

Supporting Information

O'Neill et al. 10.1073/pnas.1121549109

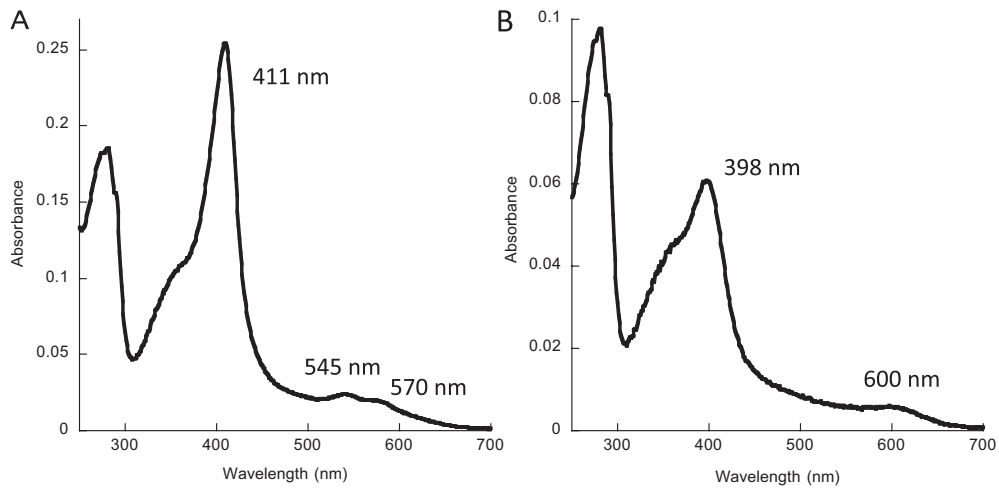


Fig. S1. Absorption spectra of holo-PhuS wild type (A) and H209/210/212A (B) proteins. Cuvettes contained 10 μ M holo-PhuS in 20 mM Tris•HCl (pH 7.5).

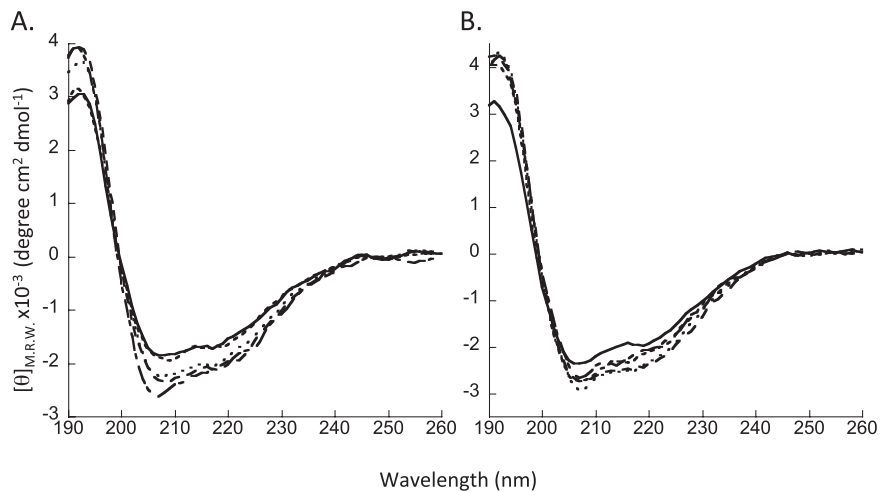


Fig. S2. Circular dichroism spectra of the wild-type and His-mutant apo-PhuS (A) and holo-PhuS (B) proteins. PhuS wild type (—), H209A (---), H210A (- - -), H212A (· · · · ·), H209/210/212A (— · — · —); spectra of PhuS (2.5 μ M) were recorded in 1 mM sodium phosphate (pH 7.4) at 25 $^{\circ}$ C.

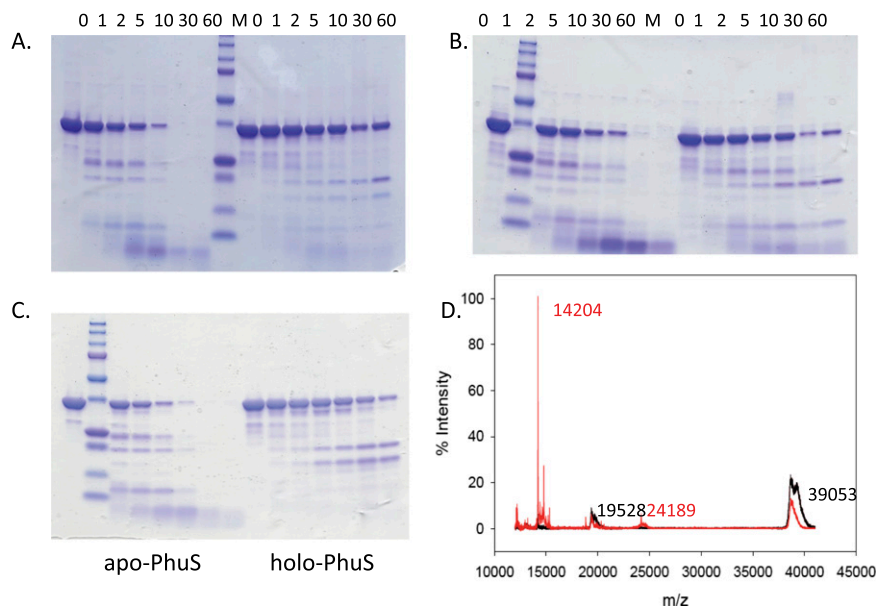


Fig. S5. Limited proteolysis and MALDI-MS spectrum of apo- and holo-PhuS mutants. SDS/PAGE of apo- and holo-PhuS H210A (A), apo- and holo-PhuS H209A (B), and apo- and holo-PhuS H212A (C), after proteolysis at intervals up to 60 min. (D) MALDI-MS spectrum of apo- and holo-PhuS H209/210/212A proteolysis after 2 min. The major peptide fragments are highlighted and color coded (red for apo-PhuS and black for holo-PhuS). The peak at ≈ 39 kDa corresponds to intact PhuS (M+1) and 19528 (M+2).

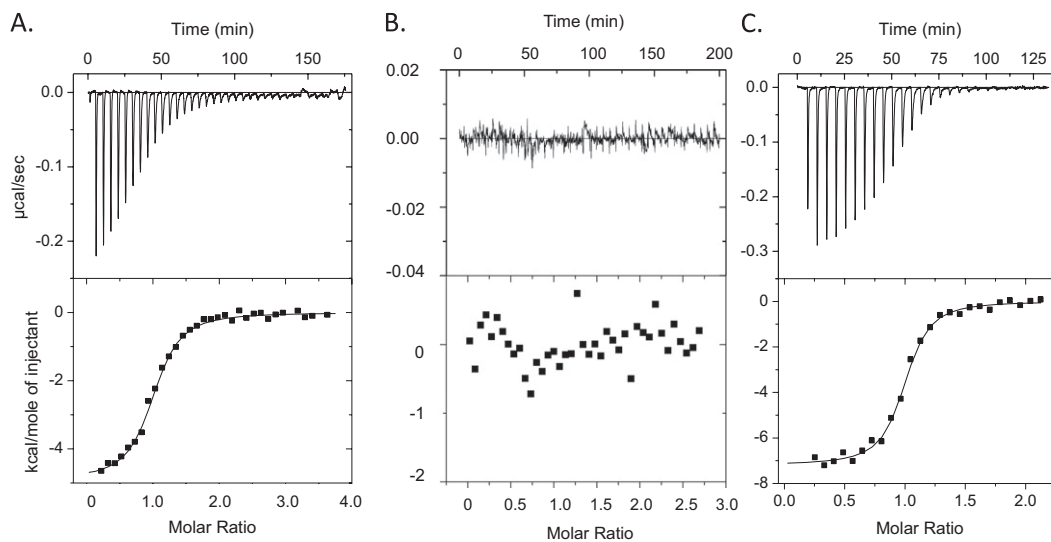


Fig. S6. Isothermal titration calorimetry (ITC) analysis of the interaction of holo-PhuS (A), apo-PhuS (B), and holo-PhuS H209A (C) with apo-HemO. Titrations were performed in 20 mM sodium phosphate (pH 7.5) at 298 K. *Upper:* Time-dependent release of heat during the titration. *Lower:* Peak integrals as a function of the molar ratio of heme to protein. The data were fit to a single binding model with Origin software, supplied by Microcal.