

SUPPLEMENTARY ONLINE DATA

Calcium/calmodulin inhibition of the *Arabidopsis* BRASSINOSTEROID-INSENSITIVE 1 receptor kinase provides a possible link between calcium and brassinosteroid signalling

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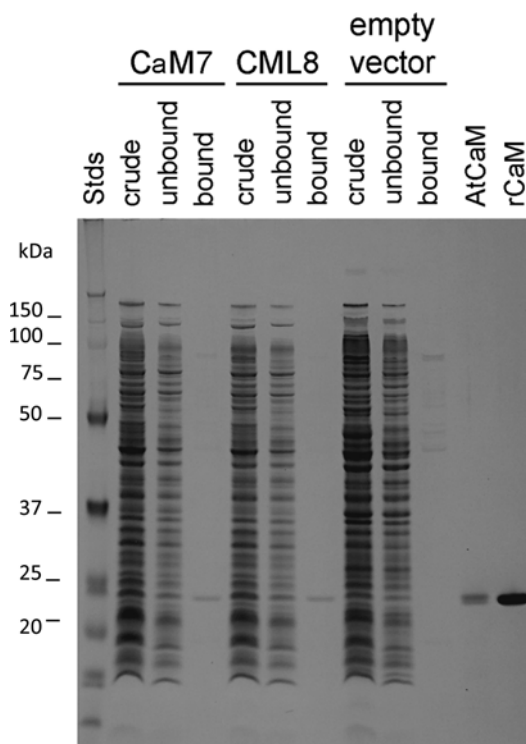


Figure S1 Constitutive expression of CaM in *E. coli* driven by pRZ528 (CaM7) and 529 (CML8)

Total soluble proteins were extracted from late-exponential-phase bacteria, clarified by centrifugation and subjected to Ca²⁺-dependent HIC (hydrophobic interaction chromatography) on phenyl-Sepharose. Samples of the crude extract, the fraction that did not bind to phenyl-Sepharose (unbound), and the fraction that eluted from the resin in the presence of EDTA (bound) were separated by SDS/PAGE and stained with Coomassie Brilliant Blue. Purified AtCaM and recombinant CaM (rCaM) were fractionated in parallel as positive controls together with molecular mass standards (Stds, sizes given in kDa).

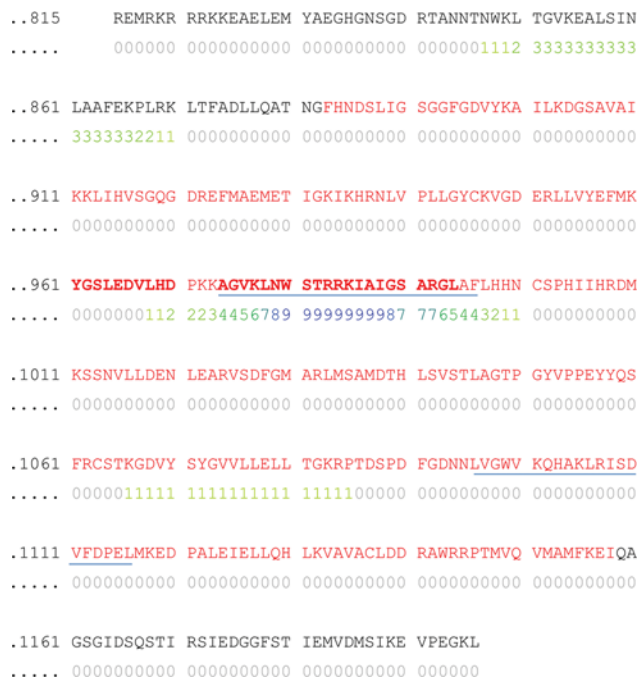


Figure S2 BRI1 cytoplasmic domain sequence showing predicted probabilities of CaM binding

The numbers below the sequence indicate the probability of CaM binding (<http://calcium.uhnres.utoronto.ca/ctdb/ctdb/home.html>) with 9 being the highest score. The kinase domain of BRI1 is shown in red, and the sequences used to generate the W980 and W1099 peptides are underlined.

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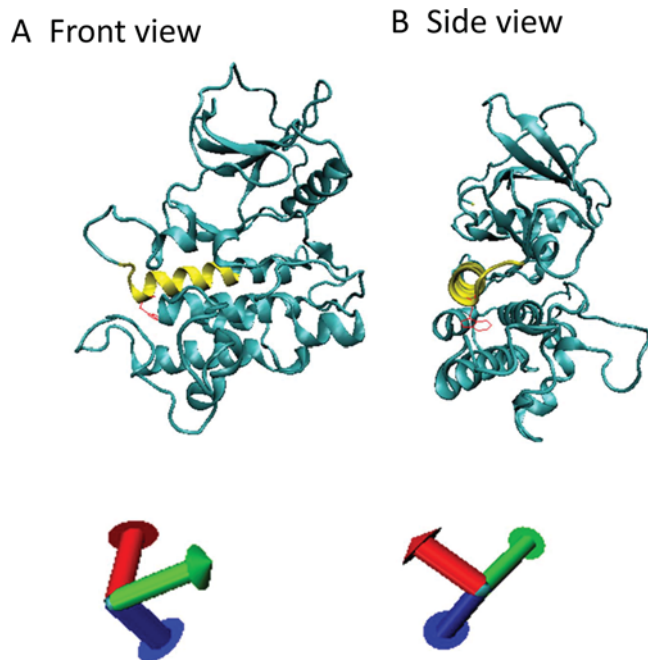


Figure S3 Model of the BRI1 kinase domain structure

The BRI1 kinase domain was modelled as described in the Experimental section of the main text. The location of the predicted CaM-binding site (Leu⁹⁷⁸–Arg⁹⁹²) is shown in yellow and the Try⁹⁶⁰ side chain is shown in red. **(A)** Front view. **(B)** Side view.

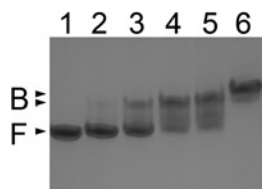


Figure S4 Gel mobility-shift analysis of AtCaM6 interaction with W980 peptide

AtCaM6 (300 pmol/reaction) was incubated with 0 (lane 1), 150 (lane 2), 300 (lane 3), 600 (lane 4) or 1200 (lane 5) pmol of W980 peptide or 300 pmol of a CaM-binding peptide from CaMKII (lane 6) then fractionated in a non-denaturing gel containing Ca^{2+} . Coomassie Brilliant Blue staining revealed the migration of free (F) and peptide-bound (B) AtCaM6. The larger size of the CaMKII peptide resulted in a slower mobility of the peptide–AtCaM6 complex compared with the W980 peptide complex.

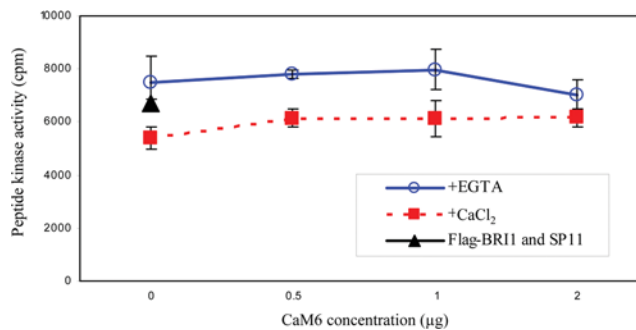


Figure S5 AtCaM6 has no effect on BRI1 peptide kinase activity *in vitro*

BRI1 peptide kinase activity was assayed using [³²P]ATP and the SP11 synthetic peptide as substrate as described in [1] with the addition of 0.1 mM CaCl_2 or 1 mM EGTA as indicated. Each reaction contained 0.5 µg of FLAG–BRI1 protein, and reactions were run for 5 min.

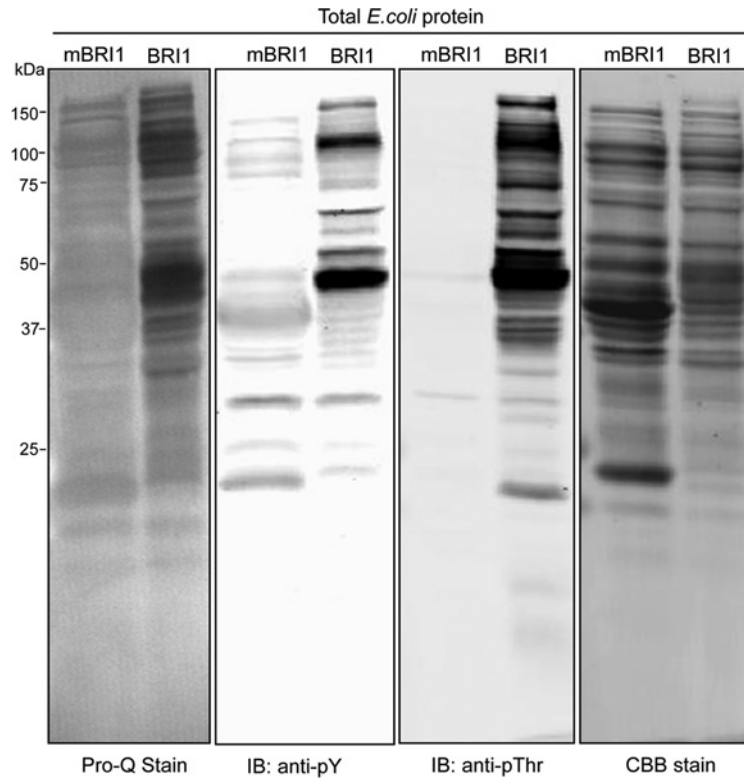


Figure S6 Kinase-inactive mBRI1 does not transphosphorylate *E. coli* proteins

BRI1 or the kinase-inactive mBRI1 (mutant BRI1) were expressed in *E. coli* and following purification of the FLAG-tagged proteins, the remaining *E. coli* proteins were analysed for phosphorylation by Pro-Q Diamond phosphoprotein staining or immunoblotting (IB) with generic anti-phosphotyrosine or anti-phosphothreonine antibodies. Molecular masses are indicated in kDa.

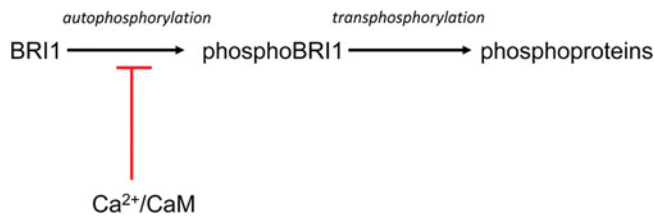


Figure S7 Working model for effects of Ca²⁺/CaM on BRI1

We propose that Ca²⁺/CaM inhibits BRI1 autophosphorylation, which then affects transphosphorylation activity. Thus certain CaM and CML isoforms inhibit BRI1 autophosphorylation when co-expressed in *E. coli* and, as a result, also inhibit the transphosphorylation of *E. coli* proteins. This provides a plausible explanation for why Ca²⁺/CaM inhibits transphosphorylation in the *in situ* system, but does not affect transphosphorylation activity *in vitro* (peptide kinase activity). The model predicts that Ca²⁺ signalling during early stages of BRI1 activation will attenuate downstream signal transduction.

Table S1 Continued

Source	Site	Name	M_r (expt)	M_r (calc)	Delta	MC	Score	Expect	Peptide
TiO ₂	Thr ³	gatZ	911.2322	911.4663	-0.2341	1	46	0.0025	-MK p TLIAR.H
TiO ₂	Ser ⁴⁷⁶	aspA	1106.1322	1106.3917	-0.2595	1	45	0.0017	K.RYTDE p SEQ.-
TiO ₂	Thr ⁶²⁶ /Thr ⁶³⁰	pnp	1082.1788	1082.4563	-0.2774	1	36	0.0098	R.VY p TGKV p TR.I
TiO ₂	Ser ³ /Thr ⁸	gatD	1592.2680	1592.6671	-0.3990	1	41	0.00036	-MK p SVVND p TDGIVR.V
TiO ₂	Thr ⁸⁷ /Thr ⁹⁷	sucB	1878.3088	1878.7761	-0.4673	2	40	0.0014	K.E p TSAKSEEKAS p TPAQR.Q
TiO ₂	Ser ⁷⁵	frr	927.1428	927.3773	-0.2344	0	39	0.0078	R.SM p SPAVEK.A
TiO ₂	Ser ²	lipA	1038.2284	1038.4933	-0.2648	0	30	0.019	M. p SKPIVMER.G
IMAC	Thr ⁶	yqjD	1316.2666	1316.5874	-0.3208	1	54	5.00×10^{-5}	M.SKEH p TTEHLR.A
IMAC	Thr ⁷	yqjD	1316.2684	1316.5874	-0.3190	1	33	0.0047	M.SKEH p TTEHLR.A
IMAC	Thr ⁶ /Thr ⁷	yqjD	1396.2338	1396.5537	-0.3199	1	67	5.80×10^{-6}	M.SKEH p T p TTEHLR.A
IMAC	Thr ¹⁷	espF	921.2758	921.4069	-0.1311	0	29	0.026	R.H p TSASR.V
IMAC	Thr ⁷⁶	grpE	1238.3146	1238.6020	-0.2873	2	28	0.039	R.RR p TELDIEK.A
IMAC	Thr ⁹⁴	tufA	2736.8350	2737.2639	-0.4289	0	25	0.024	K.NM p TGAAGMDGAILVVAATDGPMPQTR.E
P-bodies	Thr ¹⁷⁶ /Ser ¹⁸⁰	ahpC	1658.2694	1658.7205	-0.4511	0	35	0.0025	K.EGE p TL p PSLDLVGK.I

REFERENCE

- Oh, M.-H., Ray, W. K., Huber, S. C., Asara, J. M., Gage, D. A. and Clouse, S. D. (2000) Recombinant brassinosteroid insensitive 1 receptor-like kinase autophosphorylates on serine and threonine residues and phosphorylates a conserved peptide motif *in vitro*. *Plant Physiol.* **124**, 751–766

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