Site-Selective Bromination of Vancomycin

Tejas P. Pathak and Scott J. Miller*

Department of Chemistry, Yale University, New Haven, Connecticut, 06520

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General Information

Proton NMR spectra were collected on 500 MHz spectrometers at 25 °C. Proton chemical shifts are reported in ppm (δ) relative to the solvent reference relative to TMS employed as the internal standard (CDCl₃, δ 7.26 ppm; DMSO δ 2.50 ppm; D₂O, δ 4.79 ppm, CD₃OD δ 3.34 ppm). Data are reported as follows: chemical shift (multiplicity [singlet (s), doublet (d), triplet (t), quartet (q), and multiplet (m)], coupling constants [Hz], integration). Infrared spectra were obtained using a FT-IR instrument utilizing a Small Orbit FT-IR ATR (Attenuated total reflectance) module. Specific rotations were determined with a Polarimeter 341 at 20 °C on the sodium D line (path length 10.0 cm). High-resolution mass spectra were obtained from institutional providers (high resolution LCMS) and the method of ionization reported herein. All solvents were either distilled or taken from a solvent purification system except H₂O, in which case DI water was used as a solvent. Vancomycin•HCl was purchased from commercial supplier (95% purity by HPLC analysis at 280 nm) and was used without further purification. No precautions were taken to exclude air unless otherwise noted. MIC determinations were performed by Micromyx, LLC (Kalamazoo, MI, USA). All other reagents were purchased from commercial sources and used without further purification. *N*-bromopthalimide (NBP) was recrystallized and was kept away from light.

Note: Vancomycin and all the reported brominated derivatives were kept in a freezer (at -20 °C) to avoid decomposition.



Procedure for Unselective Bromination of Vancomycin (4), Figure 2:

To a round-bottom flask equipped with a stir bar were added 500.0 mg (0.336 mmol, 1.0 equiv) of vancomycin•HCl (4). Water (10.0 mL) and MeOH (2.0 mL) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min after which 150.0 mg (0.672 mmol, 2.0 equiv) of NBP were added. The reaction mixture was stirred at room temperature. After 20 h, 10 mL of EtOAc were added and the reaction was stirred for an additional 15 min. The organic layer was discarded and the aqueous layer was diluted with 2 mL of water. The reaction was analyzed by reverse phase HPLC (see below) and was purified by preparative reverse phase HPLC (see below).

Purification of the reaction mixture yielded, 60.0 mg (11% Yield) of 7_f -Br vancomycin (5), 58.0 mg (11% Yield) of 7_d -Br vancomycin (6), and 131.1 mg (23% Yield) of $7_{d,f}$ -Br₂ vancomycin (8).

Each of these isomers was characterized based on change in characteristic chemical shifts in their HSQC spectra. HSQC spectra of compound **5**, **6**, and **7** were recorded in both D_2O and DMSO- d_6 and were compared with HSQC spectrum of native vancomycin (**4**). The raw spectral data are presented, including overlays with vancomycin. In the case of 7_d -Br, we also observed ROESY correlation between 7_f -H with X_7 -H indicating the presence of 7_f -H, thus supporting substitution at the 7_d site of resorcinol ring. A similar ROESY correlation was absent in case of compound **5**.

HPLC Method for preparative purification: Symmetry prep C8 7μ m (19 X 300 mm) column, 24 mL/min, starts at 5% MeCN (0.1% HCOOH) : 95% H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min.

Analytical method: Symmetry prep C8 7 μ m column, 4 mL/min, starts at 5% MeCN (0.1% HCOOH) : H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min.

Optimization of reaction conditions (Manuscript Table 1):

	Vance		3P	_		Veee
H	Vanco Vanco Vanco Vanco Vanco Vanco Vanco H, Vanco H, Vanco H, Vanco H, Vanco H, Vanco H, Vanco H, H, HO ₂ C		Vanco 5 OH OH Br	o Van HO₂C ^y Br- ⊦	NH	Vanco J5 OH Br
Entry	Catalyst	mol%	t (h)	% conv.	5 7 _f Br:	6 8 7 _d Br : 7 _{d,f} Br ₂
1	No catalyst	NA	12	71	1.0 :	: 1.0 : 1.3 ^b
2a	N,N-dimethylacetamide	100	12	68	1.0 :	1.0 : 1.8 ^b
2b	N,N-dimethylacetamide	7100	16	64	1.0 :	1.5 : 2.9 ^b
3	Boc-Asn(Me ₂)-DAla-DAla-OH (10)	100	2	97	1.0 :	6.8 : 2.6 ^b
4	Boc-DAsn(Me ₂)-DAla-DAla-OH (11)	100	2	90	1.0 :	4.9:1.8 ^b
5	Boc-Gln(Me ₂)-DAla-DAla-OH (12)	100	2	99	1.0 :	4.0:9.3 ^b
6	Boc-DGIn(Me ₂)-DAIa-DAIa-OH (13)	100	2	99	1.0 :	3.0 : 3.4 ^b
7	Boc-Leu-DAla-DAla-OH (14)	100	12	60	1.0 :	5.7 : 2.5 ^b
8a		100	1.5	98	1.0 :	14.6 : 2.8°
8b	Boc-Asn(Me ₂)-DAla-DAla-OH (10) {	200	1.5	99	1.0 :	19.0 : 6.0 ^c
8c		50	2	97	1.0	3.4:1.1°
9	Boc-Leu-DAsn(Me ₂)-DAla-OH (15)	200	12	96	1.0 :	6.5 : 2.7 ^c
10	Boc-Leu-DAla-DAsn(Me ₂)-OH (16)	200	12	85	1.0	3.5 : 1.0 ^c
11	Boc-Asn(Me ₂)-DAla-DAla-OH (10)	100	1.5	16	1.0 :	10.5 : 0.9 ^d
12	Boc-Leu-DAla-DAsn(Me ₂)-OH (16)	100	1.5	12	1.0 :	3.5 : 1.6 ^d

a) Ratios were measured by HPLC at 280 nm wavelength. b) 2.0 equiv of NBP, 250 μ L of water, 50 μ L of MeOH, 8 μ mol of 4. c) 2.0 equiv of NBP, 1000 μ L of water, 200 μ L of MeOH, 8 μ mol of 4. d) 50 mol% NBP, 0.033 mmol of 4.

Note: Conversions and product ratios are uncorrected. The product ratios are highly dependent on reaction time, as each monobrominated product can be further functionalized. Thus vigorously following reaction by LC-MS, is recommended to obtained the most reproducible product distribution. Data presented in Table 1 were recorded at the indicated time. For larger scale reactions reaction progress was monitored by LCMS to obtain optimal product distribution. Conversion is defined as (100 – area % of 4).

Analytical method for analysis of reaction mixture: Symmetry prep C8 7 μ m column, 4 mL/min, starts at 5% MeCN (0.1% HCOOH) : H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min.

General Procedure for Optimization Table:

Entry 1: To a HPLC vial equipped with a stir bar were added 12.5 mg (0.008 mmol, 1.0 equiv) of vancomycin•HCl. Water (250 μ L) and MeOH (50 μ L) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 2.0 equiv) of NBP were added. Aliquots were collected at regular time intervals and analyzed by reverse phase HPLC.

Entry 2a: To a HPLC vial equipped with stir bar were added 12.5 mg (0.008 mmol, 1.0 equiv) of vancomycin•HCl and 250 μ L of DMA (*Standard solution was prepared as follows: 50 \muL of DMA was added to 16.8 mL of water and 250 \muL of this mixture was used as a solvent for the reaction*). To this 50 μ L of MeOH were added. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 2.0 equiv) of NBP were added. Aliquots were collected at regular time intervals and were analyzed by reverse phase HPLC.

Entry 2b: To a HPLC vial equipped with stir bar were added 12.5 mg (0.008 mmol, 1.0 equiv) of vancomycin•HCl, 250 μ L of H₂O, 50 μ L of MeOH and 50 μ L of DMA were added. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 2.0 equiv) of NBP were added. Aliquots were collected at regular time intervals and were analyzed by reverse phase HPLC.

Entry 3: To a HPLC vial equipped with stir bar were added 12.5 mg (0.008 mmol, 1.0 equiv) of vancomycin•HCl and 3.2 mg (0.008 mmol, 1.0 equiv) of peptide **10**. Water (250 μ L) and MeOH (50 μ L) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 2.0 equiv) of NBP were added. An aliquot was collected at 120 min and analyzed by reversed phase HPLC.

Entry 4: The same procedure as entry 2 was employed, except peptide 11 was used.

Entry 5: The same procedure as entry 2 was employed, except peptide 12 was used.

Entry 6: The same procedure as entry 2 was employed, except peptide 13 was used.

Entry 7: The same procedure as entry 2 was employed, except peptide 14 was used.

Entry 8a: To a HPLC vial equipped with stir bar were added 12.5 mg (0.008 mmol, 1.0 equiv) of vancomycin•HCl and 3.2 mg (0.008 mmol, 1.0 equiv) of peptide **10**. Water (1000 μ L) and MeOH (250 μ L) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 2.0 equiv) of NBP were added. An aliquot was collected at 90 min and analyzed by reverse phase HPLC.

Entry 8b: To a HPLC vial equipped with stir bar were added 12.5 mg (0.008 mmol, 1.0 equiv) of vancomycin•HCl and 6.4 mg (0.016 mmol, 2.0 equiv) of peptide **10**. Water (1000 μ L) and MeOH (250 μ L) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 2.0 equiv) of NBP were added. An aliquot was collected at 90 min and analyzed by reverse phase HPLC.

Entry 8c: To a HPLC vial equipped with stir bar were added 12.5 mg (0.008 mmol, 1.0 equiv) of vancomycin•HCl and 1.6 mg (0.004 mmol, 0.5 equiv) of peptide **10.** Water (1000 μ L) and MeOH (250 μ L) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 2.0 equiv) of NBP were added. An aliquot was collected at 90 min and analyzed by reverse phase HPLC.

Entry 9: The same procedure as entry 2 was employed, except peptide 15 was used.

Entry 10: The same procedure as entry 2 was employed, except peptide 16 was used.

Entry 11: To a HPLC vial equipped with stir bar were added 49.0 mg (0.033 mmol, 1.0 equiv) of vancomycin•HCl and 13.2 mg (0.033 mmol, 1.0 equiv) of peptide **10**. Water (4000 μ L) and MeOH (800 μ L) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 0.5 equiv) of NBP were added. An aliquot was collected at 90 min and analyzed by reverse phase HPLC.

Entry 12: To a HPLC vial equipped with stir bar were added 49.0 mg (0.033 mmol, 1.0 equiv) of vancomycin•HCl and 14.9 mg (0.033 mmol, 1.0 equiv) of peptide **16**. Water (4000 μ L) and MeOH (800 μ L) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 0.5 equiv) of NBP were added. An aliquot was collected at 90 min and analyzed by reverse phase HPLC.





The same procedure as entry 8a (Table 1) was employed, except appropriate reagents. We selected NBP because it gives conversion of vancomycin (in 90 min) compared to other brominating reagents. Moreover, the resultant phthalimide is insoluble in water and could easily be removed *via* simple wash with EtOAc, making the purification much easier on larger scale reactions.

Procedure for Bromination of Vancomycin (4) Targeting Analog 6 (Manuscript Figure 5):



To a round bottom flask equipped with a stir bar were added 100.0 mg (0.067 mmol, 1.0 equiv) of vancomycin•HCl and 26.9 mg (0.067 mmol, 1.0 equiv) of peptide **10**. Water (8.4 mL) and MeOH (1.6 mL) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min after which 30.7 mg (0.135 mmol, 2.0 equiv) of NBP were added. The reaction mixture was stirred at room temperature for *ca*. 90 min, until complete conversion of vancomycin was observed by LC-MS (over the period of time the reaction became cloudy). To the reaction vessel, 10 mL of EtOAc was added and reaction was stirred for an additional 15 min. The organic layer was discarded and the aqueous layer was diluted with 2 mL of water. The reaction was analyzed by reverse phase HPLC and was purified by preparative reverse phase HPLC. Purification of reaction mixture yielded, 43.1 mg (41% Yield) 7_d -Br vancomycin (**6**). ES-HRMS (M+H⁺): Predicted 1526.3485, observed 1526.3184.

HPLC Method for preparative purification: Symmetry prep C8 7μ m (19 X 300 mm) column, 24 mL/min, starts at 5% MeCN (0.1% HCOOH) : 95% H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min.

Analytical method: Symmetry prep C8 7 μ m column, 4 mL/min, starts at 5% MeCN (0.1% HCOOH) : H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min. *Retention time of 6 on C8 column is 15.3 min.*

HPLC trace for the reaction mixture (at 280 nm):



In lieu of tabulated NMR data, please see raw spectral data provided below.







1533.3700

1530 1532

1534 1536 1538

m/z



5

f1 (ppm)



4.5 4**S**11 f2 (ppm) 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 3.5 3.0 2.5 2.0 1.5

Procedure for Dibromination of Vancomycin (4) Targeting Analog 8 (Manuscript Figure 6):

To a round bottom flask equipped with a stir bar were added 100.0 mg (0.067 mmol, 1.0 equiv) of vancomycin•HCl and 26.9 mg (0.067 mmol, 1.0 equiv) of peptide **10**. Water (2.0 mL) and MeOH (0.4 mL) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 45.8 mg (0.201 mmol, 3.0 equiv) of NBP were added. The reaction mixture was stir at room temperature for *ca*. 60 min (over the period of time the reaction turn cloudy). To the reaction vessel, 10 mL of EtOAc was added and the reaction was stirred for an additional 15 min. The organic layer was discarded and the aqueous layer was diluted with 2 mL of water. The reaction was analyzed by reverse phase HPLC and was purified by preparative reverse phase HPLC. Purification of reaction mixture yielded 60.9 mg (55% Yield) $7_{d,f}$ –Br₂ vancomycin (**8**). ES-HRMS (M+H⁺): Predicted 1604.2590, observed 1604.2281.

HPLC Method for preparative purification: Symmetry prep C8 7 μ m (19 X 300 mm) column, 24 mL/min, starts at 5% MeCN (0.1% HCOOH) : 95% H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min.

Analytical method: Symmetry prep C8 7 μ m column, 4 mL/min, starts at 5% MeCN (0.1% HCOOH) : H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min. *Retention time of 8 on C18 column is 20.1 min and retention time on C8 column is 13.3 min.*

HPLC trace for the reaction mixture (at 280 nm):

In lieu of tabulated NMR data, please see raw spectral data provided below.

(hpm)

Procedure for tribromination of Vancomycin (4) Targeting Analog 17 (Manuscript Figure 7):

To a round bottom flask equipped with a stir bar were added 49.0 mg (0.033 mmol, 1.0 equiv) of vancomycin•HCl and 15.1 mg (0.033 mmol, 1.0 equiv) of peptide **18**. Water (1.0 mL) and MeOH (0.2 mL) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 30.1 mg (0.132 mmol, 4.0 equiv) of NBP were added. The reaction mixture was stir at room temperature for *ca*. 120 min (over the period of time the reaction became cloudy). To the reaction vessel, 10 mL of EtOAc were added and the reaction was stirred for an additional 15 min. The organic layer was discarded and the aqueous layer was diluted with 2 mL of water. The reaction was analyzed by reverse phase HPLC and was purified by preparative reverse phase HPLC. Purification of reaction mixture yielded, 20.3 mg (35% Yield) tribrominated vancomycin (**17**). ES-HRMS (M+H⁺): Predicted 1682.1695, observed 1682.1954.

HPLC Method for preparative purification: Symmetry prep C8 7 μ m column (19 X 300 mm), 24 mL/min, starts at 5% MeCN (0.1% HCOOH) : 95% H₂O (0.1% HCOOH) for 3 min, 5 to 15% MeCN over 20 min, 15 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min.

Analytical method: Symmetry prep C8 7 μ m column, 4 mL/min, starts at 5% MeCN (0.1% HCOOH) : 95% H₂O (0.1% HCOOH) for 3 min, 5 to 15% MeCN over 20 min, 15 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min. Retention time of **17** on C8 column is 20.7 min.

HPLC trace for the reaction mixture (at 280 nm)

In lieu of tabulated NMR data, please see raw spectral data provided below.

HPLC traces of uncatalyzed (top) and peptide-catalyzed (middle and bottom) tribromination of vancomycin obtained on reverse phase HPLC.

S20

DMSO-d₆, Overlay of HSQC of tri-brominated vancomycin (red) with vancomycin (gray)

f1 (ppm)

Procedure for Bromination of Vancomycin (4) Targeting Analog 5.

To a round bottom flask equipped with a stir bar were added 100.0 mg (0.067 mmol, 1.0 equiv) of vancomycin•HCl and 114.5 mg (1.206 mmol, 18.0 equiv) of peptide guanidine•HCl. MeOH (16.0 mL) was added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min after which 61.6 mg (0.27 mmol, 4.0 equiv) of NBP were added. The reaction mixture was stirred at room temperature for 4 h (solution remained cloudy through out the course of a reaction). To the reaction vessel, 10 mL of H₂O was added and reaction was stirred for an additional 15 min. The MeOH was removed in vacuo and reaction was analyzed by reverse phase HPLC and was purified by preparative reverse phase HPLC. Purification of reaction mixture yielded, 23.3 mg (21% Yield) 7_f -Br Vancomycin (**5**). ES-HRMS (M+H⁺): Predicted 1526.3485, observed 1526.3184.

HPLC Method for preparative purification: Symmetry prep C8 7μ m (19 X 300 mm) column, 24 mL/min, starts at 5% MeCN (0.1% HCOOH) : 95% H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min.

Analytical method: Symmetry prep C8 7 μ m column, 4 mL/min, starts at 5% MeCN (0.1% HCOOH) : H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 95% MeCN over 3 min, hold at 95% MeCN for 3 min, 95% to 5% over 3 min. Retention time of **5** on C8 column is 11.6 min.

HPLC trace for the reaction mixture *with guanidine* (at 280 nm):

In lieu of tabulated NMR data, please see raw spectral data provided below.

5 4.5 12 (pt 826 8.0 7.5 5.5 5.0 2.5 1.0 7.0 6.5 6.0 3.5 3.0 2.0 1.5

f1 (ppm)

f1 (ppm)

Peptide Synthesis:

Synthesis of Peptide 10: To a dry 50 mL RBF equipped with stir bar were added 541.0 mg (2.86 mmol, 1.0 equiv.) of Boc-DAla-OH, 1.00 g (3.15 mmol, 1.1 equiv) of TsOH•H-DAla-OBzl, 819.0 mg (3.15 mmol, 1.1 equiv) of EDCI•HCl and 664.0 mg (3.15 mmol, 1.1 equiv) of HOBt•H₂O. To this, 10 mL of DCM was added and the reaction mixture was stir for ca.10 min. At this point, 700 µL (4.29 mmol, 1.5 equiv) of DIPEA were added. The reaction mixture was allowed to stir overnight at room temperature and reaction progress was monitored by LCMS. Upon complete conversion of starting material, 10 mL of 10% ag. citric acid solution (w/w) was added and reaction was stirred for 5 min. The reaction mixture was transferred to separatory funnel and was diluted with 70 mL of EtOAc. The aqueous layer was removed and organic layer was washed again with 10 mL of 10% ag. citric acid solution. Similarly, the organic layer was washed with 10 mL of saturated aq. NaHCO₃ (2 times) and 20 mL of Brine. The organic layer was dried over Na_2SO_4 and concentrated in vacuo to give a white solid (S1). This was treated with 4 M HCl in dioxane (5 mL) for 30 min followed by concentration in vacuo to provide HCl salt of S2 as a white solid (700 mg). This material was carried forward without purification. In a dry 50 mL RBF equipped with stir bar were added 192.0 mg (0.77 mmol, 1.0 equiv.) of S2, 200.0 mg (0.77 mmol, 1.0 equiv) of Boc-Asn(Me₂)-OH, 162.0 mg (0.85 mmol, 1.1 equiv) of EDCI•HCl and 130.0 mg (0.85 mmol, 1.1 equiv) of HOBt \cdot H₂O. To this 5 mL of DCM was added and reaction mixture was stir for ca.10 min. At this point, 174 µL (1.00 mmol, 1.3 equiv) of DIPEA was added. The reaction mixture was allowed to stir overnight at room temperature and the reaction progress was monitored using LCMS. Upon complete conversion of starting material, 10 mL of 10% aq. citric acid solution was added and reaction was stir for 5 min. The reaction mixture was transfer to separating funnel and was diluted with 50 mL of EtOAc. The aqueous layer was removed and organic layer was washed again with 10 mL of 10% ag. citric acid solution, 10 mL of saturated ag. NaHCO₃ (2 times) and 20 mL of Brine. The organic layer was dried over Na₂SO₄ and was concentrated in vacuo to give colorless oil. The crude mixture was purified by Biotage[®] using water and acetonitrile as eluents to give 320.0 mg of S3 (84% Yield). Compound S3, 320.0 mg (0.65 mmol, 1.0 equiv) was dissolved in 10 mL of THF and carefully added to a flask containing 69.0 mg of Pd/C under N₂ atmosphere. The flask was evacuated and refilled with H₂ three times then stirred under H₂ atmosphere by using a H₂ balloon. Upon completion (by LCMS) the reaction mixture was passed through Celite[®] and was concentrated in vacuo to give peptide 10.

Boc-Asn(Me₂)-DAla-DAla-OH (10): $[\alpha]_{D}^{20}$ +342 (c 3.5, CHCl₃), ¹H NMR (500 MHz, CDCl₃): δ 7.45 (s, 1H), 6.80 (s, 1H), 5.69 (d, J = 9.4 Hz, 1H), 4.41 (m, 3H), 3.29 (dd, J = 17.1, 3.5 Hz, 1H), 2.91 (s, 3H), 2.79 (s, 3H), 2.52 (dd, J = 17.1, 4.1 Hz, 1H), 1.38 (s, 15H). ¹³C NMR (125 MHz, CDCl₃): 174.2, 173.1, 172.8, 171.3, 155.7, 49.7, 48.8, 37.4, 36.3, 35.7, 28.5, 17.4, 17.0. FT-IR: 1521, 1635, 1646, 1700, 2934, 2979, 3313. ESI-HRMS (M+H⁺): calc. 403.2124, obsrd. 403.1885.

Boc-Leu-DGln(Me)₂-DAla-OH (18): ¹H NMR (500 MHz, CD₃OD) δ 4.40 (dd, J = 8.0, 5.6 Hz, 1H), 4.34 (q, J = 7.3 Hz, 1H), 4.05 - 3.96 (m, 1H), 3.28 (dt, J = 3.3, 1.6 Hz, 3H), 3.01 (s, 3H), 2.90 (s, 3H), 2.58 - 2.45 (m, 1H), 2.45 - 2.34 (m, 1H), 2.12 (dt, J = 14.1, 7.0 Hz, 1H), 1.89 (dt, J = 13.9, 6.9 Hz, 1H), 1.65 (dt, J = 13.3, 6.6 Hz, 1H), 1.50 (dd, J = 12.0, 6.5 Hz, 2H), 1.41 (s, 12H), 0.93 (d, J = 6.6 Hz, 3H), 0.90 (d, J = 6.6 Hz, 3H). ¹³C NMR (126 MHz, CD₃OD) δ 174.35, 174.21, 173.07, 171.82, 156.43, 79.23, 53.61, 52.16, 40.46, 36.28, 34.43, 28.72, 27.30, 24.56, 21.96, 20.56, 16.04. ESI-HRMS (M+H⁺): calc. 459.2740, obsrd. 459.1924.

Compounds	S. aureus (standard) S. aureus ATCC ¹ 29213 MMX ² 100	S. aureus (MRSA) S. aureus ATCC ¹ 43300 MMX ² 2002	E. faecalis (standard) E. faecalis ATCC ¹ 29212 MMX ² 101	E. faecalis (VRE, VanB) <i>E. faecalis</i> ATCC ¹ 51299 MMX ² 202	E. faecalis (VRE, VanA) <i>E. faecalis</i> VanA MMX ² 486
6	2, 2	4, 2	4, 4	64, 64	>64, >64
5	4, 4	4, 4	8, 8	>64,>64	>64,>64
8	4, 4	8,4	2,4	>64, >64	>64,>64
17	4, 4	8, 8	2,4	64, 64	>64,>64
Vancomycin (4)	0.5, 0.5	1, 1	2, 2	16, 16	>64, >64

 Table 4. MIC Determinations for Newly Synthesized Vancomycin Analogs.

MSSA: methicillin-susceptible *S. aureus*, MRSA: methicillin-resistant *S. aureus*, VanS: vancomycin-susceptible, VanB: VanB resistance genotype/phenotype, VanA: VanA resistance genotype/phenotype

¹American Type Culture Collection

²Micromyx Isolate Number

³CLSI acceptable limits for QC organisms (*S. aureus* ATCC 29213 and *E. faecalis* ATCC 29212) are shown in parentheses where applicable

2.5 7.5 7.0 6.5 5.5 5.0 4.0 f1 (ppm) 3.5 2.0 1.5 1.0 3.0 6.0 4.5

Synthesis of Vancomycin Methyl Ester:

Procedure is adapted from; Sundram, U. N.; Griffin, J. H. J. Org. Chem., 1995, 60, 1102.

To a round bottom flask equipped with a stir bar were added 200.0 mg (0.134 mmol, 1.0 equiv) of vancomycin•HCl, 80.0 mg (0.206 mmol, 1.5 equiv) of HBTU and 29.0 mg (0.206 mmol, 1.5 equiv) of HOBt. DMSO (2.0 mL), DMF (2.0 mL) and MeOH (2.0 mL) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min after which 65.0 μ L (0.40 mmol, 3.0 equiv) of DIPEA were added. The reaction mixture was stirred at room temperature for 5 h (during the reaction the solution became clear and turned pale yellow). To the reaction vessel, 10 mL of H₂O was added and the reaction was stirred for an additional 15 min. Then, MeOH was removed *in vacuo* and the reaction mixture was analyzed by reverse phase HPLC and was purified by preparative reverse phase HPLC to give 32 mg of S4.

HPLC Method for preparative purification: Symmetry prep C8 7μ m (19 X 300 mm) column, 24 mL/min, starts at 5% MeCN (0.1% HCOOH) : 95% H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 50% MeCN over 3 min, 50 to 95% MeCN over 3 min hold at 95% MeCN for 3 min, 95% to 5% over 3 min.

Analytical method: Symmetry prep C8 7 μ m column, 4 mL/min, starts at 5% MeCN (0.1% HCOOH) : 95% H₂O (0.1% HCOOH) for 3 min, 5 to 8.5% MeCN over 15 min, 8.5 to 50% MeCN over 3 min, 50 to 95% MeCN over 3 min hold at 95% MeCN for 3 min, 95% to 5% over 3 min. Retention time for desired product is 18.6 min.

The ¹H-NMR of the product was compared to the tabulated 1H NMR data reported in the study of Nicolaou. *Nicolaou, K. C.; Mitchell, H. J.; Jain, N. F.; Bando, T.; Hughes, R.; Winssinger, N.; Natarajan, S.; Koumbis, A. E. Chem. Eur. J.* **1999**, *5*, 2648.

In addition, we performed additional NMR experiments to assess the assignment. See below for raw proton, NOESY, HSQC data.

HPLC Trace:

Bromination of Vancomycin Methyl Ester (S4):

Control: To a 2.5 dram vial equipped with a stir bar were added 12.0 mg (0.008 mmol, 1.0 equiv) of vancomycin methyl ester (**S4**). MeOH (2 mL) was added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, when 7.3 mg (0.032 mmol, 4.0 equiv) of NBP was added. Aliquots were collected at regular time intervals and analyzed by reverse phase HPLC.

With Guanidine•HCl: To a 2.5 dram vial equipped with a stir bar were added 12.0 mg (0.008 mmol, 1.0 equiv) of vancomycin methyl ester (S4), 13.6 mg of guanidine•HCl (18.0 equiv) and MeOH (2 mL). The reaction mixture was allowed to stir for ca. 5 min, after which 7.3 mg (0.032 mmol, 4.0 equiv) of NBP was added. Aliquots were collected at regular time intervals and analyzed by reverse phase HPLC.

The HRMS and HPLC analysis of both reactions revealed that major product formed in both reactions has a mass of 1664.2957 (M+H⁺). The identity of this material is not assigned; however, the isotope pattern of mass spectrum is consistent with that of a dibromide containing a vancomycin methyl ester analogue with molecular formula of $C_{69}H_{81}Br_2Cl_2N_9O_{25}$. These experiments did not deliver ready access to monobromides analogous to 7_{f} -Br over 7_{d} -Br, as their corresponding methyl esters.

Other additives that might target carboxylic acid functionality, such as ureas, also provide some selectivity for 7_{f} -Br over 7_{d} -Br upon reaction with vancomycin (4). The raw data for these promoters are presented below.

Reaction Conditions: 0.008 mmol of 4, 4 equiv of NBP, 3 equiv of urea, 1 mL of MeOH

Table 4. Optimization of Reaction Conditions Aiming Analog 5 (Manuscript Table 2)

General Procedure for Table 4:

Entry 1: To a HPLC vial equipped with a stir bar were added 12.5 mg (0.008 mmol, 1.0 equiv) of vancomycin•HCl. Water (250 μ L) and MeOH (50 μ L) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 2.0 equiv) of NBP were added. Aliquots were collected at regular time intervals and analyzed by reverse phase HPLC.

Entry 2: To a HPLC vial equipped with a stir bar were added 12.5 mg (0.008 mmol, 1.0 equiv) of vancomycin•HCl. MeOH (1.2 mL) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 3.7 mg (0.016 mmol, 2.0 equiv) of NBP were added. Aliquots were collected at regular time intervals and analyzed by reverse phase HPLC.

Entry 3: To a HPLC vial equipped with a stir bar were added 25.0 mg (0.016 mmol, 1.0 equiv) of vancomycin•HCl. MeOH (4.0 mL) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 14.6 mg (0.064 mmol, 4.0 equiv) of NBP were added. Aliquots were collected at regular time intervals and analyzed by reverse phase HPLC.

Entry 4: To a HPLC vial equipped with a stir bar were added 25.0 mg (0.016 mmol, 1.0 equiv) of vancomycin•HCl and peptide **10** (2 equiv). MeOH (4.0 mL) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 14.6 mg (0.064 mmol, 4.0 equiv) of NBP were added. Aliquots were collected at regular time intervals and analyzed by reverse phase HPLC.

Entry 5: To a HPLC vial equipped with a stir bar were added 25.0 mg (0.016 mmol, 1.0 equiv) of vancomycin•HCl and guanidine•HCl (18 equiv). MeOH (4.0 mL) were added to the reaction flask. The reaction mixture was allowed to stir for ca. 5 min, after which 14.6 mg (0.064 mmol, 4.0 equiv) of NBP were added. Aliquots were collected at regular time intervals and analyzed by reverse phase HPLC.

Entry 6: The same procedure as entry 5 was employed, except 6 equiv of guanidine•HCl was used

Figure 8. Speculative concept for bromide delivery by guanidine.