

H1⁰-ChFP

MTENSTSAPAAKPKRAKASKKSTDHPKYSDMIVAAIQAEKNRAGSSRQSIQKYIKSHYKVGENADSQIKLSIKRLVTT GVLKQTKGVGASGSFRLAKGDEPKRSVAFKKTKKEVKKVATPKKAAKPKKAASKAPSKKPKATPVKKKKKPAATPKKA KKPKVVKVKPVKASKPKKAKTVKPKAKSSAKRGSKKKAMVSKGEEDNMAIIKEFMRFKVHMEGSVNGHEFEIEGEGEG RPYEGTQTAKLKVTKGGPLPFAWDILSPQFMYGSKAYVKHPADIPDYLKLSFPEGFKWERVMNFEDGGVVTVTQDSSL QDGEFIYKVKLRGTNFPSDGPVMQKKTMGWEASSERMYPEDGALKGEIKQRLKLKDGGHYDAEVKTTYKAKKPVQLPG AYNVNIKLDITSHNEDYTIVEQYERAEGRHSTGGMDELYKSGLRSRPRRQRTAFLNIKGMG

H1⁰-GFP

MTENSTSAPAAKPKRAKASKKSTDHPKYSDMIVAAIQAEKNRAGSSRQSIQKYIKSHYKVGENADSQIKLSILVTTGV LKQTKGVGASGSFRLAKGDEPKRSVAFKKTKKEVKKVATPKKAAKPKKAASKAPSKKPKATPVKKAKKKPAATPKKAK KPKVVKVKPVKASKPKKAKTVKPKAKSSAKRGSKKKAMVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYG KLTLKFICTTGKLPVPWPTLVTTLTYGVQCFSRYPDHMKQHDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDT LVNRIELKGIDFKEDGNILGHKLEYNYNSHNVYIMADKQKNGIKVNFKIRHNIEDGSVQLADHYQQNTPIGDGPVLLP DNHYLSTQSALSKDPNEKRDHMVLLEFVTAAGITLGMDELYKSGLRSRPRRQRTAFLNIKGMG

ChFP-H1⁰

MVSKGEEDNMAIIKEFMRFKVHMEGSVNGHEFEIEGEGEGRPYEGTQTAKLKVTKGGPLPFAWDILSPQFMYGSKAYV KHPADIPDYLKLSFPEGFKWERVMNFEDGGVVTVTQDSSLQDGEFIYKVKLRGTNFPSDGPVMQKKTMGWEASSERMY PEDGALKGEIKQRLKLKDGGHYDAEVKTTYKAKKPVQLPGAYNVNIKLDITSHNEDYTIVEQERAEGRHSTGGMDELY KSGLRSRPRRQRTAFLNIAMA**TENSTSAPAAKPKRAKASKKSTDHPKYSDMIVAAIQAEKNRAGSSRQSIQKYIKSHY** KVGENADSQIKLSIKRLVTTGVLKQTKGVGASGSFRLAKGDEPKRSVAFKKTKKEVKKVATPKKAAKPKKAASKAPSK KPKATPVKKAKKKPAATPKKAKKPKVVKVKPVKASKPKKAKTVKPKAKSSAKRGSKKK

GFP-H1⁰

MVSKGEELFTGVVPILVELDGDVNGHKFSVSGEGEGDATYGKLTLKFICTTGKLPVPWPTLVTTLTYGVQCFSRYPDH MKQHDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTLVNRIELKGIDFKEDGNILGHKLEYNYNSHNVYIMAD KQKNGIKVNFKIRHNIEDGSVQLADHYQQNTPIGDGPVLLPDNHYLSTQSALSKDPNEKRDHMVLLEFVTAAGITLGM DELYKSGLRSRPRRQRTAFLNIAMA**TENSTSAPAAKPKRAKASKKSTDHPKYSDMIVAAIQAEKNRAGSSRQSIQKYI** KSHYKVGENADSQIKLSIKRLVTTGVLKQTKGVGASGSFRLAKGDEPKRSVAFKKTKKEVKKVATPKKAAKPKKAASK APSKKPKATPVKKAKKKPAATPKKAKKPKVVKVKPVKASKPKKAKTVKPKAKSSAKRGSKKK

Supplemental Fig. S1. Constructs used in this study. (a) Representative maps of expression vectors. Transcription is driven by the mouse metallothionine I promoter. The IRES sequence was derived from pIRES2AcGFP (Clontech). (b) Sequences of expressed hybrid proteins. Histone coding sequences are in bold. GFP and ChFP are appropriately colored.



Supplemental Fig. S2. Independence of ChFP and GFP signals. (a) A construct expressing only H1⁰-ChFP was stably introduced into fibroblasts. Cells were imaged under the conditions used for dual-color FRAP. No signal was seen in the GFP channel. (b) A construct expressing only H1⁰-GFP was stably introduced into fibroblasts. Cells were imaged under the conditions used for dual-color FRAP. No signal was seen in the ChFP channel.



Supplemental Fig. S3. HPLC separation of H1 proteins. H1 proteins were extracted from a cell line expressing H1⁰-ChFP and H1⁰-GFP and separated by HPLC as previously described (16). The peak containing the tagged proteins is not seen in control cells (16, data not shown).









Supplemental Fig. S4. Comparison of t_{50} values for individual cells. Grouped pairs indicate the values obtained for the ChFP-tagged and GFP-tagged H1 proteins within a single cell. Data for averaged recovery curves are shown in Fig. 2



Supplemental Fig. S5. FRAP analysis of C-ChFP/CC0-GFP. FRAP analysis with cells coexpressing WT H1c-ChFP and CC0-GFP. Values for the half time of recovery (t_{50}) were determined as previously described (35) and represent means <u>+</u> S.D. of at-least 8 independent measurements from a pool of three stable cell lines. Error bars are omitted from the plots for clarity.



Supplemental Fig. S6. Quantitative analysis of the relative binding kinetics of N-tagged H1^o and H1c and effects of swapping their terminal domains. FRAP analyses with cells co-expressing (a) ChFP-H1^o and GFP-H1^o (b) ChFP-H1^o and GFP-H1c show faster recovery kinetics for H1c than H1^o. Simultaneous recovery curves of the C-terminal switch mutants (c) GFP-00C relative to ChFP-H1^o and (d) GFP-CC0 relative to ChFP-H1^o and of the N-terminal switch mutants (e) GFP-0CC relative to ChFP-H1^o and (f) GFP-C00 relative to ChFP-H1^o. Values for the half time of recovery (t_{50}) were determined as previously described (35) and represent means <u>+</u> S.D. of at-least 6 independent measurements from a pool of three stable cell lines. Error bars are omitted from the plots for clarity. Supplemental Table 1 provides the corresponding statistical analyses.