

Supplemental Data

Primate Genome Gain and Loss: A Bone Dysplasia, Muscular Dystrophy, Bone Cancer Syndrome Resulting from Mutated Retroviral-Derived MTAP Transcripts

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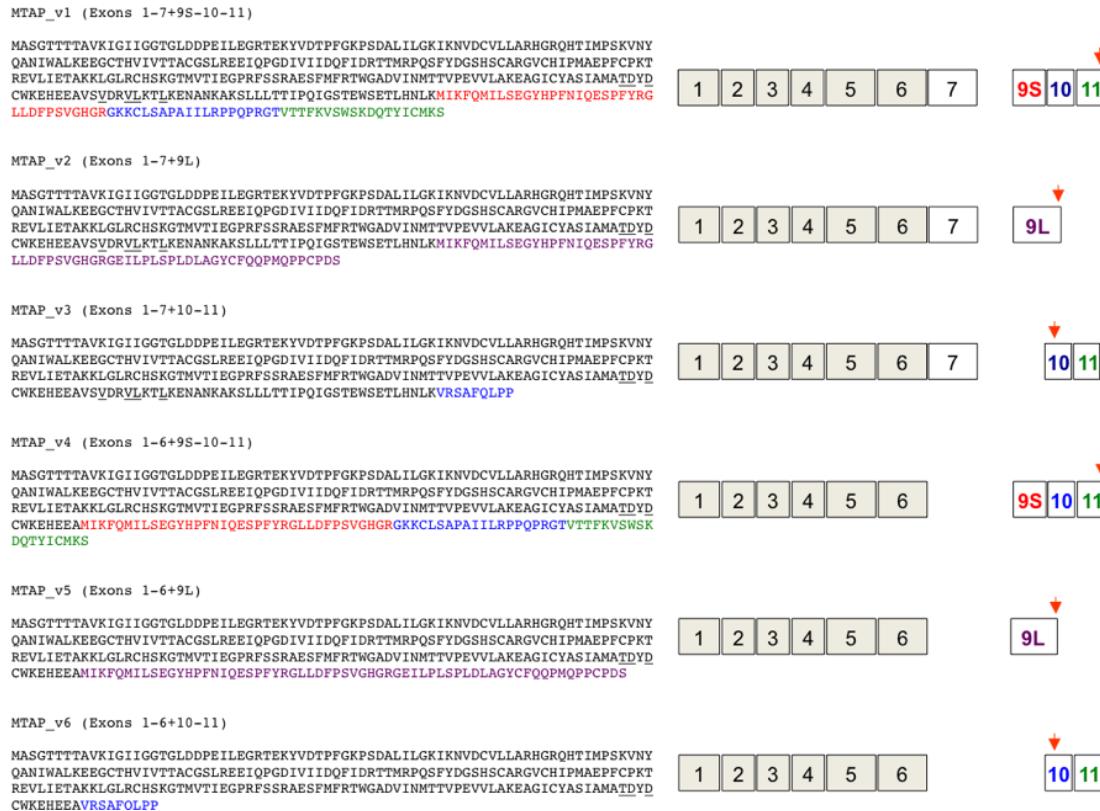


Figure S1. Amino Acid Sequence of MTAP Splice Variants

Exons 1-7 are shown in black typeface and MTA-coordinating amino acids are underlined. Exon 9S - red; 9L - purple, 10 - blue (two different reading frames are used); 11 - green. Arrows above exons depict the positions of translational stop codons.

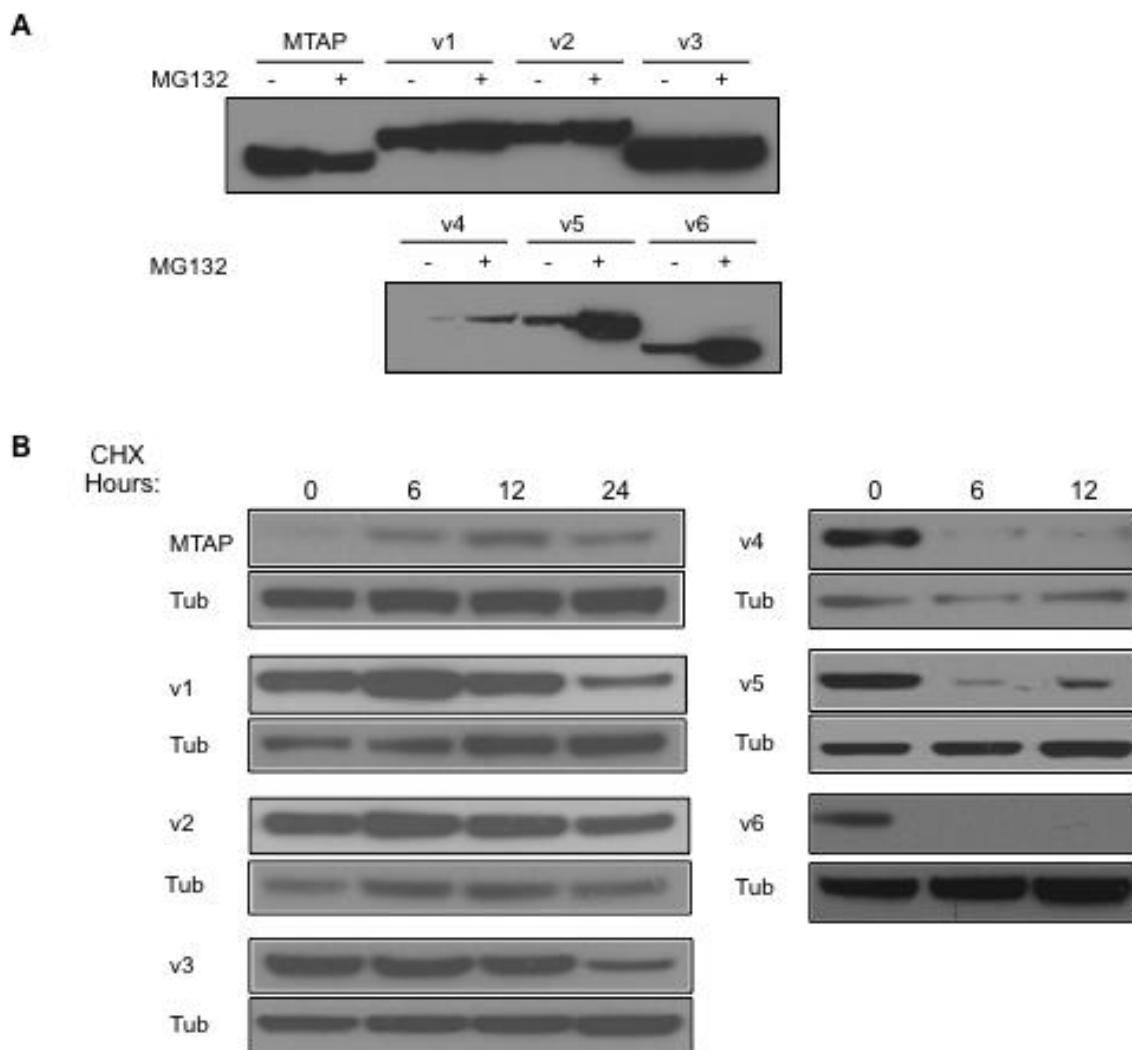


Figure S2. Protein Stability of MTAP Splice Variants

(A) Transiently transfected MCF7 cells expressing each MTAP isoform were treated with MG132 (5mM) for 4 hours.

(B) Transiently transfected MCF7 cells were exposed to cyclohexamide (10µM) for 6, 12 and 24 hours. Archetype MTAP: MTAP

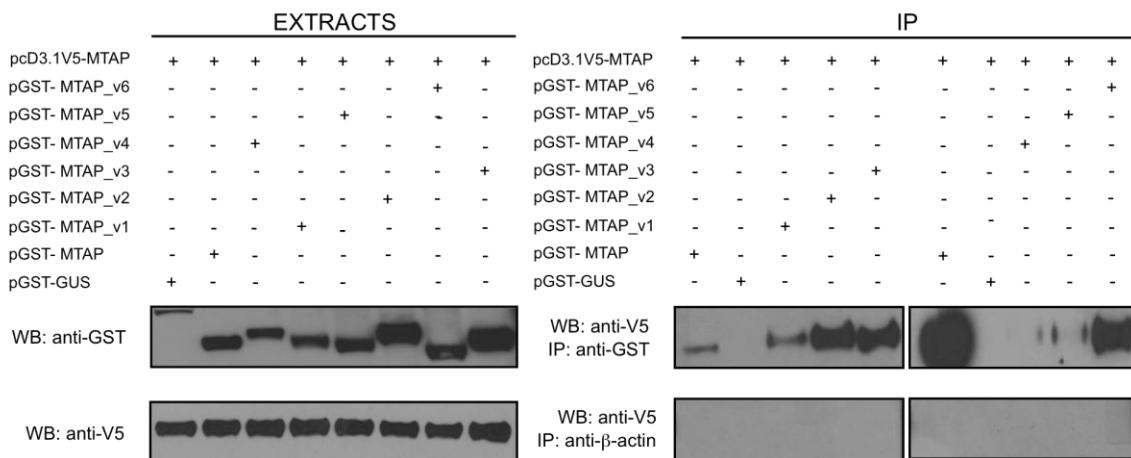


Figure S3. Coimmunoprecipitation Studies Showing Interaction between Archetype MTAP and All MTAP Splice Variants

Combinations of differentially-tagged (-V5 and -GST) isoforms of MTAP were co-expressed in PC3M cells and co-immunoprecipitated. Immunoprecipitation was performed with anti-GST and anti- β -actin as control. Lysates (left panel). Co-immunoprecipitates: MTAP, MTAP_v1, v2, v4, v5 and v6 (right panel). Archetype MTAP: MTAP

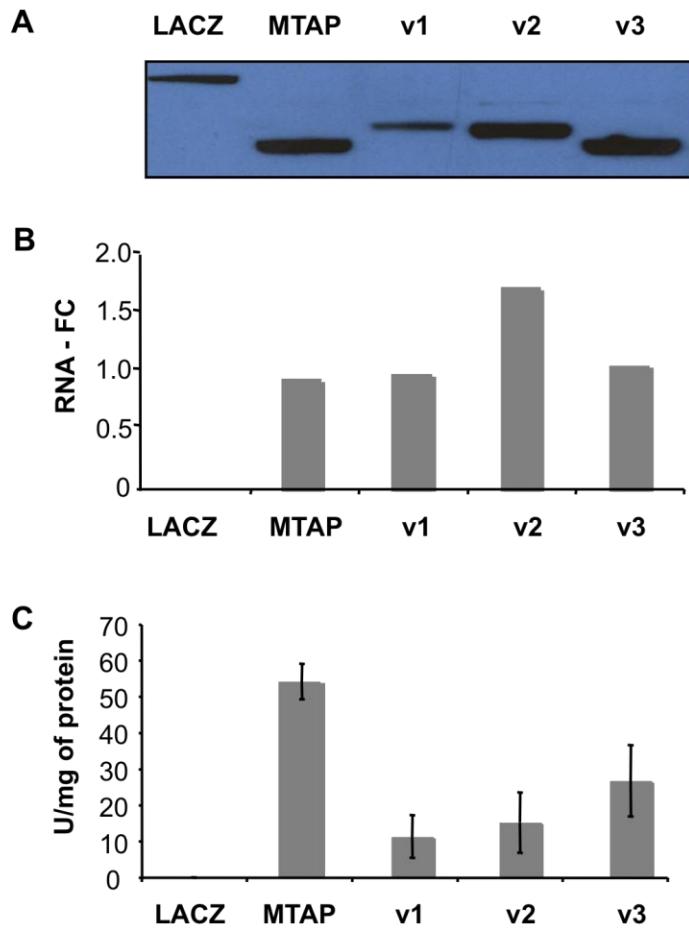


Figure S4. Determination of MTAP Activity on Cell Extracts from MCF7 Stably Expressing MTAP_v1, v2, and v3 V5-Tagged Fusion Proteins

(A) Western-blot showing protein expression.

(B) Quantitative real-time PCR results using primers that recognize all variants.

(C) Intracellular MTAP activity levels.

The error bars represent the averages of three independent experiments.

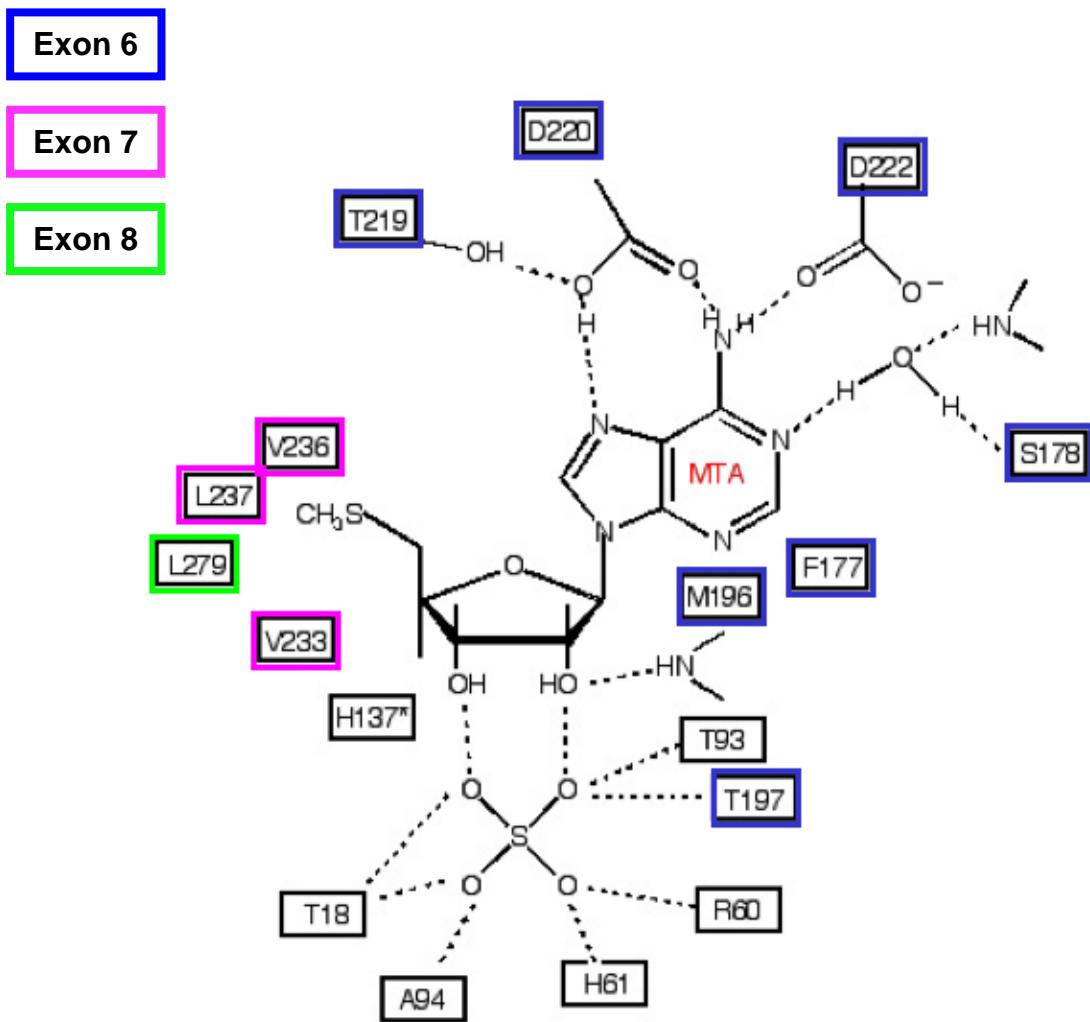


Figure S5. Amino Acids Coordinating to MTA

Note that exon 8 contributes only one amino acid in this structure.

Table S1. DNA and cDNA Amplifying Primers

MICROSATELLITE PRIMERS		5'	3'
D9SB3 Forward		CGGAATCAACCTAAATGTAG	
D9SB3 Reverse		CAGTTCTGTTCACATTGCTG	
AL624 Forward		CACCAAAGTGAGTGTGCCAT	
AL624 Reverse		GCTTTACCAGCTATCTGGGT	
AL882 Forward		CATCACAGGACATAGCAGC	
AL882 Reverse		TGAGGCTTGATGATGGTAG	
GENOMIC PRIMERS			
gExon 1 Forward		GAACGCGGGAATTCGATTGG	
gExon 1 Reverse		TGCAAGCGTACCCCTCTCCTC	
gExon 2 Forward		GGCTTGGCAGGAATGGAG	
gExon 2 Reverse		TCAAGCCAGCAAGGAGAAATG	
gExon 3 Forward		GAGTCCTGTTGTGGTTGAA	
gExon 3 Reverse		CAATGTAAGGGCAAGAGCA	
gExon 4 Forward		ACTCTAGGAGAAAACAGTTGG	
gExon 4 Reverse		ACCAGCTACAATAGCCTAAAG C	
gExon 5 Forward		AGATAAAGTTGACTCACCAAGC	
gExon 5 Reverse		CCAATTAAACCAATGCCAGAT G	
gExon 6 Forward		AGTTGTGCATGTGCTAGTATG	
gExon 6 Reverse		ACACCCATTGGCACCCAGA	
gExon 7 Forward		GCAGTGGATTAAAGTTCTAGTA	
gExon 7 Reverse		GGCATAAAATACATGAGGACCAA	
gExon 8 Forward		GTATGTTCCCTGCGTCCTCA	
gExon 8 Reverse		CATACTCTTGATAGGCAAGGG	
gExon 9 Forward		AAGTTCCAGATGATCCTCAG	
gExon 9 Reverse		CAGTCTCTGTAAGTGTTC	
CDNA PRIMERS			
Exon 1 Forward		GCACTGCTCACTCCCGCGCA	
Exon 5 Forward		CAGAGGAGTGTGCCATATTG	
Exon 6 Forward		GAGGGACCTCGTTTAGCTC	
Exon 8 Reverse		CATACTCTTGATAGGCAAGGG	
Exon 11 Reverse		ACTATAGGATGTCATGATT	
Exon 6F1		GACAGATTATGACTGCTGGAAGGAG	
REAL TIME PCR PRIMERS			
MTAP- Forward		CGGTGGACCGGGTCTAAAGAC	
MTAP-wt Reverse		ATGTCTTGGTAATAAACAGAAAAGTGG	

MTAP_v1 Forward	ACCCTCCATAACCTGAAGATG
MTAP_v1 Reverse	CTGAAAGGCACCTCTTACCTC
MTAP_v2 Forward	CTCCATAACCTGAAGATGATCA
MTAP_v2 Reverse	GACAGGGGCAGGATTCAC
MTAP_v3 Forward	ACCCTCCATAACCTGAAGGTA
MTAP_v3 Reverse	GGATGTCATGATTCATGCAAAT
MTAP_v4 Forward	AGCACGAGGAAGCAATGATC
MTAP_v4 Reverse	GCTGAAAGGCACCTCTTACCT
MTAP_v5 Forward	AGCACGAGGAAGCAATGATC
MTAP_v5 Reverse	CCAGCCAGGTCTAAGGGAG
MTAP_v6 Forward	AGGAGCACGAGGAAGCAGTA
MTAP_v6 Reverse	GGATGTCATGATTCATGCAAAT
MTAP-All Forward	ACCACCACCGCCGTGAAG
MTAP-All Reverse	CTTGCAAGGAGGACGCAATC
Actin Forward	CAATGACCCCTTCATTGACC
Actin Reverse	GATCTCGCTCCTGGAAGATG
HPRT Forward	TATTGTAATTGACCAGTCAACAG
HPRT Reverse	GGTCCTTTCACCAAGCAAG
PLASMID PRIMERS	
V5: for MTAP	ATGTCTTGGTAATAAACAGAAAACTGG
V5: for v1-v2	<u>TGATTTCATGCAAATATATGTTG</u>
V5: for v3-v4	TGGCGGGAGCTGAAAGGCACTTC
Intron 5-Sall Forward	<u>ACGCGTCGACGTCGGCTGGGAAACAGGATTATGTGA</u>
Intron 8-Nhel reverse	<u>CTAGCTAGCTAGATGTAGAGTTCCCAGCAC</u>
Intron 8-Spel Forward	<u>GGACTAGTCCAGCCTGATGATTATAACCAATG</u>
Intron 9-Xhol Reverse	<u>CTAGCTAGCTAGTCCATTATCATTTGCCTTTG</u>
Exon 9-Xhol Forward	<u>CCGCTCGAGCGGCAACAGAGCGAGAGT CTTC</u>
Intron 11-Sacl Reverse	<u>TCCCCGCGGGAACCAACCTATGAGATGCT</u>

Underlining in the PCR primers above indicates the introduced sequence including restriction sites.