

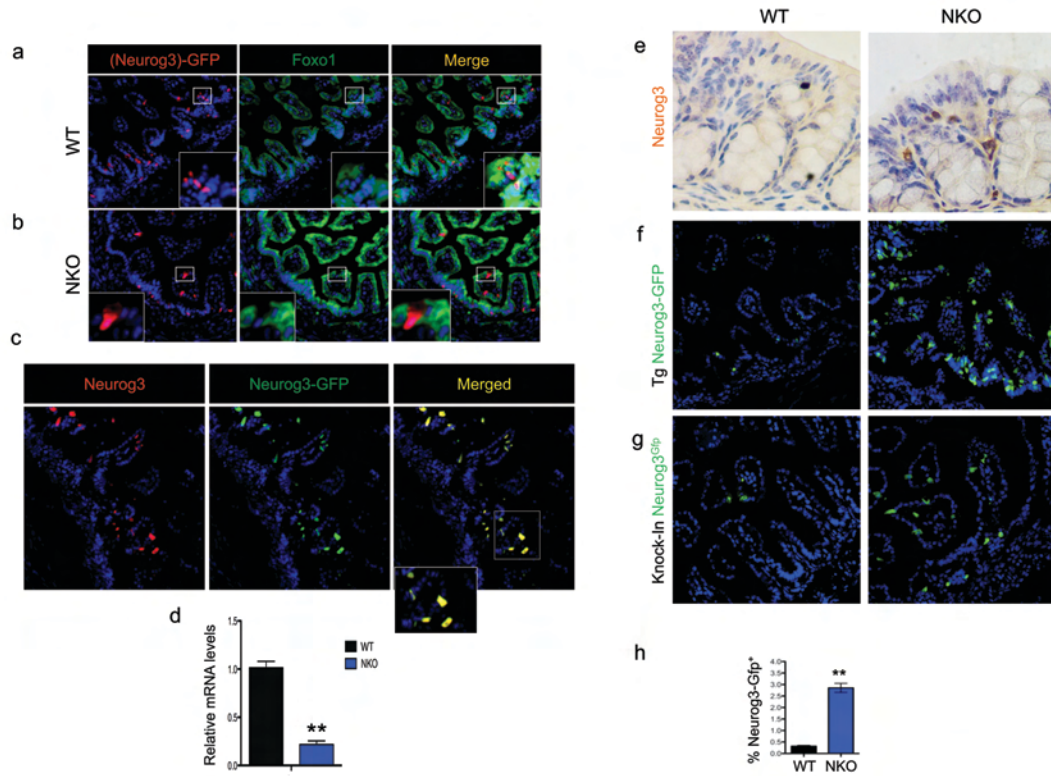
Supplemental Online Material To:

Generation Of Functional Insulin-Producing Cells In The Gut By Foxo1 Ablation

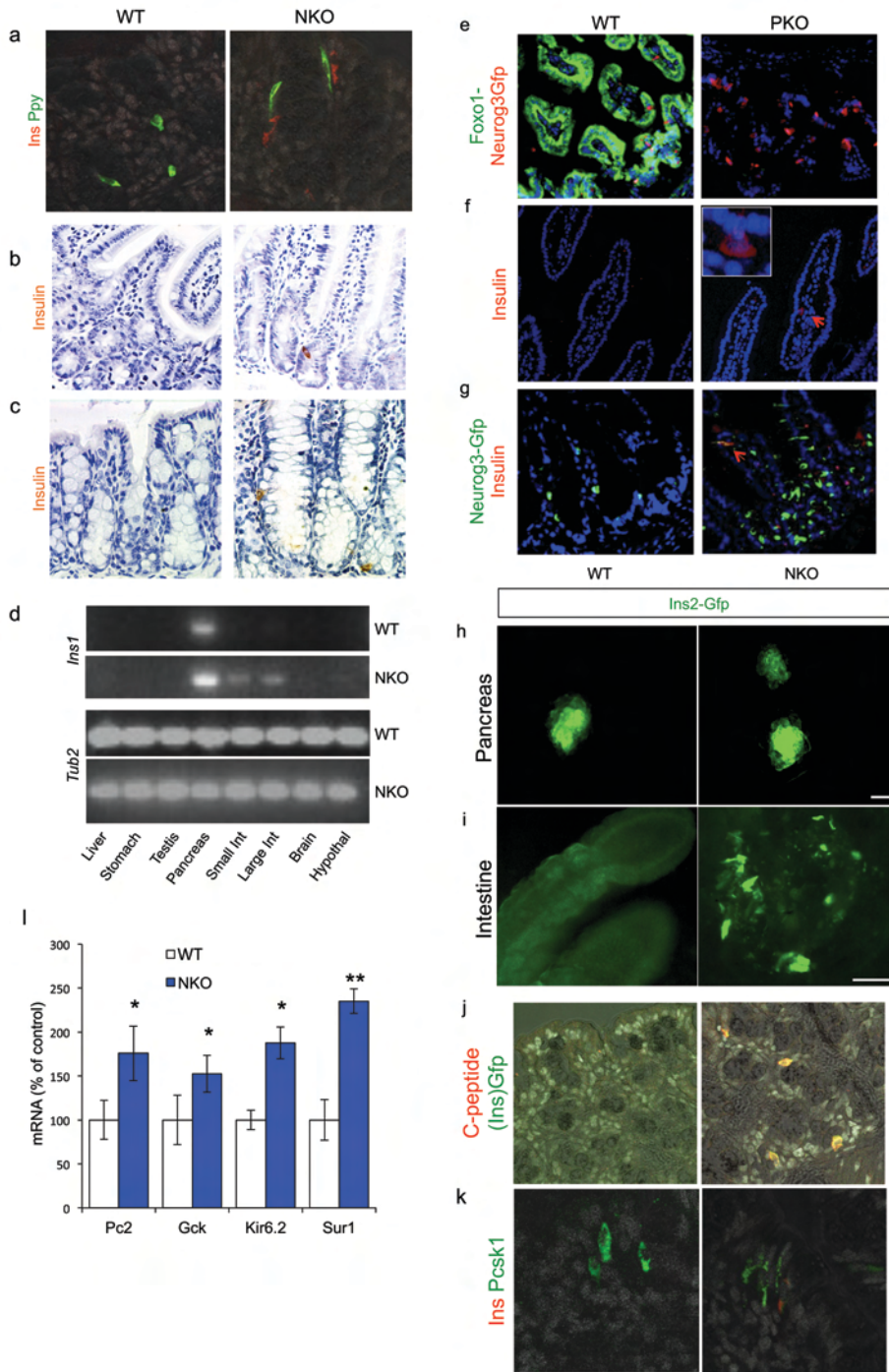
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**Contains 7 supplemental figures with figure legends**

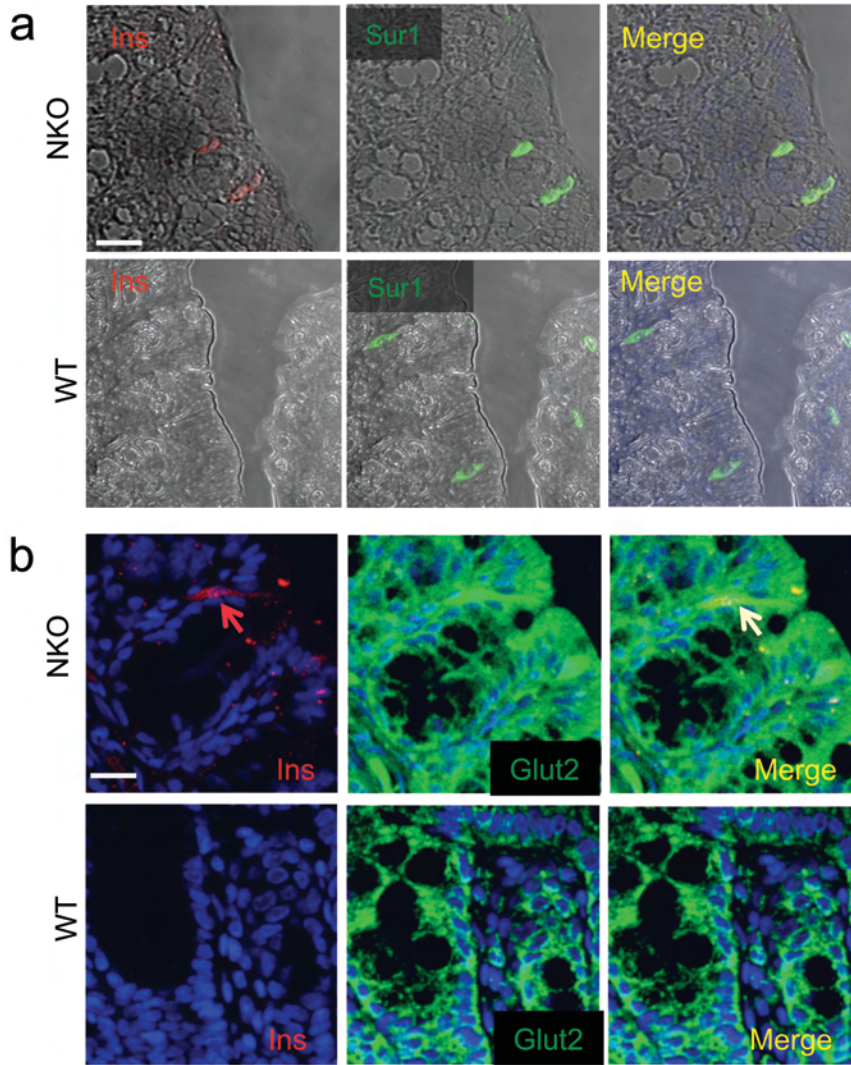


**Supplementary Figure 1** Neurog3 localization and expression. **(a)** Co-localization of Gfp (red) and Foxo1 (green) in intestines of control and *NKO* mice crossed with *Neurog3-Gfp* reporter mice. **(b)** Lack of co-localization of Neurog-Gfp with Foxo1 in *NKO* mice. **(c)** Immunofluorescence with anti-Neurog3 (red) and anti-Gfp (green) in neonatal gut from *Neurog3-Gfp* transgenic mice, indicating complete overlap between the two detection methods. **(d)** qPCR analysis of *Foxo1* mRNA expression from Neurog3-Gfp sorted cells. **(e)** Immunohistochemistry with anti-Neurog3 (brown) in adult large intestines. **(f-g)** Immunofluorescence with anti-Gfp (green) in adult gut from *Neurog3-Gfp* transgenic mice **(f)** and *Neurog3-Gfp* knock-in mice **(g)**. **(h)** Quantification of Neurog3-Gfp<sup>+</sup> cells using anti-Gfp by flow cytometry. Original magnification: 20X . n=3 for histology and n= 4 for qPCR and flow cytometry. \* =  $P < 0.05$ , \*\* =  $P < 0.01$

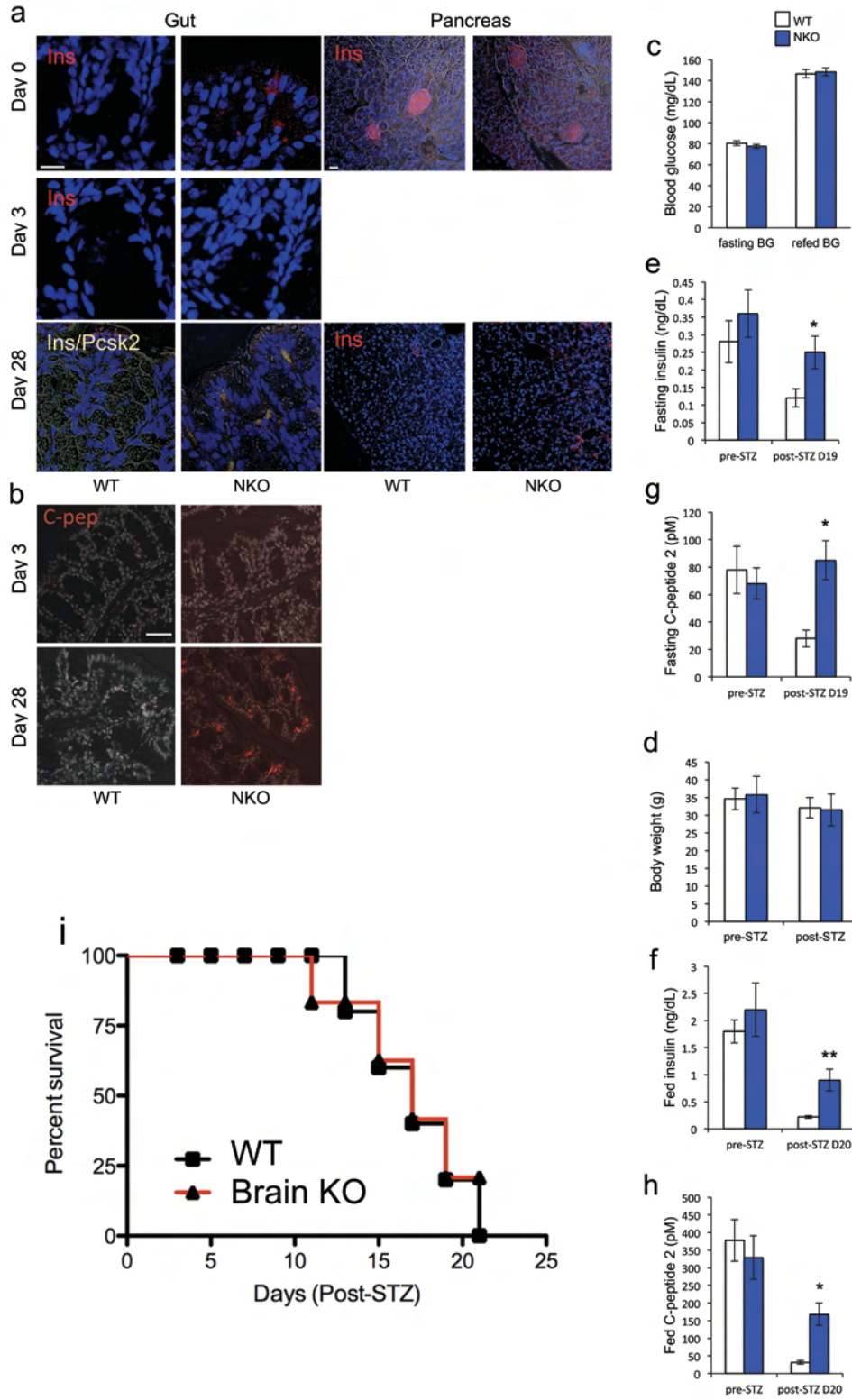


**Supplementary Figure 2** Gut immunohistochemistry. **(a)** Immunofluorescence with anti-Pancreatic polypeptide antibody (green) and insulin (red) in large intestines of adult WT and *NKO* mice. **(b-c)** Immunohistochemistry with anti-insulin (brown) in adult distal ileum **(b)** or colon **(c)**. RT-qPCR of *insulin1* and *tubulin2* expression in various tissues of

WT and *NKO* mice. **(e-g)** Generation and analysis of *PKO* mice. **(e)** Foxo1 (green) and Gfp immunochemistry (red) in 1 day-old *PKO:Neurog3-Gfp* and control mice. **(f)** Insulin immunostaining (red) in the duodenum of 9-month-old *PKO* mice. **(g)** Insulin (red) and Gfp immunostaining (green) in the duodenum of 1 day-old *PKO:Neurog3-Gfp* and *Neurog3-Gfp* control mice. **(h-i)** Direct fluorescence in pancreata **(h)** and villi **(i)** of control *Ins2-Gfp* and *NKO:Ins2-Gfp* mice. **(j)** Immunofluorescence with C-peptide (red) and Gfp (green) in *NKO:Ins2-Gfp* mice. **(k)** Co-localization of insulin (red) and Prohormone convertase 1 (Pc1) (green). **(l)** qPCR analysis of mRNA expression of markers of  $\beta$ -cell differentiation in isolated gut epithelial cell preparations from DTZ-enriched segments in *NKO* mice (blue bars) and from anatomically matched segments in control mice (black bars) (n = 8). \* =  $P < 0.05$ , \*\* =  $P < 0.01$ .

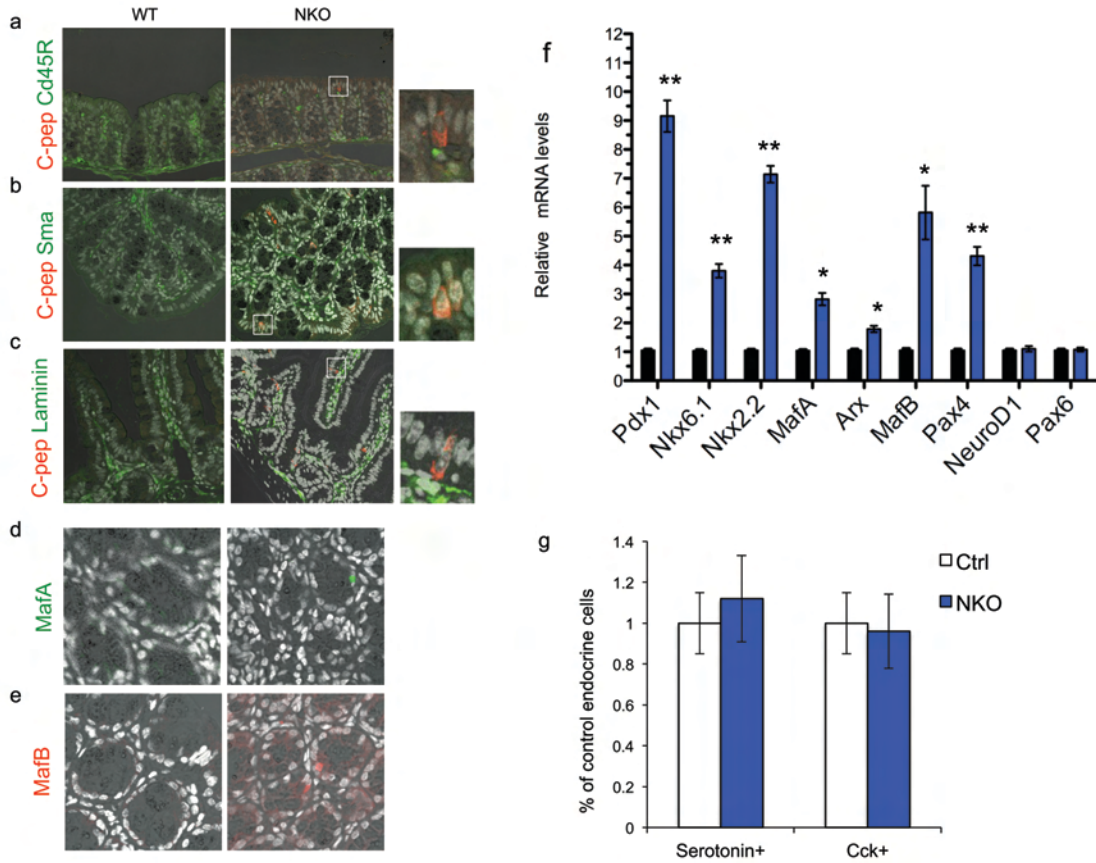


**Supplementary Figure 3** Marker analysis of gut Ins<sup>+</sup> cells by immunohistochemistry. **(a)** Insulin (red), Sulfonylurea Receptor 1 (Sur1), or **(b)** glucose transporter 2 (Glut2) antibodies (green). Scale bars: 40  $\mu\text{m}$  **(a)**, and 30  $\mu\text{m}$  **(b)**.



Supplementary Figure 4 Insulin, C-peptide, and Pcsk2 immunohistochemistry. (a)

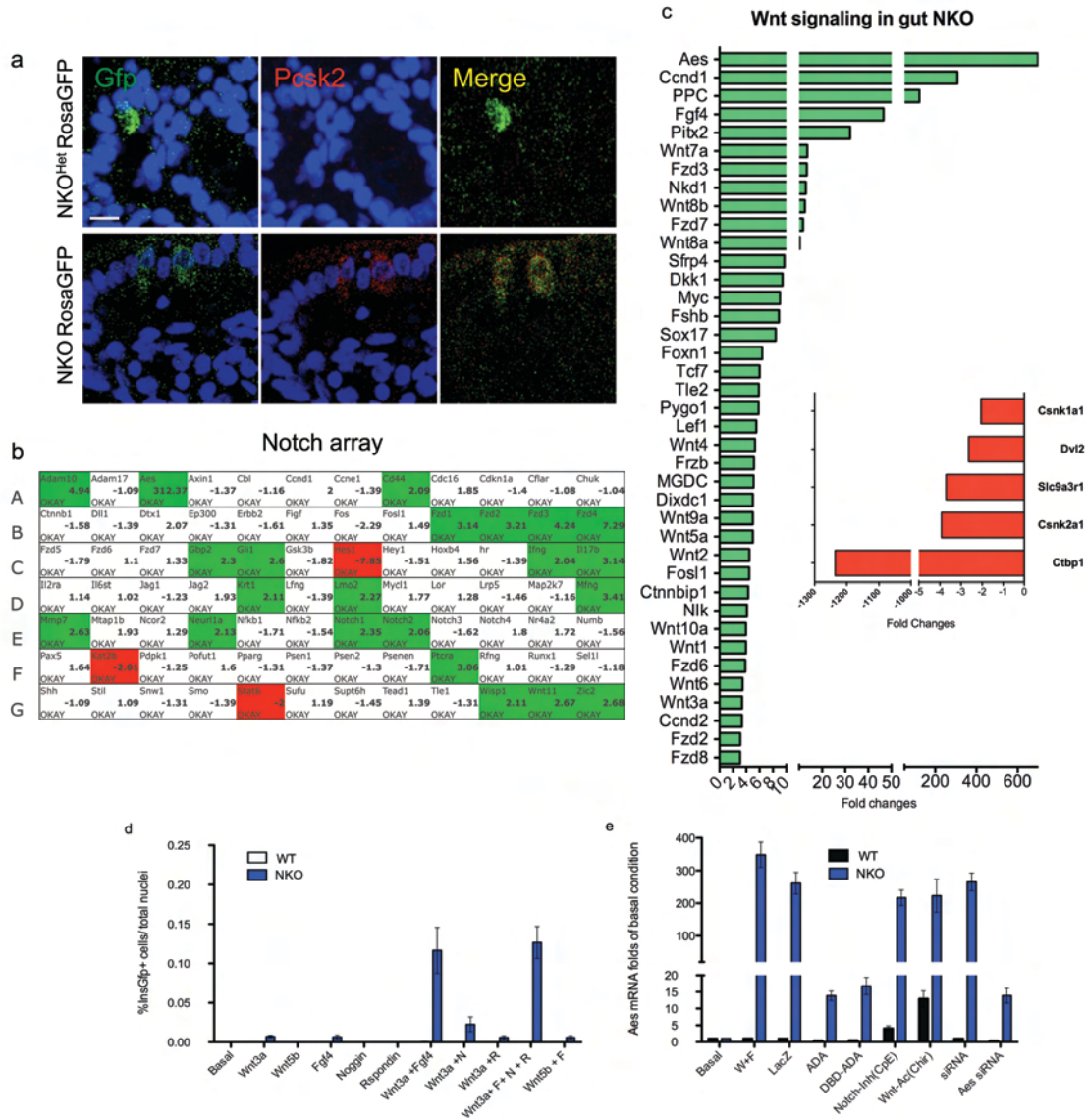
Insulin immunohistochemistry in gut (left two panels) and islet (right two panels) of *NKO* intestine before (D0), and after STZ injection (D3 and D28) with anti-insulin (red) (D0 and D3) and anti-insulin and anti-Pc2 antibodies (merged yellow fluorescence) (D28). **(b)** Immunofluorescence of anti C-peptide (red) after STZ injection day 3 (top) and day 28 (bottom). **(c-h)** metabolic profiles of WT (white bars) and *NKO* (blue bars) mice pre- and post-STZ treatment **(c)** fast and refed blood glucose levels, **(d)** body weight, **(e)** 12-hr-fasted serum insulin **(f)** fed serum insulin, **(g)** 12-hr-fasted serum C-peptide 2 **(h)** fed serum C-peptide 2. **(i)** survival plots of WT and *Foxo1* brain knock-out after STZ treatment (n=5). Scale bars: 30  $\mu\text{m}$  **(a)**, and 40  $\mu\text{m}$  **(b)**.



**Supplementary Figure 5** Analysis of epithelial and mesenchymal markers. **(a-c)**

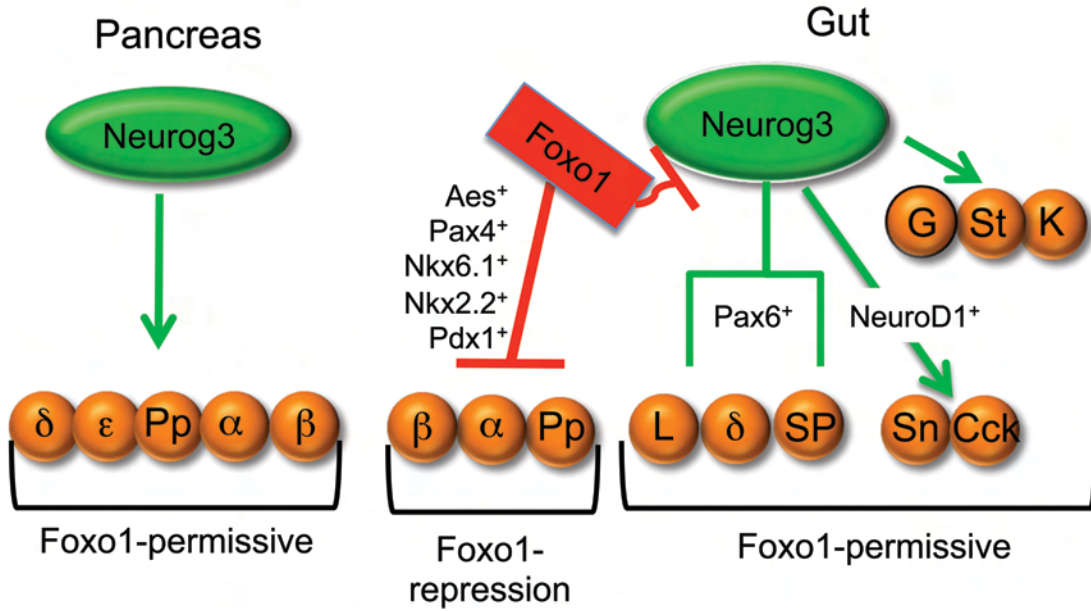
Immunohistochemical analysis of expression of C-peptide (red) and mesenchymal markers Cd45R **(a)**, smooth muscle actin (SMA)**(b)**, or laminin **(c)** (green) in adult colon and distal ileum of *NKO* mice and WT controls. **(d-e)** Immunostaining with antibodies to pancreatic islet transcription factors MafA **(d)**, and MafB **(e)**. **(f)** qPCR analysis of mRNA expression of markers of  $\beta$ -cell differentiation in isolated gut epithelial cell preparations from DTZ-enriched segments in *NKO* mice (blue bars) and from anatomically matched segments in control mice (black bars) (n = 8). **(g)** Morphometric quantitation of gut serotonin<sup>+</sup> cells and cholecystokinin<sup>+</sup> cells from small intestines normalized by total number of ChgA<sup>+</sup> cells (n=3). \* =  $P < 0.05$ , \*\* =  $P < 0.01$ .





**Supplementary Figure 6** Lineage tracing and low-density array profiling. **(a)** Lineage tracing of gut  $\text{Ins}^+$  cells post-STZ in *NKO:Rosa26Gfp* mice using Gfp (green) or Pcsk2 (red). Scale bars: 20  $\mu\text{m}$ . Gene transcript profiles of Notch signaling **(b)** and **(c)** Wnt signaling using low-density arrays with mRNA isolated from epithelial cells from DTZ-enriched gut segments. A green color indicates transcript levels increased > 2-fold in *NKO* compared to WT. A red color indicates transcript levels decreased > 2-fold in *NKO* compared to WT ( $p$ -value < 0.01). **(d)** Quantification of *Ins2-Gfp* differentiation assay of

primary gut culture: basal condition is without adding recombinant proteins, while other treatments include Wnt3a, Wnt5b, Fgf4, Noggin, R-spondin. (e) qPCR analysis of *Aes* mRNA isolated from *ex vivo* gut culture in Fig. 6f (n = 6).



**Supplementary Figure 7** Model of Foxo1 function in endocrine differentiation. In the pancreas, Foxo1 is permissive for terminal differentiation of the endocrine lineage ( $\alpha$ ,  $\beta$ ,  $\delta$ ,  $\epsilon$ , and Ppy cells). In the gut, Foxo1 actively prevents generation of pancreas-like endocrine cells ( $\beta$ ,  $\alpha$ , and Ppy), but is permissive for the generation of enteroendocrine cells. G: gastrin, K: K cells, L: L cells, SP: substance P, Sn: secretin, St: serotonin, Cck: colecystokinin.