Enantioselective Synthesis of Tryptophan Derivatives by a Tandem Friedel–Crafts Conjugate Addition/Asymmetric Protonation Reaction

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Supporting Information 1 (Experimental):

Table of Contents

1.	Materials and Methods	S3
2.	Catalyst and Substrate Preparation	S4
3.	Optimization of Reaction Parameters	
	a. General Procedure 1	S10
	b. Characterization Data	S11
4.	Optimized Conjugate Addition/Asymmetric Protonation Reaction	
	a. General Procedure 2	S13
	b. Characterization Data	S13
5.	SFC traces for racemic and enantioenriched tryptophan derivatives	S26
6.	Scale-up Procedure	S59
7.	Functionalization of tryptophan 7c and 7d	S60
8.	Scalemic (<i>R</i>)-BINOL experiment	S71
9.	Deuterium labeling studies	S72
10.	¹ H NMR Kinetics Experiment for SnCl ₄ and 9a•SnCl ₄ promoted reaction of 6a and 2c	S74
11.	Comparison of conditions for pyrroloindoline formation	S75

1. Materials and Methods. Unless otherwise stated, reactions were performed under a nitrogen atmosphere using freshly dried solvents. Methylene chloride, deuterated methylene chloride, dioxane, ether, tetrahydrofuran, and toluene were dried by passing through activated alumina. Dichloroethane and chloroform were distilled over calcium hydride. Powdered 4Å molecular sieves were flame-dried under vacuum immediately prior to use. Potassium carbonate was dried for 12 h at 130 °C under vacuum and 2,6-lutidine was distilled over AlCl₃. All other commercially obtained reagents were used as received unless specifically indicated. (R)-BINOL (9a), 2-phenylindole (6a) and 2-methylindole (6r) were purchased from Alfa Aesar, N-methyl-2phenylindole (6b) was obtained from Sigma-Aldrich, and 1 M SnCl₄ in CH₂Cl₂ was purchased from Acros Organics. (R)-3,3'-diphenvl-BINOL (9b), (R)-3,3'-dimethyl-BINOL (9c), (R)-3,3'dichloro-BINOL $(9e)^3$, (R)-3,3'-dibromo-BINOL $(9f)^4$, (R)-3,3'-dimethoxy-BINOL $(9g)^4$, (R)-6,6'-dimethyl-BINOL (9i)⁵ and (R)-6,6'-dibromo-BINOL (9i)⁶ were prepared according to literature procedures. All reactions were monitored by thin-layer chromatography using EMD/Merck silica gel 60 F254 pre-coated plates (0.25 mm). Silica gel column chromatography was performed either as described by Still et al. (Still, W. C.; Kahn, M.; Mitra, A. J. Org. Chem. 1978, 43, 2923.) using silica gel (particle size 0.032-0.063) purchased from Silicycle or using pre-packaged RediSep[®]Rf columns on a CombiSilica gel Rf system (Teledyne ISCO Inc.). Optical rotations were measured on a Jasco P-2000 polarimeter using a 100 mm path-length cell at 589 nm. ¹H and ¹³C NMR were recorded on a Varian Inova 500 (at 500 MHz and 125 MHz respectively) or a Varian Inova 600 (at 600 MHz and 150 MHz respectively) and are reported relative to internal chloroform (¹H, $\delta = 7.26$, ¹³C, $\delta = 77.0$) or internal acetonitrile (¹H, $\delta = 1.94$, ¹³C, $\delta = 1.32$). Data for ¹H NMR spectra are reported as follows: chemical shift (δ ppm) (multiplicity, coupling constant (Hz), integration). Multiplicity and qualifier abbreviations are as follows: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad. IR spectra were recorded on a Perkin Elmer Paragon 1000 spectrometer and are reported in frequency of absorption (cm⁻¹). Analytical SFC was performed with a Mettler SFC supercritical CO₂ analytical chromatography system with Chiralcel AD-H, OD-H, AS-H, OB-H, and OJ-H

¹ Zhang, X. PCT Int. Appl. WO 2002040491, 2002.

² Wu, T. R.; Shen, L.; J. M. Chong., J. M. Org. Lett. 2004, 6, 2701.

³ Ito, K.; Takahashi, M.; Hoshino, T.; Nishiki, M.; Ohba. Y. Lett. Org. Chem. 2006, 3, 735.

⁴ Ooi, T.; Kameda, M.; Maruoka, K. J. Am. Chem. Soc. 2003, 125, 5139.

⁵ Verga, D.; Percivalle, C.; Doria, F.; Porta, A.; Freccero, M. J. Org. Chem. 2011, 76, 2319.

⁶ Rueping, M.; Sugiono, E.; Steck, A.; Thiessmann, T. Adv. Synth. Catal. 2010, 352, 281.

columns (4.6 mm x 25 cm). HRMS were acquired using either an Agilent 6200 Series TOF with an Agilent G1978A Multimode source in electrospray ionization (ESI), atmospheric pressure chemical ionization (APCI) or mixed (MM) ionization mode, or obtained from the Caltech Mass Spectral Facility.

Abbreviations used: BINOL – 1,1'-bi(2-naphthol); IPA – isopropanol; Et₂O – diethyl ether; PhMe – toluene; EtOAc – ethyl acetate; DCE – dichloroethane; DCM – dichloromethane; MeCN – acetonitrile; ee – enantiomeric excess

2. Catalyst and Substrate Preparation.

Preparation of (R)-3-chloro-BINOL (9d)



To a flame-dried 100 mL flask containing MOM–protected (*R*)-BINOL **S1**² (748 mg, 2.00 mmol, 1.00 equiv) was added Et₂O (45 mL), followed by dropwise addition of *n*-BuLi as a solution in hexanes (2.5 M, 960 μ L, 2.40 mmol, 1.20 equiv) at room temperature. The mixture was then stirred at room temperature for 3 h and subsequently cooled to –78 °C, followed by addition of C₂Cl₆ (569 mg, 2.40 mmol, 1.20 equiv) in one portion. The reaction mixture was allowed to warm to room temperature over 3 h, then diluted with EtOAc (15 mL) and washed with saturated aqueous NH₄Cl (50 mL). The aqueous layer was extracted with EtOAc (45 mL) and the combined organic layers were dried (Na₂SO₄), filtered and concentrated. The crude yellow oil was purified by silica gel chromatography (0:100 to 12:88 EtOAc:hexanes) to yield 328 mg (40% yield) of **S2** as a white solid. ¹H NMR (500 MHz, CDCl₃) δ 8.05 (s, 1H), 7.97 (d, *J* = 9.0 Hz, 1H), 7.87 (d, *J* = 8.1 Hz, 1H), 7.81 (d, *J* = 8.2 Hz, 1H), 7.59 (d, *J* = 9.1 Hz, 1H), 7.42 (ddd, *J* = 8.1, 6.7, 1.3 Hz, 1H), 7.37 (ddd, *J* = 8.1, 6.8, 1.2 Hz, 1H), 7.28 (ddd, *J* = 8.2, 6.8, 1.3 Hz, 1H), 7.24 (ddd, *J* = 8.5, 6.7, 1.3 Hz, 1H), 7.18 (dddd, *J* = 8.6, 1.3, 0.7, 0.7 Hz, 1H), 7.16 (ddd, *J* = 8.5, (d, *J* = 5.6 Hz, 1H), 3.19 (s, 3H), 2.71 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 152.9, 148.9,

133.8, 132.6, 131.1, 130.0, 129.5, 128.8, 128.0, 127.9, 127.8, 127.0, 126.7, 126.4, 126.1, 125.8, 125.5, 124.2, 119.9, 116.3, 98.8, 94.9, 56.5, 55.9; IR (NaCl/thin film): 2955, 2902, 1594, 1508, 1354, 1241, 1159, 1149, 1034, 1014, 961, 922 cm⁻¹; $[\alpha]_D^{25} = +69.1$ (c = 0.90, CHCl₃). HRMS (FAB+) calc'd for M⁺ 408.1128, found 408.1128.



A 10 mL flask was charged with **S2** (305 mg, 0.75 mmol, 1.00 equiv), dioxane (3.7 mL) and aqueous HCl (12 M, 130 μ L, 1.58 mmol, 2.10 equiv), then heated to 50 °C for 2 h. The mixture was cooled to room temperature, then diluted with H₂O (30 mL) and extracted with EtOAc (6 x 20 mL). The combined organic layers were dried (Na₂SO₄), filtered and concentrated. The crude residue was purified by silica gel chromatography (0:100 to 20:80 EtOAc:hexanes) to yield 210 mg (87% yield) of (*R*)-3-chloro-BINOL (**9d**) as a white foam, which was dried over P₂O₅ under vacuum. ¹H NMR (500 MHz, CDCl₃) δ 8.09 (s, 1H), 7.97 (d, *J* = 8.9 Hz, 1H), 7.90 (d, *J* = 8.1 Hz, 1H), 7.83 (d, *J* = 8.2 Hz, 1H), 7.45 – 7.35 (m, 3H), 7.34 – 7.28 (m, 2H), 7.16 (d, *J* = 8.5 Hz, 1H), 7.11 (d, *J* = 8.4 Hz, 1H), 5.60 (s, 1H), 4.94 (s, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 152.1, 148.3, 133.1, 132.4, 131.3, 129.7, 129.32, 129.26, 128.4, 127.7, 127.5, 127.3, 125.1, 124.6, 124.1, 123.9, 122.4, 117.7, 113.6, 111.7; IR (NaCl/thin film): 3503, 3057, 1620, 1596, 1502, 1451, 1379, 1265, 1212, 1184, 1146, 828 cm⁻¹; [α]_D²⁵ = +55.4 (*c* = 1.01, CHCl₃). HRMS (MM) calc' d for [M-H]⁻ 319.0531, found 319.0549.

Preparation of (R)-6,6'-dimethoxy-BINOL (9h)



(R)-6,6'-dimethoxy-BINOL (9h) was prepared following a procedure adapted from a reported synthesis of (R)-3,3'-dimethoxy-BINOL (9g).⁴ To a 25 mL flask containing MOM-protected (R)-6,6'-dibromo-BINOL $\mathbf{83}^{6}$ (1.10 g, 2.07 mmol, 1.00 equiv) was added THF (6.3 mL). The flask was cooled to -78 °C, followed by dropwise addition of *n*-BuLi as a solution in hexanes (2.5 M, 2.50 mL, 6.20 mmol, 3.00 equiv). After stirring 1 hour at -78 °C, B(OMe)₃ (645 mg, 6.20 mmol, 3.00 equiv) was added and the reaction was allowed to warm to room temperature. After 14 hours, the reaction mixture was concentrated to give the crude borate intermediate, which was suspended in benzene (7.2 mL) and cooled to 0 °C, followed by dropwise addition of aqueous hydrogen peroxide (30 wt %, 0.61 mL, 5.98 mmol, 2.89 equiv). The suspension was heated to reflux for 4 hours, then cooled to room temperature, poured into ice-cold saturated aqueous NaSO₃ (20 mL), and extracted with EtOAc (3 x 15 mL). The combined organics were washed with brine (30 mL), dried (Na₂SO₄), filtered, and concentrated. The crude residue was purified by silica gel chromatography (0:100 to 50:50 EtOAc:hexanes) to yield 512 mg (61% yield) of S4 as a light yellow foam. ¹H NMR (500 MHz, CD₃CN) δ 7.80 (ddd, J = 9.1, 0.8, 0.4J = 9.1, 0.7, 0.7 Hz, 2H), 6.87 (dd, J = 9.1, 2.5 Hz, 2H), 5.02 (d, J = 6.7 Hz, 2H), 4.94 (d, J = 6.7Hz, 2H), 3.11 (s, 6H); ¹³C NMR (125 MHz, CD₃CN) δ 154.4, 151.6, 132.1, 129.6, 128.4, 127.8, 122.1, 119.6, 118.7, 110.1, 96.0, 56.1; IR (NaCl/thin film): 3368, 2914, 1624, 1599, 1511, 1240, 1196, 1148, 1023 cm⁻¹; $[\alpha]_D^{25} = +87.1$ (c = 1.00, MeCN). HRMS (MM) calc'd for [M-H]⁻ 405.1344, found 405.1350.



A 15 mL flask was charged with **S4** (200 mg, 0.493 mmol, 1.00 equiv) and K₂CO₃ (177 mg, 1.28 mmol, 2.60 equiv). DMF (2 mL) was added, followed by MeI (123 μ L, 1.97 mmol, 4.00 equiv) dropwise. The reaction was then heated to 55 °C for 22 hours, then cooled to room temperature and quenched with saturated aqueous NH₄Cl (2 mL) and Et₃N (3 drops). The mixture was stirred at room temperature for 6 hours, then diluted with H₂O (15 mL) and extracted with EtOAc (3 x 10 mL). The combined organics were washed with brine (15 mL), dried (Na₂SO₄), and concentrated. THF (28 mL) and IPA (9.5 mL) were added to the crude residue, followed by dropwise addition of aqueous HCl (6.0 M, 9.4 mL). The reaction was stirred at room temperature for 3 hours, then diluted with H₂O (70 mL) and extracted with EtOAc (3 x 30 mL). The combined organics were washed aqueous NaHCO₃ (2 x 45 mL) and brine (45 mL), then dried (Na₂SO₄), filtered, and concentrated. The crude oil was purified by silica gel chromatography (0:100 to 30:70 EtOAc:hexanes) to yield 62 mg (36% yield) of (*R*)-6,6²-dimethoxy-BINOL (**9h**) as a light brown solid, which was dried over P₂O₅ under hi-vacuum. Spectral data are in agreement with the literature.⁷

Preparation of 1-allyl-2-phenylindole (6c)



To a 50 mL flask was added NaH (620 mg, 15.5 mmol, 3.00 equiv) and DMF (8 mL) and the suspension was cooled to 0 $^{\circ}$ C in an ice bath. A solution of 2-phenylindole **6a** (1.00 g, 5.18 mmol, 1.00 equiv) in DMF (3 mL) was added slowly to the suspension over 15 minutes and the reaction mixture was further stirred at 0 $^{\circ}$ C for 20 minutes, followed by dropwise addition of

⁷ Yu, H.-B.; Hu, Q.-S.; Pu, L. J. Am. Chem. Soc. 2000, 122, 6500.

allyl bromide (670 μ L, 7.77 mmol, 1.50 equiv). The ice bath was then removed and the mixture was stirred for 15 minutes, then quenched by addition of saturated aqueous NH₄Cl (5 mL) and Et₃N (5 drops). After 2 hours, the reaction was diluted with H₂O (40 mL) and extracted with EtOAc (3 x 30 mL). The combined organics were washed with brine (120 mL), dried (Na₂SO₄), filtered, and concentrated. The crude was then purified by reverse phase preparatory HPLC (55:45 to 95:5 MeCN:H₂O) using an Agilent 1200 Series HPLC with an Agilent XDB-C18 5 μ M column (9.4 x 250 mm and 21.2 x 150 mm) to yield 687 mg (57% yield) of 1-allyl-2-phenylindole (**6c**) as a yellow solid and 331 mg (23% yield) of 1,3-diallyl-2-phenylindole (**S5**) as a yellow oil.

1-allyl-2-phenylindole (6c):

¹H NMR (500 MHz, CDCl₃) δ 7.65 (ddd, J = 7.8, 1.2, 0.8 Hz, 1H), 7.55 – 7.51 (m, 2H), 7.48 –

7.43 (m, 2H), 7.42 – 7.38 (m, 1H), 7.33 (br d, J = 8.2 Hz, 1H), 7.22 (ddd, J = 7.0, 7.0, 1.3 Hz, 1H), 7.15 (ddd, J = 7.0, 7.0, 1.0 Hz, 1H), 6.60 (br s, 1H), 6.02 (ddt, J = 17.2, 10.5, 4.4 Hz, 1H), 5.22 (dtd, J = 10.5, 1.8, 1.1 Hz, 1H), 5.00 (dtd, J = 17.1, 2.0, 1.2 Hz, 1H), 4.74 (dt, J = 4.2, 1.9 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 141.5, 137.8, 133.8, 132.7, 129.1, 128.5, 128.1, 128.0, 121.7, 120.5, 120.0, 116.5, 110.3, 102.0, 46.5; IR (NaCl/thin film): 3055, 2917, 1602, 1462, 1443, 1392, 1345, 1317, 1162 cm⁻¹; HRMS (APCI) calc'd for [M+H]⁺ = 234.1277, found 234.1284.

1,3-diallyl-2-phenylindole (S5):

¹H NMR (500 MHz, CDCl₃) δ 7.65 (ddd, J = 7.8, 1.2, 0.7 Hz, 1H), 7.50 – 7.40 (m, 5H), 7.33



(ddd, J = 8.1, 0.9, 0.9 Hz, 1H), 7.24 (ddd, J = 7.0, 7.0, 1.2 Hz, 1H), 7.16 (ddd, J = 7.0, 7.0, 1.1 Hz, 1H), 6.05 (ddt, J = 17.0, 10.1, 5.9 Hz, 1H), 5.91 (ddt, J = 17.1, 10.4, 4.7 Hz, 1H), 5.14 (dtd, J = 10.4, 1.8, 1.2 Hz, 1H), 5.08 – 5.02 (m, 2H), 4.92 (dtd, J = 17.1, 1.9, 1.3 Hz, 1H), 4.62 (dt, J = 4.6, 1.9 Hz, 2H), 3.46 (dt, J = 6.0, 1.7 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 138.0, 137.9, 136.7.

133.9, 131.8, 130.4, 128.3, 128.2, 128.1, 128.0, 121.7, 119.34, 119.30, 116.2, 114.6, 110.9, 110.1, 46.4, 29.2; IR (NaCl/thin film): 3056, 2915, 1637, 1463, 1443, 1408, 1360, 1340, 1191 cm⁻¹; HRMS (MM) calc'd for $[M+H]^+ = 274.1590$, found 274.1591.

Preparation of 2-(2-fluorophenyl)indole (6p)



2-(2-fluorophenyl)indole (6p) was prepared by an analogous procedure to that reported by Sakai et. al.8 A flame-dried flask was charged with 2-iodoaniline (S6, 200 mg, 0.90 mmol, 1.00 equiv), ethynyl-2-fluorobenzene (S7, 133 mg, 1.10 mmol, 1.20 equiv), Pd(PPh₃)₂Cl₂ (13 mg, 0.02 mmol, 0.02 equiv), copper (I) iodide (2.0 mg, 0.025 mmol, 0.01 equiv) and Et₃N (4 mL). The mixture was stirred overnight at room temperature, then filtered through a plug of silica, concentrated and redissolved in PhMe (5 mL). InBr₃ (16 mg, 0.05 mmol, 0.05 equiv) was added in one portion and the mixture was heated to 110 °C for 5 h, then cooled to room temperature, filtered through celite, and concentrated. The crude residue was purified by silica gel chromatography (10:90 EtOAc:hexanes) to vield 148 mg (77% vield) of 2-(2-fluorophenyl)indole (6p) as a white solid. ¹H NMR (500 MHz, CDCl₃) δ 8.89 (br s, 1H), 7.80 (ddd, J = 7.8, 7.8, 1.8 Hz, 1H), 7.66 (dddd, J= 2.5, 1.3, 0.8, 0.8 Hz, 1H), 7.43 (ddd, J = 8.1, 1.5, 0.8 Hz, 1H), 7.32 - 7.26 (m, 1H), 7.26 - 7.16(m, 3H), 7.14 (ddd, J = 8.0, 7.0, 1.0 Hz, 1H), 6.97 (d, J = 1.9 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 159.3 (d, J_{C-F} = 246.4 Hz), 134.6 (d, J_{C-F} = 501.8 Hz), 128.8 (d, J_{C-F} = 8.8 Hz), 128.1, 128.0 (d, $J_{C-F} = 4.1$ Hz), 124.8 (d, $J_{C-F} = 3.2$ Hz), 122.7, 120.6, 120.2, 119.9 (d, $J_{C-F} = 11.0$ Hz), 116.6, 116.4, 111.0, 101.6 (d, J_{C-F} = 3.0 Hz); IR (NaCl/thin film): 3469, 3042, 2918, 2848, 1577, 1472, 1460, 1212, 1178, 1109, 928 cm⁻¹; HRMS (MM) calc'd for [M+H]⁺ 212.0870, found 212.0869.

Preparation of 2-(ethylphthalimide)indole (6v)



2-(ethylphthalimide)indole (1v) was prepared by an analogous procedure to that reported by Sakai et. al.⁸ A flame-dried flask was charged with 2-iodoaniline (**S6**, 500 mg, 2.30 mmol, 1.00 equiv), 2-(but-3-yn-1-yl)isoindoline-1,3-dione (**S8**, 550 mg, 2.75 mmol, 1.20 equiv),

⁸ Sakai, N.; Annaka, K.; Fujita, A.; Sato, A.; Konakahara, T. J. Org. Chem. 2008, 73, 4160.

Pd(PPh₃)₂Cl₂ (32 mg, 0.05 mmol, 0.02 equiv), copper (I) iodide (4.5 mg, 0.025 mmol, 0.01 equiv) and Et₃N (8 mL). The mixture was stirred overnight at room temperature, then filtered through a plug of silica, concentrated and redissolved in PhMe (10 mL). InBr₃ (40 mg, 0.1 mmol, 0.05 equiv) was added in one portion and the mixture was heated to 110 °C for 5 h, then cooled to room temperature, filtered through celite and concentrated. The crude residue was purified by silica gel chromatography (60:40 EtOAc:hexanes) to yield 302 mg (45% yield) of 2- (ethylphthalimide)indole (**6v**) as a light yellow solid. ¹H NMR (500 MHz, CDCl₃) δ 8.26 (br s, 1H), 7.83 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.71 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.51 (d, J = 7.8 Hz, 1H), 7.33 (d, J = 8.1 Hz, 1H), 7.13 (ddd, J = 8.2, 7.1, 1.2 Hz, 1H), 7.06 (ddd, J = 7.5, 7.5, 1.0 Hz, 1H), 6.33 (d, J = 1.2 Hz, 1H), 4.06 (t, *J* = 7.5 Hz, 2H), 3.21 (t, J = 7.4 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 168.3, 136.1, 134.9, 134.1, 131.9, 128.6, 123.4, 121.4, 120.0, 119.7, 110.6, 101.1, 37.1, 27.4.; IR (NaCl/thin film): 3366, 1772, 1707, 1653, 1617, 1466, 1395, 1363, 1293 cm⁻¹; HRMS (MM) calc'd for [M+H]⁺ 291.1128, found 291.1138.

3. Optimization of Reaction Parameters.

a. General Procedure 1

An oven-dried vial was charged with 2-phenylindole (**6a**, 0.20 mmol, 1.00 equiv), the acrylate (0.24 mmol, 1.20 equiv) and an (*R*)-BINOL derivative and pumped into a glove box. The vial was charged with solvent to an indole concentration of 0.12 M, and SnCl₄ (1.00 equiv, as a 1.0 M solution in DCM) was added. The reaction was stirred at 20 °C for 2 hours, after which time it was removed from the glove box and quenched by dilution with 1 M HCl (5 mL) and MeCN (1 mL). The aqueous layer was extracted with EtOAc (2 x 5 mL) and the combined organic layers were washed with saturated aqueous NaHCO₃ (5 mL), dried (Na₂SO₄), filtered, and concentrated. The crude residue was purified by silica gel chromatography.

Additive screens. Reactions were performed following General Procedure 1 using 0.20 equiv (*R*)-BINOL. After the vial was pumped into the glove box, one of the following additives was added:

- flame-dried powdered 4Å molecular sieves (200 wt % relative to indole)
- $K_2CO_3(1.00 \text{ equiv})$

• 2,6-lutidine (1.00 equiv)

Upon addition of the additive, DCM was added to an indole concentration of 0.12 M and the reaction was further conducted as described above.

Catalyst screens. Reactions were performed following General Procedure 1 using flame-dried powdered 4Å molecular sieves (200 wt % relative to indole) as an additive and DCM as a solvent.

b. Characterization Data

(S)- N_{α} -Trifluoroacetyl-2-phenyltryptophan benzyl ester (7a)



Prepared from benzyl 2-trifluoroacetamidoacrylate⁹ (2a, 65.5 mg, 0.24 mmol) following General Procedure 1. The crude residue was purified by silica gel chromatography (30:70 to 70:30 DCM:hexanes) to yield 11.1 mg (12% yield) of 7a as a yellow solid. The enantiomeric excess was

determined to be 35% by chiral SFC analysis (OB-H, 2.5 mL/min, 15% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 11.0 min, $t_{\rm R}$ (minor) = 12.9 min. ¹H NMR (500 MHz, CDCl₃) δ 8.14 (br s, 1H), 7.57 (ddd, J = 7.9, 1.8, 0.7 Hz, 1H), 7.54 – 7.50 (m, 2H), 7.50 – 7.45 (m, 2H), 7.42 – 7.36 (m, 2H), 7.34 – 7.29 (m, 3H), 7.24 (ddd, J = 8.1, 7.1, 1.1 Hz, 1H), 7.16 (ddd, J = 8.0, 7.1, 1.0 Hz, 1H), 7.11 – 7.07 (m, 2H), 6.67 (br d, J = 7.6 Hz, 1H), 4.95 (d, J = 12.2 Hz, 1H), 4.88 (dt, J = 7.8, 6.0 Hz, 1H), 4.53 (d, J = 12.2 Hz, 1H), 3.65 – 3.56 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ ¹³C NMR (125 MHz, CDCl₃) δ 170.1, 156.6 (q, $J_{C-F} = 37.8$ Hz), 136.3, 135.6, 134.6, 132.4, 129.2, 128.9, 128.5, 128.44, 128.38, 128.2, 128.1, 122.8, 120.3, 118.6, 115.3 (q, $J_{C-F} = 287.9$ Hz), 111.0, 105.6, 67.5, 53.3, 26.7; IR (NaCl/thin film): 3391, 3061, 2924, 1714, 1542, 1457, 1210, 1173 cm⁻¹; [α]_D²⁵ = +3.5 (c = 0.44, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 467.1577, found 467.1580.

⁹ Synthesis of benzyl 2-trifluoroacetamidoacrylate (2a): Crossley, M.; Stamford, A. Aust. J. Chem. 1994, 47, 1695.

(S)- N_{α} -Trifluoroacetyl-2-phenyltryptophan methyl ester (7b)



Prepared from methyl 2-trifluoroacetamidoacrylate¹⁰ (**2b**, 47.3 mg, 0.24 mmol) following General Procedure 1. The crude residue was purified by silica gel chromatography (0:100 to 5:95 EtOAc:toluene, then 0:100 to 20:80 EtOAc:hexanes) to yield 9.0 mg (12% yield) of **7b** as a yellow

solid. The enantiomeric excess was determined to be 42% by chiral SFC analysis (AS-H, 2.5 mL/min, 10% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 8.7 min, $t_{\rm R}$ (minor) = 7.7 min. ¹H NMR (500 MHz, CDCl₃) δ 8.17 (br s, 1H), 7.58 – 7.52 (m, 3H), 7.52 – 7.47 (m, 2H), 7.43 – 7.39 (m, 1H), 7.38 (ddd, J = 8.1, 0.9, 0.9 Hz, 1H), 7.23 (ddd, J = 8.2, 7.0, 1.2 Hz, 1H), 7.16 (ddd, J = 8.0, 7.0, 1.0 Hz, 1H), 6.65 (br d, J = 7.3 Hz, 1H), 4.83 (dt, J = 7.8, 5.6 Hz, 1H), 3.66 – 3.56 (m, 2H), 3.34 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.5, 156.6 (q, $J_{C-F} = 37.7$ Hz), 136.3, 135.6, 132.5, 129.2, 129.0, 128.4, 128.2, 122.8, 120.3, 118.5, 115.3 (q, $J_{C-F} = 287.7$ Hz), 111.0, 105.5, 53.2, 52.5, 26.4; IR (NaCl/thin film): 3391, 3057, 2917, 2849, 1718, 1542, 1458, 1449, 1211, 1170 cm⁻¹; $[\alpha]_{\rm D}^{25} = +22.3$ (c = 0.39, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 391.1264, found 391.1267.

¹⁰ Synthesis of methyl 2-trifluoroacetamidocacrylate (**2b**): Navarre, L.; Martinez, R.; Genet, J.; Darses, S. *J. Am. Chem. Soc.* **2008**, *130*, 6159.

3. Optimized Conjugate Addition/Asymmetric Protonation.

a. General Procedure 2

An oven-dried vial was charged with the indole (1.00 equiv), methyl 2-acetamidoacrylate (2c, 1.20 equiv)¹¹ and (R)-3,3'-dibromo-BINOL (9f, 0.20 equiv) and pumped into a glove box. To the vial was added flame-dried powdered 4Å molecular sieves (200 wt % relative to indole). The vial was charged with DCM to an indole concentration of 0.12 M, and SnCl₄ (1.00 equiv unless specifically indicated, as a 1 M solution in DCM) was added. The reaction was stirred at 20 °C for 2 hours, after which time it was removed from the glove box and quenched by dilution with 1 M HCl (5 mL) and MeCN (1 mL). The aqueous layer was extracted with EtOAc (2 x 5 mL) and the combined organic layers were washed with saturated aqueous NaHCO₃ (5 mL), dried (Na₂SO₄), filtered, and concentrated. The crude residue was purified by silica gel chromatography.

b. Characterization Data

(S)- N_{α} -Acetyl-2-phenyltryptophan methyl ester (7c)



Prepared from 2-phenylindole (**6a**, 19.0 mg, 0.10 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 25.6 mg (76% yield) of **7c** as a white foam. The enantiomeric excess was determined to

be 93% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm). $t_R(major) = 5.7 \text{ min}, t_R(minor) = 6.9 \text{ min}. [\alpha]_D^{25} = +37.7 (c = 0.94, CHCl_3)$. Spectral data matches that reported in the literature.¹²

(S)- N_{α} -Acetyl-1-methyl-2-phenyltryptophan methyl ester (7d)



Prepared from 1-methyl-2-phenylindole (**6b**, 41.4 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (0:100 to 55:45 EtOAc:hexanes) to yield 43.4 mg (63% yield) of **7d** as a yellow solid. The enantiomeric excess was

¹¹ Methyl 2-acetamidoacrylate (**2c**) is commercially available, or can be prepared according to Crestey, F.; Collot, V.; Steibing, S.; Rault, S. *Synthesis* **2006**, *20*, 3506.

¹² Angelini, E.; Balsamini, C.; Bartoccini, F.; Lucarini, S.; Piersanti, G. J. Org. Chem. 2008, 73, 5654.

determined to be 85% by chiral SFC analysis (AD-H, 2.5 mL/min, 20% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}({\rm major}) = 4.6$ min, $t_{\rm R}({\rm minor}) = 3.9$ min. ¹H NMR (500 MHz, CDCl₃) δ 7.60 (ddd, J = 7.9, 1.2, 0.7 Hz, 1H), 7.56 – 7.49 (m, 2H), 7.48 – 7.44 (m, 1H), 7.42 – 7.38 (m, 2H), 7.34 (ddd, J = 8.2, 0.9, 0.9 Hz, 1H), 7.26 (ddd, J = 8.2, 7.0, 1.2 Hz, 1H), 7.17 (ddd, J = 8.0, 7.0, 1.1 Hz, 1H), 5.72 (br d, J = 7.8 Hz, 1H), 4.74 (dt, J = 8.0, 5.6 Hz, 1H), 3.57 (s, 3H), 3.39 (s, 3H), 3.41 (dd, J = 14.7, 5.7 Hz, 1H), 3.34 (dd, J=14.8, 5.6 Hz, 1H), 1.73 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 169.5, 139.2, 136.9, 131.6, 130.7, 128.7, 128.4, 127.9, 122.0, 119.7, 118.7, 109.5, 106.7, 52.8, 52.0, 30.8, 26.6, 23.0.; IR (NaCl/thin film): 3288, 3055, 2950, 1743, 1657, 1539, 1469, 1441, 1368, 1238, 1212 cm⁻¹; [α]_D²⁵ = +21.3 (c = 0.91, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 351.1703, found 351.1708.

(S)- N_{α} -Acetyl-1-allyl-2-phenyltryptophan methyl ester (7e)



Prepared from 1-allyl-2-phenylindole (**6c**, 46.6 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (0:100 to 55:45 EtOAc:hexanes) to yield 51.3 mg (68% yield) of **7e** as a yellow foam. The enantiomeric excess was determined to be 85% by chiral SFC analysis (AS-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda =$

254 nm): $t_{\rm R}$ (major) = 2.9 min, $t_{\rm R}$ (minor) = 2.4 min. ¹H NMR (500 MHz, CDCl₃) δ 7.62 (ddd, J = 7.8, 1.0, 1.0 Hz, 1H), 7.53 – 7.47 (m, 2H), 7.47 – 7.42 (m, 1H), 7.42 – 7.37 (m, 2H), 7.30 (ddd, J = 8.1, 0.9, 0.9 Hz, 1H), 7.23 (ddd, J = 8.2, 7.0, 1.2 Hz, 1H), 7.17 (ddd, J = 8.0, 7.0, 1.1 Hz, 1H), 5.85 (ddt, J = 17.1, 10.3, 4.7 Hz, 1H), 5.76 (br d, J = 7.9 Hz, 1H), 5.11 (dtd, J = 10.4, 1.7, 1.2 Hz, 1H), 4.82 (dtd, J = 17.1, 1.9, 1.3 Hz, 1H), 4.76 (dt, J = 8.0, 5.8 Hz, 1H), 4.56 (dt, J = 4.7, 1.8 Hz, 2H), 3.39 (s, 3H), 3.36 (dd, J = 14.7, 5.7 Hz, 1H), 3.29 (dd, J = 14.7, 5.9 Hz, 1H), 1.75 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 169.5, 139.0, 136.3, 133.5, 131.5, 130.5, 128.7, 128.5, 128.1, 122.0, 119.8, 118.8, 116.3, 110.2, 107.2, 52.8, 52.0, 46.3, 26.8, 23.0; IR (NaCl/thin film): 3435, 3287, 3056, 2950, 2926, 2851, 1744, 1658, 1538, 1500, 1408, 1367, 1219, 1196, 1134; $[\alpha]_{\rm D}^{25}$ = +13.8 (c = 2.96, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 377.1860, found 377.1865.

(S)- N_{α} -Acetyl-4-methyl-2-phenyltryptophan methyl ester (7f)



Prepared from 4-methyl-2-phenylindole¹³ (**6d**, 21.0 mg, 0.10 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 30.8 mg (88% yield) of **7f** as a white foam. The enantiomeric excess was

determined to be 96% by chiral SFC analysis (AD-H, 2.5 mL/min, 25% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 9.9 min, $t_{\rm R}$ (minor) = 8.9 min. ¹H NMR (500 MHz, CDCl₃) δ 8.32 (br s, 1H), 7.55 – 7.45 (m, 4H), 7.44 – 7.37 (m, 1H), 7.19 (d, J = 8.0 Hz, 1H), 7.08 (m, 1H), 6.91 (m, 1H), 5.44 (br d, J = 7.6 Hz, 1H), 4.63 (td, J = 8.2, 5.0 Hz, 1H), 3.69 – 3.45 (m, 2H), 3.44 (s, 3H), 2.78 (s, 3H), 1.64 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.3, 169.7, 136.3, 136.1, 133.1, 130.5, 129.2, 128.9, 128.3, 126.9, 122.5, 122.3, 109.0, 107.6, 54.2, 52.1, 27.6, 22.8, 20.5; IR (NaCl/thin film): 3295, 3052, 2952, 1741, 1659, 1602, 1547, 1514, 1492, 1449, 1372, 1218; [α]_D²⁵ = -29.0 (c = 0.63, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 351.1703, found 351.1698.

(S)-Na-Acetyl-5-methyl-2-phenyltryptophan methyl ester (7g)



Prepared from 5-methyl-2-phenylindole⁸ (6e, 42.0 mg, 0.20 mmol)
 NHAC following General Procedure 2. The crude residue was purified by silica gel chromatography (30:70 to 100:0 EtOAc:hexanes) to yield 58.0 mg (83% yield) of 7g as a white foam. The enantiomeric excess

was determined to be 95% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 4.9 min, $t_{\rm R}$ (minor) = 6.4 min. ¹H NMR (500 MHz, CDCl₃) δ 8.12 (br s, 1H), 7.54 (ddd, J = 10.1, 6.1, 4.2 Hz, 2H), 7.50 – 7.43 (m, 2H), 7.40 – 7.33 (m, 2H), 7.24 (d, J = 8.3 Hz, 1H), 7.06 – 7.00 (m, 1H), 5.78 (br d, J = 8.1 Hz, 1H), 4.83 (dt, J = 8.1, 5.4 Hz, 1H), 3.53 – 3.51 (m, 2H), 3.31 (s, 3H), 2.46 (s, 3H), 1.66 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.1, 169.5, 136.1, 134.0, 133.3, 129.7, 129.2, 129.1, 128.2, 128.0, 124.1, 118.6, 110.6, 106.3, 52.7, 51.2, 26.5, 22.8, 21.5; IR (NaCl/thin film): 3379, 3365, 2948, 1737, 1658, 1439, 1372, 1306, 1217 cm⁻¹; $[\alpha]_{\rm D}^{25} = +33.8$ (c = 0.26, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 351.1703, found 351.1680.

¹³ Zhao, J.; Zhang, Y.; Cheng, K. J. Org. Chem. 2008, 73, 7428.

(S)- N_{α} -Acetyl-6-methyl-2-phenyltryptophan methyl ester (7h)



Prepared from 6-methyl-2-phenylindole¹³ (**6f**, 21.0 mg, 0.10 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 27.9 mg (80% yield) of **7h** as a colorless oil. The enantiomeric

excesses was determined to be 89% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 9.1$ min, $t_R(minor) = 10.1$ min. ¹H NMR (500 MHz, CDCl₃) δ 8.01 (br s, 1H), 7.55 (ddd, J = 5.8, 4.0, 2.1 Hz, 2H), 7.48 – 7.44 (m, 3H), 7.39 – 7.33 (m, 1H), 7.14 (s, 1H), 6.97 (dd, J = 8.3, 1.5 Hz, 1H), 5.78 (br d, J = 7.8 Hz, 1H), 4.83 (dt, J = 8.0, 5.4 Hz, 1H), 3.55 – 3.49 (m, 2H), 3.30 (s, 3H), 2.47 (s, 3H), 1.67 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) 172.1, 169.6, 136.1, 135.2, 133.3, 132.4, 129.1, 128.2, 127.9, 127.3, 121.8, 118.5, 110.9, 106.5, 52.7, 52.0, 26.6, 22.9, 21.7; IR (NaCl/thin film): 3292, 3052, 2958, 2908, 1741, 1658, 1545, 1530, 1511, 1446, 1375, 1216; $[\alpha]_D^{25} = +39.3$ (c = 0.38, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 351.1703, found 351.1698.

(S)- N_{α} -Acetyl-7-methyl-2-phenyltryptophan methyl ester (7i)



Prepared from 7-methyl-2-phenylindole¹³ (**6g**, 21.0 mg, 0.10 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (30:70 to 100:0 EtOAc:hexanes) to yield 33.0 mg

Me⁶ H (94% yield) of 7i as a white foam. The enantiomeric excess was determined to be 94% by chiral SFC analysis (AD-H, 2.5 mL/min, 25% IPA in CO₂, λ = 254 nm): *t*_R(major) = 5.6 min, *t*_R(minor) = 5.0 min. ¹H NMR (500 MHz, CDCl₃) δ 8.23 (br s, 1H), 7.61 – 7.54 (m, 2H), 7.51 – 7.45 (m, 2H), 7.42 (d, *J* = 8.1 Hz, 1H), 7.40 – 7.35 (m, 1H), 7.11 – 7.04 (m, 1H), 7.03 – 6.97 (m, 1H), 5.79 (br d, *J* = 8.1 Hz, 1H), 4.82 (dt, *J* = 8.1, 5.7 Hz, 1H), 2.55 (dd, *J* = 12.5, 3.1 Hz, 1H), 3.51 (dd, *J* = 12.5, 3.1 Hz, 1H), 3.30 (s, 3H), 2.50 (s, 3H), 1.65 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.1, 169.6, 135.8, 135.3, 133.3, 129.1, 128.9, 128.4, 128.0, 123.1, 120.20, 120.18, 116.5, 107.1, 52.7, 51.9, 26.6, 22.8, 16.6; IR (NaCl/thin film): 3283, 3053, 2950, 1736, 1659, 1518, 1438, 1372, 1306, 1266, 1219, 1137, 1043; [α]_D²⁵ = +26.5 (*c* = 0.20, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 351.1703, found 351.1708.

(S)- N_{α} -Acetyl-5-methoxy-2-phenyltryptophan methyl ester (7j)



Prepared from 5-methoxy-2-phenylindole¹³ (6h, 45.0 mg, 0.20 mmol)
following General Procedure 2. The crude residue was purified by
silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield
62.0 mg (85% yield) of 7j as a colorless oil. The enantiomeric excess

was determined to be 91% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 4.7 min, $t_{\rm R}$ (minor) = 6.5 min. ¹H NMR (500 MHz, CDCl₃) δ 8.24 (br s, 1H), 7.58 – 7.49 (m, 2H), 7.50 – 7.41 (m, 2H), 7.36 (dd, J = 7.4, 7.4 Hz, 1H), 7.24 (d, J = 8.7 Hz, 1H), 7.05 (d, J = 2.3 Hz, 1H), 6.90 – 6.80 (m, 1H), 5.82 (br d, J = 7.9 Hz, 1H), 4.82 (td, J = 7.9, 5.4 Hz, 1H), 3.87 (s, 3H), 3.49 (m, 2H), 3.29 (s, 3H), 1.67 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 169.6, 154.4, 136.7, 133.2, 130.8, 129.8, 129.1, 128.2, 128.0, 112.7, 111.7, 106.5, 100.5, 55.9, 52.7, 52.0, 26.6, 22.9; IR (NaCl/thin film): 3291, 3057, 2926, 1739, 1652, 1558, 1539, 1520, 1483, 1455, 1374, 1218, 1178; $[\alpha]_{\rm D}^{25} = +32.6$ (c = 0.93, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 367.1652, found 367.1658.

(S)-N_a-Acetyl-5-bromo-2-phenyltryptophan methyl ester (7k)



Prepared from 5-bromo-2-phenylindole¹⁴ (**6i**, 54.0 mg, 0.20 mmol) with 1.6 equiv SnCl₄ following General Procedure 2. The crude residue was purified by silica gel chromatography (30:70 to 100:0 EtOAc:hexanes) to yield 49.5 mg (60% yield) of **7k** as a white foam.

The enantiomeric excess was determined to be 93% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 5.3$ min, $t_R(minor) = 7.9$ min. ¹H NMR (500 MHz, CDCl₃) δ 8.42 (br s, 1H), 7.66 (d, J = 2.0 Hz, 1H), 7.56 – 7.50 (m, 2H), 7.49 – 7.43 (m, 2H), 7.42 – 7.34 (m, 1H), 7.28 – 7.24 (m, 1H), 7.22 – 7.18 (m, 1H), 5.75 (br d, J = 8.1 Hz, 1H), 4.82 (dt, J = 8.1, 5.7 Hz, 1H), 3.53 (dd, J = 14.9, 5.5 Hz, 1H), 3.46 (dd, J = 14.9, 4.8 Hz, 1H), 3.36 (s, 3H), 1.63 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 171.9, 169.6, 137.2, 134.2, 132.6, 131.1, 129.2, 128.3, 128.2, 125.2, 121.6, 113.1, 112.4, 106.4, 52.6, 52.1, 26.5, 22.8; IR (NaCl/thin film): 3417, 3369, 3282, 1734, 1654, 1521, 1466, 1437, 1374, 1215; [α]_D²⁵ = +47.2 (c = 1.04, CHCl₃).

¹⁴ Prepared from 4-bromo-2-iodoaniline by an analogous procedure to that reported by Sakai et. al. (reference 8). Spectral data matches that reported in the literature: Homes, T. P.; Mattner, F.; Keller, P. A.; Katsifis, A. *Bioorg. Med. Chem.* **2006**, *14*, 3938.

HRMS (MM) calc'd for [M+H]⁺ 415.0652, found 415.0653.

(S)- N_{α} -Acetyl-5-fluoro-2-phenyltryptophan methyl ester (7l)



Prepared from 5-fluoro-2-phenylindole¹³ (**6j**, 42.0 mg, 0.20 mmol) with 1.6 equiv SnCl₄ following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 44.7 mg (63% yield) of **7l** as a colorless oil. The enantiomeric

excess was determined to be 92% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 3.8$ min, $t_R(minor) = 5.2$ min. ¹H NMR (500 MHz, CDCl₃) δ 8.30 (br s, 1H), 7.60 – 7.52 (m, 2H), 7.50 – 7.43 (m, 2H), 7.42 – 7.34 (m, 1H), 7.27 – 7.24 (m, 1H), 7.21 (dd, J = 9.8, 2.6 Hz, 1H), 6.94 (ddd, J = 9.0, 9.0, 2.6 Hz, 1H), 5.77 (br d, J = 7.8 Hz, 1H), 4.82 (dt, J = 8.1, 5.4 Hz, 1H), 3.53 (dd, J = 14.9, 5.6 Hz, 1H), 3.47 (dd, J = 14.9, 5.0 Hz, 1H), 3.35 (s, 3H), 1.64 (s, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 172.7, 169.8, 168.3, 135.6, 134.2, 132.5, 131.9, 128.6, 123.5, 121.8, 119.7, 118.2, 110.8, 107.4, 52.9, 52.4, 37.0, 27.0, 25.3, 23.1; IR (NaCl/thin film): 3275, 3062, 2952, 1733, 1652, 1584, 1558, 1539, 1520, 1486, 1456, 1436, 1374, 1266, 1217, 1180; $[\alpha]_D^{25} = +49.9$ (c = 1.25, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 355.1452, found 355.1455.

(S)-N_α-Acetyl-2-(4-methylphenyl)tryptophan methyl ester (7m)



Prepared from 2-(4-methylphenyl)indole¹⁵ (**6k**, 41.0 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 60.1 mg (86% yield) of **7m** as a white foam. The enantiomeric excess was

determined to be 94% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm). $t_{\rm R}$ (major) = 6.6 min, $t_{\rm R}$ (minor) = 8.8 min. ¹H NMR (500 MHz, CDCl₃) δ 8.20 (br s, 1H), 7.56 (d, J = 8.1 Hz, 1H), 7.45 (d, J = 8.1, 2H), 7.34 (d, J = 8.1, 1H), 7.28 (d, J = 8.1, 2H), 7.19 (ddd, J = 7.8, 7.1, 1.2 Hz, 1H), 7.15 – 7.09 (m, 1H), 5.77 (br d, J = 8.1, 1H), 4.82 (dt, J = 7.8, 5.5 Hz, 1H), 3.54 (dd, J = 13.1, 4.0 Hz, 1H), 3.50 (dd, J = 13.1, 3.7 Hz, 1H), 3.33 (s, 3H), 2.40 (s,

¹⁵ Prepared from 2-iodoaniline by an analogous procedure to that reported by Sakai et. al. (reference 8). Spectral data matches that reported in the literature: Shen, M.; Leslie, B. E.; Driver, T. G. *Angew. Chem., Int. Ed.* **2008**, *47*, 5056.

3H), 1.66 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 169.6, 138.0, 136.1, 135.6, 130.2, 129.8, 129.4, 128.1, 122.3, 119.9, 118.7, 110.9, 106.4, 52.8, 52.0, 26.6, 22.8, 21.2; IR (NaCl/thin film): 3365, 3271, 3052, 2951, 1737, 1657, 1519, 1460, 1439, 1375, 1305, 1217 cm⁻¹; $[\alpha]_D^{25} = 43.2$ (c = 0.74, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 351.1703, found 351.1700.

(S)- N_{α} -Acetyl-2-(2-methylphenyl)tryptophan methyl ester (7n)



Prepared from 2-(2-methylphenyl)indole¹⁶ (**6l**, 21.0 mg, 0.1 mmol) following General Procedure 2. The crude residue was purified by flash chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 9.2 mg (26% yield) of **7n**. The enantiomeric excess was determined to be 87% by chiral SFC analysis (AD-H, 2.5 mL/min, 25% IPA in CO₂, $\lambda = 254$ nm):

 $t_{\rm R}$ (major) = 4.3 min, $t_{\rm R}$ (minor) = 4.9 min. ¹H NMR (500 MHz, CDCl₃) δ 8.03 (br s, 1H), 7.62 – 7.55 (dd, J = 7.6, 0.9 Hz, 1H), 7.38 – 7.32 (m, 4H), 7.31 – 7.27 (m, 1H), 7.22 (ddd, J = 8.1, 5.6, 2.1 Hz, 1H), 7.16 (ddd, J = 7.1, 5.6, 1.1 Hz, 1H), 5.71 (br d, J = 7.9 Hz, 1H), 4.82 – 4.68 (dt, J = 7.9, 5.4 Hz, 1H), 3.38 – 3.29 (m, 4H), 3.28 – 3.16 (m, 1H), 2.28 (s, 3H), 1.73 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.1, 169.6, 137.3, 135.8, 135.5, 132.1, 130.9, 130.8, 128.9, 128.7, 126.0, 122.3, 119.9, 118.8, 110.8, 107.6, 52.8, 52.0, 26.6, 23.0, 20.0; IR (NaCl/thin film): 3385, 3271, 3062, 2924, 2853, 1734, 1653, 1559, 1539, 1521, 1457, 1437, 1374; [α]_D²⁵ = +21.5 (c = 0.29, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 351.1703, found 351.1709.

(S)-Na-Acetyl-2-(4-chlorophenyl)tryptophan methyl ester (70)



Prepared from 2-(4-chlorophenyl)indole¹³ (**6m**, 45.0 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 55.2 mg (75% yield) of **70** as a colorless oil. The enantiomeric excess was

determined to be 93% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 6.1 min, $t_{\rm R}$ (minor) = 7.0 min. ¹H NMR (500 MHz, CDCl₃) δ 8.45 (br s, 1H), 7.56 (d, J = 8.1 Hz, 1H), 7.49 – 7.43 (m, 2H), 7.43 – 7.37 (m, 2H), 7.33 (ddd, J = 8.1, 8.1, 1.0

¹⁶ Prepared from 2-iodoaniline by an analogous procedure to that reported by Sakai et. al. (reference 8). Spectral data matches that reported in the literature: Zhao, J.; Zhang, Y.; Cheng, K. *J. Org. Chem.* **2008**, *73*, 7428.

Hz, 1H), 7.23 – 7.18 (m, 1H), 7.14 (ddd, J = 8.0, 7.1, 1.1 Hz, 1H), 5.85 (br d, J = 8.1 Hz, 1H), 4.83 (dt, J = 8.1, 5.5 Hz, 1H), 3.55 – 3.38 (m, 2H), 3.34 (s, 3H), 1.69 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.1, 169.6, 135.8, 134.6, 133.9, 131.5, 129.4, 129.3, 122.7, 120.1, 118.9, 111.1, 107.1, 52.8, 52.1, 29.6, 26.7, 22.9; IR (NaCl/thin film): 3280, 3058, 2948, 1737, 1657, 1519, 1487, 1458, 1439, 1373, 1310, 1216, 1093 cm⁻¹; $[\alpha]_D^{25} = +40.8$ (c = 0.96, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 371.1157, found 371.1158.

(S)- N_{α} -Acetyl-2-(4-fluorophenyl)tryptophan methyl ester (7p)



Prepared from 2-(4-fluorophenyl)indole⁸ (**6n**, 42.0 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc/hexanes) to yield 55.6 mg (78% yield) of 7p as a colorless oil. The enantiomeric excess was

determined to be 92% by chiral SFC analysis (AD-H, 2.5 mL/min, 25% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 6.1 min, $t_{\rm R}$ (minor) = 6.9 min. ¹H NMR (500 MHz, CDCl₃) δ 8.19 (d, J = 47.9 Hz, 1H), 7.57 (dd, J = 7.9, 1.1 Hz, 1 H), 7.54 – 7.51 (m, 2H), 7.36 (ddd, J = 8.1, 8.1, 0.9 Hz, 1H), 7.23 – 7.10 (m, 4H), 5.82 (d, J = 8.1 Hz, 1H), 4.83 (dt, J = 8.1, 5.5 Hz, 1H), 3.55 – 3.40 (m, 2H), 3.34 (s, 3H), 1.71 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 169.5, 135.6, 135.0, 130.1, 130.1, 129.4, 122.7, 120.2, 118.9, 116.2, 116.1, 110.9, 106.9, 52.8, 52.0, 26.7, 22.9.; IR (NaCl/thin film): 3364, 3271, 3061, 2925, 2853, 1738, 1661, 1553, 1505, 1460, 1440, 1373, 1221, 1158; [α]_D²⁵ = +38.2 (*c* = 0.65, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 355.1452, found 355.1460.

(S)-Na-Acetyl-2-(3-fluorophenyl)tryptophan methyl ester (7q)



Prepared from 2-(3-fluorophenyl)indole¹⁵ (60, 42.0 mg, 0.20 mmol)
following General Procedure 2. The crude residue was purified by silica
gel chromatography (40:60 to 100:0 ethyl acetate/hexanes) to yield 50.6
mg (76% yield) of 7q as a white foam. The enantiomeric excess was
determined to be 92% by chiral SFC analysis (AD-H, 2.5 mL/min, 30%)

IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 3.8 min, $t_{\rm R}$ (minor) = 4.6 min. ¹H NMR (500 MHz, CDCl₃) δ 8.65 (br s, 1H), 7.57 (d, J = 8.1 Hz, 1H), 7.41 – 7.37 (m, 1H), 7.33-7.31 (m, 2H), 7.27-7.24 (m, 1H), 7.19 (ddd, J = 8.2, 7.0, 1.0 Hz, 1H), 7.13 (ddd, J = 7.9, 7.0, 1.0 Hz, 1H), 7.07 – 7.03 (m, 1H), 5.89 (br d, J = 8.1 Hz, 1H), 4.84 (dt, J = 8.1, 5.5 Hz, 1H), 3.53 (dd, J = 13.6, 4.7 Hz, 1H), 3.49 (dd, J = 13.6, 4.2 Hz, 1H), 3.34 (s, 3H), 1.69 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.1, 169.7, 162.9 (d, $J_{C-F} = 246.3$ Hz), 135.8, 135.2 (d, $J_{C-F} = 7.5$ Hz), 134.5 (d, $J_{C-F} = 2.5$ Hz), 130.6 (d, $J_{C-F} = 8.8$ Hz), 129.2, 123.9 (d, $J_{C-F} = 3.8$ Hz), 122.8, 120.0, 118.9, 115.1 (d, $J_{C-F} = 21.2$ Hz), 114.7 (d, $J_{C-F} = 21.2$ Hz), 111.1, 107.3, 52.8, 52.0, 26.7, 22.8; IR (NaCl/thin film): 3370, 3275, 3060, 2952, 1735, 1655, 1614, 1585, 1522, 1438, 1374, 1266, 1200, 1155 cm⁻¹; [α]_D²⁵ = +37.6 (*c* = 1.21, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 355.1452, found 355.1450.

(S)- N_{α} -Acetyl-2-(2-fluorophenyl)tryptophan methyl ester (7r)



Prepared from 2-(2-fluorophenyl)indole (**6p**, 21.0 mg, 0.10 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 12.4 mg (35% yield) of **7r**. The enantiomeric excesses was determined to be 92%

by chiral SFC analysis (AD-H, 2.5 mL/min, 25% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 9.5$ min, $t_R(minor) = 8.4$ min. ¹H NMR (500 MHz, CDCl₃) δ 8.28 (s, 1H), 7.61 (d, J = 7.9 Hz, 1H), 7.55 (ddd, J = 7.5, 7.5, 1.8 Hz, 1H), 7.45 – 7.35 (m, 2H), 7.29 (ddd, J = 7.5, 7.5, 1.2 Hz, 1H), 7.25 – 7.20 (m, 1H), 7.19 – 7.10 (m, 1H), 5.83 (br d, J = 7.6 Hz, 1H), 4.85 (dt, J = 7.9, 5.5 Hz, 1H), 3.55 – 3.39 (m, 2H), 3.36 (s, 2H), 1.73 (s, 3H).; ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 169.5, 159.8 (d, $J_{C-F} = 246.3$ Hz), 135.9, 131.4 (d, $J_{C-F} = 3.8$ Hz) 130.2 (d, $J_{C-F} = 8.8$ Hz), 129.73, 128.65, 124.8 (d, $J_{C-F} = 3.8$ Hz), 122.84, 120.6 (d, $J_{C-F} = 15.0$ Hz), 120.0, 119.0, 116.4 (d, $J_{C-F} = 21.3$ Hz), 111.0, 108.8, 52.5, 52.0, 26.8, 26.8, 22.9; IR (NaCl/thin film): 3275, 3058, 2925, 2853, 1734, 1653, 1523, 1490, 1457, 1437, 1374, 1245, 1216, 1130, 1104; [α]_D²⁵ = +39.8 (*c* = 0.41, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 355.1452, found 355.1463.

(S)- N_{α} -Acetyl-2-(3-methoxyphenyl)tryptophan methyl ester (7s)



Prepared from 2-(3-methoxyphenyl)indole¹⁷ (6q, 45.0 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (30:70 to 100:0 EtOAc:hexanes) to yield 65.0 mg

¹⁷ Prepared from 2-iodoaniline by an analogous procedure to that reported by Sakai et. al. (reference 8). Spectral data matches that reported in the literature: Yang, S.-D.; Sun, C. L.; Fang, Z.; Li, B.-J.; Li, Y.-Z.; Shi, Z.-J. *Angew. Chem., Int. Ed.* **2008**, *47*, 1473.

(88% yield) of **7s** as a colorless oil. The enantiomeric excess was determined to be 92% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 5.9$ min, $t_R(minor) = 7.6$ min. ¹H NMR (500 MHz, CDCl₃) δ 8.40 (br s, 1H), 7.55 (d, J = 8.1 Hz, 1H), 7.40 – 7.31 (m, 2H), 7.19 (ddd, J = 8.1, 7.1, 1.2 Hz, 1H), 7.16 – 7.10 (m, 2H), 7.08 (dd, J = 2.6, 1.6 Hz, 1H), 6.91 (ddd, J = 8.3, 2.6, 0.8 Hz, 1H), 5.82 (br d, J = 7.8 Hz, 1H), 4.83 (dt, J = 7.8, 5.5 Hz, 1H), 3.85 (s, 3H), 3.57 – 3.49 (m, 2H), 3.35 (s, 3H), 1.65 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.2, 169.6, 160.0, 135.8, 135.6, 134.4, 130.2, 129.3, 122.5, 120.6, 119.9, 118.8, 113.8, 113.5, 111.0, 106.7, 55.4, 52.8, 52.0, 26.6, 22.8; IR (NaCl/thin film): 3282, 3058, 2951, 1738, 1658, 1603, 1520, 1462, 1439, 1373, 1218, 1040; $[\alpha]_D^{25} = +40.3$ (c = 1.16, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 367.1652, found 367.1656.

(S)-N_α-Acetyl-2-methyltryptophan methyl ester (7t)



Prepared from 2-methylindole (**6r**, 26.0 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (50:50 to 100:0 EtOAc:hexanes) to yield 31.0 mg (61% yield) of **7t** as a white foam. The enantiomeric excess was determined to

be 85% by chiral SFC analysis (AD-H, 2.5 mL/min, 25% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 3.9 min, $t_{\rm R}$ (minor) = 2.7 min. [α]_D²⁵ = +25.9 (c = 0.99, CHCl₃). Spectral data matches that reported in the literature.⁹

(S)- N_{α} -Acetyl-2-butyltryptophan methyl ester (7u)



Prepared from 2-butylindole¹⁸ (**6s**, 35.0 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 45.8 mg (72% yield) of **7u** as a colorless oil. The enantiomeric excess was determined to

be 91% by chiral SFC analysis (AD-H, 2.5 mL/min, 20% IPA in CO₂, $\lambda = 254$ nm): t_R (major) = 5.1 min, t_R (minor) = 4.2 min. ¹H NMR (500 MHz, CDCl₃) δ 8.03 (br s, 1H), 7.46 – 7.40 (m, 1H), 7.31 – 7.24 (m, 1H), 7.15 – 6.99 (m, 2H), 6.00 (br d, J = 7.8 Hz, 1H), 4.88 (dt, J = 8.1, 5.7 Hz,

¹⁸ Prepared from 2-iodoaniline by an analogous procedure to that reported by Sakai et. al. (reference 8). Spectral data matches that reported in the literature: Ambrogio, I.; Cacchi, S.; Fabrizi, G.; Prastaro, A. *Tetrahedron* **2009**, *65*, 8916.

1H), 3.65 (s, 3H), 3.26 (dd, J = 5.7, 0.9 Hz, 2H), 2.69 (td, J = 7.8 2.2 Hz, 2H), 1.93 (s, 3H), 1.66 – 1.57 (m, 2H), 1.45 – 1.31 (m, 2H), 0.95 (t, J = 7.3 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.6, 169.6, 137.4, 135.2, 128.8, 121.3, 119.5, 117.9, 110.4, 105.26, 105.29, 53.0, 52.3, 31.8, 26.8, 25.7, 23.2, 22.6, 13.9; IR (NaCl/thin film): 3296, 3058, 2955, 2871, 1737, 1658, 1562, 1530, 1463, 1439, 1376, 1217, 1129; $[\alpha]_D^{25} = +16.3$ (c = 0.83, CHCl₃). HRMS (MM) calc'd for $[M+H]^+$ 317.1860, found 317.1855.

(S)-Na-Acetyl-2-isopropyltryptophan methyl ester (7v)



Prepared from 2-isopropylindole¹⁹ (**6t**, 32.0 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 39.6 mg (66% yield) of 7v as a colorless oil. The enantiomeric excess was determined to

be 92% by chiral SFC analysis (AD-H, 2.5 mL/min, 15% IPA in CO₂, $\lambda = 254$ nm). t_R (major) = 6.4 min, t_R (minor) = 5.6 min. ¹H NMR (500 MHz, CDCl₃) δ 8.16 (br s, 1H), 7.48 – 7.41 (m, 1H), 7.30 – 7.27 (m, 1H), 7.15 – 7.02 (m, 2H), 6.04 (br d, J = 8.0 Hz, 1H), 4.89 (dt, J = 8.1, 5.7 Hz, 1H), 3.66 (s, 3H), 3.29 (dd, J = 12.7, 4.0 Hz, 1H), 3.26 (dd, J = 12.7, 3.4 Hz, 1H), 3.18 (m, 1H), 1.93 (s, 3H1.31 (d, J = 3.3 Hz, 3H), 1.30 (d, J = 3.3 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.6, 169.7, 142.7, 135.2, 128.7, 121.3, 119.5, 117.9, 110.6, 103.6, 53.0, 52.3, 26.7, 25.3, 23.2, 23.0; IR (NaCl/thin film): 3305, 2962, 1734, 1700, 1653, 1559, 1539, 1506, 1457, 1436, 1374, 1299, 1217 cm⁻¹; [α]_D²⁵ = +22.2 (*c* = 0.35, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 303.1703, found 303.1709.

(S)- N_{α} -Acetyl-2-(tert-butyl)tryptophan methyl ester (7w)



Prepared from 2-(tert-butyl)indole¹³ (**6u**, 35.0 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 18.1 mg (29% yield) of **7w** as a yellow oil. The enantiomeric excess was determined to

be 84% by chiral SFC analysis (OD-H, 2.5 mL/min, 10% IPA in CO₂, $\lambda = 254$ nm): t_R (major) =

¹⁹ Prepared from 2-iodoaniline by an analogous procedure to that reported by Sakai et. Al (reference 8). Spectral data matches that reported in the literature: Smith, A. B.; Visnick, M.; Haseltine, J. N.; Sprengeler, P. A. *Tetrahedron* **2007**, *42*, 2957.

12.8 min, $t_{\rm R}$ (minor) = 14.2 min. ¹H NMR (500 MHz, CDCl₃) δ 8.07 (br s, 1H), 7.47 (dd, J = 14.0, 7.1 Hz, 1H), 7.27 (dd, J = 5.8, 4.8 Hz, 1H), 7.15 – 7.03 (m, 2H), 6.06 (br d, J = 7.4 Hz, 1H), 4.84 (m, 1H), 3.54 (s, 3H), 3.38 – 3.29 (m, 2H), 1.86 (s, 3H), 1.49 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 173.2, 169.6, 143.4, 133.9, 129.8, 121.3, 119.4, 117.7, 110.4, 104.3, 53.7, 52.2, 33.2, 30.7, 28.6, 23.0; IR (NaCl/thin film): 3326, 3047, 2961, 2918, 2868, 1734, 1653, 1539, 1457, 1436, 1374, 1303, 1254, 1211, 1128; $[\alpha]_{\rm D}^{25}$ = +12.4 (*c* = 0.36, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 317.1860, found 317.1856.

(S)- N_{α} -Acetyl-2-(ethylphthalimide)tryptophan methyl ester (7x)



Prepared from 2-(ethylphthalimide)indole (6v, 29.0 mg, 0.10 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (70:30 to 100:0 EtOAc:hexanes) to yield 34.6 mg (80% yield) of 7x as a yellow foam. The enantiomeric excess was determined to be 90% by chiral SFC analysis (AD-H, 2.5 mL/min, 25%)

IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 7.3$ min, $t_R(minor) = 6.3$ min. ¹H NMR (500 MHz, CDCl₃) δ 8.47 (br s, 1H), 7.83 (dd, J = 5.4, 2.9 Hz, 2H), 7.72 (dd, J = 5.5, 3.1 Hz, 2H), 7.46 (d, J = 8.1 Hz, 1H), 7.31 (ddd, J = 8.1, 8.1, 1.0 Hz, 1H), 7.13 (ddd, J = 8.1, 7.1, 1.2 Hz, 1H), 7.07 (ddd, J = 10.5, 5.8, 2.2 Hz, 1H), 6.13 (br d, J = 8.1 Hz, 1H), 4.92 (dt, J = 8.2, 6.0 Hz, 1H), 4.05 – 3.89 (m, 2H), 3.66 (s, 3H), 3.33 – 2.98 (m, 4H), 1.93 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.7, 169.8, 168.3, 135.6, 134.2, 132.5, 131.9, 128.6, 123.5, 121.8, 119.7, 118.2, 110.8, 107.4, 52.9, 52.4, 37.0, 27.0, 25.3, 23.1; IR (NaCl/thin film): 3369, 3280, 3052, 2948, 1770, 1738, 1711, 1659, 1530, 1438, 1397, 1371; $[\alpha]_D^{25} = +14.8$ (c = 0.96, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 355.1452, found 355.1455.

(S)- N_{α} -Acetyltryptophan methyl ester and indole dimer

Prepared from indole (23.4 mg, 0.20 mmol) following General Procedure 2. The crude residue was purified by silica gel chromatography (0:100 to 100:0 EtOAc:hexanes) to yield 17.9 mg



(contains 9 wt % EtOAc, 31% corrected yield) of (*S*)- N_{α} -acetyltryptophan methyl ester as a light pink oil and 7.0 mg (30% yield) of an indole dimer as a light yellow oil. The enantiomeric excess of (*S*)- N_{α} acetyltryptophan methyl ester was determined to be 67% by chiral SFC analysis (OD-H, 2.5 mL/min, 15% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 11.4 min, $t_{\rm R}$ (minor) = 10.6 min. [α]_D²⁵ = +39.3 (c = 0.83, CHCl₃).

Spectral data for both (S)- N_{α} -acetyltryptophan methyl ester²⁰ and the indole dimer²¹ are in agreement with the literature.

²⁰ Ruis-Rodríguez, J.; Albericio, F.; Lavilla, R. Chem. Eur. J. 2010, 16, 1124.

²¹ Xu, X.-H.; Liu, G.-K.; Azuma, A.; Tokunaga, E.; Shibata, N. Org. Lett. 2011, 13, 4854.

5. SFC traces for racemic and enantioenriched tryptophan derivatives.

Optimization of Reaction Parameters

7a (Table 2, entry 1): racemic



7a (Table 2, entry 1): enantioenriched, 35% ee



7b (Table 2, entry 2): racemic



7b (Table 2, entry 2): enantioenriched, 42% ee



7c: racemic



7c (Table 2, entry 3, no additive, DCM as solvent): enantioenriched, 78% ee





7c (Table 2, entry 6, with K₂CO₃, DCM as solvent): enantioenriched, 78% ee

7c (Table 2, entry 8, with 4Å MS, DCM as solvent): enantioenriched, 81% ee







7c (Table 2, entry 10, with 4Å MS, CHCl₃ as solvent): enantioenriched, 72% ee







7c (Table 3, entry 3, (R)-3,3'-dimethyl-BINOL (9c)): enantioenriched, 87% ee







7c (Table 3, entry 5, (R)-3,3'-dichloro-BINOL (9e)): enantioenriched, 90% ee







7c (Table 3, entry 7, (R)-3,3'-dimethoxy-BINOL (9g)): enantioenriched, 1% ee





7c (Table 3, entry 8, (R)-6,6'-dimethoxy-BINOL (9h)): enantioenriched, 54% ee

7c (Table 3, entry 9, (R)-6,6'-dimethyl-BINOL (9i)): enantioenriched, 78% ee





7c (Table 3, entry 10, (R)-6,6'-dibromo-BINOL (9j)): enantioenriched, 78% ee

7c (Table 3, entry 12, 15 mol % 9f): enantioenriched, 93% ee





7c (Table 3, entry 13, 10 mol % 9f): enantioenriched, 92% ee

7c (Table 3, entry 14, 5 mol % 9f): enantioenriched, 88% ee


Substrate scope of the conjugate addition/asymmetric protonation



7d (Table 4, entry 2): racemic

7d (Table 4, entry 2): enantioenriched, 85% ee



7e (Table 4, entry 3): racemic



7e (Table 4, entry 3): enantioenriched, 85% ee







3e (Table 2, entry 3): enantioenriched, 96% ee







7g (Table 4, entry 5): enantioenriched, 95% ee







7h (Table 4, entry 6): enantioenriched, 89% ee







7i (Table 4, entry 7): enantioenriched, 94% ee







7j (Table 4, entry 8): enantioenriched, 91% ee



7k (Table 4, entry 9): racemic



7k (Table 4, entry 9): enantioenriched, 93% ee



7l (Table 4, entry 10): racemic



7l (Table 4, entry 10): enantioenriched, 92% ee



7m (Table 4, entry 11): racemic



7m (Table 4, entry 11): enantioenriched, 94% ee



7n (Table 4, entry 12): racemic



7n (Table 4, entry 12): enantioenriched, 87% ee



70 (Table 4, entry 13): racemic



70 (Table 4, entry 13): enantioenriched, 93% ee



7p (Table 4, entry 14): racemic



7p (Table 4, entry 14): enantioenriched, 93% ee



7q (Table 4, entry 15): racemic



7q (Table 4, entry 15): enantioenriched, 92% ee



7r (Table 4, entry 16): racemic



7r (Table 4, entry 16): enantioenriched, 92% ee



7s (Table 4, entry 17): racemic



7s (Table 4, entry 17): enantioenriched, 92% ee



7t (Table 4, entry 18): racemic



7t (Table 4, entry 18): enantioenriched, 85% ee



7u (Table 4, entry 19): racemic



7u (Table 4, entry 19): enantioenriched, 91% ee



7v (Table 4, entry 20): racemic



7v (Table 4, entry 20): enantioenriched, 92% ee



7w (Table 4, entry 21): racemic



7w (Table 4, entry 21): enantioenriched, 84% ee



3x (Table 4, entry 22): racemic



3x (Table 4, entry 22): enantioenriched, 90% ee



(S)-Na-Acetyltryptophan methyl ester: racemic



(S)-Na-Acetyltryptophan methyl ester: enantioenriched, 67% ee



6. Scale-up Procedure.



To a flame-dried flask under nitrogen containing freshly activated powdered 4Å molecular sieves (200 wt %) was added 2-phenylindole (**1a**, 1.00 g, 5.20 mmol, 1.00 equiv), methyl 2acetamidoacrylate (**2c**, 890 mg, 6.20 mmol, 1.20 equiv), and (*R*)-3,3'-dibromo-BINOL (**9f**, 457 mg, 1.00 mmol, 0.20 equiv). The flask was charged with DCM (40 mL) and SnCl₄ (1 M in DCM, 5.20 mL, 5.20 mmol, 1.00 equiv) was added. The reaction was stirred at room temperature for 2 hours, then quenched by addition of 1 M HCl (50 mL). The aqueous layer was extracted with EtOAc (2 x 50 mL) and the combined organic layers were washed with saturated aqueous NaHCO₃ (50 mL), dried (Na₂SO₄), filtered and concentrated. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes) to yield 1.33 g (77% yield) of **7c** as a pale yellow foam. The enantiomeric excess was determined to be 93% by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 5.7$ min, $t_R(minor) = 6.9$ min.



7. Functionalization of tryptophans 7c and 7d.

Acetamide hydrolysis of 7c²²



A vial was charged with (S)- N_{α} -acetyl-2-phenyltryptophan methyl ester (7c, 30.0 mg, 0.09 mmol), MeOH (1 mL), H₂O (1 mL) and aqueous HCl (12 M, 1 mL). The reaction was heated to 75 °C for 12 hours, then concentrated, redissolved in DCM (10 mL) and washed with saturated aqueous NaHCO₃ (3 X 5 mL). The aqueous layers were combined and extracted with DCM (4 X 5 mL). The combined organic layers were washed with brine, dried (Na₂SO₄), filtered and concentrated. The crude residue was purified by silica gel chromatography (99:1 CH₂Cl₂:MeOH) to yield 20.0 mg (76% yield) of 10 as a light yellow oil. The enantiomeric excess was determined by chiral SFC analysis of the corresponding methylcarbamate S9 (see below). ¹H NMR (500 MHz, CDCl₃) δ 8.18 (br s, 1H), 7.67 (dd, J = 7.6, 0.7 Hz, 1H), 7.62 – 7.60 (m, 2H), 7.50 - 7.43 (m, 2H), 7.41 - 7.34 (m, 2H), 7.22 (ddd, J = 8.1, 7.1, 1.2 Hz, 1H), 7.15(ddd, J = 7.9, 7.0, 1.0 Hz, 1H), 3.89 (dd, J = 8.4, 5.0 Hz, 1H), 3.56 (s, 3H), 3.47 - 3.38 (m, 1H), 3.27 – 3.14 (m, 1H), 1.69 (br s, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 175.5, 136.1, 135.8, 132.9, 129.1, 129.0, 128.3, 128.0, 122.5, 119.9, 119.2, 110.9, 108.2, 55.2, 51.9, 30.2; IR (NaCl/thin film): 3367, 3062, 2948, 1732, 1603, 1489, 1457, 1207; $[\alpha]_D^{25} = -12.4$ (c = 0.85, CHCl₃). HRMS (MM) calc'd for $[M+H]^+$ 295.1441, found 295.1446.

²² For an analogous hydrolysis procedure, see: Messina, F.; Botta, M.; Corelli, F.; Schneider, M.; Fazio, F. J. Org. Chem. **1999**, 64, 3767.

Methylcarbamate Protection



A flame-dried flask was charged with free amine **10** (19.5 mg, 0.70 mmol, 1.00 equiv), Et₃N (19 μ L, 0.13 mmol, 2.0 equiv) and DCM (5 mL). Methylchloroformate (6.0 μ L, 0.73 mmol, 1.10 equiv) was added and the solution was stirred at room temperature for 3 hours, then quenched with saturated aqueous NH₄Cl (5 mL) and extracted with EtOAc (2 X 5 mL). The combined organic layers were dried (Na₂SO₄), filtered and concentrated. The crude residue was purified by silica gel chromatography (25:75 EtOAc:hexanes) to yield 18.5 mg (80% yield) of methylcarbamate **S9** as a colorless oil. The enantiomeric excess was determined to be 93% by chiral SFC analysis (OD-H, 2.5 mL/min, 15% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 16.7 min, $t_{\rm R}$ (minor) = 15.6 min. ¹H NMR (500 MHz, CDCl₃) δ 8.11 (br s, 1H), 7.61 (d, J = 7.9 Hz, 1H), 7.57 – 7.52 (m, 1H), 7.48 – 7.45 (m, 2H), 7.40 – 7.35 (m, 2H), 7.25 – 7.19 (m, 1H), 7.16 (m, 1H), 5.06 (br d, J = 7.7 Hz, 1H), 4.63 – 4.59 (m, 1H), 3.54 (s, 3H), 3.50 (m, 2H), 3.38 (s, 3H); 1³C NMR (125 MHz, CDCl₃) δ 172.3, 156.1, 136.2, 135.7, 132.9, 129.2, 129.0, 128.3, 128.0, 122.5, 120.0, 118.9, 110.9, 106.7, 54.5, 52.12, 52.07, 27.1; IR (NaCl/thin film) 3338, 2953, 2923, 2852, 1718, 1701, 1507, 1457, 1363, 1213, 1072 cm⁻¹; [α]_D²⁵ = +22.6 (*c* = 0.10, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 353.1496, found 353.1497.

Methylcarbamate (S9): racemic



Methylcarbamate (S9): enantioenriched, 93% ee



Methyl ester hydrolysis²³



A 10 mL flask was charged with (S)- N_{α} -acetyl-2-phenyltryptophan methyl ester 7c (67.2 mg, 0.20 mmol, 1.00 equiv) and THF (0.9 mL) then cooled to 0 °C, followed by dropwise addition of aqueous LiOH (1.75 M, 230 µL, 0.40 mmol, 2.00 equiv). The reaction was vigorously stirred at 0 °C for 2 hours, then diluted with H₂O (15 mL) and extracted with EtOAc (2 x 10 mL). The aqueous layer was acidified to pH = 1.5 and extracted with EtOAc (5 x 15 mL). The combined organic layers from the acidic aqueous extraction were dried (Na₂SO₄), filtered, and concentrated. The crude residue was purified by silica gel chromatography (0:99:1 to 15:84:1 MeOH:DCM:AcOH) to yield 59.2 mg (92% yield) of carboxylic acid 11 as a pale yellow foam. The enantiomeric excess was determined to be 92% by chiral SFC analysis (AS-H, 2.5 mL/min, 28% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 4.5 min, $t_{\rm R}$ (minor) = 8.0 min. ¹H NMR (500 MHz, CDCl₃) δ 8.21 (br s, 1H), 7.63 (d, J = 7.8 Hz, 1H), 7.56 – 7.51 (m, 2H), 7.47 (dd, J = 7.6, 7.6 Hz, 2H), 7.40 (m, 1H), 7.37 (ddd, J = 8.0, 0.8, 0.8 Hz, 1H), 7.21 (ddd, J = 8.1, 7.1, 1.1 Hz, 1H), 7.14 (ddd, J = 8.0, 7.1, 1.0 Hz, 1H), 5.72 (br d, J = 7.4 Hz, 1H), 4.73 (td, J = 7.1, 5.4 Hz, 1H), 3.56 $(dd, J = 14.9, 5.2 Hz, 1H), 3.49 (dd, J=15.0, 6.9 Hz, 1H), 1.62 (s, 3H); {}^{13}C NMR (125 MHz, 125 MHz)$ CDCl₃) § 174.7, 170.9, 136.2, 135.7, 132.9, 129.13, 129.05, 128.3, 128.2, 122.6, 120.1, 118.8, 111.0, 106.8, 53.1, 26.2, 22.6; IR (NaCl/thin film): 3391, 3306, 3055, 3011, 2921, 2850, 1717, 1615, 1527, 1457, 1448, 1215 cm⁻¹; $[\alpha]_D^{25} = +9.2$ (*c* = 1.05, MeCN). HRMS (MM) calc'd for [M+H]⁺ 323.1390, found 323.1390.

²³ For an analogous hydrolysis procedure, see: Morieux, P.; Stables, J.P.; Kohn, H. *Bioorg. Med. Chem.* **2008**, *16*, 8968.

11: racemic



11: enantioenriched, 92% ee



Preparation of bromo-dehydroindoline 12.



A solution of (S)- N_{α} -acetyl-2-phenyltryptophan methyl ester 7c (101 mg, 0.30 mmol, 1.00 equiv) in DCM (8.4 mL) was cooled to -50 °C in an acetonitrile/dry ice bath. NBS (53.4 mg, 0.30 mmol, 1.00 equiv) was then added, followed by TFA (900 µL). The reaction was stirred in the dark at -50 °C for 3 hours, then poured onto ice, quenched with aqueous ammonia (1.5 mL) and extracted with DCM (3 x 25 mL). The combined organics were washed (40 mL H₂O, then 40 mL brine), dried (Na₂SO₄), filtered, and concentrated. The product **12** was formed in a 1:1 ratio of diastereomers (determined by ¹H NMR analysis of the crude reaction mixture) and was purified by silica gel chromatography (30:70 to 70:30 EtOAc:hexanes) to yield 98 mg (79% yield) of the combined diastereomers as a bright yellow foam. The enantiomeric excesses of the two diastereomers were determined to be 92% and 90% by chiral SFC analysis (AS-H, 2.5 mL/min, 20% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 3.8$ min, $t_R(minor) = 4.1$ min; $t_R(major) =$ 4.6 min, $t_{\rm R}$ (minor) = 6.0 min. Spectral data and optical rotation are reported for the mixture of diastereomers. ¹H NMR (500 MHz, CDCl₃) δ 8.42 - 8.32 (m, 4H), 7.70 - 7.64 (m, 2H), 7.57 -7.49 (m, 8H), 7.47 - 7.40 (m, 2H), 7.39 - 7.30 (m, 2H), 5.37 (br d, J = 7.4 Hz, 1H), 5.05 (br d, J= 8.5 Hz, 1H), 4.33 (dt, J = 7.5, 5.5 Hz, 1H), 3.95 (td, J = 8.9, 4.0 Hz, 1H), 3.56 (dd, J = 14.8, 5.2 Hz, 1H), 3.47 - 3.41 (m, 4H), 3.38 - 3.32 (m, 4H), 3.23 (dd, J = 14.6, 9.3 Hz, 1H), 1.45 (s, 3H), 1.27 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 175.6, 174.8, 170.7, 170.0, 169.4, 169.2, 151.82, 151.76, 139.8, 139.6, 131.6, 131.4, 131.3, 130.5, 130.4, 128.81, 128.80, 128.71, 128.70, 127.2, 126.6, 123.2, 122.5, 121.9, 121.7, 59.16, 59.14, 52.5, 52.3, 50.3, 49.8, 41.6, 41.4, 22.3, 22.0; IR (NaCl/thin film): 3271, 3062, 2952, 2924, 2853, 1747, 1661, 1525, 1444, 1372, 1264, 1216 cm⁻¹; $[\alpha]_D^{25} = +17.1$ (c = 0.50, CHCl₃). HRMS (MM) calc'd for M⁺ 415.0652, found 415.0652.



Bromo-dehydroindoline 12: 1:1 mixture of diastereomers, racemic

Bromo-dehydroindoline 12: 1:1 mixture of diastereomers, enantioenriched, 92:90% ee



Preparation of 3-hydroxypyrroloindoline 13



A 15 mL flask containing (S)- N_{α} -acetyl-1-methyl-2-phenyltryptophan methyl ester 7d (52.5 mg. 0.150 mmol, 1.00 equiv) was flushed with argon and then charged with MeCN (3.3 mL). TFA was added as a solution in MeCN (1.3 M, 125 µL, 0.150 mmol, 1.00 equiv), followed by NCS as a solution in MeCN (0.2 M, 0.75 mL, 0.150 mmol, 1.00 equiv). The flask was then sealed under argon and the solution was stirred in the dark at room temperature. After 3 hours, the reaction was quenched with aqueous ammonia (1.5 mL), poured onto ice, and extracted with DCM (3 x 15 mL). The combined organics were washed (20 mL H₂O, then 20 mL brine), dried (Na₂SO₄), filtered, and concentrated to give the crude mixture of 3-chloropyrroloindoline diastereomers (detected by HRMS direct injection (MM) calc'd for [M+H]⁺ 385.1313, found 385.1320). The crude residue was redissolved in MeCN (2 mL), then H₂O (1.2 mL) and SiO₂ (2.5 mL) were added. The mixture was vigorously stirred open to air at room temperature for 30 minutes, then filtered through a 1.5 mL silica plug with EtOAc (50 mL), dried (Na₂SO₄), filtered and concentrated. The 3-hydroxypyrroloindoline 13 existed in a 6:1 ratio of diastereomers, favoring the endo diastereomer (determined by ¹H NMR analysis of the crude reaction mixture) and was purified by silica gel chromatography (0:100 to 10:90 EtOAc:hexanes) to yield 30.8 mg (contains 18 wt % CHCl₃, 46% corrected yield) of the endo diastereomer as a yellow oil. The exo diastereomer, obtained post chromatography in a mixture with (S)-N_a-acetyl-1-methyl-2phenyltryptophan methyl ester 7d, was subjected to reverse phase preparatory HPLC (30:70 to 90:10 MeCN:H₂O) using an Agilent 1200 Series HPLC with an Agilent XDB-C18 5 µM column (9.4 x 250 mm) to yield 3.5 mg (6% yield) of the exo diastereomer as a yellow oil.

Endo diastereomer:

The enantiomeric excess was determined to be 84% by chiral SFC analysis (AD-H, 2.5 mL/min, 25% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 7.4$ min, $t_R(minor) = 4.7$ min. The relative stereochemistry was assigned by 2D NMR analysis. ¹H NMR (500 ŌН MHz, CD₃CN; compound exists as a 15:1 mixture of rotamers, the CO₂Me major rotamer is reported) δ 7.40 – 7.35 (m, 2H), 7.34 – 7.26 (m, 3H), Me h Ac 7.20 (ddd, J = 7.9, 7.5, 1.3 Hz, 1H), 7.12 (ddd, J = 7.2, 1.3, 0.5 Hz, endo-13 1H), 6.66 (ddd, J = 7.3, 7.3, 1.0 Hz, 1H), 6.51 (d, J = 7.9 Hz, 1H), 4.79 (d, J = 8.8 Hz, 1H), 3.19 (s, 3H), 2.97 (s, 3H), 2.90 (br s, 1H), 2.82 (d, J = 12.7 Hz, 1H), 2.59 (ddd, J = 12.7, 8.8, 1.1 Hz, 1H), 1.95 (s, 3H); ¹³C NMR (125 MHz, CD₃CN; compound exists as a 15:1 mixture of rotamers, the major rotamer is reported) δ 172.0, 171.3, 153.1, 138.0, 131.6, 128.9, 128.6, 128.3, 125.2, 118.0, 107.1, 95.3, 88.3, 61.3, 52.7, 39.0, 32.7, 23.6; IR (NaCl/thin film): 3292, 3010, 2948, 1735, 1653, 1648, 1610, 1491, 1448, 1388, 1313, 1220 cm⁻¹; $[\alpha]_{D}^{25} = +264.0$ (*c* = 1.35, CHCl₃). HRMS (MM) calc'd for $[M+H]^{+}$ 367.1652, found 367.1650. Exo diastereomer:



The enantiomeric excess was determined to be 85% by chiral SFC analysis (OD-H, 2.5 mL/min, 20% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 6.2 min, $t_{\rm R}$ (minor) = 4.0 min. The relative stereochemistry was assigned by 2D NMR analysis. ¹H NMR (500 MHz, CD₃CN; compound exists as a 1.5:1 mixture of rotamers, the major rotamer is

denoted by *, the minor rotamer by [§]) δ 7.60 – 7.22 (m, 6H*, 7H[§]), 7.17 (ddd, J = 7.3, 0.6, 0.6 Hz, 1H*), 6.79 (dd, J = 7.5, 7.5 Hz, 1H[§]), 6.70 (dd, J = 7.5, 7.5 Hz, 1H*), 6.65 (d, J = 7.9 Hz, 1H[§]), 6.54 (d, J = 7.9 Hz, 1H*), 4.49 (dd, J = 8.0, 6.7 Hz, 1H*), 4.07 (dd, J = 10.0, 6.9 Hz, 1H[§]), 3.81 (s, 3H*), 3.71 (s, 3H[§]), 3.34 (s, 1H[§]), 3.01 (s, 1H*), 2.963 (s, 3H*), 2.958 (s, 3H[§]), 2.71 (dd, J = 13.0, 8.1 Hz, 1H*), 2.68 (dd, J = 12.6, 7.0 Hz, 1H[§]), 2.34 (dd, J = 12.9, 6.7 Hz, 1H*), 2.07 (dd, J = 12.7, 10.0 Hz, 1H[§]), 1.89 (s, 3H*), 1.80 (s, 3H[§]); ¹³C NMR (125 MHz, CD₃CN) δ 174.1, 173.6, 172.3, 171.8, 151.2, 151.1, 136.3, 136.2, 131.6, 131.3, 130.3, 129.60, 129.57, 129.4, 128.7, 128.6, 124.4, 123.9, 119.3, 118.2, 108.0, 106.4, 98.8, 96.1, 90.1, 88.5, 61.2, 60.3, 53.3, 52.6, 40.9, 37.2, 33.4, 32.4, 24.6, 23.8; IR (NaCl/thin film): 3305, 2924, 1747, 1646, 1610, 1491, 1448, 1381, 1311, 1207 cm⁻¹; [α]_D²⁵ = -138.2 (c = 0.33, CHCl₃). HRMS (MM) calc'd for [M+H]⁺ 367.1652, found 367.1655.





Endo-13: enantioenriched, 84% ee







Exo-13: enantioenriched, 85% ee



8. Scalemic (R)-BINOL Experiment.



A flame-dried flask was charged with 2-phenylindole (**6a**, 19.0 mg, 0.10 mmol, 1.00 equiv) and methyl 2-acetamidoacrylate (**2c**, 14.0 mg, 0.10 mmol, 1.00 equiv). Stock solutions of (*R*)-BINOL (**9a**) and racemic BINOL were prepared (0.0134 M in DCM) and the appropriate volume of each solution was added to the flask (0.02 mmol, 0.20 equiv). The reaction was charged with SnCl₄ as a solution in DCM (1 M, 120 μ L, 0.12 mmol, 1.20 equiv) and stirred at room temperature for 2 hours, then quenched by dilution with 1 M HCl (5 mL) and MeCN (1 mL). The aqueous layer was extracted with EtOAc (2 x 5 mL) and the combined organic layers were washed with saturated aqueous NaHCO₃ (5 mL), dried (Na₂SO₄), filtered, and concentrated. The crude residue was purified by silica gel chromatography (40:60 to 100:0 EtOAc:hexanes). The enantiomeric excess of each experiment was determined by chiral SFC analysis (AD-H, 2.5 mL/min, 30% IPA in CO₂, $\lambda = 254$ nm).



9. Deuterium labeling studies.

Preparation of N-deuteroacrylate (S10).



Acrylate **2c** was dissolved in MeOD (1 mL) under nitrogen. After stirring for 1 minute, the solution was concentrated under high vacuum. This procedure was repeated three times to give >99% deuterium incorporation.


Preparation of per-deutero-2-phenylindole (S11).



To MeOD (1 mL) in a microwave vial was added acetyl chloride (100 μ L), followed by 2phenylindole (**6a**, 50 mg) and D₂O (1 mL). The vial was sealed and heated in a microwave to 140 °C for 1 hour. Upon cooling, the heterogenous solution was diluted with DCM. The phases were separated and the aqueous was extracted with DCM (2 x 5 mL). The combined organic layers were dried (Na₂SO₄), filtered and concentrated to give per-deutero-2-phenylindole (**S11**) with 90% deuterium incorporation.



10. ¹H NMR Kinetics Experiment for SnCl₄ and (*R*)-BINOL (9a)•SnCl₄ promoted reaction of 6a and 2c.



An oven-dried vial was charged with 2-phenylindole (**6a**, 19.0 mg, 0.10 mmol, 1.00 equiv), methyl 2-acetamidoacrylate (**2c**, 14.0 mg, 0.10 mmol, 1.00 equiv), (*R*)-BINOL if necessary (**9a**, 6.0 mg, 0.02 mmol, 0.20 equiv) and 1,4-diethylbenzene (4.7 μ L, 0.03 mmol, 0.30 equiv) as the internal standard. The vial was pumped into a glove box and charged with CD₂Cl₂ (0.75 mL, to an indole concentration of 0.12 M), then transferred to a screw-cap NMR tube. A ¹H NMR spectrum (1 scan) was taken to determine the initial ratio of acrylate and 1,4-diethylbenzene. SnCl₄ (1 M in CD₂Cl₂, 120 μ L, 0.12 mmol, 1.20 equiv) was then added through the septum of the screw-cap and the NMR tube was inverted once and quickly inserted into the spectrometer. The concentration of acrylate was monitored by ¹H NMR over 9 hours and was determined by integration of its resonance at 3.83 ppm relative to 1,4-diethylbenzene's resonance at 2.74 ppm.





11. Comparison of conditions for pyrroloindoline formation.

Table 5, entry 1:



To a flame-dried 10 mL flask was added 1,3-dimethylindole (1a, 29.0 mg, 0.20 mmol, 1.00 equiv), acrylate 2c (28.6 mg, 0.20 mmol, 1.00 equiv), and (R)-BINOL (9a, 11.4 mg, 0.04 mmol, 0.20 equiv). The flask was charged with DCM (1.5 mL), followed by addition of $SnCl_4$ (1 M in DCM, 240 µL, 0.24 mmol, 1.20 equiv) and the reaction mixture was stirred at room temperature for 4 hours, then guenched by diluting with MeCN (1 mL) and 1 M HCl (5 mL). The agueous layer was extracted with EtOAc (3 x 5 mL) and the combined organic layers were washed with saturated aqueous NaHCO₃ (15 mL) The aqueous layer was back extracted with EtOAc (10 mL) and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated. The product **3k** was formed in a 5:1 ratio of diastereomers favoring the *exo* diastereomer (determined by 1 H NMR analysis of the crude reaction mixture) and purified by silica gel chromatography (0:100 to 100:0 EtOAc: hexanes) to yield 40.1 mg (70% yield) of the combined diastereomers as a yellow oil. The enantiomeric excess of the exo diastereomer was determined to be 66% by chiral SFC analysis (AD-H, 2.5 mL/min, 10% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 9.0$ min, $t_R(minor) =$ 5.8 min. The enantiomeric excess of the endo diastereomer was determined to be 80% by chiral SFC analysis (AD-H, 2.5 mL/min, 10% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 3.8$ min, $t_R(minor)$ = 4.5 min. Spectral data are in agreement with the literature.²⁴

²⁴ Repka, L. M.; Ni, J.; J. Reisman. S. E. J. Am. Chem. Soc. 2010, 132, 14418.





3k (Table 5, entry 1): enantioenriched, exo: 66% ee, endo: 80% ee





An oven-dried vial was charged with 1,3-dimethylindole (1a, 29.0 mg, 0.20 mmol, 1.00 equiv), acrylate 2c (34.3 mg, 0.24 mmol, 1.20 equiv), and (R)-3,3'-dibromo-BINOL (9f, 17.8 mg, 0.04 mmol, 0.20 equiv) and pumped into a glove box. To the vial was added flame-dried powdered 4Å molecular sieves (200 wt % relative to 1a). The vial was charged with DCM (1.5 mL) and SnCl₄ (1 M in DCM, 200 µL, 0.20 mmol, 1.00 equiv) was added. The reaction was stirred at 20 °C for 4 hours, after which time it was removed from the glove box and quenched by dilution with 1 M HCl (5 mL) and MeCN (1 mL). The aqueous layer was extracted with EtOAc (3 x 5 mL) and the combined organic layers were washed with saturated aqueous NaHCO₃ (15 mL). The aqueous was back extracted with EtOAc (10 mL) and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated. The product 3k was formed in a 8:1 ratio of diastereomers favoring the exo diastereomer (determined by ¹H NMR analysis of the crude reaction mixture) and purified by silica gel chromatography (0:100 to 100:0 EtOAc:hexanes) to yield 33.5 mg (58% yield) of the combined diastereomers as a yellow oil. The enantiomeric excess of the exo diastereomer was determined to be 87% by chiral SFC analysis (AD-H, 2.5 mL/min, 10% IPA in CO₂, $\lambda = 254$ nm): $t_R(major) = 8.9$ min, $t_R(minor) = 5.7$ min. The enantiomeric excess of the endo diastereomer was determined to be 85% by chiral SFC analysis (AD-H, 2.5 mL/min, 10% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 3.7 min, $t_{\rm R}$ (minor) = 4.4 min. Spectral data are in agreement with the literature.²⁴



3k (Table 5, entry 2): enantioenriched, exo: 87% ee, endo: 85% ee



(7:1 dr, 98:92% ee)

An oven-dried vial was charged with 1,3-dimethylindole (1a, 29.0 mg, 0.20 mmol, 1.00 equiv), acrylate 2a (65.5 mg, 0.24 mmol, 1.20 equiv), and (R)-3,3'-dibromo-BINOL (9f, 17.8 mg, 0.04 mmol, 0.20 equiv) and pumped into a glove box. To the vial was added flame-dried powdered 4Å molecular sieves (200 wt % relative to 1a). The vial was charged with DCM (1.5 mL) and SnCl₄ (1 M in DCM, 200 µL, 0.20 mmol, 1.00 equiv) was added. The reaction was stirred at 20 °C for 4 hours, after which time it was removed from the glove box and quenched by dilution with 1 M HCl (5 mL) and MeCN (1 mL). The aqueous layer was extracted with EtOAc (3 x 5 mL) and the combined organic layers were washed with saturated aqueous NaHCO₃ (15 mL). The aqueous was back extracted with EtOAc (10 mL) and the combined organic layers were dried (Na₂SO₄), filtered, and concentrated. The product **3a** was formed in a 7:1 ratio of diastereomers favoring the exo diastereomer (determined by ¹H NMR analysis of the crude reaction mixture) and purified by silica gel chromatography (0:100 to 10:90 EtOAc:hexanes) to yield 32.7 mg (39% yield) of the combined diastereomers as a yellow oil. The enantiomeric excess of the exo diastereomer was determined to be 98% by chiral SFC analysis (OJ-H, 2.5 mL/min, 3% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 12.5 min, $t_{\rm R}$ (minor) = 10.9 min. The enantiomeric excess of the endo diastereomer was determined to be 92% by chiral SFC analysis (OJ-H, 2.5 mL/min, 3% IPA in CO₂, $\lambda = 254$ nm): $t_{\rm R}$ (major) = 5.8 min, $t_{\rm R}$ (minor) = 5.0 min. Spectral data are in agreement with the literature.²⁴

²⁵ For more information regarding Table 5, entry 3, see reference 24.





3a (Table 5, entry 4): enantioenriched, exo: 98% ee, endo: 92% ee

