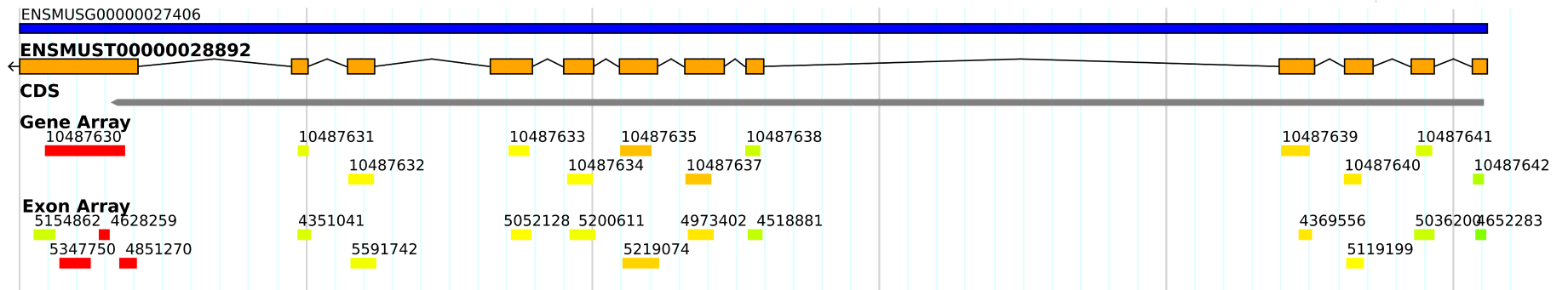


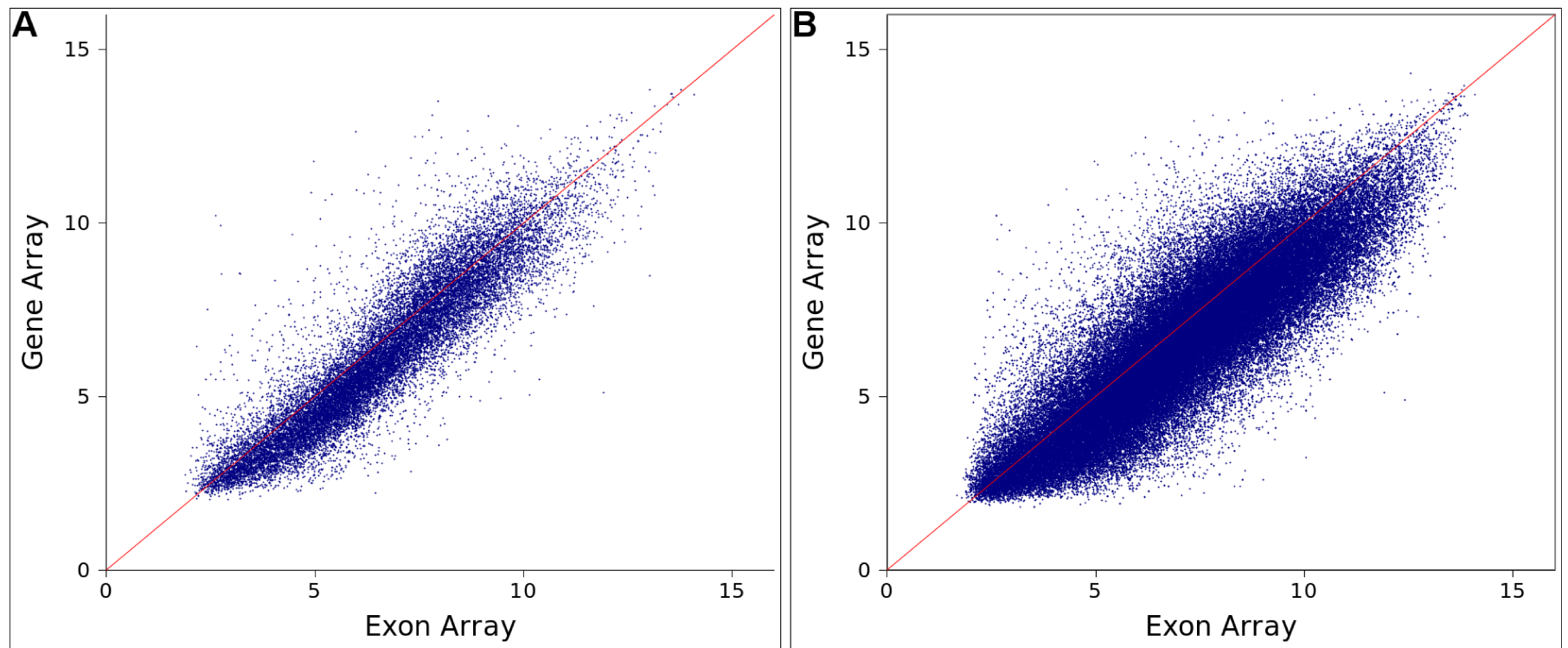
Supplementary Figure S1: Regulated splicing components

DEG (F p-value < 0.001 during the heart development or the cardiomyocyte differentiation) were selected and filtered by the Gene Ontology term GO:0008380 “RNA splicing”. We found three genes regulated during cardiomyocyte differentiation (red box) and 34 during the heart development. Gene expression signals were normalized and clustered by MeV using Pearson correlation with average linkage clustering.



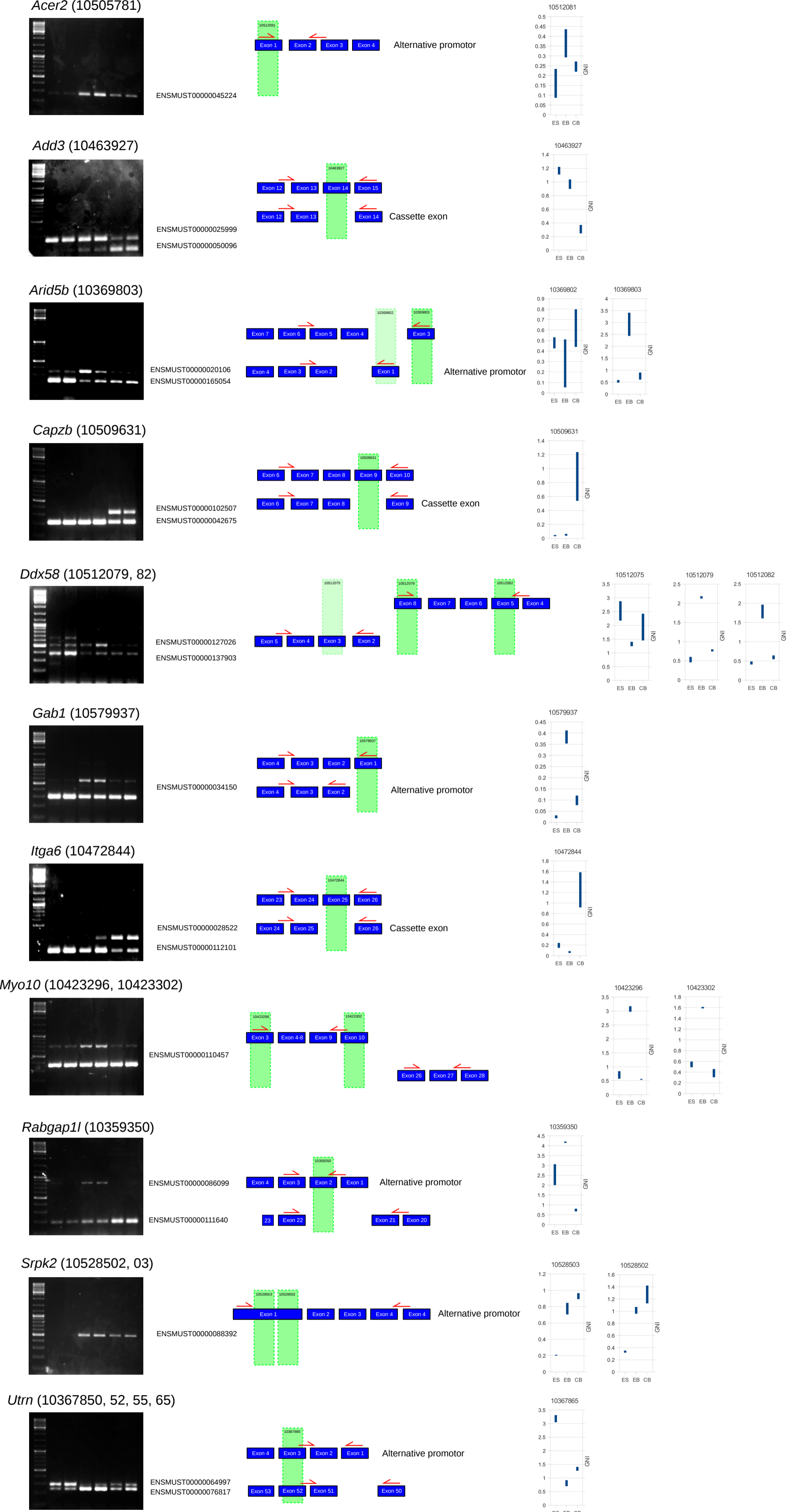
Supplementary Figure S2: Probe set coverage of gene and exon arrays

The majority of probe sets targeting well-known exons are identical on gene and exon arrays. However, in some cases, there are differences in the coverage and resolution between both array types. The lower resolution of gene arrays prevents the detection of the internal spliced exon of *Idh3b* while all other exons have similar probe sets on both arrays.



Supplementary Figure S3: Correlation between exon and gene arrays

Scatter plot of the data set for ES cells between gene and exon arrays at the gene (A) and exon (B) levels. We detected on both levels a very high correlation (0.91 and 0.87, respectively). We processed the arrays by using APT and the custom CDF files “ENSG” and “ENSE” from BrainArray (<http://brainarray.mbni.med.umich.edu>, release #14). These CDF files group probes in an ENSEMBL gene and ENSEMBL exon centric manner, respectively. This allows a direct comparison between the two types of arrays. Only ENSEMBL genes (n=21,094) and exons (n=125,768) present on both arrays were retained. The mean log₂ values of the biological replicates were used. Red lines indicate the x-y diagonal.



Supplementary Figure S4: RT-PCR results

RT-PCR results of 11 splice events. Red arrows indicate used primers, and green boxes regulated probe sets. Light green boxes represent probe sets that are not significantly regulated, but have a high SI which helped to identify the expressed isoforms. Box plots on the right show GNIs of regulated probe sets.

Supplementary Table S3: Comparison of validated exon array splice events with gene arrays.

Gene Symbol	Validated splice event identified by exon arrays	Observation with gene arrays	UCSC	AceView
<i>Idh3b</i>	Internal splice sites in 3' UTR	Alternative 3' exon	-	Cassette exon, alternative 3', intron retention
<i>Kif1b</i>	Internal 3' splice site and cassette exon	Same as exon array	Alternative 3', cassette exon	Alternative 3', cassette exon
<i>Ldb3</i>	Cassette exon	Gene filtered (gene not significantly expressed in non-heart group)	-	-
<i>Mfn2</i>	Cassette exon	Same as exon array	Cassette exon	Cassette exon
<i>Myom1</i>	Cassette exon	Probe set filtered (very high SI suggesting a false positive)	-	-
<i>Slc8a1</i>	Cassette exons	Not significant p-value, but high SI	-	Cassette exon, double cassette exon
<i>Svil</i>	Cassette exon	Same as exon array (only 1 probe set instead of 2)	Cassette exon	-
<i>Trip10</i>	Cassette exon	Same as exon array	Cassette exon	Cassette exon

Eight validated heart-specific splice events were identified by exon arrays compared to a similar analysis with gene arrays.

In four cases, both chip types identified exactly the same splice events. The splice events of *Idh3b* were not detected correctly due to the lack of sufficient probe sets. Heart-enriched exons of *Slc8a1* were identified with a high SI on both chip types but no significant p-values were recorded on gene arrays. No splice events were detected for *Ldb3* and *Myom1* because both genes were filtered out by the GAA.

Supplementary Table S6: RT-PCR primer pairs

Gene	Forward primer	Reverse primer
<i>Acer2</i>	GAGAAGCTGCTCCGATGCT	CAGATCCAATCCCCACTACAA
<i>Add3</i>	CTTCGACCATGCAGTTTGAC	GGAGGTGAAGCTCTTGGAAA
<i>Arid5b</i>	CCACACCAAGACCAGACTCC	GCCTCTCCTTTCACCTTGCT
	GAAACCAACGTGATTGTGCT	GCCTCTCCTTTCACCTTGCT
<i>Capzb</i>	CCAGACAGATGGAGAAAGACG	CCTCCACCAGGTCGTTCTTA
<i>Ddx58</i>	GATAAAGGTTTCAAAGGGCTGA	GATGGCTCCGTTGTTGAGAT
	CACAAAGCGTGCTCAGTGTT	CCATTTCTTCAGAGCATCCA
<i>Gab1</i>	CCTCTTCAGTCCGCGAATC	GGATCTTCTTCTGTGGGATTG
	CGTGGAAGAGAAGGTGGTTT	GGATCTTCTTCTGTGGGATTG
<i>Itga6</i>	GGCACTCAGGTTTCGAGTGA	GGTATCGGGGAATGCTGTC
<i>Myo10</i>	GGACTTCGACTCCAGGTTTG	GCTTGTGTTGTCTATGATCTCCTT
	GGGCTCCATCATGTACAACC	GACTTCCATTGCCGTGATAA
<i>Rabgap11</i>	GGAATTCAAAGAATTACTCAGAAGG	CCGCATCGCCTGATATTC
	GCTCTTCTAAAGACCTCAAAGGA	CCGCATCGCCTGATATTC
<i>Srpk2</i>	GGAGCCGTTTCCCTTTATAGC	CCTTGCATATCCCAGCACA
<i>Utrn</i>	GGAGAATGCTCTTCAGGACAG	CCAAATGGGCCCTGATGCTA
	CTTAAAATGATGGCGGTGGT	CCAAATGGGCCCTGATGCTA