

## SUPPLEMENTARY DATA

**Supplementary Table 1.** Forward (F) and reverse (R) primers used for PCR quantification of mRNA transcripts encoding Gapdh, both isoforms of the beta chain constant region, V $\beta$ 9, V $\beta$ 13, V $\beta$ 15, and V $\beta$ 16. Gapdh was used as housekeeping control. Additional details are given in Methods.

Genetic Element	Sequence
Real-time Primers	
Gapdh (F)	5'-GCACCACCAACTGCTTAGCC-3'
Gapdh (R)	5'-CTGAGTGGCAGTGATGGCAT-3'
C $\beta$ (F)	5'-TGCCAAGTGCAGTTCTATGG-3'
C $\beta$ (R)	5'-TTGTTGATAGGATGCTGAGGT-3'
V $\beta$ 4 (F)	5'-GTTGCCTGGTCCTCTGTCTC-3'
V $\beta$ 4 (R)	5'-TTGTGTCCTAGATGCTGTCG-3'
V $\beta$ 9 (F)	5'-TGTGATCTTCTGTCTTGCAG-3'
V $\beta$ 9 (R)	5'-CACTGGTTTCCAACCTTGC-3'
V $\beta$ 13 (F)	5'-GGGCAGTGCTCTGTCTCCT-3'
V $\beta$ 13 (R)	5'-TGGGTGACTCCAGTTTCAGA-3'
V $\beta$ 15 (F)	5'-TGCTGCTCTGCTACTTCTGG -3'
V $\beta$ 15 (R)	5'-CCTGAACACCCTCAGTGTGAC-3'
V $\beta$ 16 (F)	5'-GGCATCCAGACCCCTGTGTT-3'
V $\beta$ 16 (R)	5'-TGAUTCCAGCATCTGCGTTA-3'
V $\beta$ 13 for J $\beta$ (F)	5'-TTGCTGTGAGGACTGAAGGA-3'
V $\beta$ 16 for J $\beta$ (F)	5'-CGATGGTAAAATTGGGGAGA-3'
J $\beta$ 1.1 (R)	5'-AACTGTCAGTCTCGTTCTTACC-3'
J $\beta$ 1.2 (R)	5'-GGGCCGAAGGTGTAGTCATA-3'
J $\beta$ 1.3 (R)	5'-CTTCCTTCACCAAAATAGAGCAC-3'
J $\beta$ 1.4 (R)	5'-CATGGCCGAAAAACAATCTT-3'
J $\beta$ 1.5 (R)	5'-CCGAGTCCCTCTCCAAAAT-3'
J $\beta$ 1.6 (R)	5'-GTGCCTGCTGCAAAGTAGAG-3'
J $\beta$ 2.1 (R)	5'-GTGTCCCTGATCCGAAGAA-3'
J $\beta$ 2.2 (R)	5'-CAGCACTGTCAGCTTGAGC-3'
J $\beta$ 2.3 (R)	5'-AACGGTCAGTCTGGTTCTG-3'
J $\beta$ 2.4 (R)	5'-CTGCACCGAAGAACAAAGGT-3'
J $\beta$ 2.5 (R)	5'-TGCCTGGTCCAAAGTACTGG-3'
J $\beta$ 2.6 (R)	5'-CGGGACCGAAATACTGCT-3'
Cloning and Sequencing Primers	
V $\beta$ 13 Cloning 1 (F)	5'-GGGCAGTGCTCTGTCTCCT-3'
V $\beta$ 13 Cloning 1 (R)	5'-TTGTTGATAGGATGCTGAGGT-3'
V $\beta$ 13 Cloning 2 (F)	5'-TCGACACAGTTCTGTCTGAAAC-3'
V $\beta$ 13 Cloning 2 (R)	5'-CCTGTTGAGCCATCAGAAC-3'

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**Supplementary Figure 1.** Correlation plot of J $\beta$  usage from direct sequencing and qRT-PCR analysis. To exclude the possibility that skewing of individual CDR3 regions, as determined by direct sequencing of bacterial clones, was an artifact of PCR amplification of the insert or bacterial clone expansion we analyzed J $\beta$  isotype abundance by qRT-PCR as described in Methods. Percent frequency of J $\beta$  determined by sequencing was plotted against the percent frequency of J $\beta$  determined by qRT-PCR. We observe a significant positive correlation of J $\beta$  abundance by these two methods (Islets: R $^2$ =0.4425, p=0.018; spleen poly I:C: R $^2$ =0.5871, p=0.0037; spleen untreated: R $^2$ =0.3683, p=0.036). The significant positive correlation between J $\beta$  usage determined by direct sequencing of clones and J $\beta$  usage as determined by qRT-PCR indicates that our measurements of skewing and diversity from direct sequencing give an accurate measure of the true TCR repertoire diversity

