

Supporting Information for

Enhanced Epimerization of Glycosylated Amino Acids During Solid Phase Glycopeptide Synthesis

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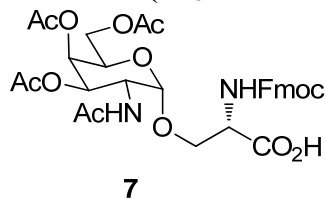
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General methods

Unless otherwise stated, reagents were purchased from commercial suppliers and used without purification. 2-Chlorotriyl chloride resin, Fmoc-Ser(Trt)-OH, Fmoc-D-Ser(Trt)-OH, Fmoc-OSu were purchased from AnaSpec, Inc. (San Jose, CA). Fmoc-6-aminohexanoic acid (Fmoc-Hex-OH), Fmoc-Gly-OH, and Fmoc-Pro-OPfp were purchased from Novabiochem (Darmstadt, Germany). 2-(7-Aza-1H-benzotriazol-1-yl)-*N,N,N',N'*-tetramethyl uronium hexafluorophosphate (HATU), 1-hydroxy-7-azabenzotriazole (HOAt), 2-(1H-benzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium hexafluorophosphate (HBTU), *N*-hydroxybenzotriazole (HOBt), *N,N'*-Dicyclohexylcarbodiimide (DCC), benzotriazol-1-yl-oxy-tris-pyrrolidino-phosphonium hexafluorophosphate (PyBOP), *N,N*-dimethylformamide (DMF), *N*-methyl-2-pyrrolidone (NMP), *N,N*-diisopropylethylamine (DIEA), *N*-methylmorpholine (NMM), 2,4,6-trimethylpyridine (TMP) were purchased from Sigma (St. Louis, MO). All RP-HPLC purifications were carried using a preparative C-18 reversed phase column (VYDAC[®] 218 TP Protein & Peptide C18 22 mm × 250 mm) on a Waters PropLC 4000 system (HPLC) with a waters 2996 photodiode array detector at room temperature. All ¹H and ¹³C NMR spectra were recorded on a 400 MHz or 500 MHz Varian spectrometer. The mass spectra were recorded on Shimadzu Axima-CFR MALDI-TOF or on Agilent LC/MSD SL mass spectrometers. Analysis of yields and epimerization were recorded on an Agilent 1200 Series HPLC using Waters XTerra[®] RP 18 (5μm 4.6 mm × 250 mm) reversed phase column.

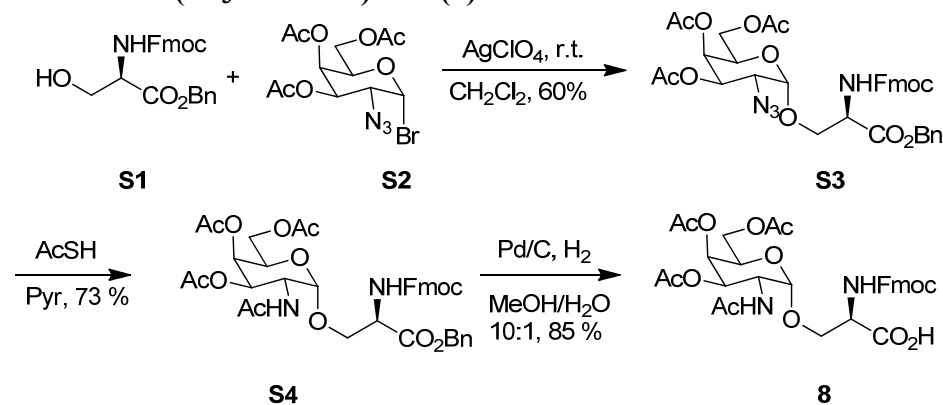
Preparation of Fmoc-protected glyco-amino acids

Fmoc-Ser(Ac₃GalNAc α)-OH (7)



N-(9H-fluoren-9-ylmethoxycarbonyl)-*O*-(2-acetyl-2-deoxy-3,4,6-tri-*O*- α -D-galactopyranosyl)-L-serine (**7**) was prepared according to the previously reported method.¹ ¹H-NMR (400 MHz, *N,N*-dimethylformamide-*d*₇) δ 7.93 (d, *J* = 7.5 Hz, 2H, H-Fmoc), 7.88 (d, *J* = 8.8 Hz, 1H, NH-S), 7.83 (d, *J* = 8.7 Hz, 1H, NH-GalNAc), 7.74 (dd, *J* = 7.3, 2.3 Hz, 2H, H-Fmoc), 7.44 (t, *J* = 7.5 Hz, 2H, H-Fmoc), 7.34 (t, *J* = 7.5 Hz, 2H, H-Fmoc), 5.41 (d, *J* = 2.3 Hz, 1H, H4'), 5.17 (dd, *J* = 11.7, 3.2 Hz, 1H, H3'), 5.01 (d, *J* = 3.5 Hz, 1H, H1'), 4.47-4.50 (m, 1H, S ^{α}), 4.33-4.40 (m, 4H, H2' {4.38}, H5' {4.33}, CH₂-Fmoc {4.33}), 4.29 (m, 1H, H-Fmoc), 4.08-4.16 (m, 2H, H6'), 3.97-4.04 (m, 2H, S ^{β}), 2.14 (s, 3H, CH₃-Ac), 1.99 (s, 3H, CH₃-Ac), 1.93 (s, 3H, CH₃-Ac), 1.89 (s, 3H, CH₃-Ac); ¹³C-NMR (100 MHz, *N,N*-dimethylformamide-*d*₇) δ 176.1, 174.6, 174.4, 174.2, 160.9, 148.6, 145.5, 132.1, 131.5, 129.7, 124.5, 102.9, 72.8, 72.4, 71.8, 71.2, 70.8, 66.2, 59.0, 51.8, 51.4, 26.5, 24.3, 24.25, 24.2. HRMS calcd for C₃₂H₃₆N₂O₁₃ [M + Na]⁺ 679.2110, measured 679.2115.

Fmoc-D-Ser(Ac₃GalNAc α)-OH (8)



The unnatural glyco-amino acid, Fmoc-D-Ser(Ac₃GalNAc α)-OH **8**, was synthesized with similar procedures reported for **7**. Briefly, the bromide donor **S2** (2.64 g, 6.70 mmol), *N*-Fmoc-D-serine

benzyl ester **S1** (2.33 g, 5.58 mmol), and 2 g 4 Å molecular sieve were added to a flask under argon. The mixture was dissolved in 20 mL CH₂Cl₂, then silver perchlorate (2.31 g, 11.2 mmol) was added and the reaction mixture was stirred at room temperature for 2 h. The mixture was filtered, concentrated, and the residue was purified by flash chromatography on silica gel (120g) with EtOAc/Hexane (1/3) to give compound **S3** (2.44 g, 60 %, α:β=5:1) as a white solid. Reductive acetylation using thiolacetic acid/pyridine (2/1, v/v, 3 mL) produced the fully protected Fmoc-D-Ser(Ac₃GalNAcα)-OBn **S4** in a 73% yield. Hydrogenation of **S4** with 10% Pd/C in MeOH/H₂O/Formic acid (10/1/1, v/v/v, 12 mL) followed by RP- HPLC purification using a 0-48% acetonitrile/H₂O (0.1% TFA) gradient over 45 min to provide compound **8** in 71% yield as white solid.

N-(9H-fluoren-9-ylmethoxycarbonyl)-D-serine benzyl ester (S1). ¹H-NMR (400 MHz, CDCl₃) δ 7.73 (d, *J* = 7.6 Hz, 2H), 7.56 (d, *J* = 7.4 Hz, 2H), 7.37 (t, *J* = 7.5 Hz, 2H), 7.26-7.39 (m, 7H), 5.69 (d, *J* = 7.0 Hz, 1H), 5.20 (Br, 2H), 4.35-4.47 (m, 3H), 4.18 (t, *J* = 6.7 Hz, 1H), 3.99 (d, *J* = 8.5 Hz, 1H), 3.91 (d, *J* = 10.3 Hz, 1H); ¹³C-NMR (100 MHz, CDCl₃) δ 170.1, 155.5, 143.6, 141.3, 141.2, 135.0, 128.6, 128.5, 128.2, 127.7, 127.1, 127.0, 125.0, 120.0, 119.9, 67.5, 67.2, 63.3, 56.1, 47.1. ESI MS calcd for C₂₅H₂₃NO₅ [M + Na]⁺ 440.16, measured 440.1.

N-(9H-fluoren-9-ylmethoxycarbonyl)-O-(2-azido-2-deoxy-3,4,6-tri-O-α-D-galactopyranosyl)-D-serine benzyl ester (S3). ¹H-NMR (400 MHz, CDCl₃) δ 7.73 (d, *J* = 7.5 Hz, 2H), 7.57 (d, *J* = 7.4 Hz, 2H), 7.26-7.39 (m, 9H), 5.84 (d, *J* = 8.4 Hz, 1H), 5.35 (d, *J* = 12.0 Hz, 1H), 5.19 (d, *J* = 3.0 Hz, 1H), 5.10 (d, *J* = 11.9 Hz, 1H), 4.91 (d, *J* = 3.5 Hz, 1H), 4.61-4.64 (m, 1 H), 4.33-4.42 (m, 2H), 4.17-4.22 (m, 2H), 3.79-3.90 (m, 2H), 3.69 (dd, *J* = 10.1, 3.1 Hz, 1H), 3.63 (t, *J* = 6.1 Hz, 1H), 3.55 (dd, *J* = 11.2, 3.6 Hz, 1H), 2.09 (s, 3H), 2.05 (s, 3H), 1.97 (s, 3H); ¹³C-NMR (100 MHz, CDCl₃) δ 170.2, 169.2, 155.9, 143.6, 141.3, 135.1, 128.7, 125.1, 119.9, 169.5, 169.9, 98.7, 67.5, 67.0, 61.9, 54.1, 47.1, 20.61, 20.6, 20.5. MALDI MS calcd for C₃₇H₃₈N₄O₁₂ [M + Na]⁺ 753.2378, measured 753.2382.

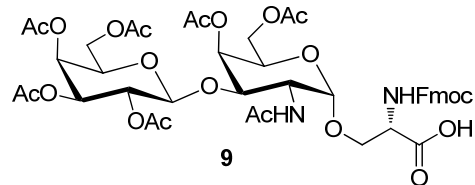
N-(9H-fluoren-9-ylmethoxycarbonyl)-O-(2-acetyl-2-deoxy-3,4,6-tri-O-α-D-galactopyranosyl)-D-serine benzyl ester (S4). ¹H-NMR (400 MHz, CDCl₃) δ 7.72 (d, *J* = 7.5 Hz, 2H), 7.56 (d, *J* = 7.2 Hz, 2H), 7.25-7.38 (m, 9H), 6.02 (d, *J* = 9.3 Hz, 1H), 5.77 (d, *J* = 8.0

Hz, 1H), 5.29 (d, $J = 11.9$ Hz, 1H), 5.10-5.14 (m, 2 H), 4.81-4.88 (m, 2H), 4.58-4.61 (m, 1H), 4.50-4.52 (m, 1H), 4.43 (d, $J = 6.5$ Hz, 2H), 4.18 (t, $J = 6.6$ Hz, 1H), 3.97 (dd, $J = 10.3, 4.2$ Hz, 1H), 3.90 (d, $J = 6.3$ Hz, 2H), 3.71-3.74 (m, 2H), 2.10 (s, 3H), 1.95 (s, 3H), 1.94 (s, 3H), 1.88 (s, 3H); ^{13}C -NMR (100 MHz, CDCl_3) δ 170.7, 170.32, 170.3, 170.2, 169.7, 156.0, 143.6, 141.3, 134.8, 128.9, 128.8, 128.7, 127.7, 127.1, 124.8, 120.0, 98.5, 68.7, 68.1, 67.9, 67.2, 67.1, 66.9, 62.1, 54.4, 47.5, 47.0, 23.0, 20.7, 20.66, 20.6. MALDI MS calcd for $\text{C}_{39}\text{H}_{42}\text{N}_2\text{O}_{13}$ $[\text{M} + \text{Na}]^+$ 769.2579, measured 769.2585.

***N*-(9H-fluoren-9-ylmethoxycarbonyl)-*O*-(2-acetyl-2-deoxy-3,4,6-tri-*O*- α -D-**

galactopyranosyl)-D-serine (8). ^1H -NMR (400 MHz, *N,N*-dimethylformamide- d_7) δ 7.94 (d, $J = 7.6$ Hz, 2H, H-Fmoc), 7.45 (d, $J = 7.5$ Hz, 2H, H-Fmoc), 7.75-7.79 (m, 2H, H-Fmoc), 7.34 (t, $J = 7.4$ Hz, 2H, H-Fmoc), 5.37 (d, $J = 3.1$ Hz, 1H, H4'), 5.05 (dd, $J = 11.6, 3.2$ Hz, 1H, H3'), 4.99 (d, $J = 3.5$ Hz, 1H, H1'), 4.57-4.61 (m, 1H, S $^{\alpha}$), 4.48-4.54 (m, 1H, H2'), 4.40 (dd, $J = 9.9, 7.0$ Hz, 1H, H6a'), 4.19-4.34 (m, 5H, H6' {4.32}, S $^{\beta}$ {4.26}, CH $_2$ -Fmoc {4.20}), 4.01-4.07 (m, 1H, CH-Fmoc), 3.74 (dd, $J = 9.5, 3.3$ Hz, 1H, H5'), 2.15 (s, 3H, CH $_3$ -Ac), 2.02 (s, 3H, CH $_3$ -Ac), 1.95 (s, 3H, CH $_3$ -Ac), 1.93 (s, 3H, CH $_3$ -Ac). ^{13}C -NMR (100 MHz, *N,N*-dimethylformamide- d_7) δ 176.0, 174.6, 174.4, 174.36, 174.1, 160.9, 148.7, 148.5, 145.5, 145.52, 132.1, 131.5, 131.4, 124.5, 124.4, 102.2, 72.7, 72.6, 71.8, 71.4, 70.8, 66.1, 58.4, 51.4, 26.5, 24.3, 24.2. HRMS calcd for $\text{C}_{32}\text{H}_{36}\text{N}_2\text{O}_{13}$ $[\text{M} + \text{Na}]^+$ 679.2115, found 679.2116.

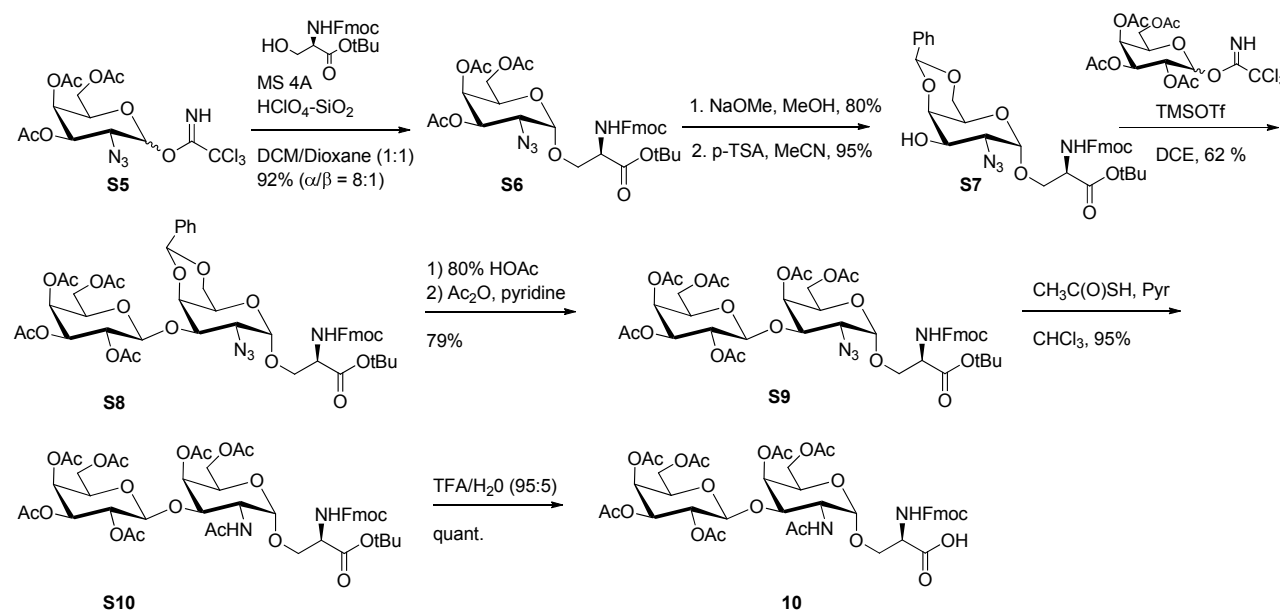
Fmoc-Ser(Ac $_4$ Gal β 1,3Ac $_2$ GalNAc α)-OH (9)



N-(9H-fluoren-9-ylmethoxycarbonyl)-*O*-[*O*-(2',3',4',6'-tetra-*O*-acetyl- β -D-galactopyranosyl)-(1,3)-2-acetamido-4,6-di-*O*-acetoxy-2-deoxy- α -D-galactopyranosyl]-L-serine (9) was prepared according to a reported method.¹ ^1H NMR (400 MHz, *N,N*-dimethylformamide- d_7) δ 7.95 (d, $J = 7.5$ Hz, 2H), 7.66-7.79 (m, 4H), 7.47 (t, $J = 7.5$ Hz, 2H), 7.34-7.38 (m, 2H), 5.45 (d, $J = 3.2$ Hz, 1H), 5.38 (d, $J = 3.6$ Hz, 1H), 5.11-5.13 (m, 1H), 5.01-5.03 (m, 2H), 4.89 (d, $J = 3.6$ Hz, 1H), 3.95-4.49 (m, 15H), 2.14 (s, 3H), 2.10 (s, 3H), 2.08 (s, 3H), 2.02-2.03 (m, 6H), 1.95 (s,

6H); ^{13}C NMR (100 MHz, *N,N*-dimethylformamide- d_7) δ 171.9, 170.3, 170.31, 170.1, 168.8, 169.7, 169.5, 156.7, 144.4, 144.3, 127.9, 127.3, 125.5, 125.47, 120.3, 101.1, 98.9, 73.5, 70.9, 70.4, 70.1, 68.9, 67.5, 66.7, 63.0, 61.1, 54.9, 49.0, 47.3, 22.6, 20.2, 20.15, 20.1, 20.0, 19.9. HRMS calcd for $\text{C}_{44}\text{H}_{52}\text{N}_2\text{O}_{21}$ $[\text{M} + \text{H}]^+$ 945.3141, measured 945.3136.

Fmoc-D-Ser(Ac₄Gal β 1,3Ac₂GalNAc α)-OH (10)



S6: Trichloroacetimidate donor **S5** (3.50 g, 7.36 mmol) and *D*-serine *tert*-butyl ester (2.17 g, 5.66 mmol) were dissolved in anhydrous CH₂Cl₂ / 1,4-dioxane (1:1, 70 mL) at room temperature. Activated molecular sieves (4Å, 3.50 g) were added and the mixture was stirred for 0.5 h at room temperature. The reaction mixture was cooled to 0°C and HClO₄-SiO₂ (0.4 mmol/g, 1.32 g) was added in one portion with rapid stirring. The reaction was stirred for 1 h at 0°C and then neutralized by addition of a diluted DIEA solution (92 μL , 0.53 mmol in 10 mL of CH₂Cl₂). The solids were filtered off and the solvent was removed under reduced pressure. The crude residue was purified by silica-gel flash chromatography (EtOAc in hexanes 20 – 45 %) to yield the alpha-glycosylated serine **S6** (4.20 g, 82 %), together with the beta product (512 mg, 10 %) as colorless foams. ^1H NMR (400 MHz, CDCl₃) δ 7.74 (d, $J = 7.5$ Hz, 2H), 7.58 (d, $J = 7.4$ Hz, 2H), 7.40 – 7.25 (m, 4H), 5.74 (d, $J = 8.7$ Hz, 1H), 5.40 (d, $J = 3.0$ Hz, 1H), 5.26 (dd, $J = 11.1$, 3.0 Hz, 1H), 4.98 (d, $J = 3.3$ Hz, 1H), 4.47 (ddd, $J = 8.7$, 2.9, 2.9 Hz, 1H), 4.37 (d, $J = 7.2$ Hz, 2H), 4.24 – 4.02 (m, 5H), 3.68 (dd, $J = 9.8$, 2.9 Hz, 1H), 3.58 (dd, $J = 11.1$, 3.4 Hz, 1H), 2.12 (s,

3H), 2.04 (s, 3H), 2.02 (s, 3H), 1.49 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 170.3, 170.0, 169.66, 168.2, 155.8, 143.9, 143.7, 141.3, 141.26, 127.7, 127.0, 125.1, 119.9, 98.7, 83.2, 69.0, 67.7, 67.4, 67.1, 61.6, 57.4, 54.5, 47.1, 27.9, 20.6, 20.57. ESI-MS calcd for $\text{C}_{34}\text{H}_{40}\text{N}_4\text{O}_{20}$ $[\text{M} + \text{Na}]^+$ 719.2, measured: 719.1.

S7: Deacetylation of **S6** using Zemplén conditions (sodium methoxide in methanol) produced the triol product as a colorless foam in 80 % yield. Triol product (1.00 g, 1.75 mmol) and α,α -dimethoxytoluene (0.502 mL, 3.51 mmol) were dissolved in anhydrous MeCN (20 mL) and a catalytic amount of *p*-toluenesulfonic acid (*p*-TSA, 80 mg, 0.421 mmol) was added in one portion. The reaction was stirred for 1 h at room temperature and then neutralized by slow addition of a diluted diisopropylethylamine solution (DIEA, 74 μL , 0.421 mmol in 10 mL of MeCN). The solvent was removed *in vacuo* and the residue was purified by flash chromatography on silica gel (EtOAc in hexanes, 20 – 45 %) to yield the benzylidene protected product **S7** (1.10 g, 95 %) as a colorless foam. ^1H NMR (400 MHz, CDCl_3) δ 7.78 (d, $J = 7.6$ Hz, 2H), 7.59 (dd, $J = 7.2, 2.3$ Hz, 2H), 7.47 – 7.44 (m, 2H), 7.40–7.35 (m, 5H), 7.31–7.26 (m, 2H), 5.77 (d, $J = 9.0$ Hz, 1H), 5.55 (s, 1H), 4.99 (d, $J = 3.3$ Hz, 1H), 4.46 (ddd, $J = 8.8, 2.6, 2.6$ Hz, 1H), 4.40–4.32 (m, 2H), 4.30–4.18 (m, 4H), 4.08 (bd, $J = 10.5$ Hz, 1H), 4.05 (dd, $J = 12.6, 1.6$ Hz, 1H), 3.70 (s, 1H), 3.66 (dd, $J = 9.7, 2.8$ Hz, 1H), 3.53 (dd, $J = 10.5, 3.3$ Hz, 1H), 2.40 (bs, 1H), 1.45 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 168.7, 155.9, 143.9, 143.7, 141.3, 141.25, 137.1, 129.4, 128.3, 127.7, 127.0, 126.2, 125.2, 119.9, 101.3, 99.5, 82.7, 75.3, 69.2, 69.0, 67.2, 63.1, 60.7, 54.6, 47.1, 28.0. ESI-MS calcd for $\text{C}_{35}\text{H}_{38}\text{N}_4\text{O}_9$ $[\text{M} + \text{Na}]^+$ 681.2, measured: 681.1.

S8: A mixture of **S7** (1.10 g, 1.67 mmol) and galactose donor (1.32 g, 2.67 mmol) were coevaporated with anhydrous toluene (2 \times 5.0 mL) and then dissolved in anhydrous dichloroethane (DCE, 20.0 mL). The reaction mixture was cooled to 10°C and TMSOTf (48 μL , 0.267 mmol) was added. After stirring for 45 min at 10°C TLC analysis (EtOAc / hexanes (4:6)) showed complete consumption of starting material and the reaction was neutralized by slow addition of diluted DIEA (47 μL , 0.267 mmol in 10 mL of CH_2Cl_2). The solvent was evaporated off and the crude was purified by flash chromatography on silica gel (EtOAc in hexanes, 30–50%) to yield the beta glycosylated product **S8** (1.05 g, 62%) as a colorless foam. ^1H NMR (400 MHz, CDCl_3) δ 7.74 (d, $J = 7.5$ Hz, 2H), 7.59 (dd, $J = 7.2, 2.1$ Hz, 2H), 7.50 (dd, $J = 7.7, 1.8$ Hz,

2H), 7.40–7.25 (m, 7H), 5.74 (d, $J = 8.6$ Hz, 1H), 5.51 (s, 1H), 5.38 (d, $J = 2.6$ Hz, 1H), 5.26 (dd, $J = 10.0, 8.1$ Hz, 1H), 5.03–4.98 (m, 2H), 4.70 (d, $J = 7.8$ Hz, 1H), 4.50–4.45 (m, 1H), 4.42–4.35 (m, 2H), 4.32 (d, $J = 2.6$ Hz, 1H), 4.26–4.20 (m, 2H), 4.16 (dd, $J = 9.7, 2.4$ Hz, 1H), 4.13–4.08 (m, 2H), 3.98–4.02 (m, 2H), 3.87 (dd, $J = 6.6, 6.6$ Hz, 1H), 3.78 (dd, $J = 10.8, 3.3$ Hz, 1H), 3.68 (dd, $J = 9.7, 2.6$ Hz, 1H), 3.63 (s, 1H), 2.14 (s, 3H), 2.04 (s, 3H), 2.00 (s, 3H), 1.96 (s, 3H), 1.49 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 170.2, 170.17, 170.1, 169.4, 168.8, 155.9, 143.8, 143.7, 141.3, 141.27, 137.5, 128.9, 128.1, 127.7, 127.0, 126.1, 125.1, 120.0, 102.6, 100.6, 99.4, 82.6, 76.7, 75.7, 70.9, 70.86, 69.0, 68.6, 67.2, 66.8, 63.4, 61.2, 58.7, 54.6, 47.1, 28.0, 20.7, 20.5. HRMS calcd for $\text{C}_{49}\text{H}_{56}\text{N}_4\text{O}_{18} [\text{M} + \text{Na}]^+$ 1011.3487, measured 1011.3496.

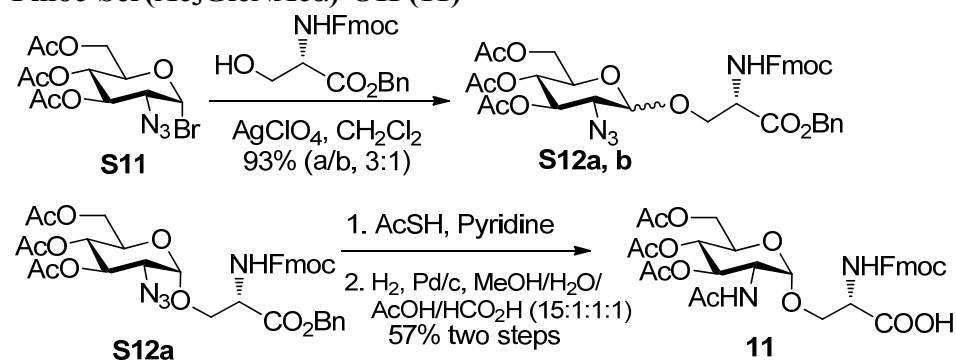
S9: Benzylidene protected disaccharide **S8** (1.00 g, 1.01 mmol) was dissolved in 80% aqueous acetic acid solution (15 mL) and stirred at 80°C for 2h. After cooling to room temperature, the solvent was evaporated off and the residue was evaporated with toluene (3×10 mL). The residue was dissolved in a mixture of anhydrous pyridine (5.0 mL) and acetic anhydride (5.0 mL). The reaction was stirred overnight at room temperature. The solvent was evaporated off and the residue was dissolved in CH_2Cl_2 (100 mL). The organic phase was extracted with water (50 mL) and 1 M hydrochloric acid (50 mL). The combined aqueous washings were back-extracted with CH_2Cl_2 (50 mL). The combined organic extracts were dried with MgSO_4 and concentrated. The crude was purified by flash chromatography on silica gel (EtOAc in hexanes, 20 – 45 %) to yield the peracetylated disaccharide **S9** (786 mg, 79%) as a colorless foam. ^1H NMR (400 MHz, CDCl_3) δ 7.74 (d, $J = 7.5$ Hz, 2H), 7.58 (dd, $J = 7.3, 3.0$ Hz, 2H), 7.38 (vt, $J = 7.5$ Hz, 2H), 7.29 (tt, $J = 7.5, 1.0$ Hz, 2H), 5.74 (d, $J = 8.7$ Hz, 1H), 5.40 (d, $J = 3.1$ Hz, 1H), 5.34 (d, $J = 3.2$ Hz, 1H), 5.14 (dd, $J = 10.5, 7.8$ Hz, 1H), 4.96 (dd, $J = 10.5, 3.2$ Hz, 1H), 4.93 (d, $J = 3.3$ Hz, 1H), 4.61 (d, $J = 7.8$ Hz, 1H), 4.45–4.50 (m, 1H), 4.41 (dd, $J = 10.6, 7.7$ Hz, 1H), 4.36 (dd, $J = 10.6, 7.3$ Hz, 1H), 4.21 (dd, $J = 7.2, 7.2$ Hz, 1H), 4.00–4.13 (m, 5H), 3.94–4.00 (m, 2H), 3.85 (dd, $J = 6.7, 6.7$ Hz, 1H), 3.66 (dd, $J = 9.8, 2.9$ Hz, 1H), 3.60 (dd, $J = 10.7, 3.5$ Hz, 1H), 2.12 (s, 3H), 2.09 (s, 3H), 2.05 (s, 3H), 2.03 (s, 3H), 2.02 (s, 3H), 1.95 (s, 3H), 1.48 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 170.3, 170.29, 170.2, 170.0, 169.6, 169.5, 168.46, 155.8, 143.8, 143.7, 141.3, 141.3, 127.7, 127.0, 125.1, 120.0, 101.4, 98.5, 82.8, 74.3, 70.8, 70.7, 69.1, 68.8, 68.7, 67.8, 67.1, 66.7, 62.5, 60.9, 59.5, 54.7, 47.1, 28.0, 20.7, 20.66, 20.6, 20.5. ESI-MS calcd for $\text{C}_{46}\text{H}_{56}\text{N}_4\text{O}_{20} [\text{M} + \text{Na}]^+$ 1007.3, measured: 1007.1.

S10: To a solution of compound **S9** (780 mg, 0.792 mmol) in chloroform (7.0 mL) were added pyridine (7.0 mL) and thioacetic acid (7.0 mL). The reaction was stirred overnight at room temperature. The solvent was evaporated off and the residue was dissolved in EtOAc (100 mL). The organic phase was extracted sequentially with 1 M aqueous HCl (50 mL), saturated aqueous NaHCO₃ (50 mL) and water (50 mL). The individual aqueous washings were back-extracted with EtOAc (25 mL, respectively). The combined organic extracts were dried with MgSO₄ and concentrated. The crude was purified by flash chromatography on silica gel (EtOAc in hexanes 70 – 90 %) to yield the acetyl amine **S10** (753 mg, 95%) as a hygroscopic solid. ¹H NMR (400 MHz, CDCl₃) δ 7.76 (d, *J* = 7.5 Hz, 2H), 7.57 (dd, *J* = 9.2, 8.2 Hz, 2H), 7.39 (dt, *J* = 7.5, 3.5 Hz, 2H); 7.31 (dt, *J* = 7.4, 1.2 Hz, 2H), 6.31 (d, *J* = 8.8 Hz, 1H), 5.64 (d, *J* = 8.0 Hz, 1H), 5.24 (d, *J* = 2.7 Hz, 1H), 5.22 (d, *J* = 2.8 Hz, 1H), 4.99 (dd, *J* = 10.2, 8.0 Hz, 1H), 4.87 (d, *J* = 3.0 Hz, 1H), 4.72 (dd, *J* = 10.5, 3.2 Hz, 1H), 4.50 – 4.35 (m, 4H), 4.21 (dd, *J* = 6.9, 6.9 Hz, 1H), 4.20 (d, *J* = 7.8 Hz, 1H), 4.11 (dd, *J* = 10.9, 4.3 Hz, 1H), 4.03 – 3.90 (m, 4H), 3.84 (dd, *J* = 10.3, 6.1 Hz, 1H), 3.71 (dd, *J* = 10.3, 2.5 Hz, 1H), 3.63 (dd, *J* = 11.0, 3.0 Hz, 1H), 3.61 – 3.57 (m, 1H), 2.09 (s, 3H), 2.07 (s, 3H), 2.02 (s, 3H), 1.98 (s, 3H), 1.96 (s, 3H), 1.91 (s, 3H), 1.47 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 170.3, 170.28, 170.1, 170.12, 170.0, 169.5, 168.8, 156.1, 143.6, 141.3, 141.26, 127.9, 127.2, 127.15, 124.9, 124.8, 120.2, 100.7, 98.9, 83.4, 73.6, 70.7, 70.5, 69.5, 68.5, 68.4, 67.7, 67.2, 66.6, 62.8, 60.7, 54.8, 48.6, 47.1, 27.9, 23.1, 20.7, 20.69, 20.6, 20.5. HRMS calcd for C₄₈H₆₀N₂O₂₁ [M + H]⁺ 1001.3767, measured 1001.3771.

***N*-(Fluoren-9-ylmethoxycarbonyl)-*O*-[*O*-(2',3',4',6'-tetra-*O*-acetyl-β-D-galactopyranosyl)-(1,3)-2-acetamido-4,6-di-*O*-acetoxy-2-deoxy-α-D-galactopyranosyl]-D-serine(10)**, Compound **S10** (750 mg, 0.749 mmol) was dissolved in a TFA-water mixture (95:5, 10 mL) and stirred at room temperature for 1 h. TLC-analysis of the reaction (EtOAc/hexanes 3:1) showed complete consumption of the starting material. The solvent was evaporated off and the residue was coevaporated with toluene (3 × 10 mL). The residue was dissolved in a water-acetonitrile mixture (1:1, 10 mL) and lyophilized to yield the carboxylic acid **10** (708 mg, quant.) as a colorless powder. The acid was purified with RP-HPLC as the same method for purifying **9**. ¹H NMR (400 MHz, *N,N*-dimethylformamide-*d*₇) δ 7.98 (d, *J* = 9.7 Hz, 1H), 7.95 (d, *J* = 7.6 Hz, 2H), 7.77 (t, *J* = 7.6 Hz, 2H), 7.53-7.45 (m, 3H), 7.34-7.38 (m, 2H), 5.41 (d, *J* = 3.2 Hz, 1H),

5.38 (d, $J = 3.6$ Hz, 1H), 5.12 (dd, $J = 10.4, 3.6$ Hz, 1H), 5.02 (dd, $J = 10.3, 7.8$ Hz, 1H), 4.93 (d, $J = 7.9$ Hz, 1H), 4.88 (d, $J = 3.7$ Hz, 1H), 4.58 – 4.48 (m, 2H), 4.45-4.35 (m, 2H), 4.32-4.21 (m, 2H), 4.20 – 4.04 (m, 7H), 3.91 (dd, $J = 11.3, 7.4$ Hz, 1H), 3.75 (dd, $J = 9.8, 3.9$ Hz, 1H), 2.13 (s, 3H), 2.10 (s, 3H), 2.07 (s, 3H), 2.04 (s, 6H), 1.98 (s, 3H), 1.96 (s, 3H); ^{13}C NMR (100 MHz, *N,N*-dimethylformamide- d_7) δ 171.8, 170.34, 170.31, 170.2, 170.1, 169.8, 169.4, 156.7, 144.5, 141.3, 127.9, 127.3, 125.5, 120.3, 101.1, 98.3, 73.9, 70.8, 70.5, 70.0, 68.9, 68.0, 66.6, 62.9, 61.2, 54.2, 48.7, 47.3, 22.6, 20.2, 20.15, 19.95, 19.9. HRMS calcd for $\text{C}_{44}\text{H}_{52}\text{N}_2\text{O}_{21}$ $[\text{M} + \text{H}]^+$ 945.3141, measured 945.3147.

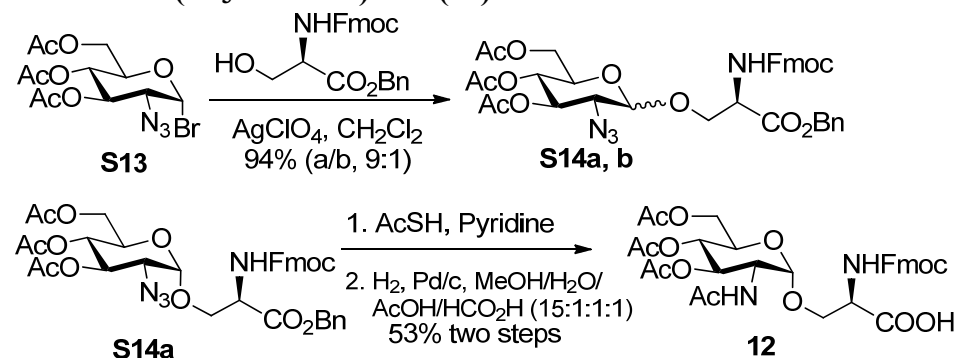
Fmoc-Ser(Ac₃GlcNAc α)-OH (**11**)



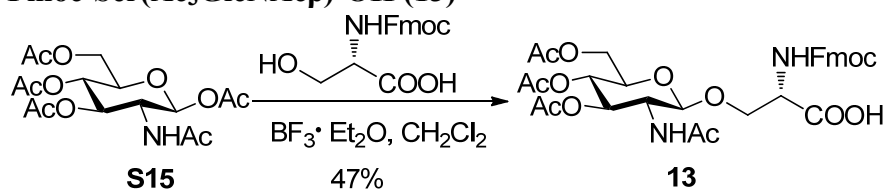
N-(9H-fluoren-9-ylmethoxycarbonyl)-*O*-(2-acetyl-2-deoxy-3,4,6-tri-*O*- α -D-glucopyranosyl)-L-serine **11** was synthesized following a reported method.¹ Peracetylated 2-azido-2-deoxy-D-Glucopyranosyl bromide (**S11**)² (0.85 g, 2.04 mmol), Fmoc-L-serine benzyl ester (1.12 g, 2.84 mmol), and 4 Å molecular sieves were placed in a flask under argon. Anhydrous DCM (30 mL) was added, and the reaction mixture was stirred at room temperature for 20 min. AgClO_4 (0.85 g, 4.10 mmol) was added to the reaction mixture was allowed to stir at room temperature for 3h, and monitored by TLC Hex/EtOAc (2:1). The reaction mixture was diluted with DCM, filtered over celite, and concentrated. The residue was filtered through a silica gel column with Hex/EtOAc (3:1) to give compounds **S12a** and **S12b** (α/β , 3:1) in 93% yield. Compound **S12a** (1.08 g, 1.48 mmol) was dissolved in pyridine (4mL) followed by addition of thioacetic acid (8mL). The reaction mixture was stirred at room temperature overnight and monitored by TLC Hex/EtOAc (1:2). The reaction mixture was diluted with DCM, filtered, concentrated, and filtered through a silica gel column to provide Fmoc-L-Ser(GlcNAc α)OBn (1.07 g, 97%). Fmoc-L-Ser(GlcNAc α)OBn (0.62 g, 0.83 mmol) was dissolved in MeOH (15 mL). Water (1 mL), acetic acid (1mL), and formic acid (1mL) were added, followed by addition of Pd/C 10%

(0.15g). The reaction mixture was purged with H₂ and stirred under H₂ for 2h at room temperature. The reaction mixture was filtered over celite, concentrated, and purified by RP-HPLC followed by silica gel column to provide compound **11** as a white solid (0.40g, 73%). ¹H NMR (400 MHz, *N,N*-dimethylformamide-*d*₇) δ 7.96-7.93 (m, 3H), 7.75 (d, *J* = 7.4Hz, 2H), 7.70 (d, *J* = 9.4 Hz, 1H), 7.45 (t, *J* = 7.4 Hz, 2H), 7.36 (t, *J* = 7.5 Hz, 2H), 5.27 (t, *J* = 9.5 Hz, 1H), 5.01 (t, *J* = 9.7 Hz, 1H), 4.95 (d, *J* = 3.6 Hz, 1H), 4.53-4.49 (m, 1H), 4.37–4.06 (m, 9H), 2.06 (s, 3H), 2.02 (s, 3H), 1.97 (s, 3H), 1.91 (s, 3H); ¹³C NMR (100MHz, *N,N*-dimethylformamide-*d*₇) δ 172.0, 170.4, 170.2, 170.1, 156.8, 144.40, 144.36, 141.35, 141.32, 127.9, 127.3, 125.6, 125.5, 120.3, 98.6, 71.0, 69.2, 68.6, 68.1, 66.7, 62.1, 54.9, 51.6, 47.3, 22.2, 20.1. HRMS calcd for C₃₂H₃₆N₂O₁₃[M + Na]⁺ 679.2115, measured 679.2104.

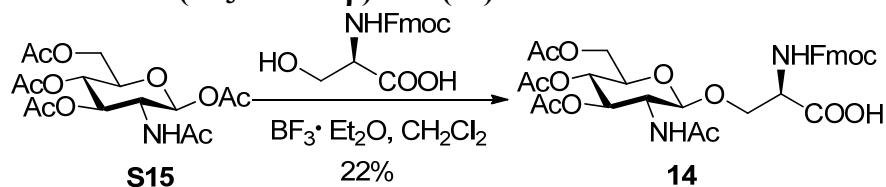
Fmoc-D-Ser(Ac₃GlcNAcα)-OH (**12**)



N-(9H-fluoren-9-ylmethoxycarbonyl)-*O*-(2-acetyl-2-deoxy-3,4,6-tri-*O*- α -D-glucopyranosyl)-*D*-serine **12** was synthesized in a similar method described for preparing **11** starting from peracetylated 2-azido-2-deoxy-*D*-Glucopyranosyl bromide (**S13**)¹ and Fmoc-*D*-serine benzyl ester (**S1**). ¹H NMR (400 MHz, *N,N*-dimethylformamide-*d*₇) δ 8.00-7.90 (m, 4H), 7.80-7.75 (m, 2H), 7.45 (t, *J* = 7.4 Hz, 2H), 7.34 (td, *J* = 1.0, 7.5 Hz, 2H), 5.17 (t, *J* = 9.5 Hz, 1H), 5.00 (t, *J* = 9.7 Hz, 1H), 4.94 (d, *J* = 3.6 Hz, 1H), 4.65-4.61(m, 1H), 4.39–4.02 (m, 8H), 3.74 (dd, *J* = 3.1, 9.6 Hz, 1H), 2.07 (s, 3H), 1.99 (s, 3H), 1.96 (s, 3H), 1.93 (s, 3H); ¹³C NMR (100MHz, *N,N*-dimethylformamide-*d*₇) δ 171.7, 170.4, 170.3, 169.9, 169.6, 156.8, 144.5, 144.3, 141.3, 127.9, 127.30, 127.29, 125.6, 125.5, 120.3, 97.8, 71.2, 68.9, 68.7, 68.2, 66.7, 62.0, 54.2, 51.4, 47.3, 22.2, 20.1, 20.0. HRMS calcd for C₃₂H₃₆N₂O₁₃ [M + Na]⁺ 679.2115, measured 679.2115.

Fmoc-Ser(Ac₃GlcNAcβ)-OH (13)

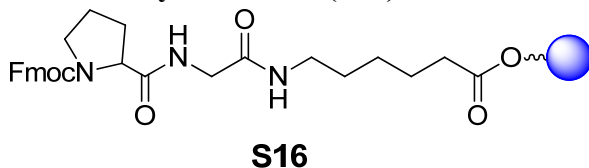
N-(9H-fluoren-9-ylmethoxycarbonyl)-*O*-(2-acetyl-2-deoxy-3,4,6-tri-*O*-β-D-glucopyranosyl)-*L*-serine **13** was synthesized according to reported method³. PeracetylatedGlcNAc **S15** (1.08 g, 2.77 mmol) with 4 Å molecular sieves were placed in a flask under argon, and anhydrous DCM (15 mL) was added. The reaction mixture was cooled to 0 °C and $\text{BF}_3 \cdot \text{Et}_2\text{O}$ (1.00 mL, 8.10 mmol) was added dropwise and stirred at room temperature overnight. The reaction mixture was cooled to 0 °C and Et_3N (0.40 mL, 1 equiv.) was added and stirred for 10 min. A solution of Fmoc-serine (1.20g, 3.67 mmol) in DCM/MeCN (1:2) was added, and the reaction was allowed to stir at room temperature for 4 days and monitored by TLC $\text{CDCl}_3/\text{MeOH}/\text{AcOH}$ (10:1:0.2). The reaction mixture with neutralized with Et_3N , diluted with DCM, filtered over celite, and concentrated. The residue was first filtered through a silica gel column with $\text{CHCl}_3/\text{MeOH}$ (40:1-10:1), then purified by RP-HPLC, and filtered through silica gel column DCM/MeOH (30:1-10:1) to provide compound **13** as a white solid (0.86g, 47%). ¹H NMR (400 MHz, *N,N*-dimethylformamide-*d*₇) δ 8.00 (d, *J* = 8.8Hz, 1H, NH-GalNAc), 7.93 (d, *J* = 7.6 Hz, 2H, H-Fmoc), 7.80–7.77 (m, 2H, H-Fmoc), 7.46–7.33 (m, 4H, H-Fmoc), 7.28 (d, *J* = 8.4Hz, 1H, NH-S), 5.28 (t, *J* = 9.2 Hz, 1H, H4'), 4.99-4.93 (m, 2H, H3' {4.97}, H1' {4.94}), 4.46–4.42 (m, 1H, S^α), 4.35–4.17 (m, 5H, H2' {4.35}, CH₂-Fmoc {4.33}, H6a' {4.19}, S^βa {4.18}), 4.11 (dd, *J* = 2.4, 12.4 Hz, 1H, H6b'), 3.97–3.85 (m, 3H, S^βb {3.95}, H-Fmoc {3.87}, H5'), 2.04 (s, 3H, CH₃-Ac), 2.02 (s, 3H, CH₃-Ac), 1.97 (s, 3H, CH₃-Ac), 1.85 (s, 3H, CH₃-Ac); ¹³C NMR (100MHz, *N,N*-dimethylformamide-*d*₇) δ 171.5, 170.4, 170.01, 169.7, 156.5, 144.40, 144.35, 141.33, 141.31, 127.9, 127.3, 125.63, 125.60, 120.3, 101.2, 73.0, 71.7, 69.2, 66.7, 62.3, 54.7, 54.0, 47.2, 22.5, 20.13, 20.08. HRMS calcd for C₃₂H₃₆N₂O₁₃ [M + Na]⁺ 679.2115, measured 679.2091.

Fmoc-D-Ser(Ac₃GlcNAcβ)-OH (14)

The synthesis of *N*-(9H-fluoren-9-ylmethoxycarbonyl)-*O*-(2-acetyl-2-deoxy-3,4,6-tri-*O*- β -D-glucopyranosyl)-D-serine (**14**) was carried out according to reported method³ in similar procedure described for compound **13** to provide compound **14** as a white solid (0.350g, 22%). ¹H NMR (400 MHz, *N,N*-dimethylformamide-*d*₇) δ 7.93 (d, *J* = 7.3 Hz, 3H, H-Fmoc), 7.79 (d, *J* = 7.4 Hz, 2H, H-Fmoc), 7.46–7.34 (m, 5H, H-Fmoc), 5.27 (t, *J* = 9.8 Hz, 1H, H4'), 4.96 (t, *J* = 9.6 Hz, 1H, H3'), 4.86 (d, *J* = 8.4 Hz, 1H, H1'), 4.45–4.25 (m, 5H, S^a {4.43}, CH₂-Fmoc {4.32}, H2' {4.32}, H6a' {4.25}), 4.14–3.86 (m, 5H, H6b' {4.12}, S^b {4.12}, {4.03}, H-Fmoc {3.91}, H5' {3.88}), 2.14–1.96 (m, 9H, CH₃-Ac), 1.83 (s, 3H, CH₃-Ac); ¹³C NMR (100MHz, *N,N*-dimethylformamide-*d*₇) δ 171.5, 170.4, 170.01, 169.95, 169.7, 156.5, 144.43, 144.41, 141.34, 141.31, 127.9, 127.3, 125.63, 125.60, 120.2, 101.7, 73.1, 71.6, 69.9, 69.3, 66.6, 62.4, 54.9, 54.0, 47.2, 22.5, 20.13, 20.10, 20.07. HRMS calcd for C₃₂H₃₆N₂O₁₃ [M + Na]⁺ 679.2115, measured 679.2092.

Preparation of peptide and glycopeptide standards

Fmoc-Pro-Gly-Hex-resin (S16)



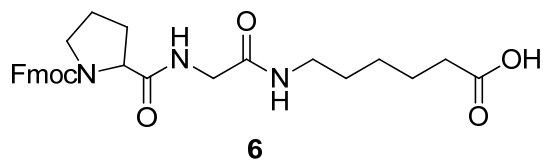
The solid-phase synthesis of Fmoc-Pro-Gly-Hex **S16** was carried out using 2-Chlorotriethyl chloride resin (1.0g, 1.55 mmol/g) manually functionalized with Fmoc-6-aminohexanoic acid (0.17 g, 0.48 mmol) and DIEA (0.34 mL, 1.92 mmol) in DCM (8.0 mL), followed by capping the resin with DCM/MeOH/DIEA (17:2:1, v/v/v, 3×5 mL) for 20 min. The resin was washed with DMF (2×5 mL), DCM (3×5 mL) and dried overnight under vacuum. The loading density of the Fmoc-6-aminohexanoic acid was calculated using Fmoc test by measuring piperidine-dibenzylfulvene adduct absorption at 290 nm with the following equation:

$$\text{Loading Density} \left(\frac{\text{mmol}}{\text{g}} \right) = \frac{A \times V (\mu\text{L})}{5800 \times m (\text{mg})}$$

where A is the absorption at 290 nm, V is the volume of 20% piperidine-DMF solution, and m is the weight of the loaded resin.⁴

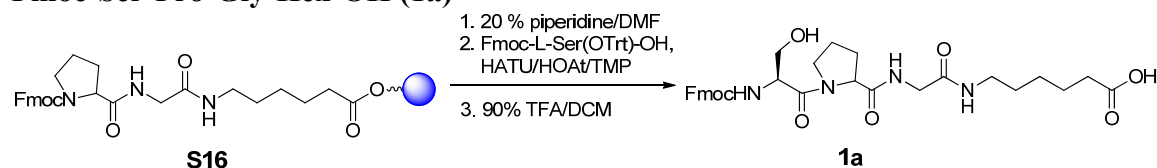
Fmoc groups on resin were removed with 20% piperidine in DMF (2×5 mL) for 15 min and then washed with DCM (3×5 mL) and DMF (2×5 mL). The first amino acid Fmoc-Gly-OH was preactivated with HBTU (0.67 g, 1.8 mmol), HOBt (0.28 g, 1.8 mmol), and DIEA (0.31 mL, 1.8 mmol) in DMF (8.0 mL) for 30 min, then coupled to the resin (1.0 g, 0.36 mmol) at room temperature for 1.5 h. The unreacted amino groups on the resin were capped with 10% acetic anhydride/10% pyridine in THF (2×5 mL) for 20 min. The resin was then deprotected with 20% piperidine, washed and then coupled with Fmoc-Pro-OPfp (0.89 g, 1.77 mmol) and DIEA (0.31 mL, 1.77 mmol) in DMF (8.0 mL) at room temperature for 2.0 h, followed by capping the resin with 10% acetic anhydride/10% pyridine in THF. The resin was washed with DMF (2×5 mL), DCM (3×5 mL) and dried under vacuum overnight.

Fmoc-Pro-Gly-Hex-OH (6)



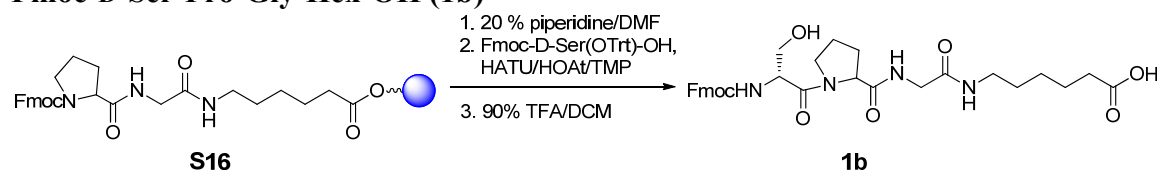
The Fmoc-Pro-Gly-Hex resin was treated with TFA/DCM (9/1, v/v, 2 mL) at room temperature and the crude product was purified by RP-HPLC. The flow rate is 10 mL/min and elute was monitored at 290 nm. Buffer A was MilliQ water containing 0.1% TFA, and buffer B was acetonitrile containing 0.1% TFA. For other peptides and glycopeptides, the percentage of B was increased after 1 min according to a linear gradient of 1%-35% in 18 min, kept at 35% for 4 min, increased from 35% to 48% in 56 min, increase to 100% B in 5 min, kept at 100 for 4 min, and then dropped back to 1% over 4 min and kept at 1% for 3 min (95 min total time). The trace amount of TFA was removed from the glycopeptides by a silica flash chromatography using a mixture of MeOH and DCM (10/90, v/v) to give **6** as a white foam. ¹H-NMR (400 MHz, CD₃OD) δ 7.81 (d, *J* = 7.4 Hz, 2H), 7.63 (t, *J* = 6.9 Hz, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.28-7.35 (m, 2H), 4.37-4.50 (m, 2H), 4.11-4.31 (m, 2H), 3.69-3.91 (m, 2H), 3.41-3.62 (m, 2H), 3.08-3.19 (m, 2H), 2.14-2.28 (m, 3H), 1.84-2.11 (m, 3H), 1.38-1.59 (m, 4H), 1.23-1.33 (m, 2H); ¹³C-NMR (100 MHz, CD₃OD) δ 178.2, 176.4, 172.3, 158.0, 146.1, 143.5, 129.7, 129.1, 121.8, 69.7, 63.2, 44.5, 41.2, 35.6, 32.0, 30.7, 28.3, 26.5. HRMS calcd for C₂₈H₃₃N₃O₆ [M + Na]⁺ 530.2265, measured 530.2250.

Fmoc-Ser-Pro-Gly-Hex-OH (**1a**)



Compound **1a** was synthesized starting from **S16**. Prior to coupling the resin (100 mg) was swollen in DCM for 0.5 h, followed by removal of the Fmoc-group with 20% piperidine in DMF (2× 2.0 mL) for 15 min, and then washed with DCM (3× 2.0 mL) and DMF (2× 2.0 mL). The appropriate amount of Fmoc-serine (2 eq), HATU (2 eq), HOAt (2 eq), and TMP (2eq) in DMF were mixed and immediately added to the resin. The reaction was allowed to mix at room temperature for 2 h, followed by washing and capping the unreacted amino groups with FmocOSu (10 eq) and DIEA (10 eq) in DMF at room temperature for 2 h. The resin was washed with DMF (2×5 mL), DCM (3×5 mL), and the peptide was cleaved off with a mixture TFA/DCM (90:10) (1.5 mL) at room temperature for 1.5 h. The crude product was purified by RP-HPLC using the same method for purifying **6**. The trace amount of TFA was removed from the glycopeptides by a silica flash chromatography using a mixture of methanol and dichloromethane to give as white foam. ¹H-NMR (400 MHz, CD₃OD) δ 7.79 (d, *J* = 7.5 Hz, 2H), 7.65 (d, *J* = 8.3 Hz, 2H), 7.38 (t, *J* = 7.4 Hz, 2H), 7.30 (t, *J* = 7.4 Hz, 2H), 4.57 (t, *J* = 6.6 Hz, 1H), 4.32-4.46 (m, 3H), 4.21 (t, *J* = 6.7 Hz, 1H), 3.70-3.87 (m, 6H), 3.15-3.25 (m, 2H), 2.19-2.30 (m, 3H), 1.93-2.11 (m, 3H), 1.50-1.66 (m, 4H), 1.31-1.41 (m, 2H); ¹³C-NMR (100 MHz, CD₃OD) δ 176.1, 173.4, 171.1, 169.9, 156.9, 143.7, 141.1, 127.3, 124.7, 119.5, 66.5, 54.5, 42.1, 38.8, 33.3, 28.9, 28.5, 25.9, 24.5, 24.2. HRMS calcd for C₃₁H₃₈N₄O₈ [M + Na]⁺ 617.2582, measured 617.2553.

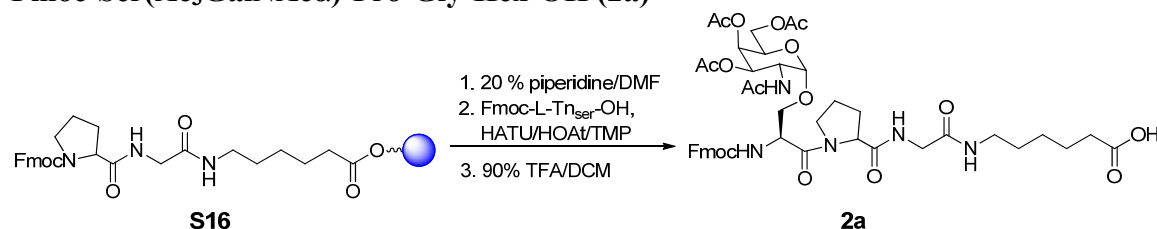
Fmoc-D-Ser-Pro-Gly-Hex-OH (**1b**)



Compound **1b** was synthesized and purified according to the general procedure described for **1a**. ¹H-NMR (400 MHz, CD₃OD) δ 7.79 (d, *J* = 7.5 Hz, 2H), 7.64 (dd, *J* = 7.5, 2.6 Hz, 2H), 7.38 (t, *J* = 7.4 Hz, 2H), 7.30 (t, *J* = 7.4 Hz, 2H), 4.47 (t, *J* = 6.7 Hz, 1H), 4.36-4.44 (m, 1H), 4.29 (dd, *J* = 10.5, 6.4 Hz, 1H), 3.66-3.93 (m, 6H), 2.96-3.19 (m, 2H), 2.22-2.28 (m, 3H), 1.93-2.09 (m, 3H),

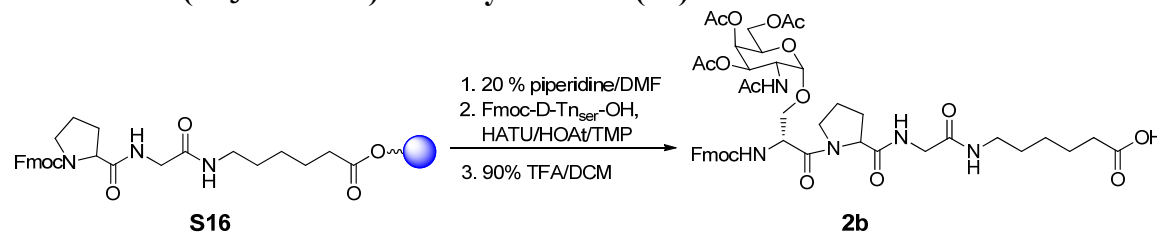
1.43-1.59 (m, 4H), 1.24-1.36 (m, 2H); ^{13}C -NMR (100 MHz, CD_3OD) δ 176.1, 173.3, 171.5, 169.9, 157.2, 143.7, 143.6, 141.2, 127.4, 126.7, 119.5, 66.6, 54.9, 42.1, 38.8, 33.4, 28.9, 28.3, 25.9, 24.3, 24.2. HRMS calcd for $\text{C}_{31}\text{H}_{38}\text{N}_4\text{O}_8$ $[\text{M} + \text{H}]^+$ 595.2766, measured 595.2754.

Fmoc-Ser($\text{Ac}_3\text{GalNAc}\alpha$)-Pro-Gly-Hex-OH (**2a**)



Fmoc-Ser($\text{Ac}_3\text{GalNAc}\alpha$)-Pro-Gly-Hex-OH (**2a**) was synthesized and purified according to the general procedure described for **6**. ^1H -NMR (400 MHz, CD_3OD) δ 7.80 (d, $J = 7.5$ Hz, 2H), 7.65 (t, $J = 6.6$ Hz, 2H), 7.39 (t, $J = 7.4$ Hz, 2H), 7.31 (t, $J = 7.0$ Hz, 2H), 5.40 (d, $J = 2.2$ Hz, 1H), 5.17 (dd, $J = 11.5, 3.0$ Hz, 1H), 4.93 (d, $J = 3.4$ Hz, 1H), 4.72 (t, $J = 5.5$ Hz, 1H), 4.40-4.51 (m, 3H), 4.37 (dd, $J = 12.4, 5.8$ Hz, 1H), 4.21-4.26 (m, 2H), 4.09 (dd, $J = 11.0, 6.4$ Hz, 1H), 3.88-4.04 (m, 3H), 3.78-3.87 (m, 1H), 3.63-3.78 (m, 3H), 3.02-3.30 (m, 2H), 2.27 (t, $J = 7.4$ Hz, 2H), 2.19-2.24 (m, 1H), 2.12 (m, 4H), 1.88-2.05 (m, 11H), 1.47-1.68 (m, 4H), 1.30-1.43 (m, 2H); ^{13}C -NMR (100 MHz, CD_3OD) δ 177.5, 173.7, 172.2, 172.1, 171.9, 171.5, 158.3, 145.2, 142.7, 128.9, 128.2, 126.2, 121.1, 99.8, 69.8, 68.9, 68.6, 68.2, 68.0, 62.9, 62.3, 54.2, 43.7, 40.4, 34.8, 30.5, 30.1, 27.5, 25.8, 22.8, 20.7, 20.6. HRMS calcd for $\text{C}_{45}\text{H}_{57}\text{N}_5\text{O}_{16}$ $[\text{M} + \text{Na}]^+$ 946.3700, measured 946.369.

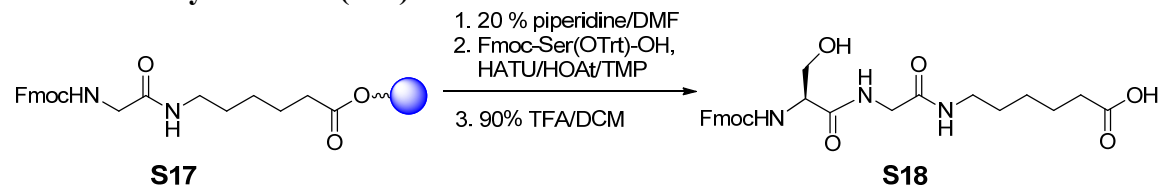
Fmoc-D-Ser($\text{Ac}_3\text{GalNAc}\alpha$)-Pro-Gly-Hex-OH (**2b**)



Compound **2b** was synthesized and purified according to the general procedure described for **6**. ^1H -NMR (400 MHz, CD_3OD) δ 7.79 (d, $J = 7.5$ Hz, 2H), 7.66 (dd, $J = 14.9, 7.2$ Hz, 2H), 7.39 (t, $J = 7.5$ Hz, 2H), 7.30-7.33 (m, 2H), 5.34-5.38 (m, 1H), 5.06-5.10 (m, 1H), 4.92-4.95 (m, 1H), 4.61-4.65 (m, 2H), 4.31-4.51 (m, 3H), 4.15-4.24 (m, 2H), 4.01-4.12 (m, 2H), 3.77-3.82 (m, 4H), 3.55-3.61 (m, 2H), 3.06-3.20 (m, 2H), 2.24-2.29 (m, 3H), 2.12-2.13 (m, 3H), 1.92-1.98 (m, 12H),

1.47-1.62 (m, 4H), 1.28-1.40 (m, 2H). HRMS calcd for C₄₅H₅₇N₅O₁₆ [M + Na]⁺ 946.3700, measured 946.3700.

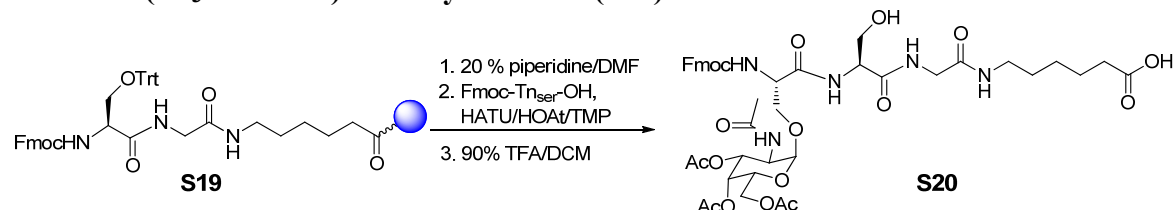
Fmoc-Ser-Gly-Hex-OH (S18)



Compound **S18** was synthesized from **S17** according to the general procedure described for **6**.

The crude product was purified by flash chromatography MeOH/CH₂Cl₂ (5% then 20%). The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.83 (d, *J* = 7.6 Hz, 2H), 7.70 (t, *J* = 6.9 Hz, 2H), 7.42 (t, *J* = 7.2 Hz, 2H), 7.37-7.33 (m, 2H), 4.49-4.39 (m, 2H), 4.27 (t, *J* = 6.9 Hz, 1H), 4.17 (t, *J* = 5.3 Hz, 1H), 3.88-3.79 (m, 4H), 3.20-3.17 (m, 2H), 2.26 (t, *J* = 7.3 Hz, 2H), 1.63-1.48 (m, 4H), 1.39-1.31 (m, 2H); ¹³C-NMR (100 MHz, CD₃OD) δ 174.6, 172.3, 159.6, 146.1, 143.5, 129.7, 129.1, 127.1, 121.8, 69.0, 63.8, 59.7, 44.5, 41.1, 30.7, 28.3, 26.8. HRMS calcd for C₂₆H₃₁N₃O₇ [M + Na]⁺ 520.2060, measured 520.2046.

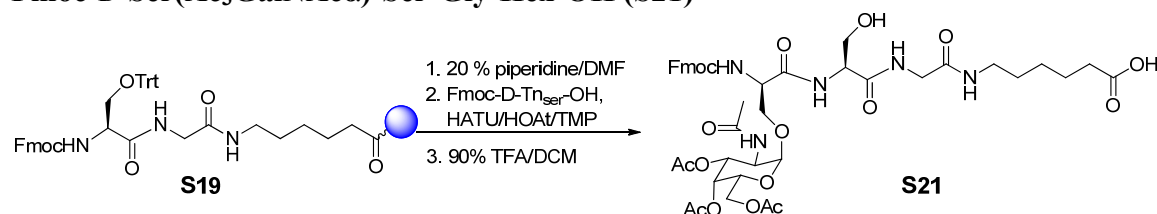
Fmoc-Ser(Ac₃GalNAc)-Ser-Gly-Hex-OH (S20)



Compound **S20** was synthesized from **S19** according to the general procedure described for **6**.

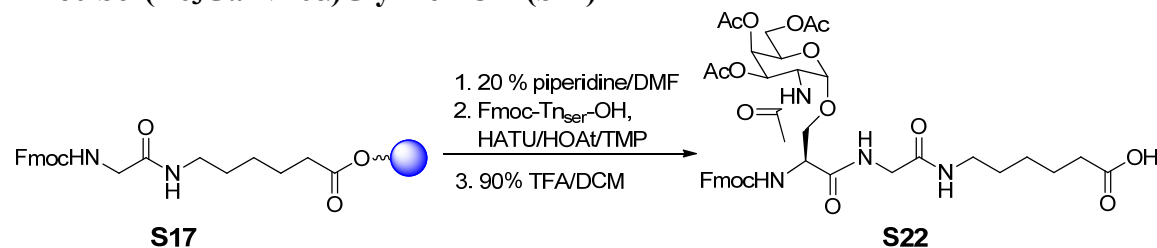
The crude product was purified by flash chromatography MeOH/CH₂Cl₂ (5% then 20%). The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.83 (d, *J* = 7.5 Hz, 2H), 7.71 (t, *J* = 7.7 Hz, 2H), 7.43 (t, *J* = 7.4 Hz, 2H), 7.36-7.34 (m, 2H), 5.41 (d, *J* = 3.0 Hz, 1H), 5.18 (dd, *J* = 11.5, 3.5 Hz, 1H), 4.49-4.44 (m, 5H), 4.27 (t, *J* = 6.5 Hz, 1H), 4.10-4.06 (m, 3H), 3.96-3.85 (m, 6H), 2.27 (t, *J* = 7.4 Hz, 2H), 2.16 (s, 3H), 1.98 (br, 9H), 1.67-1.50 (m, 4H), 1.40-1.32 (m, 2H), ¹³C-NMR (100 MHz, CD₃OD) δ 171.21, 170.8, 170.7, 170.6, 170.4, 169.9, 161.94, 161.6, 157.1, 143.8, 143.7, 141.2, 141.17, 127.4, 126.8, 124.7, 119.5, 118.1, 115.2, 98.4, 68.2, 67.83, 67.24, 66.72, 61.45, 61.35, 55.58, 54.81, 42.14, 38.84, 28.47, 25.99, 24.42, 21.30, 19.21, 19.13, 19.07. HRMS calcd for C₄₃H₅₅N₅O₁₇ [M + Na]⁺ 936.345, measured 936.352.

Fmoc-D-Ser(Ac₃GalNAc)-Ser-Gly-Hex-OH (S21)



Compound **S21** was synthesized from **S19** according to the general procedure described for **5**. The crude product was purified by flash chromatography MeOH/CH₂Cl₂ (5% then 20%). The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.83 (d, *J* = 7.5 Hz, 2H), 7.70 (t, *J* = 6.3 Hz, 2H), 7.42 (t, *J* = 7.3 Hz, 2H), 7.37-7.33 (m, 2H), 5.40 (d, *J* = 2.9 Hz, 1H), 5.13 (dd, *J* = 11.5, 3.2 Hz, 1H), 4.59-4.45 (m, 5H), 4.28 (t, *J* = 6.2 Hz, 1H), 4.20-3.83 (m, 8H), 3.72-3.69 (m, 1H), 3.24-3.11 (m, 2H), 2.28 (t, *J* = 7.4 Hz, 2H), 2.16 (s, 3H), 2.02 (s, 3H), 1.97 (s, 3H), 1.95 (s, 3H), 1.65-1.48 (m, 4H), 1.39-1.33 (m, 2H); ¹³C-NMR (100 MHz, CD₃OD) δ 173.1, 172.9, 172.7, 172.2, 146.0, 143.52, 143.51, 129.7, 129.1, 129.07, 126.9, 121.9, 99.8, 70.5, 69.6, 69.0, 63.8, 63.6, 58.1, 57.0, 44.5, 41.1, 30.8, 28.3, 26.7, 23.6, 21.5, 21.4. HRMS calcd for C₄₃H₅₅N₅O₁₇ [M + Na]⁺ 936.349, measured 936.353.

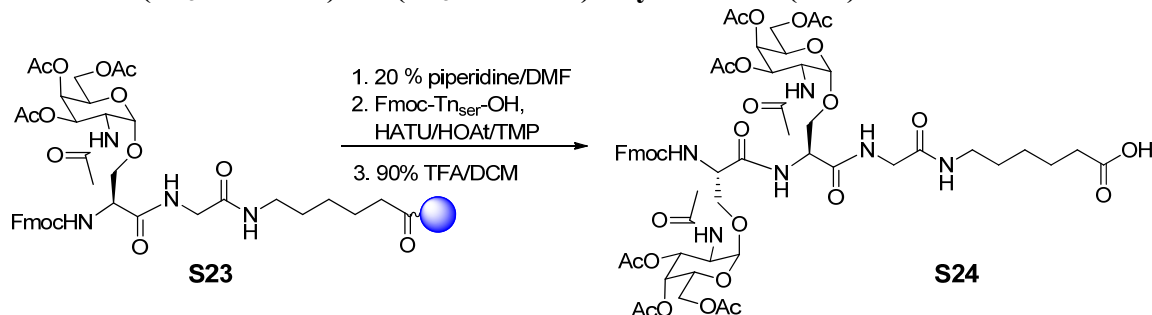
Fmoc-Ser(Ac₃GalNAc)Gly-Hex-OH (S22)



Compound **S22** was synthesized from **S17** according to the general procedure described for **6**. The crude product was purified by flash chromatography MeOH/CH₂Cl₂ (5% then 20%). The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.84 (d, *J* = 7.5 Hz, 2H), 7.71 (t, *J* = 6.8 Hz, 2H), 7.43 (t, *J* = 7.4 Hz, 2H), 7.39-7.36 (m, 2H), 5.42 (dd, *J* = 3.1, 1.1 Hz, 1H), 5.19 (dd, *J* = 11.6, 3.2 Hz, 1H), 4.94 (t, *J* = 3.6 Hz, 1H), 4.57-4.46 (m, 3H), 4.42 (t, *J* = 5.0 Hz, 1H), 4.30-4.24 (m, 2H), 4.09-4.03 (m, 2H), 3.99-3.80 (m, 4H), 3.22-3.18 (m, 2H), 2.28 (t, *J* = 7.4 Hz, 2H), 2.17, (s, 3H), 1.99 (s, 3H), 1.96 (s, 6H), 1.65-1.50 (m, 4H), 1.41-1.32 (m, 2H); ¹³C-NMR (100 MHz, CD₃OD) δ 174.5, 173.4, 173.35, 173.0, 172.96, 172.7, 171.9, 146.1, 146.0, 129.7, 129.1, 127.0, 121.9, 100.6, 70.6, 69.9, 69.5, 69.1, 69.0, 63.8, 57.4, 44.2,

41.2, 30.9, 28.4, 26.8, 23.6, 21.5, 21.4, 21.38. HRMS calcd for $C_{40}H_{50}N_4O_{15}$ $[M + Na]^+$ 849.317, measured 849.314.

Fmoc-Ser(Ac₃GalNAc)-Ser(Ac₃GalNAc)-Gly-Hex-OH (S24)

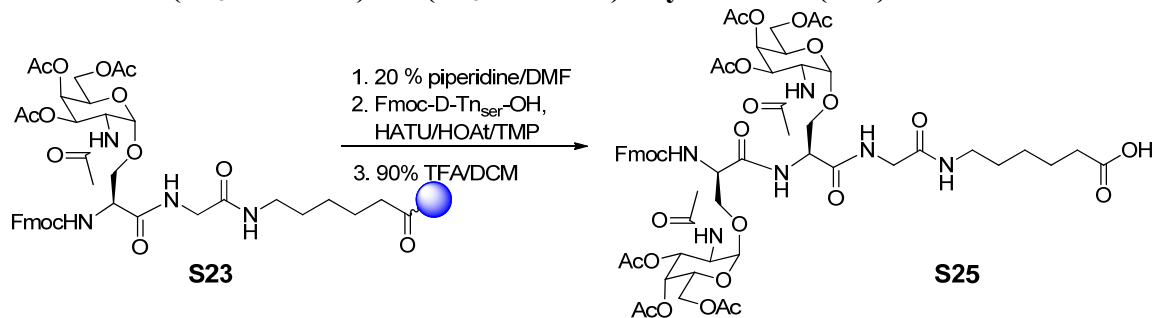


Compound **S24** was synthesized from **S24** according to the general procedure described for **5**.

The crude product was purified by flash chromatography with MeOH/CH₂Cl₂ (5% then 20%).

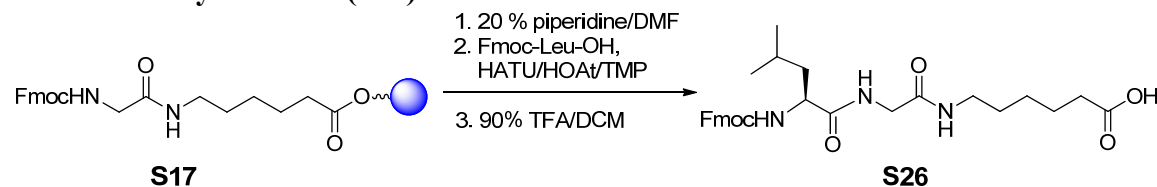
The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.80 (d, *J* = 7.3 Hz, 2H), 7.69-7.66 (m, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.34-7.31 (m, 2H), 5.40-5.37 (m, 2H), 5.19-5.13 (m, 2H), 4.95-4.93 (m, 2H), 4.69 (t, *J* = 5.1 Hz, 1H), 4.78-4.43 (m, 5H), 4.28-4.21 (m, 3H), 4.11-3.93 (m, 6H), 3.87-3.79 (m, 4 H), 3.25-3.11 (m, 2 H), 2.28 (t, *J* = 7.4 Hz, 2H), 2.13, (s, 3H), 2.11 (s, 3H), 1.95 (s, 15H), 1.88 (s, 3H), 1.65-1.57 (m, 2H), 1.55-1.48 (m, 2H), 1.40-1.33 (m, 2H); ¹³C-NMR (100 MHz, CD₃OD) δ 178.4, 174.5, 174.45, 173.1, 173.0, 172.4, 171.6, 159.3, 146.2, 146.0, 143.5, 129.7, 129.1, 129.08, 127.1, 127.0, 121.9, 100.8, 100.5, 70.6, 70.5, 70.2, 69.8, 69.6, 69.5, 69.0, 69.01, 68.98, 63.8, 63.7, 57.0, 55.5, 44.2, 41.2, 35.8, 30.9, 28.3, 26.6, 23.7, 23.6, 21.6, 21.51, 21.50, 21.40, 21.38. HRMS calcd for $C_{57}H_{74}N_6O_{25}$ $[M + Na]^+$ 1265.460, measured 1265.464.

Fmoc-D-Ser(Ac₃GalNAc)-Ser(Ac₃GalNAc)-Gly-Hex-OH (S25)



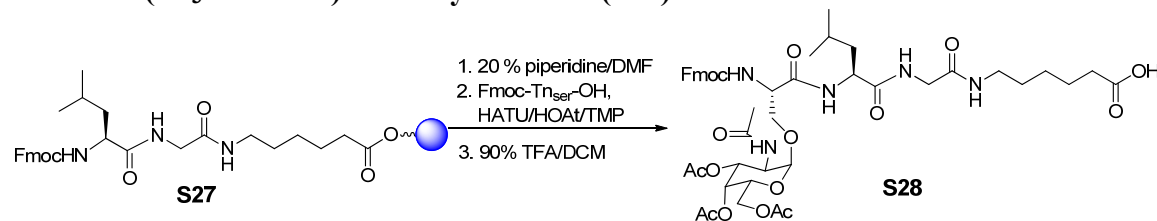
Compound **S25** was synthesized from **S23** according to the general procedure described for **6**. The crude product was purified by flash chromatography MeOH/CH₂Cl₂ (5% then 20%). The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.83 (d, *J* = 7.6 Hz, 2H), 7.75-7.72 (m, 2H), 7.43 (t, *J* = 7.4 Hz, 2H), 7.38-7.34 (m, 2H), 5.46 (br, 1H), 5.39 (d, *J* = 3.1 Hz, 1H), 5.20 (dd, *J* = 11.5, 3.3 Hz, 1H), 5.10 (dd, *J* = 11.4, 3.2 Hz, 1H) 4.96-4.94 (m, 2H), 4.73 (t, *J* = 4.4 Hz, 1H), 4.65-4.61 (m, 1H), 4.52-4.48 (m, 4H), 4.31 (t, *J* = 6.4 Hz, 2H), 4.17-3.90 (m, 10 H), 3.78-3.75 (m, 1 H), 3.26-3.13 (m, 2 H), 2.30 (t, *J* = 7.4 Hz, 2H), 2.16, (s, 3H), 2.15 (s, 3H), 1.99 (s, 3H), 2.02-1.96 (m, 18H), 1.68-1.50 (m, 4H), 1.39-1.32 (m, 2H); ¹³C-NMR (100 MHz, CD₃OD) δ 174.5, 174.4, 173.1, 172.9, 172.8, 172.7, 172.6, 172.5, 171.7, 146.2, 146.1, 143.5, 129.7, 129.14, 129.10, 127.0, 126.99, 121.9, 100.4, 99.7, 70.54, 70.50, 69.6, 69.14, 69.11, 69.06, 63.9, 63.8, 57.4, 55.3, 44.2, 41.2, 36.0, 30.9, 28.4, 26.7, 23.6, 21.6, 21.53, 21.52, 21.4. HRMS calcd for C₅₇H₇₄N₆O₂₅ [M + Na]⁺ 1265.460, measured 1265.460.

Fmoc-Leu-Gly-Hex-OH (**S26**)



Compound **S26** was synthesized from **S17** according to the general procedure described for **6**. The crude product was purified by flash chromatography MeOH/CH₂Cl₂ (5% then 15%). The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.82 (d, *J* = 7.6 Hz, 2H), 7.68 (t, *J* = 7.4 Hz, 2H), 7.41 (t, *J* = 7.5 Hz, 2H), 7.35-7.31 (m, 2H), 4.50-4.46 (m, 2H), 4.25 (t, *J* = 6.8 Hz, 1H), 4.10-4.06 (m, 1H), 3.94 (d, *J* = 16.8 Hz, 1H), 3.73 (t, *J* = 16.8 Hz, 1H), 3.21-3.06 (m, 2H), 2.23 (t, *J* = 7.4 Hz, 2H), 1.73-1.46 (m, 7H), 1.34-1.27 (m, 2H), 0.98 (dd, *J* = 13.7, 6.4 Hz, 6H); ¹³C-NMR (100 MHz, CD₃OD) δ 178.4, 176.9, 172.2, 159.7, 146.0, 143.5, 129.7, 129.1, 129.0, 127.1, 127.0, 121.8, 68.8, 56.3, 44.4, 42.2, 41.3, 41.1, 35.7, 30.8, 28.3, 26.7, 26.6, 24.2, 22.9, 19.1. HRMS calcd for C₂₉H₃₇N₃O₆ [M + H]⁺ 524.2761, measured 524.2731.

Fmoc-Ser(Ac₃GalNAc)-Leu-Gly-Hex-OH (**S28**)



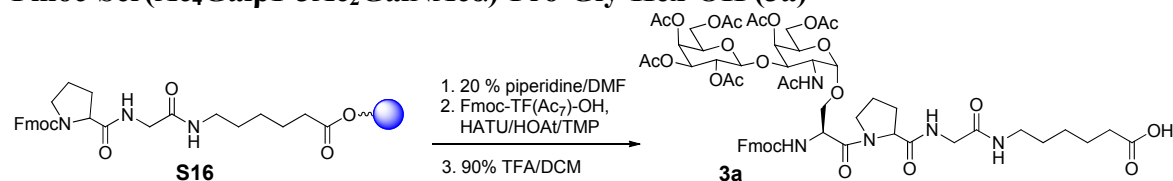
Compound **S28** was synthesized from **S27** according to the general procedure described for **6**. The crude product was purified by flash chromatography MeOH/CH₂Cl₂ (5% then 15%). The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.71 (d, *J* = 7.5 Hz, 2H), 7.57 (t, *J* = 7.5 Hz, 2H), 7.30 (t, *J* = 7.3 Hz, 2H), 7.25-7.21 (m, 2H), 5.29 (d, *J* = 3.0 Hz, 1H), 5.05 (dd, *J* = 11.5, 3.0 Hz, 1H), 4.83 (d, *J* = 3.2 Hz, 1H), 4.37-4.32 (m, 4H), 4.25 (dd, *J* = 8.8, 5.9 Hz, 1H), 4.14 (d, *J* = 6.6 Hz, 2H), 4.02-3.89 (m, 2H), 3.84-3.80 (m, 2H), 3.72-3.63 (m, 2H), 3.17-3.04 (m, 2H), 2.18 (t, *J* = 7.4 Hz, 2H), 2.04 (s, 3H), 1.86 (s, 3 H), 1.85 (s, 3 H), 1.84 (s, 3 H), 1.57-1.40 (m, 7H), 1.29-1.19 (m, 2H), 0.85 (dd, *J* = 11.3, 5.9 Hz, 6H); ¹³C-NMR (100 MHz, CD₃OD) δ 176.1, 173.4, 172.1, 170.8, 170.4, 169.8, 156.9, 143.8, 143.7, 141.2, 127.4, 126.8, 124.7, 119.5, 98.5, 68.3, 68.0, 67.2, 66.7, 66.66, 61.4, 54.6, 52.3, 42.0, 40.1, 38.9, 33.4, 28.6, 26.0, 24.4, 24.3, 19.2, 19.1, 19.07, . HRMS calcd for C₄₆H₆₁N₅O₁₆ [M + Na]⁺ 962.401, measured 962.408.

Fmoc-D-Ser(Ac₃GalNaca)-Leu-Gly-Hex-OH (**S29**)



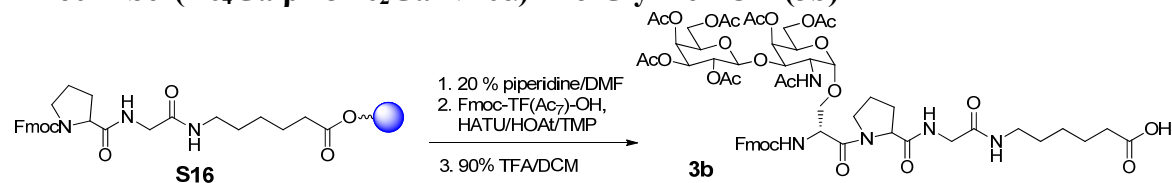
Compound **S29** was synthesized from **S27** according to the general procedure described for **6**. The crude product was purified by flash chromatography MeOH/CH₂Cl₂ (5% then 15%). The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.71 (d, *J* = 7.5 Hz, 2H), 7.61-7.56 (m, 2H), 7.30 (t, *J* = 7.4 Hz, 2H), 7.25-7.21 (m, 2H), 5.27 (d, *J* = 3.1 Hz, 1H), 4.99 (dd, *J* = 11.5, 3.2 Hz, 1H), 4.46-4.35 (m, 5H), 4.15 (t, *J* = 6.1 Hz, 1H), 4.05-4.00 (m, 2H), 3.93-3.85 (m, 2H), 3.78-3.67 (m, 2H), 3.60 (dd, *J* = 10.0, 3.2 Hz, 2H), 3.14-2.99 (m, 2 H), 2.16 (t, *J* = 7.4 Hz, 1H), 2.04 (s, 3H), 1.88 (s, 3 H), 1.85 (s, 3 H), 1.81 (s, 3 H), 1.58-1.51 (m, 5H), 1.42-1.36 (m, 2H), 1.26-1.19 (m, 2H), 0.85 (dd, *J* = 8.2, 5.6 Hz, 1H); ¹³C-NMR (100 MHz, CD₃OD) δ 178.4, 175.8, 174.4, 173.2, 173.0, 172.9, 172.7, 172.1, 159.4, 146.1, 146.05, 143.6, 143.5, 129.7, 129.1, 129.1, 126.9, 126.8, 121.9, 99.9, 70.5, 70.0, 69.5, 69.0, 68.9, 63.7, 57.0, 54.6, 44.4, 42.4, 41.1, 35.7, 30.8, 28.3, 26.9, 26.6, 24.3, 23.6, 22.8, 21.5, 21.4. HRMS calcd for C₄₆H₆₁N₅O₁₆ [M + Na]⁺ 962.401, measured 962.408.

Fmoc-Ser(Ac₄Galβ1-3Ac₂GalNAcα)-Pro-Gly-Hex-OH (**3a**)



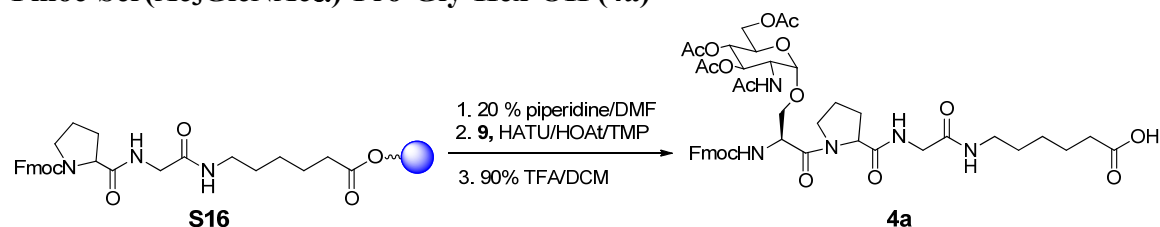
Compound **3a** was synthesized from **S16** according to the general procedure described for **6**. The crude product was purified by flash chromatography MeOH/CH₂Cl₂ (5% then 15%). The product was lyophilized to give a white solid. ¹H-NMR (400 MHz, CD₃OD) δ 7.71 (d, *J* = 7.5 Hz, 2H), 7.54 (d, *J* = 7.5 Hz, 2H), 7.30 (t, *J* = 7.3 Hz, 2H), 7.21 (t, *J* = 7.4 Hz, 2H), 5.29 (br, 1H), 5.25 (d, *J* = 3.3 Hz, 1H), 5.03-4.99 (m, 1H), 4.89 (dd, *J* = 10.2, 8.04 Hz, 1H), 4.65-4.60 (m, 1H), 4.52 (d, *J* = 7.7 Hz, 1H), 4.39-3.50 (m, 18H), 3.31-3.24 (m, 2H), 3.03-2.91 (m, 2H), 2.17 (t, *J* = 7.4 Hz, 3H), 2.02 (s, 3H), 2.01 (s, 3H), 1.93-1.89 (m, 13H), 1.83 (s, 3H), 1.57-1.46 (m, 4H), 1.34-1.26 (m, 2H); ¹³C-NMR (100 MHz, CD₃OD) δ 178.3, 175.9, 173.9, 173.2, 172.9, 172.9, 172.8, 172.3, 172.28, 171.9, 171.2, 159.0, 146.0, 146.0, 143.5, 129.8, 129.1, 127.0, 126.9, 121.9, 103.6, 100.3, 76.7, 73.2, 72.4, 72.1, 71.1, 69.9, 69.5, 69.1, 68.9, 64.9, 63.2, 54.8, 50.9, 49.3, 44.7, 41.4, 35.7, 31.3, 31.0, 28.4, 27.2, 26.6, 24.0, 21.9, 21.7, 21.6, 21.61, 21.3. HRMS calcd for C₅₇H₇₃N₅O₂₄ [M + Na]⁺ 1234.454, measured 1234.457.

Fmoc-D-Ser(Ac₄Galβ1-3Ac₂GalNAcα)-Pro-Gly-Hex-OH (**3b**)



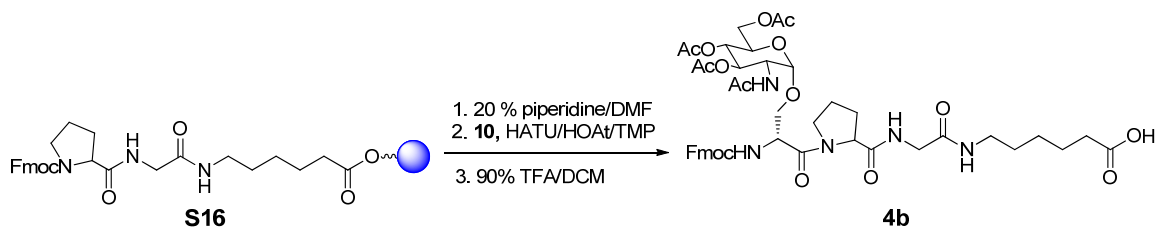
Compound **3b** was synthesized and purified according to the general procedure described for **6**. ¹H-NMR (400 MHz, *N,N*-dimethylformamide-*d*₇) δ 7.96-7.94 (m, 2H), 7.82-7.74 (m, 2H), 7.48-7.44 (m, 2H), 7.39-7.34 (m, 2H), 5.44-5.36 (m, 2H), 5.22-4.73 (m, 5H), 4.64-3.61 (m, 19H), 3.28-3.03 (m, 2H), 2.29-2.24 (m, 3H), 2.13-1.95 (m, 23H), 1.58-1.51 (m, 4H), 1.38-1.30 (m, 2H). HRMS calcd for C₅₇H₇₃N₅O₂₄ [M + Na]⁺ 1234.454, measured 1234.458.

Fmoc-Ser(Ac₃GlcNAc α)-Pro-Gly-Hex-OH (4a)



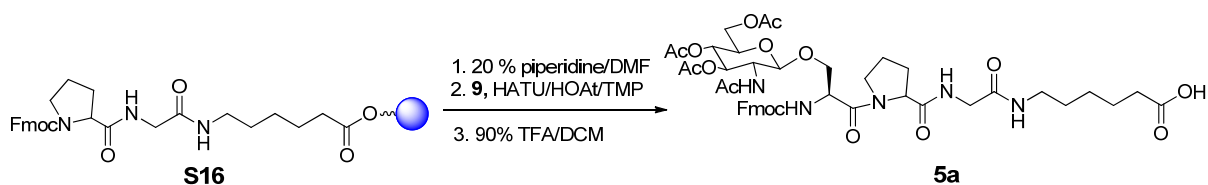
Glycopeptide **4a** was synthesized and purified according to the general procedure described for **6**. ¹H NMR (400 MHz, CD₃OD) δ 7.81 (d, J = 7.5 Hz, 2H), 7.7.66 (d, J = 7.4 Hz, 2H), 7.40 (t, J = 7.4 Hz, 2H), 7.32 (t, J = 7.5 Hz, 2H), 5.21 (t, J = 9.9 Hz, 1H), 5.00 (t, J = 10.0 Hz, 1H), 4.90-3.68 (m, 16H), 3.27-3.13 (m, 2H), 2.29-1.92 (m, 17H), 1.66-1.35 (m, 6H); ¹³C NMR (100 MHz, CD₃OD) δ 176.0, 172.1, 170.9, 170.4, 170.1, 169.8, 168.9, 156.8, 143.8, 143.7, 141.2, 127.4, 126.8, 124.6, 119.5, 98.0, 71.0, 68.7, 67.8, 67.2, 66.5, 61.8, 60.8, 52.8, 51.4, 42.2, 38.9, 33.3, 28.9, 28.6, 26.0, 24.8, 24.3, 21.2, 19.20, 19.16. HRMS calcd for C₄₅H₅₇N₅O₁₆[M + Na]⁺ 946.370, measured 946.363.

Fmoc-D-Ser(Ac₃GlcNAc α)-Pro-Gly-Hex-OH (4b)



Compound **4b** was synthesized and purified according to the general procedure described for **6**. ¹H NMR (400 MHz, CD₃OD) δ 7.80 (d, J = 7.5 Hz, 2H), 7.70-7.64 (m, 2H), 7.40 (t, J = 7.4 Hz, 2H), 7.32 (t, J = 7.3 Hz, 2H), 5.20 (t, J = 10.1 Hz, 1H), 5.00 (t, J = 9.8 Hz, 1H), 4.75-3.54 (m, 16H), 3.25-3.08 (m, 2H), 2.28-1.82 (m, 17H), 1.63-1.29 (m, 6H); ¹³C NMR (100 MHz, CD₃OD) δ 176.0, 173.3, 171.93, 171.92, 170.9, 170.5, 169.9, 169.7, 169.1, 157.2, 143.9, 143.6, 141.3, 127.42, 127.40, 126.8, 124.74, 124.69, 119.5, 97.2, 70.8, 68.6, 67.8, 67.1, 66.8, 66.4, 61.8, 61.3, 52.6, 51.4, 42.7, 42.2, 38.9, 33.3, 28.9, 28.7, 28.4, 26.0, 24.7, 24.2, 19.2., 19.1, 19.0. HRMS calcd for C₄₅H₅₇N₅O₁₆[M + Na]⁺ 946.370, measured 946.364.

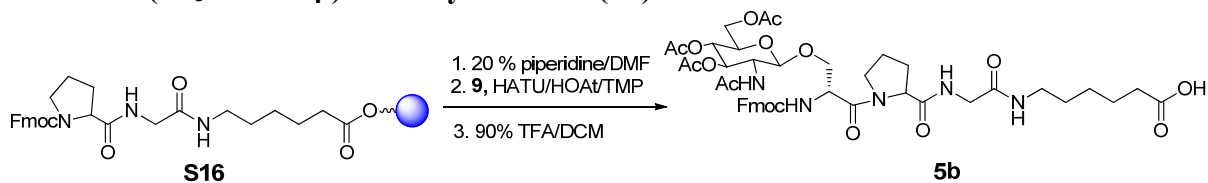
Fmoc-Ser(Ac₃GlcNAc β)-Pro-Gly-Hex-OH (5a)



Compound **5a** was synthesized and purified according to the general procedure described for **6**.

^1H NMR (400 MHz, CD_3OD) δ 7.80 (d, $J = 7.5\text{Hz}$, 2H), 7.65 (d, $J = 7.5\text{ Hz}$, 2H), 7.39 (t, $J = 7.4\text{ Hz}$, 2H), 7.31 (t, $J = 7.3\text{ Hz}$, 2H), 5.20 (t, $J = 9.8\text{ Hz}$, 1H), 4.98 (t, $J = 9.8\text{ Hz}$, 1H), 4.69-4.10 (m, 8H), 3.95-3.13 (m, 10H), 2.31-1.86(m, 17H), 1.64-1.34 (m, 6H); ^{13}C NMR (100MHz, CD_3OD) δ 176.0, 173.3, 172.0, 170.9, 170.4, 170.2, 169.9, 169.8, 156.8, 143.7, 141.2, 127.4, 126.8, 124.7, 119.5, 100.5, 72.7, 71.7, 68.6, 68.2, 66.5, 61.8, 61.0, 53.8, 52.7, 42.1, 38.9, 33.3, 29.0, 28.5, 26.0, 24.7, 24.3, 21.5, 19.2, 19.11, 19.09. HRMS calcd for $\text{C}_{45}\text{H}_{57}\text{N}_5\text{O}_{16}[\text{M} + \text{Na}]^+$ 946.370, measured 946.367.

Fmoc-D-Ser($\text{Ac}_3\text{GlcNAc}\beta$)-Pro-Gly-Hex-OH (**5b**)



Compound **5b** was synthesized and purified according to the general procedure described for **6**.

^1H NMR (400 MHz, CD_3OD) δ 7.79 (d, $J = 7.5\text{Hz}$, 2H), 7.64 (d, $J = 7.5\text{ Hz}$, 2H), 7.41-7.29 (m, 4H), 5.17 (t, $J = 9.3\text{ Hz}$, 1H), 4.98 (t, $J = 9.8\text{ Hz}$, 1H), 4.64-3.68 (m, 16H), 3.19-2.98 (m, 2H), 2.27-1.88 (m, 17H), 1.60-1.30 (m, 6H); ^{13}C NMR (100MHz, CD_3OD) δ 176.0, 173.2, 172.0, 170.9, 170.7, 170.4, 169.9, 169.8, 157.0, 143.7, 143.6, 141.25, 141.20, 127.4, 126.8, 124.7, 124.6, 119.5, 119.6, 100.9, 72.6, 71.6, 68.5, 66.5, 61.6, 61.3, 53.7, 53.0, 42.1, 38.8, 33.4, 29.0, 28.4, 25.9, 24.4, 24.2, 21.4, 19.3, 19.11, 19.07. HRMS calcd for $\text{C}_{45}\text{H}_{57}\text{N}_5\text{O}_{16}[\text{M} + \text{Na}]^+$ 946.370, measured 946.364.

HPLC analysis of products in coupling reactions

HPLC trace of Fmoc-Ser-Pro-Gly-Hex-OH

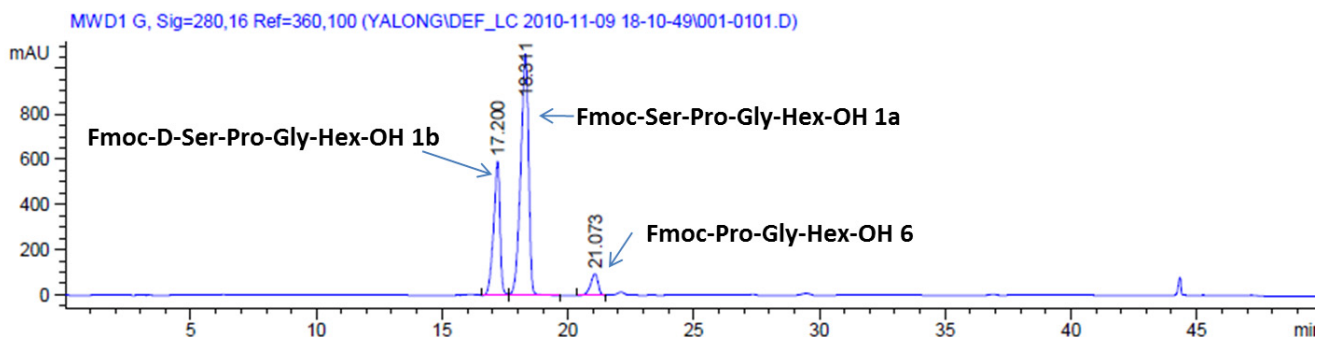


Figure S1. HPLC trace of Fmoc-Ser-Pro-Gly-Hex-OH **1a**, its racemized D isomer **1b** and the unreacted starting peptide **6**. Analyses were performed with Waters XTerra[®] RP 18 (4.6 mm × 250 mm) reverse phase column. UV absorption was measured at 280 nm and the flow rate was 1 mL/min. The percentage of B was increased according to a linear gradient of 1%-35% in 9 min, kept at 35% for 3 min, and then increased from 35% to 48% in 27 min.

HPLC trace of Fmoc-Ser(Ac₃GalNAc α)-Pro-Gly-Hex-OH

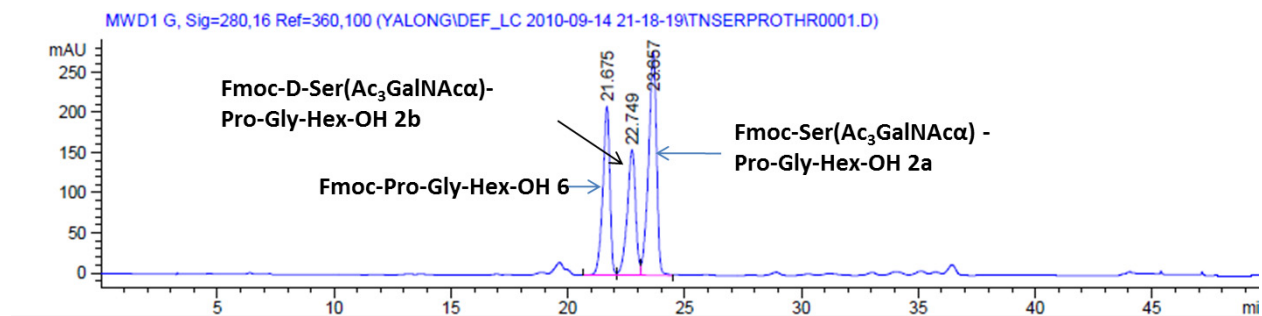


Figure S2. HPLC trace of Fmoc-Ser(Ac₃GalNAc α)-Pro-Gly-Hex-OH **2a**, its epimerization D isomer **2b** and the unreacted starting peptide **6**. Analyses were performed with the same conditions as those for obtaining Figure S1.

HPLC trace of Fmoc-Ser(Ac₃GalNAc α)-Ser-Gly-Hex-OH

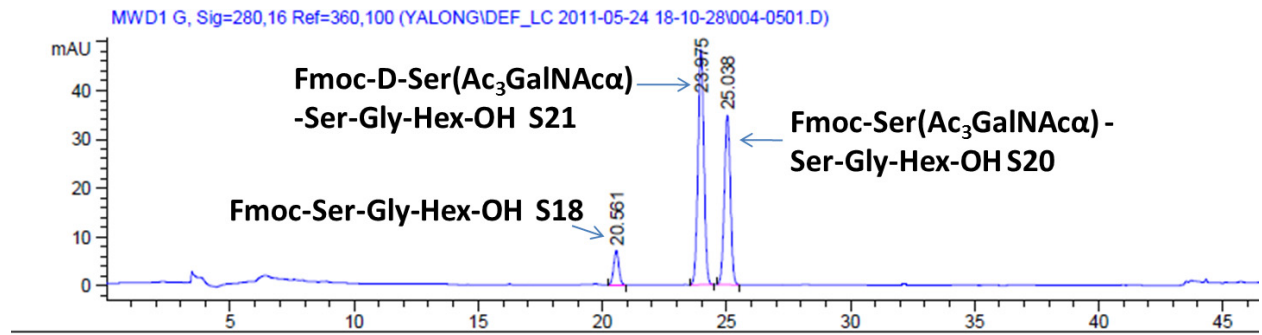


Figure S3. HPLC trace of Fmoc-Ser(Ac₃GalNAc α)-Ser-Gly-Hex-OH **S20**, its epimerization D isomer **S21** and the unreacted starting peptide **S18**. Analyses were performed with the same conditions as those for obtaining Figure S1.

HPLC trace of Fmoc-Ser(Ac₃GalNAc α)-Ser(Ac₃GalNAc α)-Gly-Hex-OH

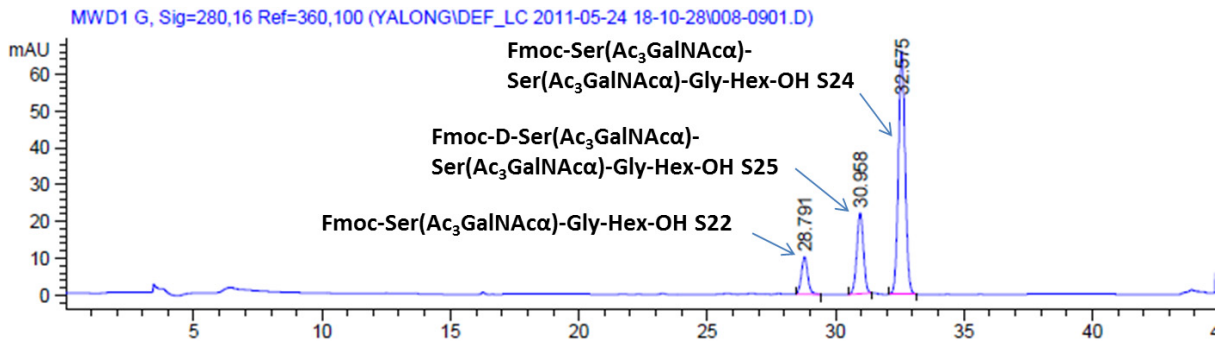


Figure S4. HPLC trace of Fmoc-Ser(Ac₃GalNAc α)-Ser(Ac₃GalNAc α)-Gly-Hex-OH **S24**, its epimerization D isomer **S25** and the unreacted starting glycopeptide **S22**. Analyses were performed with the same conditions as those for obtaining Figure S1.

HPLC trace of Fmoc-Ser(Ac₃GalNAc α)-Leu-Gly-Hex-OH

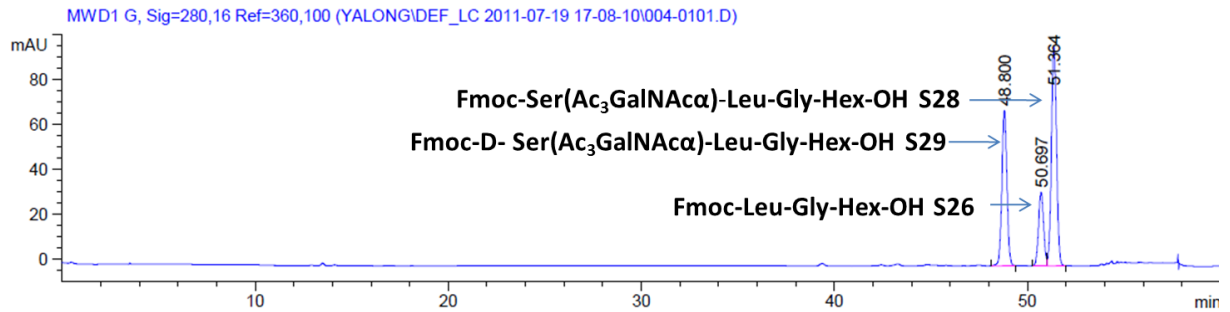


Figure S5. HPLC trace of Fmoc-Ser(Ac₃GalNAc α)-Leu-Gly-Hex-OH **S28**, its epimerization D isomer **S29** and the unreacted starting peptide **S26**. Analyses were performed with Waters XTerra[®] RP 18 (4.6 mm \times 250 mm) reverse phase column. UV absorption was measured at 280 nm and the flow rate was 1 mL/min. The percentage of B was increased according to a linear gradient of 1%-25% in 9 min, kept at 25% for 1 min, increased from 25% to 29% in 8 min, kept at 29% for 1.5 min, increased from 29% to 32% for 3.5 min, and then increased from 32% to 45% in 16 min.

HPLC trace of Fmoc-Ser(Ac₄Galβ1-3Ac₂GalNAcα)-Pro-Gly-Hex-OH

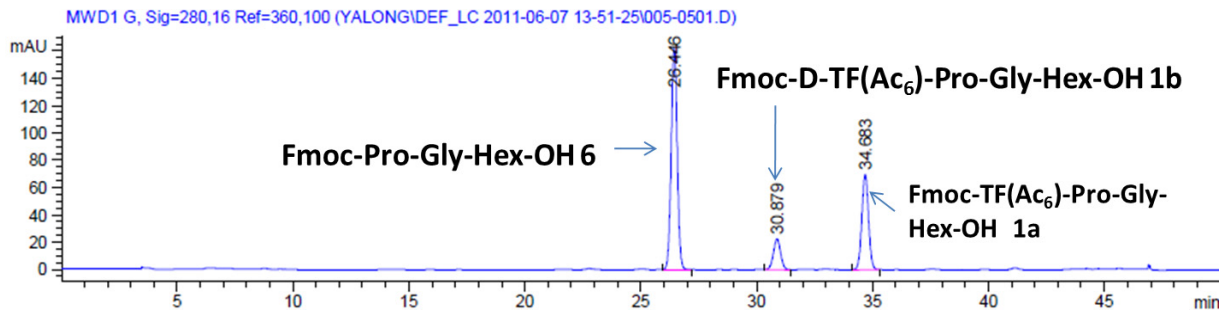


Figure S6. HPLC trace of Fmoc-Ser(Ac₄Galβ1-3Ac₂GalNAcα)(TF(Ac₆))-Pro-Gly-Hex-OH **3a**, its epimerization D isomer **3b** and the unreacted starting peptide **6**. Analyses Waters XTerra[®] RP 18 (4.6 mm × 250 mm) reverse phase column. UV absorption was measured at 280 nm and the flow rate was 1 mL/min. The percentage of B was increased according to a linear gradient of 1%-35% in 7 min, kept at 35% for 3 min, increased from 35% to 48% in 27 min.

HPLC trace of Fmoc-Ser(Ac₃GlcNAc α)-Pro-Gly-Hex-OH

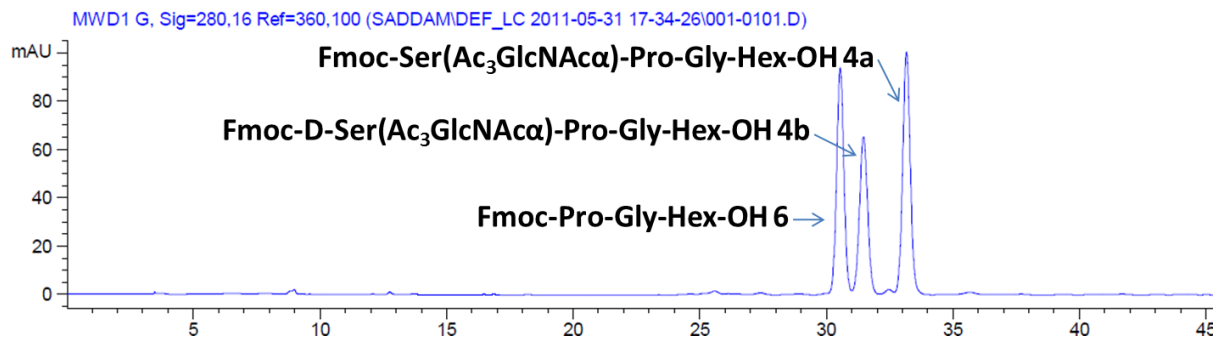


Figure S7. HPLC trace of Fmoc-Ser(Ac₃GlcNAc α)-Pro-Gly-Hex-OH **4a**, its epimerization D isomer **4b** and the unreacted starting peptide **6**. Analyses were performed with Waters XTerra[®] RP 18 (4.6 mm \times 250 mm) reverse phase column. UV absorption was measured at 280 nm and the flow rate was 1 mL/min. The percentage of B was increased according to a linear gradient of 1%-35% in 9 min, kept at 35% for 3 min, increased from 35% to 48% in 27 min.

HPLC trace of Fmoc-Ser(Ac₃GlcNAcβ)-Pro-Gly-Hex-OH

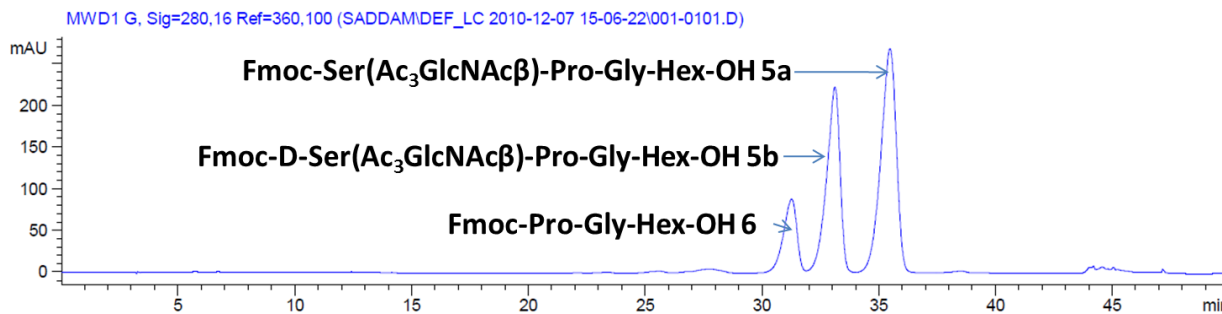
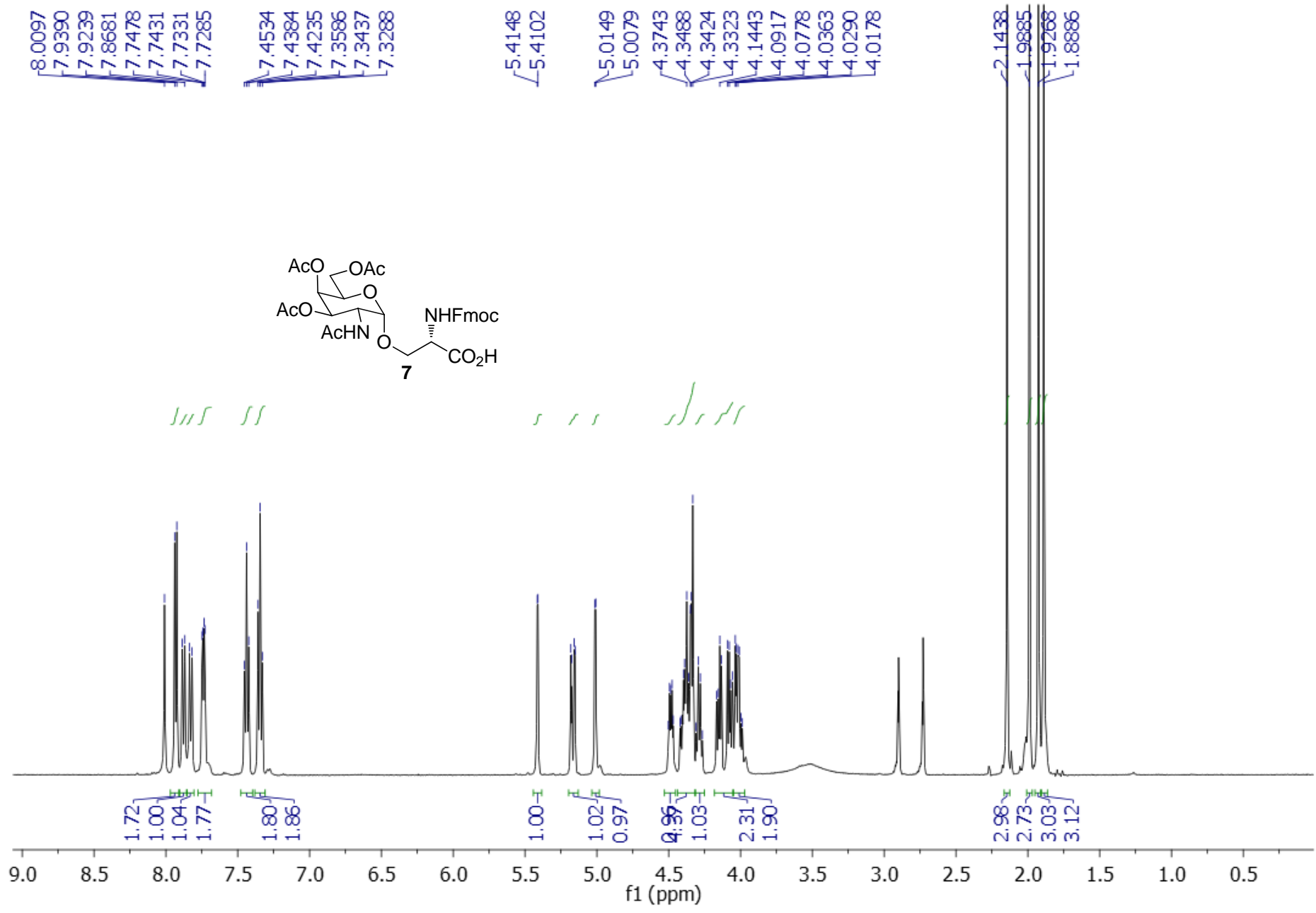
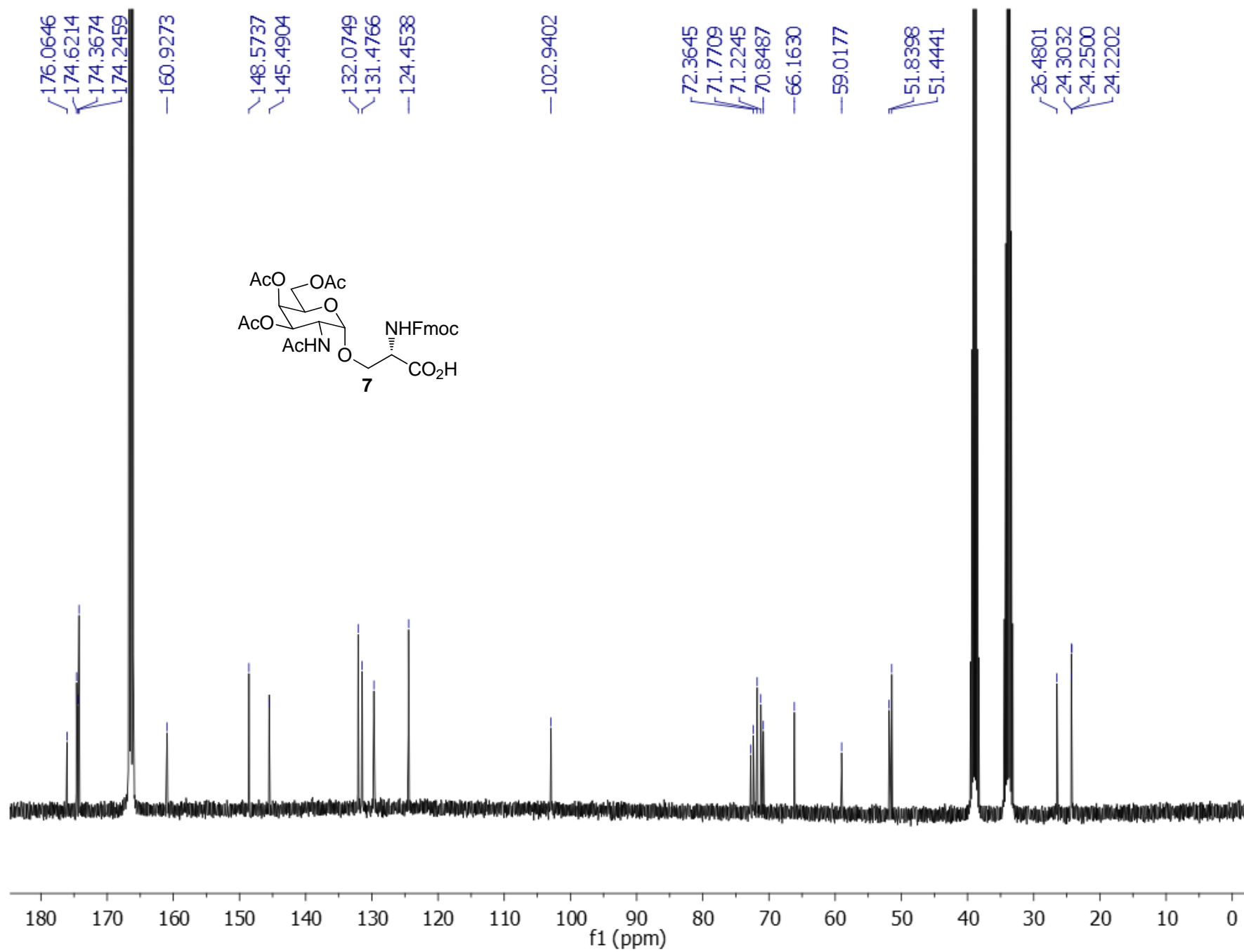


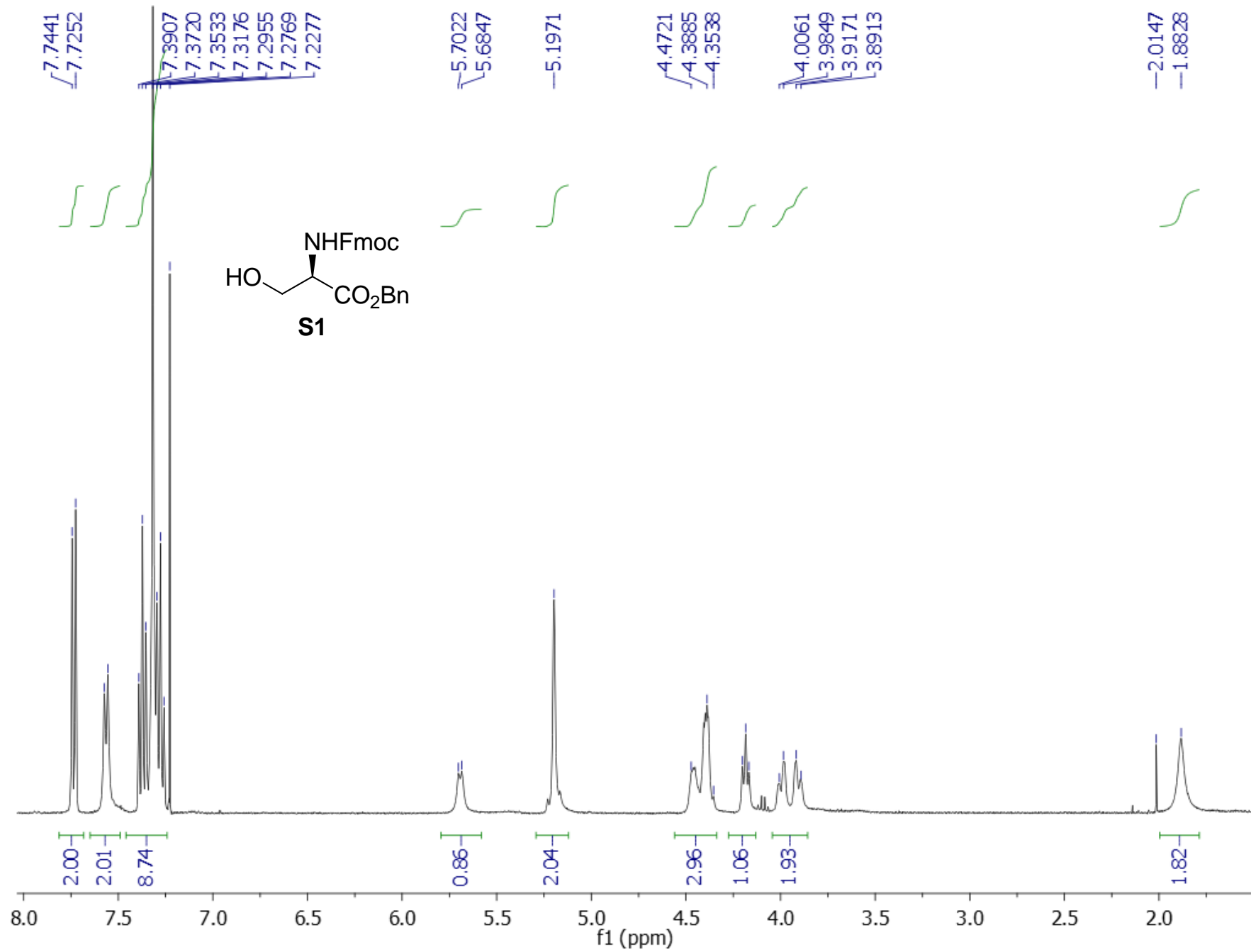
Figure S8. HPLC trace of Fmoc-Ser(Ac₃GlcNAcβ)-Pro-Gly-Hex-OH **5a**, its epimerization D isomer **5b** and the unreacted starting peptide **6**. Analyses were performed with Waters XTerra[®] RP 18 (4.6 mm × 250 mm) reverse phase column. UV absorption was measured at 280 nm and the flow rate was 1 mL/min. The percentage of B was increased according to a linear gradient of 1%-31% in 7 min, kept at 31% for 5 min, increased from 38% to 48% in 27 min

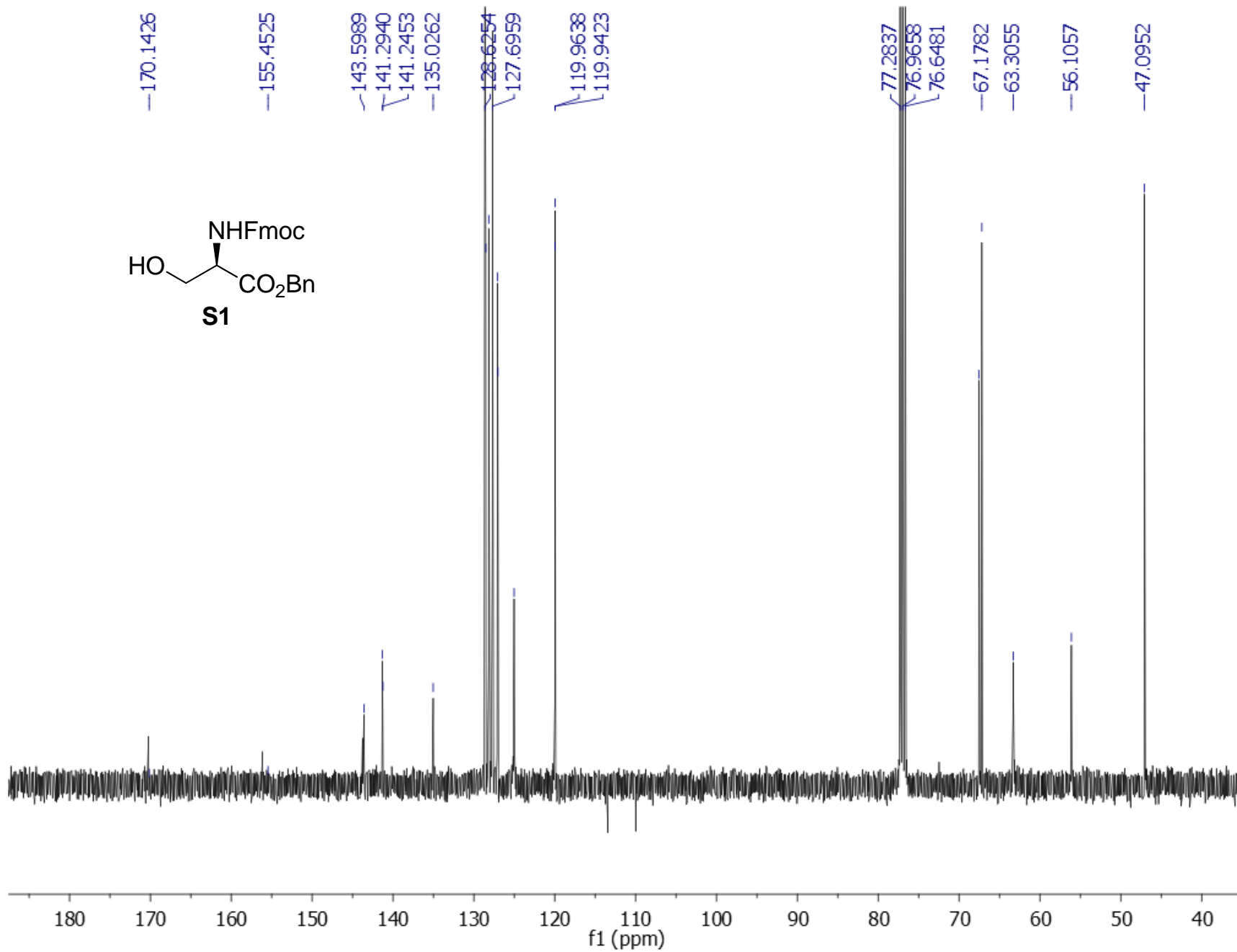
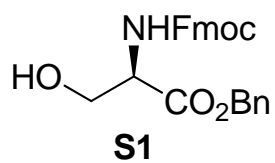
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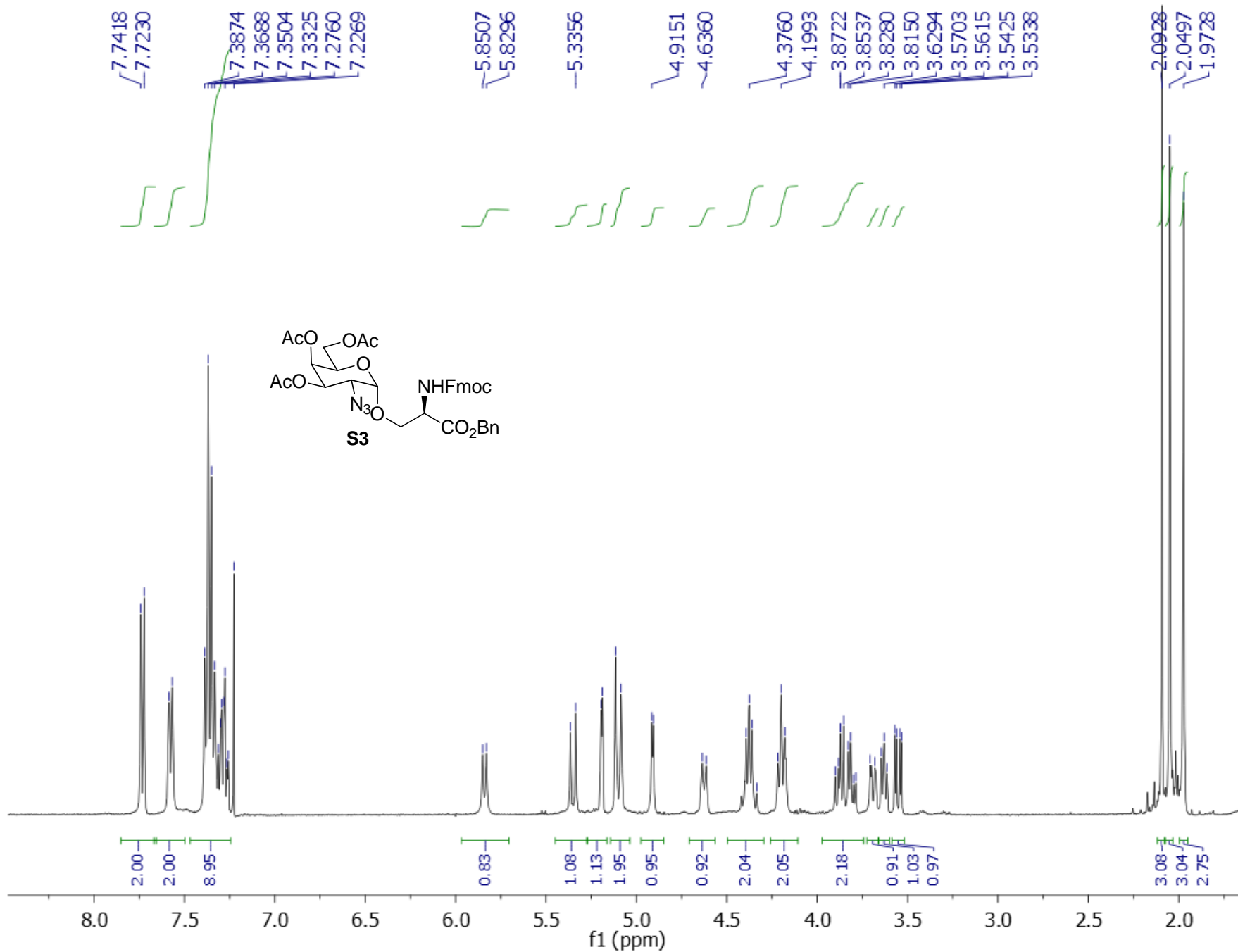
- (1) Kuduk, S. D.; Schwarz, J. B.; Chen, X.-T.; Glunz, P. W.; Sames, D.; Ragupathi, G.; Livingston, P. O.; Danishefsky, S. J. *J. Am. Chem. Soc.* **1998**, *120*, 12474.
- (2) Montero, J. L.; Winum, J. Y.; Leydet, A.; Kamal, M.; Pavia, A. A.; Roque, J. P. *Carbohydrate Research* **1997**, *297*, 175.
- (3) Arsequell, G.; Krippner, L.; Dwek, R. A.; Wong, S. Y. C. *Journal of the Chemical Society-Chemical Communications* **1994**, 2383.
- (4) Kay, C.; Lorthioir, O. E.; Parr, N. J.; Congreve, M.; McKeown, S. C.; Scicinski, J. J.; Ley, S. V. *Biotechnol Bioeng* **2000**, *71*, 110.











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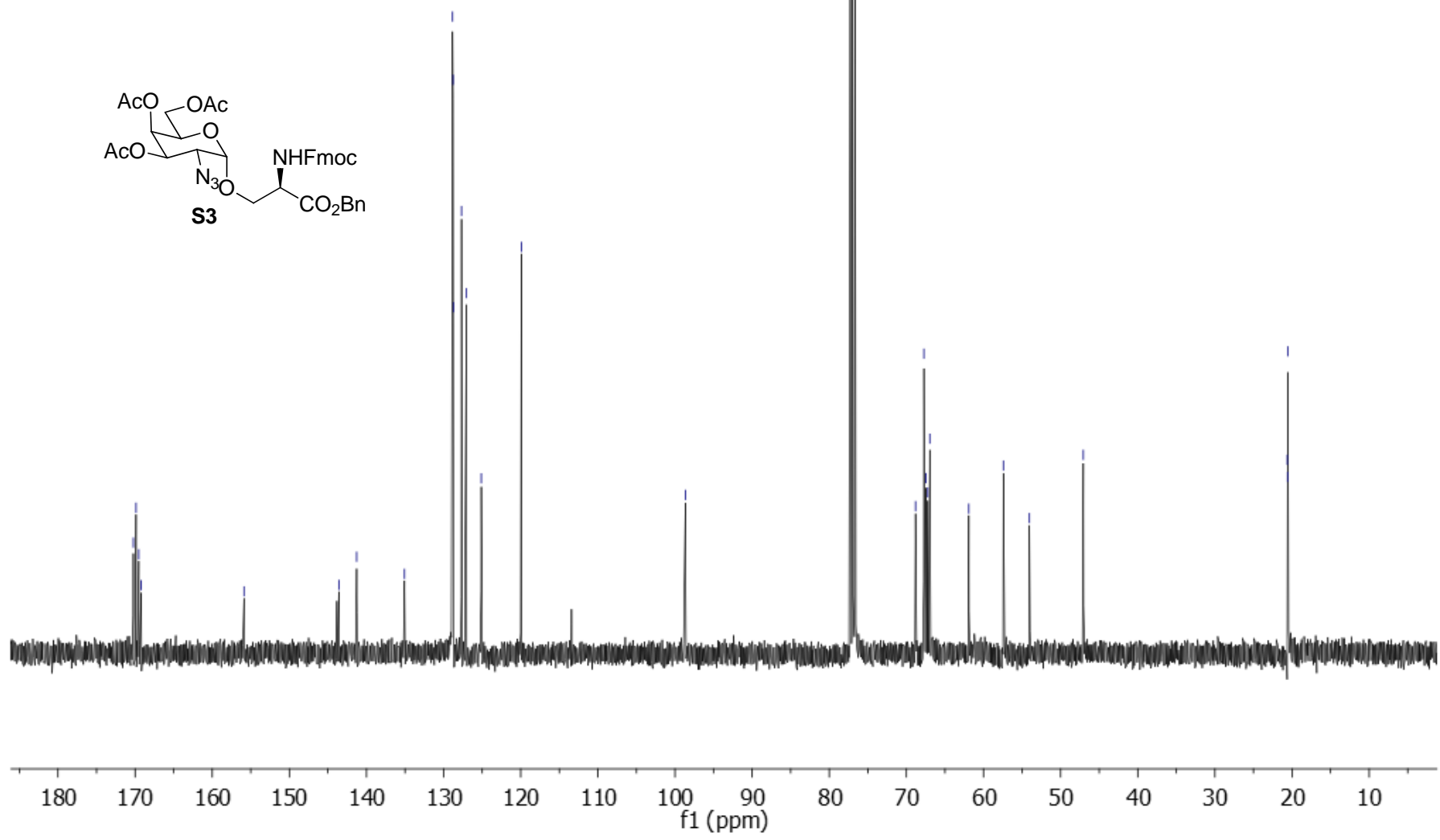
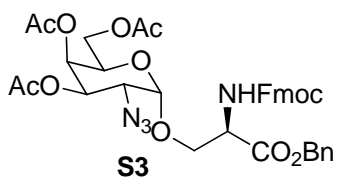
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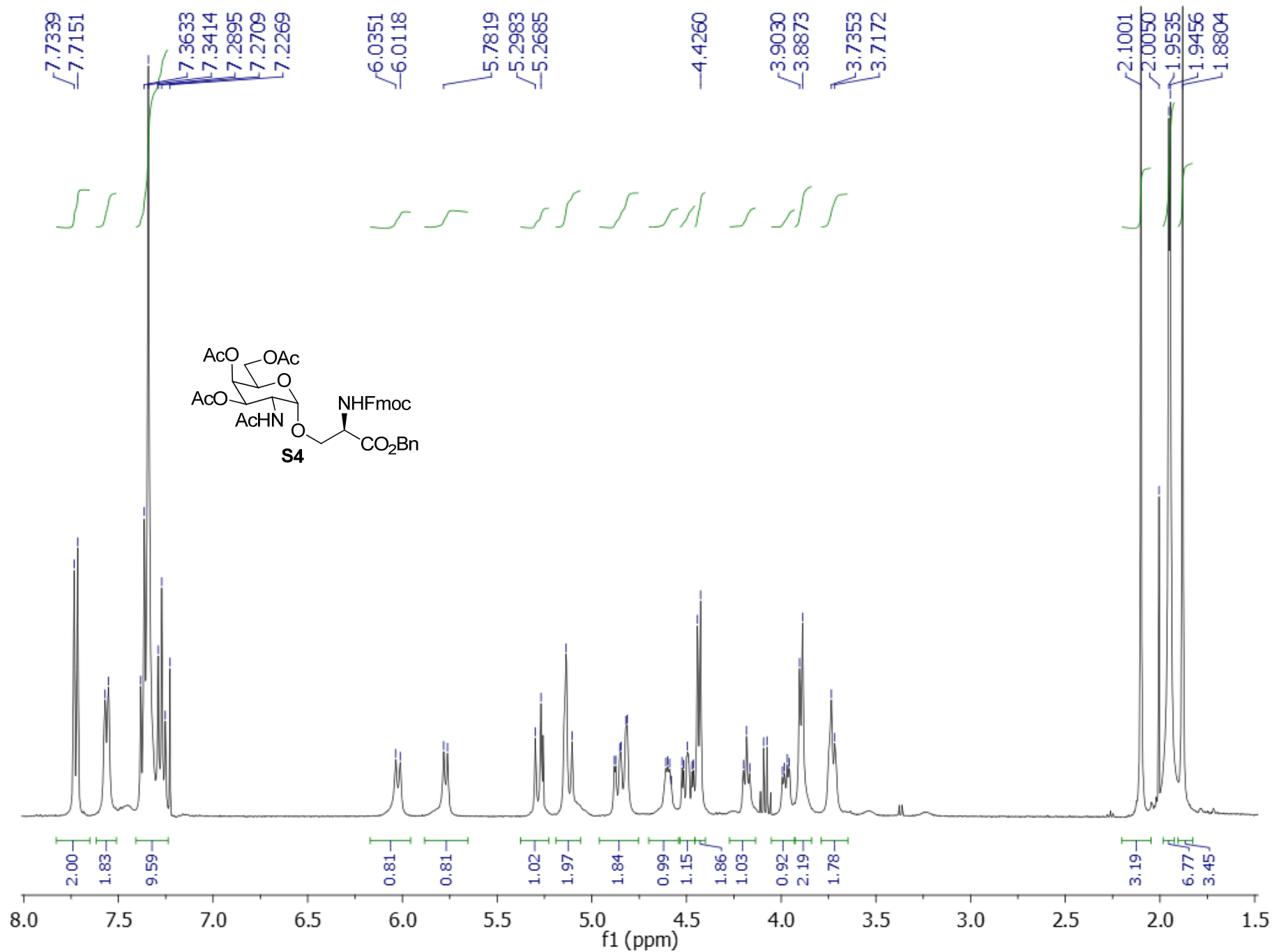
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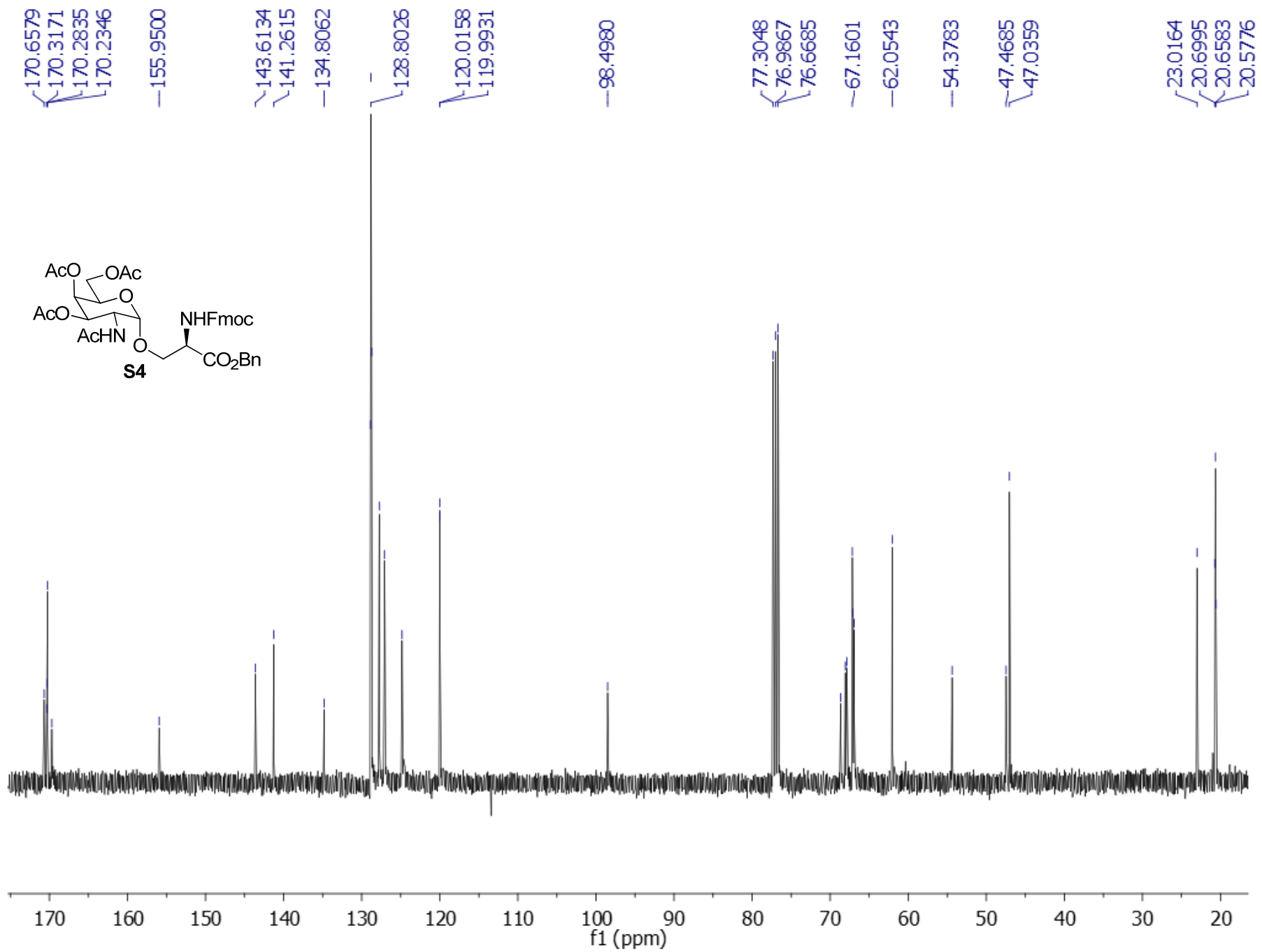
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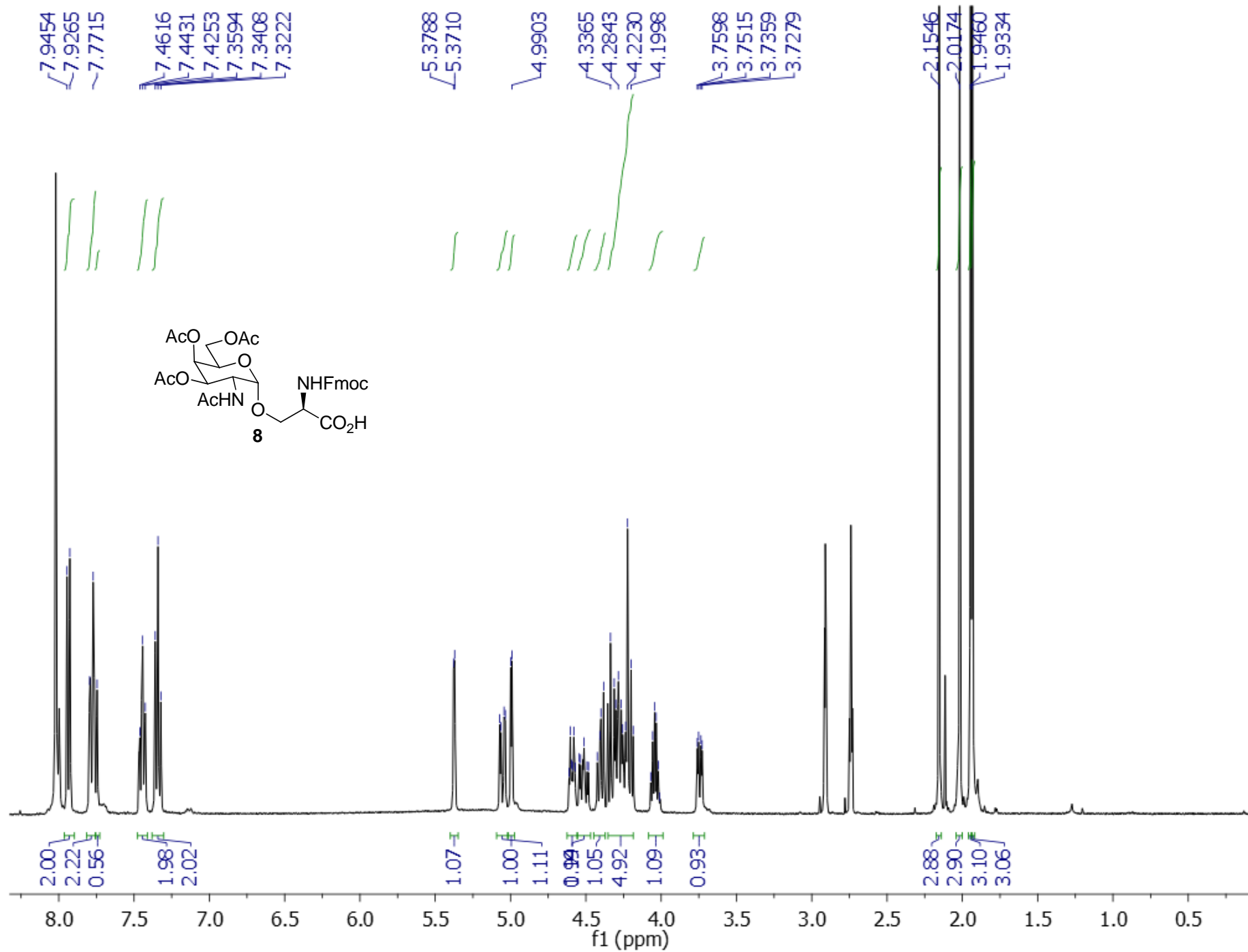
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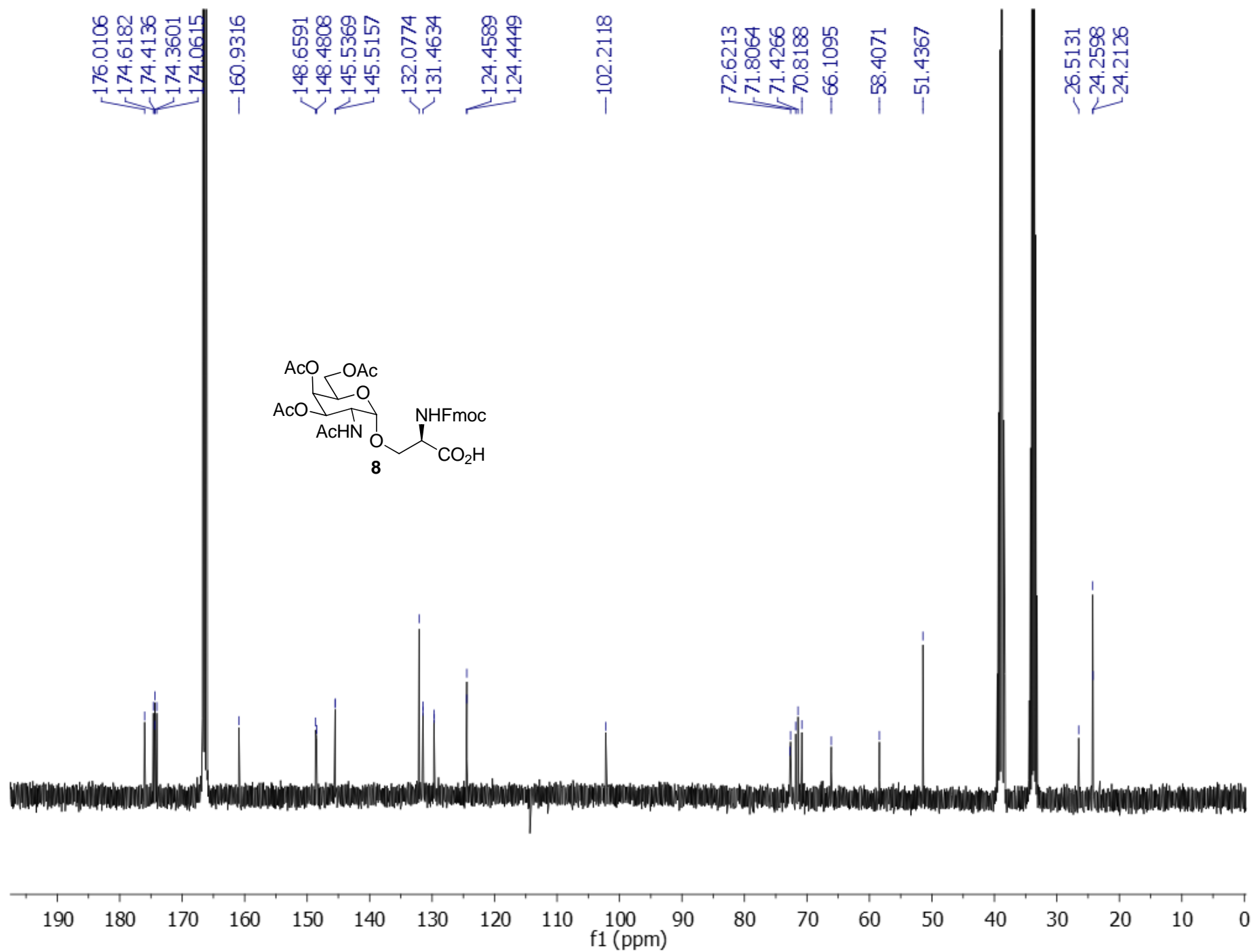
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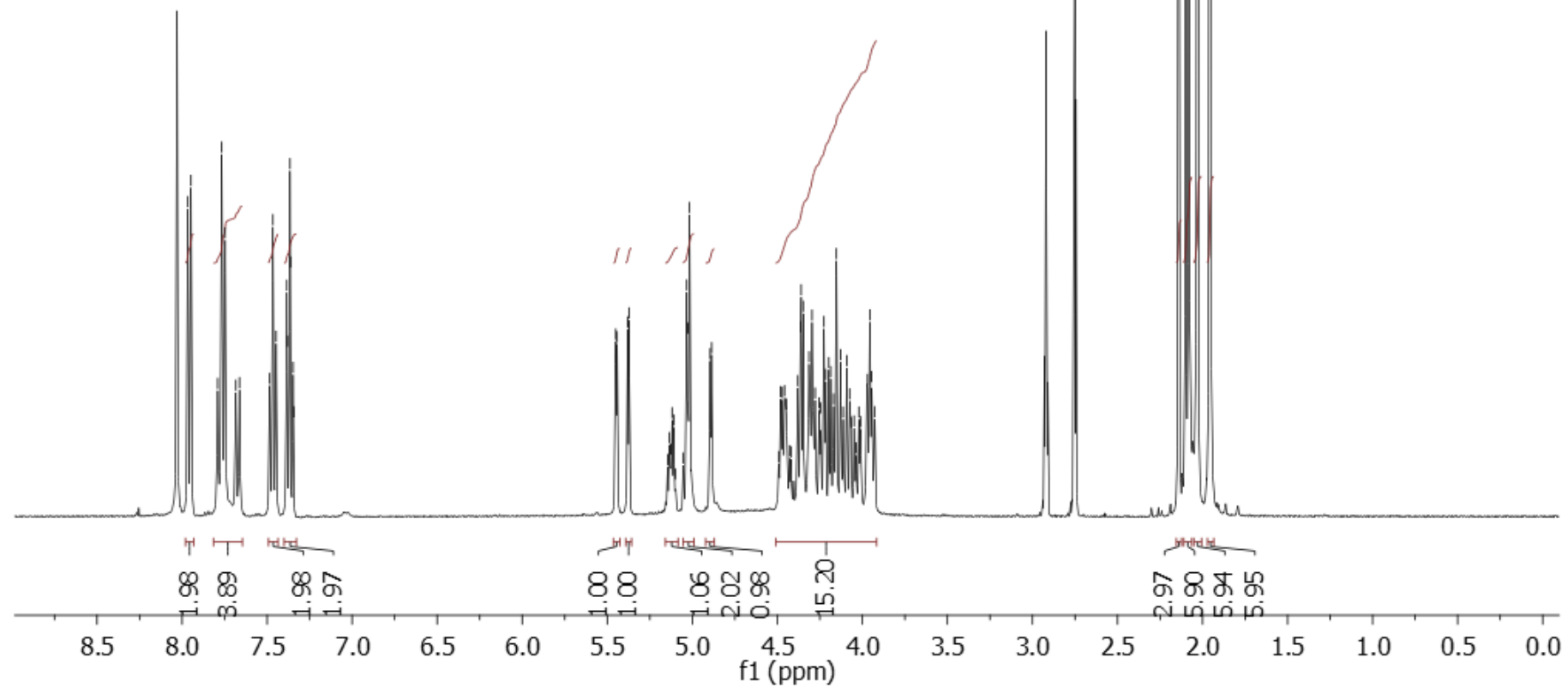
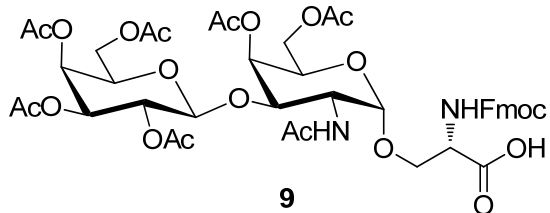
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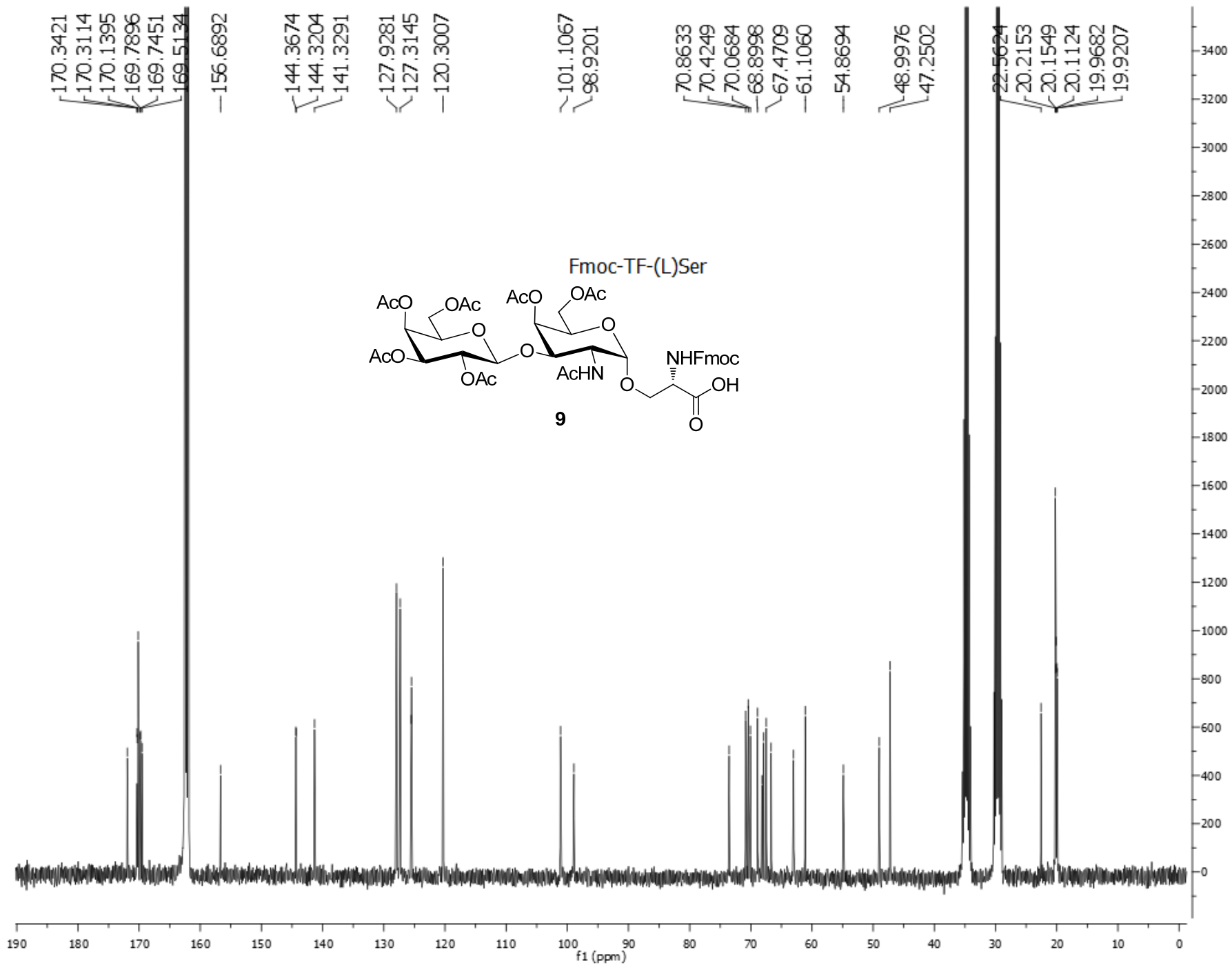
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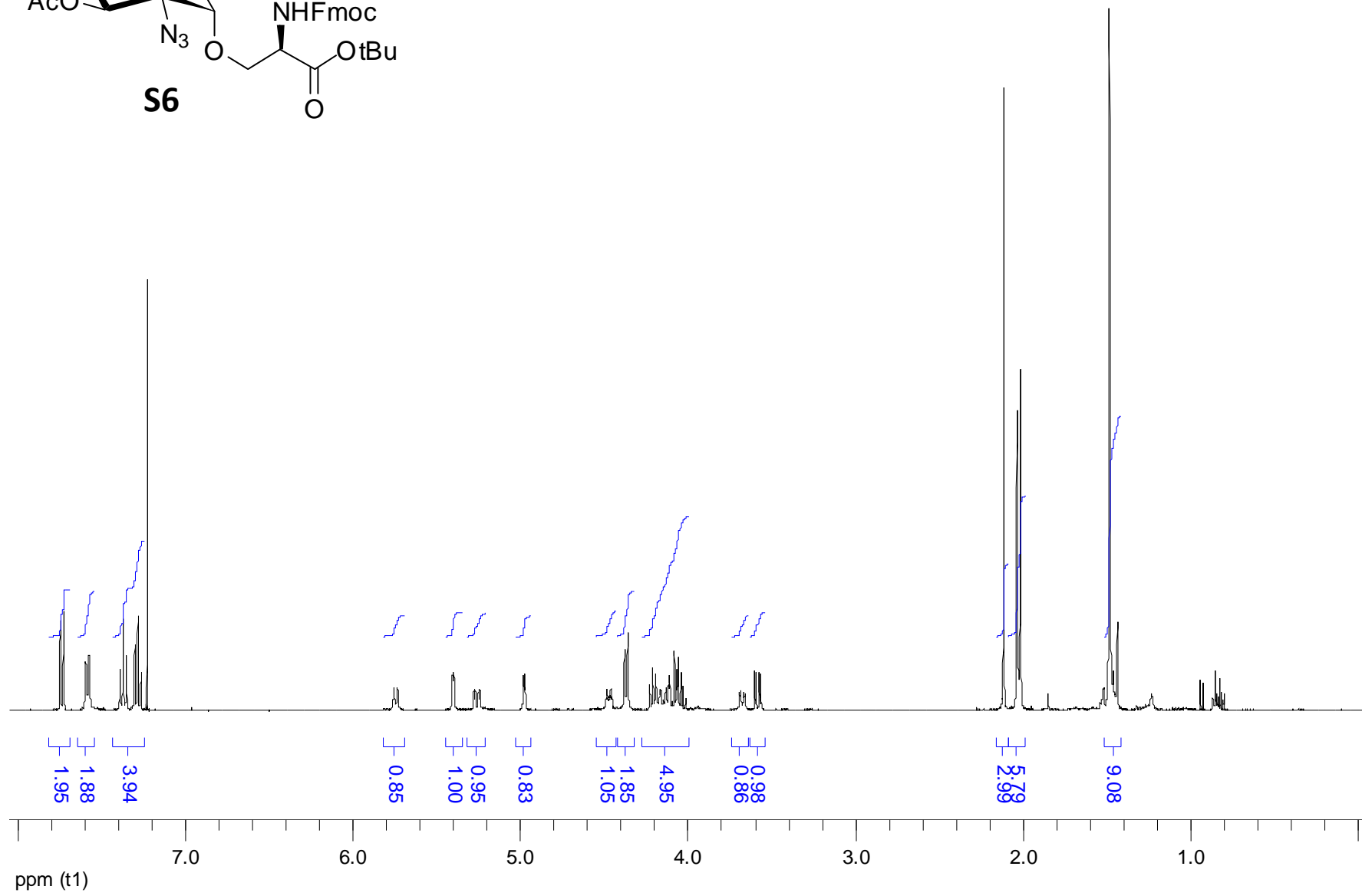
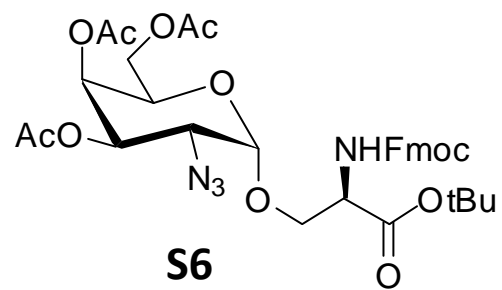
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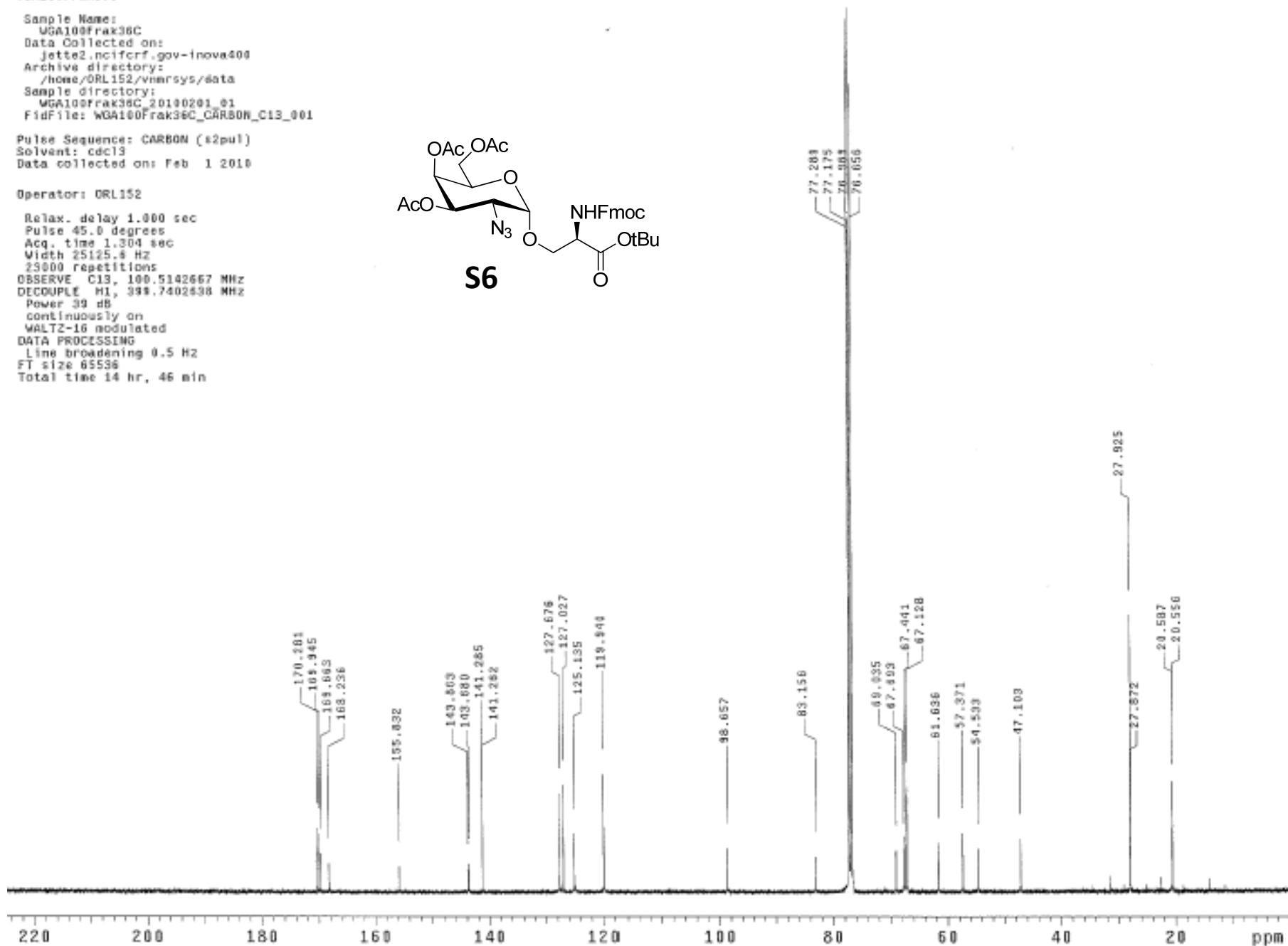
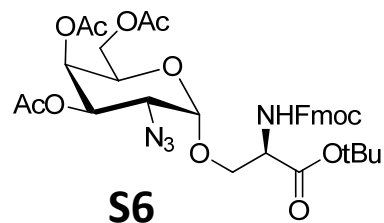
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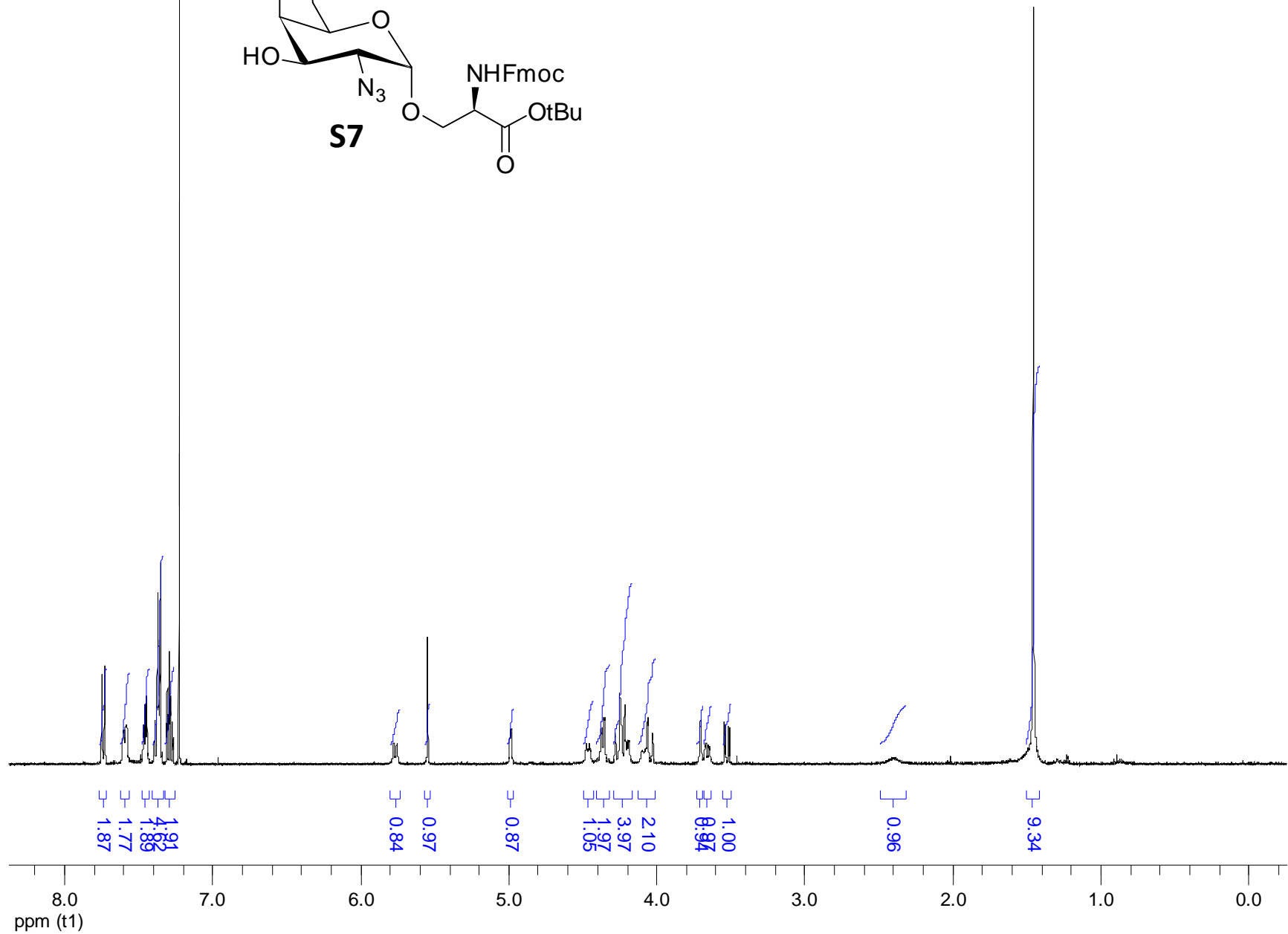
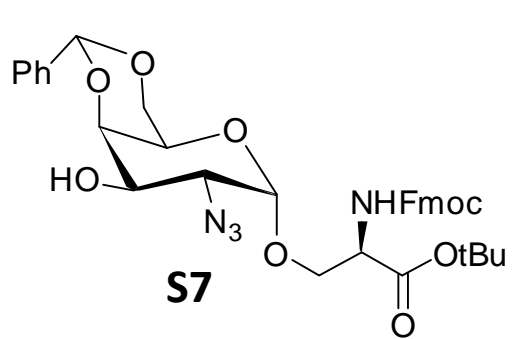
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Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.304 sec
Width 25125.8 Hz
23000 repetitions
OBSERVE C13, 100.5142667 MHz
DECOUPLE H1, 399.7402438 MHz
Power 39 dB
continuously on
WALTZ-16 modulated
DATA PROCESSING
Line broadening 0.5 Hz
FT size 65536
Total time 14 hr, 46 min





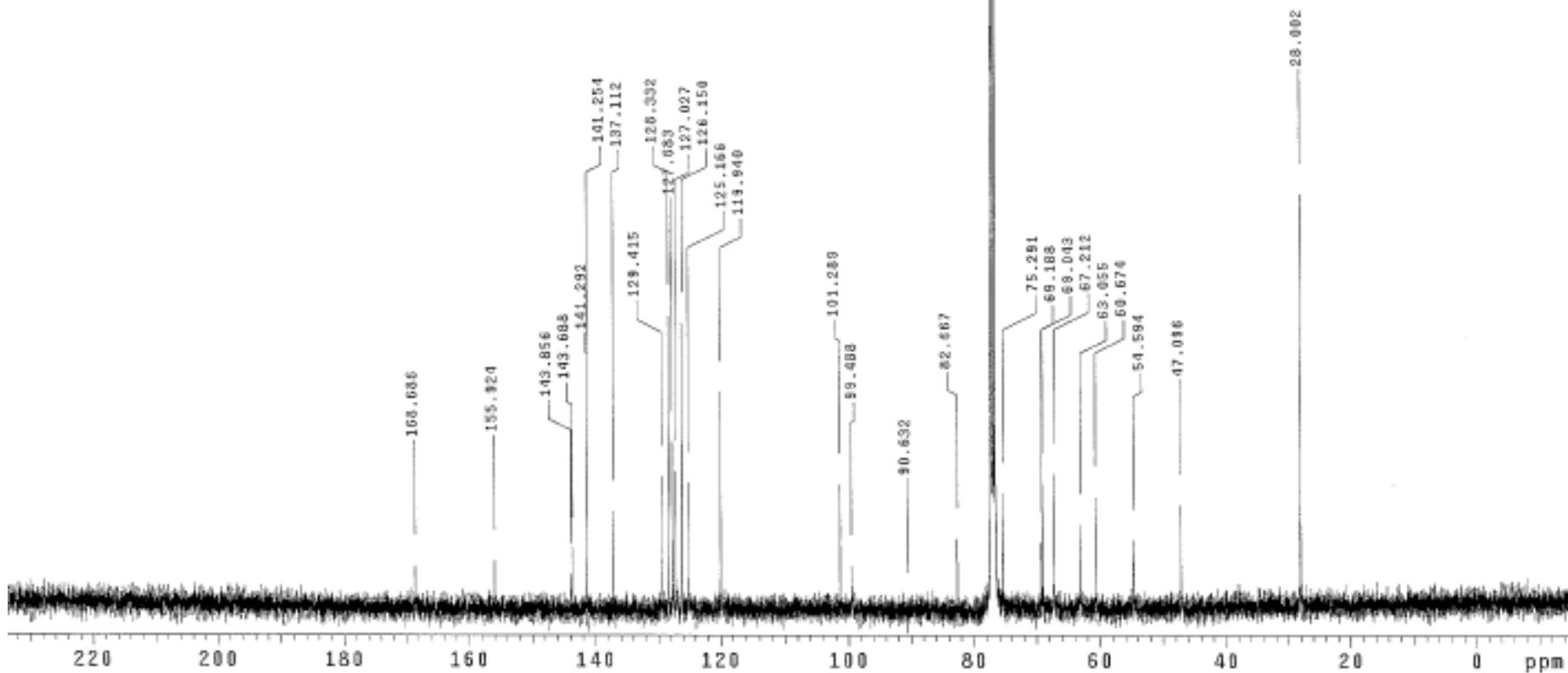
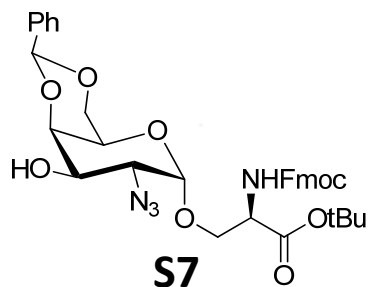
152F038C

Sample Name:
152F038C
Data Collected on:
jette2.ncifcrf.gov-inova400
Archive directory:
/home/ORL152/vnersys/data
Sample directory:
152F038C_20100325_01
FidFile: 152F038C_CARBON_C13_001

Pulse Sequence: CARBON (s2pu1)
Solvent: cdc13
Data collected on: Mar 25 2010

Operator: ORL152

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.304 sec
Width 25125.6 Hz
20000 repetitions
OBSERVE C13, 100.5142667 MHz
DECOUPLE H1, 399.7402630 MHz
Power 39 dB
continuously on
WALTZ-16 modulated
DATA PROCESSING
Line broadening 0.5 Hz
FT size 85536
Total time 12 hr, 51 min



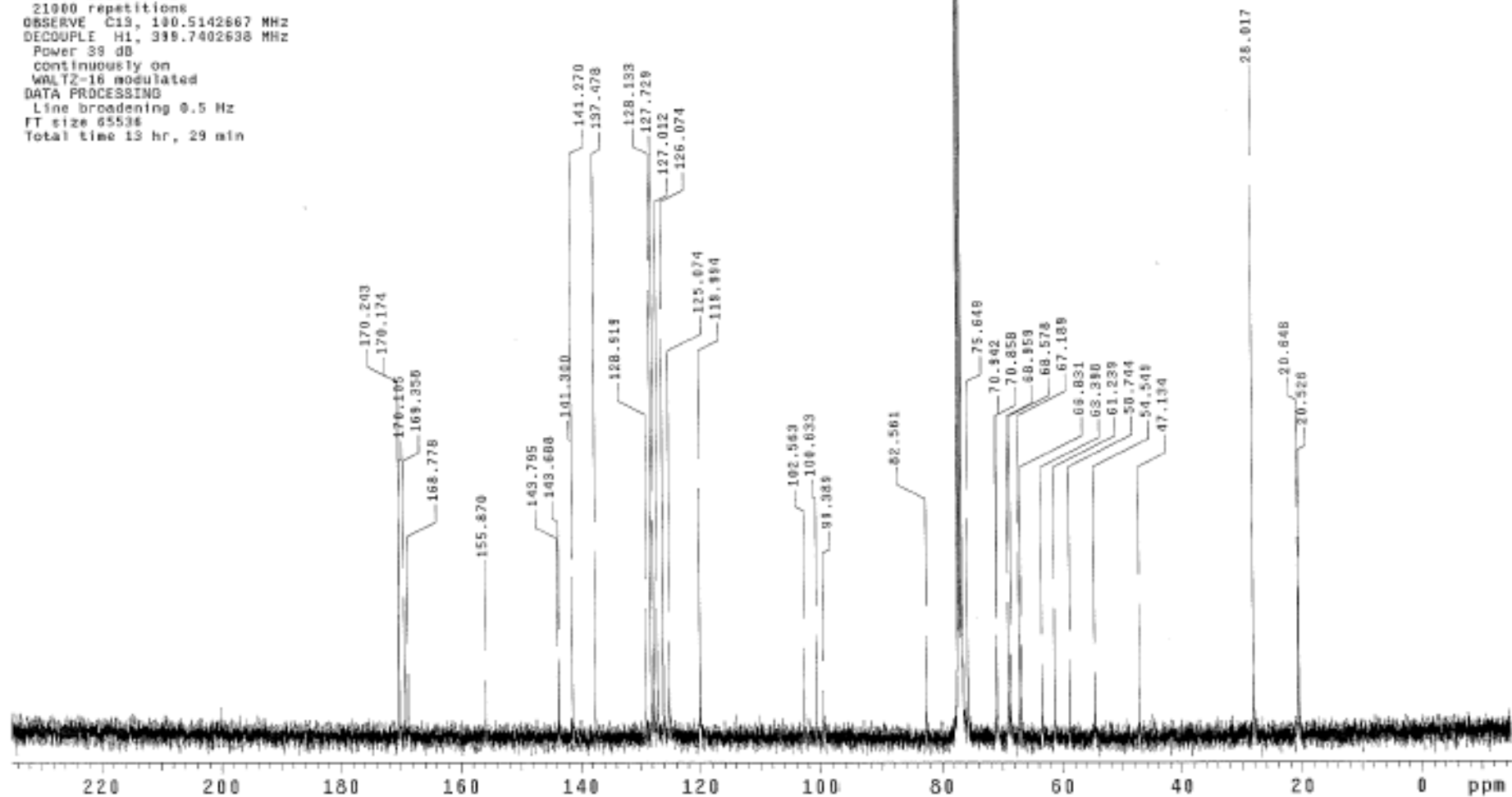
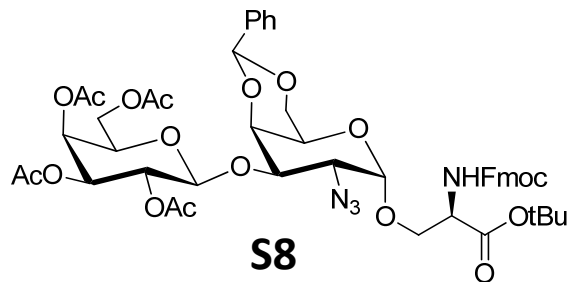
152F039C

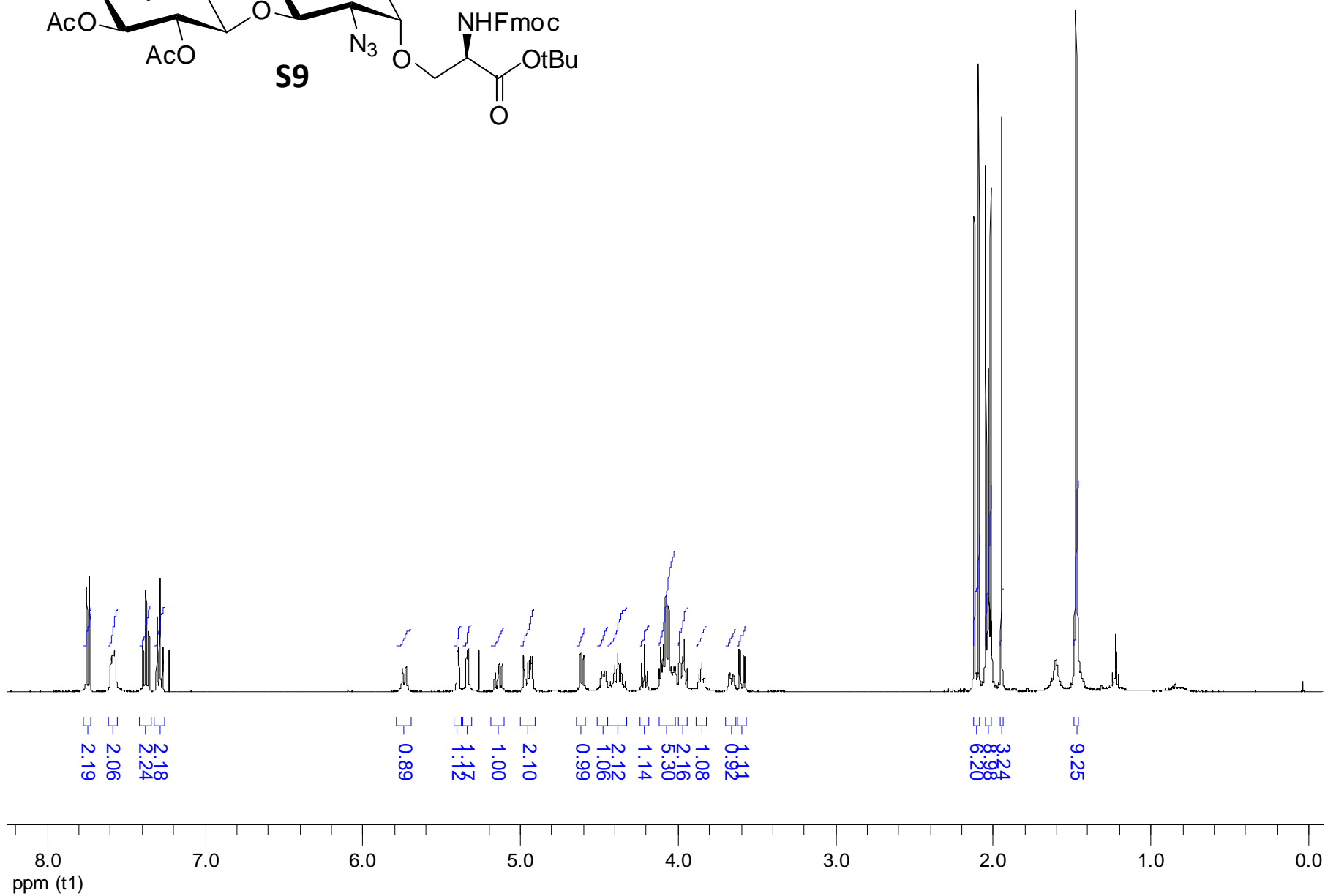
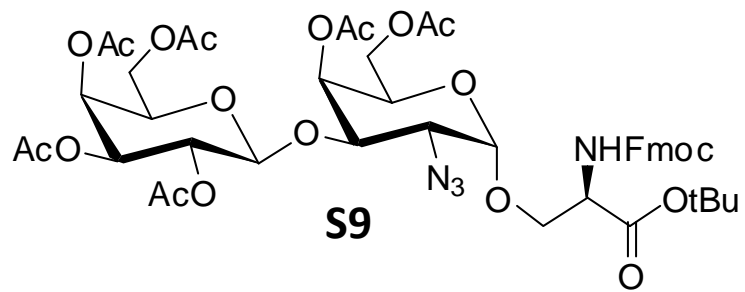
Sample Name:
152F039C
Data Collected on:
jette2.ncifcrf.gov-inova400
Archive directory:
/home/ORL152/vnmrsys/data
Sample directory:
152F039C_20100421_02
FidFile: 152F039C_CARBON_C13_001

Pulse Sequence: CARBON (t2pu1)
Solvent: cdc13
Data collected on: Apr 21 2010

Operator: ORL152

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.304 sec
Width 25125.6 Hz
2100 repetitions
OBSERVE C13, 100.5142667 MHz
DECOUPLE H1, 399.7402638 MHz
Power 39 dB
continuously on
WALTZ-16 modulated
DATA PROCESSING
Line broadening 0.5 Hz
FT size 65536
Total time 13 hr, 29 min





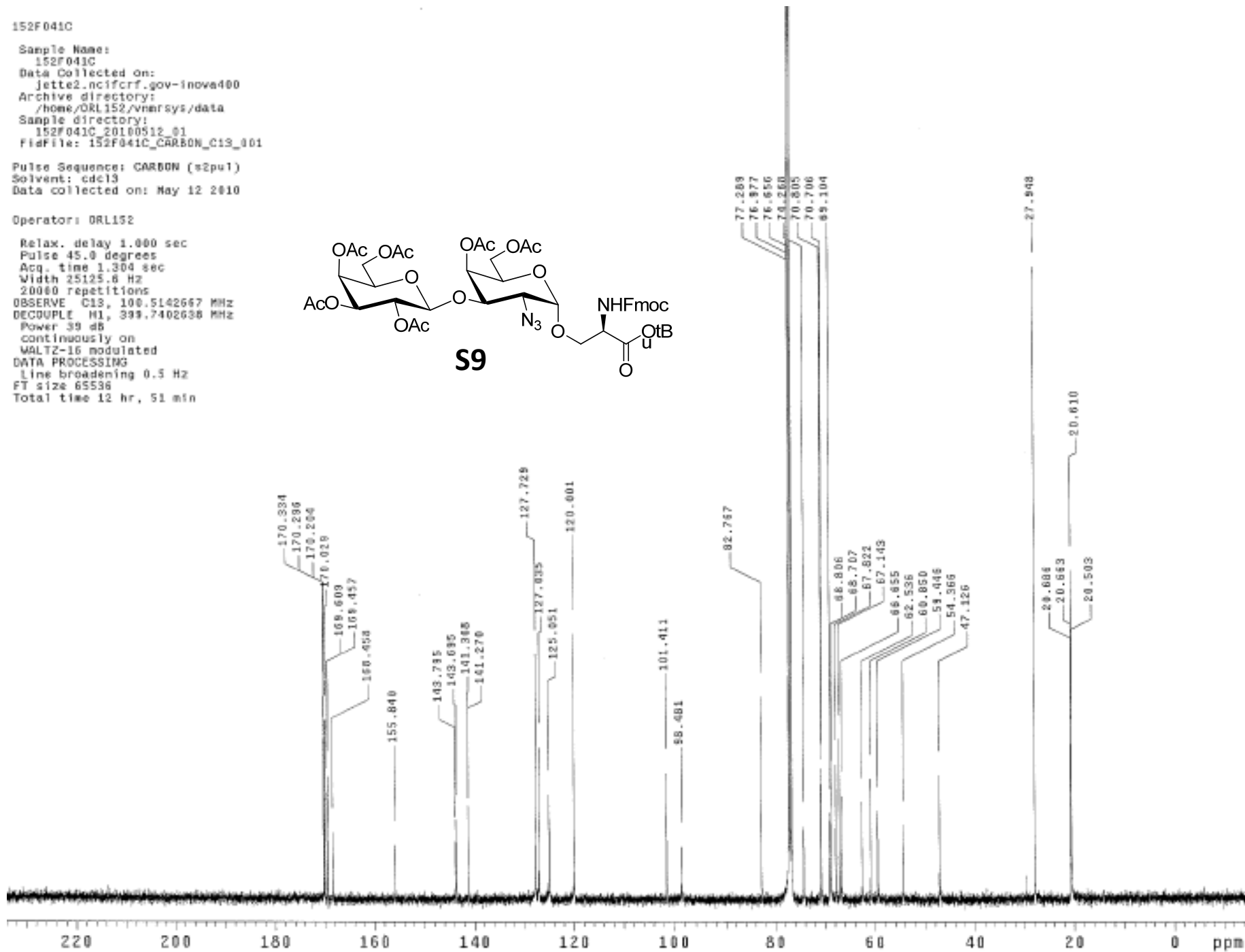
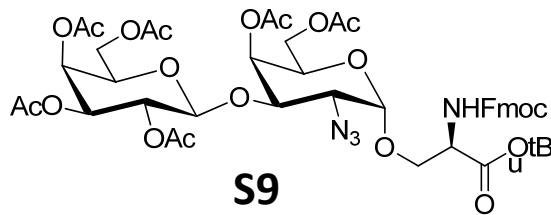
152F041C

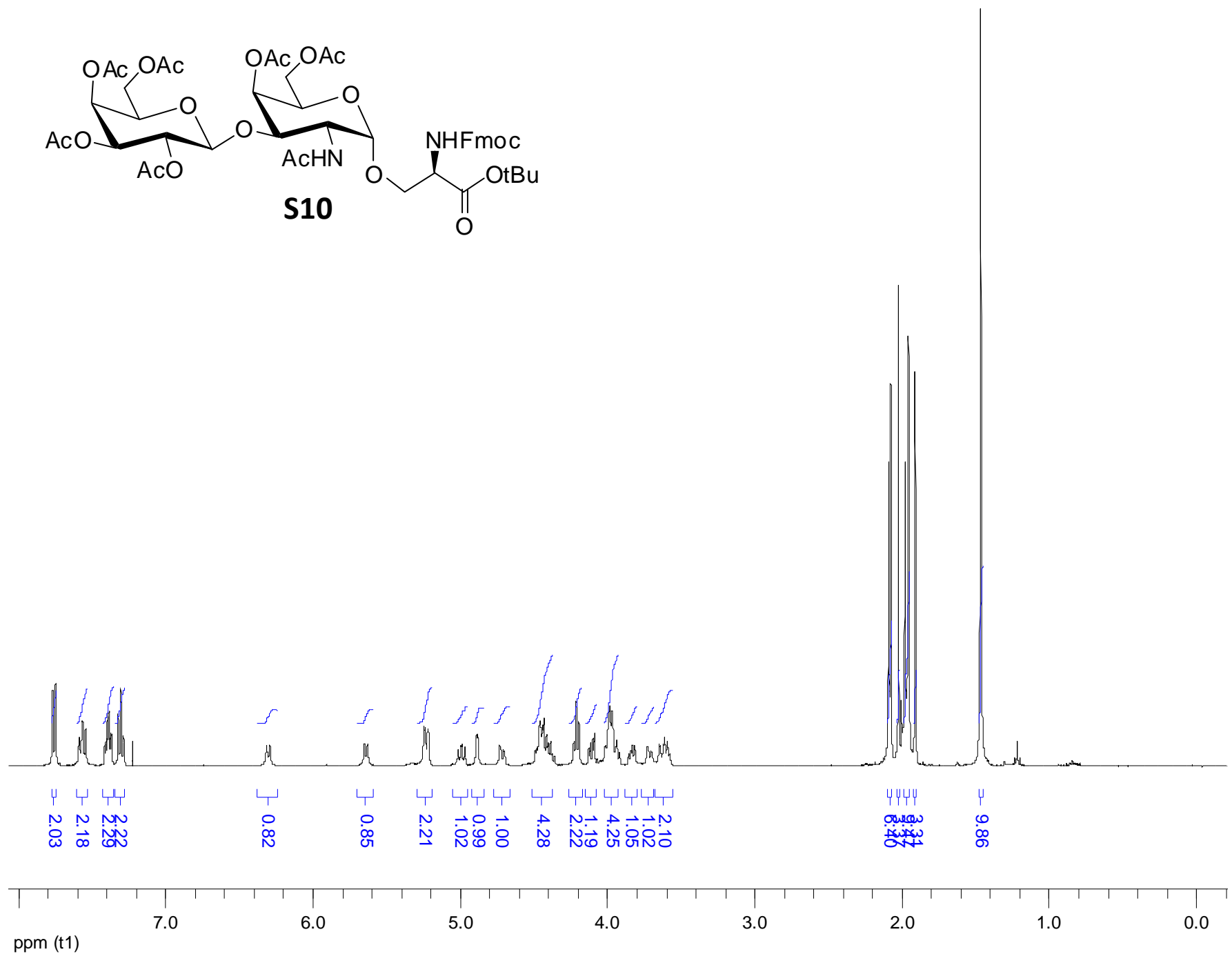
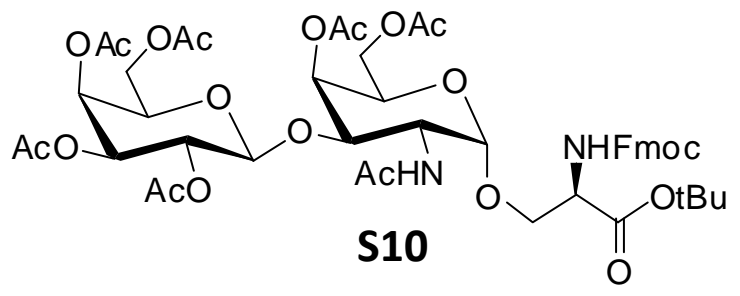
Sample Name:
152F041C
Data Collected on:
jette2.ncifcrf.gov-inova400
Archive directory:
/home/ORL152/vnersys/data
Sample directory:
152F041C_20100512_01
FidFile: 152F041C_CARBON_C13_001

Pulse Sequence: CARBON (s2pu1)
Solvent: cdcl3
Data collected on: May 12 2010

Operator: ORL152

Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.304 sec
Width 25125.6 Hz
20000 repetitions
OBSERVE C13, 100.5142667 MHz
DECOUPLE H1, 399.7402630 MHz
Power 39 dB
continuously on
WALTZ-16 modulated
DATA PROCESSING
Line broadening 0.5 Hz
FT size 65536
Total time 12 hr, 51 min





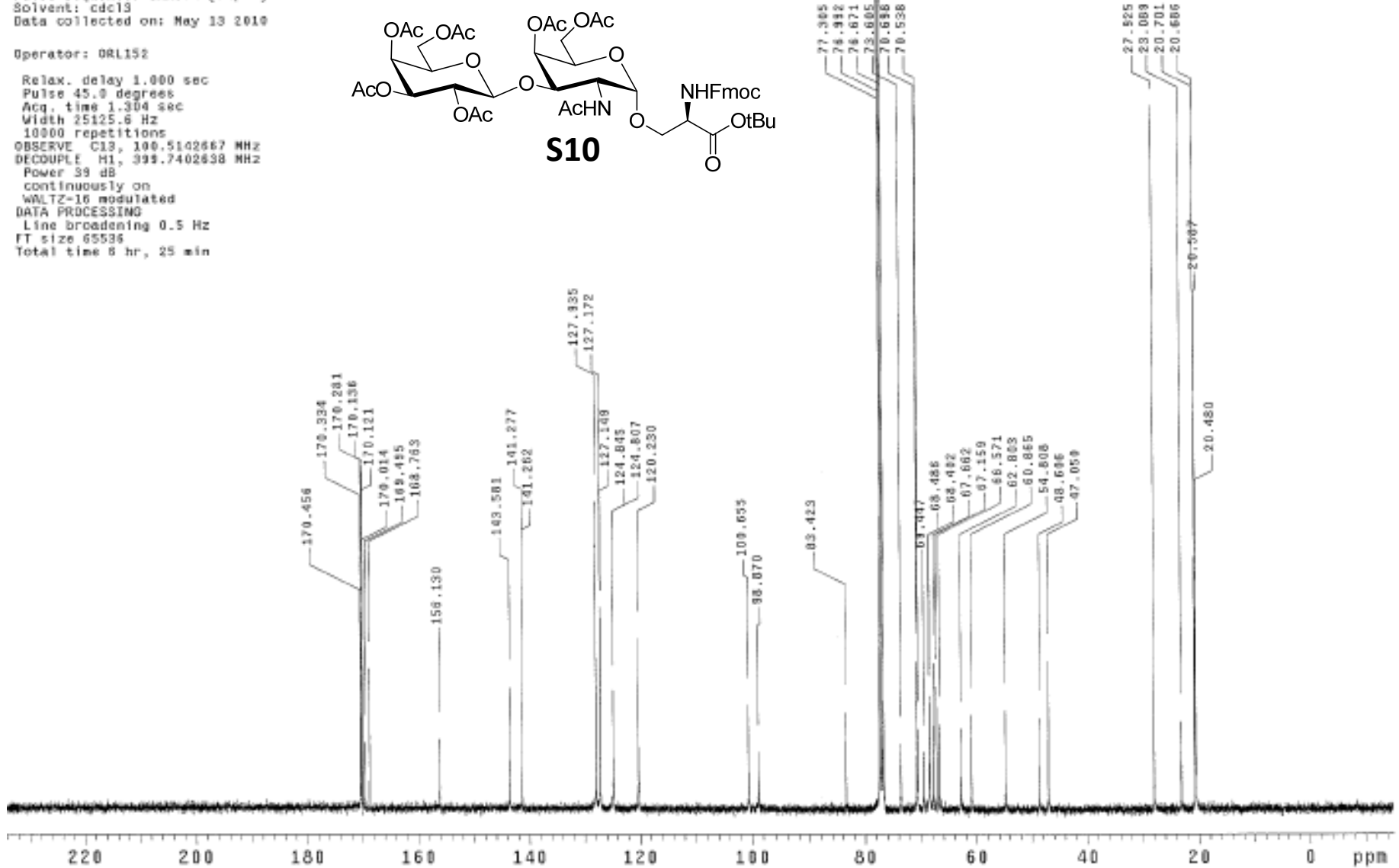
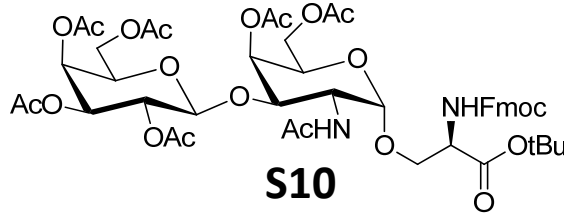
152F042C

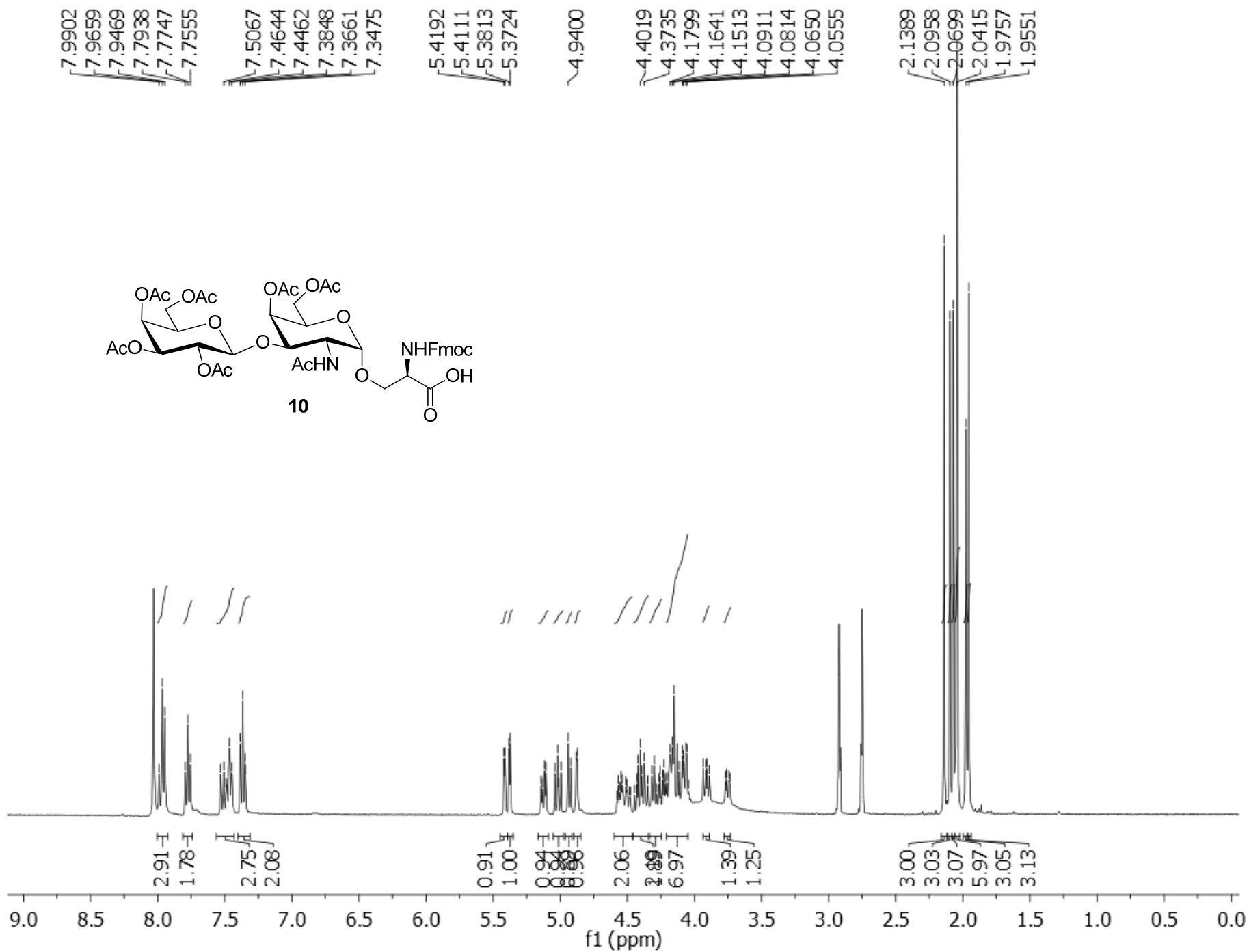
Sample Name:
152F042C
Data Collected on:
jette2.ncifcrf.gov-inova400
Archive directory:
/home/ORL152/vnmrsys/data
Sample directory:
152F042C_20180513_01
FidFile: 152F042C_CARBON_C13_001

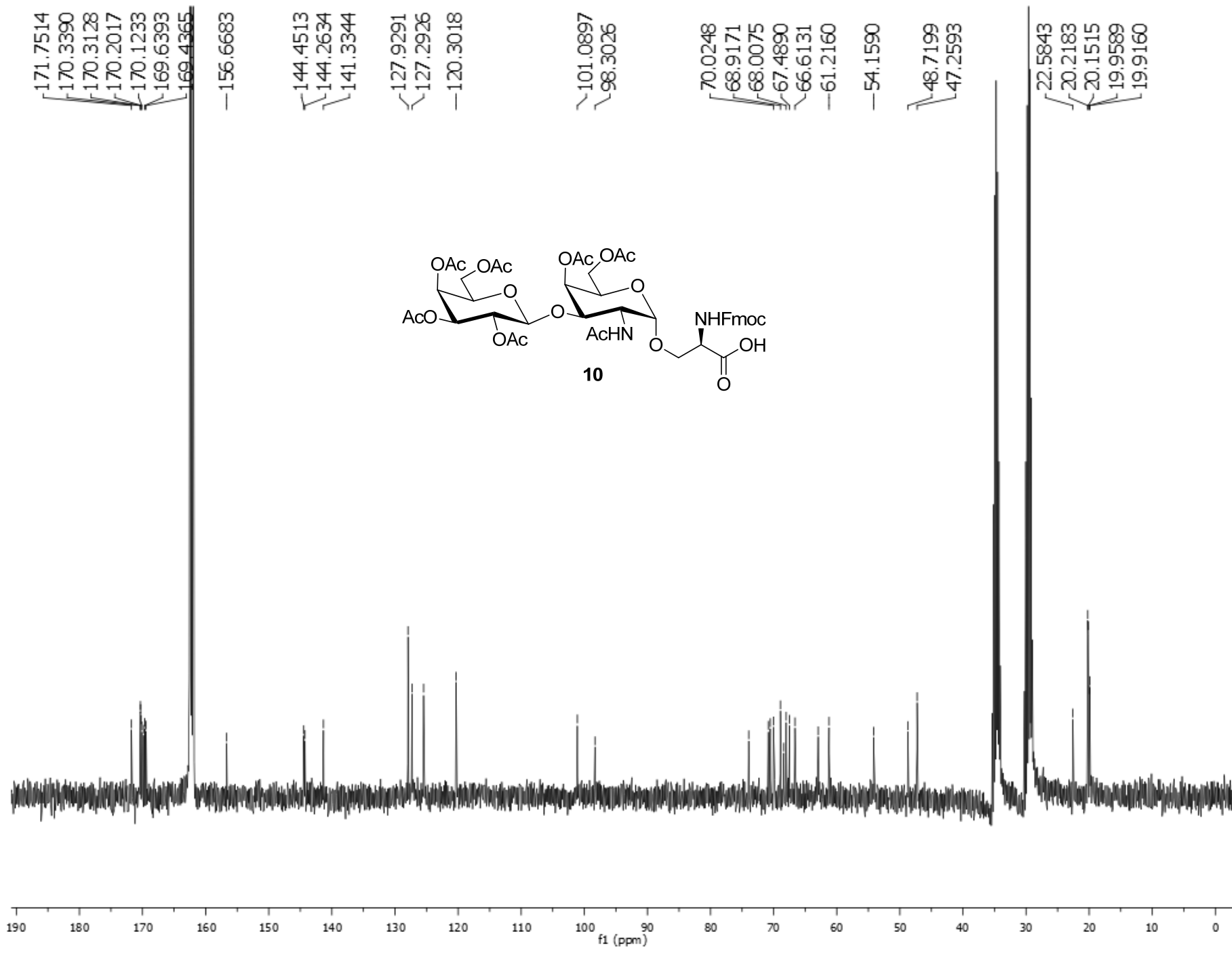
Pulse Sequence: CARBON (s2pul)
Solvent: cdc13
Data collected on: May 13 2018

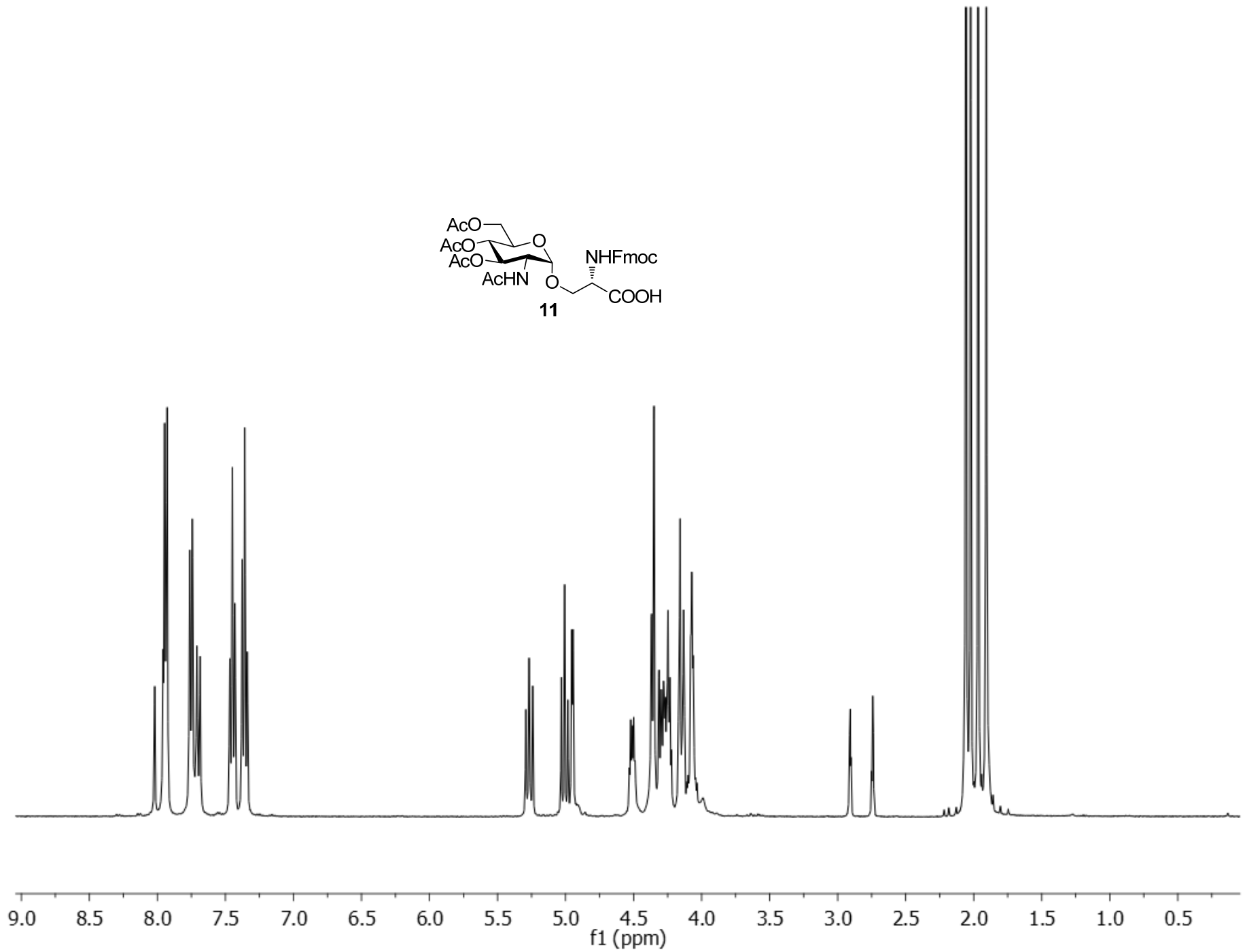
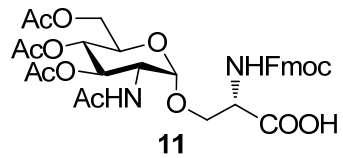
Operator: ORL152

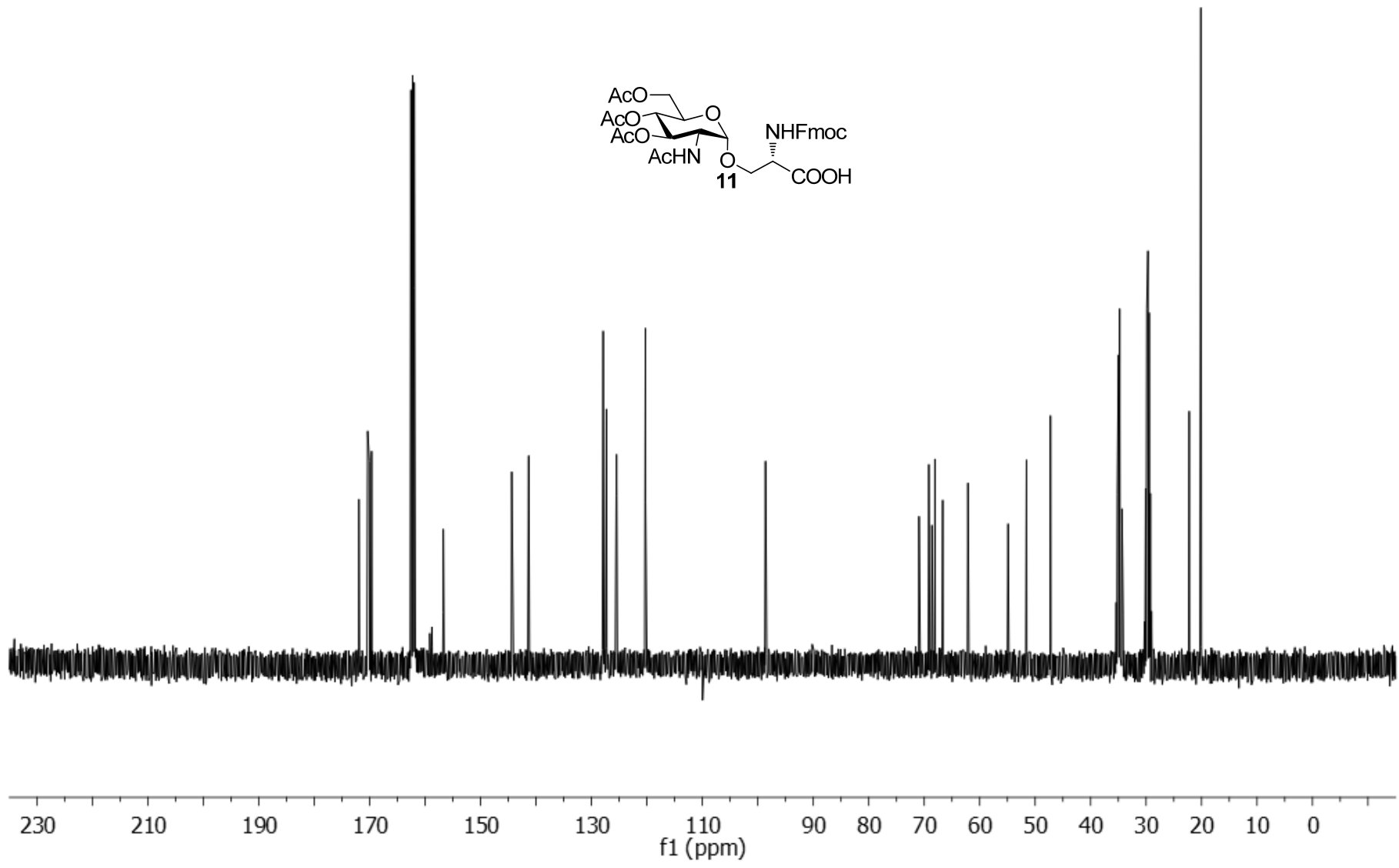
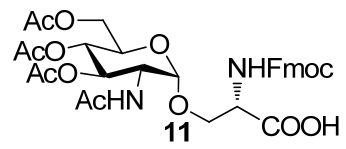
Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 1.304 sec
Width 25125.6 Hz
10900 repetitions
OBSERVE C13, 100.5102667 MHz
DECOUPLE H1, 399.7402638 MHz
Power 39 dB
continuously on
WALTZ-16 modulated
DATA PROCESSING
Line broadening 0.5 Hz
FT size 65536
Total time 8 hr, 25 min

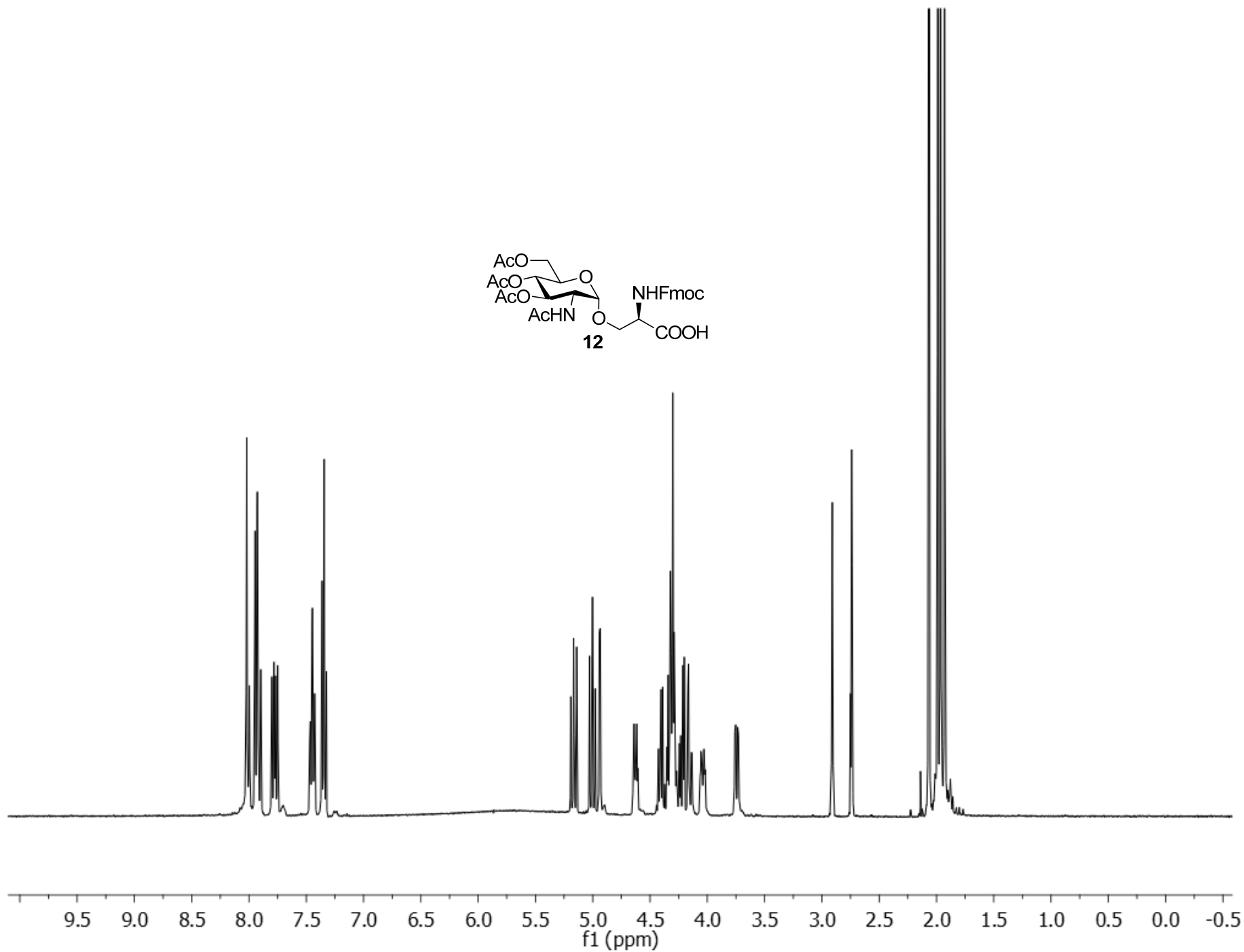
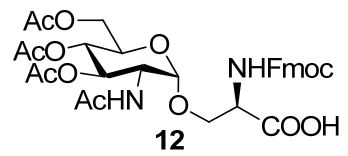


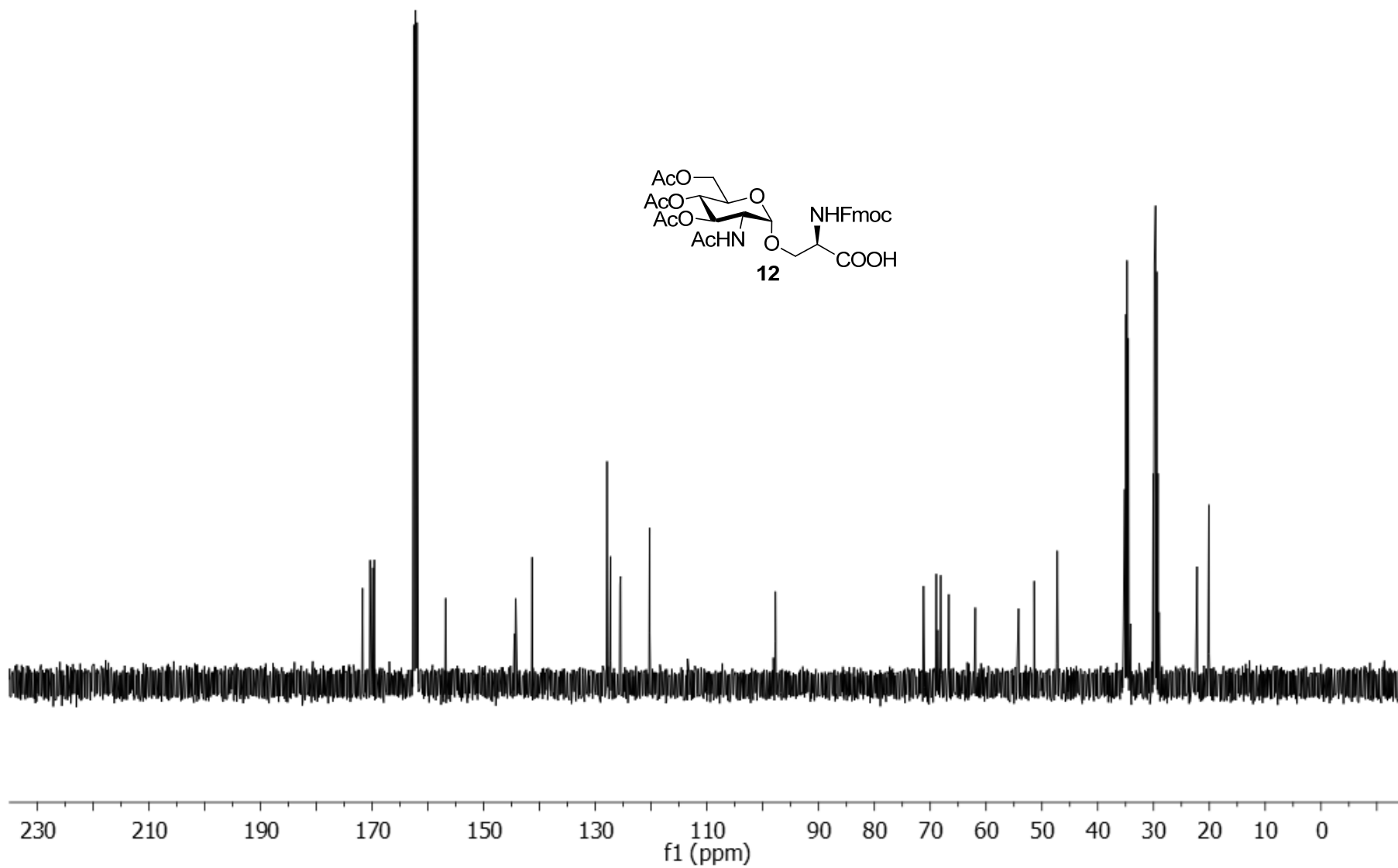
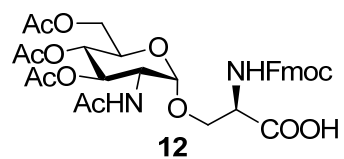


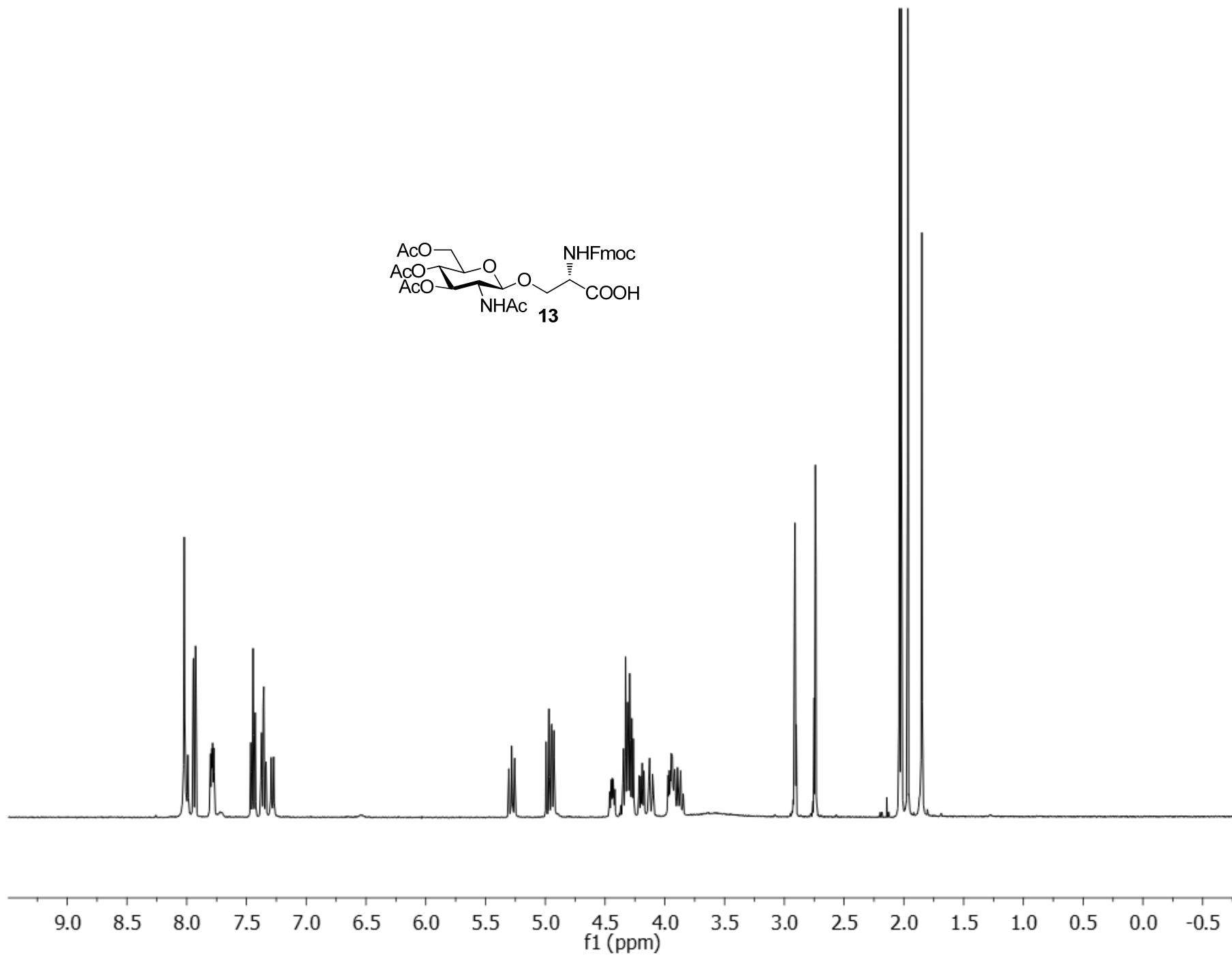
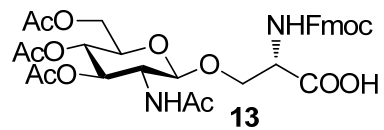


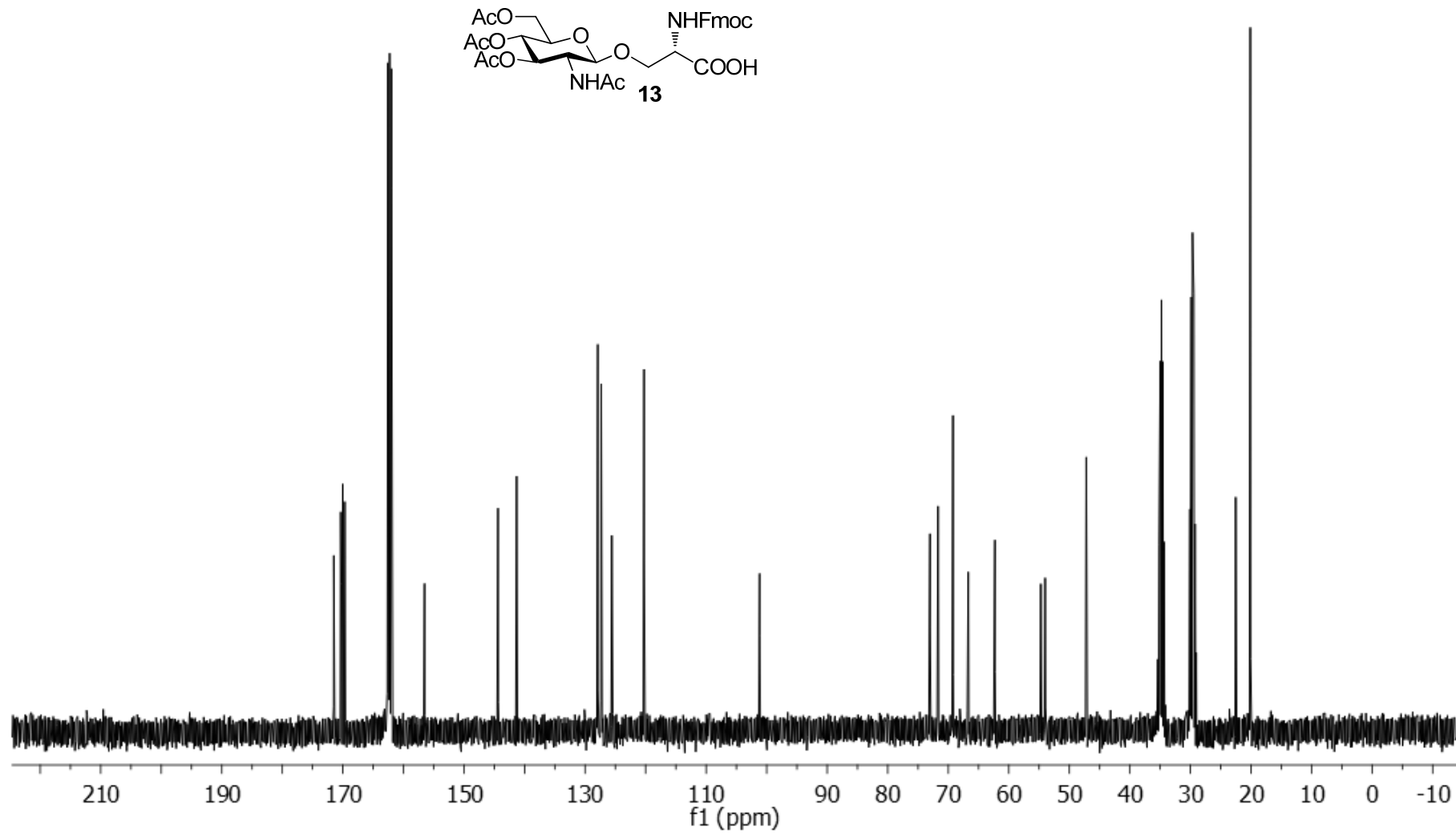
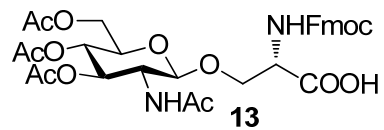


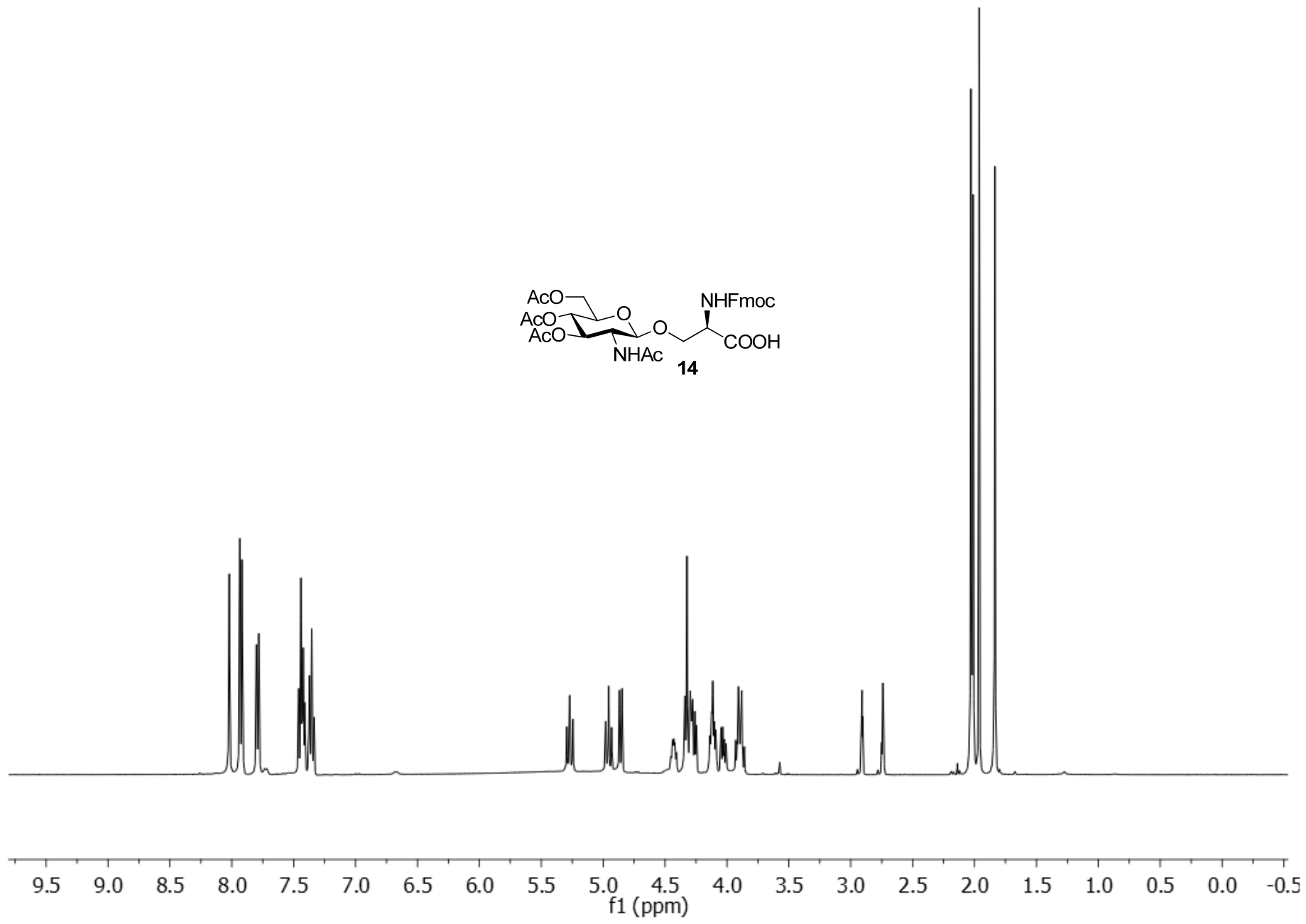
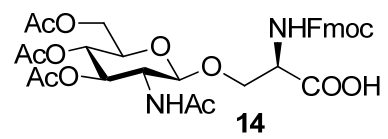


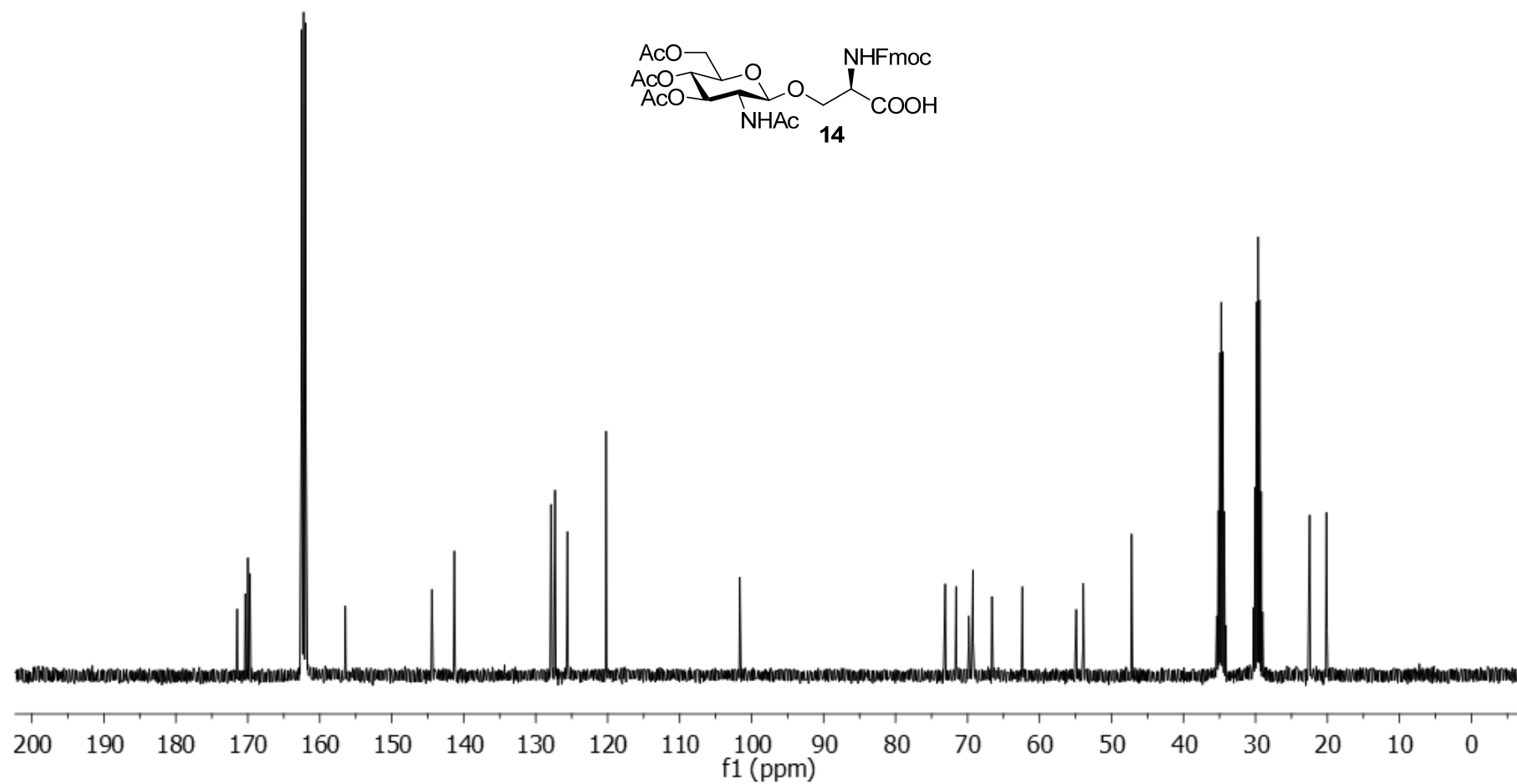
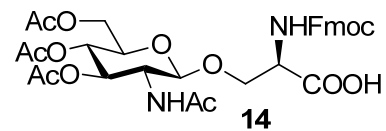


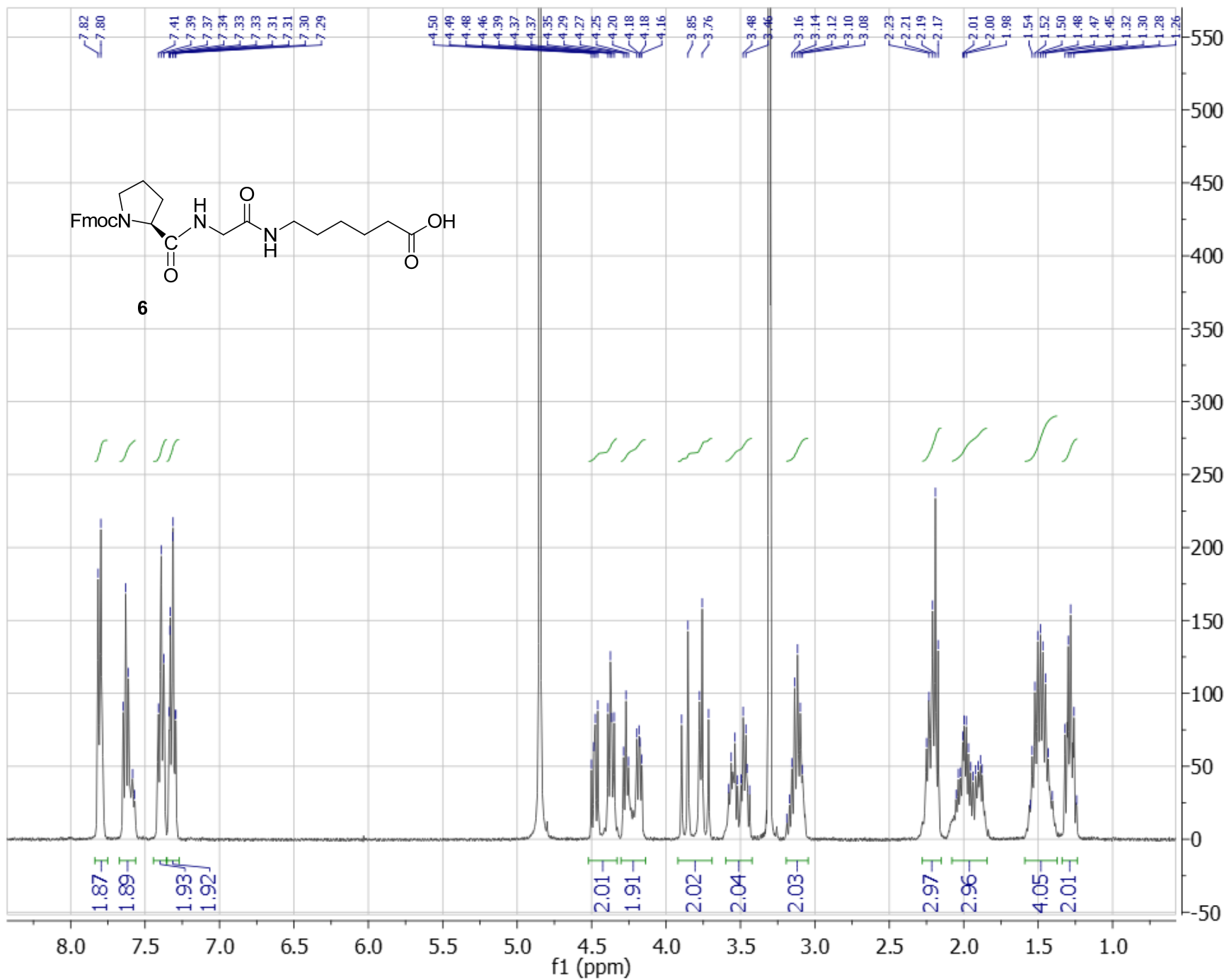


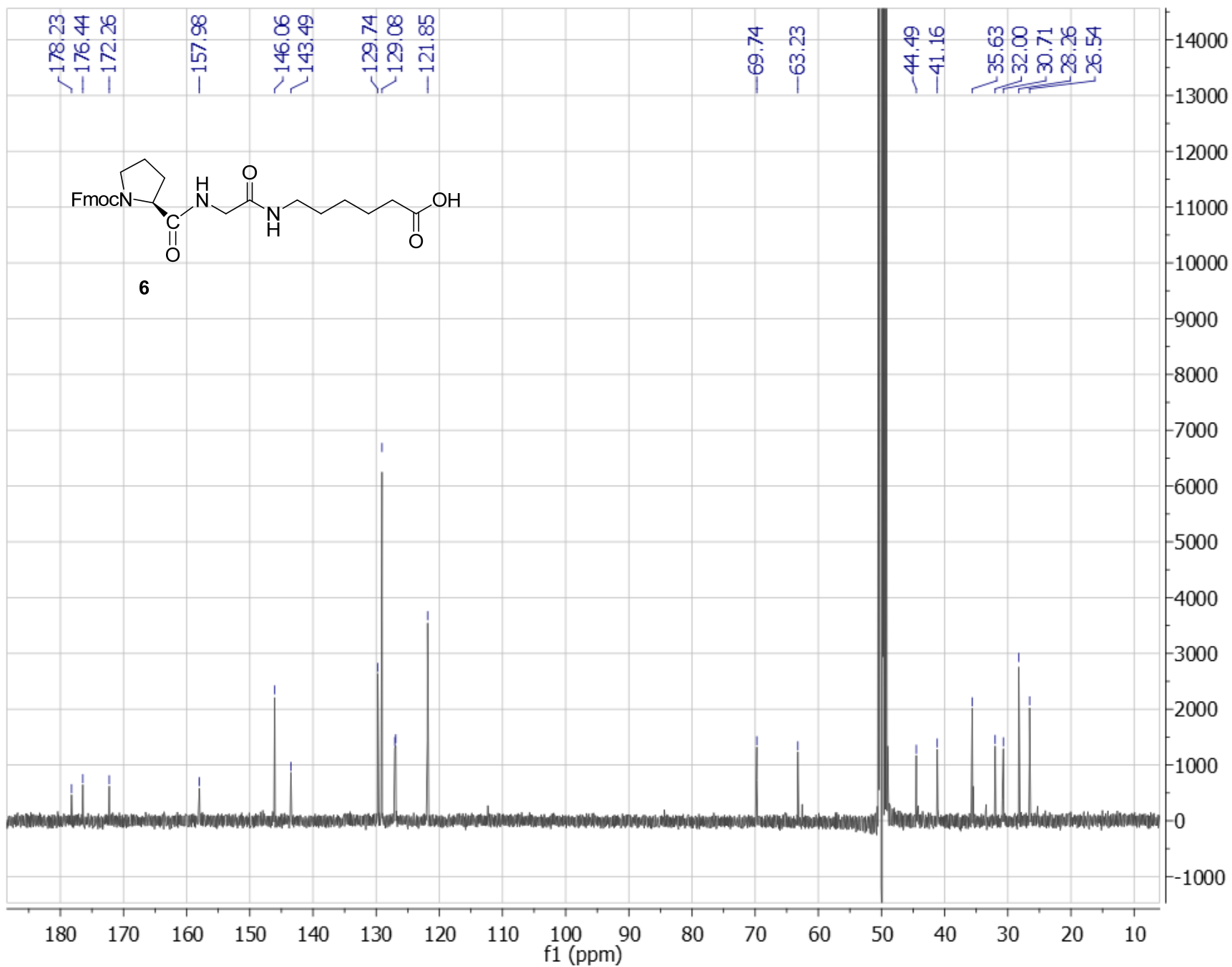


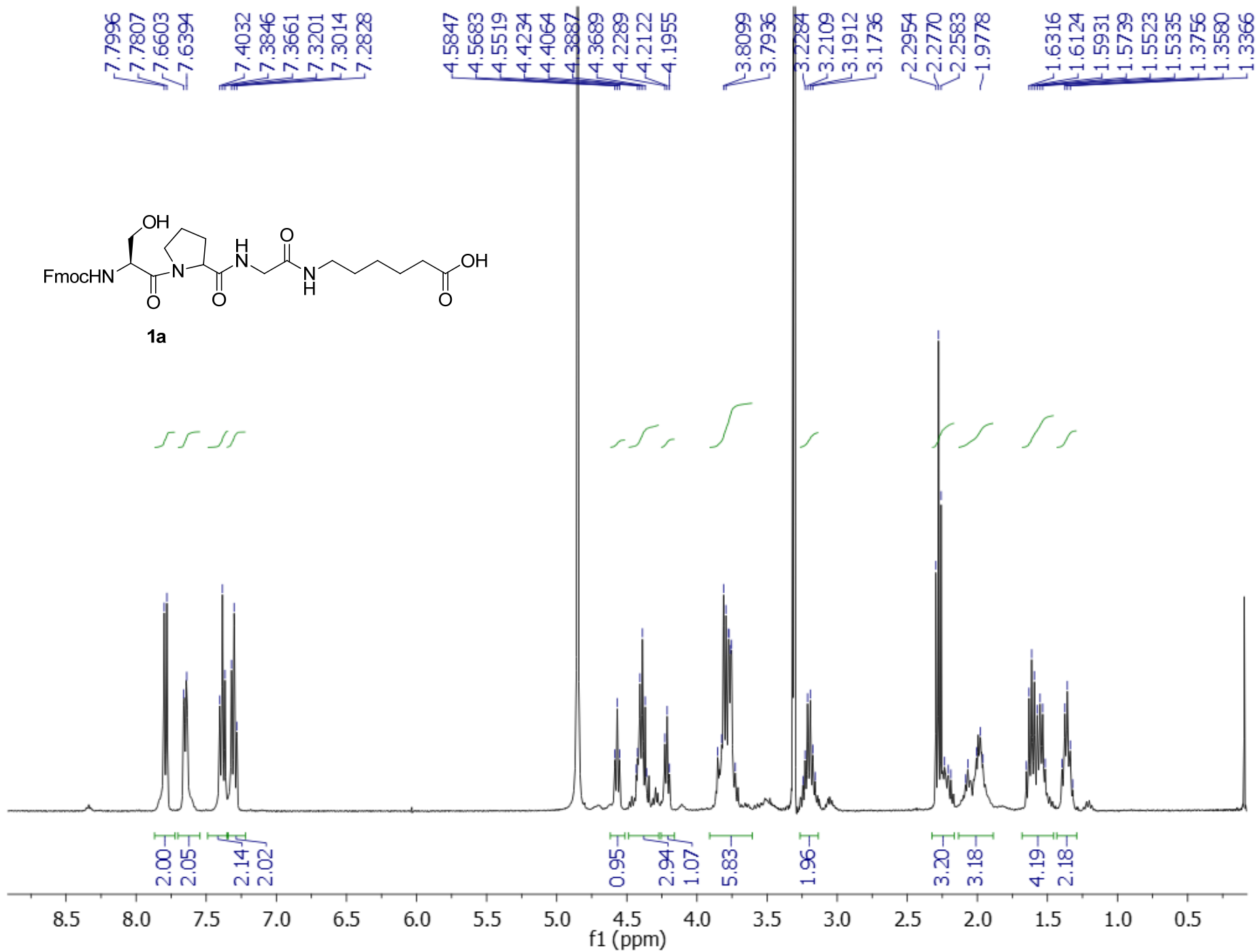
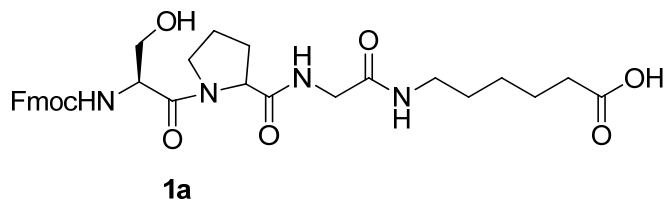


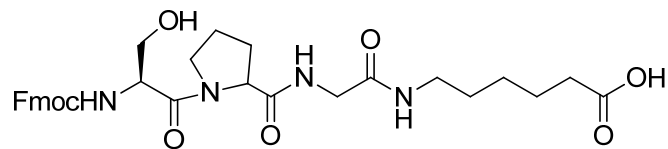




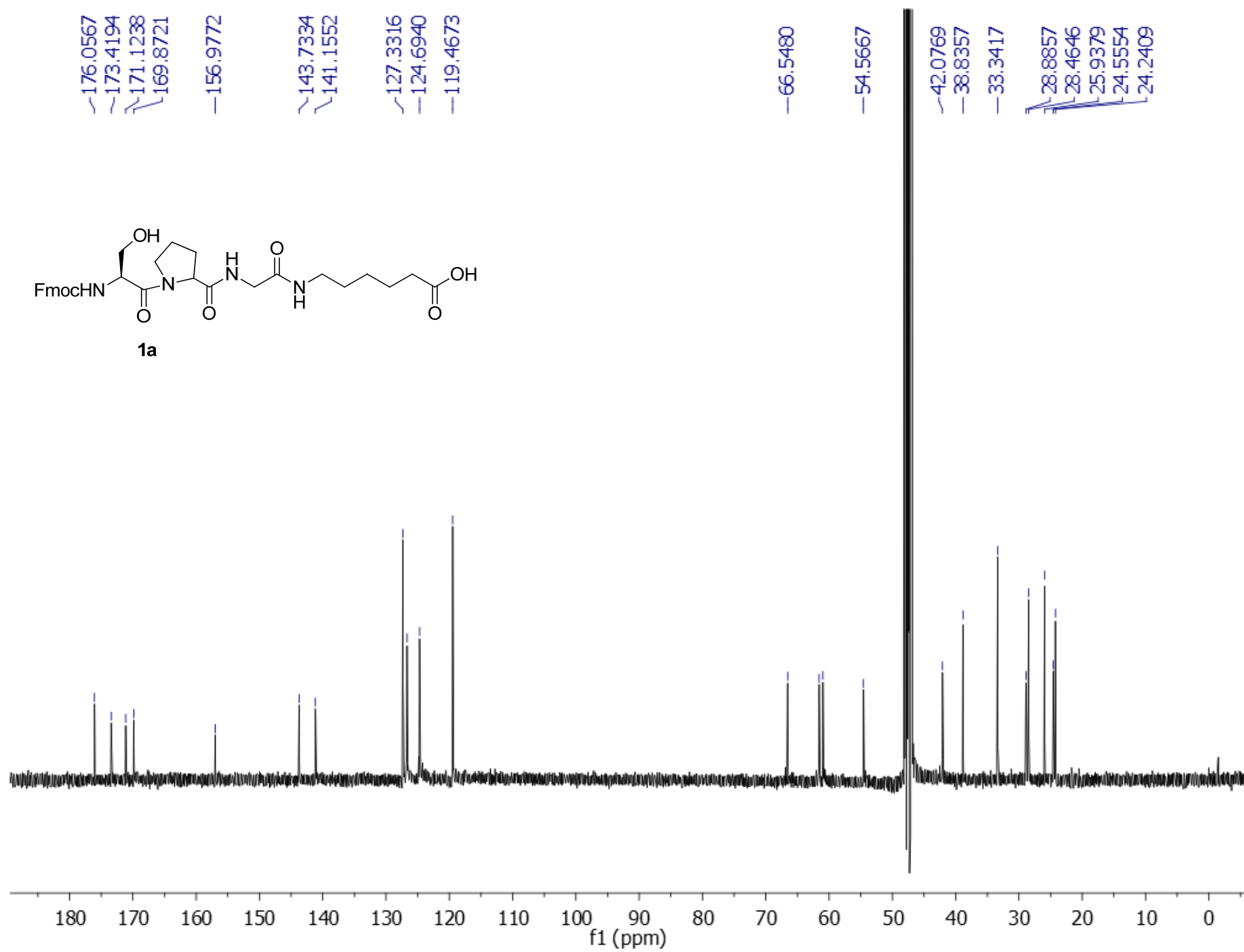


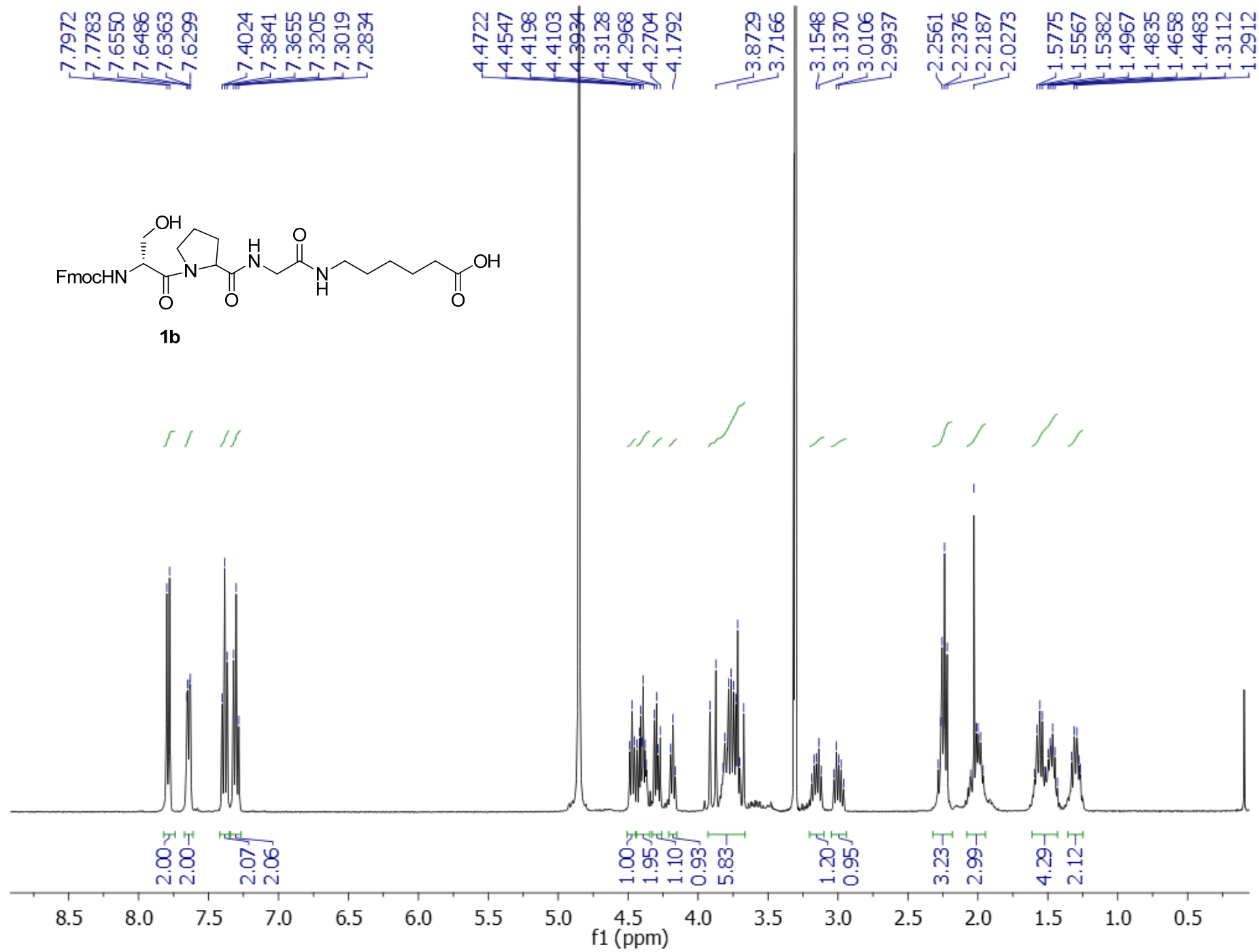


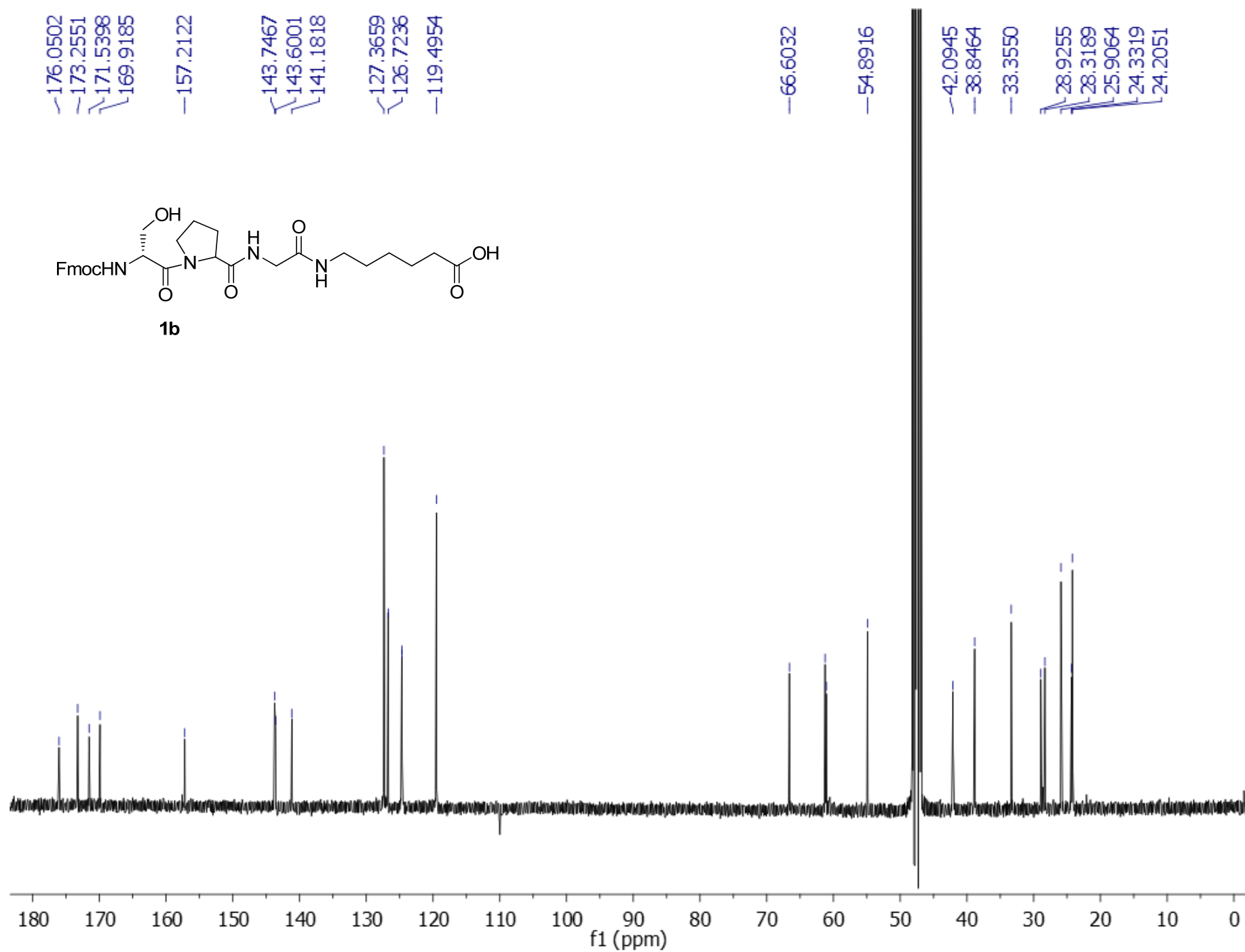


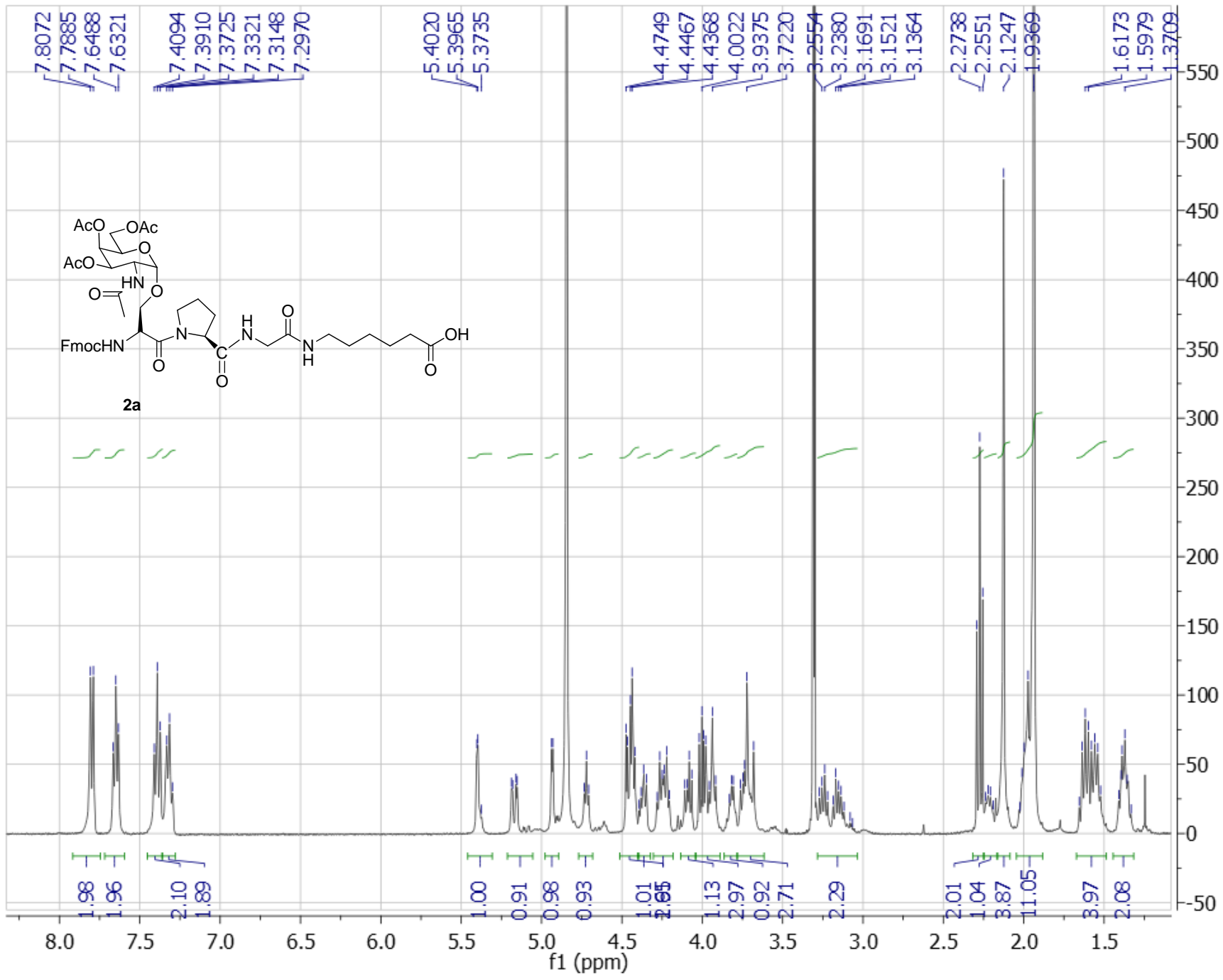


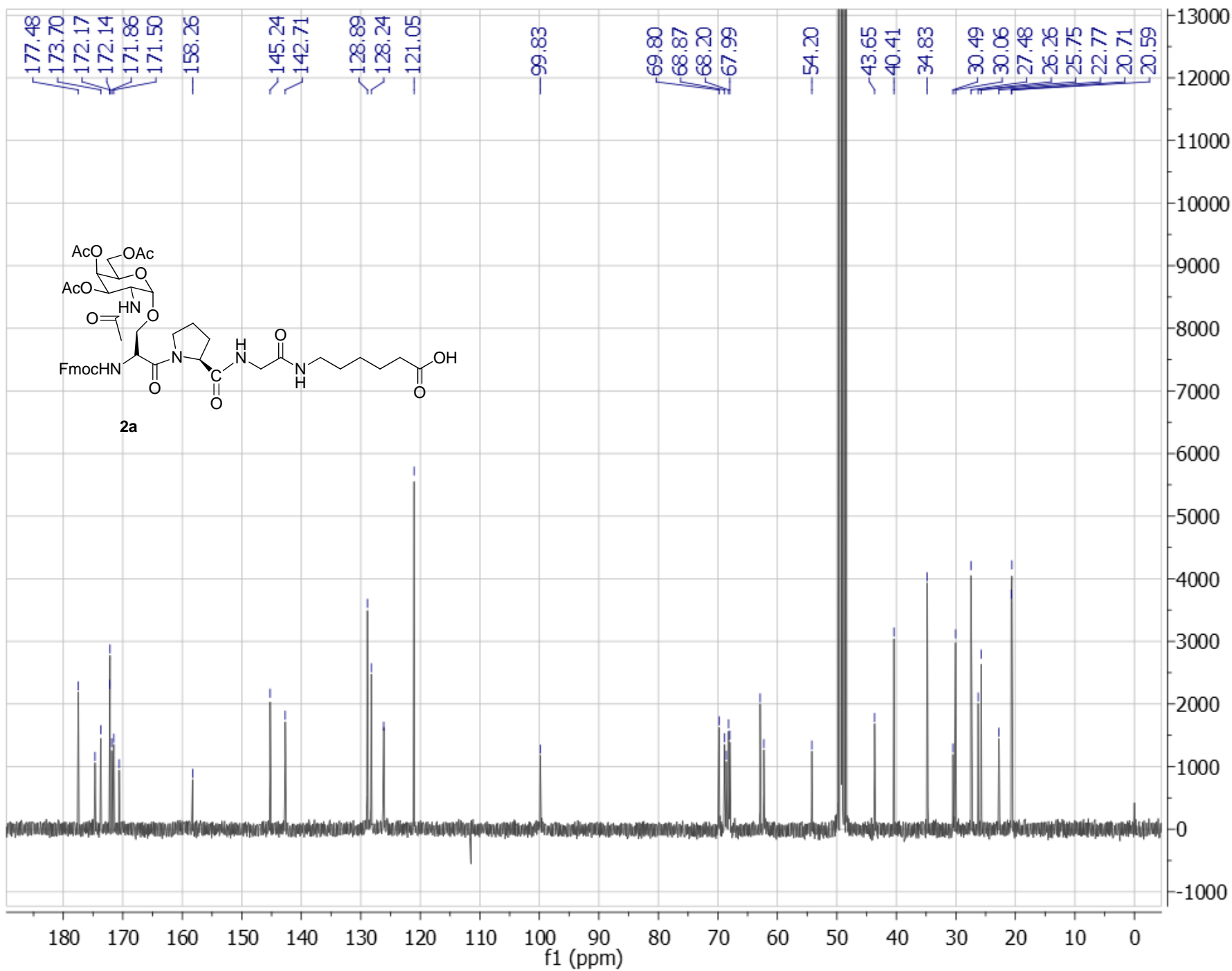
1a

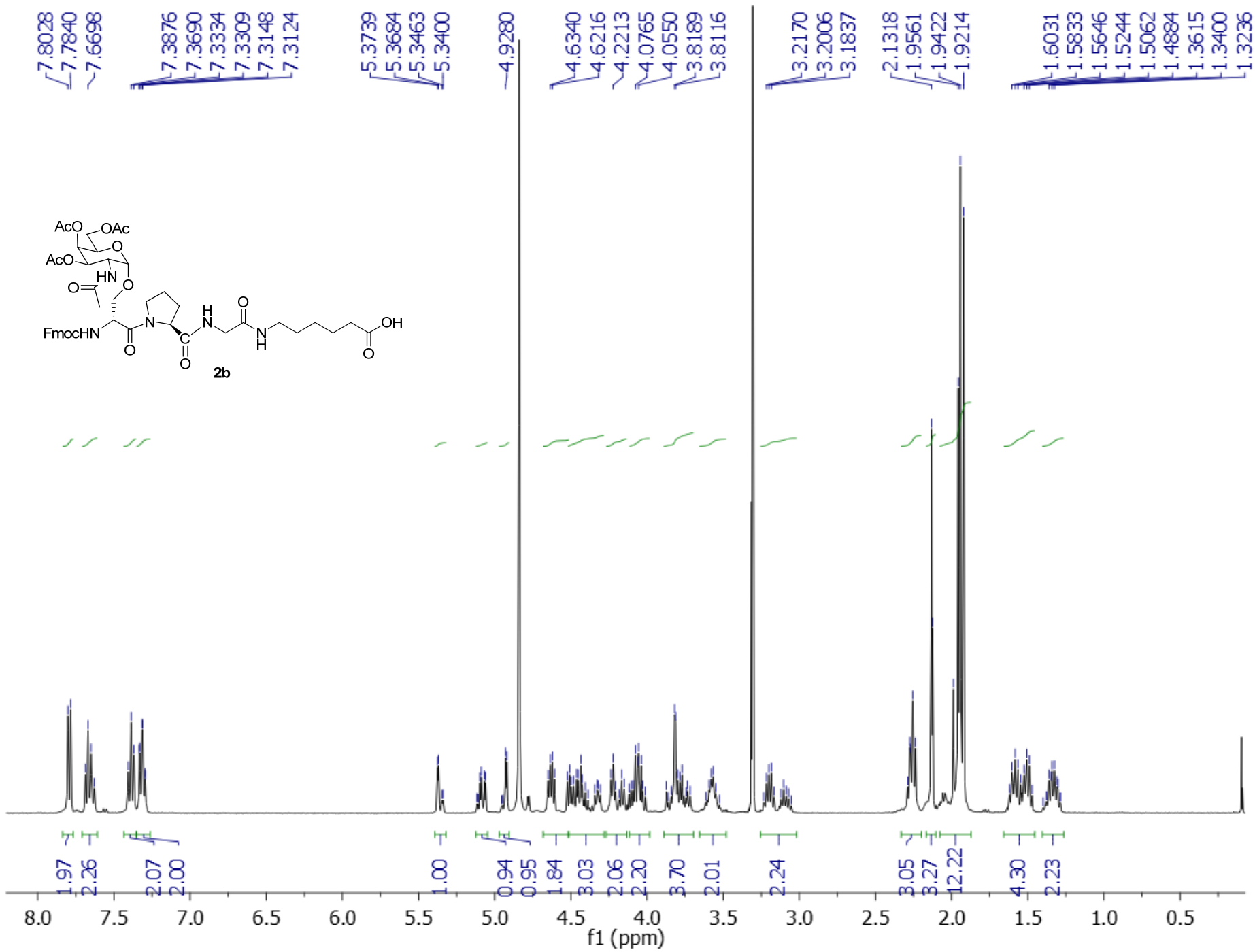


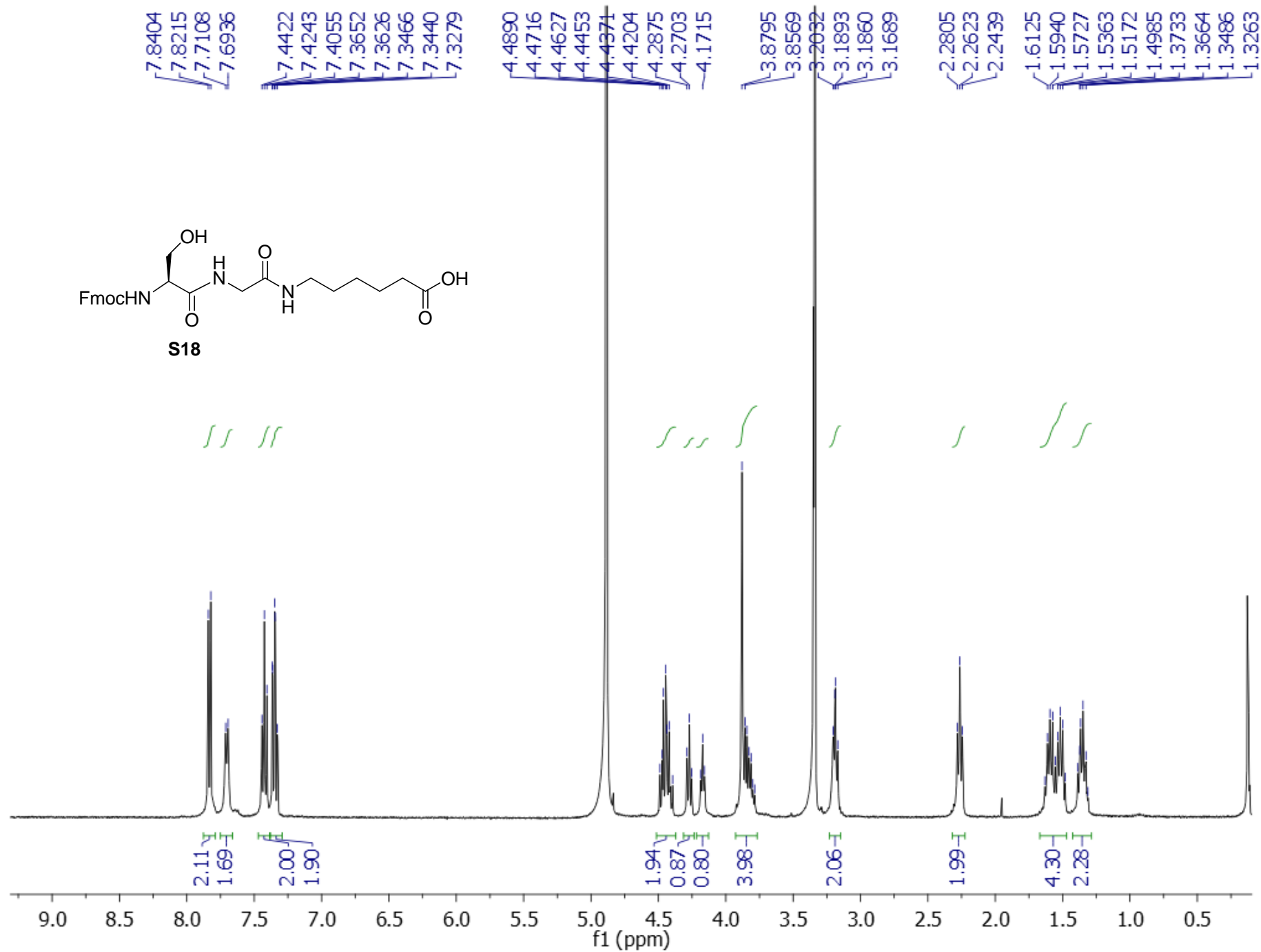
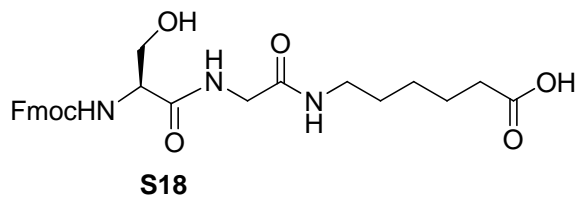


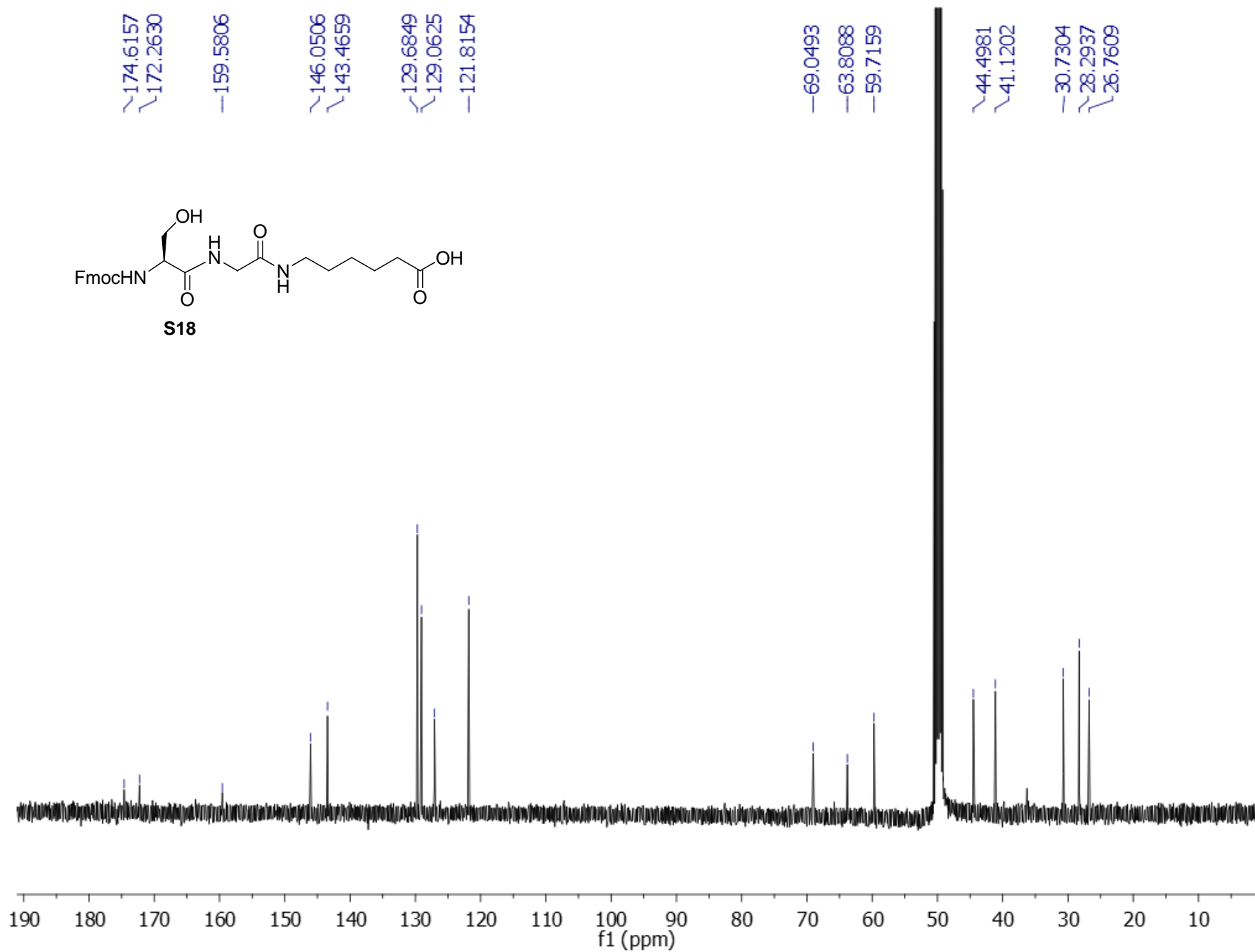
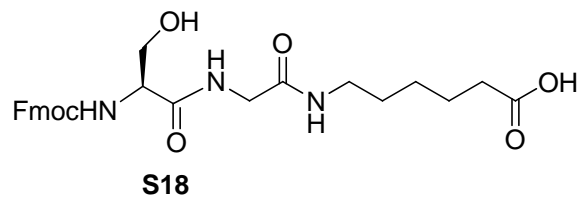


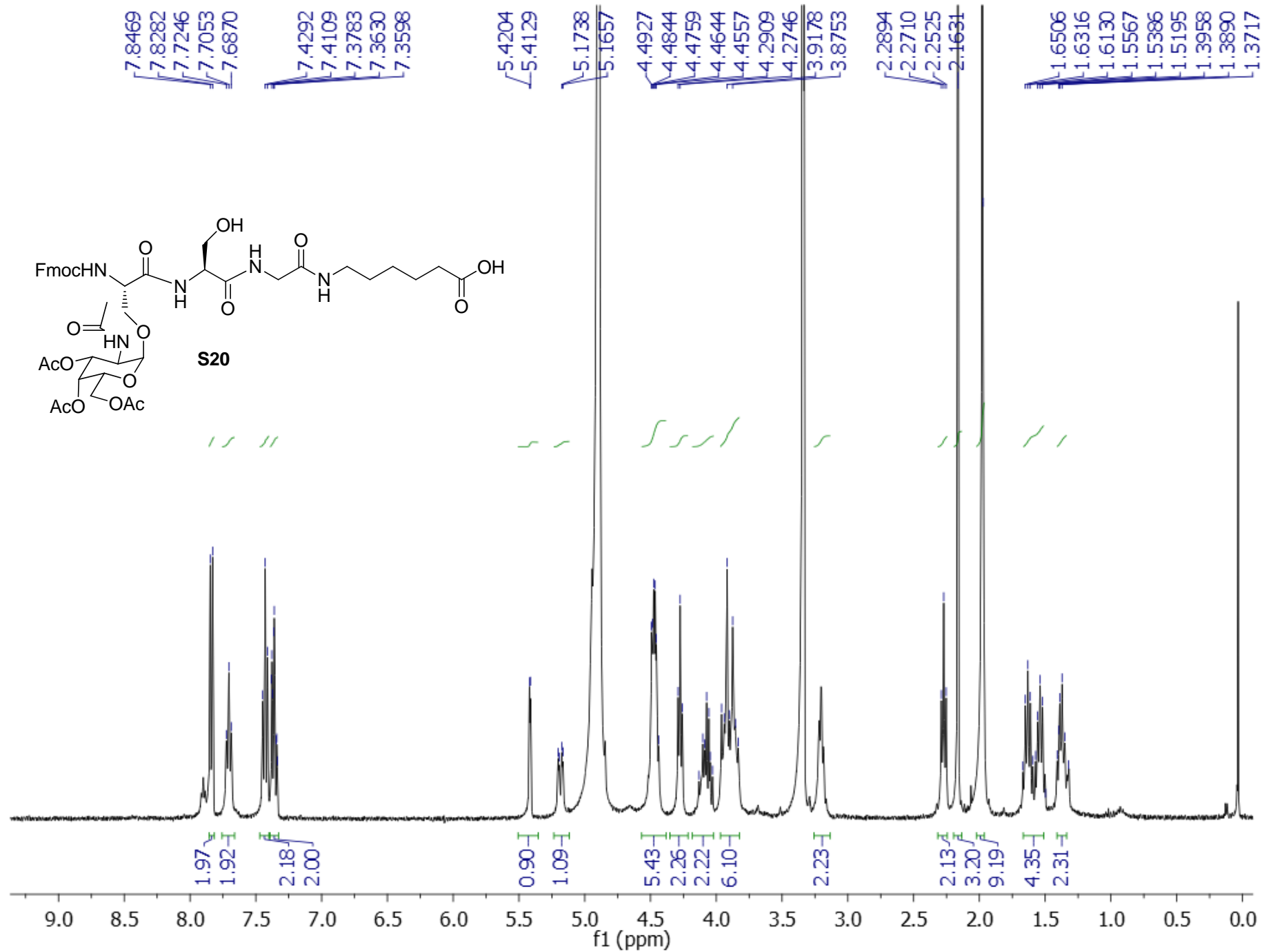












172.2129
171.2073
170.8280
170.7055
170.6288
170.3966
169.9096
161.9376
157.0770

143.7881
143.6866
141.1829
141.1675

127.3965
126.7716

119.5449
118.0648
115.1667

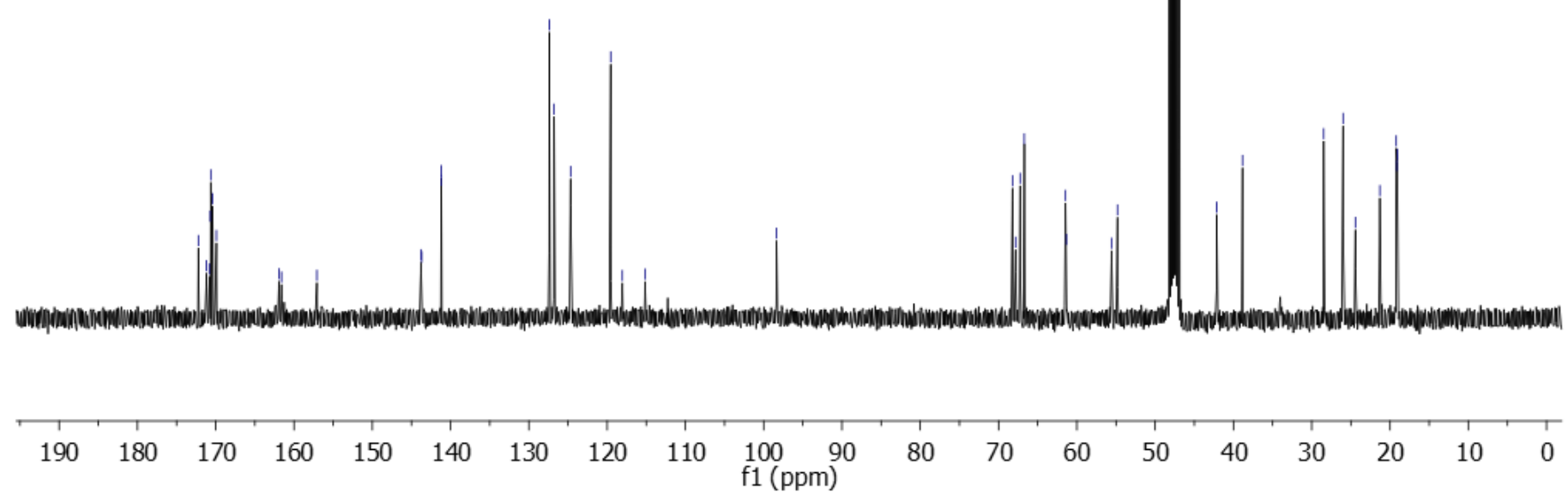
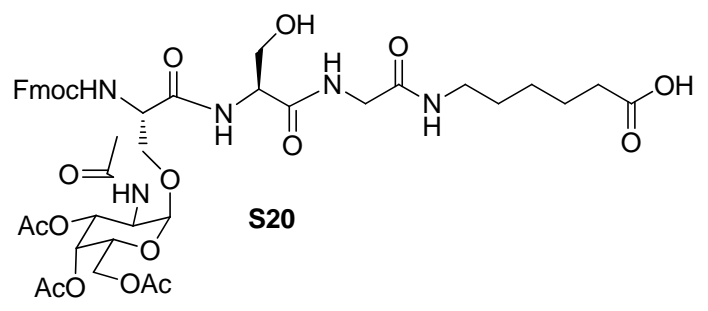
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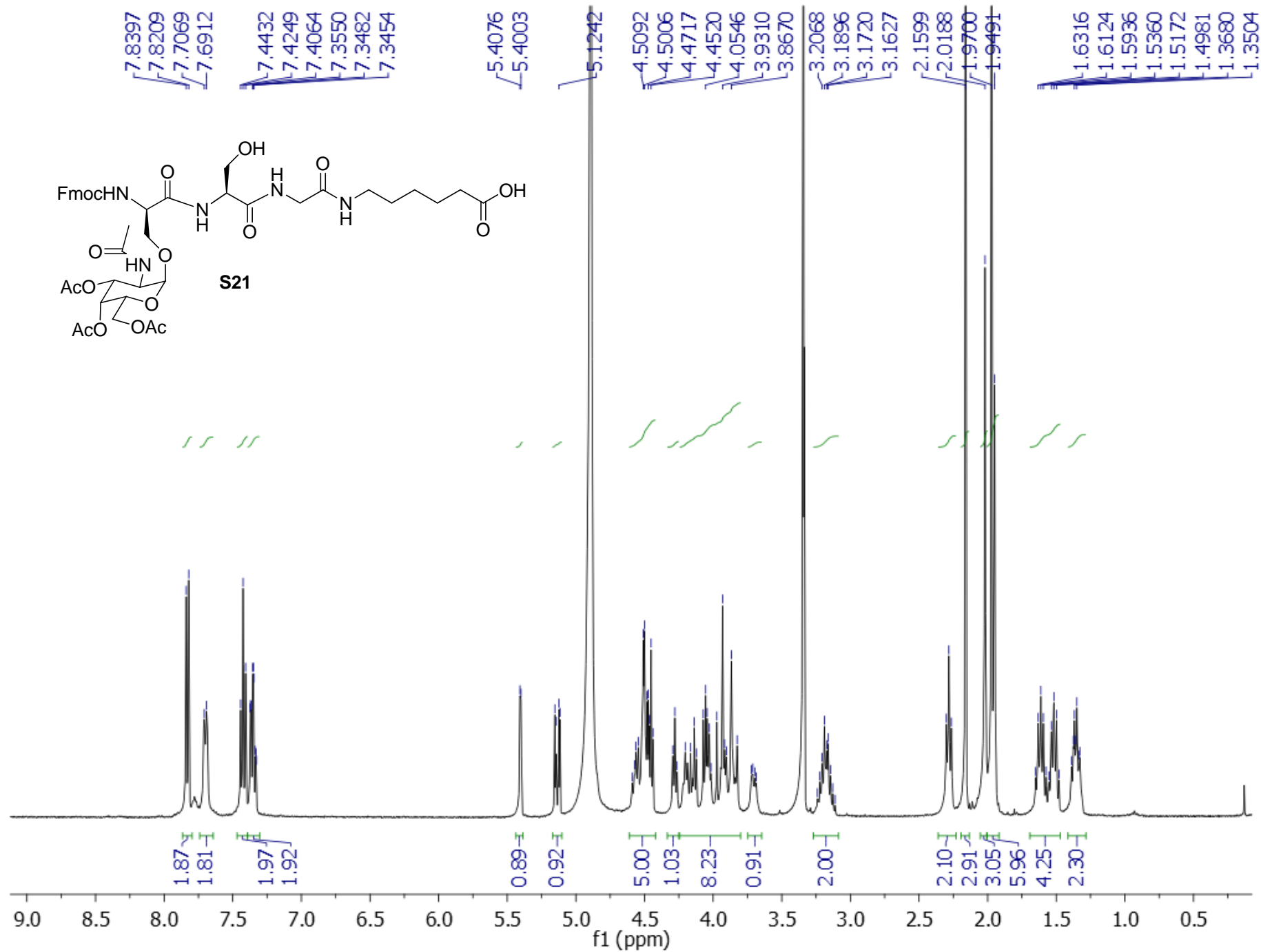
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67.8340
67.2384
66.7202

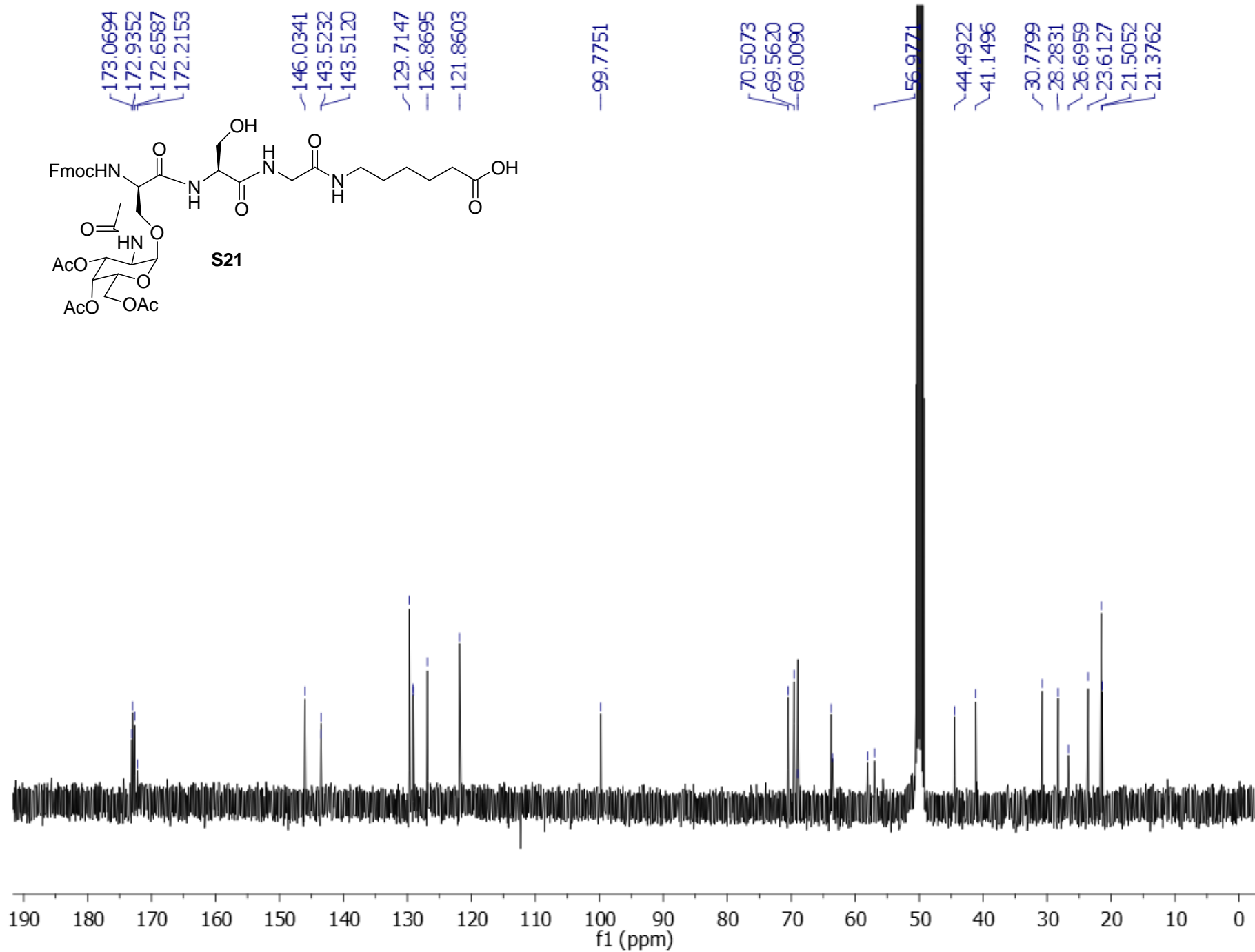
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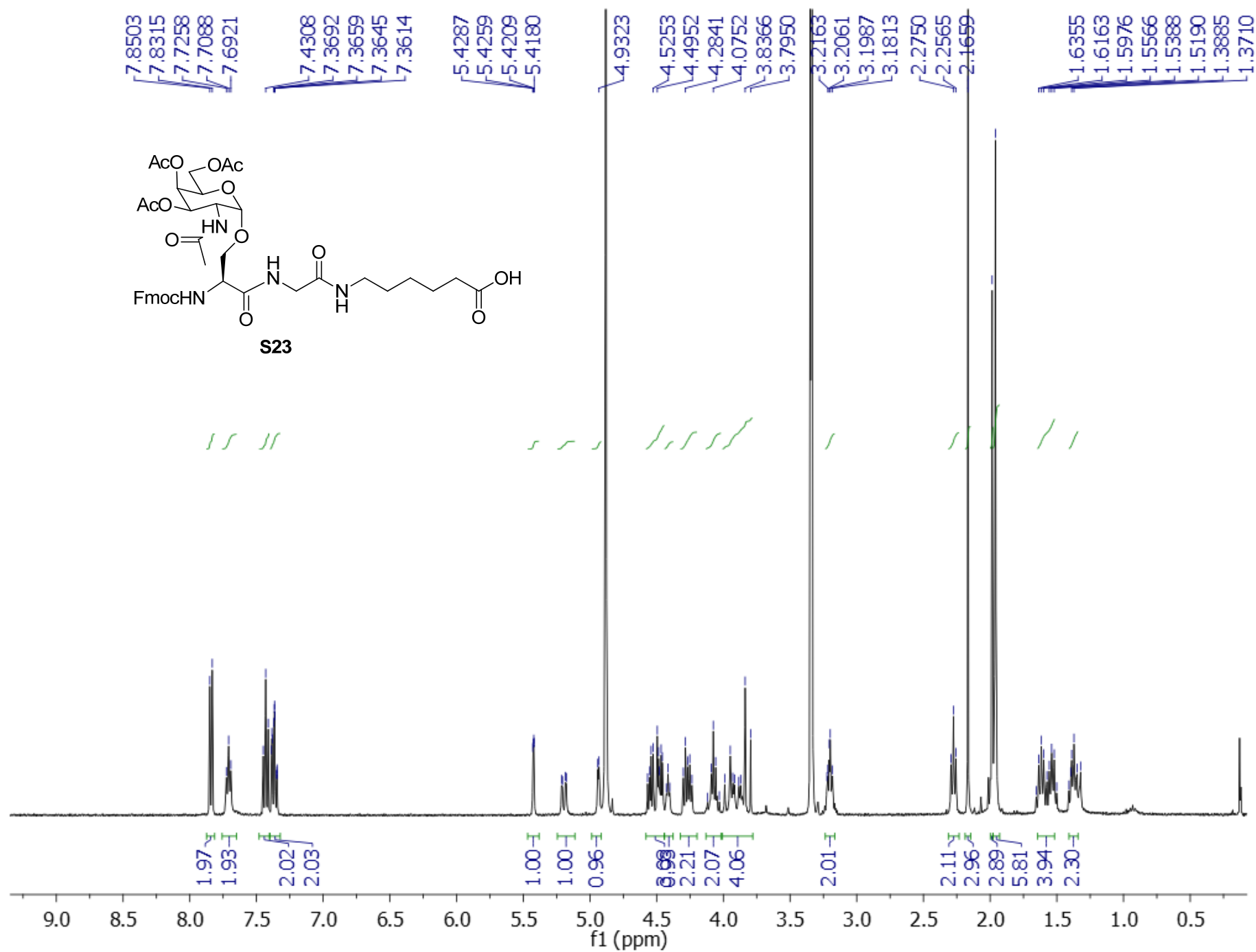
42.1437
38.8385

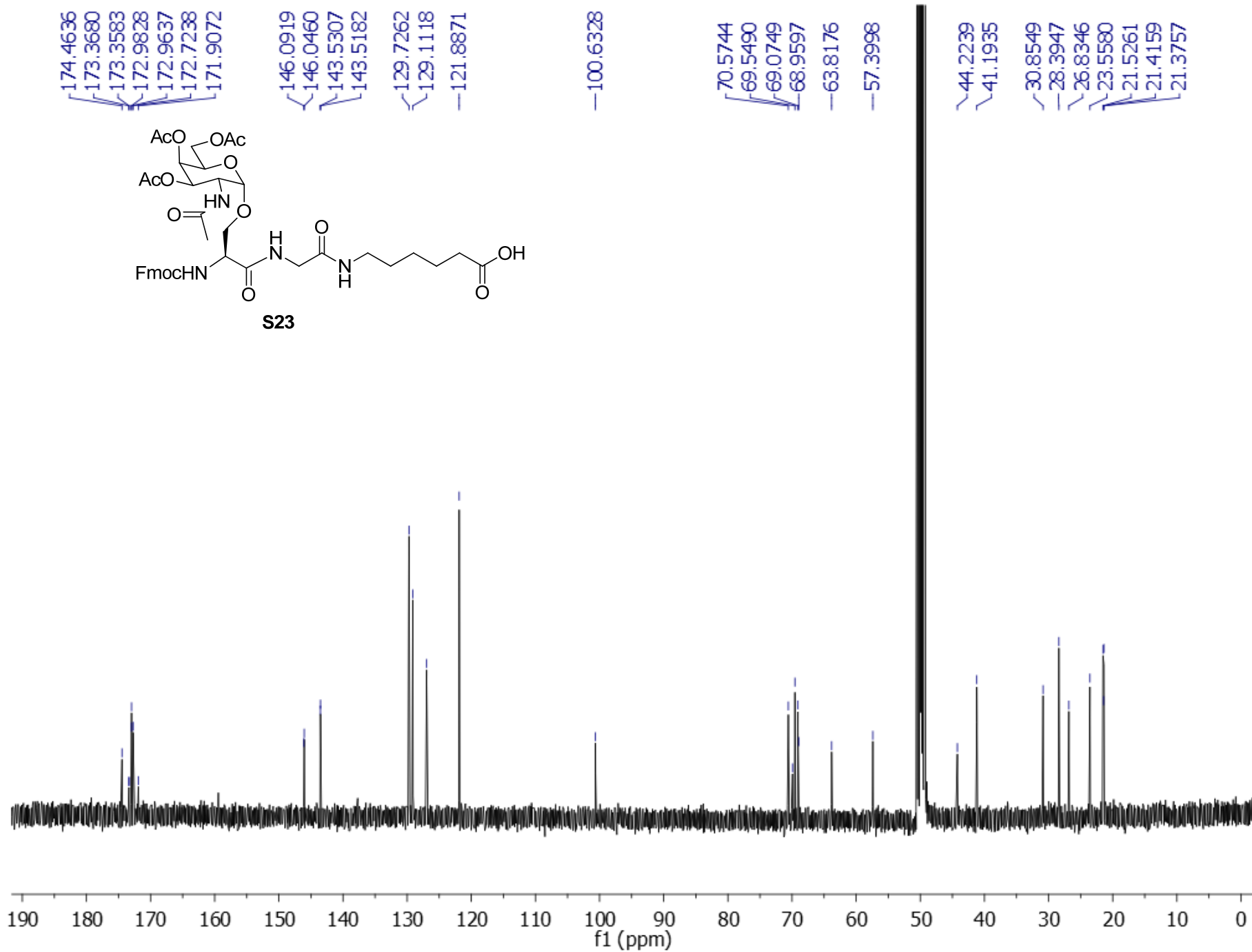
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25.9933
24.4185
21.2963
19.2073
19.1340
19.0684

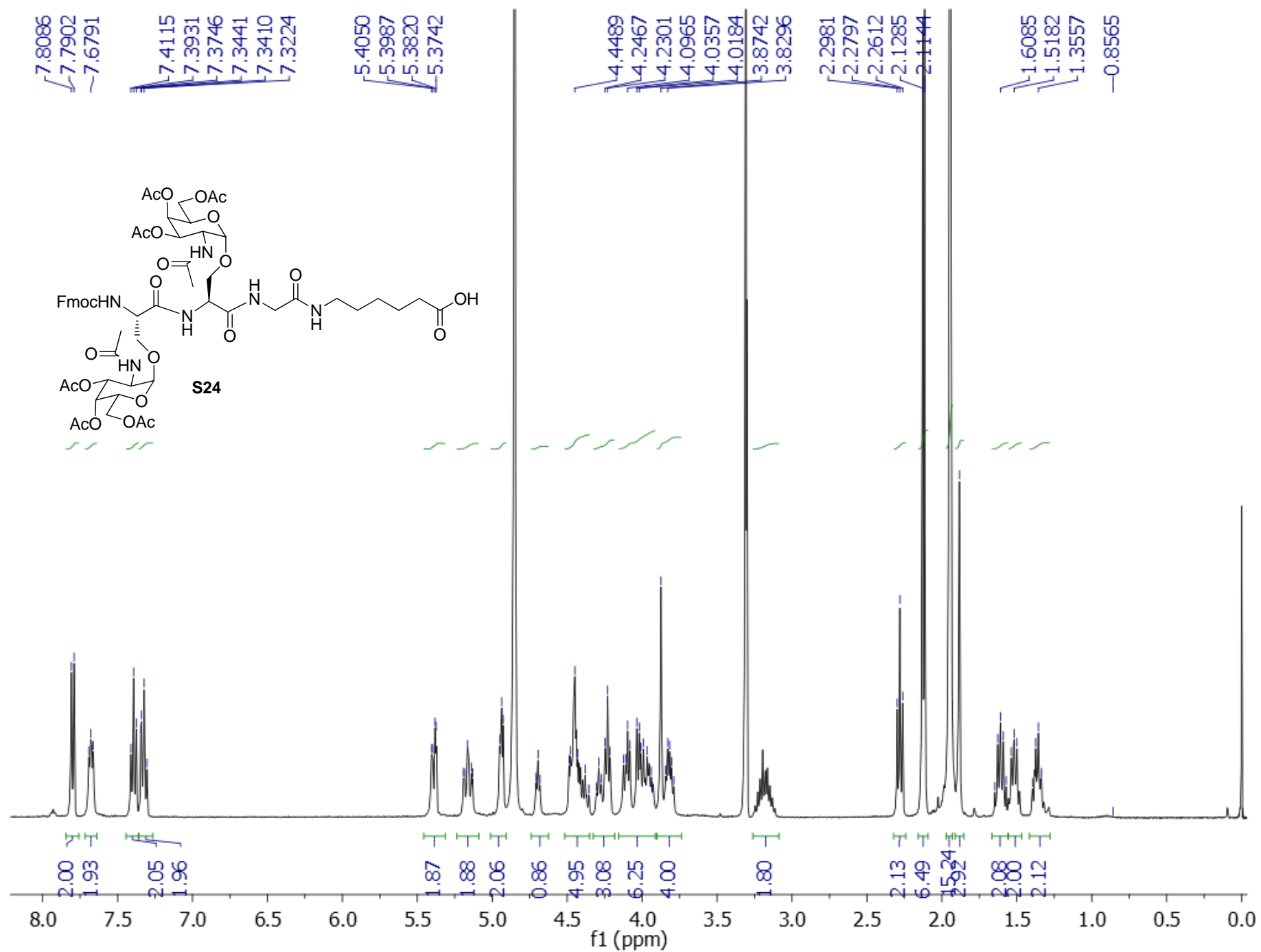


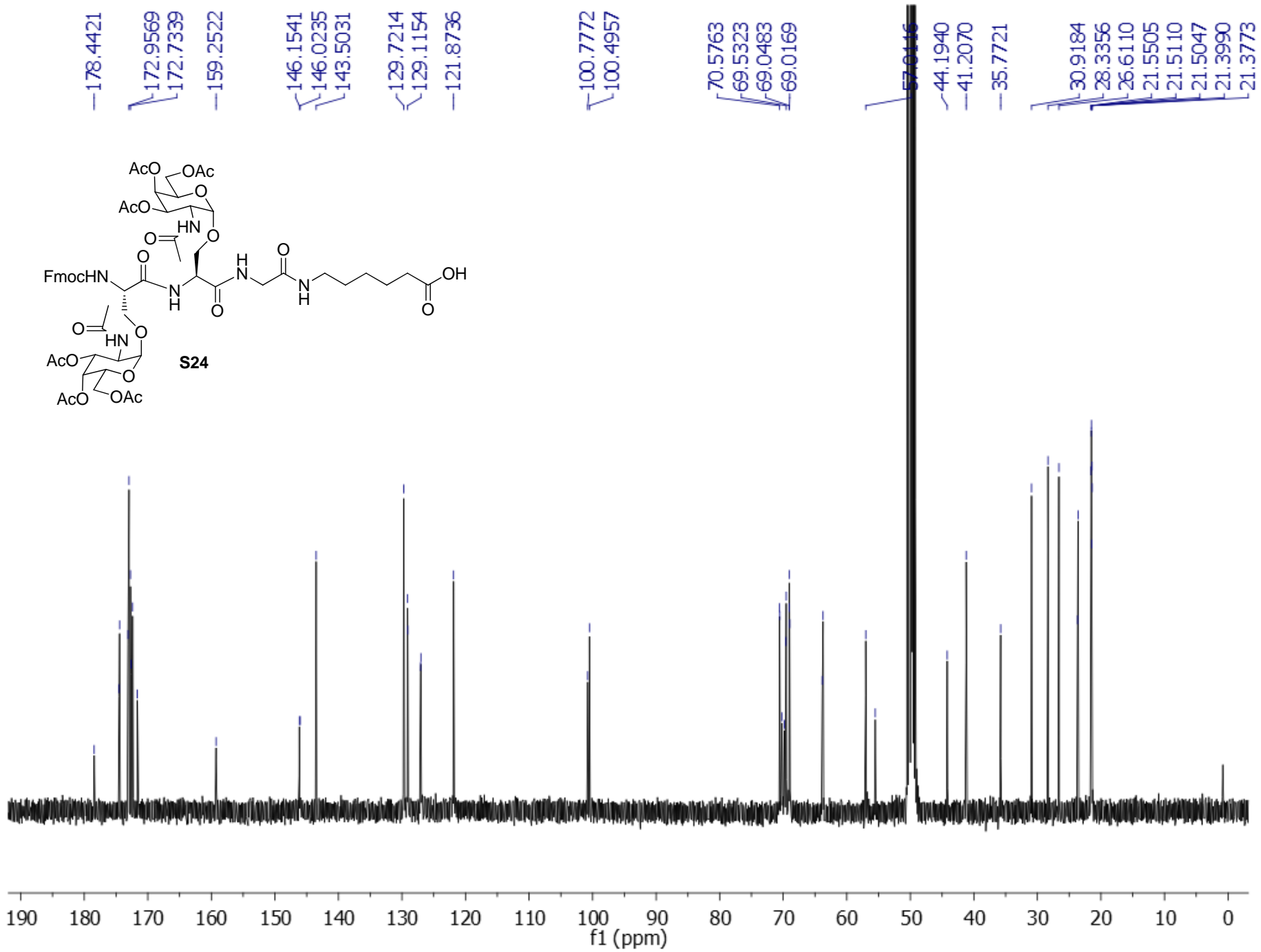


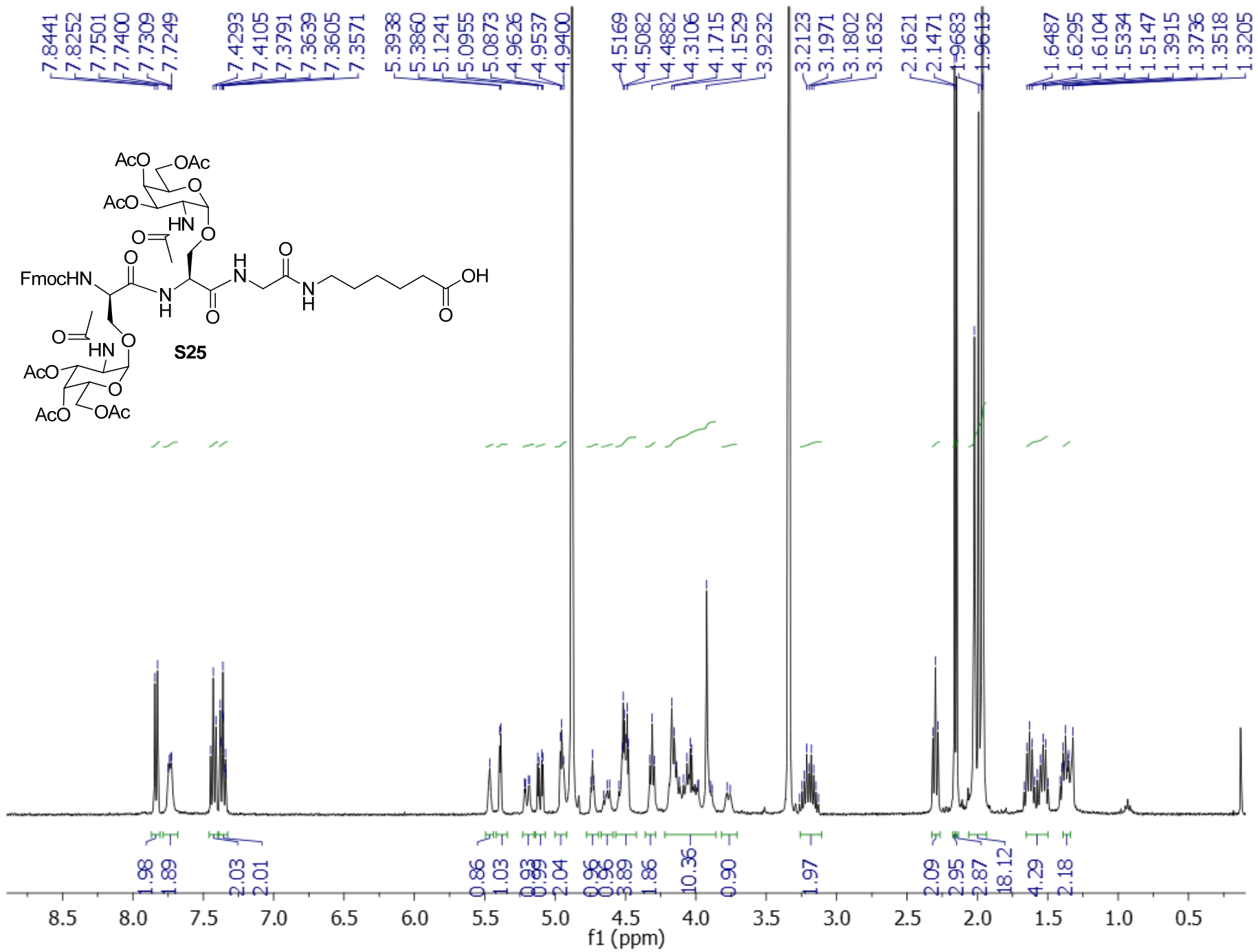


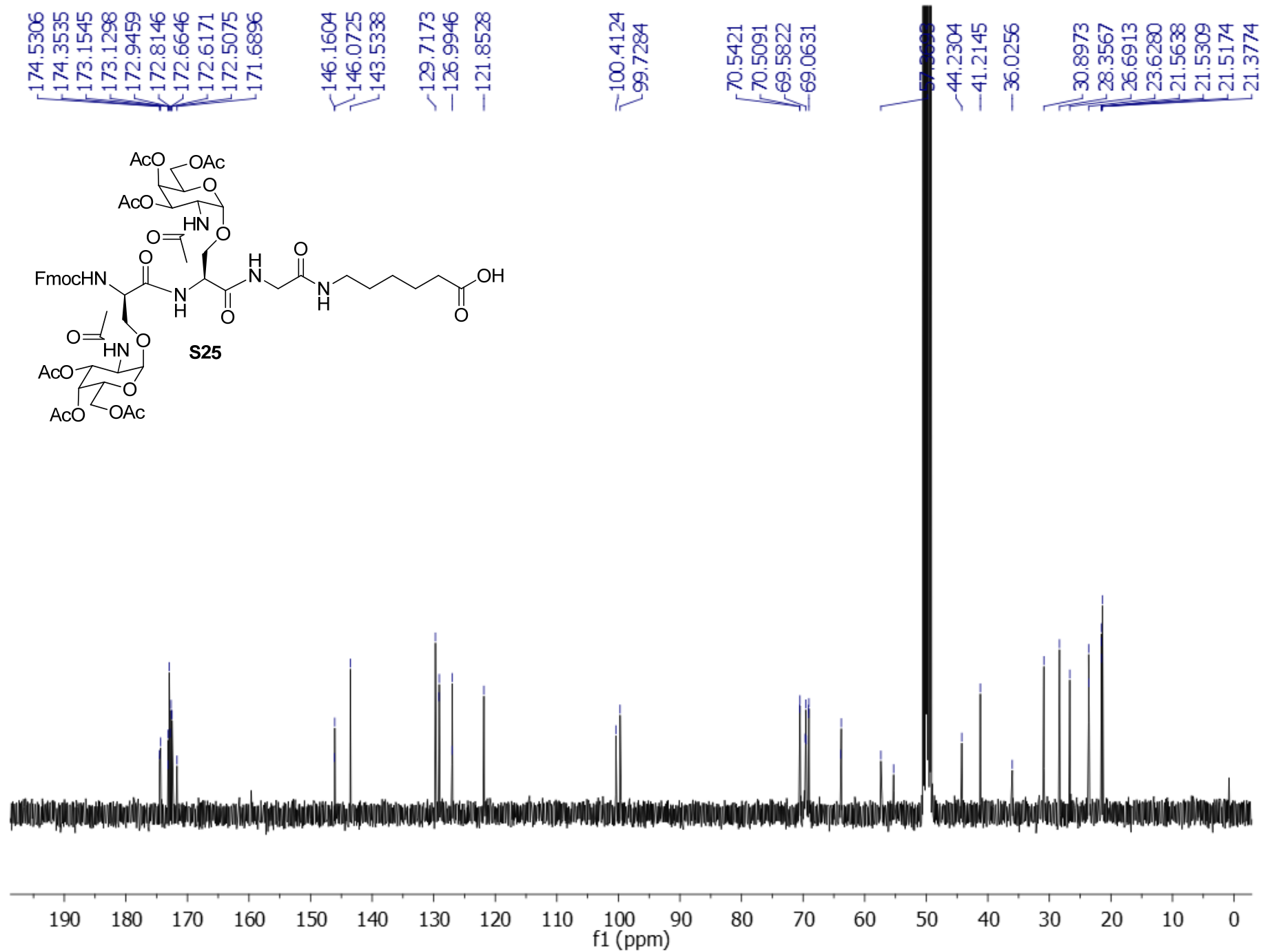


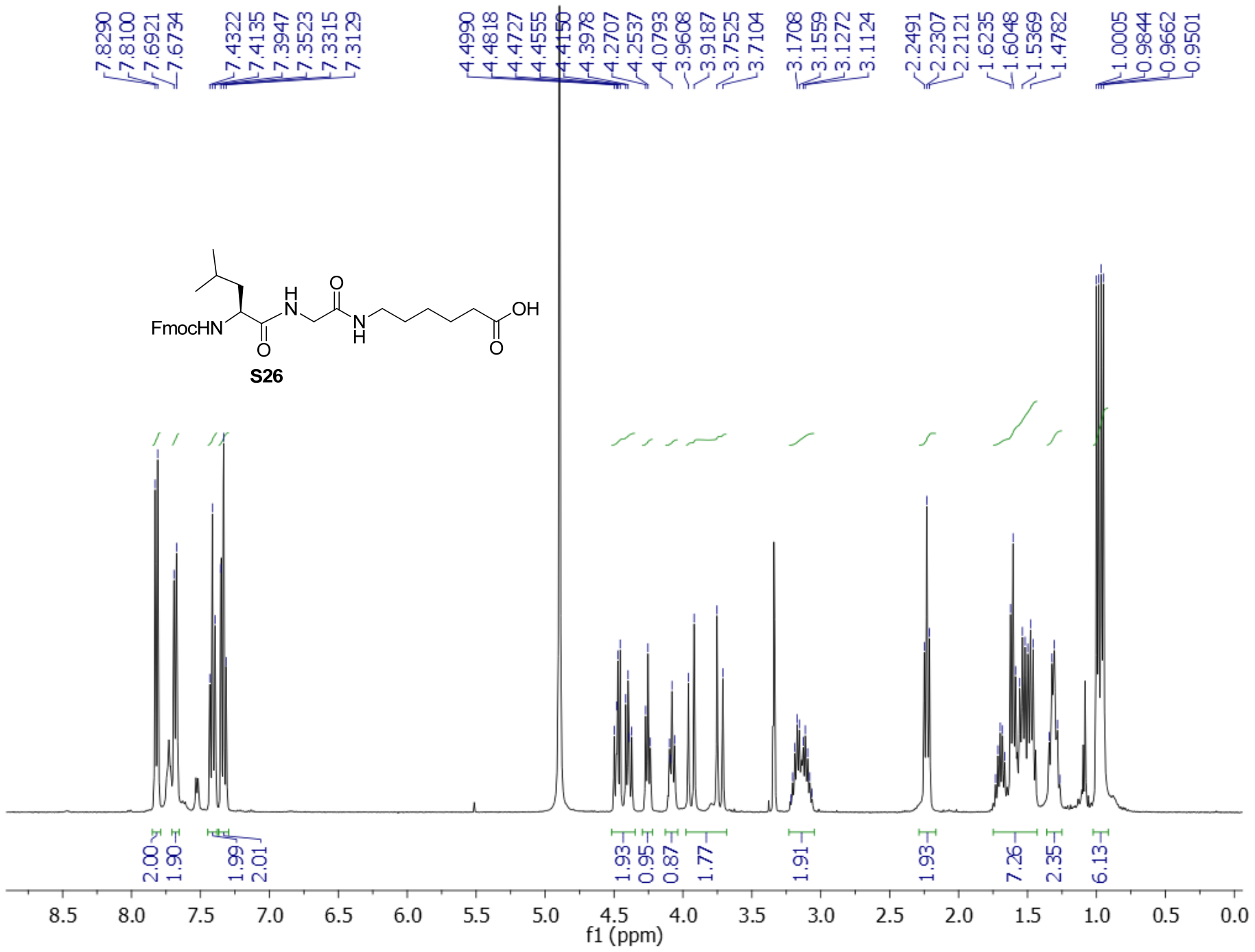
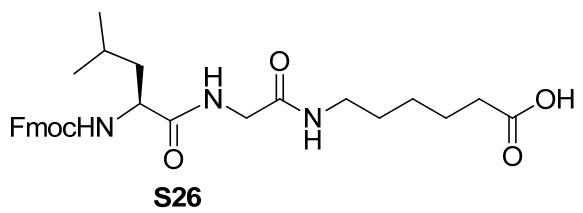


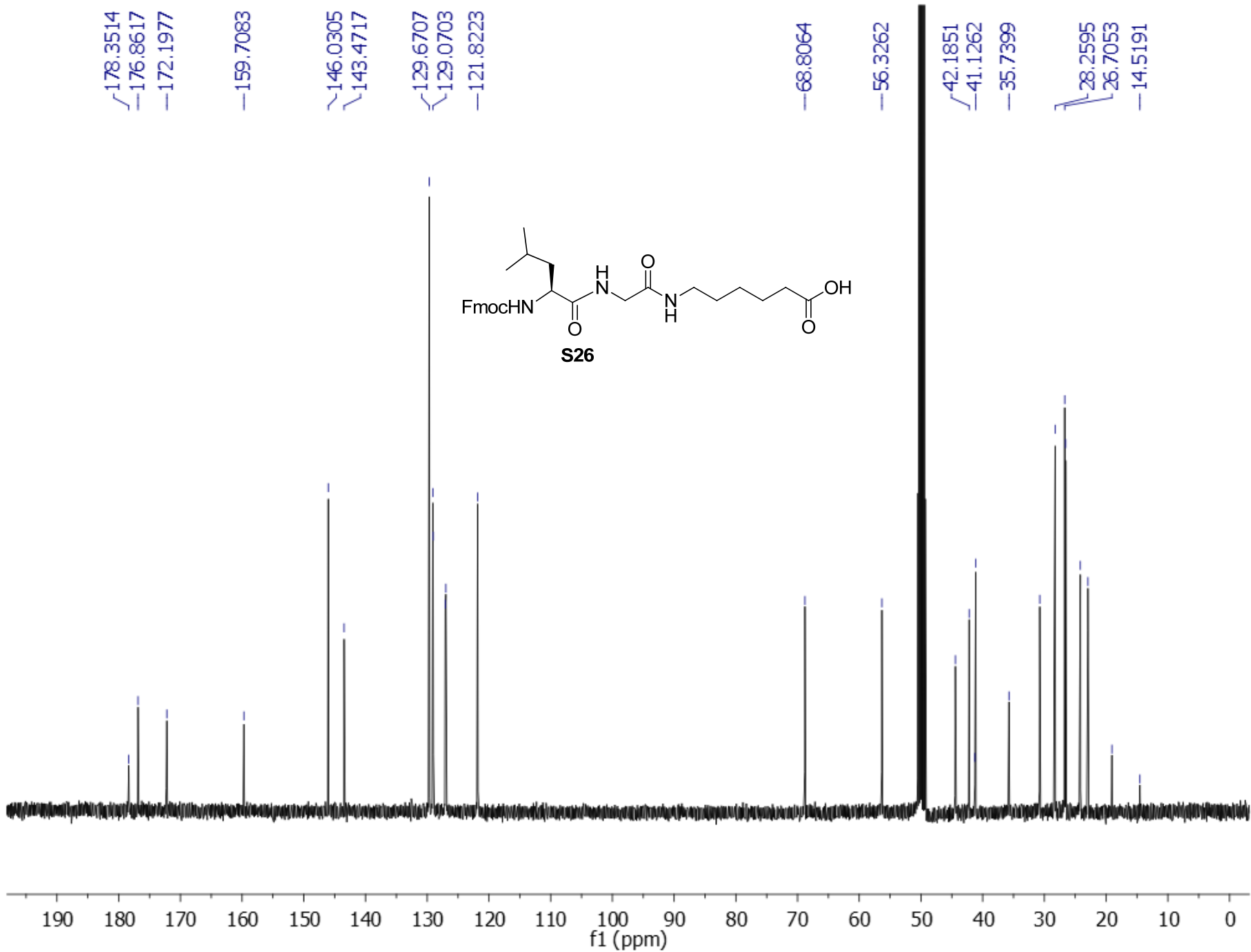


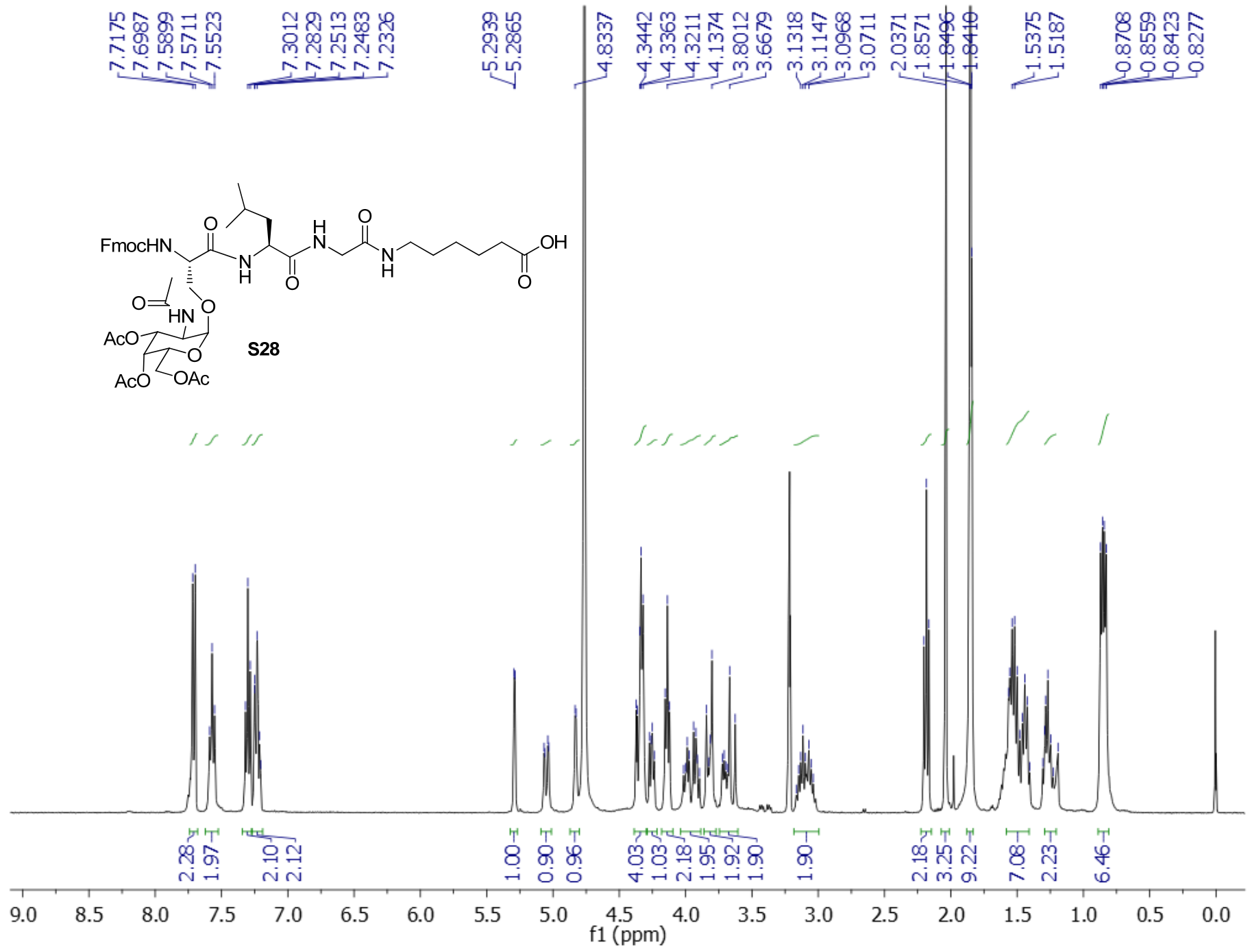


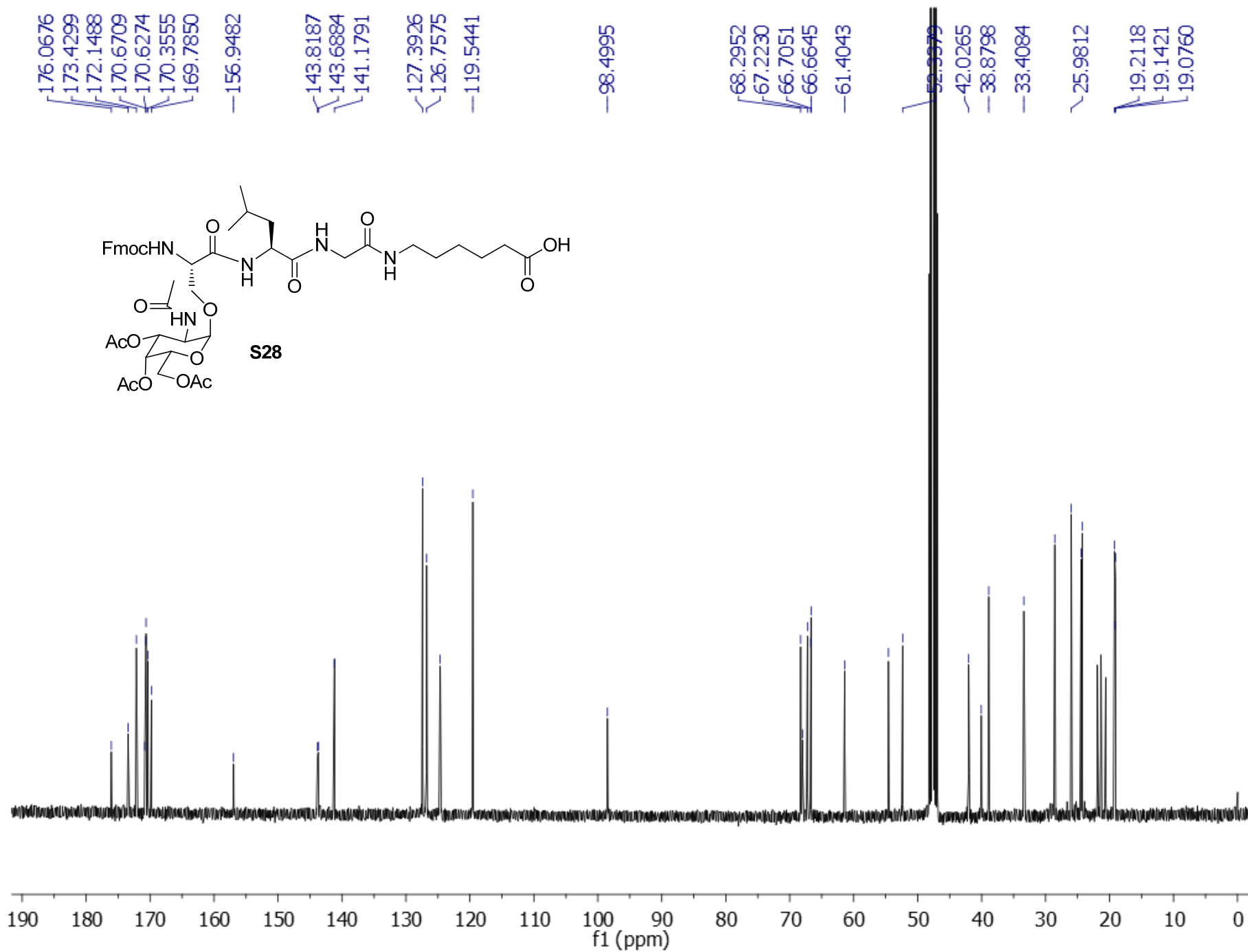


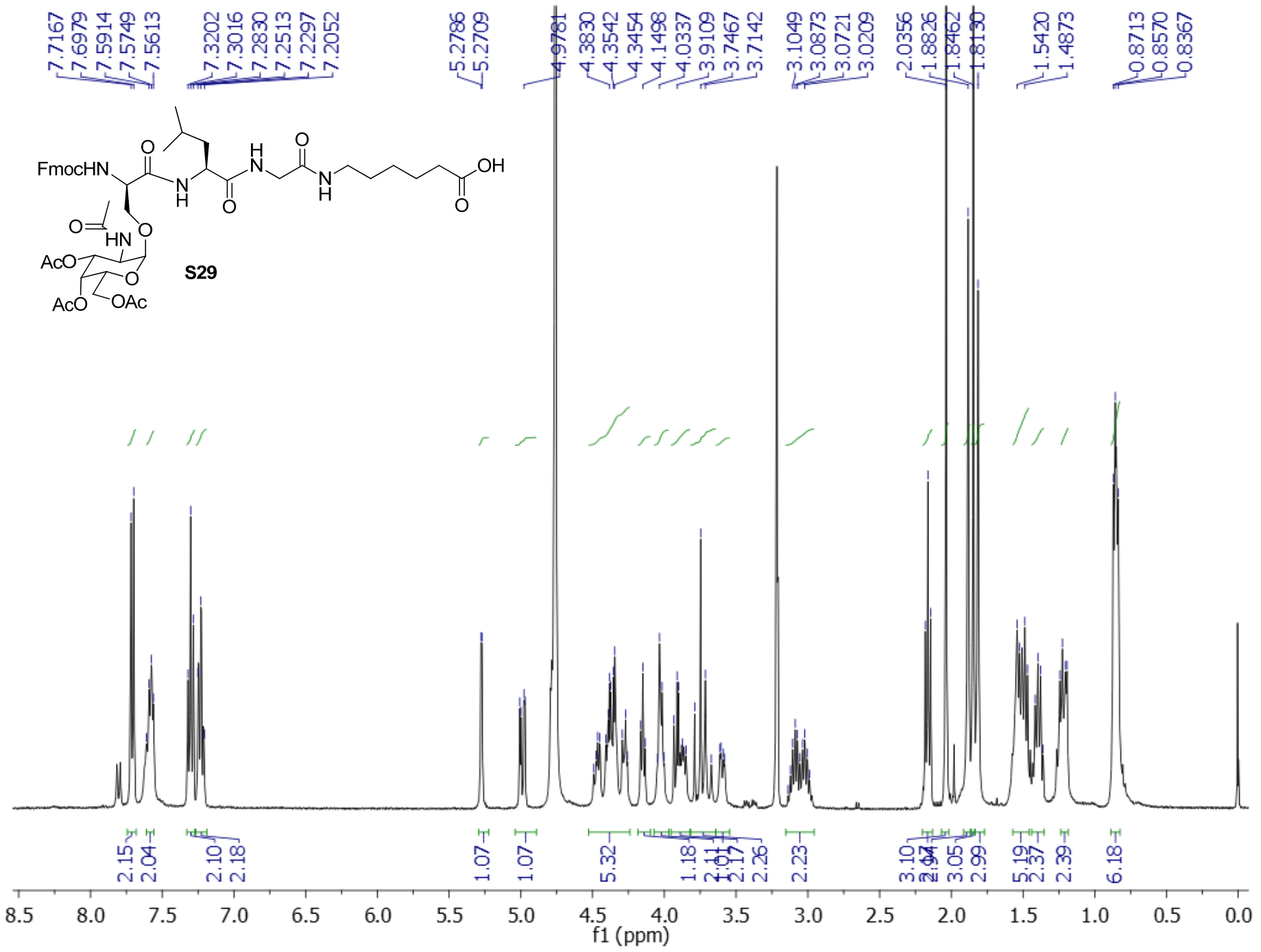


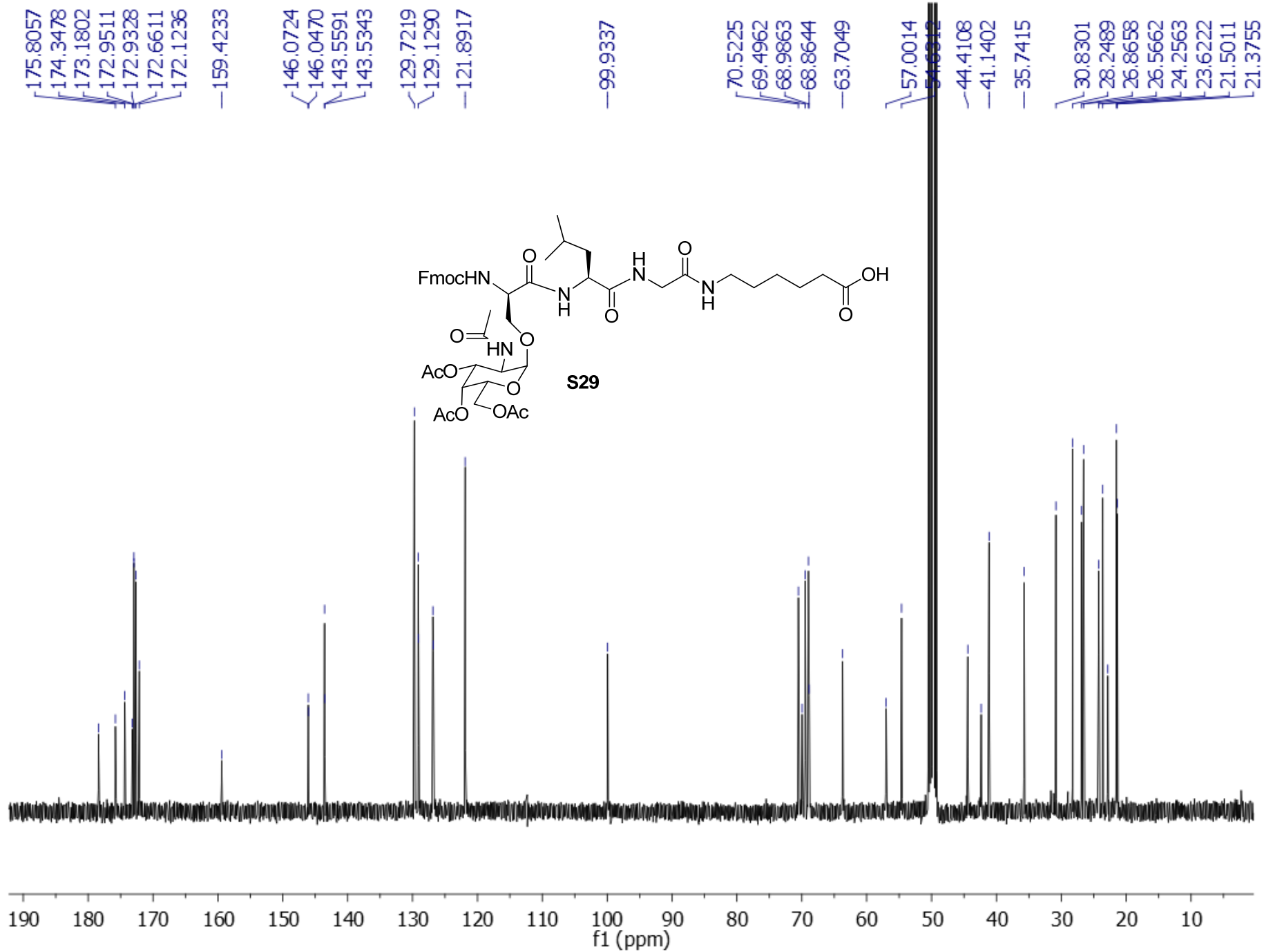


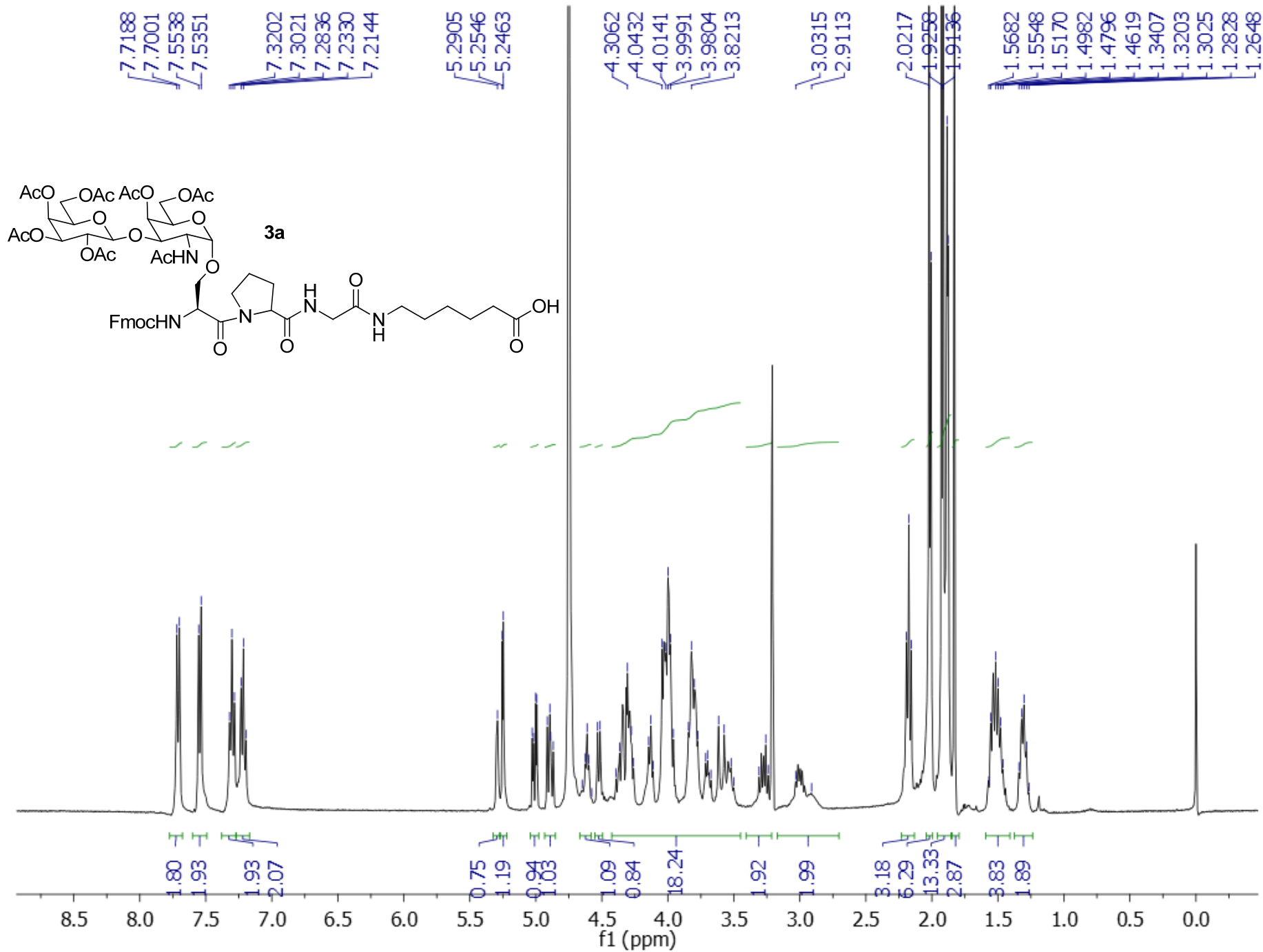


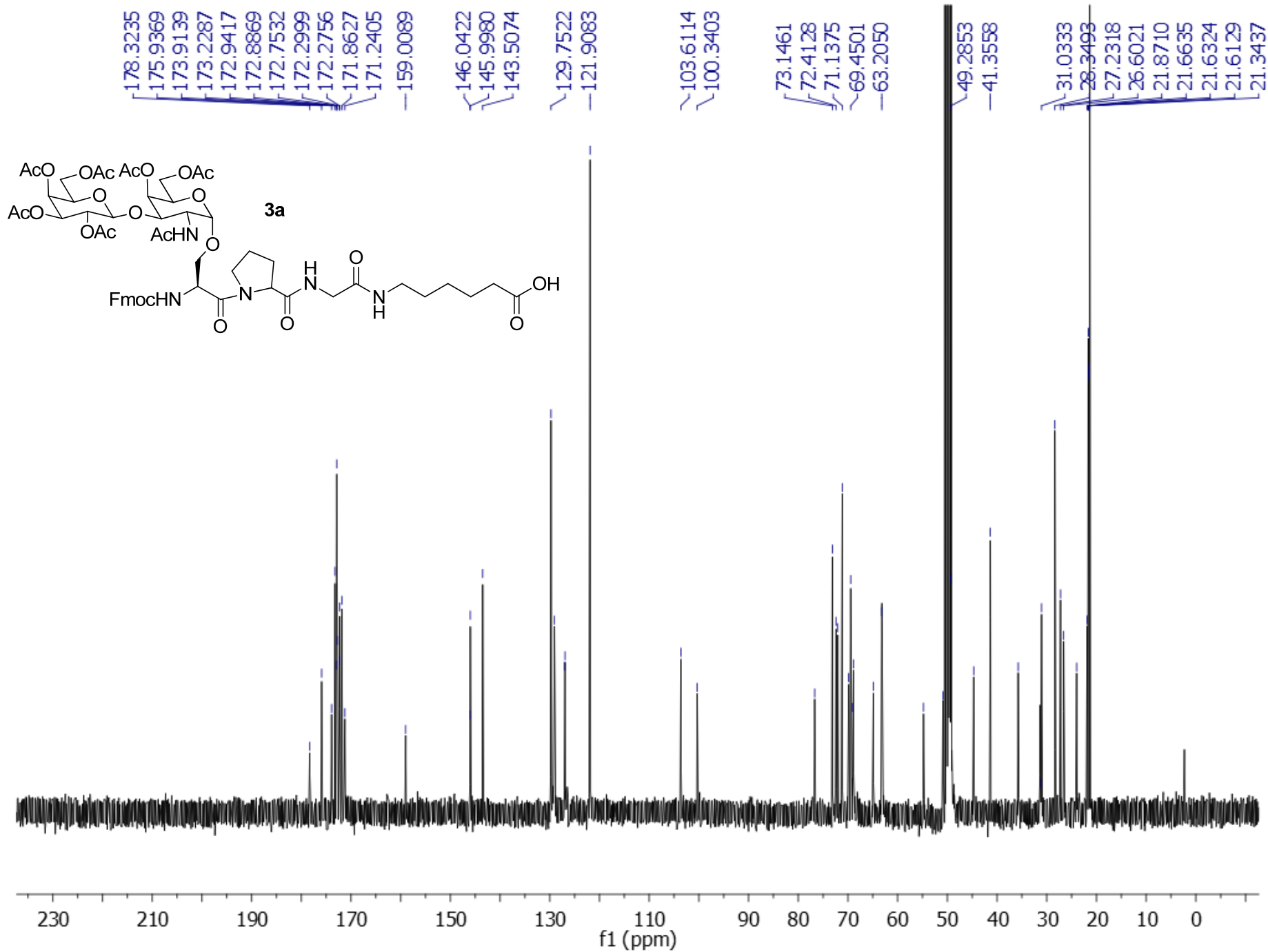


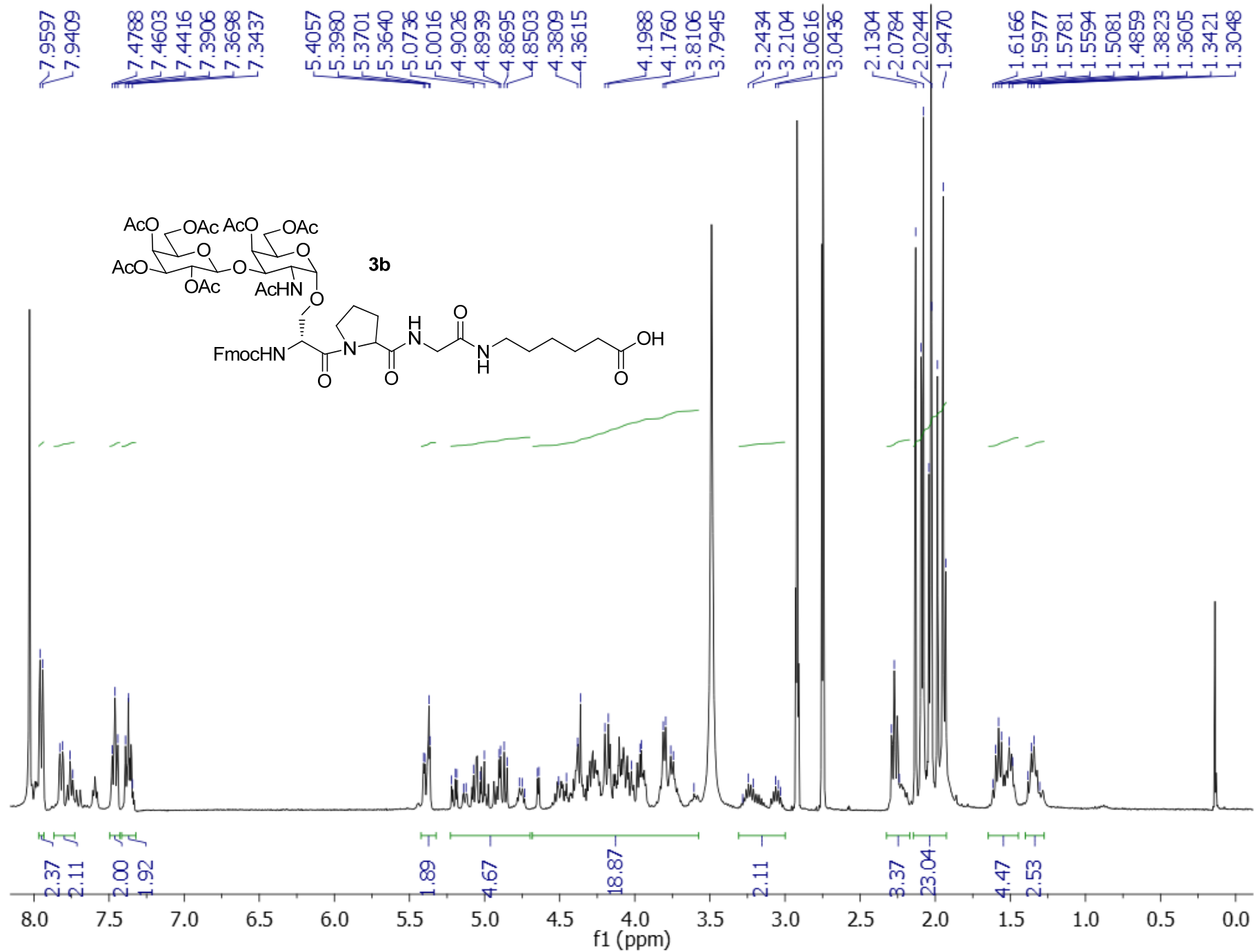


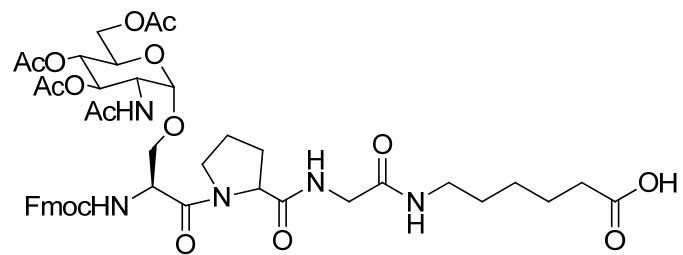




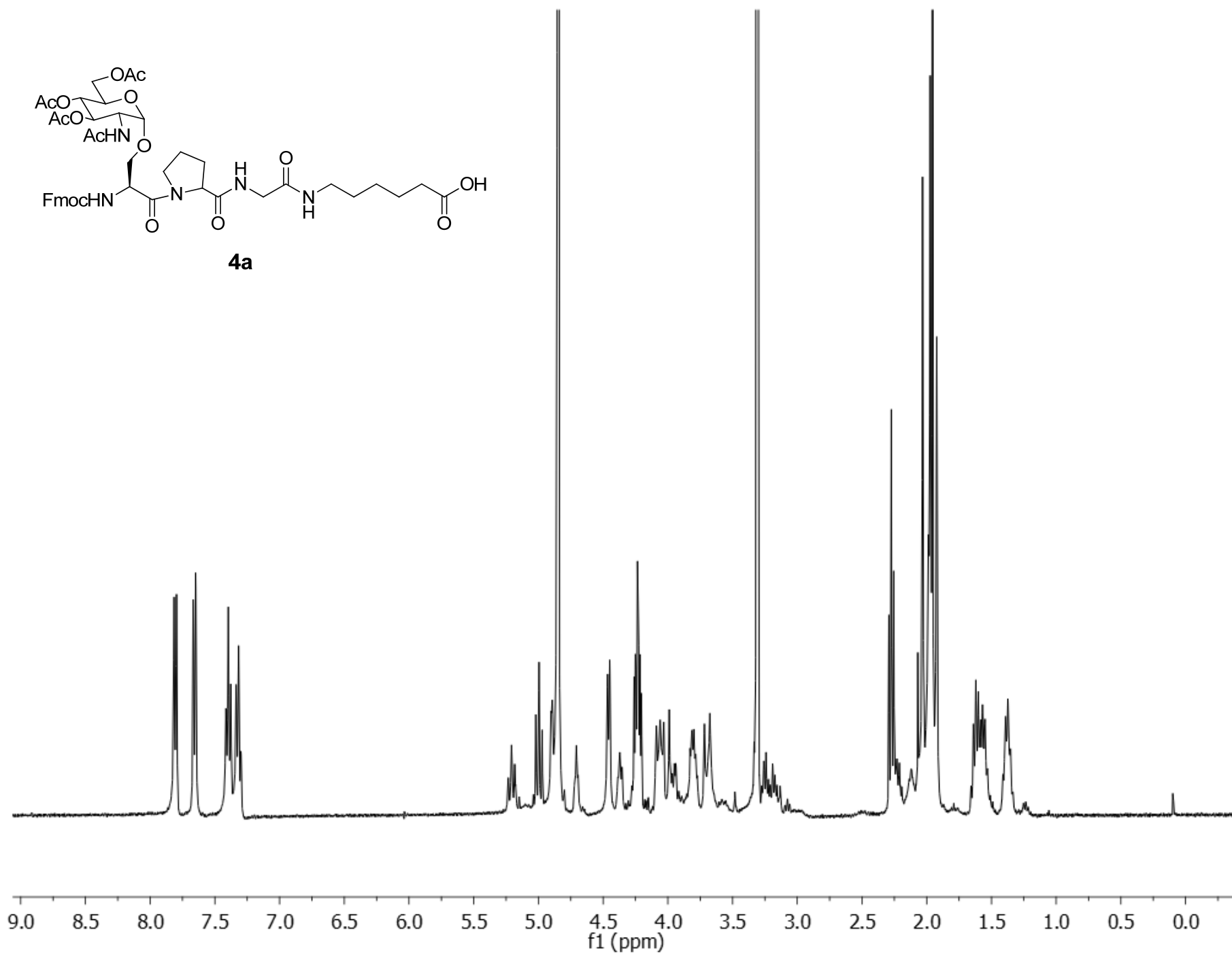


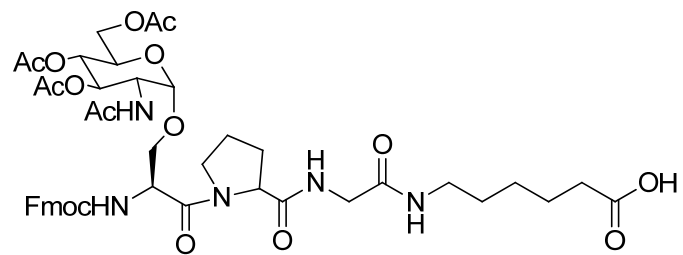




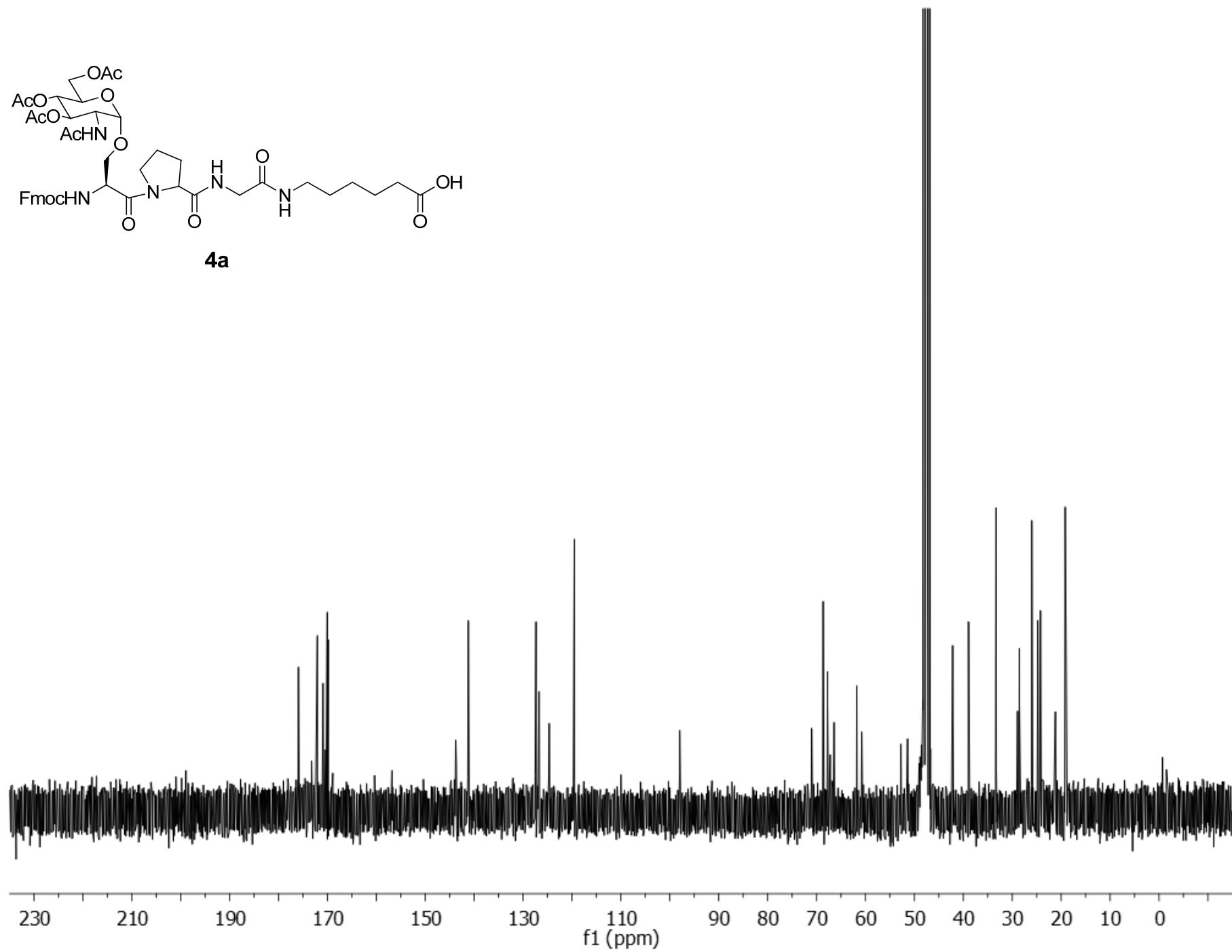


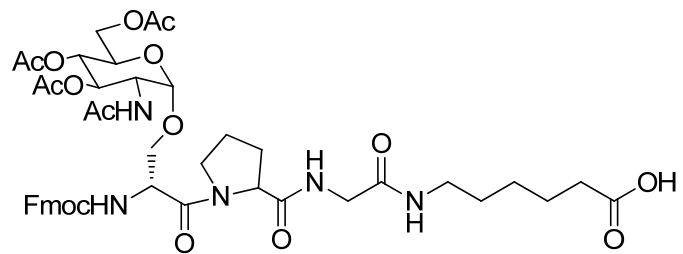
4a



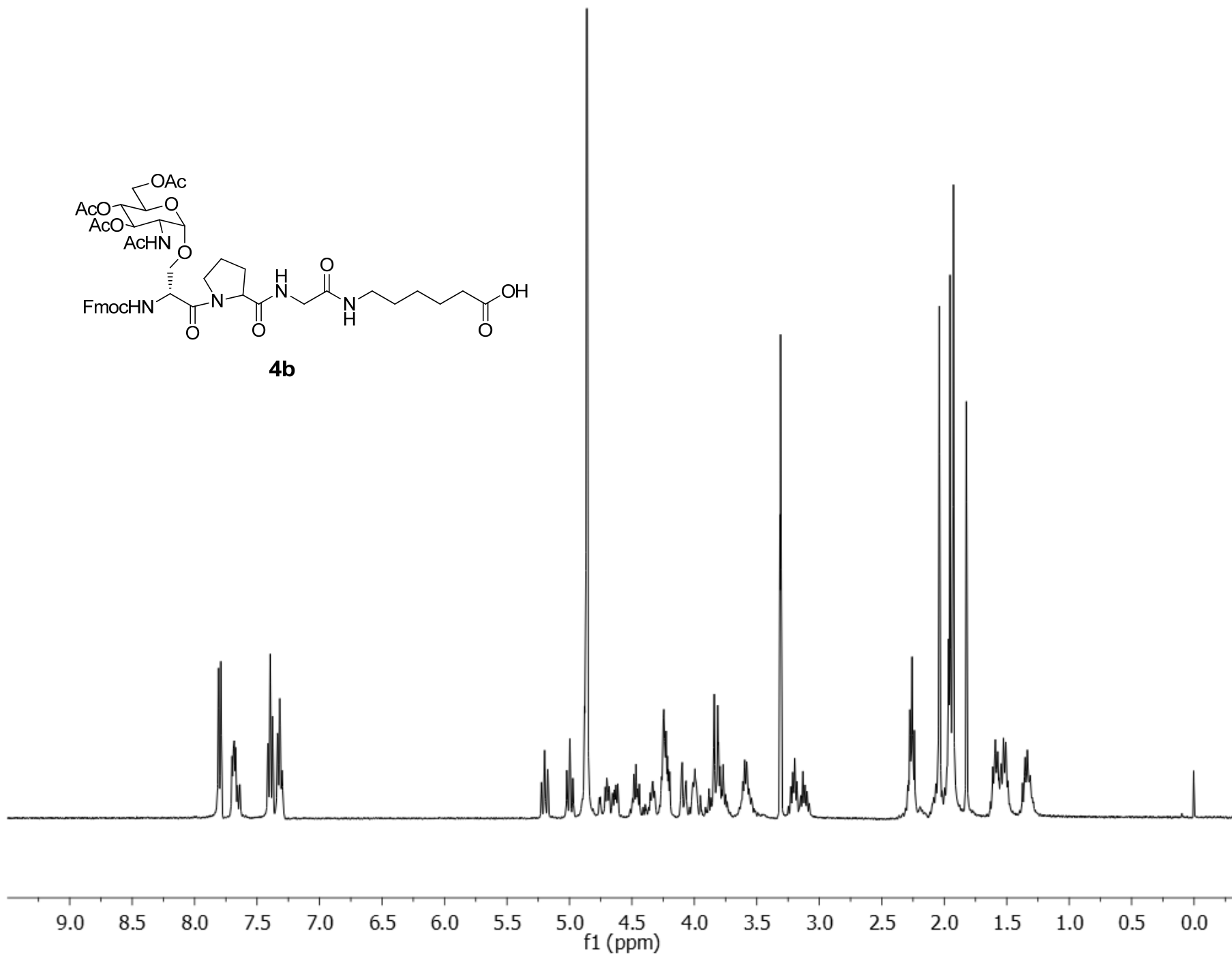


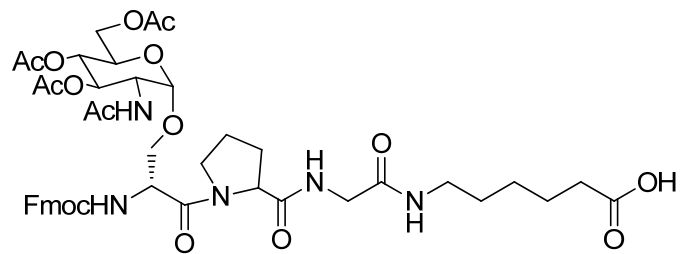
4a



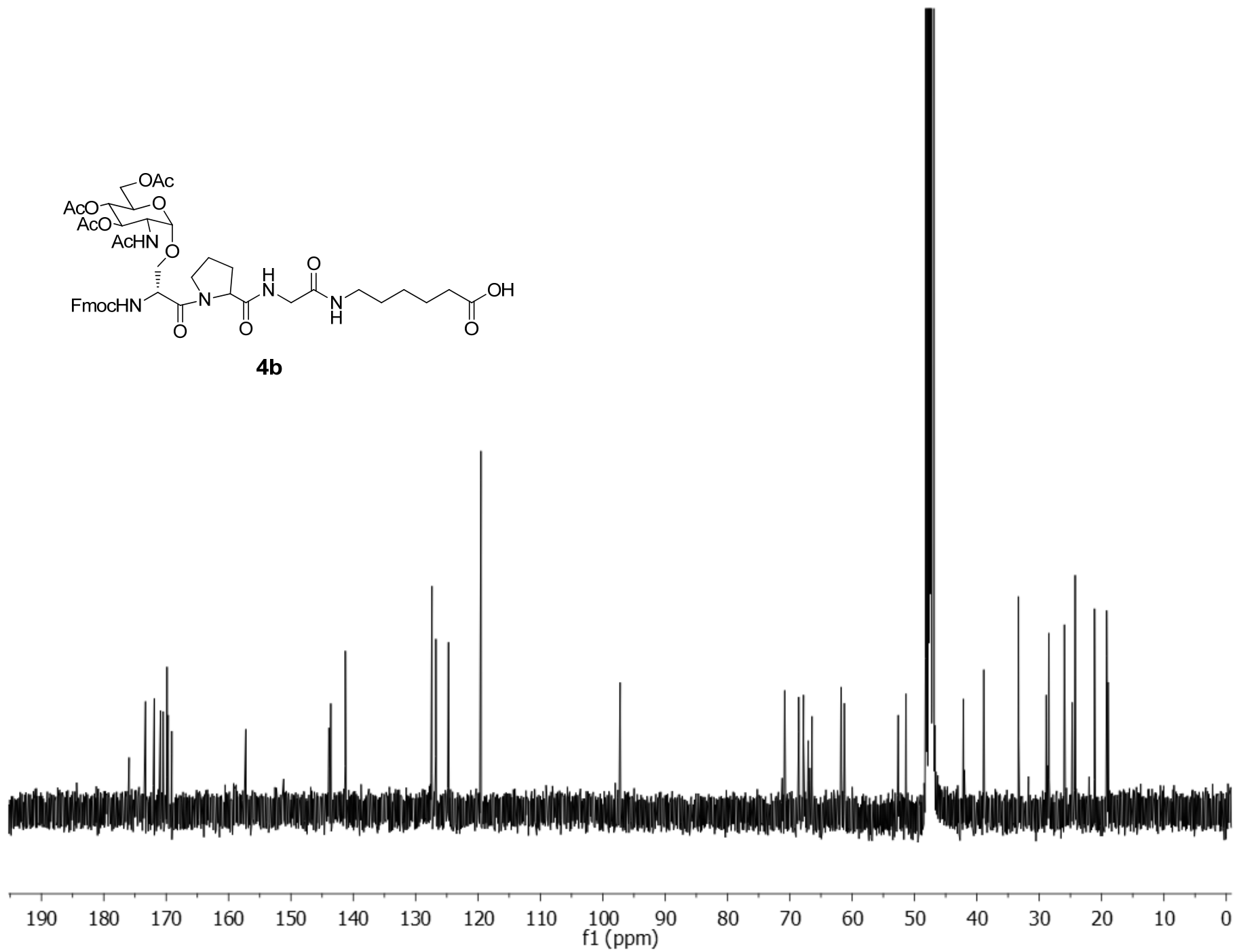


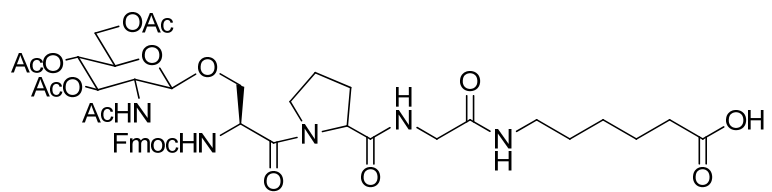
4b



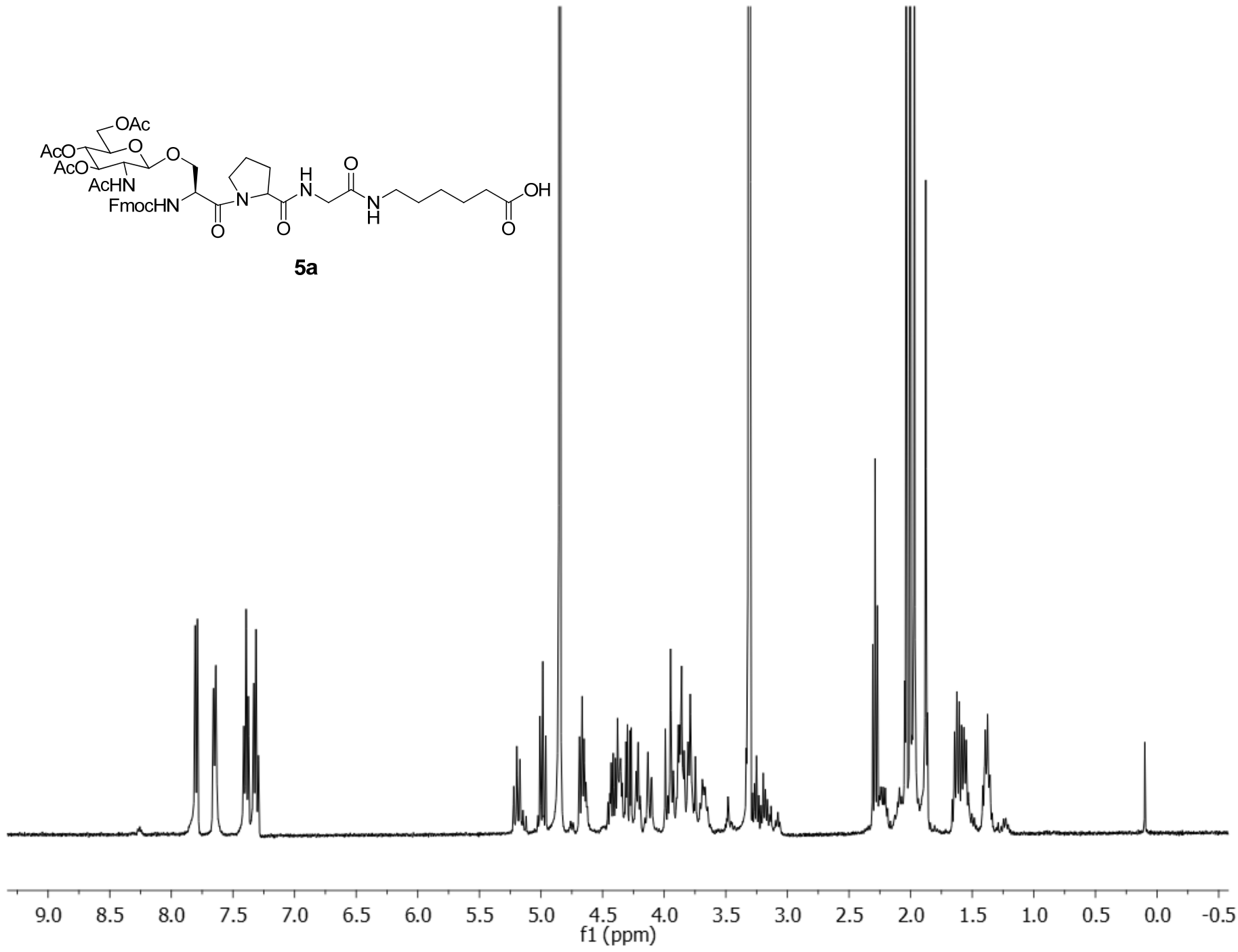


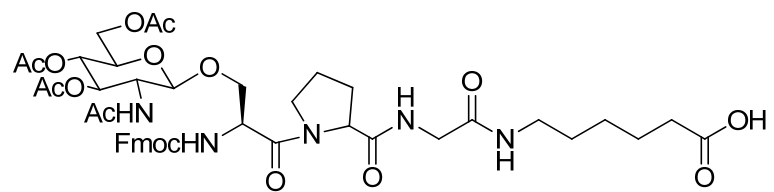
4b



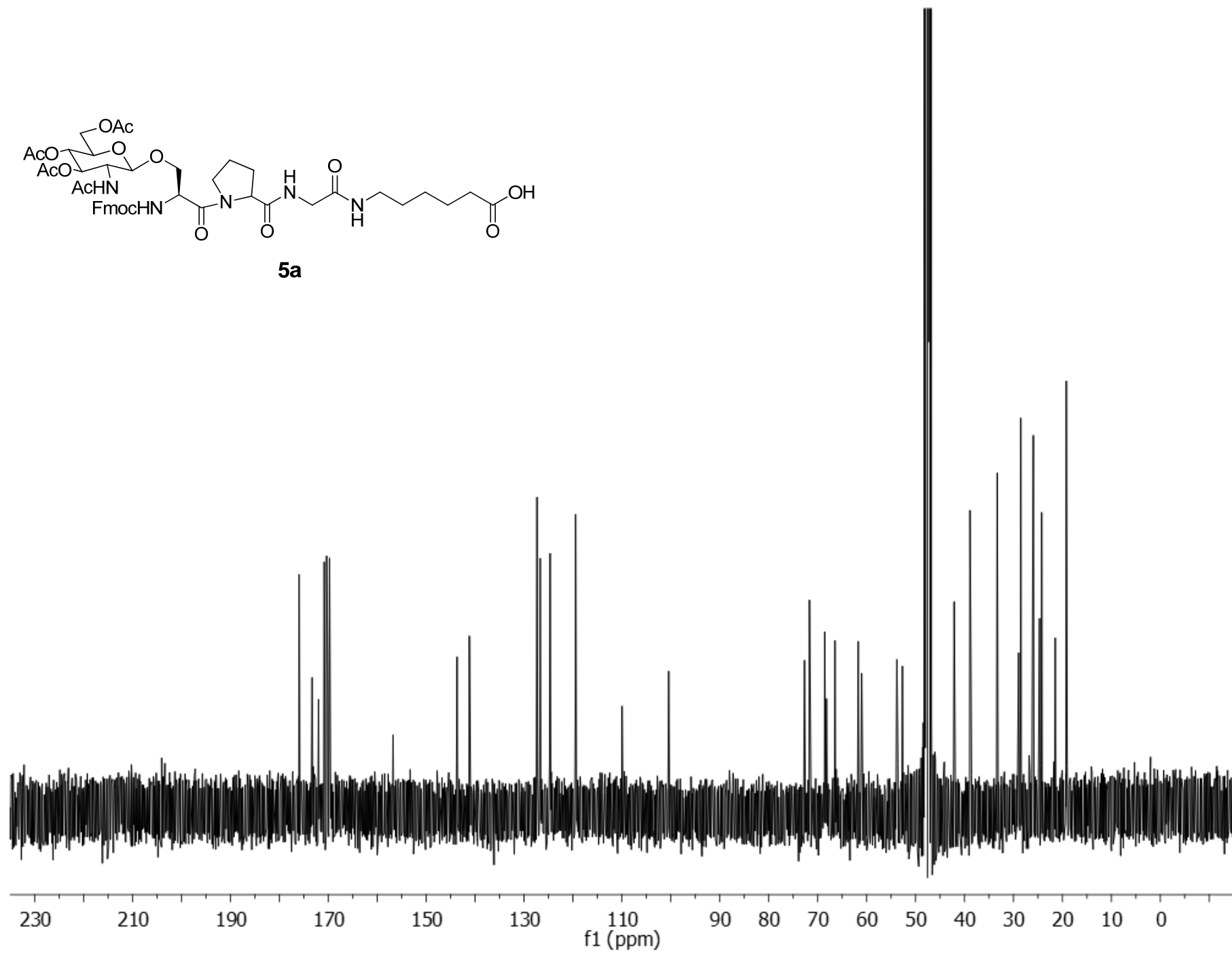


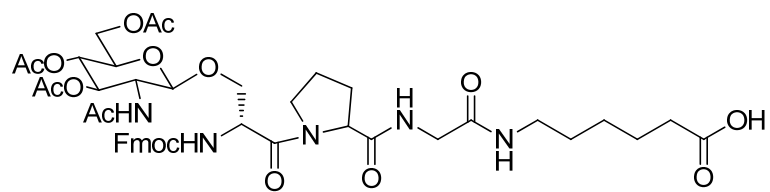
5a



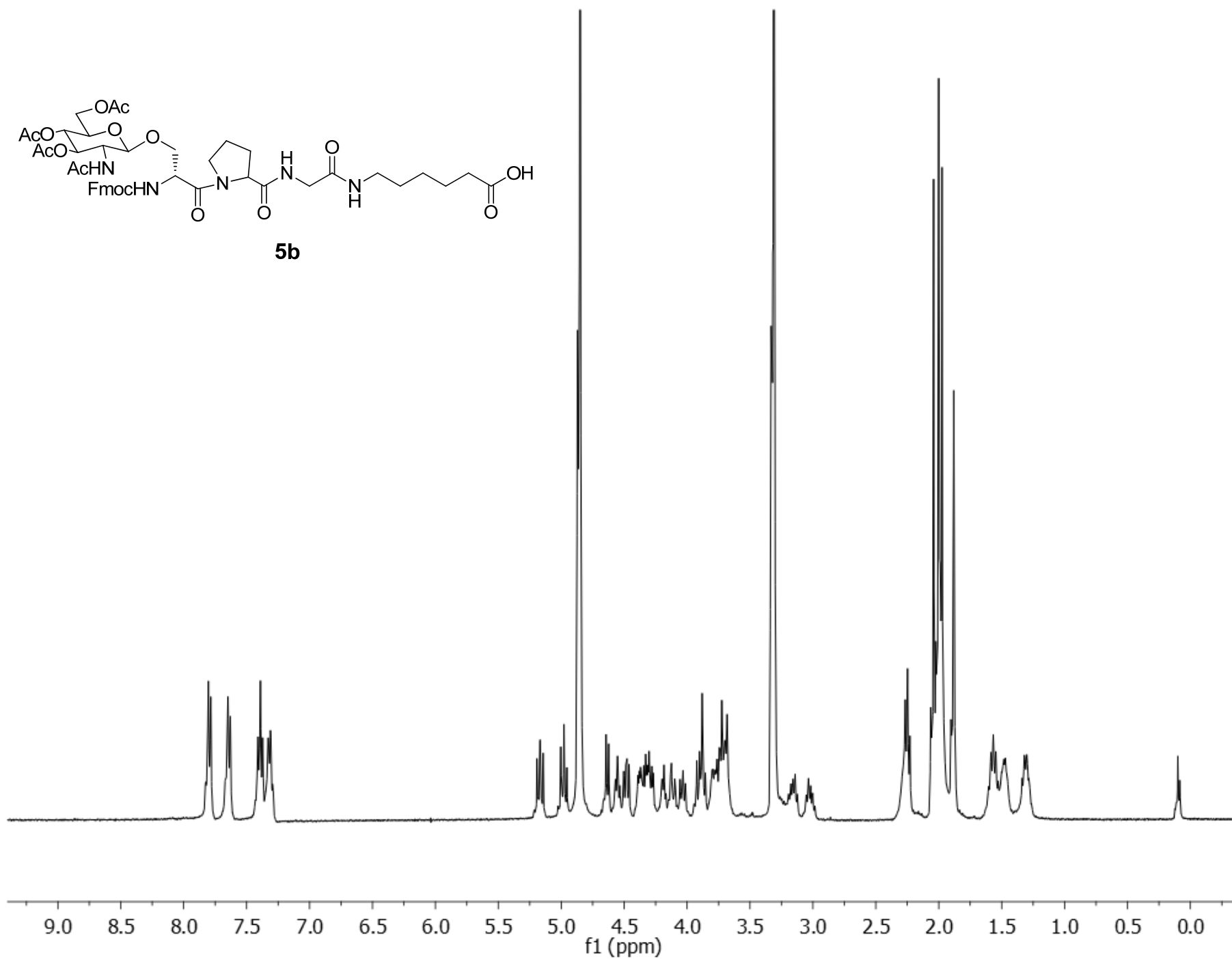


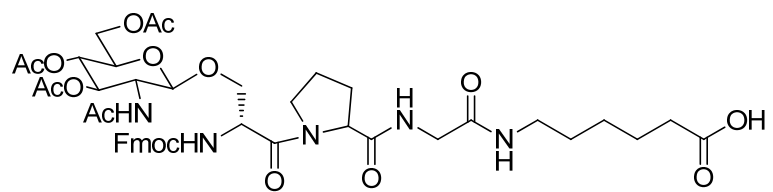
5a





5b





5b

