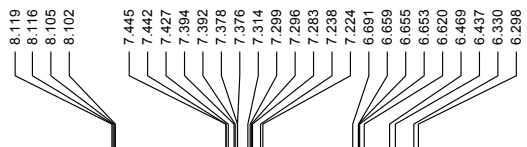


Substrate for Table 2, entry 5

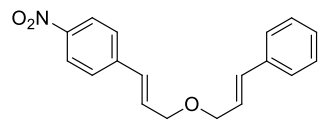


$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$

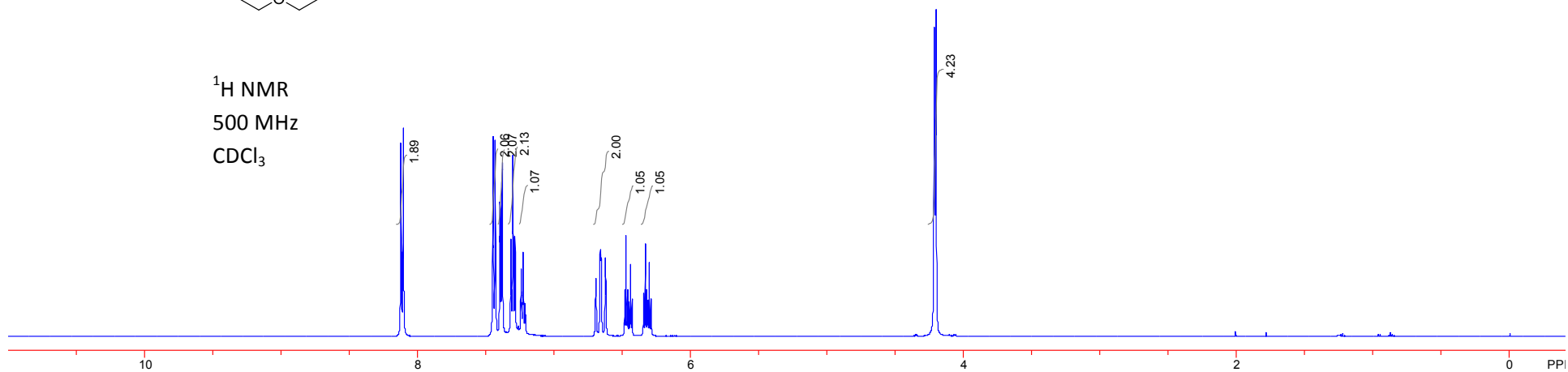




(*E*)-*p*-Nitrocinnamyl (*E*)-cinnamyl ether

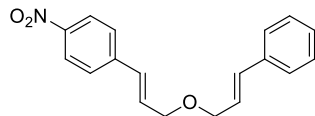


<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>

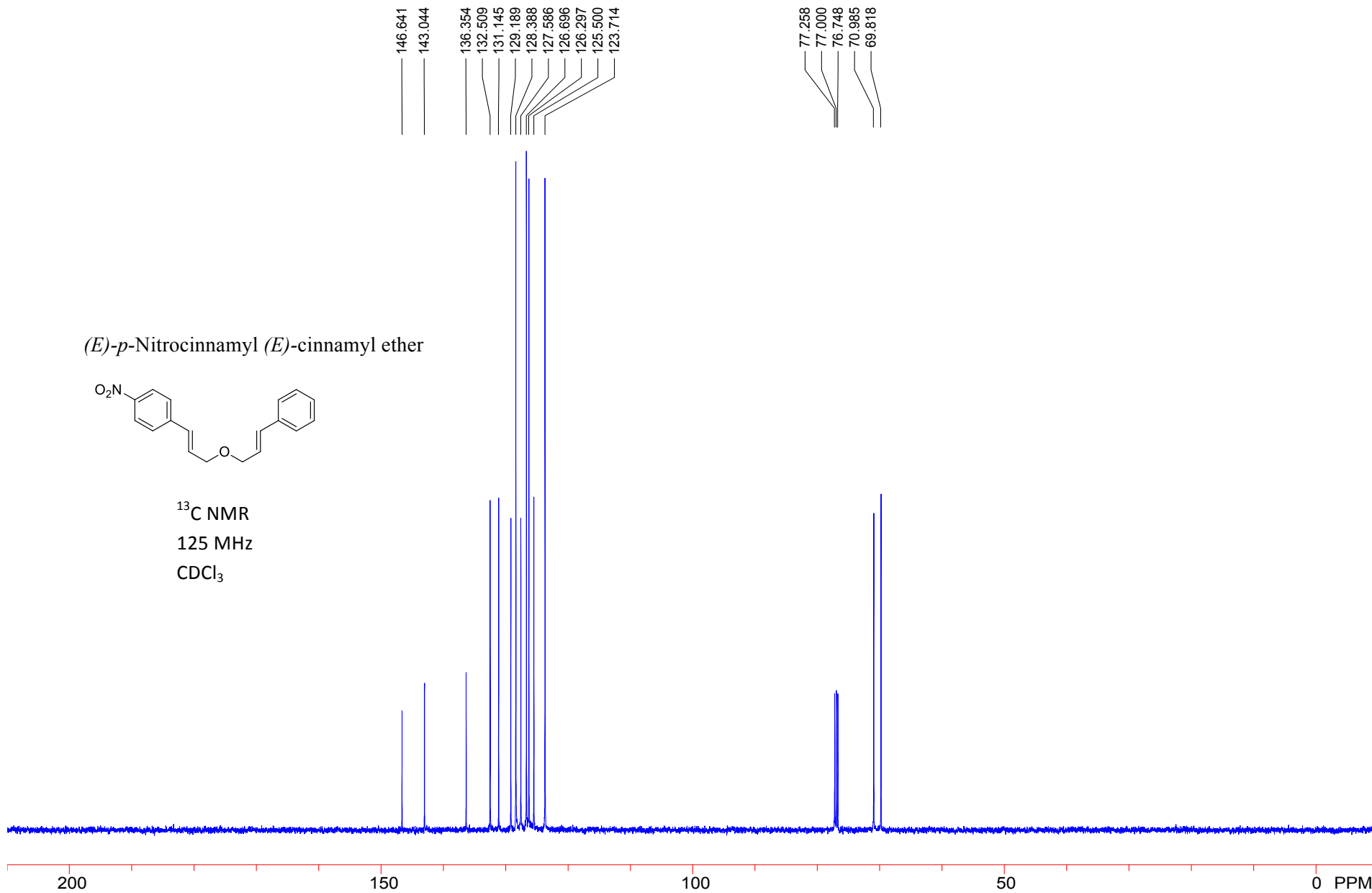


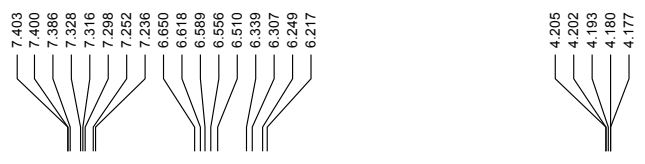


*(E)*-*p*-Nitrocinnamyl *(E)*-cinnamyl ether

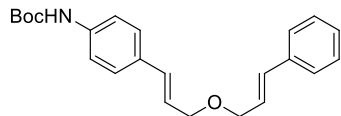


<sup>13</sup>C NMR  
125 MHz  
CDCl<sub>3</sub>

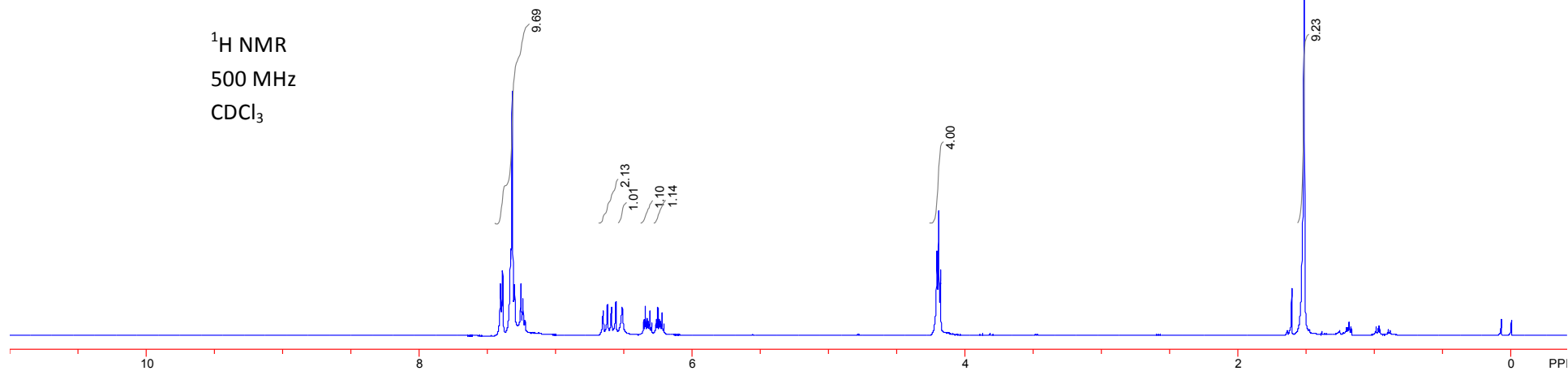




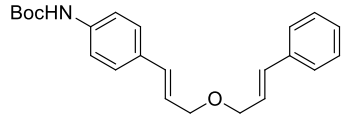
Substrate for Table 2, entry 9



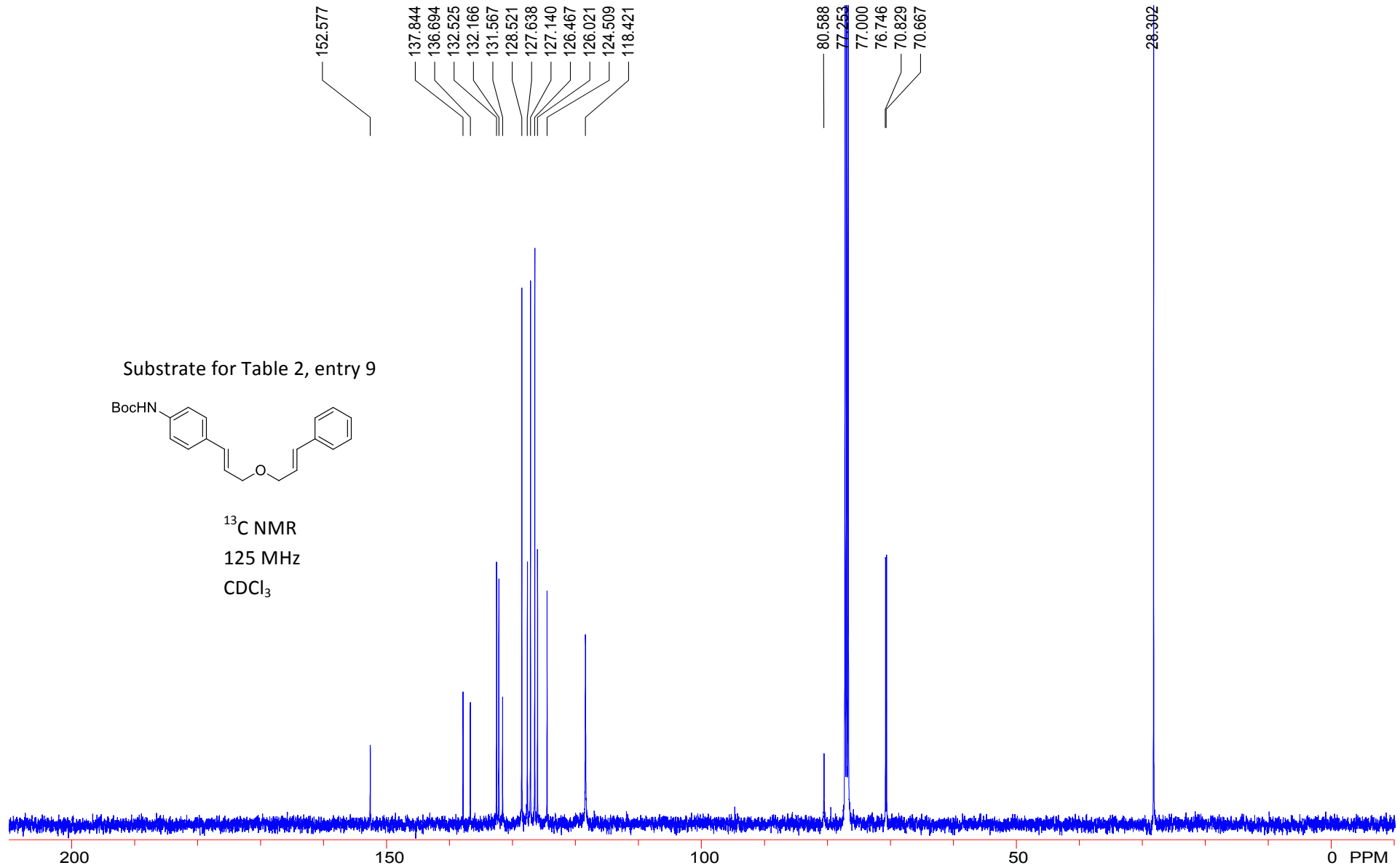
<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>



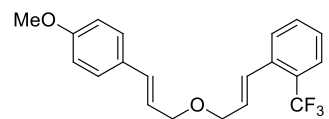
Substrate for Table 2, entry 9



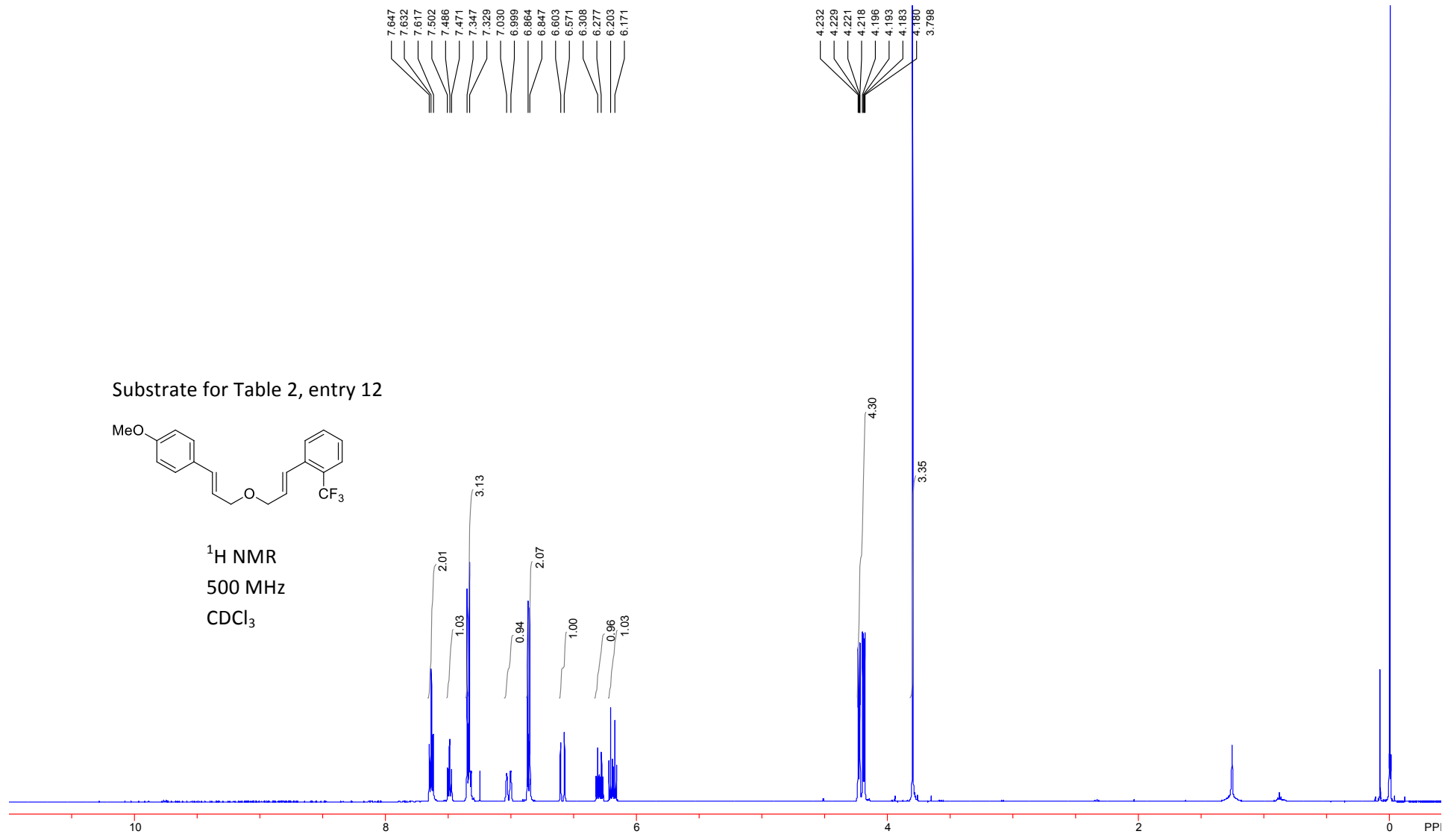
<sup>13</sup>C NMR  
125 MHz  
CDCl<sub>3</sub>

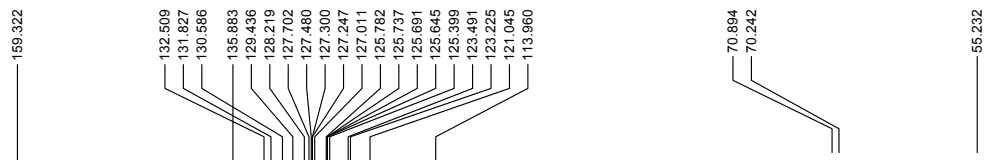


Substrate for Table 2, entry 12

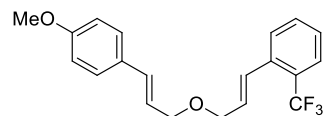


<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>

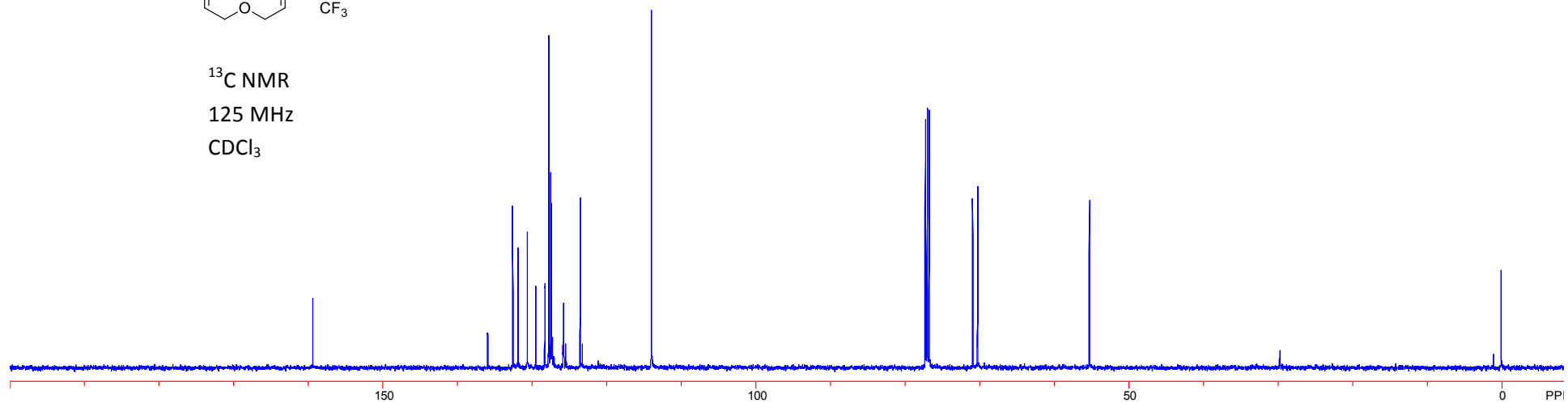




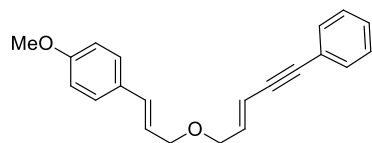
Substrate for Table 2, entry 12



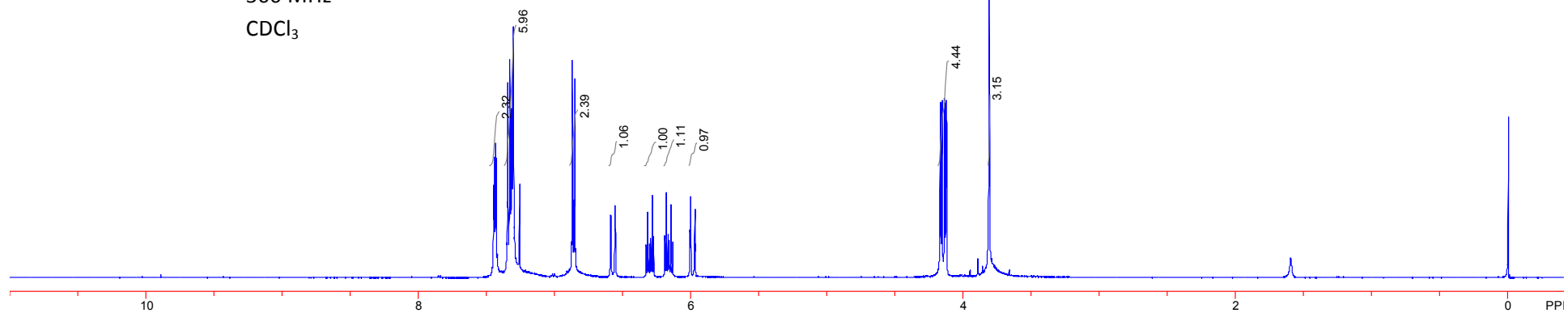
<sup>13</sup>C NMR  
125 MHz  
CDCl<sub>3</sub>



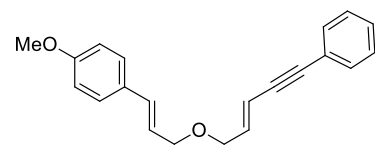
Substrate for Table 2, entry 14



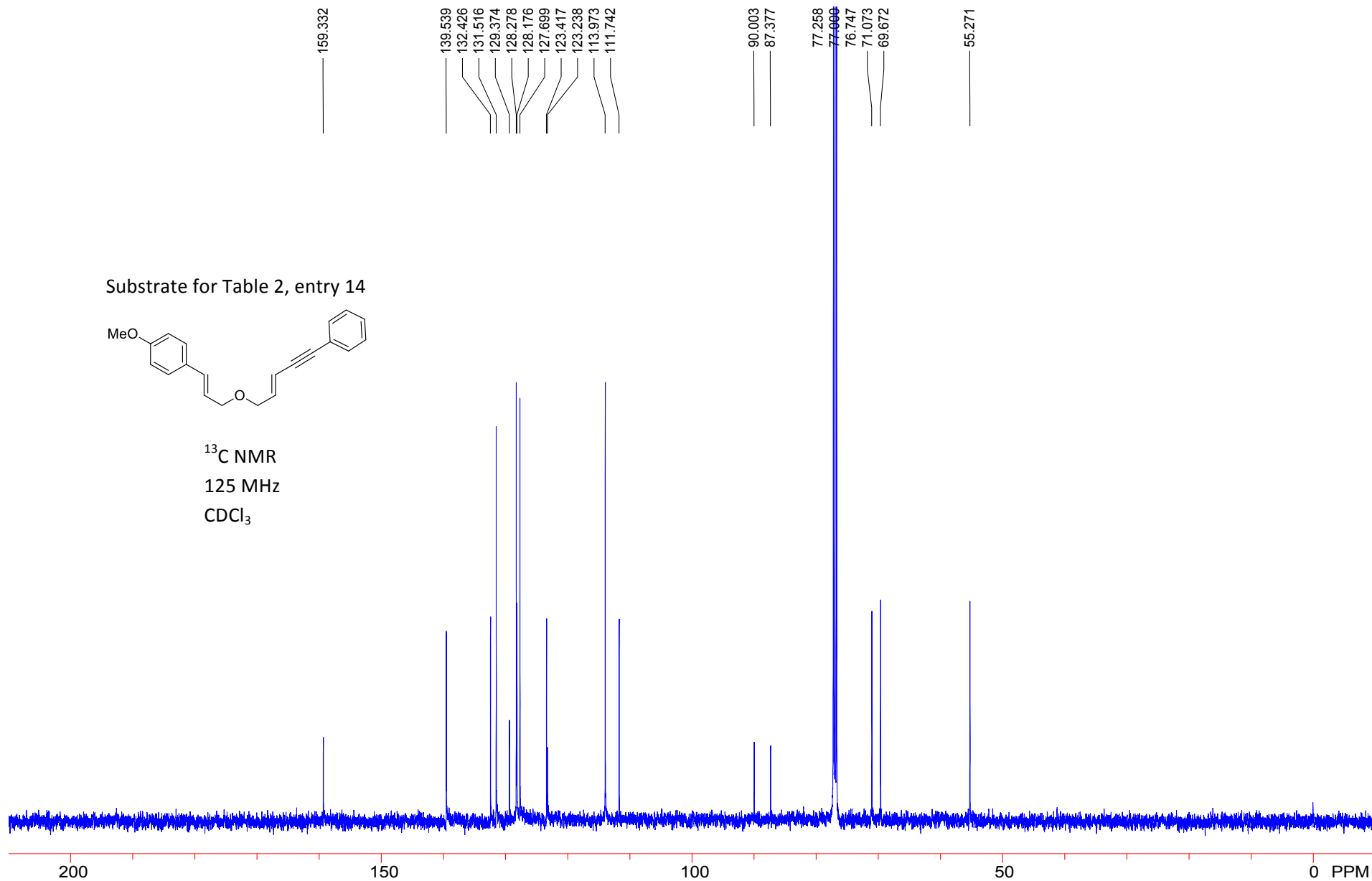
$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$



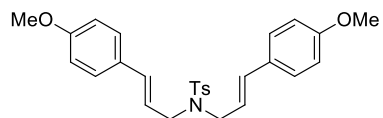
Substrate for Table 2, entry 14



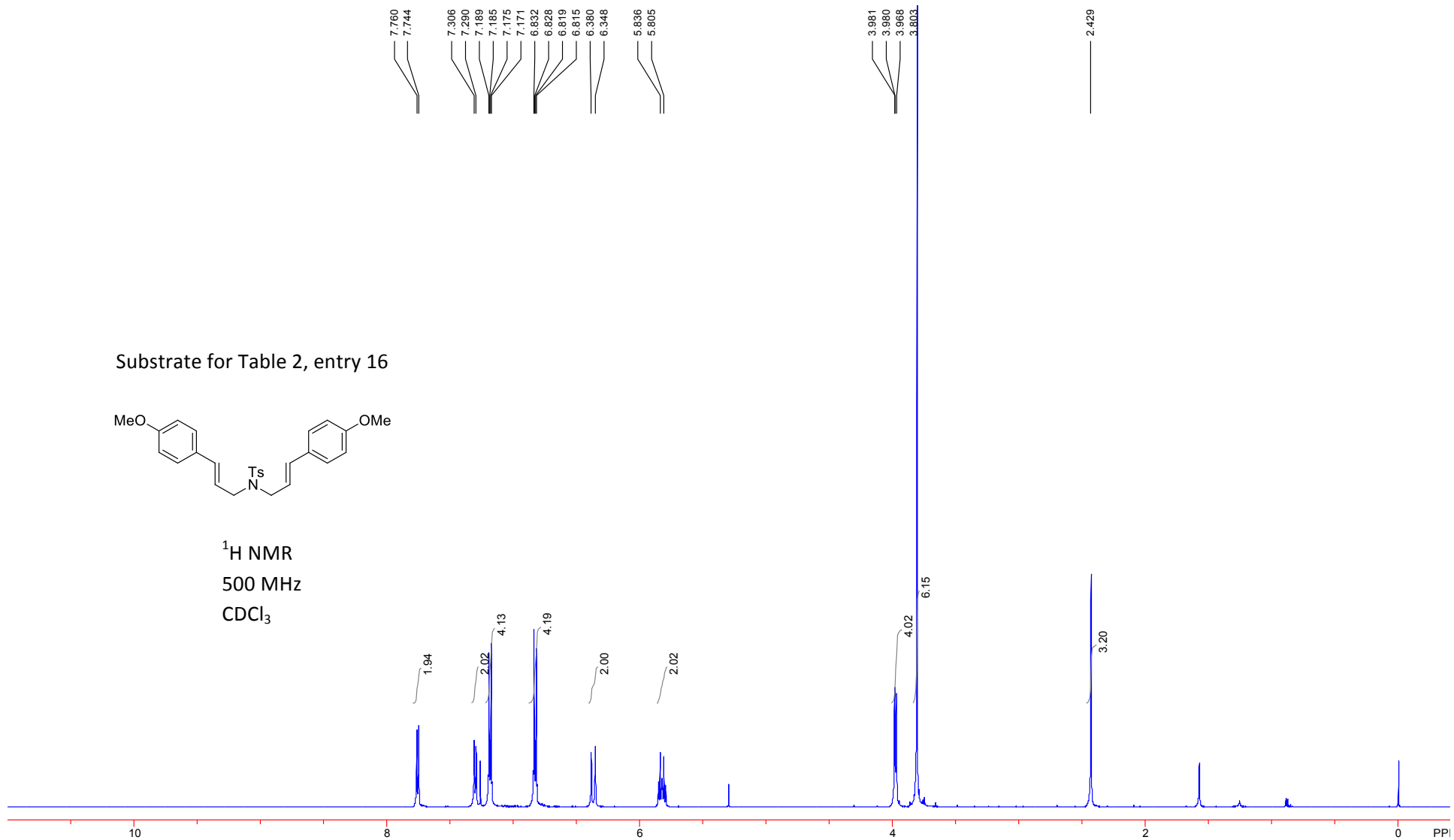
$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$



Substrate for Table 2, entry 16

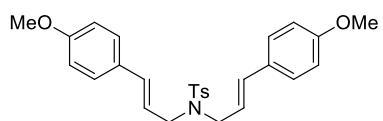


$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$

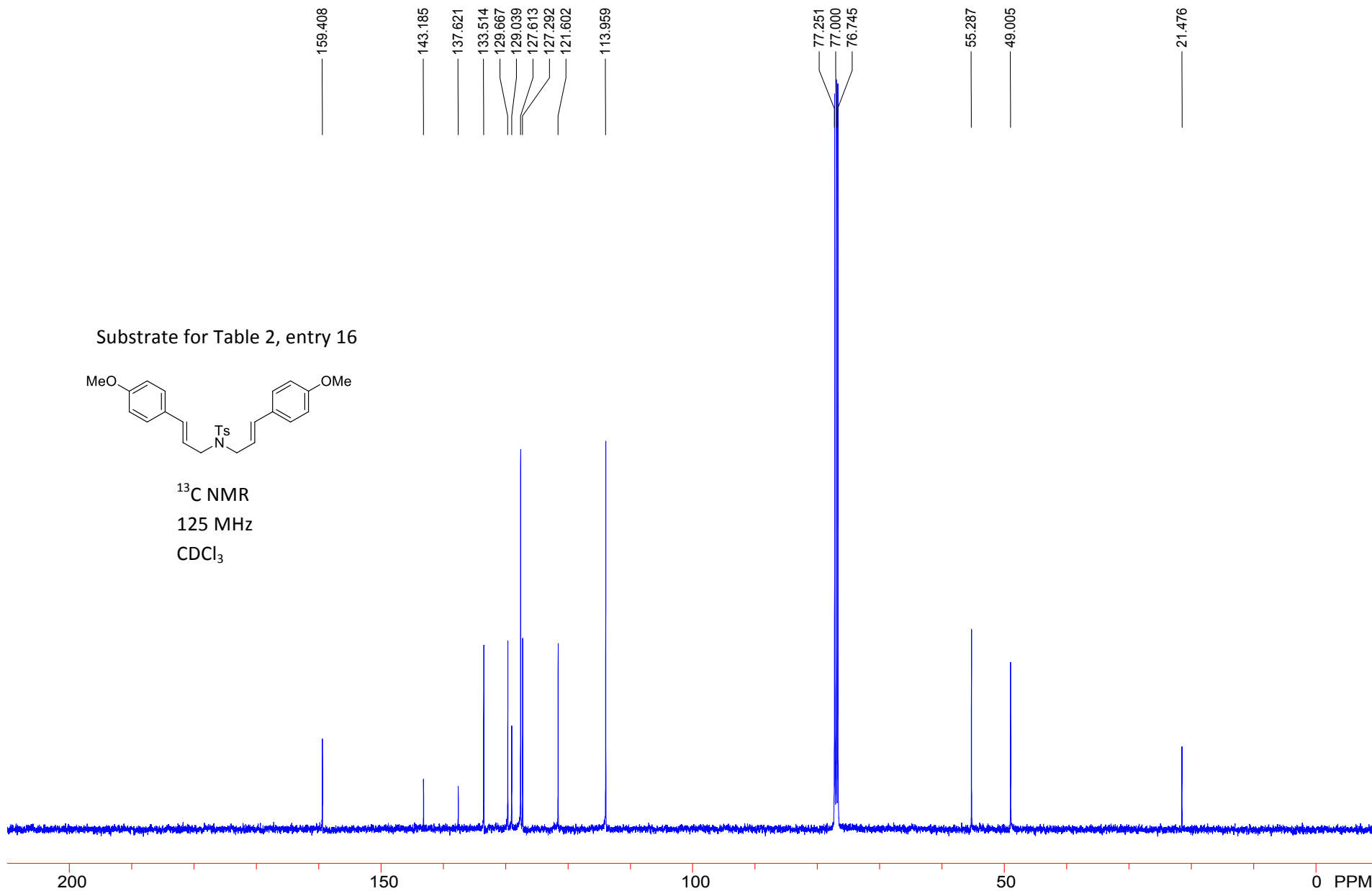




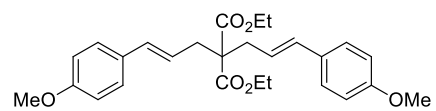
Substrate for Table 2, entry 16



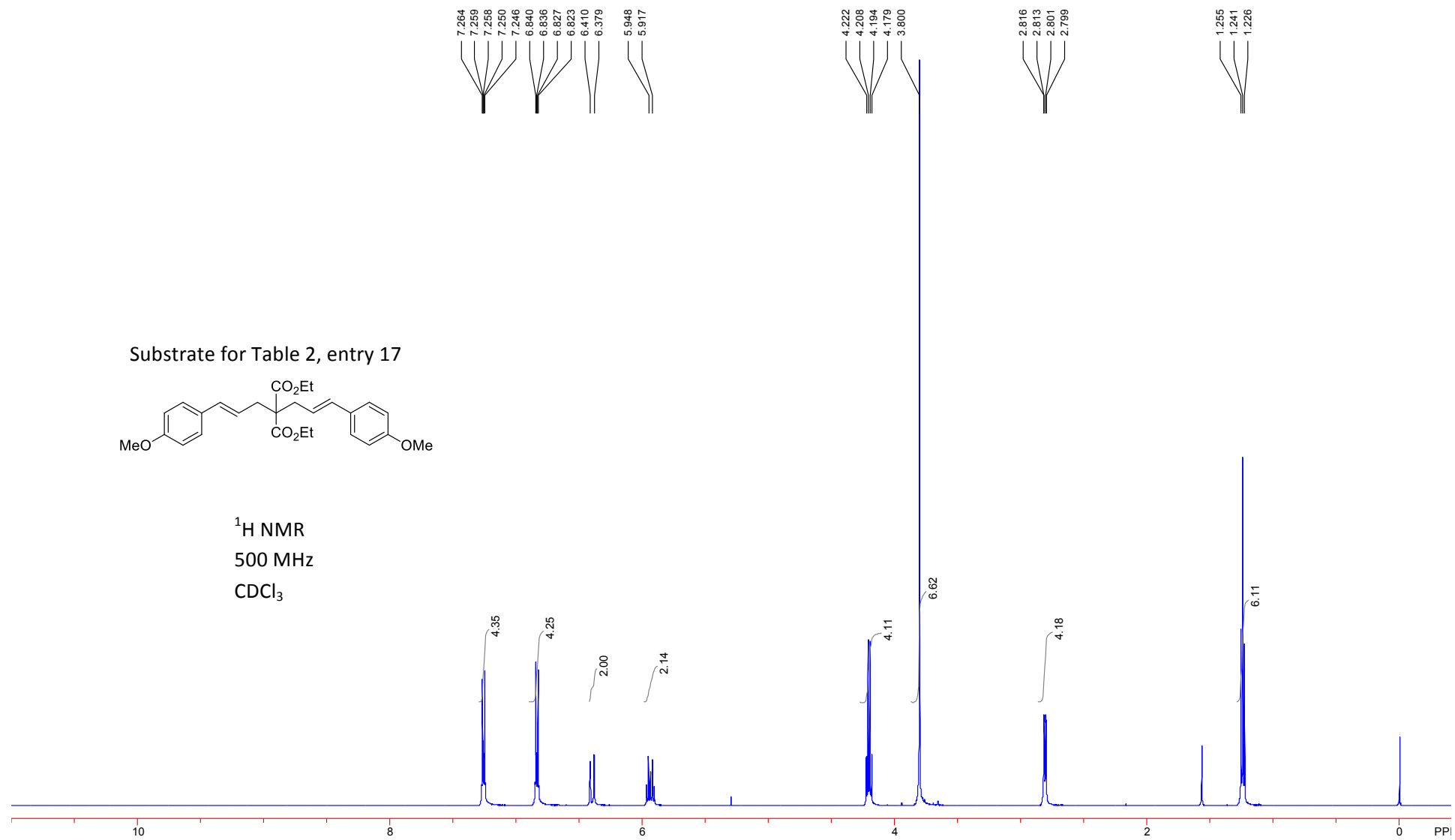
$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$

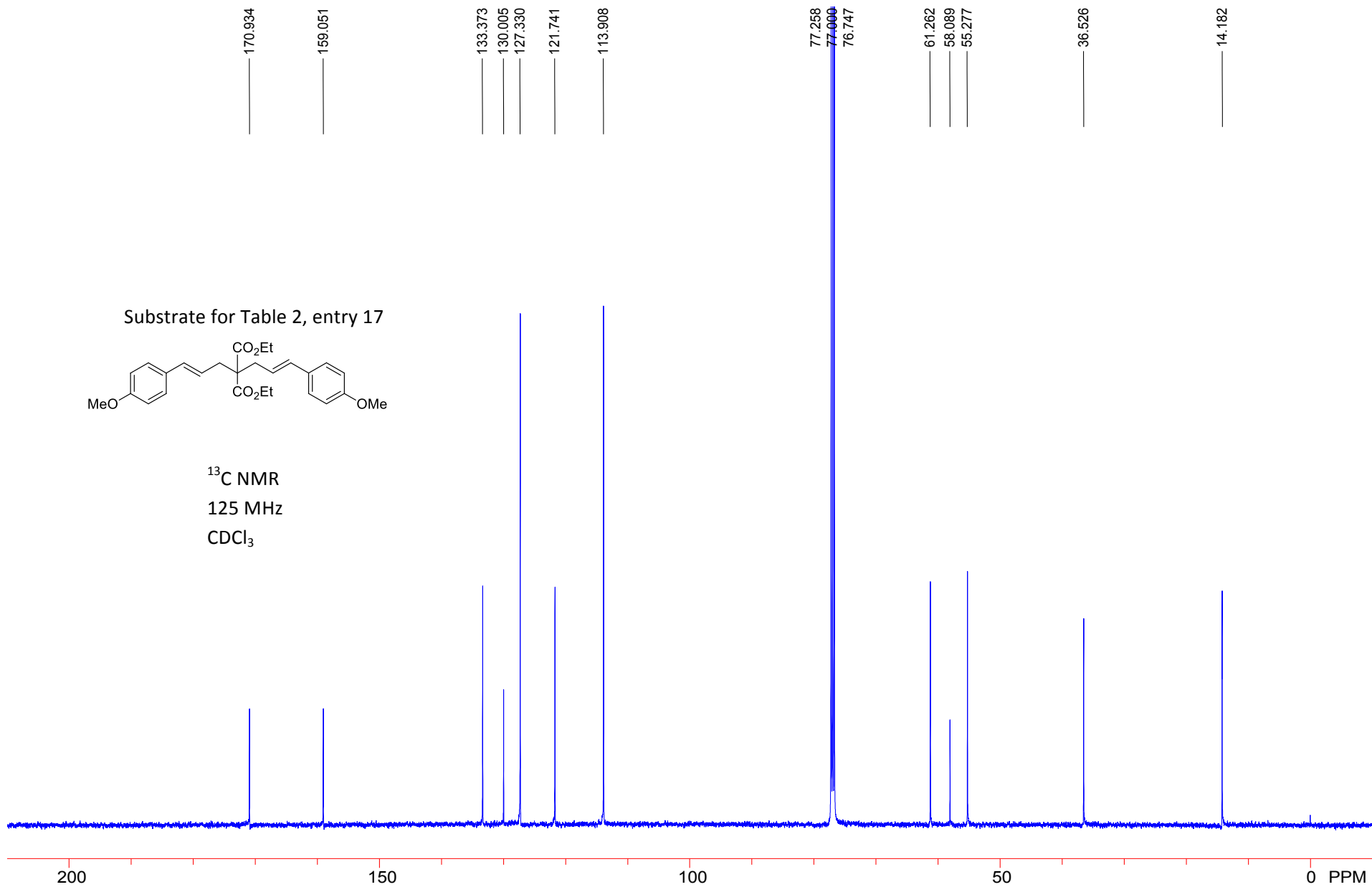


Substrate for Table 2, entry 17

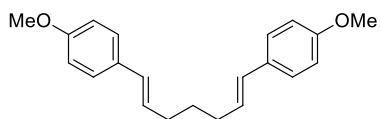


$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$

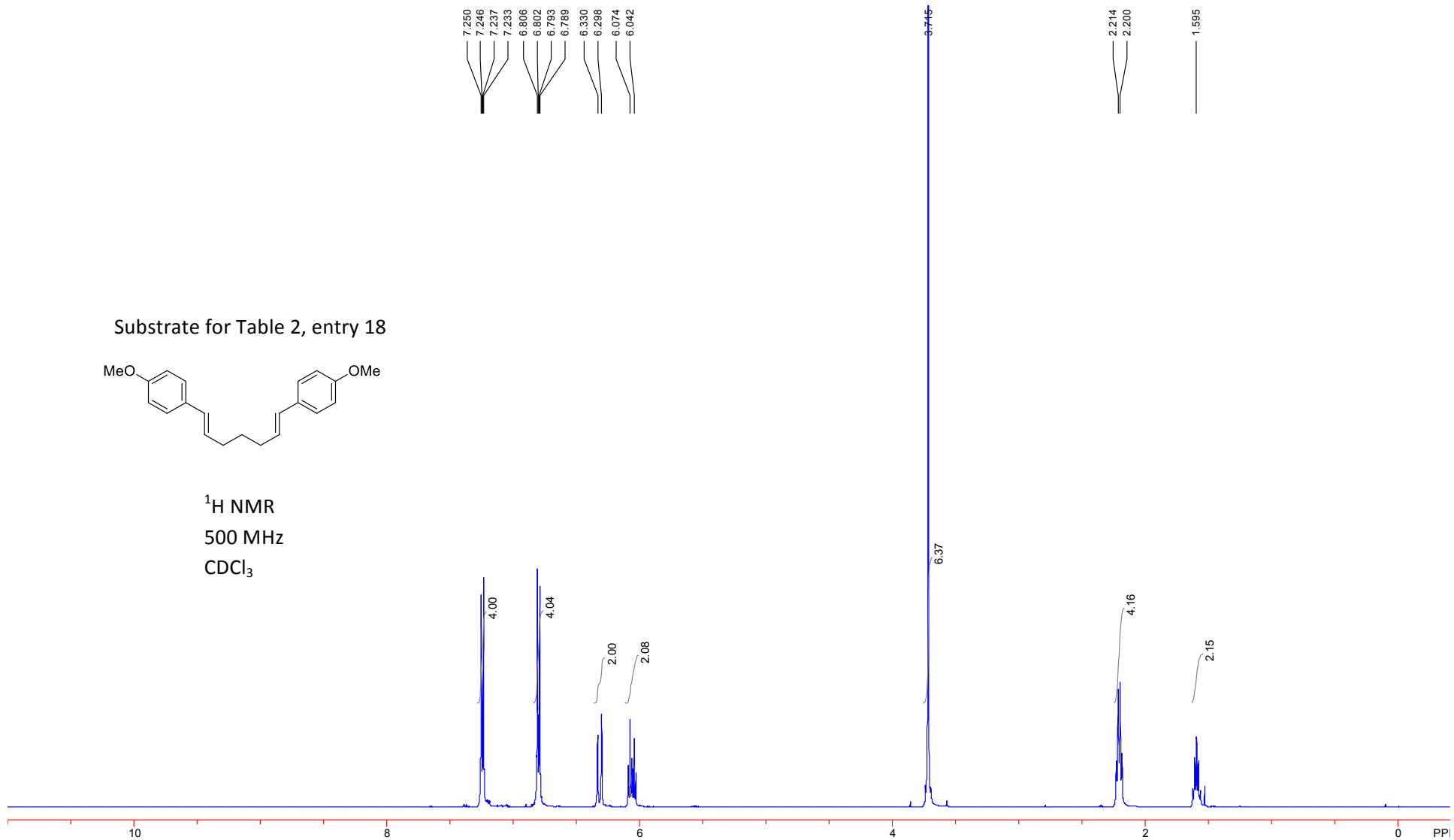




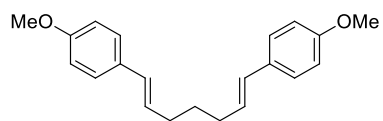
Substrate for Table 2, entry 18



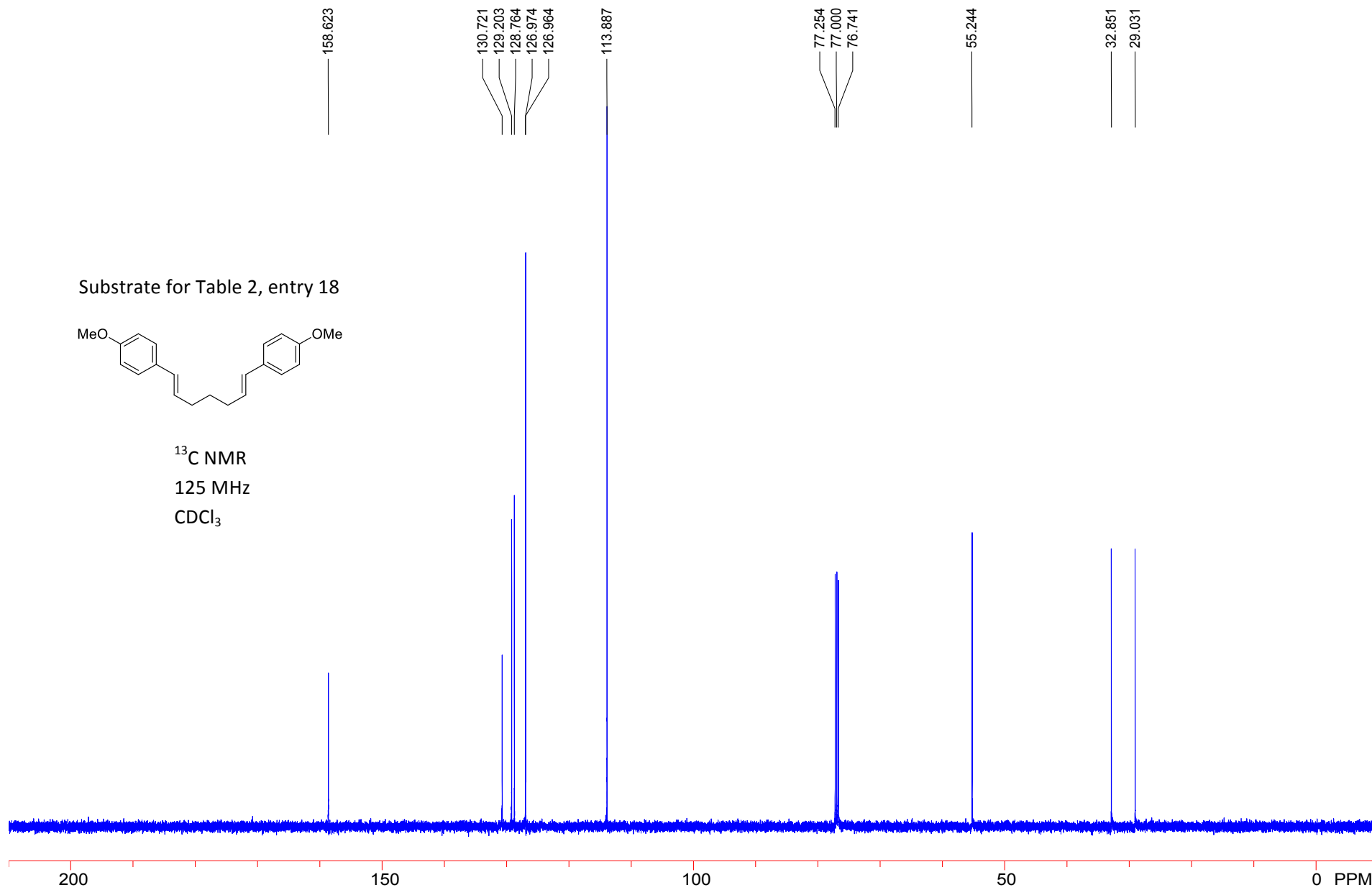
$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$



Substrate for Table 2, entry 18

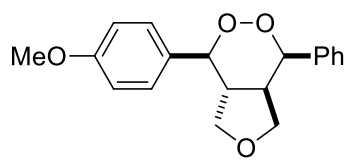


$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$

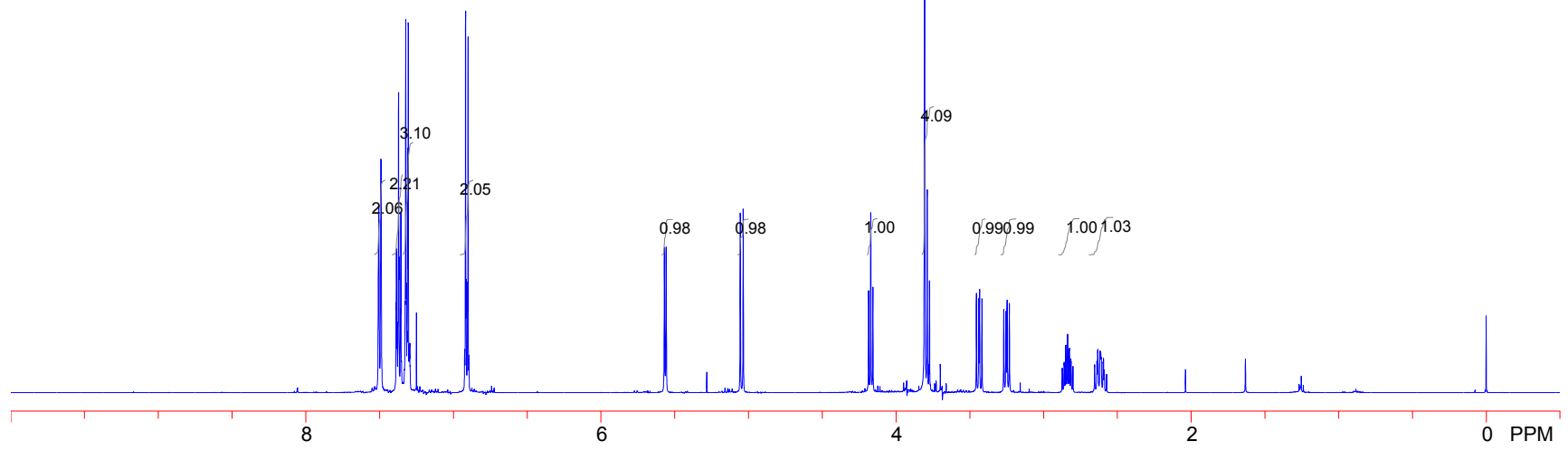


7.505  
7.491  
7.388  
7.386  
7.382  
7.371  
7.369  
7.359  
7.356  
7.352  
7.327  
7.323  
7.319  
7.314  
7.310  
7.306  
7.299  
7.296  
7.293  
7.250  
6.924  
6.918  
6.914  
6.904  
6.901  
6.894  
5.570  
5.559  
5.056  
5.036  
4.186  
4.172  
4.157  
3.807  
3.789  
3.778  
3.774  
3.456  
3.440  
3.435  
3.419  
3.270  
3.255  
3.248  
3.233  
2.862  
2.860  
2.850  
2.838  
2.827  
2.825  
2.816  
2.814  
2.802  
2.654  
2.639  
2.633  
2.618  
2.614  
2.610  
2.598  
2.595  
-0.000

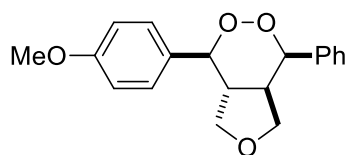
Major product for Table 2, entry 1



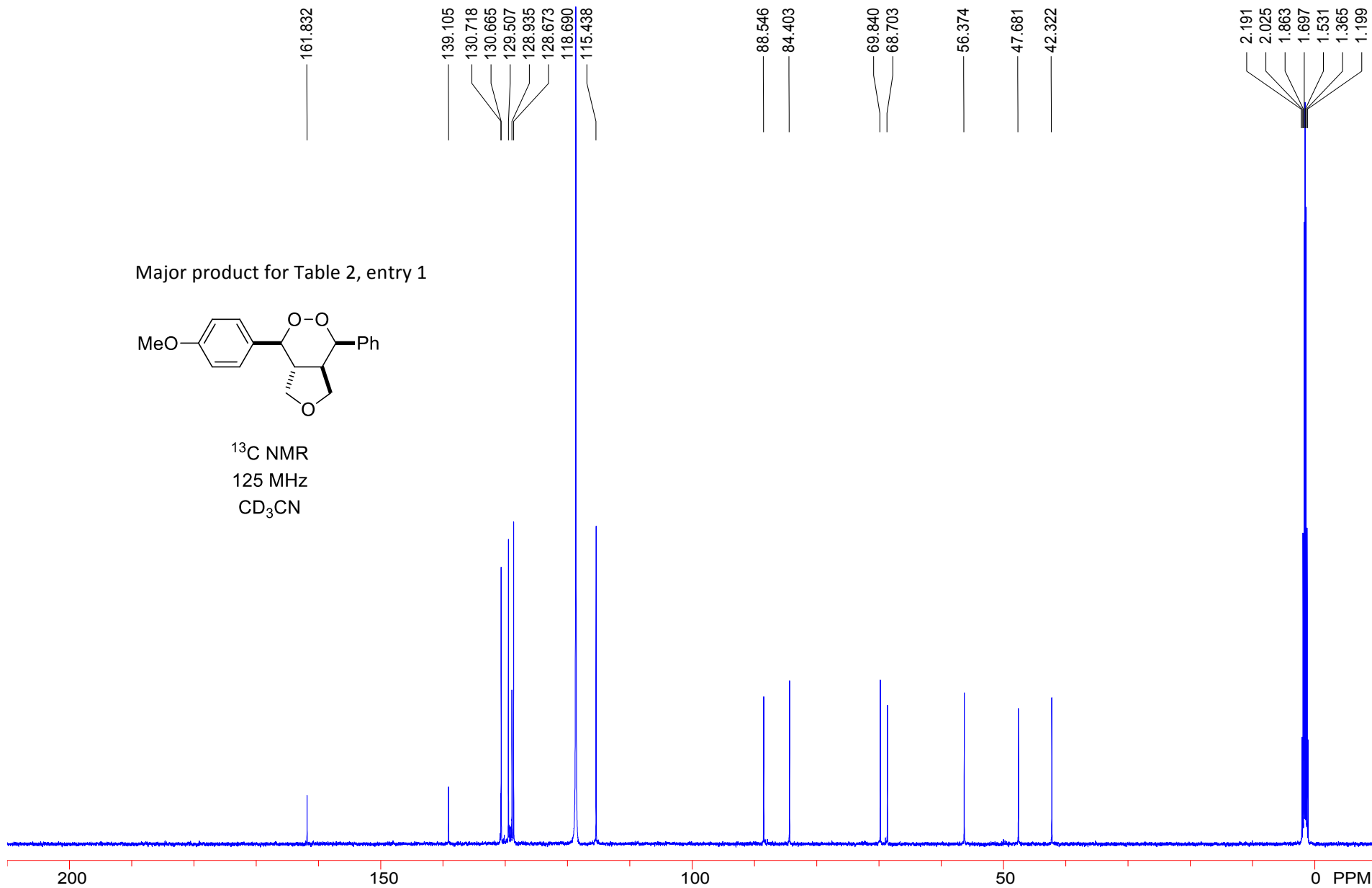
<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>



Major product for Table 2, entry 1

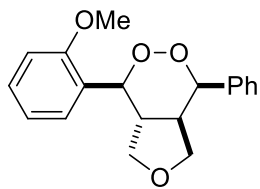


$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CD}_3\text{CN}$

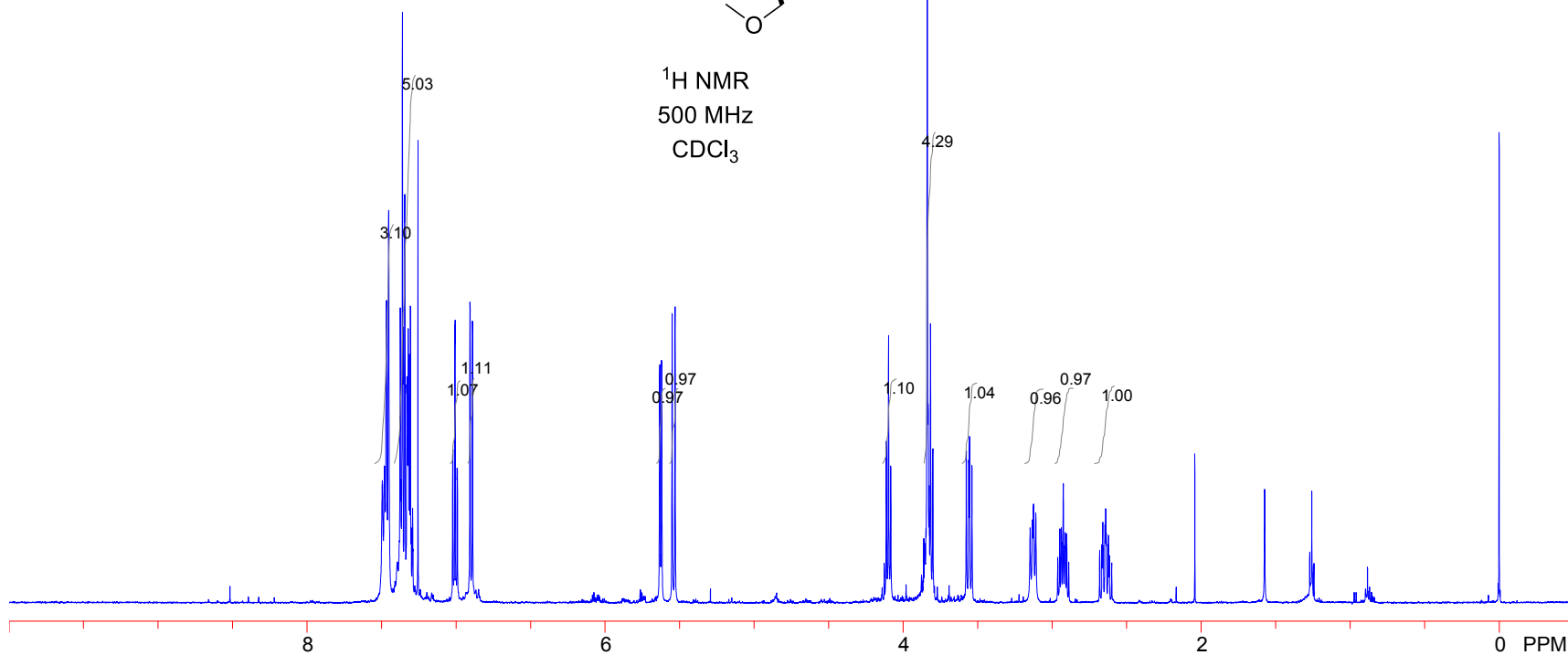


7.467  
7.452  
7.370  
7.360  
7.357  
7.344  
7.331  
7.329  
7.318  
7.314  
7.256  
7.023  
7.021  
7.008  
7.006  
6.993  
6.991  
6.907  
6.905  
6.890  
5.633  
5.620  
5.550  
5.531  
4.113  
4.098  
4.084  
3.839  
3.817  
3.801  
3.576  
3.560  
3.555  
3.539  
3.148  
3.132  
3.126  
3.111  
2.949  
2.939  
2.936  
2.926  
2.916  
2.912  
2.902  
2.681  
2.666  
2.660  
2.657  
2.646  
2.641  
2.637  
2.625  
2.621

Major product for Table 2, entry 3

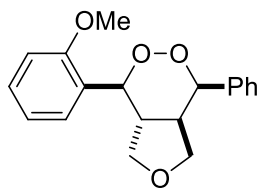


<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>

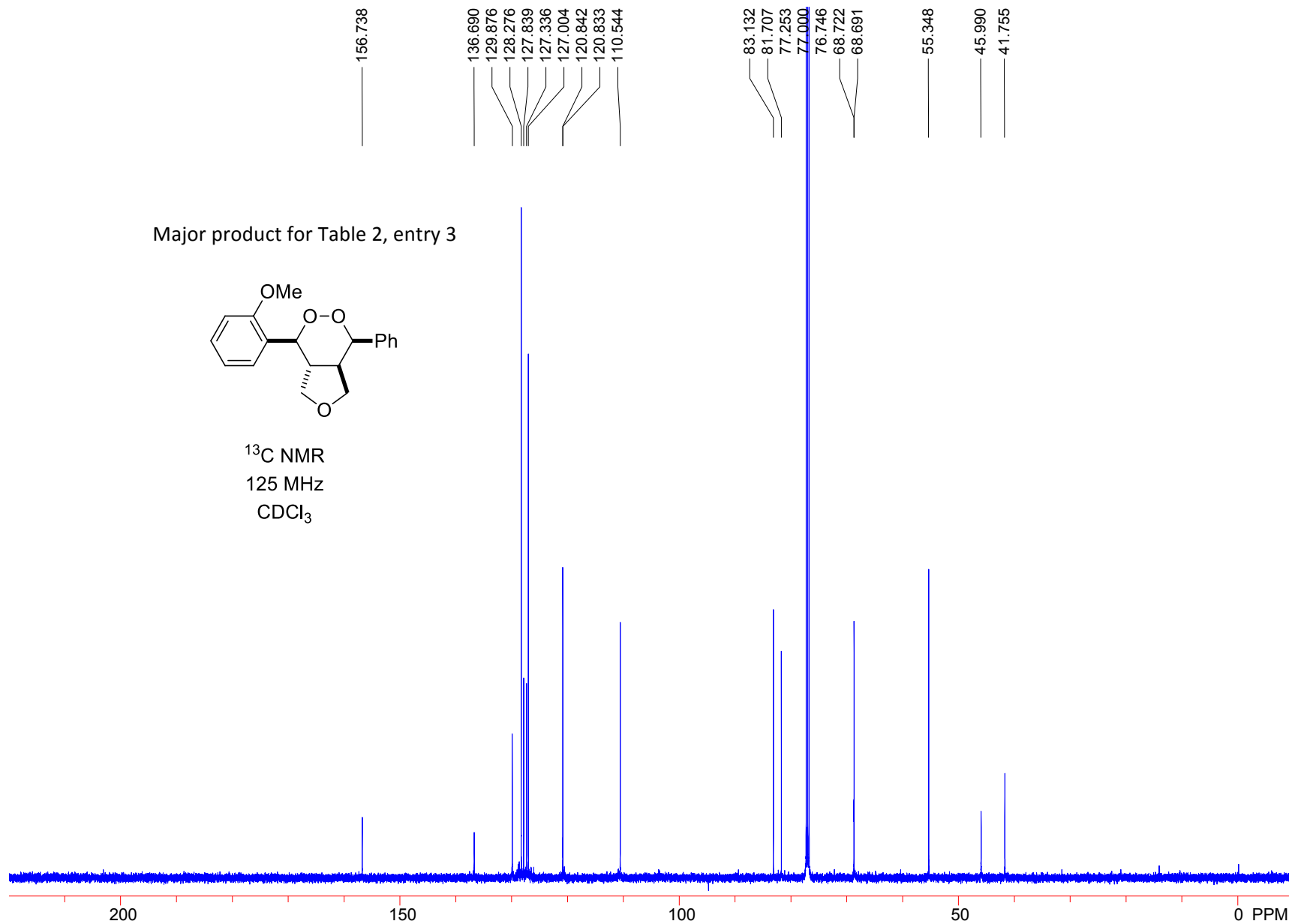


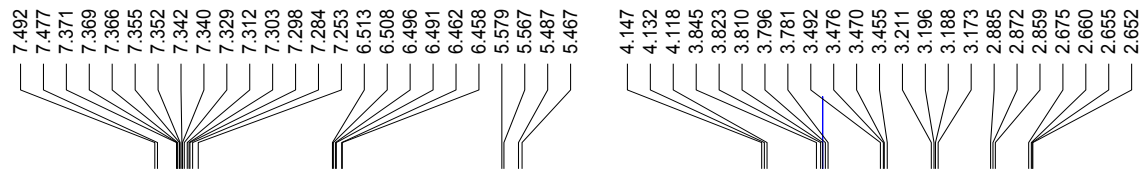


Major product for Table 2, entry 3

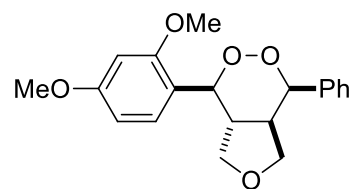


$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$

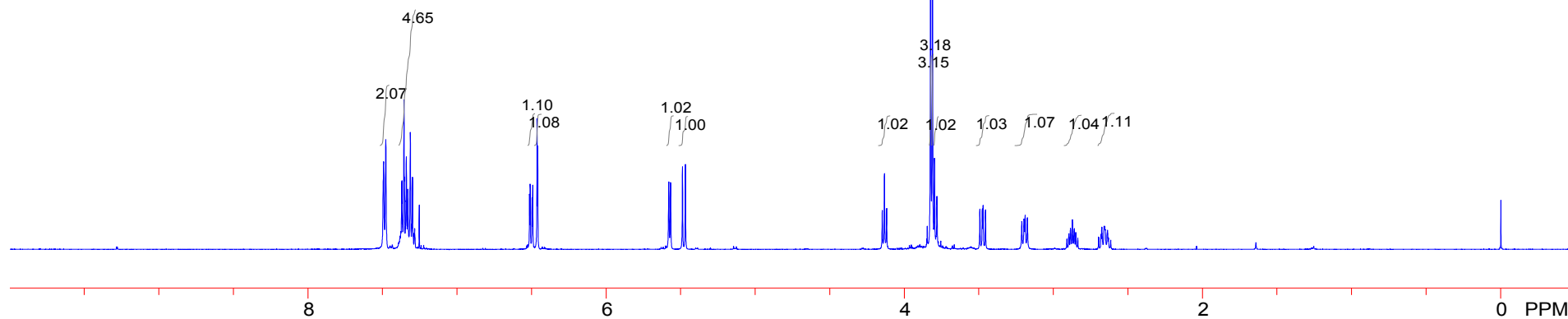


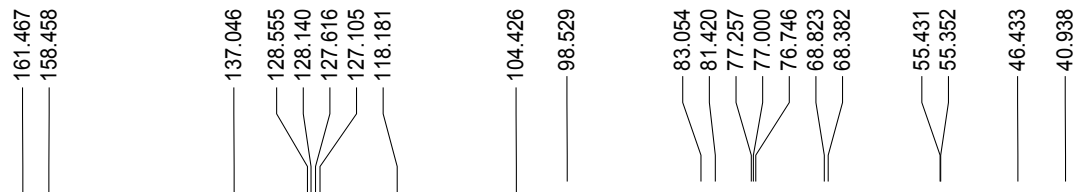


Major product for Table 2, entry 5

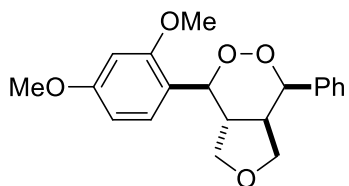


<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>

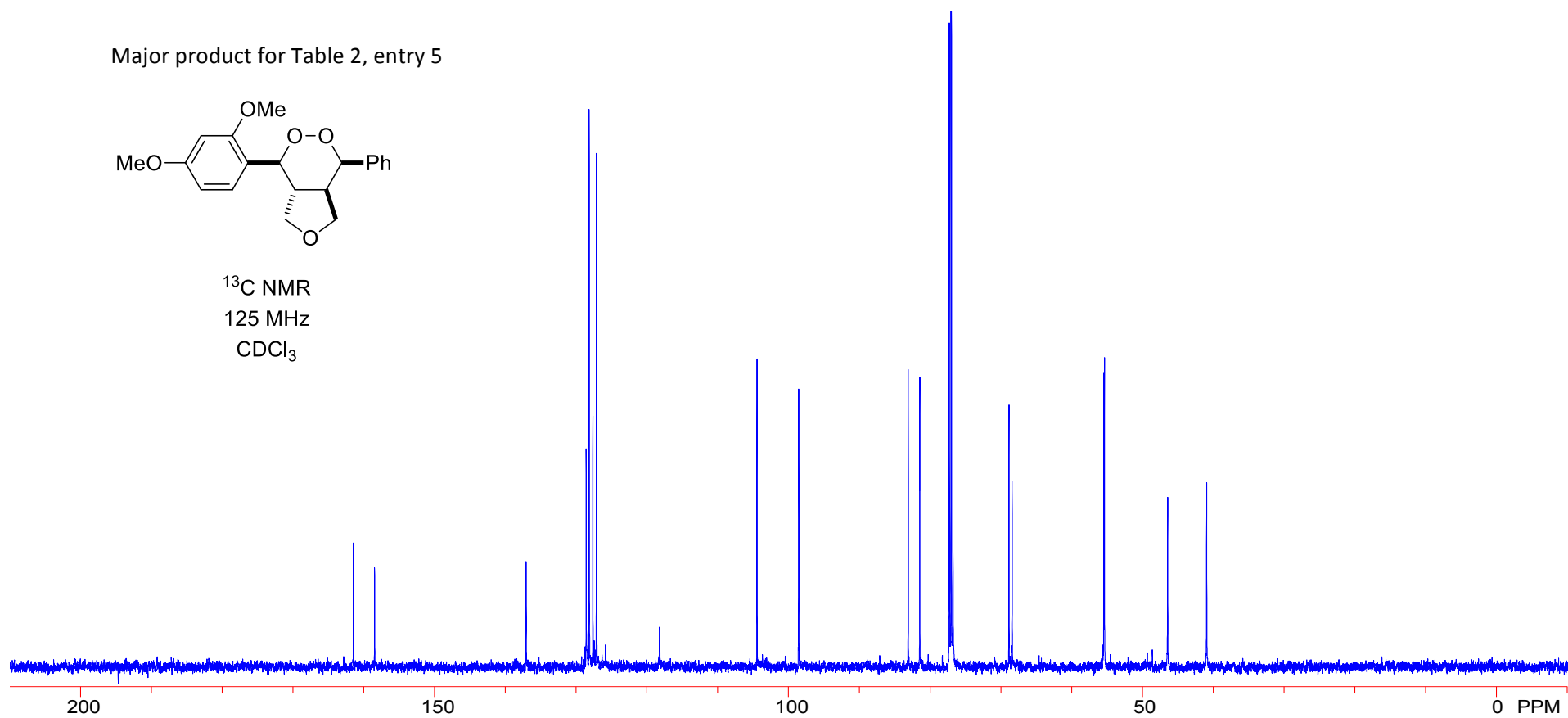




Major product for Table 2, entry 5



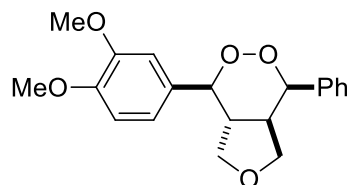
$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$



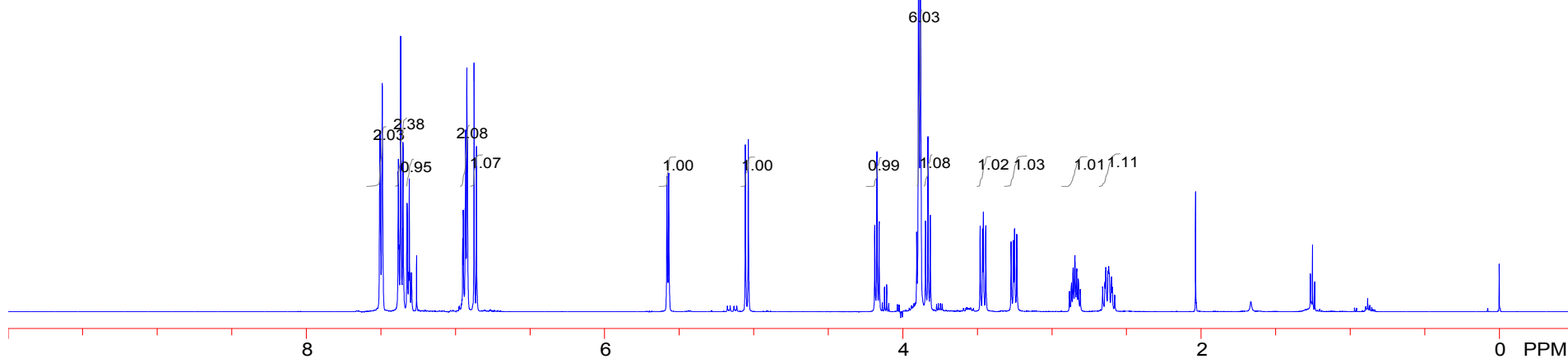
7.506  
7.491  
7.383  
7.382  
7.379  
7.368  
7.355  
7.352  
7.327  
7.324  
7.322  
6.951  
6.948  
6.935  
6.931  
6.924  
6.920  
6.875  
6.859  
5.582  
5.570  
5.056  
5.037  
4.189  
4.174  
4.159  
3.907  
3.883  
3.832  
3.817  
3.482  
3.466  
3.460  
3.444  
3.275  
3.259  
3.252  
3.237  
2.859  
2.847  
2.833  
2.824  
2.645  
2.640  
2.625  
2.620  
2.617  
2.601

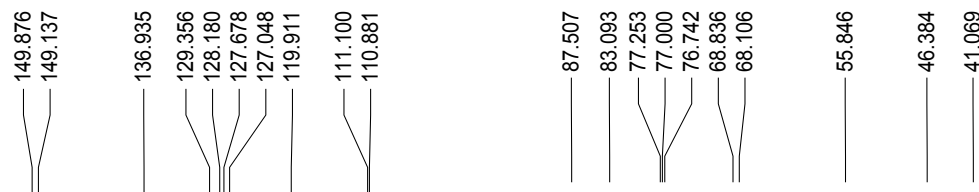
-0.000

Major product for Table 2, entry 6

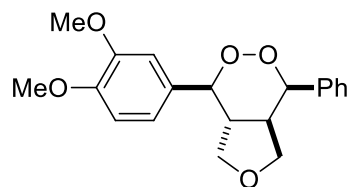


<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>

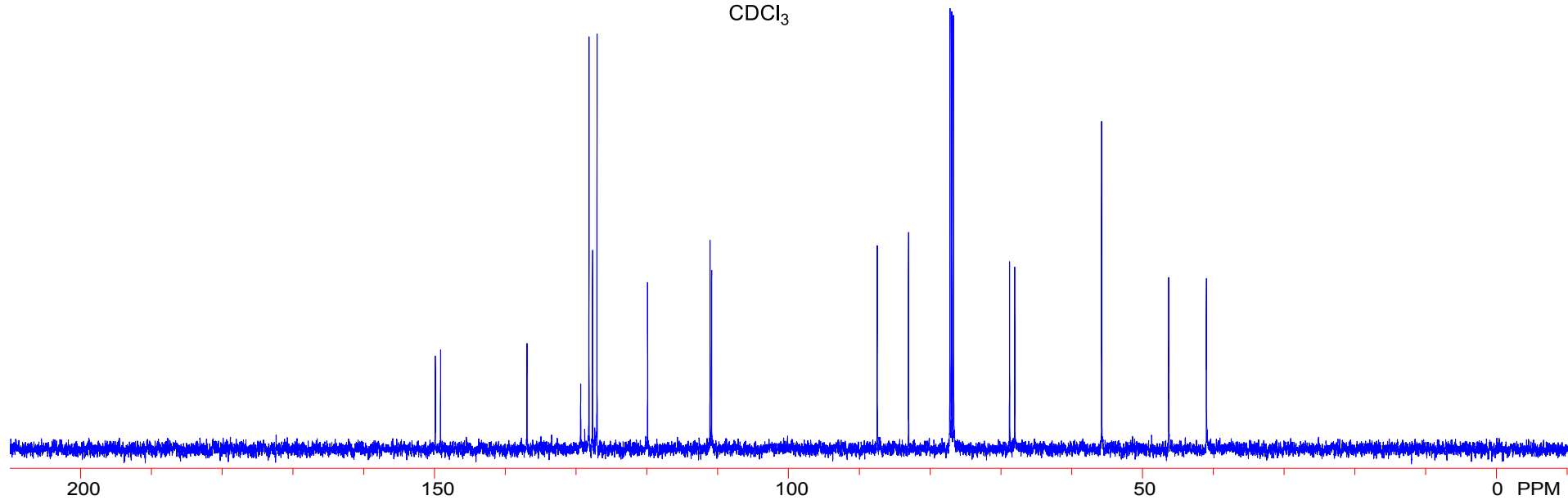


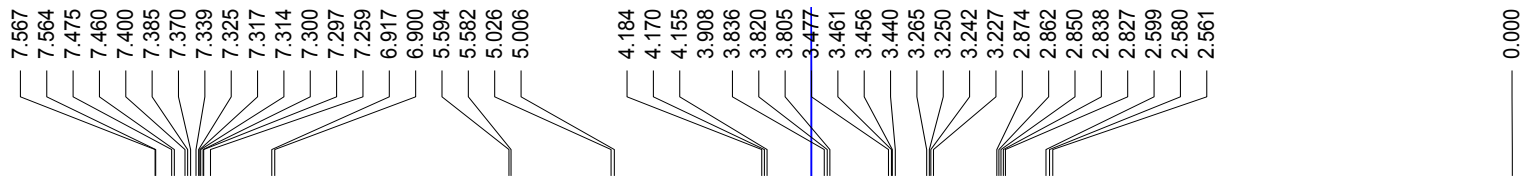


Major product for Table 2, entry 6

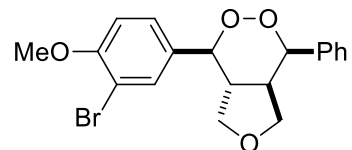


$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$

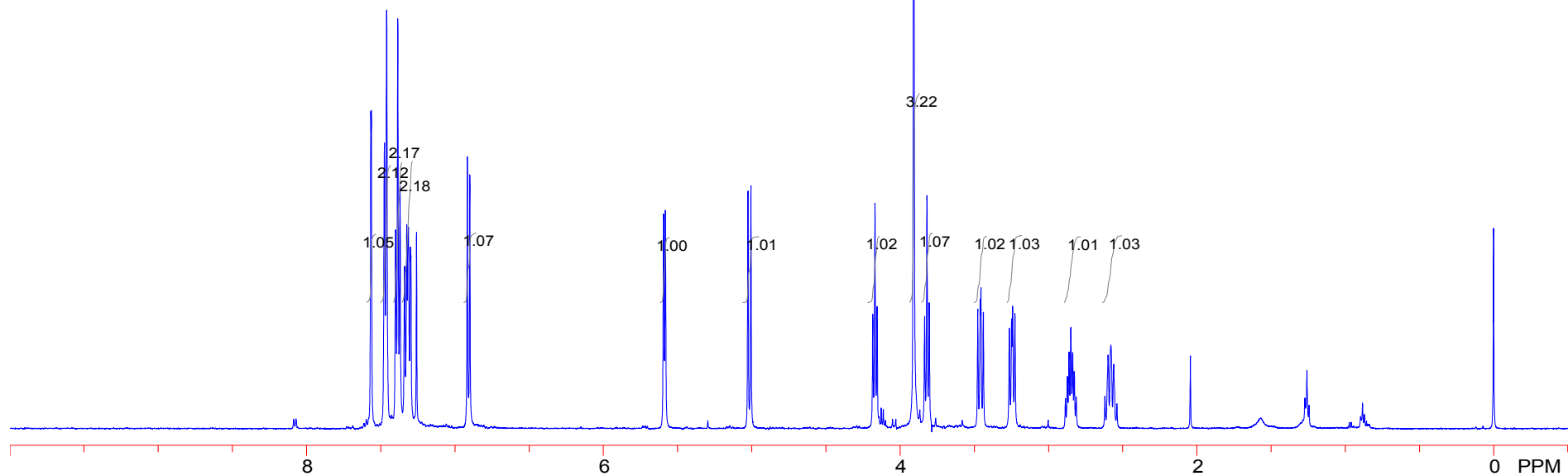




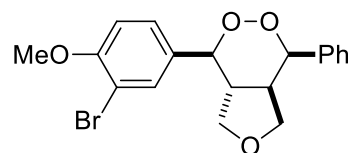
Major product for Table 2, entry 7



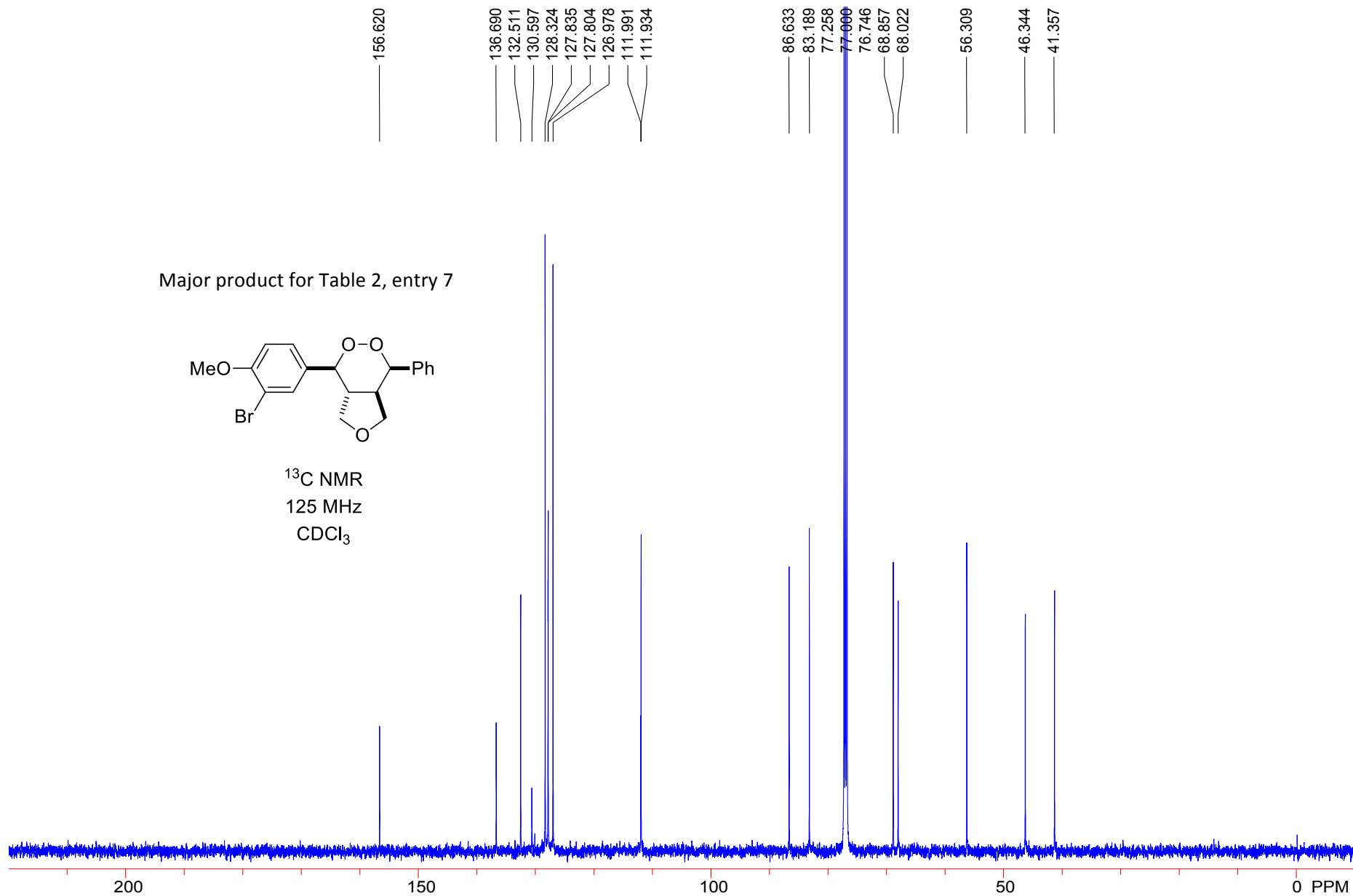
<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>



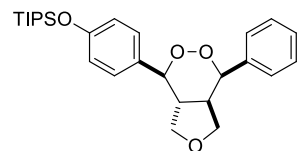
Major product for Table 2, entry 7



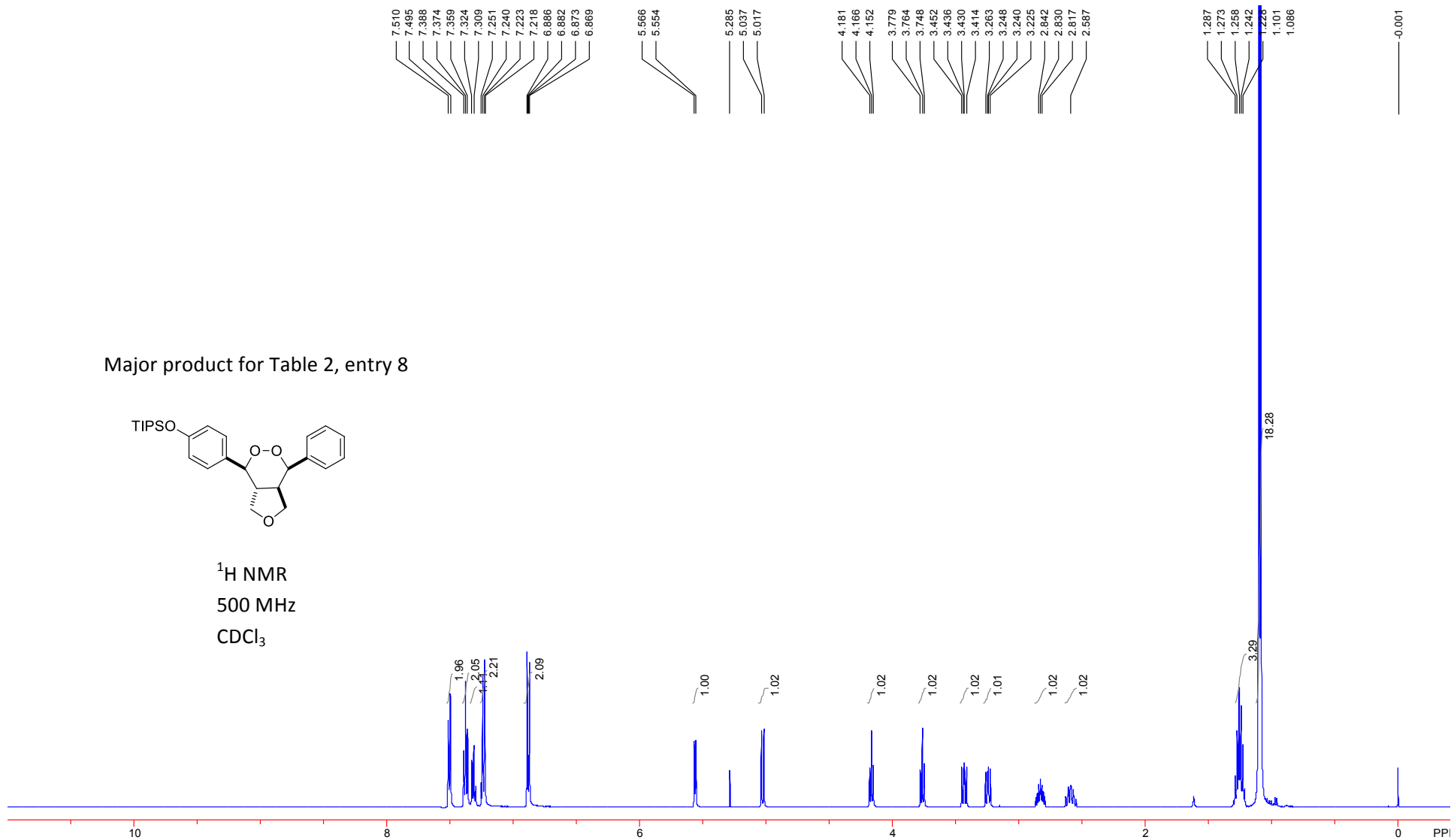
$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$



Major product for Table 2, entry 8

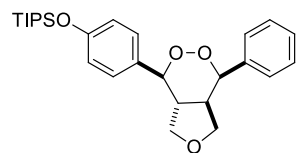


$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$

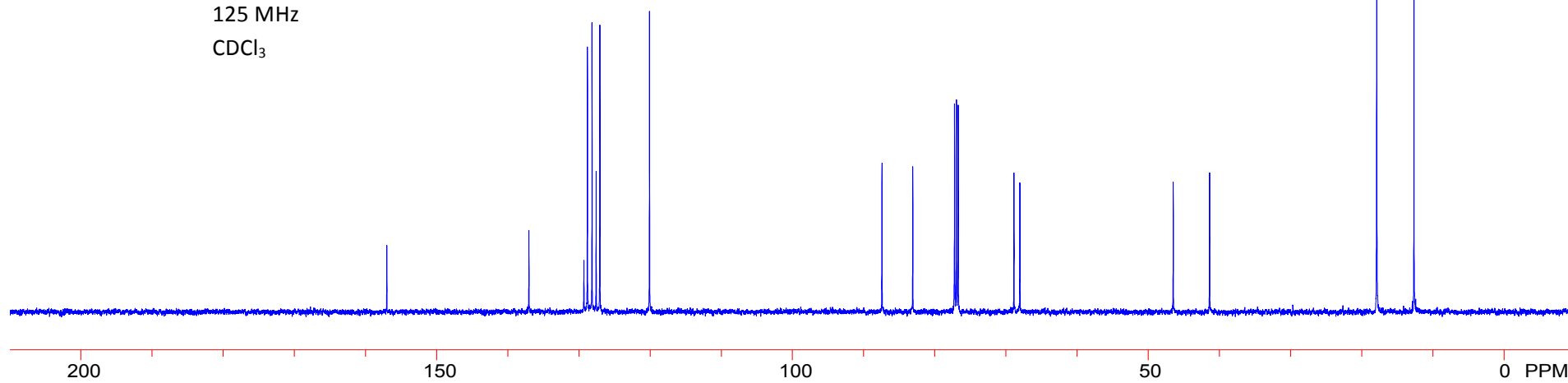


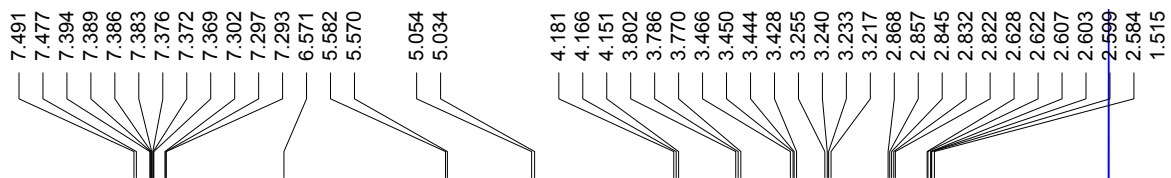


Major product for Table 2, entry 8

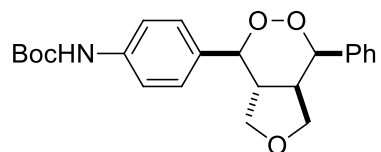


$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$

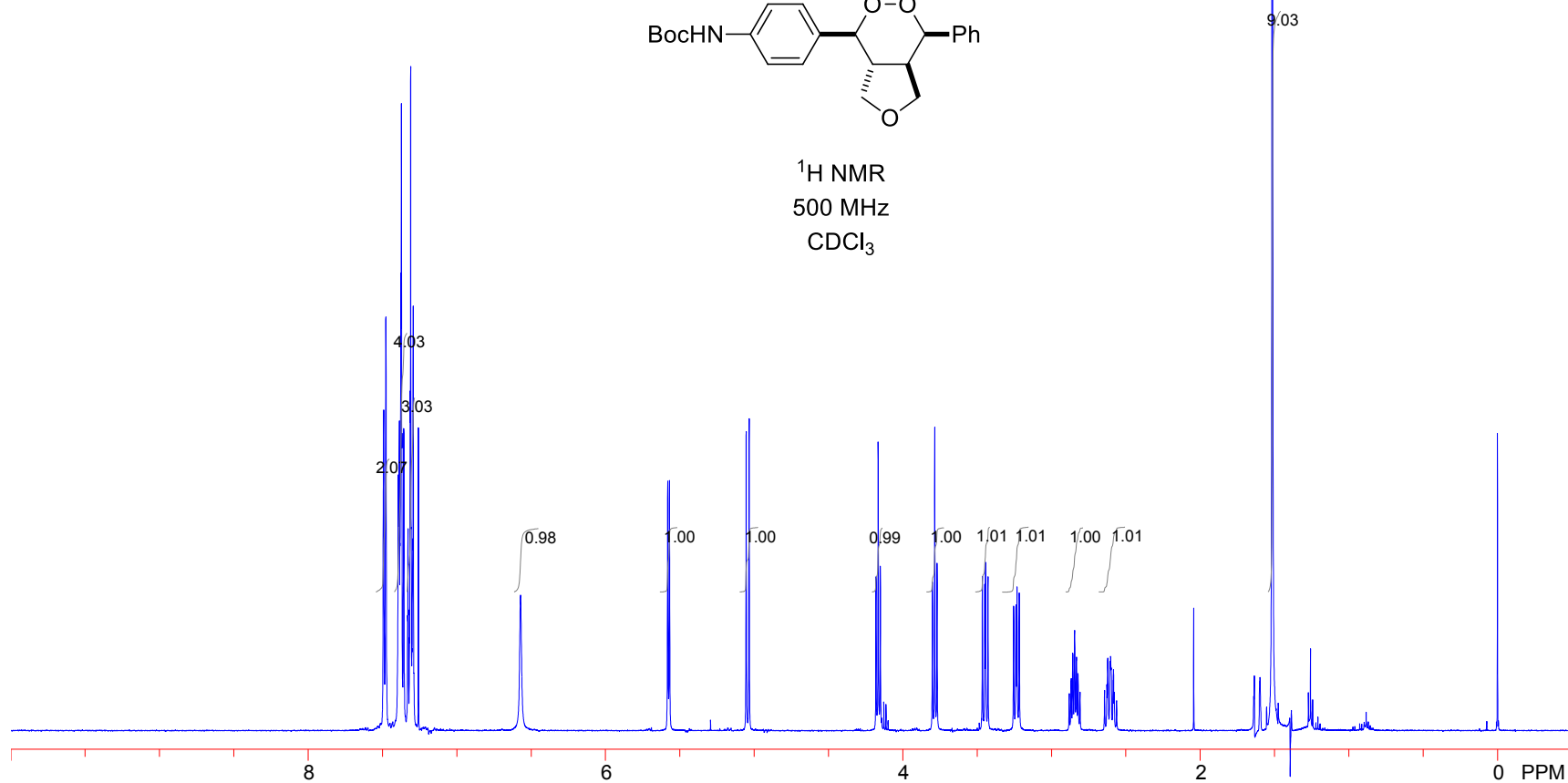




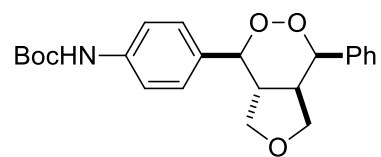
Major product for Table 2, entry 9



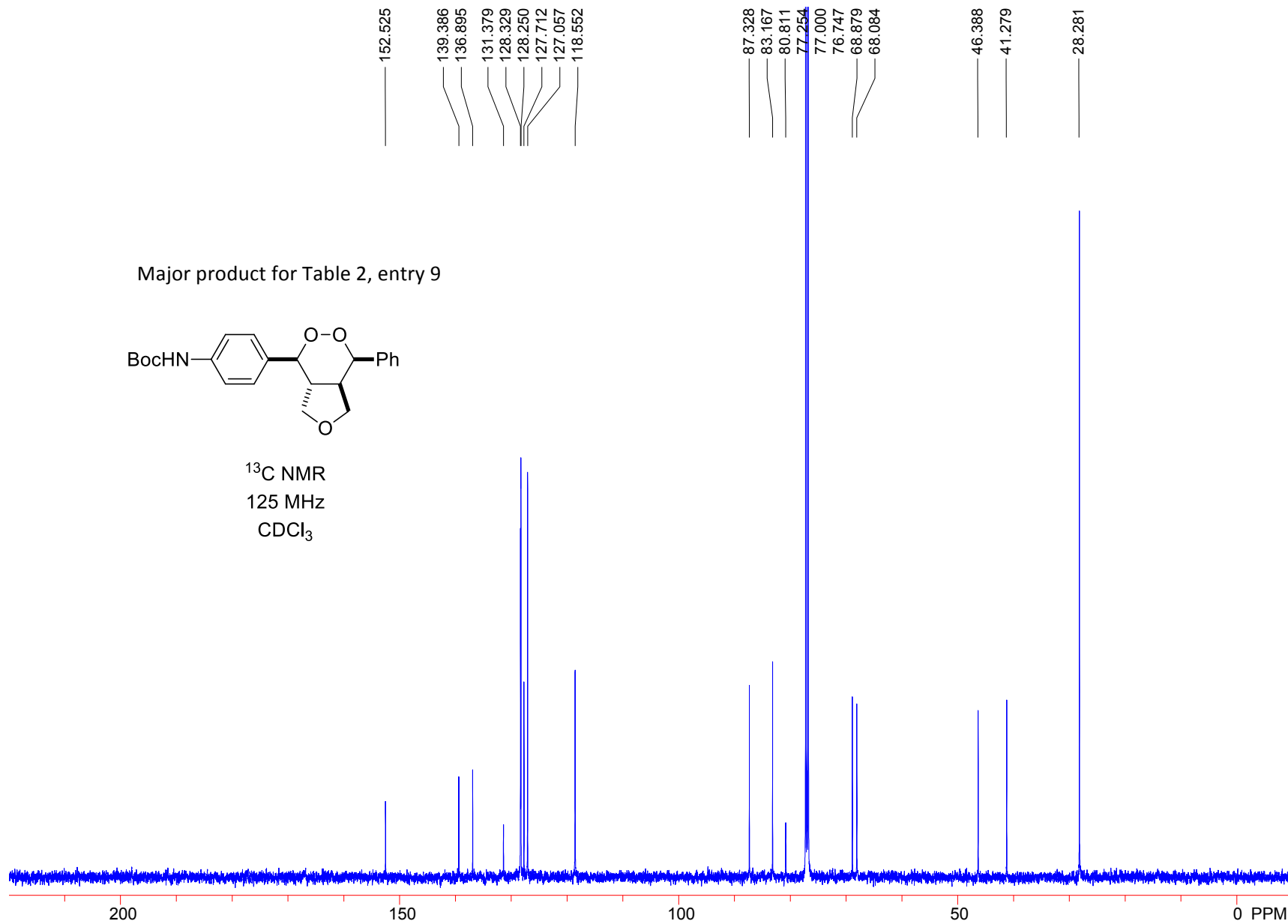
<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>



Major product for Table 2, entry 9



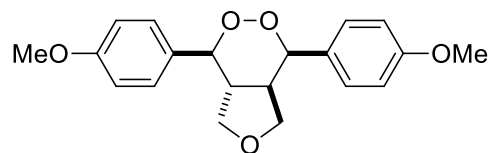
$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$



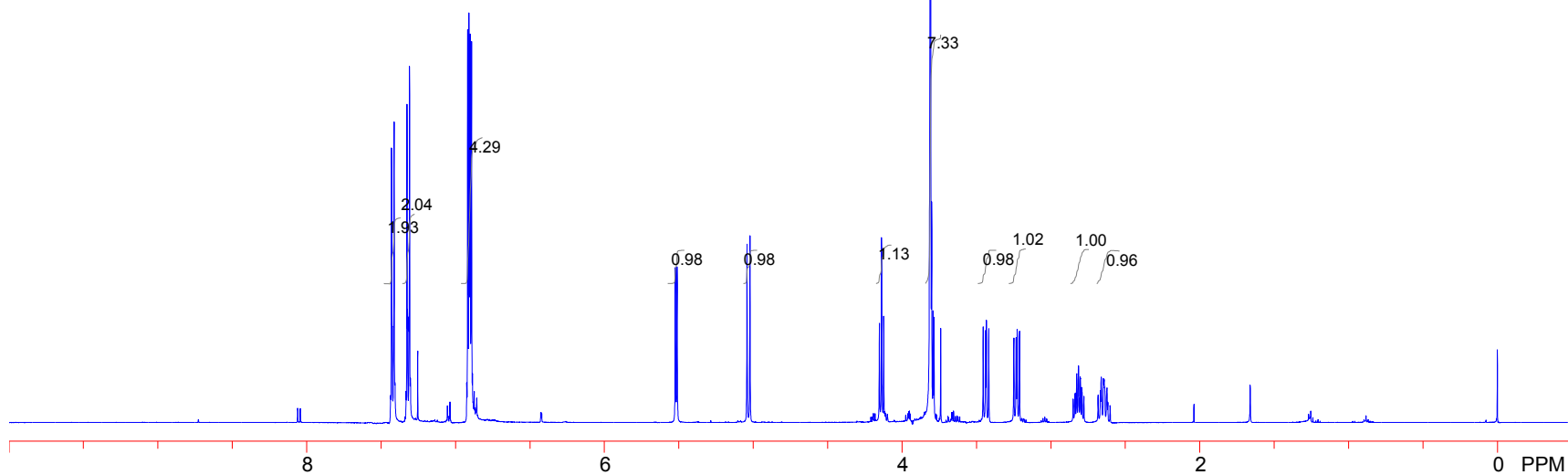
7.430  
7.426  
7.416  
7.413  
7.407  
7.326  
7.322  
7.313  
7.309  
7.303  
6.924  
6.919  
6.915  
6.910  
6.905  
6.901  
6.896  
6.892  
6.886  
5.523  
5.512  
5.042  
5.022  
4.153  
4.138  
4.123  
3.813  
3.808  
3.802  
3.792  
3.786  
3.455  
3.440  
3.434  
3.418  
3.250  
3.234  
3.227  
3.212  
2.827  
2.815  
2.803  
2.802  
2.792  
2.662  
2.647  
2.642  
2.639  
2.624

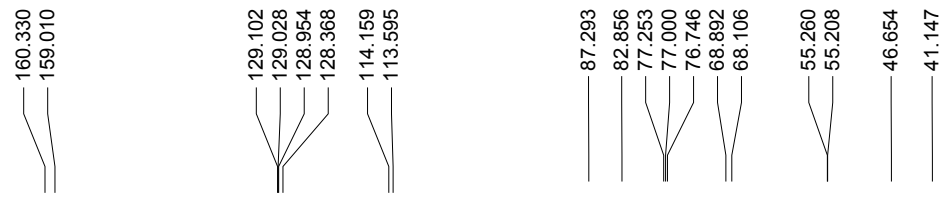
0.000

Major product for Table 2, entry 11

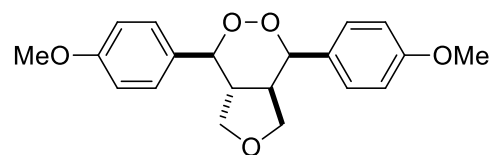


<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>

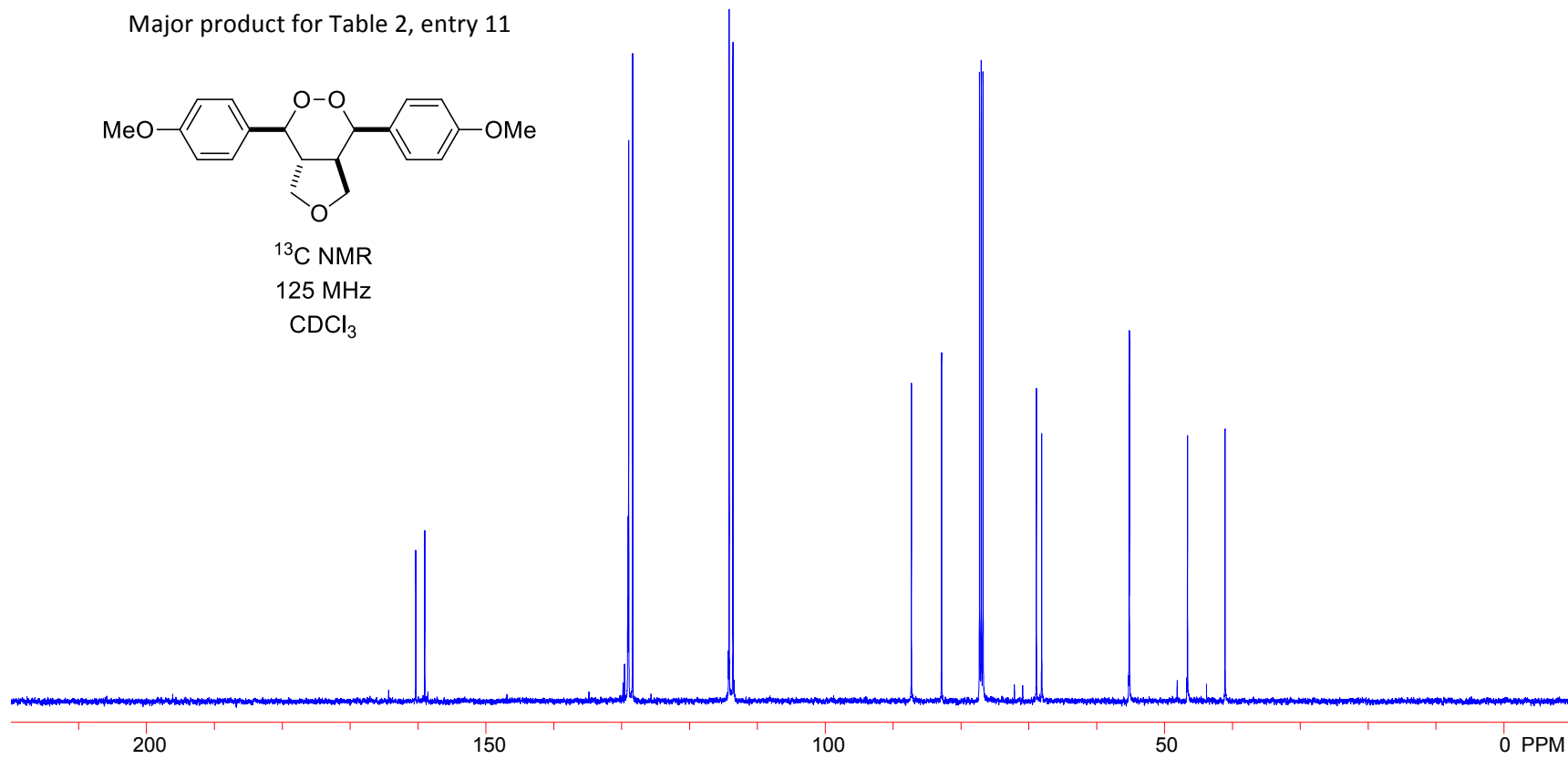




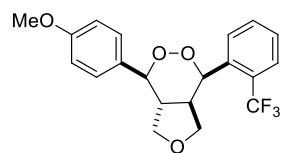
Major product for Table 2, entry 11



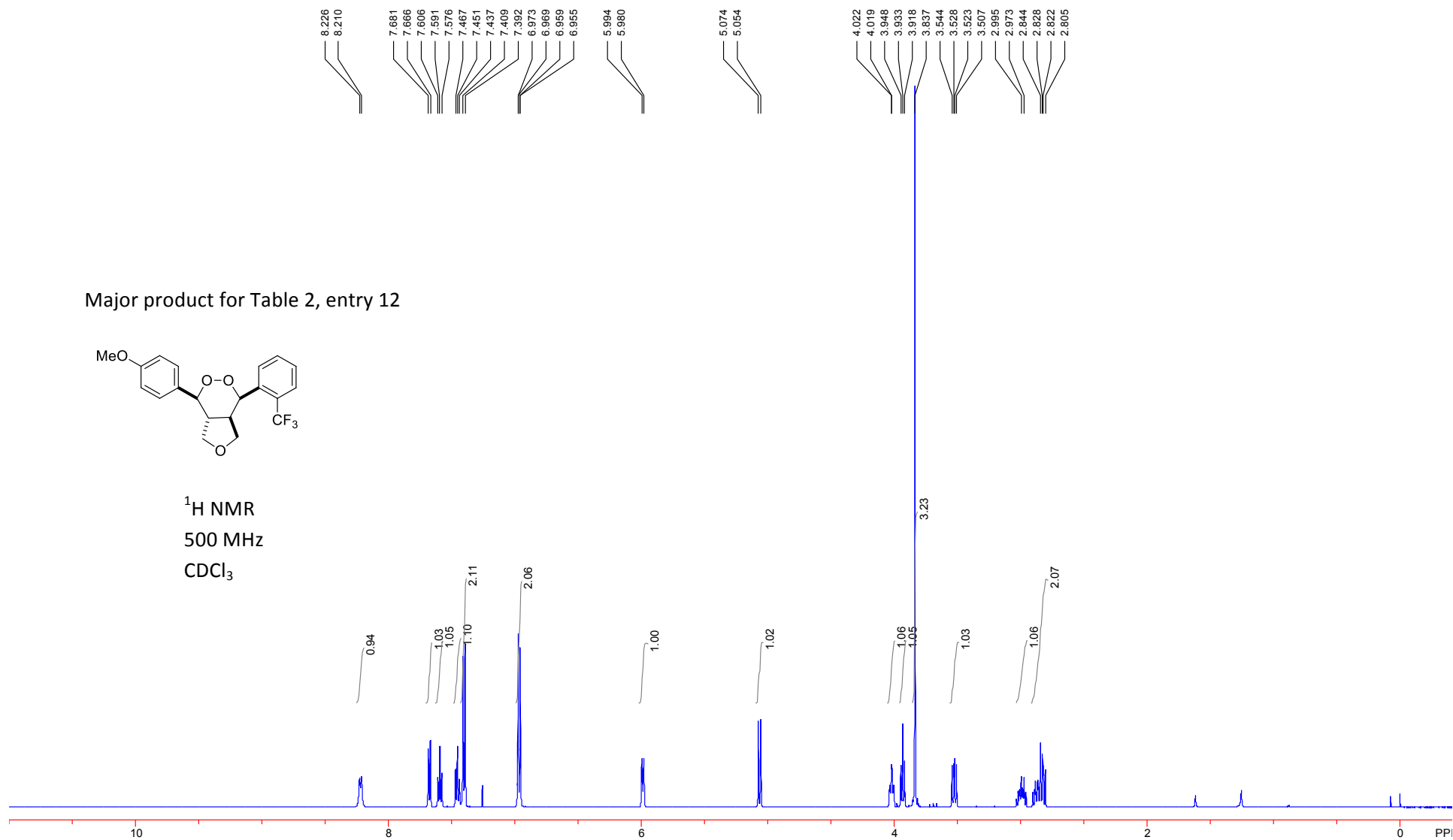
<sup>13</sup>C NMR  
125 MHz  
CDCl<sub>3</sub>



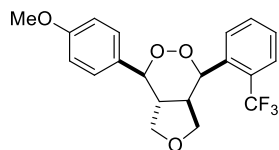
Major product for Table 2, entry 12



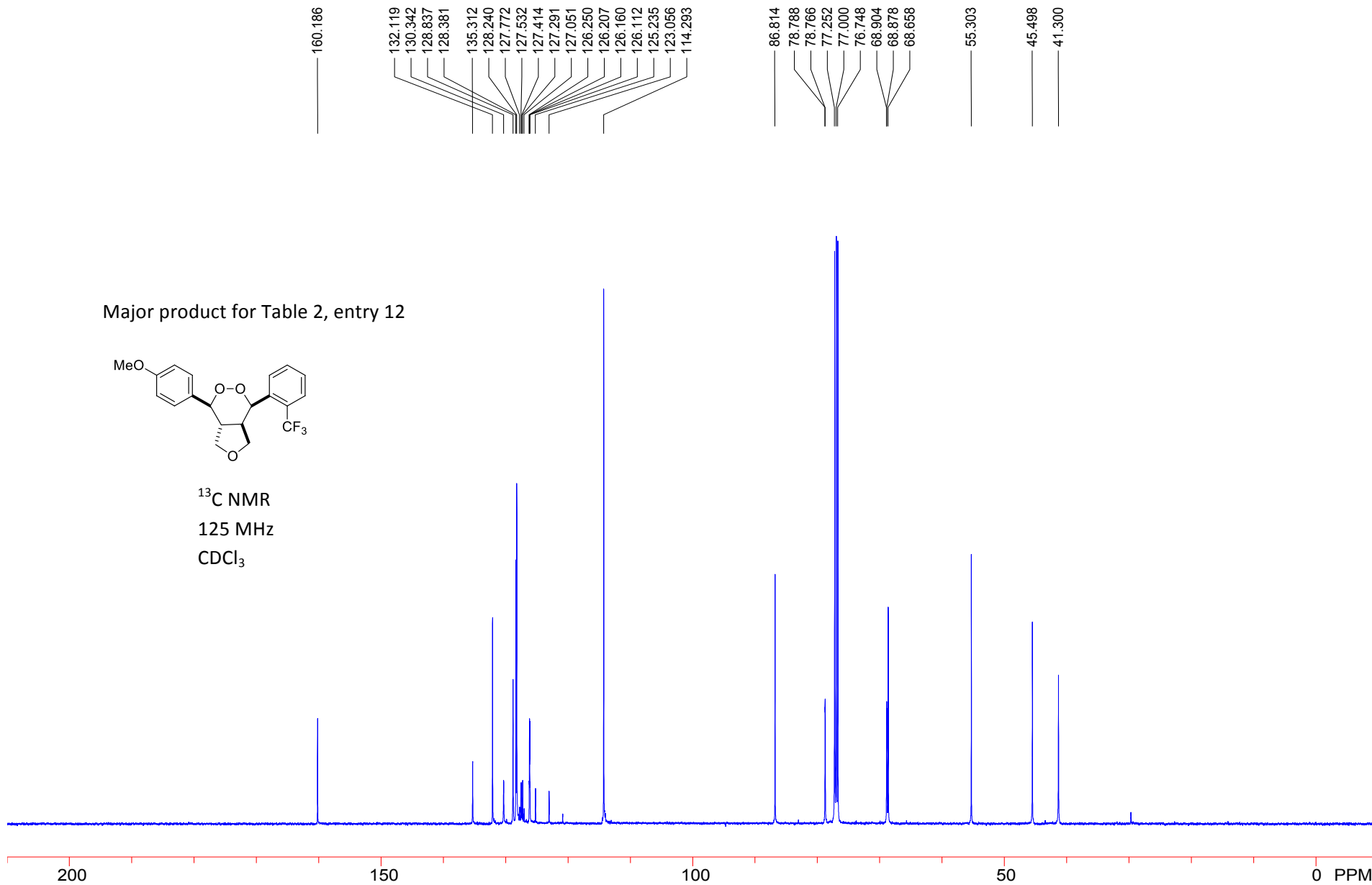
$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$



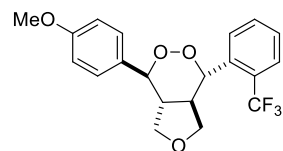
Major product for Table 2, entry 12



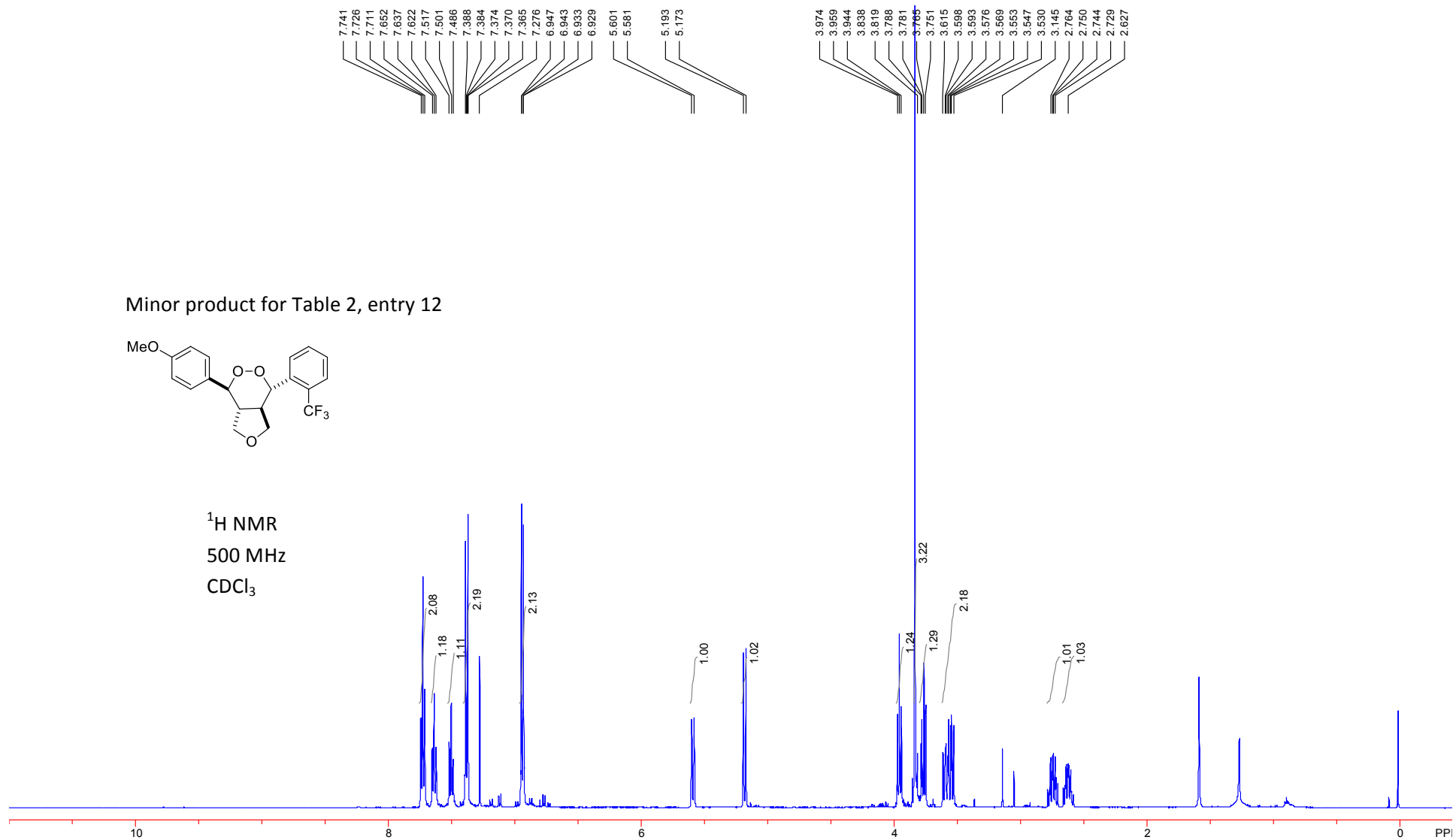
$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$



Minor product for Table 2, entry 12

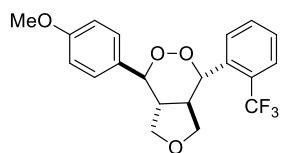


$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$

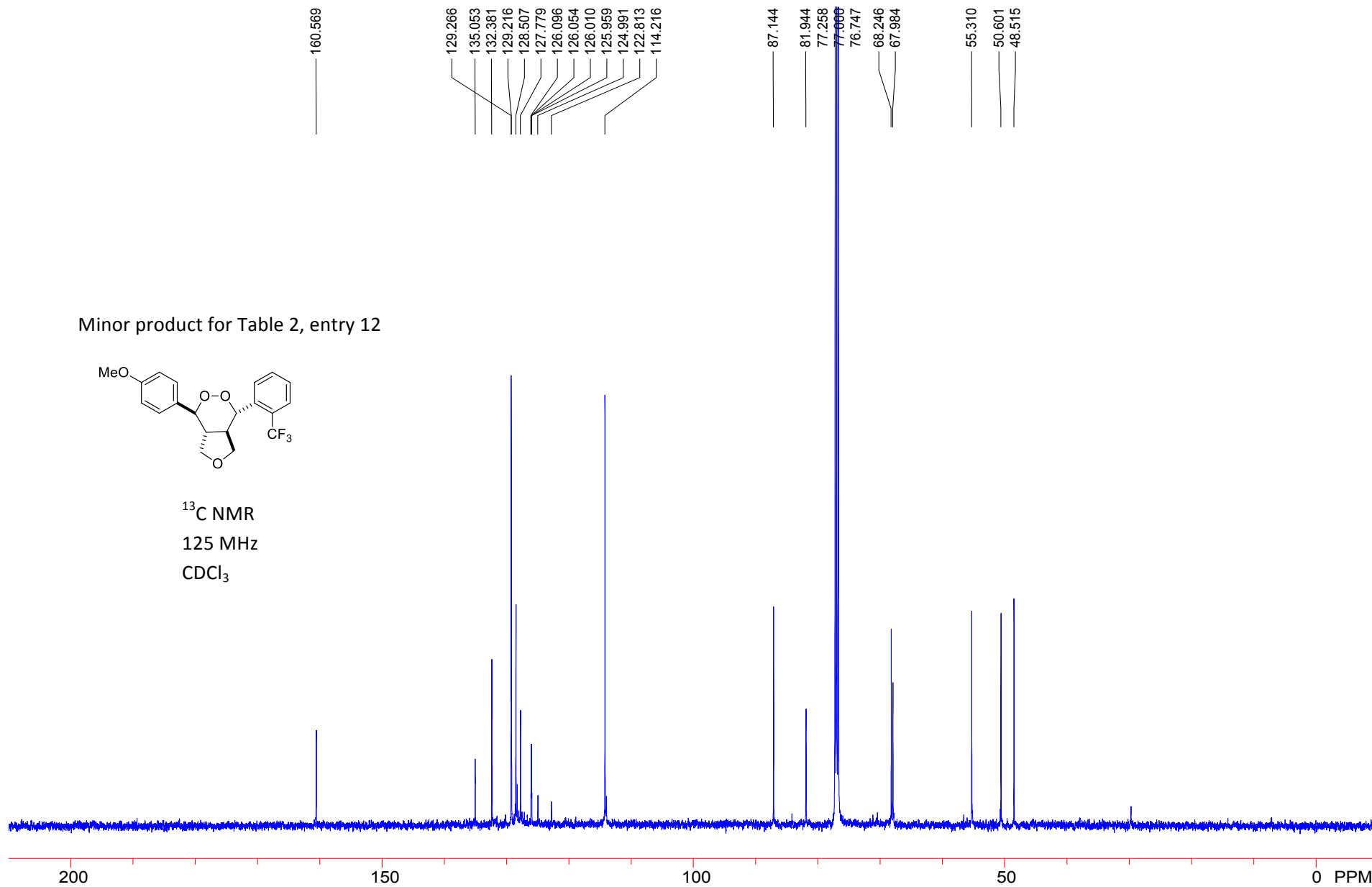




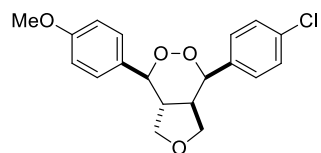
Minor product for Table 2, entry 12



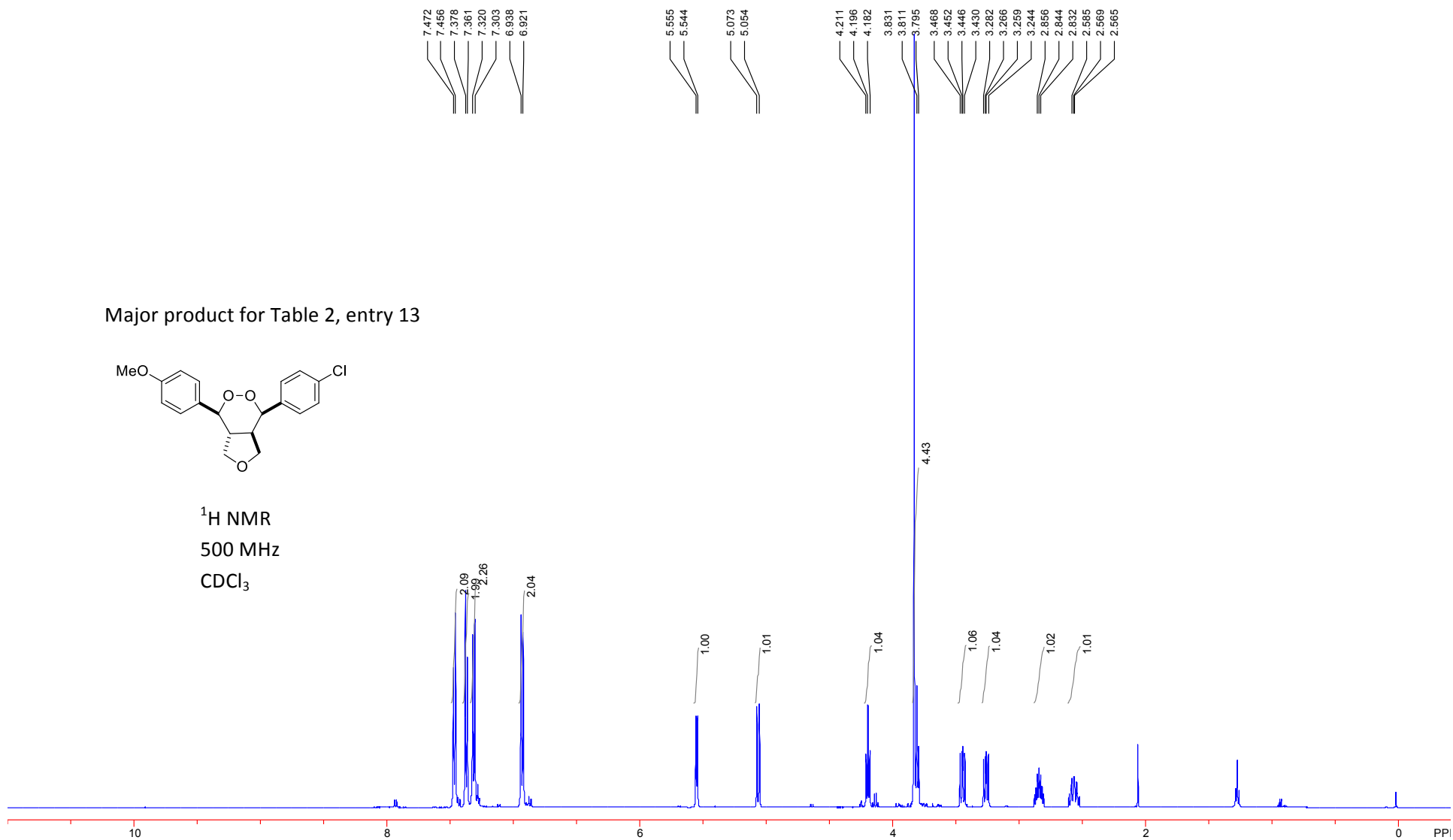
$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$



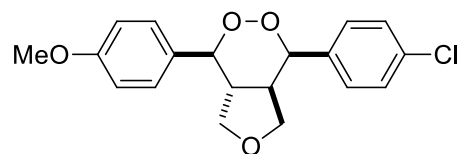
Major product for Table 2, entry 13



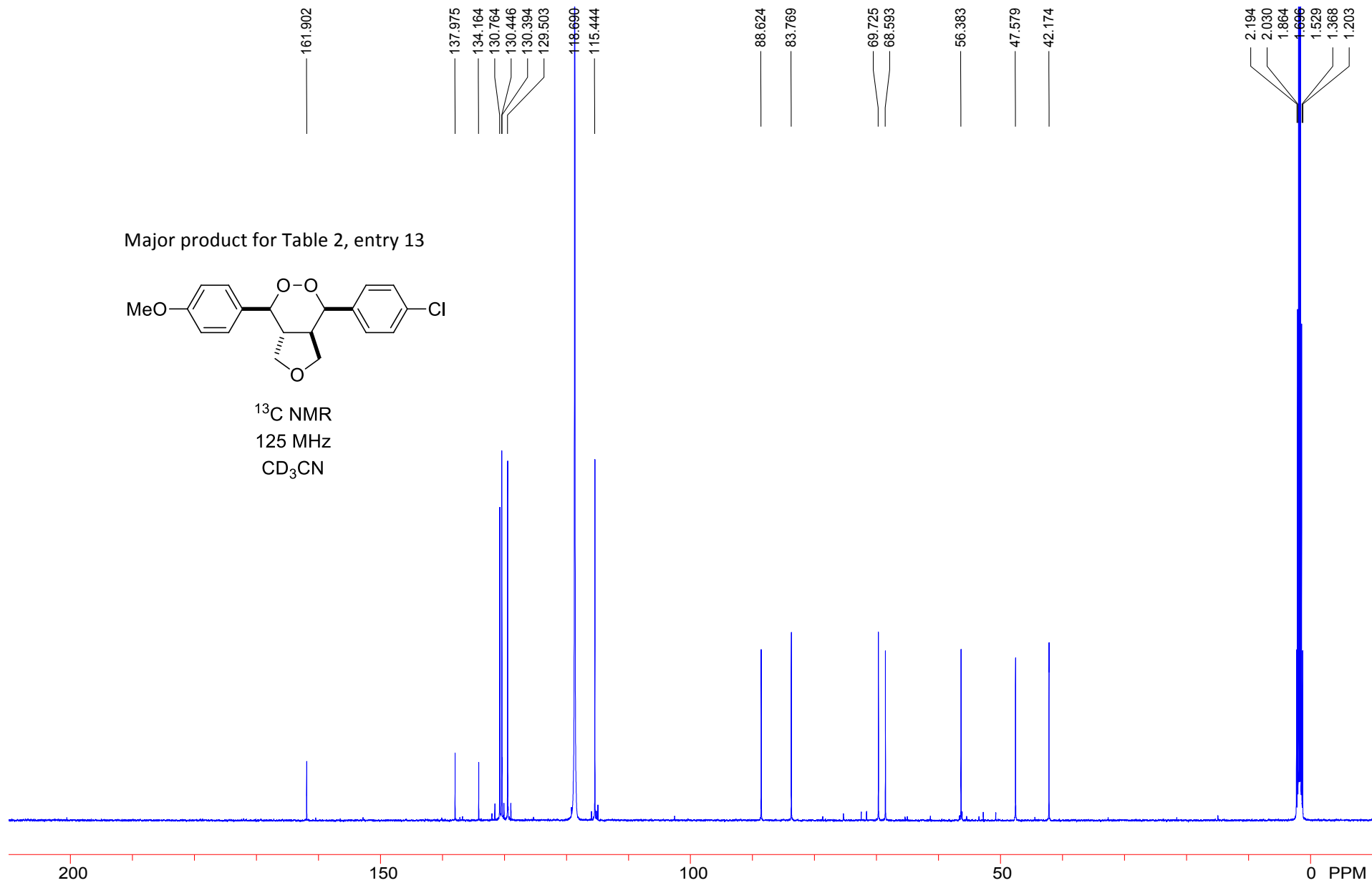
$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$



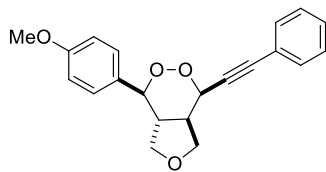
Major product for Table 2, entry 13



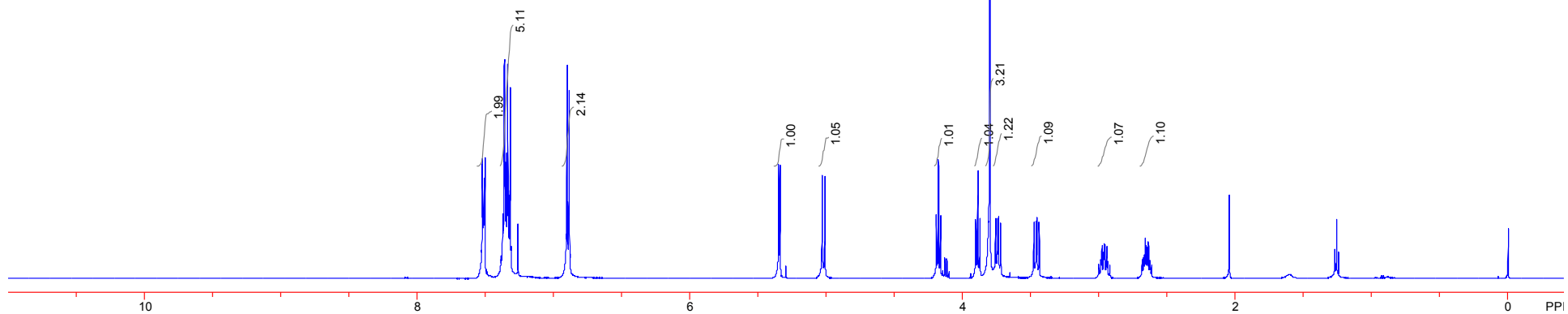
$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CD}_3\text{CN}$



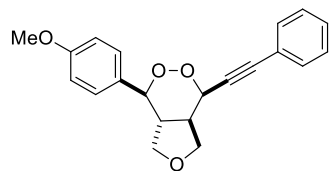
Major product for Table 2, entry 14



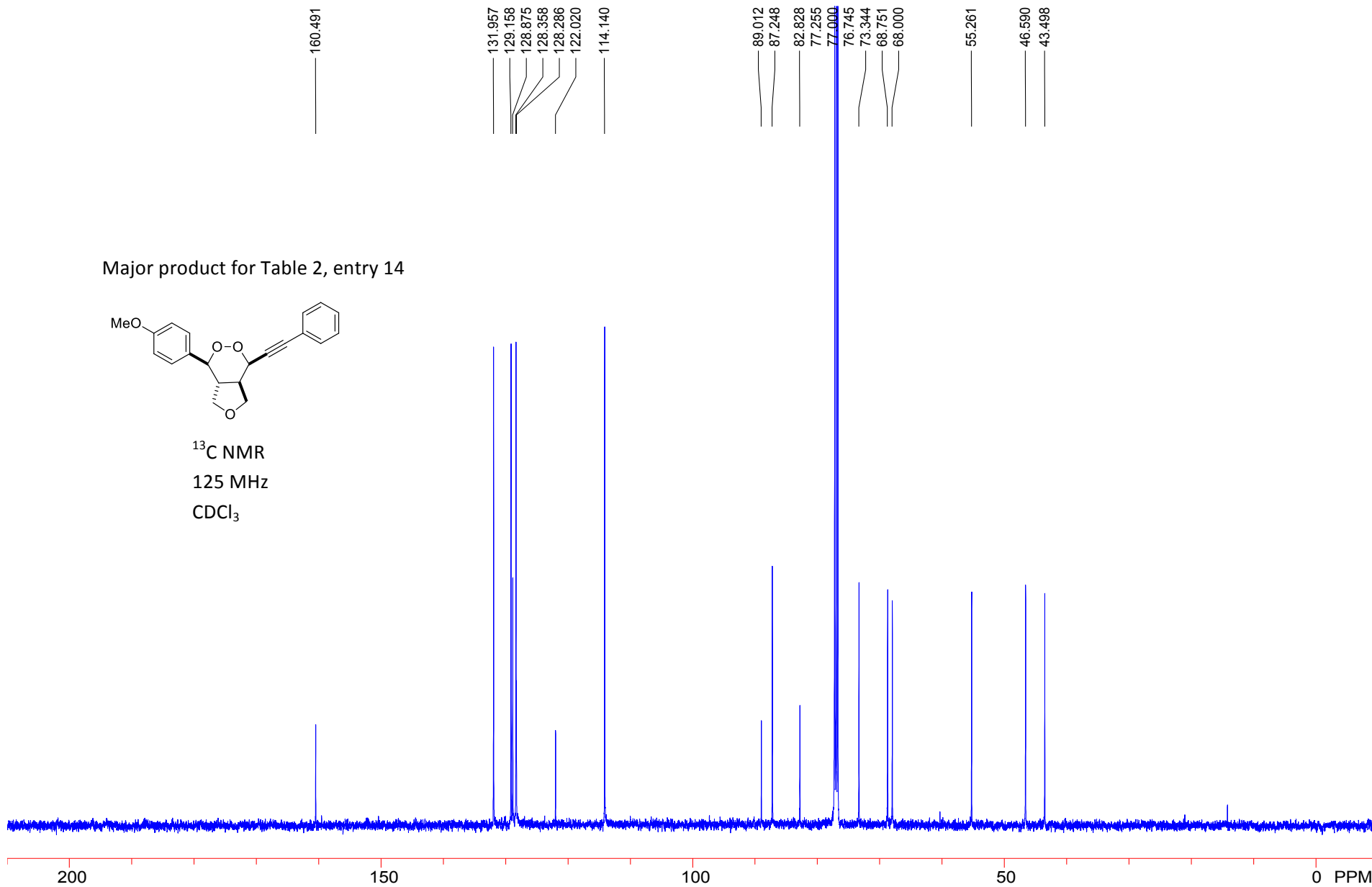
<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>



Major product for Table 2, entry 14

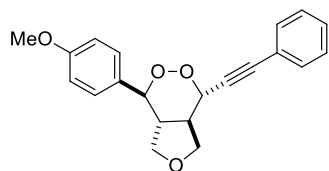


$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$

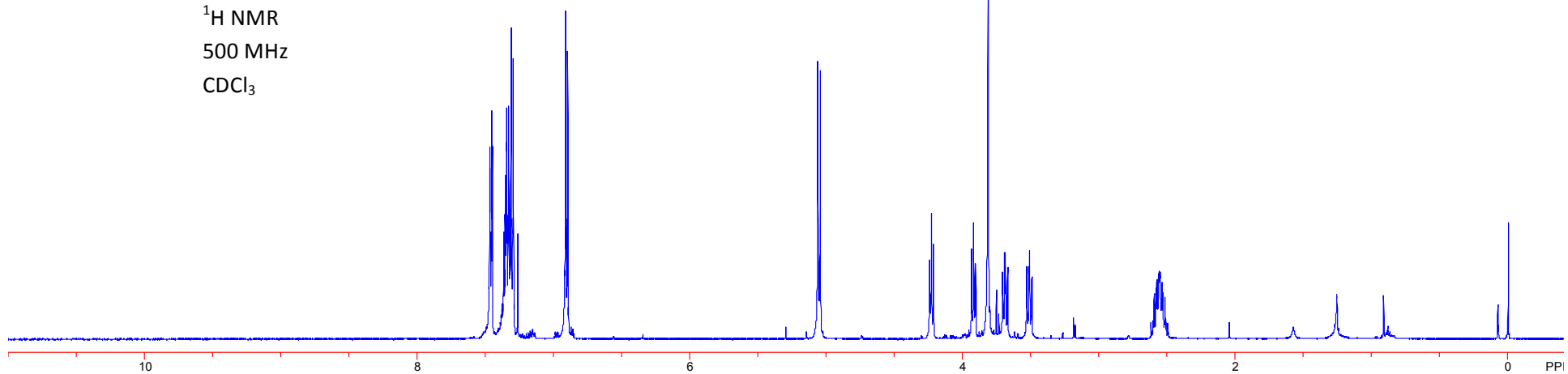


7.466  
7.462  
7.460  
7.454  
7.450  
7.446  
7.362  
7.356  
7.352  
7.348  
7.345  
7.341  
7.340  
7.337  
7.329  
7.325  
7.321  
7.315  
7.308  
7.305  
7.295  
7.291  
7.259  
6.916  
6.910  
6.906  
6.897  
6.892  
5.059  
5.040  
4.241  
4.225  
4.211  
3.931  
3.917  
3.901  
3.809  
3.704  
3.688  
3.683  
3.667  
3.527  
3.511  
3.506  
2.559  
2.557  
2.551

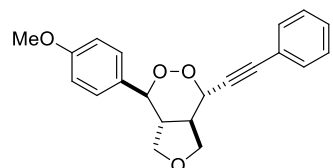
Minor product for Table 2, entry 14



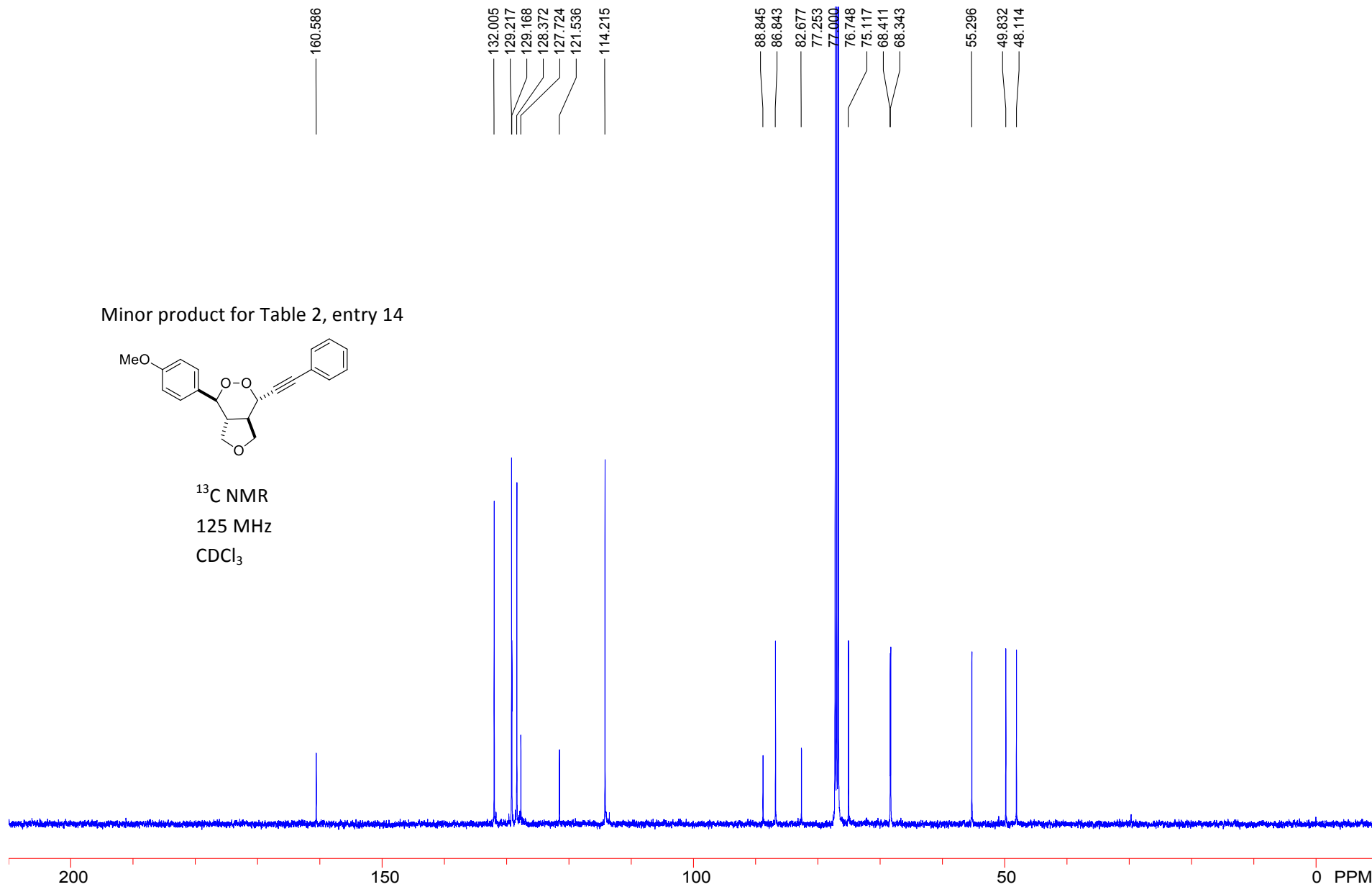
$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$

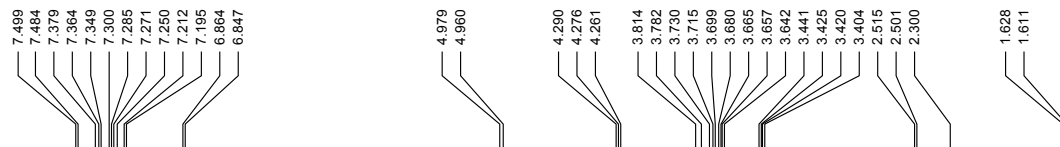


Minor product for Table 2, entry 14

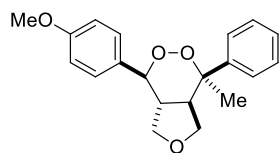


$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$

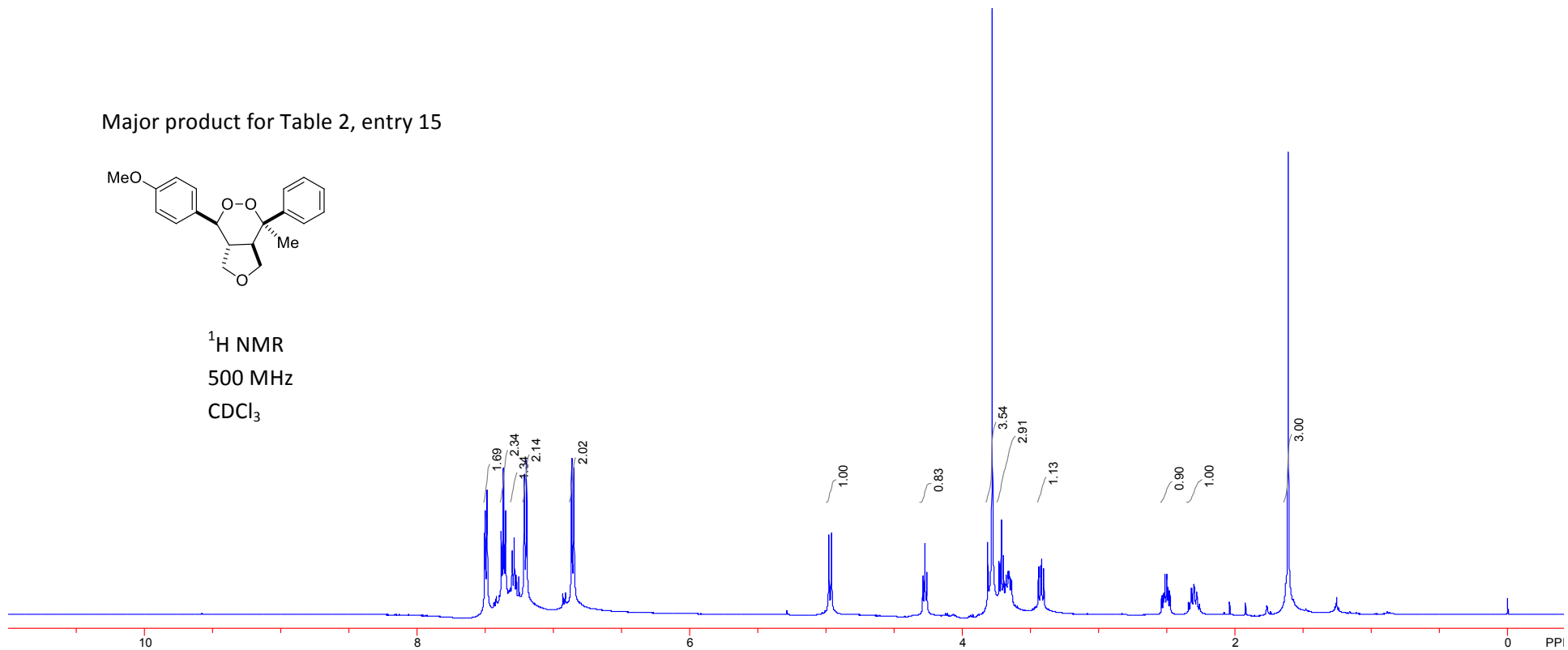




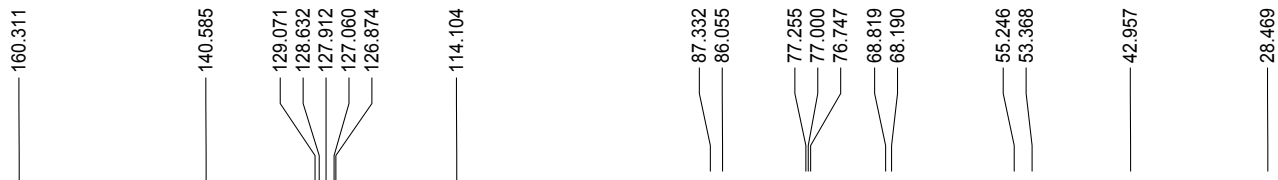
Major product for Table 2, entry 15



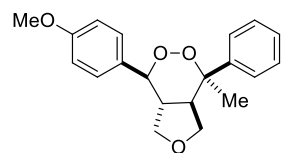
$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$



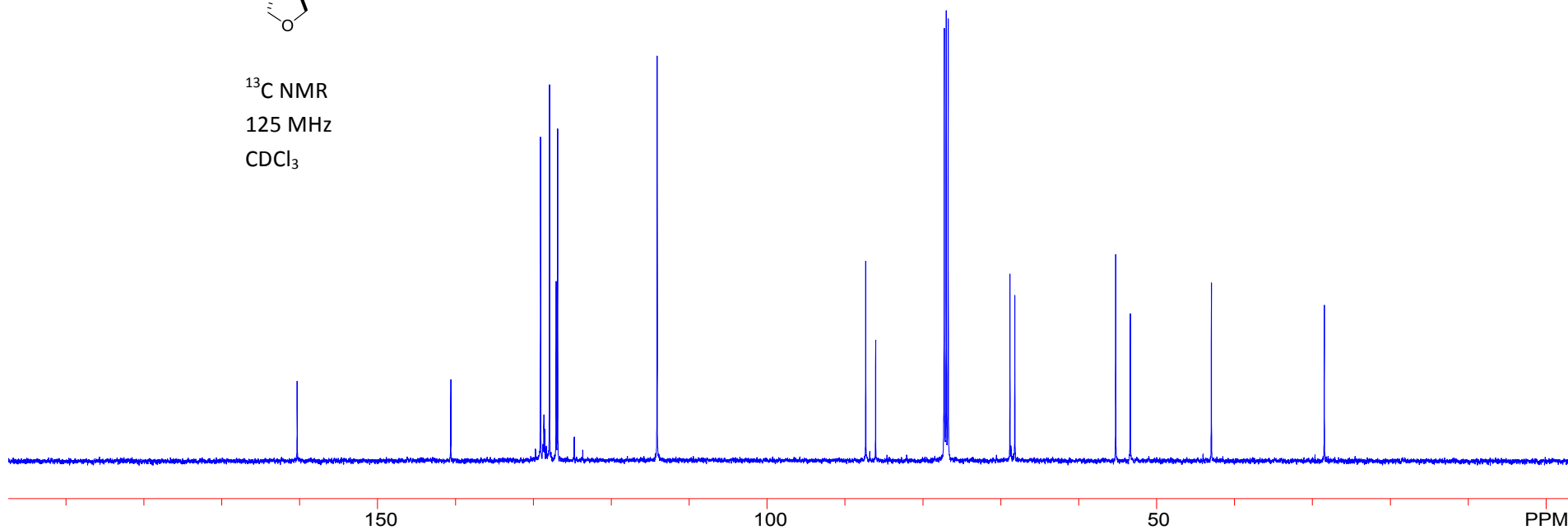




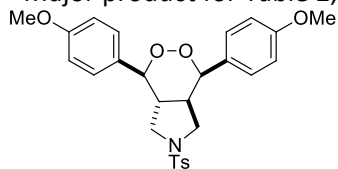
Major product for Table 2, entry 15



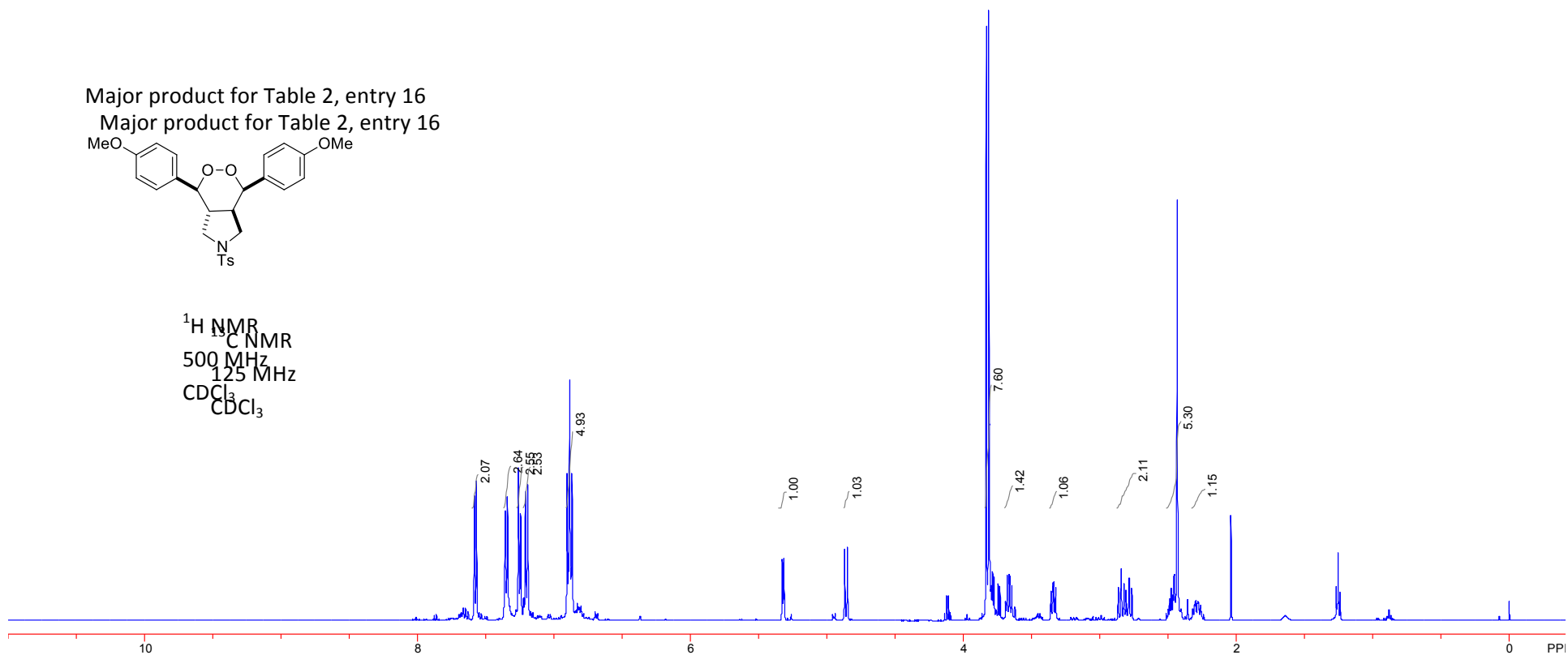
$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$



Major product for Table 2, entry 16  
 Major product for Table 2, entry 16



<sup>1</sup>H NMR  
<sup>13</sup>C NMR  
 500 MHz  
 125 MHz  
 CDCl<sub>3</sub>  
 CDCl<sub>3</sub>



160.498  
 159.182

143.526

134.196  
 129.715  
 129.045  
 128.963  
 128.543  
 128.363  
 127.138  
 114.214  
 113.664

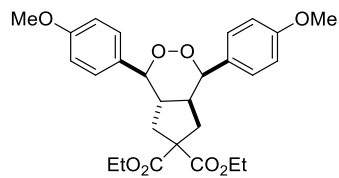
S-58

86.951  
 82.682  
 77.256  
 77.000  
 76.745

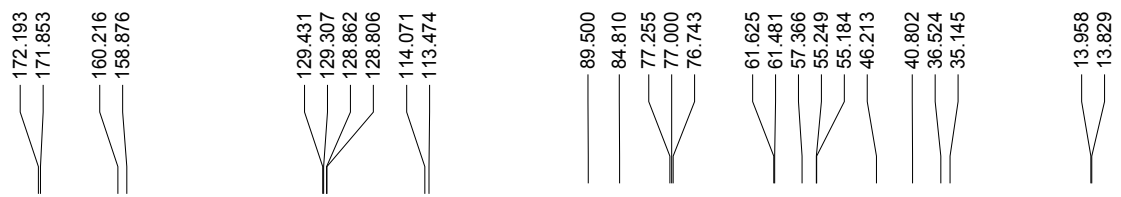
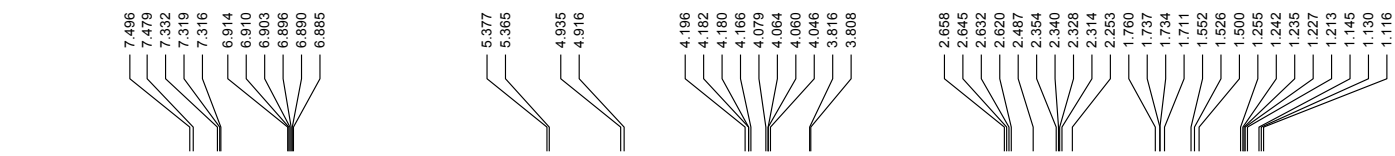
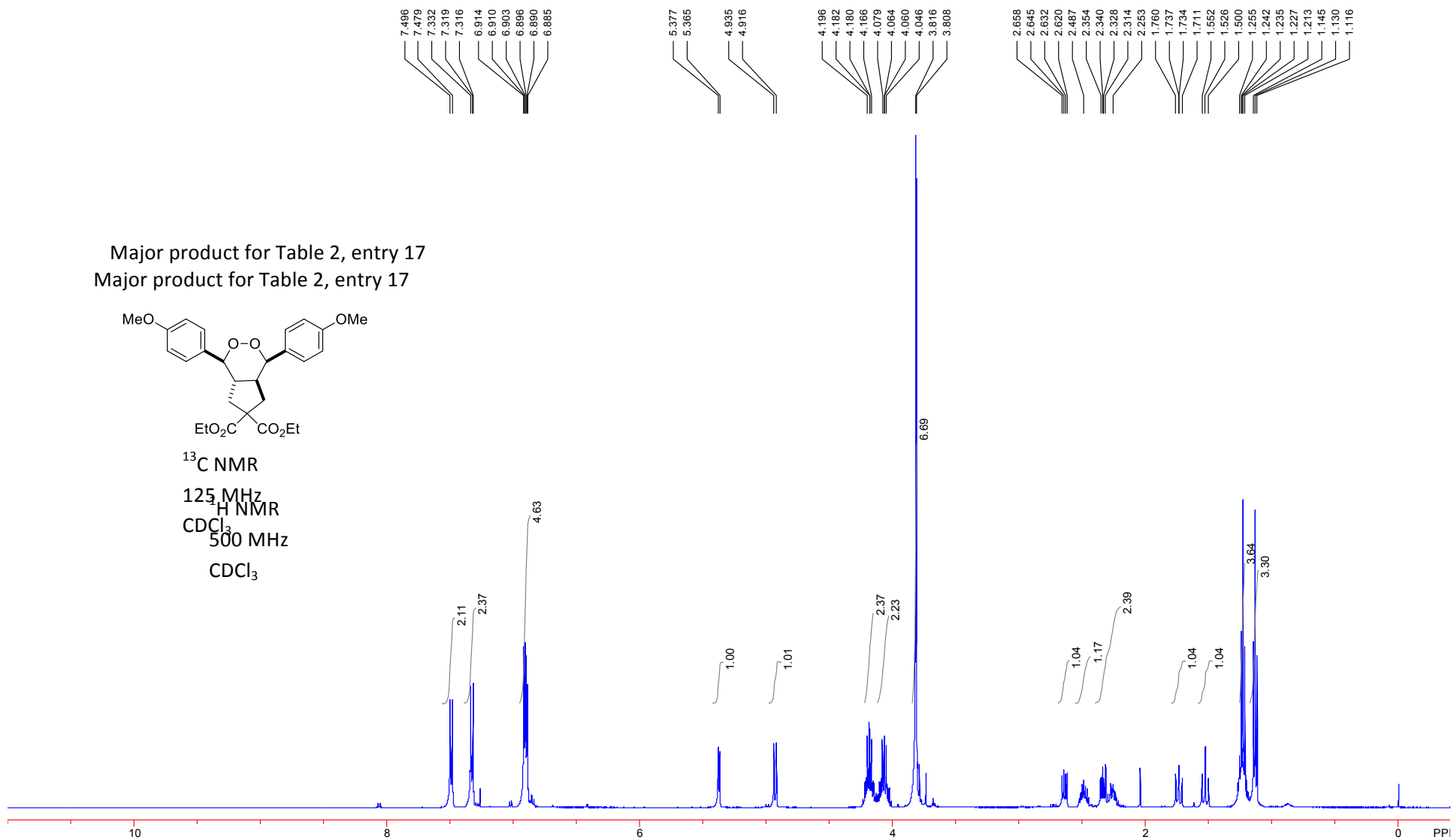
55.268  
 55.216  
 49.545  
 48.922  
 45.141  
 39.747

21.443

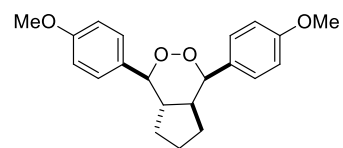
Major product for Table 2, entry 17  
 Major product for Table 2, entry 17



$^{13}\text{C}$  NMR  
 125 MHz  
 $^1\text{H}$  NMR  
 500 MHz  
 $\text{CDCl}_3$   
 $\text{CDCl}_3$

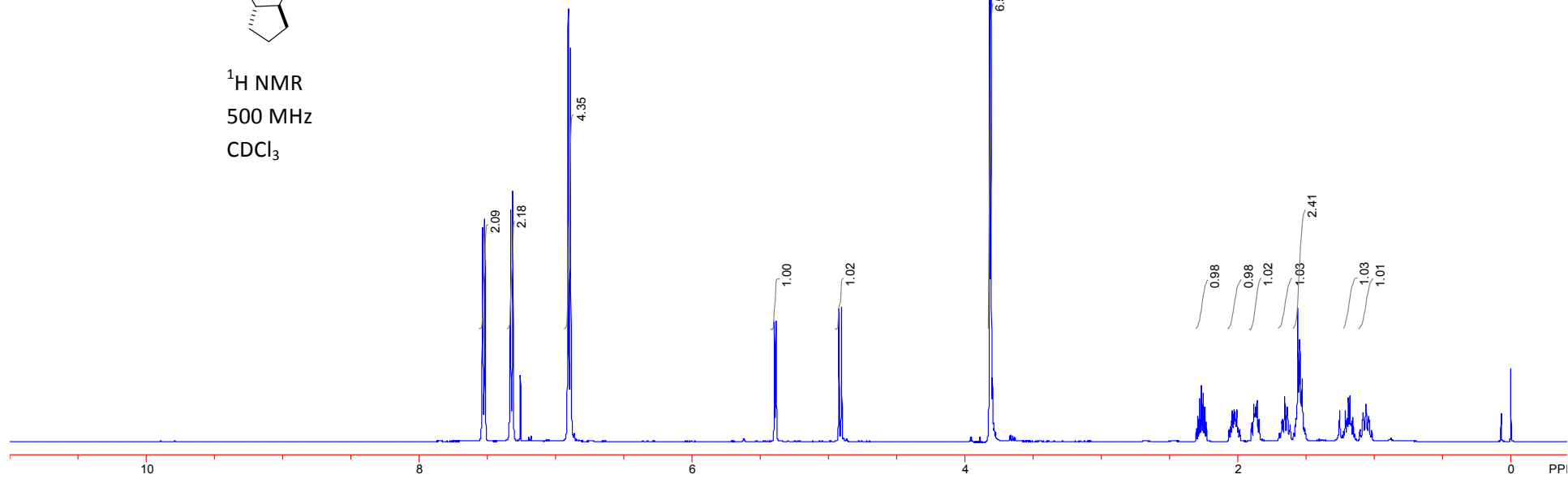


Major product for Table 2, entry 18

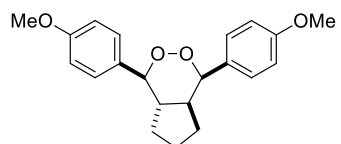


$^1\text{H}$  NMR  
500 MHz  
 $\text{CDCl}_3$

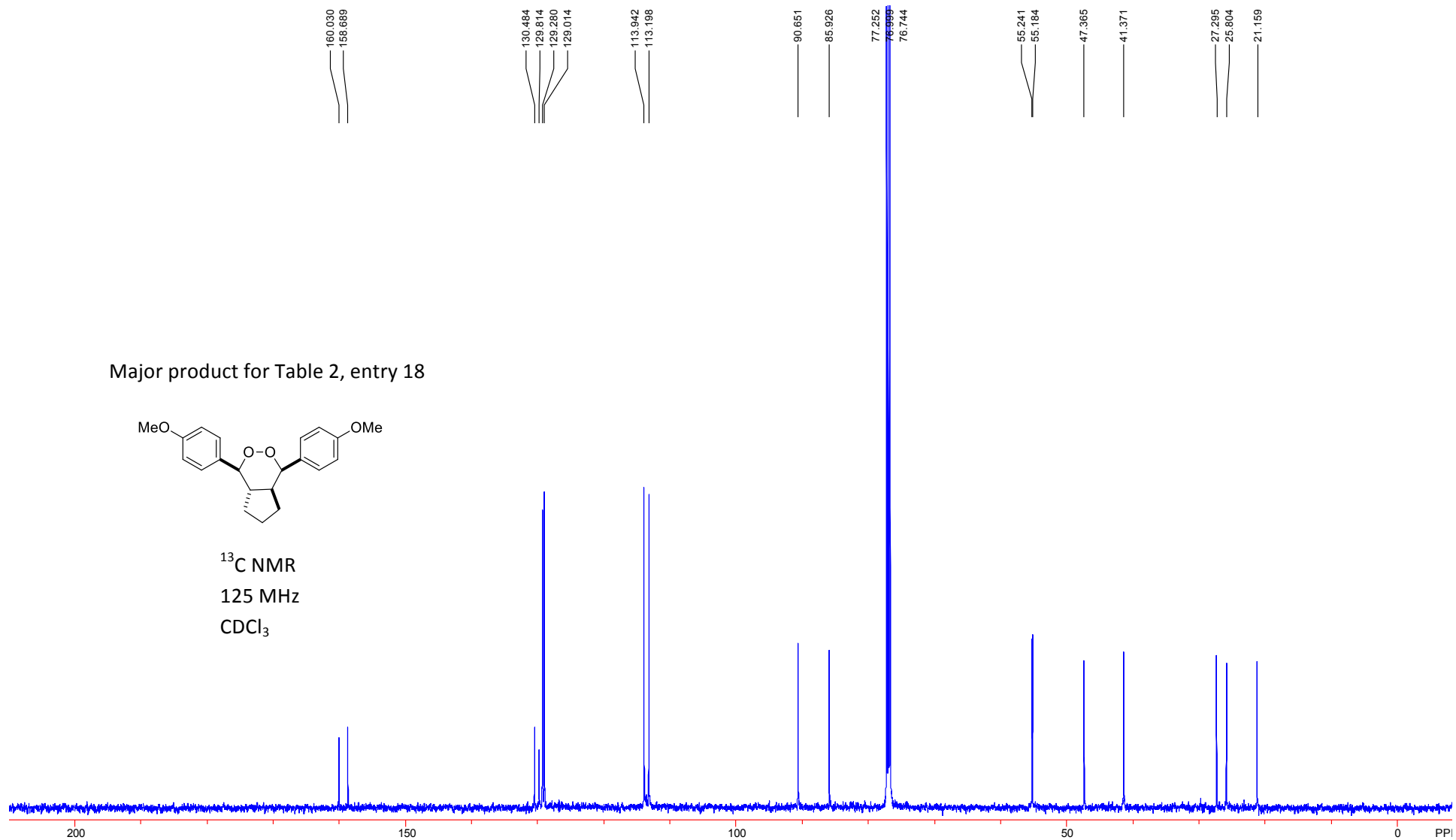
7.534  
7.516  
7.334  
7.329  
7.312  
7.256  
6.906  
6.889  
  
5.395  
5.383  
  
4.924  
4.903  
  
3.820  
3.810  
3.800  
  
2.280  
2.268  
2.256  
2.244  
1.884  
1.872  
1.860  
1.857  
1.636  
1.561  
1.547  
1.530  
1.191  
1.180  
1.062

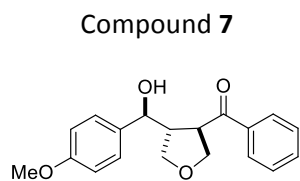


Major product for Table 2, entry 18

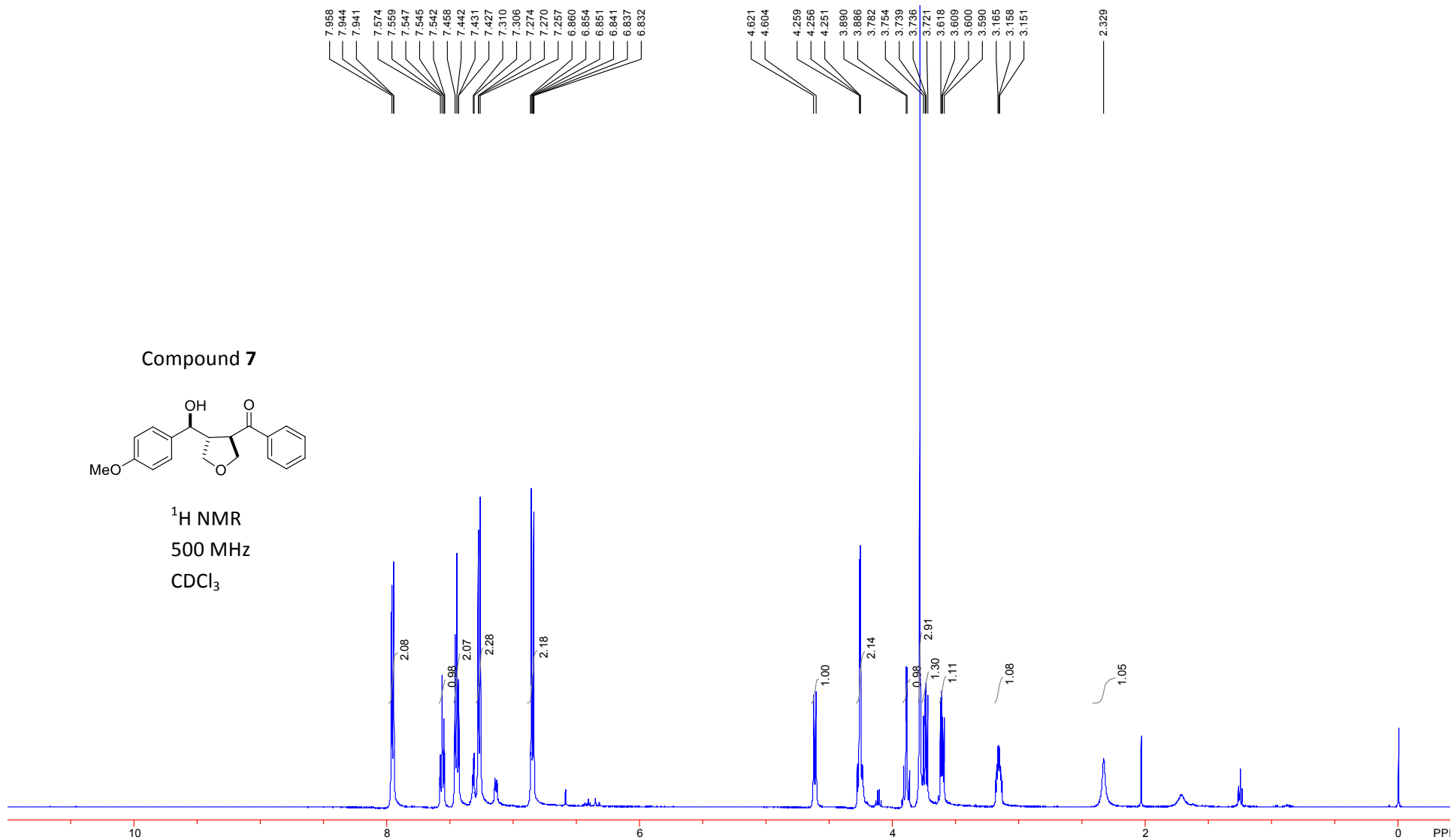


$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$





<sup>1</sup>H NMR  
500 MHz  
CDCl<sub>3</sub>



200.013

159.398

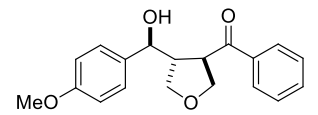
136.438  
134.851  
133.230  
128.622  
128.604  
127.608

114.028

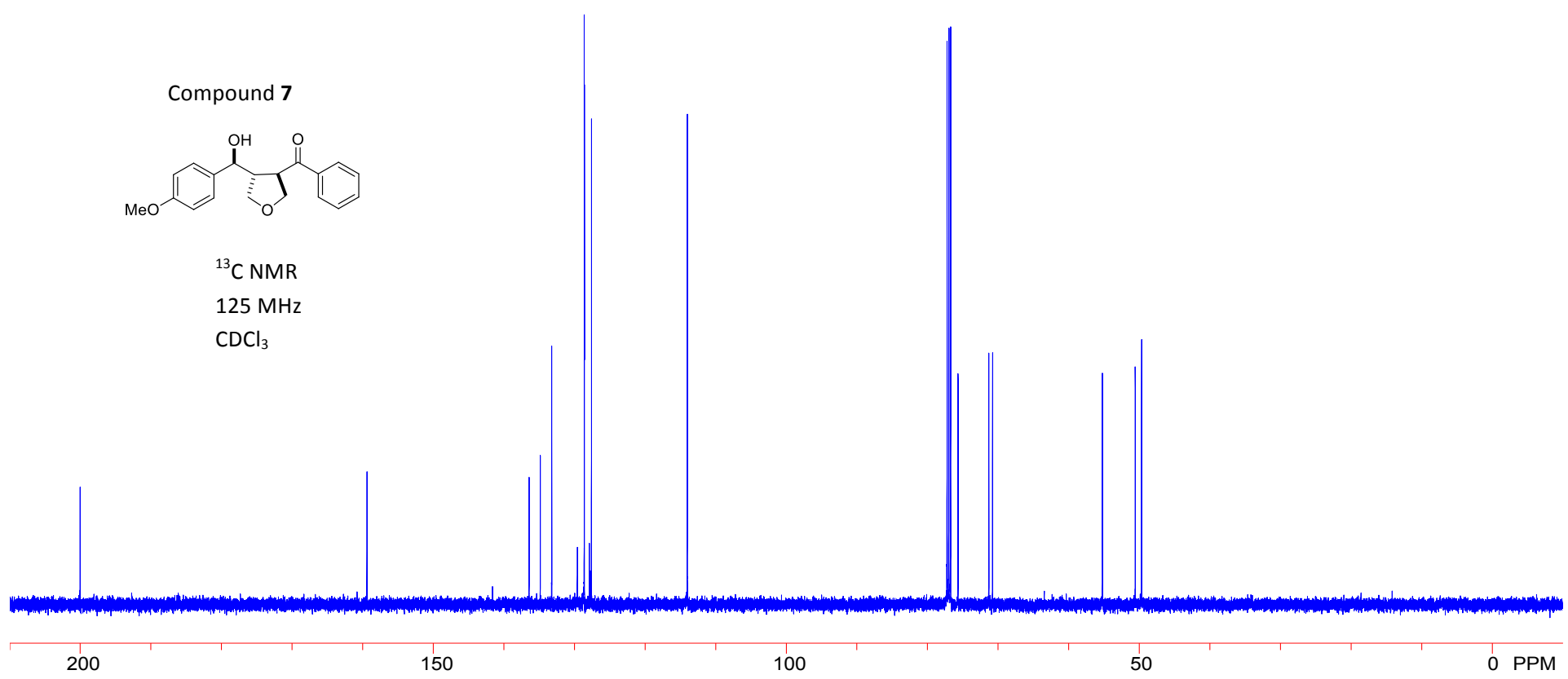
77.257  
77.000  
76.748  
75.701  
71.343  
70.823

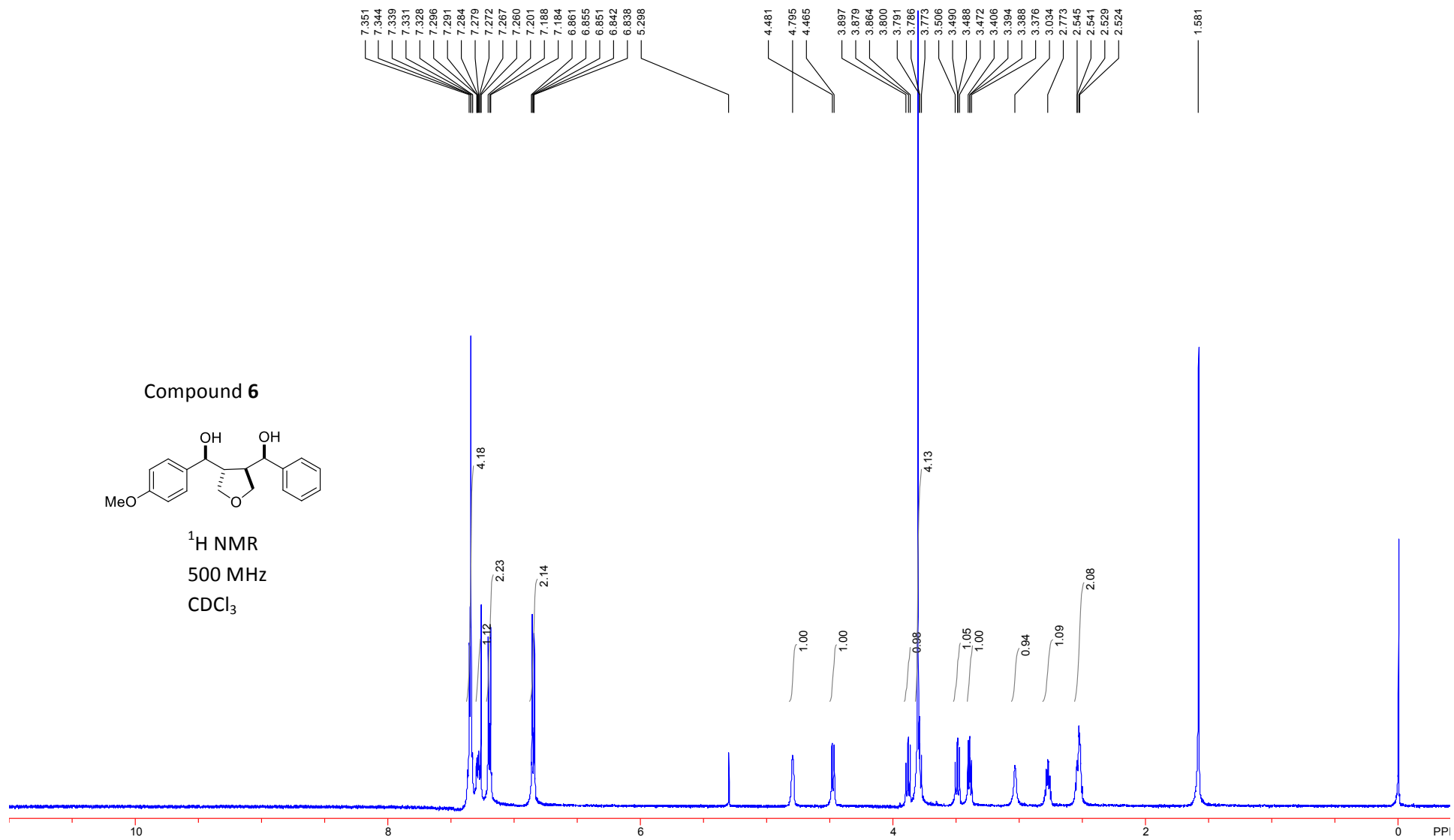
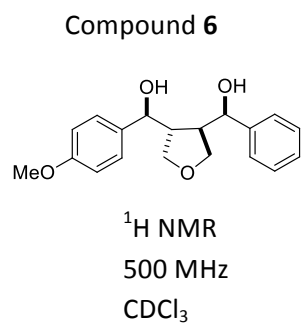
55.235  
50.584  
49.685

Compound 7

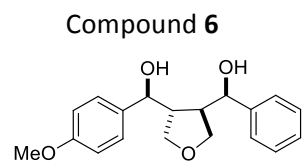


<sup>13</sup>C NMR  
125 MHz  
CDCl<sub>3</sub>









$^{13}\text{C}$  NMR  
125 MHz  
 $\text{CDCl}_3$

