

Supporting Information

Title: Role of a SpoVA Protein in Dipicolinic Acid Uptake into Developing Spores of *Bacillus subtilis*

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Supporting Information List

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Table S1. Structural homologs of the *B. subtilis* SpoVAD; nonredundant top-400 DaLi results.

PDB id	Z-score	Rmsd (Å)	Aligned residues #	Protein
3H76	24.8	3.3	261	PQS biosynthetic enzyme PqsD
2GYO	24.7	3.3	254	β -ketoacyl-[acyl-carrier-protein] synthase III
3OYT	24.2	3.1	263	β -ketoacyl-[acyl-carrier-protein] synthase I
1WL4	24.1	2.8	258	acetyl-CoA thiolase
3E60	23.8	3.2	271	β -ketoacyl-[acyl-carrier-protein] synthase II
3HHD	23.5	3.2	271	human fatty acid synthase
3EUO	23.1	3.2	264	fungus type III polyketide synthase
1U0W	22.7	3.3	271	stilbene synthase
3A5Q	22.6	3.2	266	benzalacetone synthase
1EE0	22.6	3.2	267	2-pyrone synthase

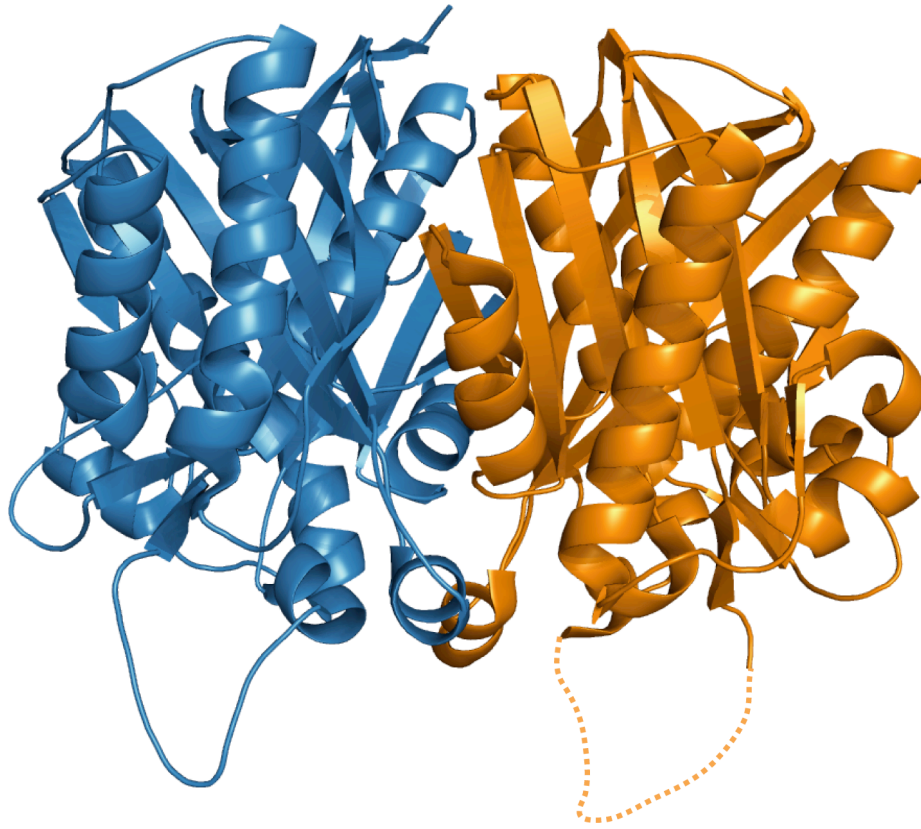


Figure S1. **Ribbon diagram of the SpoVAD crystallographic dimer (PDB code 3LM6).** The two monomers are colored blue and orange, respectively. Dotted lines represent disordered regions.

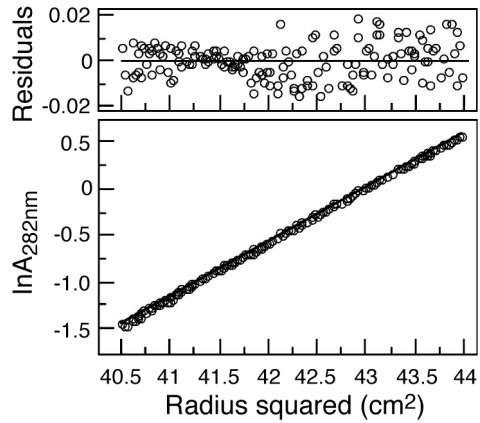


Figure S2. **SpoVAD sediments as a monomer in solution.** Equilibrium sedimentation data (16,000 rpm) of SpoVAD (60 μM) at 20°C in a buffer containing 15 mM Tris-HCl (pH 7.6), 100 mM NaCl and 2 mM dithiothreitol. The data fit closely to a monomer. (Upper) The deviation in the data from the linear fit for a monomeric model is plotted.

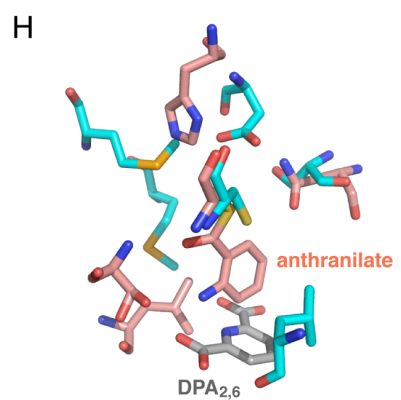
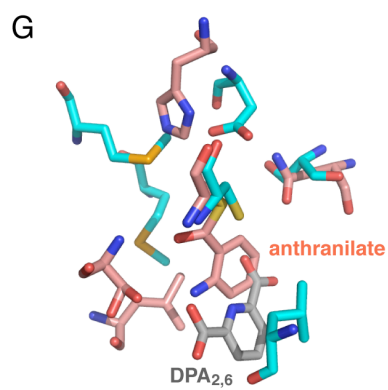
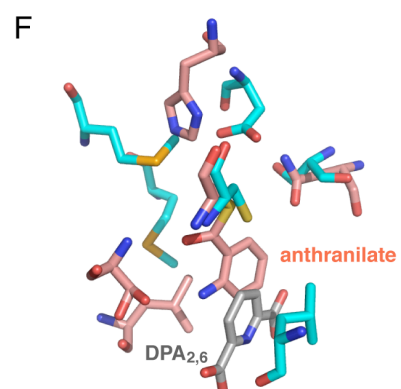
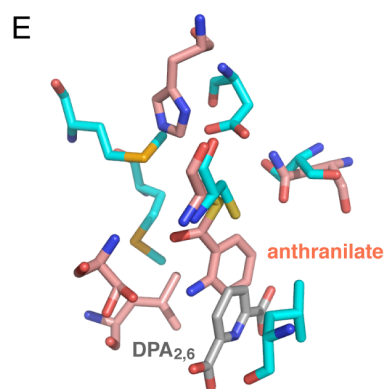
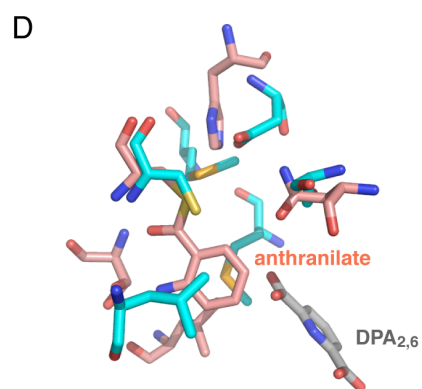
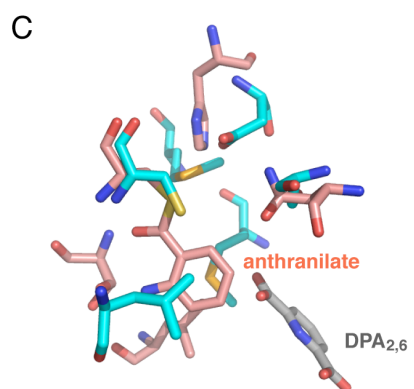
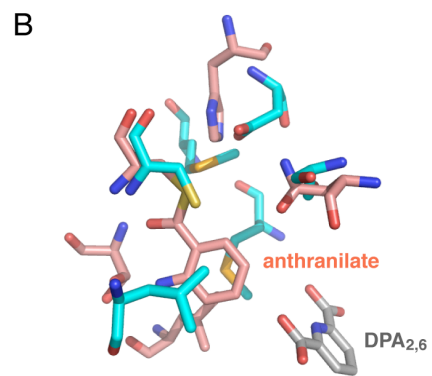
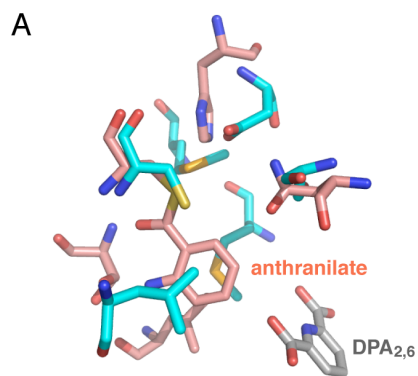
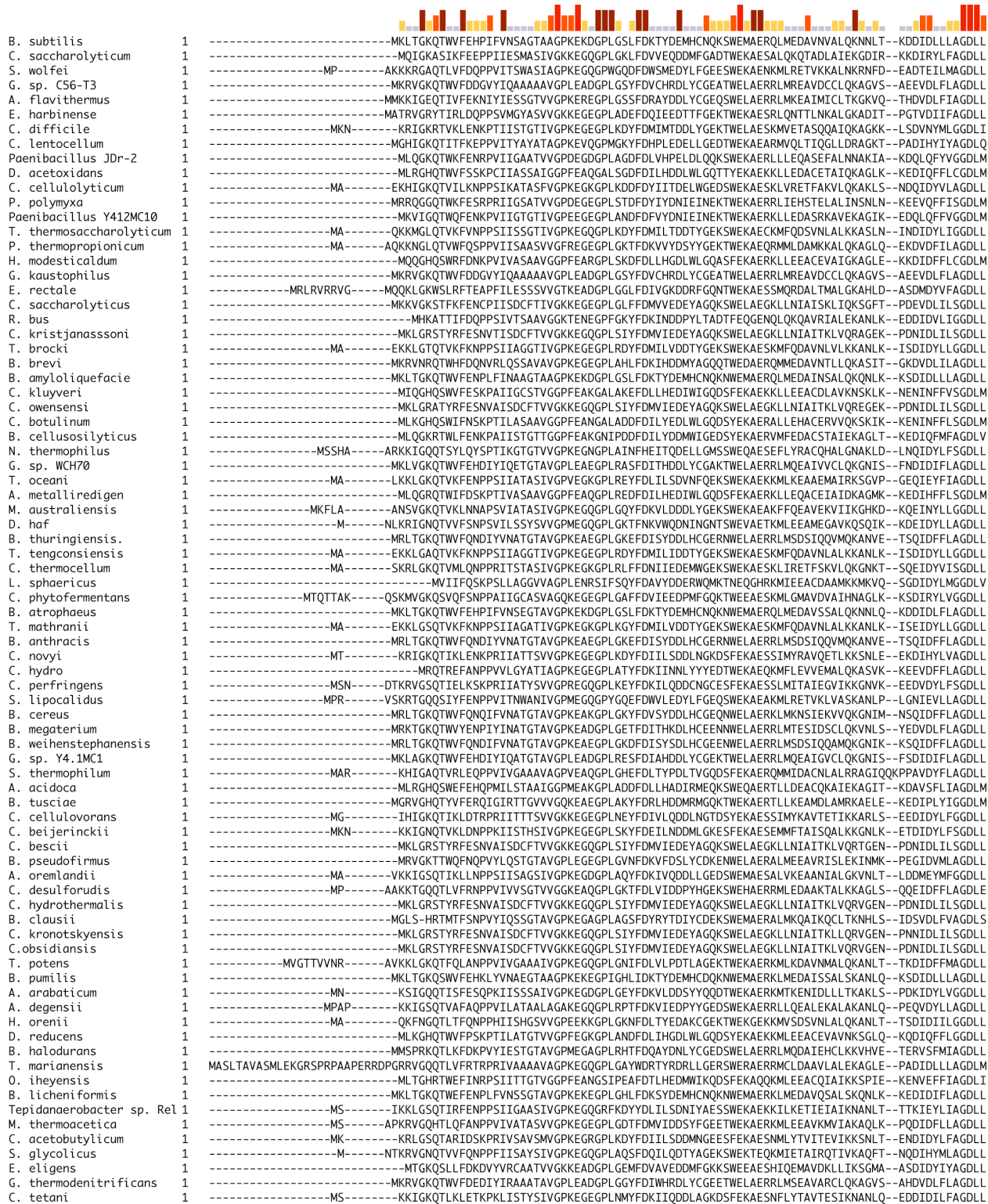



Figure S3. **The docking conformations of the DPA_{2,6} molecule to SpoVAD.** A-H, Eight of the top nine docking solutions for the DPA_{2,6} binding to SpoVAD (PDB code 3LM6) using AutoDock Vina. DPA_{2,6} was docked as a rigid body and all the nine solutions found the DPA_{2,6} molecule in the same presumed binding site in SpoVAD.



Conservation: 100-80-60-40-20%

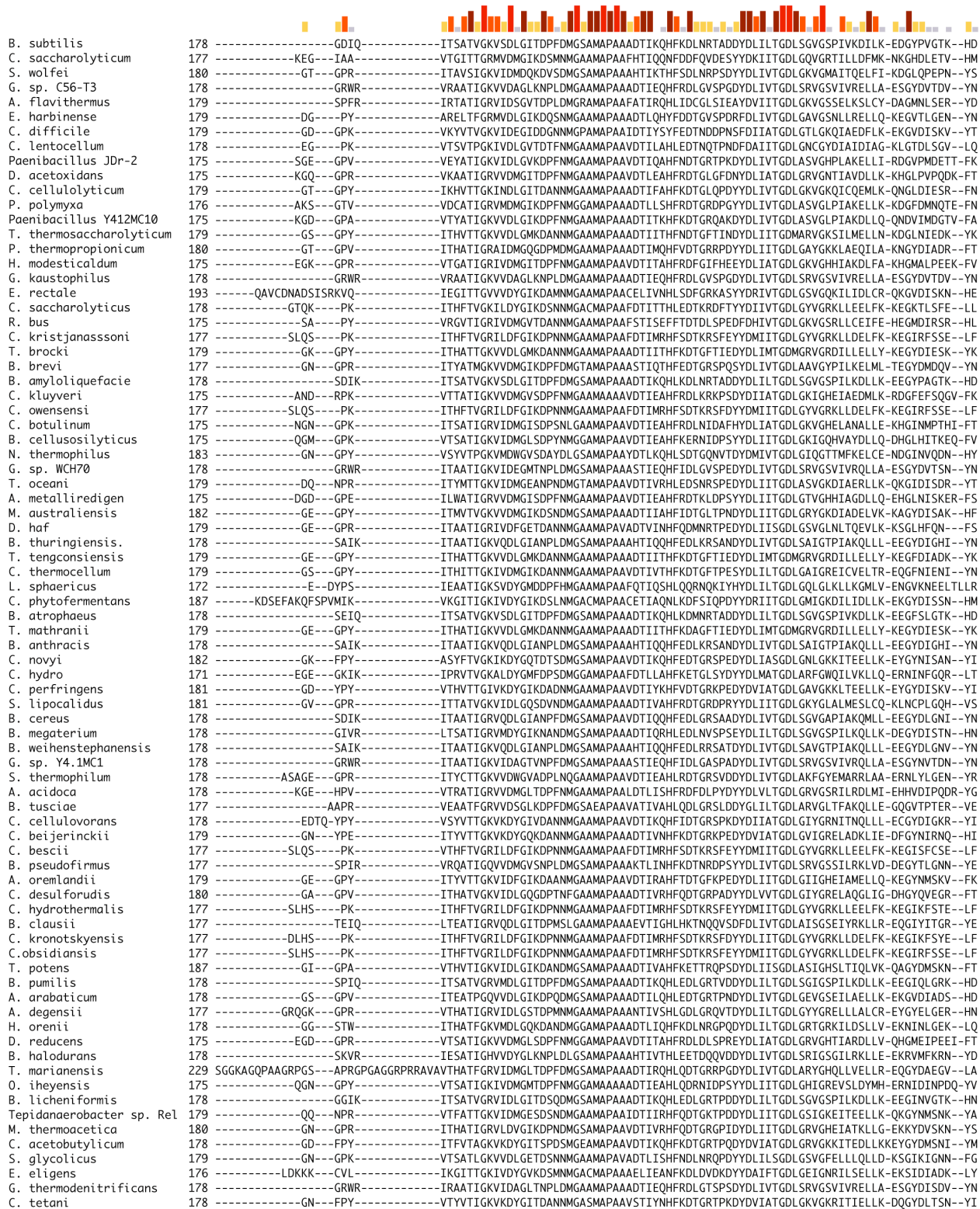
Figure S4 (1 of 4) (Hao)



B. subtilis	87	NQNVNTANYVARHLKIPFLCMFGACSTSMETVAVASALVDGGFAKRALAATSSHNATAERQFRYPTEYGGQKPDATSTVTTGSGAVVISQTP
C. saccharolyticum	86	GQLIATISFGVDLEIPLFGLFGACSTMGEALNLGAMTVAGGYADKVMMAASSHFATAEKQFRFPLESYGNQRPYASWTVTGAGAVLAKHR
S. wolfei	90	NQLVSSNYAARELGIPFLGLYGACSTMESLALLAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP
G. sp. C56-T3	87	NQNTANSYVARELPIPLCLFGACSTMQLAAAAFVAGGLADRALAATSSHNATAERQFRYPTEYGGQKPKTATFTVTGAGAVLGRAP
A. flavithermus	88	NQTVTANYVASELHPIPLGVFACSTMTEAMLVASTFDGAFHRVLAATSSHYGAERQFRFPTEYGGQKESASWTVTGAGAAALSTTR
E. harbinense	88	NQICGTSYGLREFGIPFAGLYGACSTMESLAMAALVFEVGGFANVAALTSCHFSAERQYRPLLEYGQRAPSAQWTVTGAGAVLAPN
C. difficile	89	NQIVPASFARELAIPFLGIYGACSTMAGLCTGSMIDGGFADLVLSGTSYHCTAERQFRFPLELGNQKPMTAQWTVTGAGAVLAPN
C. lentocellum	87	NQVIASFTGKDFNIPFFGLYGACSTMESITLASMCIAGGMADLVIAGTSSHNCAAEKQFRFPLEYGGQRTETQWTVTASGFVLLSRGC
Paenibacillus JDr-2	86	NQIISNSFAARTLGVNPLGVFACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVVAA
D. acetoxidans	86	NQIVSSSFAARTLGVNPLGVFACSTMESLALALLDKAAKYVLCATASHNASVEKQFRYPTEYGGQKPPAQWTVTGAGAAI LAP
C. cellulolyticum	89	NQICIAANGLRETEVPFLGLYGACSTMESLMSISLMEVGGFAENLVCMTSSHFCSAEKQFRFPLELGSORTPTSQWTVTGAGAVLSSK
P. polymyxa	87	NQIVSSSFAAKLIPYLVGFACSTMESLALALVDSAGDYVMAGTSHNCTVEKQFRYPTEYGGQKPPAQWTVTGAGAAIVGH
Paenibacillus Y412MC10	86	NQIISSSFAARQMGVPIYGVFACSTMESLAVASMLVDSGAGQYVMAGTASHNCTVEKQFRYPTEYGGQKPPYQYITGAGCSVSH
T. thermosaccharolyticum	89	NQIITSSFAARQFNVPFLGLYGACSTMAGLCTGSMIDGGFADYVIVTSSHFSTAERQYRFPLEGGQRPPTAQWTVTGAGAVLSS
P. thermopropionicum	90	NQIISANFTASSMSVPLGLYGACSTMIEGMALGAMLDGGFADRVLAATSSHYSTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
H. modesticaldum	86	NQITASSFAARTMAVPYLVGFACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVVAP
G. kaustophilus	87	NQNTANSYVARELPIPLCLFGACSTMQLAAAAFVAGGLADRALAATSSHNATAERQFRYPTEYGGQKPKTATFTVTGAGAVLGRAP
E. rectale	97	GSMAITTFGVENFRIPWFGVYACSTMESLALAAAFVAGGYARHVAVAVTSSHFSAEKQFRFPNQYANQRPVSAWTVTGAGAVVSGAGNET
C. saccharolyticus	87	NQITSSFAARDLNIPFLGIYGACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
R. bus	84	DQCVGTYTVGRDYLPFLGIYAACTSMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
C. kristjanassoni	86	NQIISSSFAARDLDIPFGLYGACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
T. brocki	89	NQIISANFAARQLNVPFHLGLYGACSTMTEGLTLGAMLDGGYADYIAATSSHFSTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
B. brevis	87	NQNTANSYVARELPIPLCLFGACSTMESLALAAAFVAGGLADRALAATSSHNATAERQFRYPTEYGGQKPKTATFTVTGAGAVLGRAP
B. amyloliquefacie	87	NQNVNTANYVARHLKIPFLCMFGACSTSMETVAVASALVDGGFAKRALAATSSHNATAERQFRNPTEYGGQKPDATSTVTTGAGAVISQNP
C. kluyveri	86	NQIITSSFAARTLKIPLYGVFACSSMEGLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
C. owensensi	86	NQIISSSFAARDLDIPFGLYGACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
C. botulinum	86	NQIVSSSFAARTLGIYVYFGLYGACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
B. cellulosilyticus	86	NQLTPTSSFAARTLAAPYLVGFACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVVAP
N. thermophilus	93	NQIVSSSNLVAKLLQIPYLVGFACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVVAP
G. sp. WCH70	87	NQNVNTANYVAQTIPFLCMFGACSTSMETVAVASALVDGGFAKRALAATSSHNATAERQFRYPTEYGGQKPKTATFTVTGAGAVLGRAP
T. oceanii	89	NQIITASSFAARELGPFFGIYGACSTMAGLCTGSMIDGGFADYVIVTSSHFSTAERQYRFPLEGGQRPPTAQWTVTGAGAVLARE
A. metalliregigen	86	NQIITSSSFAARTLGMPLYGVFACSSMEGLALAAALVDSGAGKNAALAAASHNAAVEKQFRYPTEYGGQKPPAQWTVTGAGAVVAP
M. australiensis	92	NQIISDFAAARQLGIPFGLYGACSTMESAMALGAMLDGGFADYVIVTSSHFSTAERQYRFPLELGVQRPTAHWTVTGAGAAVNSQ
D. haf	89	NQIISANFAARTMGIPFLGLYGACSTMALMSVGTMLIDGGFARKVLAATSSHHTAERQYRFPTEYGGQKPPAQWTVTGAGAVLGR
B. thuringiensis	87	NQNVNTANYVARKNIPFLGMFACSTSMETLAVGSAFIDGGFANRVLATVSSHNTAERQFRYPTEYGGQKPGTANSTVTTGAGAVLSQTP
T. tengconsiensis	89	NQIISANFAARQLNVPFHLGLYGACSTMTEGLTLGAMLDGGYADYIAATSSHFSTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
C. thermocellum	89	NQICIAANGLRESDVPFFGVYACSTIAESMISIGAMLDVGGFADNVCTSSHFCSAEKQFRFPLELGTQRPPSAQWTVTGAGAVLARE
L. sphaericus	81	NQMTPTNFARLDLIPFGLYGACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
C. phytofermentans	95	GQLIATISFGIENFLEIPLGVYACSTMESLALAAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP
B. atrophaeus	87	NQNVNTANYVARHLKIPFLCMFGACSTSMETVAVASALVDGGFAKRALAATSSHNATAERQFRNPTEYGGQKPDATSTVTTGAGAVISQNP
T. mathranii	89	NQIISANFAARQLNVPFHLGLYGACSTMTEGLTLGAMLDGGYADYIAATSSHFSTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
B. anthracis	87	NQTVTANYVARKNIPFLGMFACSTSMETLAVGSAFIDGGFANRVLATVSSHNTAERQFRYPTEYGGQKPGTANSTVTTGAGAVLSQTP
C. novyi	88	NQITASSFAARDVDIPFLGLYGACSTMESLAVASMLVDSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
C. hydro	83	NQIISANFAARQLGIPFGLYNACTFYEGLLGAMLDGGFADYVIVTSSHFSTAERQYRFPLEGGQRPPTAQWTVTGAGAVLARE
C. perfringens	91	NQIISNSFAARELKIPLYGVFACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
S. lipocalidus	91	NQIISANFAARELKIPLYGVFACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
B. cereus	87	NQNVNTANYVARDMIPFLGMFACSTSMETLAVGSAFIDGGFANRVLATVSSHNTAERQFRYPTEYGGQKPPAQWTVTGAGAVLARE
B. megaterium	87	NQNVNTANFVARHNIQIPFLCMFGACSTSMETLAVGSAFIDGGFANRVLATVSSHNTAERQFRNPTEYGGQKPDATSTVTTGAGAVISQNP
B. weihenstephanensis	87	NQNVNTANYVAREYIPFLGMFACSTSMETLAVGSAFIDGGFANRVLATVSSHNTAERQFRNPTEYGGQKPDATSTVTTGAGAVISQNP
G. sp. Y4.1MC1	87	NQNVNTANYVARTVAPFLCMFGACSTSMETLAVGSAFIDGGFANRVLATVSSHNTAERQFRNPTEYGGQKPDATSTVTTGAGAVISQNP
S. thermophilum	91	NQLITATFASGLGLVPLGLYGACSTMTEGLTLGAMLDGGYADYIAATSSHFSTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
A. acidoca	86	NQIISNSFAARELKIPLYGVFACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
B. tusciae	87	DQITTTNFAASFLGSMYLVGFACSTMESLALAAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP
C. cellulovorans	88	NQLTATSFARVRLNIPFGLYGACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
C. beijerinckii	89	NQIISNSFAARELKIPLYGVFACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
C. besicii	86	NQIISNSFAARELKIPLYGVFACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
B. pseudofirmus	86	NQIVTSSNYAARQLSIPFLGMFACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
A. oremlandii	89	NQIMSTSSFAARDMKVPFLGLYGACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
C. desulfuridis	90	NQIILASSFAACSLGPHLGLYGACSTMESLALAAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP
C. hydrothermalis	86	NQLSSSFAARDLDIPFGLYGACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
B. clausii	86	NQITASSFVARDLQPLGYGACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
C. kronotskyensis	86	NQIISNSFAARELKIPLYGVFACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
C. obsidiansis	86	NQIISNSFAARELKIPLYGVFACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
T. potens	97	NQIISANFTARDLGIPLYGVFACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRYPTEYGGQKPPAQWTVTGAGAVLARE
B. pumilis	87	NQNVNTANYVARELKIPLYGVFACSTMESLALASLMDVSGAGQYVLAGTCSHNTAEKQFRNPTEYGGQKPDATSTVTTGAGAVISQNP
A. arabaticum	88	NQVVTNSFAVKFIPYVGIYACSTLAELGLGAVLDGGFANRVLATVSSHNTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
A. degensii	90	NQIVANFAARALGIPYVGLYGACSTIYEGLLGAMLDGGFADYVIVTSSHFSTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
H. oreii	88	NQLITANFVARDYVPLGLYGACSTMESLALAAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP
D. reducus	86	NQIISNSFAARELKIPLYGVFACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
B. halodurans	87	NQNVNTANYVARLGIPLYGVFACSTMESLALAAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP
T. marianensis	114	NQLITCANFTAELDIPFLGLFAACATWAAALGLAAMAADGAYATVLCITSSHHTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
O. iheyensis	86	NQITPTSSFAARTVGPYLVGFACSTMESLALAAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP
B. licheniformis	87	NQNVNTANYVARHLKIPFLCMFGACSTSMETVAVASALVDGGFAKRALAATSSHNATAERQFRNPTEYGGQKPDATSTVTTGAGAVISQNP
Tepidanaerobacter sp. Rel	89	NQIISAGFAARELKIPLYGVFACSSMEGLALAAQIEKAAKYVITAAASHNAAAEKQFRYPTEYGGQKPPAQWTVTGAGAVLGR
M. thermoacetica	90	NQIISANYAARSLGIPLYGVFACSTMIEGMALGAMLDGGFADRVLAATSSHYSTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
C. acetobutylicum	88	NQIITSSSFAARDKIPYLVGFACSTMESLALAAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP
S. glycolicus	89	NQIISANFTARQLLPIPLYGVFACSTMALTTALGMLIDGGFANRVLATVSSHHTAERQYRFPTEYGGQKPPAQWTVTGAGAVLARE
E. eligens	85	NQLIATSFGLMRYEIPMFLYGACSTMESLALAAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP
G. thermodenitrificans	87	NQNTANSYVARELPIPLCLFGACSTMQLAAAAFVAGGLADRALAATSSHNATAERQFRYPTEYGGQKPKTATFTVTGAGAVLGRAP
C. tetani	88	NQLSSSFAARDMNIPIPLYGVFACSTMESLALAAALVDGFEFFERVAIAAASHHYTAERQFRFPTEYGGQKTPSSQWTAAGAVLLEVP

Conservation: 100-80-60-40-20%

Figure S4 (2 of 4) (Hao)



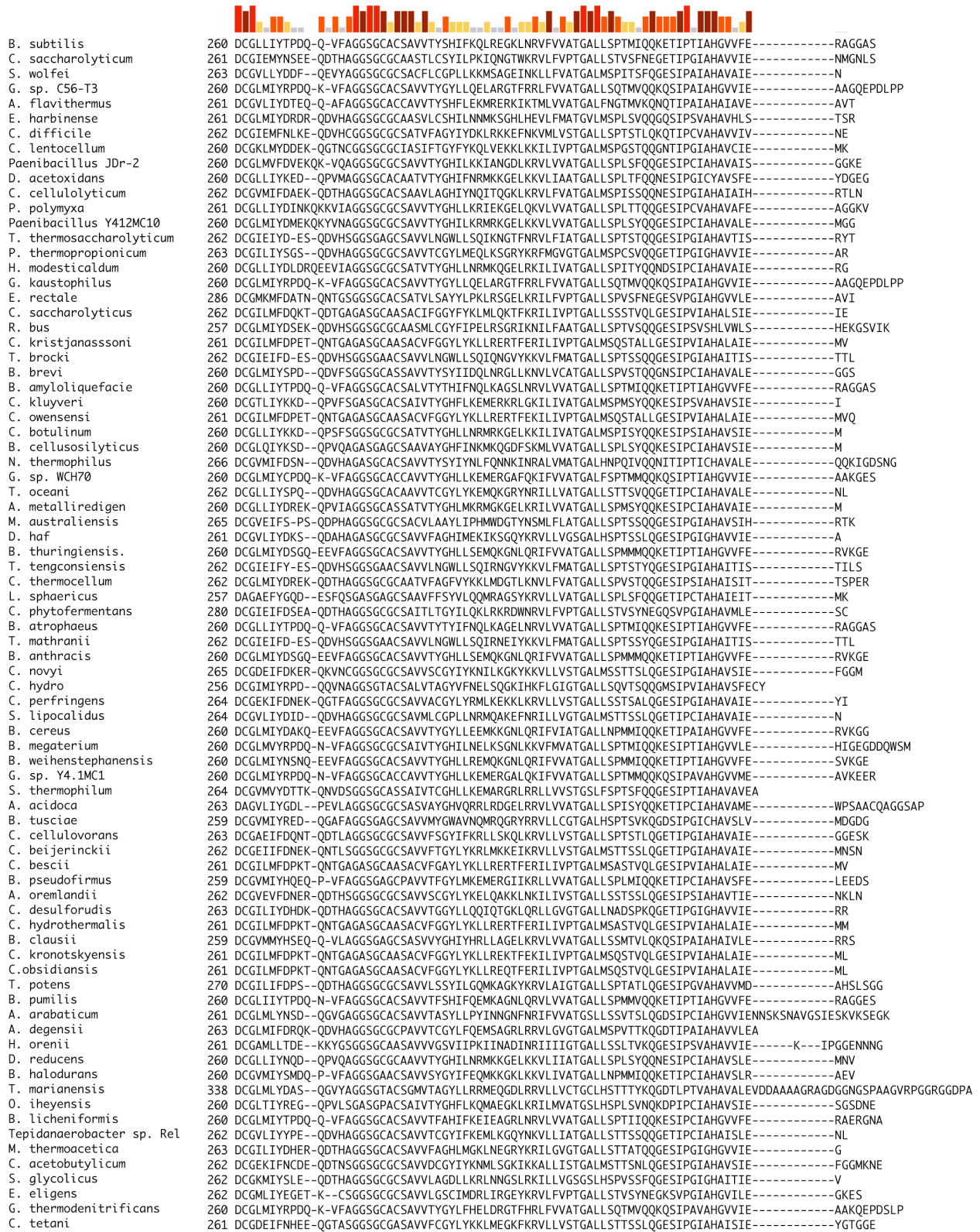


Figure S4 (4 of 4) (Hao)

Figure S4. **Sequence alignment of SpoVAD orthologs in the *Bacillus* and *Clostridium* species.** A ClustalW alignment of 80 SpoVAD orthologs was performed using DNASTAR Lasergene suite 8 (DNASTAR Inc., Madison, WI). Sequence conservation is shown as a bar graph, with red bars indicating identity among SpoVAD homologs.

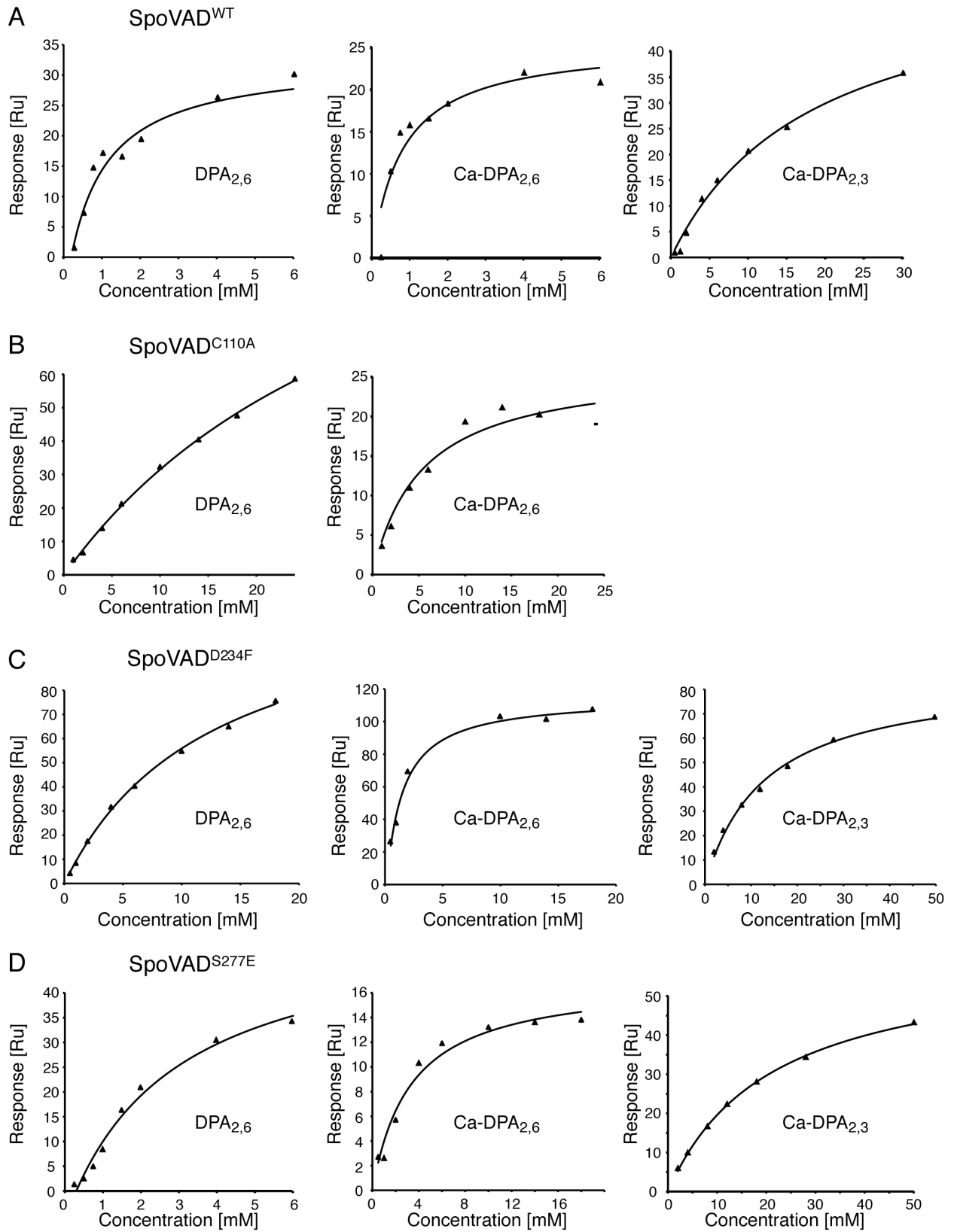


Figure S5. **Binding of DPA and Ca-DPA to mutant SpoVAD proteins.** An overlay of the representative Biacore SPR sensorgrams displaying the association curves of 2 mM DPA (DPA_{2,6}, DPA_{2,3}, DPA_{2,5} and DPA_{3,5}; left) and Ca-DPA (Ca-DPA_{2,6}, Ca-DPA_{2,3}, Ca-DPA_{2,5} and Ca-DPA_{3,5}; right) with the SpoVAD^{C110A} (A), SpoVAD^{D234F} (B) and SpoVAD^{S277E} (C) mutant proteins.

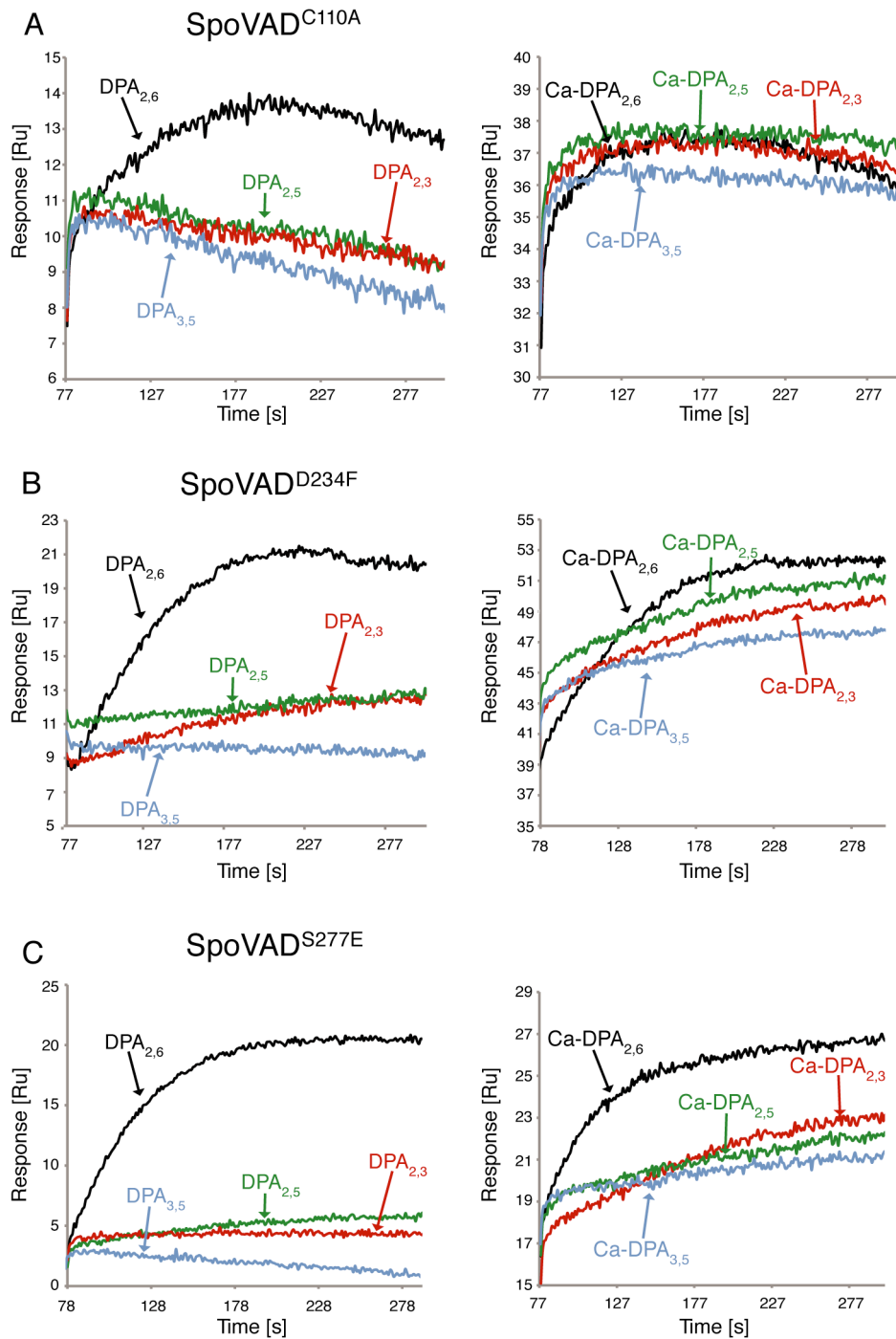


Figure S6. **Measurement of binding affinity of DPA/Ca-DPA to SpoVAD by SPR.** Multiple concentrations of DPA_{2,6}, Ca-DPA_{2,6} or Ca-DPA_{2,3} were used to determine their binding affinity (K_D) to the wild-type SpoVAD (A), SpoVAD^{C110A} (B), SpoVAD^{D234F} (C) and SpoVAD^{S277E} (D) (also see Table 1). The titration data were analyzed by nonlinear curve fitting to a steady-state

model using Biacore T100 analysis software (GE Healthcare, Piscataway, NJ) and OriginPro 7.5 (OriginLab Corporation, Northampton, MA). The binding of Ca-DPA_{2,3} to SpoVAD^{C110A} was too weak to be measured.

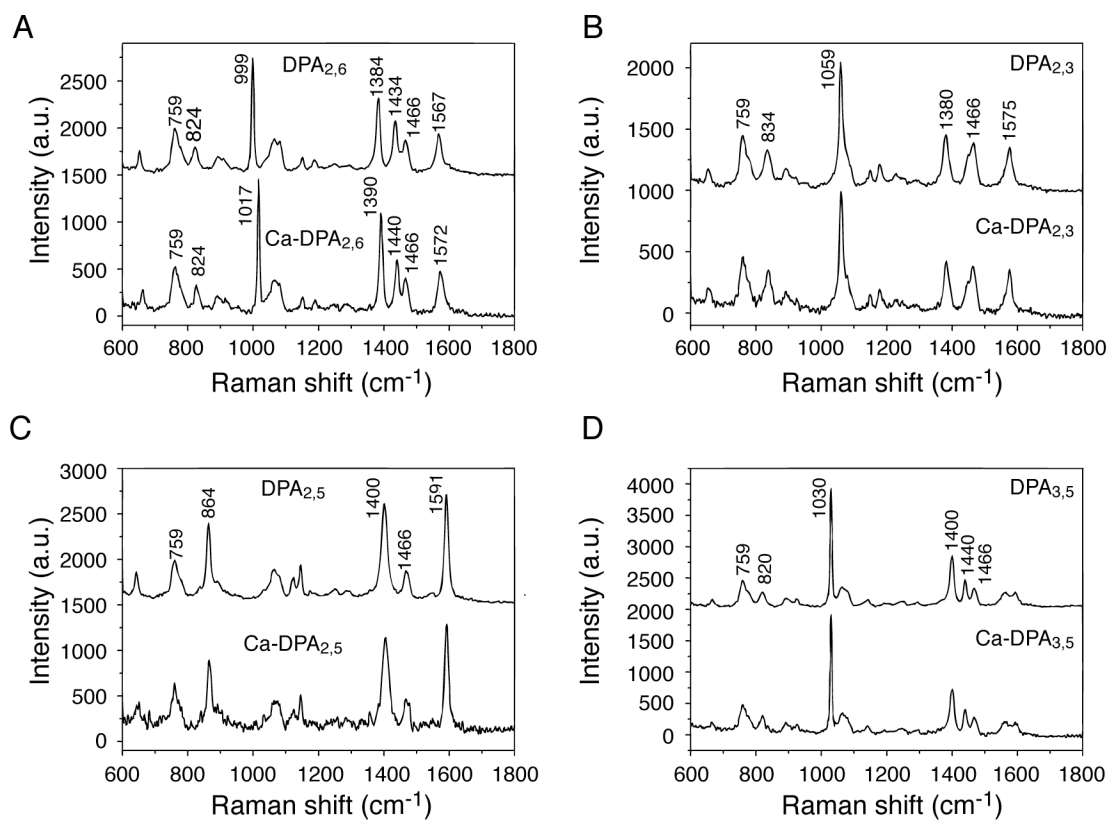


Figure S7. **Raman spectra of DPA_{2,6}, Ca-DPA_{2,6} and their isomers.** For each measurement, 60 mM of DPA_{2,6} or Ca-DPA_{2,6} (A), DPA_{2,3} or Ca-DPA_{2,3} (B), DPA_{2,5} or Ca-DPA_{2,5} (C) and DPA_{3,5} or Ca-DPA_{3,5} (D) were used. The laser power used was 30 mW and the integration time for Raman measurement was 20 s.

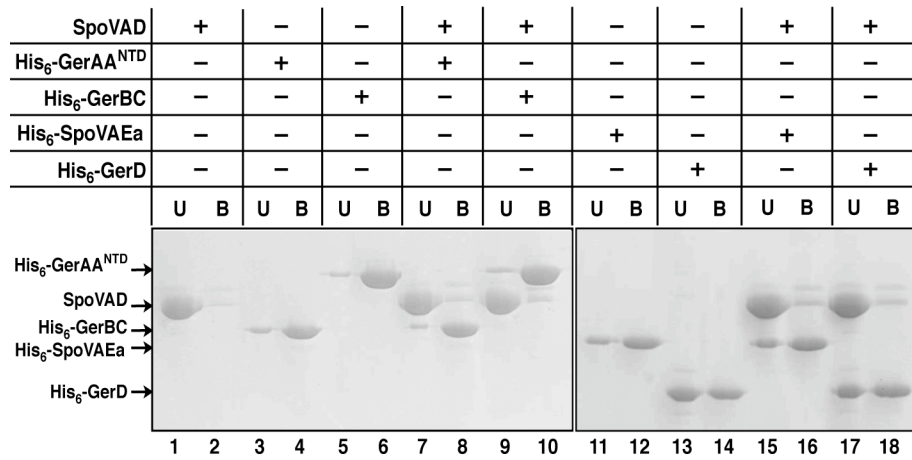


Figure S8. **SpoVAD does not interact with other germinant proteins including SpoVAEa, GerD, GerBC and GerAA^{NTD} *in vitro*.** A Ni²⁺-NTA pull-down assay was used to characterize the interaction of SpoVAD with the His₆-tagged SpoVAEa, GerD, GerBC and GerAA^{NTD}. Indicated proteins were incubated and reactions were precipitated with Ni²⁺-NTA resin. The unbound (U) and eluted bound (B) fractions were analyzed by SDS-PAGE and Coomassie staining.