

Electronic supplementary material for:

Unraveling the Function of *Arabidopsis thaliana* OS9 in the Endoplasmic Reticulum-Associated Degradation of Glycoproteins

Plant Molecular Biology

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Table S1. Oligonucleotide sequences as used in this study.

Fig. S1. LC-ESI-MS of OS9-GFPglyc purified from *N. benthamiana* leaves.

Fig. S2. Matrix-assisted laser desorption ionization time-of-flight mass spectrometry of N-glycans extracted from *A. thaliana* leaves.

Fig. S3. *os9-1* partially suppresses the *bri1-5* phenotype.

Fig. S4. Expression of OS9-GFP and OS9-GFPglyc in *os9-1 bri1-5* plants restores the *bri1-5* phenotype.

Fig. S5. *os9-1* partially suppresses the *bri1-9* phenotype.

Fig. S6. *os9-1* does not suppress the *bri1-6* phenotype.

Fig. S7. BRI1-5-GFP is retained in the ER.

Fig. S8. Expression of OS9R201A-GFPglyc does not restore the *bri1-5* phenotype in *os9-1 bri1-5* plants.

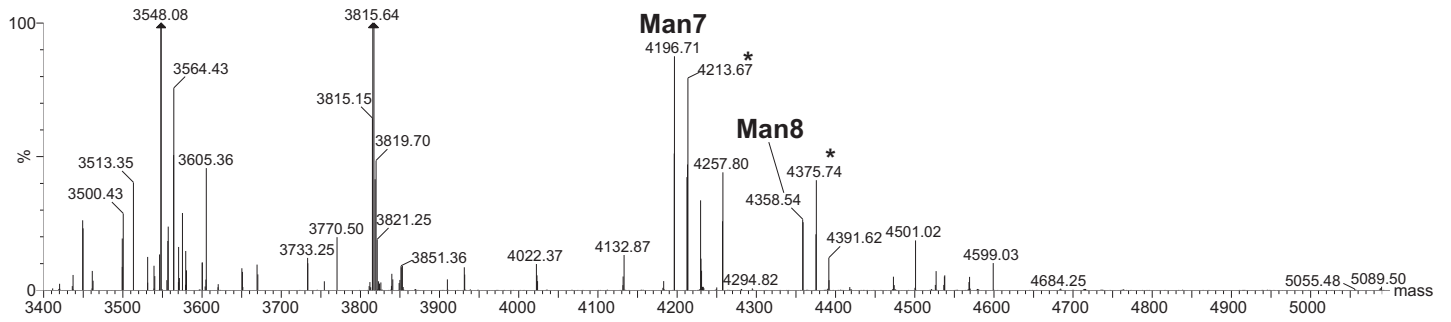
Fig. S9. *os9-1* seedlings are sensitive to elf18.

Fig. S10. Effect of tunicamycin and paraquat on *os9-1* seedlings.

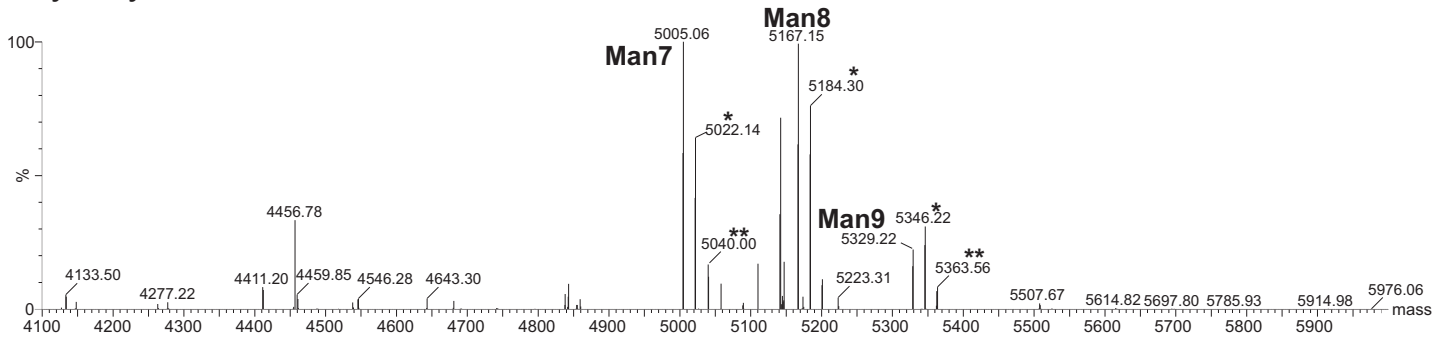
Table S1. Oligonucleotide sequences as used in this study

Name	Sequence (5' – 3')
LBa1	TGGTTCACGTAGTGGGCCATCG
At5g35080_1F	TCTAATGGTATGGCCCTTGTATCCTG
At5g35080_2R	GTTCTTCCTTGTTCCTTGTTCGGTCTT
At5g35080_3F	AAAAAGCGACGAGTGGATGGAC
At5g35080_4F	TTCTAGAATGAGAATCACGCAGATCTTGTT
At5g35080_5R	TGGATCCAGAATCAGCTATCATCTTAGGTG
At5g35080_6F	CCGTCCGGTGCTATTCTCCTTC
At5g35080_7R	AGCGGGTAACTGAAAATGGAGATA
At5g35080_21R	tattCTCGAGTCAGAGTTCGTCGTGAGAATCAGCTATCATCTTAGGTG
At5g35080_22F	tataGGATCCATGAGAATCACGCAGATCTTGTT
At5g35080-rev-Sall	TATAGTCGACTGCTTGTTCCTTGTTCCTTCT
At5g35080_23F	ATCTTACAGGATCACCTGCCGAAGTCGAGGTGAGGT
At5g35080_23R	ACCTCACCTCGACTTCGGCAGGTGATCCTGTAAGAT
At1g18260_1F	AGTAGCAGAAAGGGGGCCGTGGAGT
At1g18260_2R	GAGAATCAGGCGACAAGTATCAAGTTT
At1g18260_10F	TACTAGTATGAGAATATTAAGCTACGGAATCG
At1g18260_11R	TAGATCTCCGTGGGAACGCAGCGAGGTGTT
BRI1_1F	tGGATCCATGAAGACTTTTTCAAGCTTCTTTCTCT
BRI1_2R	tGGATCCTAATTTTCCTTCAGGAACTTCTTTTATAC
BRI1_7F	GTTTCTGGACATGTTTTACAACATGTTGT
BRI1_7R	ACAACATGTTGTAAAACATGTCCAGAAAC
BRI1_10F	TACGTTGTTGAAGATGAGAGGACT
BRI1_13F	CAAAAACGATGGGATGAAGAAAGAGT
BRI1_14R	ACACCGCGGAAGAGGATAACCACAG
BRI1_15F	TTCTTCTTCTCCTTCTTTTCTCTTTCATT
BRI1_16R	ACGACGTTAGCACCGGAGATTGAA
BRI1_17F	tataACTAGTATGAAGACTTTTTCAAGCTTCTTTCTC
BRI1_18R	tataGGATCCGATTTTGTTCGCTAATCGCTAA
Sc_CPY_1F	CCATCGGGAACCCTTACTCTTG
Sc_CPY_5R	CACAGGCCATTGGTTCGTAA
Sc_Yos9_1F	ATTCTATATTTGGTTGGGTTTTTAAG
Sc_Yos9_2R	ATGAACAAATTCTATGCCAGGACAGTA
Sc_Yos9_7F	AGGATCCATGCAAGCTAAAATTATATATGCTCTG
Sc_Yos9_8R	ACTCGAGTTAAAGCTCATCATGTTTCGATGAA
Sc_Yos9-fw-Sall	TAATGCCGTCGACTATGAACCGATTTTTTTAG
KanB	CTGCAGCGAGGAGCCGTAAT
KanC	TGATTTTGATGACGAGCGTAAT
KanD	CCCCGGCAAACAGCATTCC
KanE	TCACCGAGGCAGTTCATAGG

Glycosylation site 1, ATSGWTSSQQNISTVMMETQQLVK



Glycosylation site 2, IVQEFFLGTFDPEATAAFNQTVS DASTDASQR



Glycosylation site 3, YHSHVYTNGTTCDLTGSPR

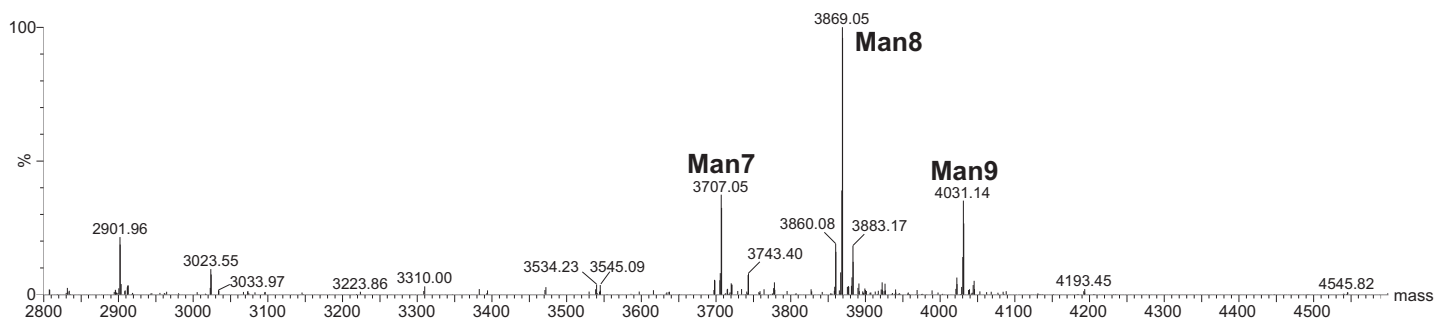


Fig. S1. LC-ESI-MS of OS9-GFPglyc purified from *N. benthamiana* leaves. Mass spectra of the three glycopeptides derived from *A. thaliana* OS9 are shown. Man7, Man8 and Man9 are oligomannosidic peaks corresponding to Man7GlcNAc₂, Man8GlcNAc₂ and Man9GlcNAc₂. The asterisks indicate ammonia adducts of the same peaks. The whole procedure for purification and detection of glycopeptides has been described previously (Schoberer et al., 2009).

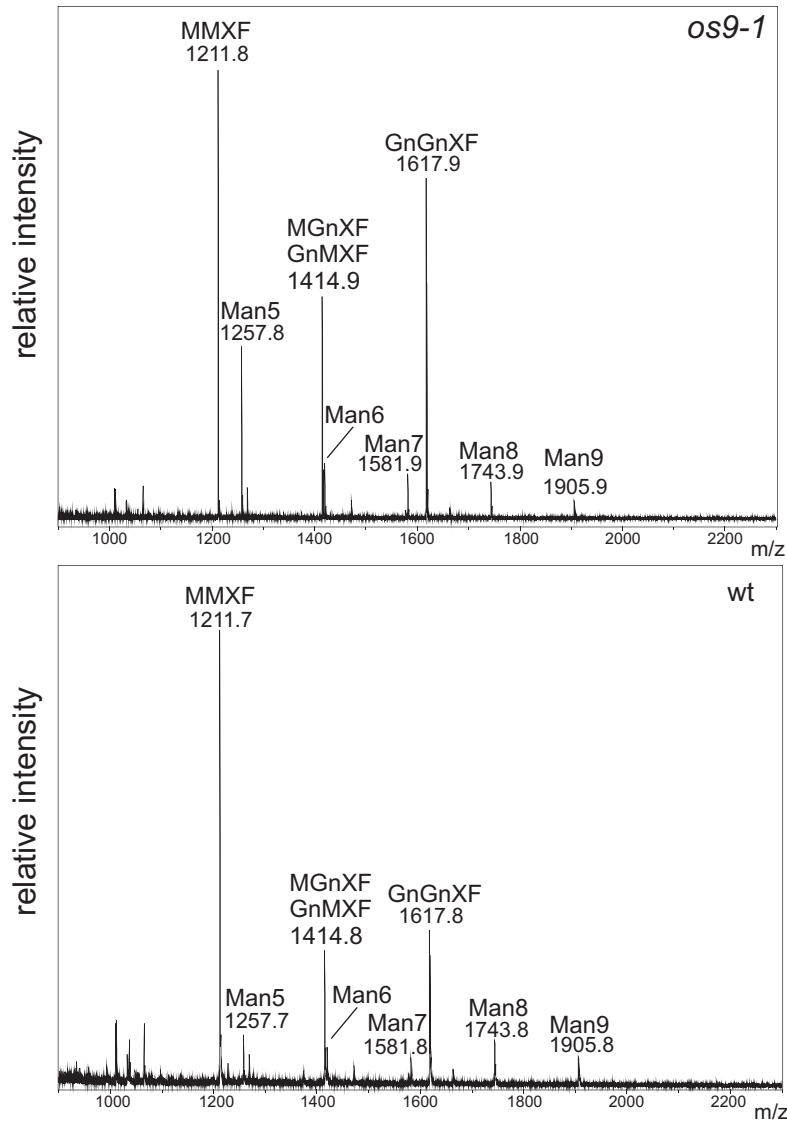


Fig. S2. Matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF-MS) of total N-glycans extracted from leaves of *A. thaliana* plants. Spectra of *os9-1* and wild-type plants are shown. The following oligosaccharide structures are indicated: MMXF: Man3XylFucGlcNAc₂; Man5: Man5GlcNAc₂; MGnXF/GnMXF: GlcNAcMan3XylFucGlcNAc₂; Man6: Man6GlcNAc₂; Man7: Man7GlcNAc₂; GnGnXF: GlcNAc₂Man3XylFucGlcNAc₂; Man8: Man8GlcNAc₂; Man9: Man9GlcNAc₂.

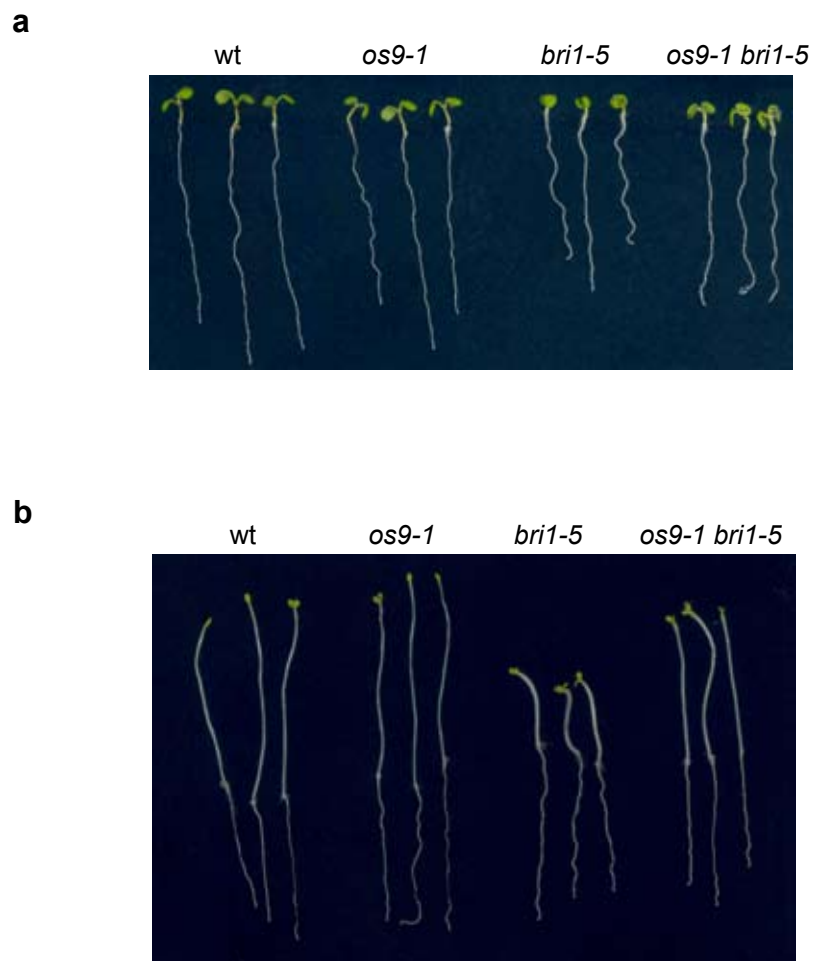


Fig. S3. *os9-1* suppresses the *bri1-5* phenotype. Growth of different plants on 1x MS medium with 1.5% sucrose for 8 days at 22°C **(a)** under long-day conditions (16h-light/8h-dark) or **(b)** in the dark.

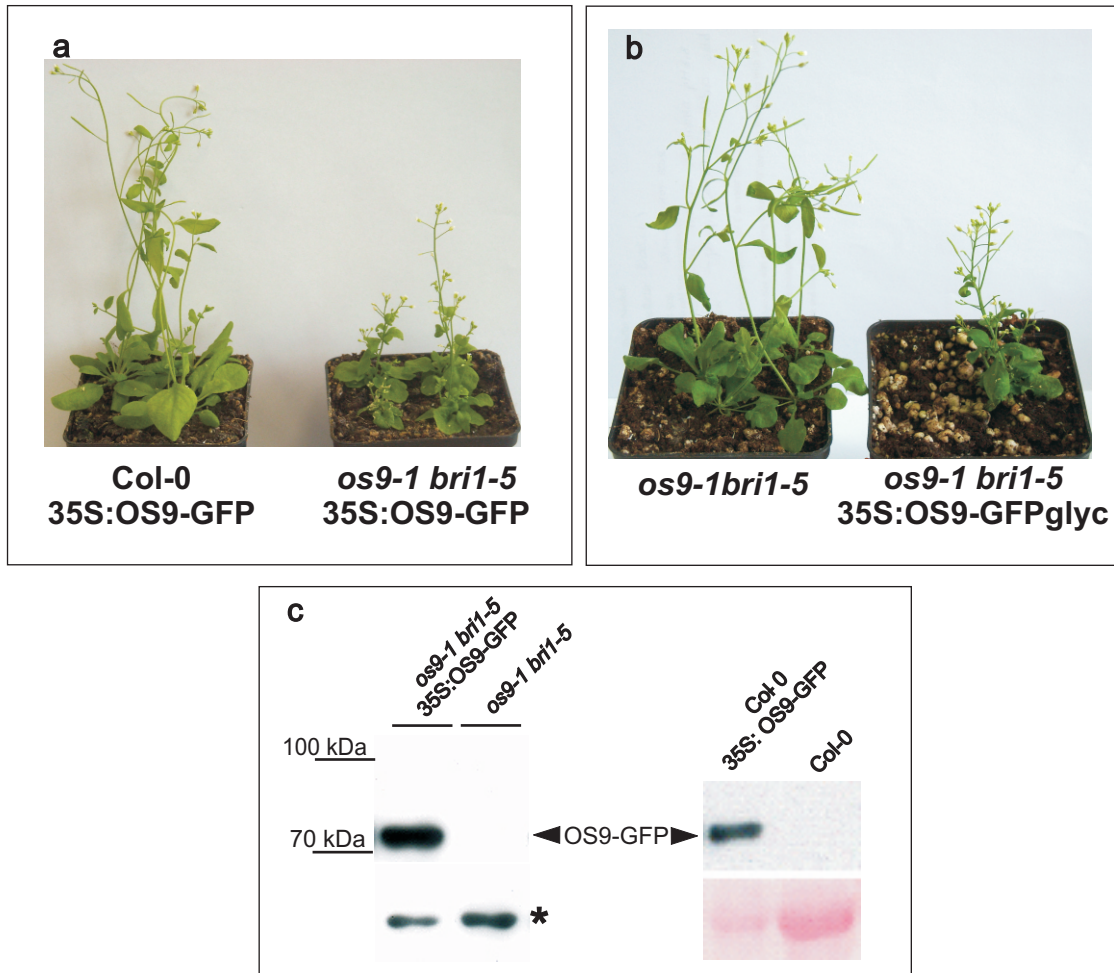


Fig. S4. Expression of OS9-GFP or OS9-GFPglyc in *os9-1 bri1-5* plants restores the *bri1-5* growth phenotype. *os9-1 bri1-5* double mutants were floral dipped with **(a)** 35S:OS9-GFP or **(b)** 35S:OS9-GFPglyc. **(c)** Positive transformants were analyzed by immunoblotting with anti-OS9 antibodies for the presence of OS9-GFP expression or OS9-GFPglyc. The asterisk on the immunoblot denotes an unspecific band that served as a loading control.

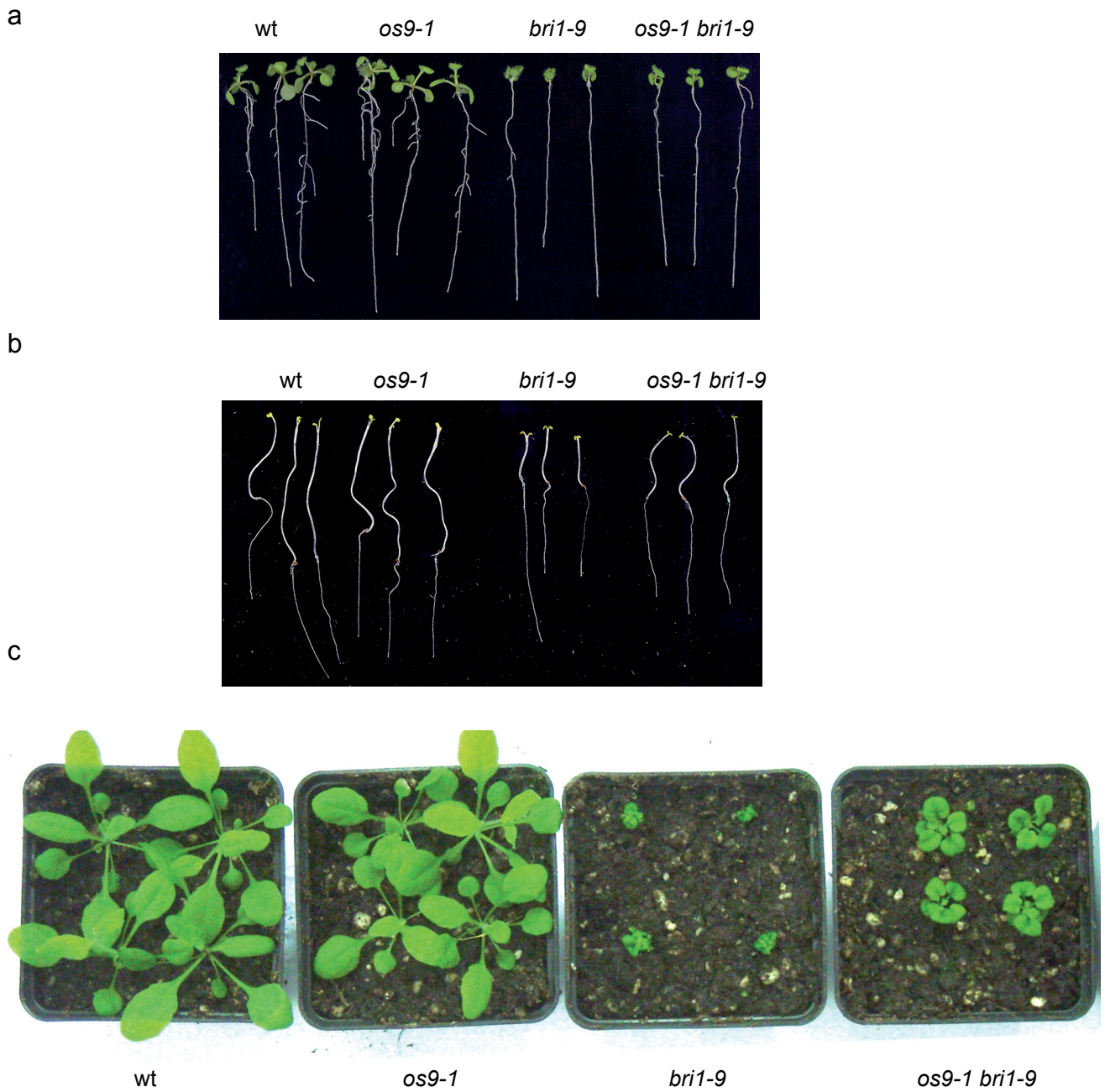


Fig. S5. *os9-1* partially suppresses the *bri1-9* phenotype. Growth of different plants on 1x MS medium with 1.5% sucrose for 2 weeks at 22°C **(a)** under long-day conditions (16h-light/8h-dark) or **(b)** in the dark. **(c)** Growth phenotype of 24 day-old soil-grown plants.

a



b

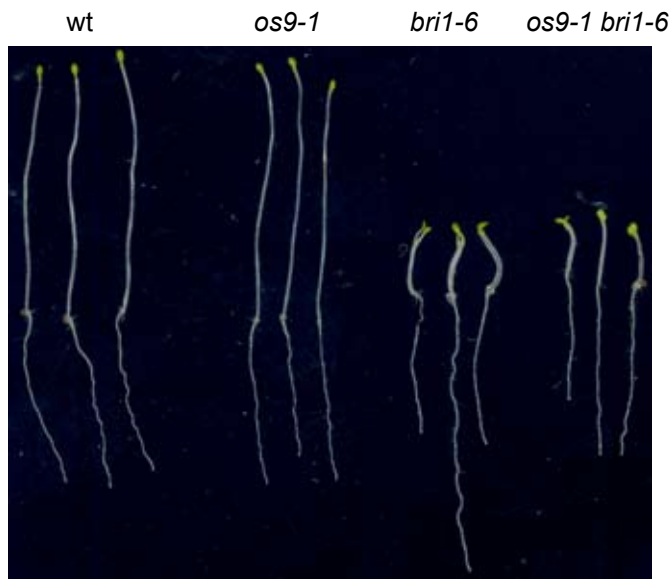


Fig. S6. *os9-1* does not suppress the *bri1-6* phenotype. Growth of different plants on 1x MS medium with 1.5% sucrose for 8 days at 22°C **(a)** under long-day conditions (16h-light/8h-dark) or **(b)** in the dark.

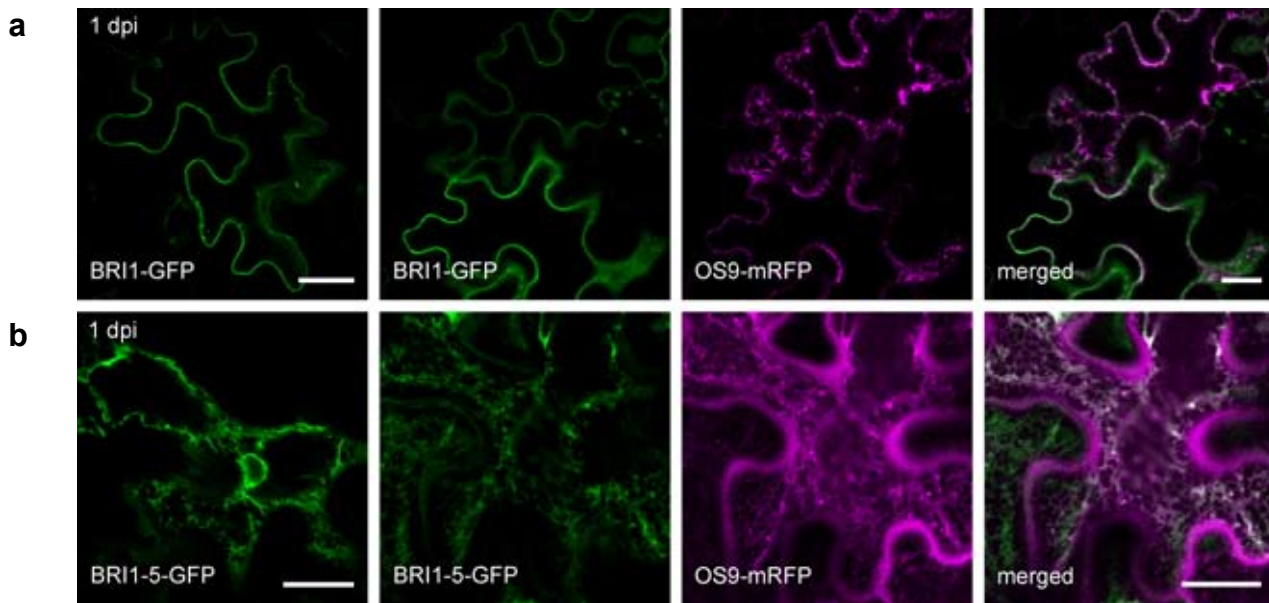


Fig. S7. BRI1-5-GFP is retained in the ER. Co-expression of different BRI1-forms in *N. benthamiana* leaf epidermal cells. Confocal images were taken 1 day after infiltration ($OD_{600} = 0.15$). Scale bars = 20 μ m. **(a)** Co-expression of BRI1-GFP (in green) and OS9-mRFP (in magenta). The first image shows BRI1-GFP alone, followed by the images showing co-expression. **(b)** Co-expression of BRI1-5-GFP (in green) and OS9-mRFP (in magenta). The first image shows BRI1-5-GFP alone, followed by the images showing co-expression. **(c)** Immunoblot analysis suggests that BRI1-5 carries only oligomannosidic N-glycans indicative of ER-localization of BRI1-5 and OS9. Transient expression was done in *N. benthamiana* Δ XF plants, which contain low amounts of core α 1,3-fucose. Complex N-glycans from this plant are therefore sensitive to PNGase F digestion.



Fig. S8. Expression of OS9R201A-GFPglyc does not restore the *bri1-5* growth phenotype in *os9-1 bri1-5* plants. *os9-1 bri1-5* double mutants, *os9-1* mutants and wild-type (wt) were floral dipped with 35S:OS9R201A-GFPglyc. Positive transformants were grown on soil (**a** and **b**) and OS9R201A-GFPglyc expression was confirmed by (**c**) immunoblotting with anti-OS9 antibodies for the presence of OS9-GFPglyc expression.

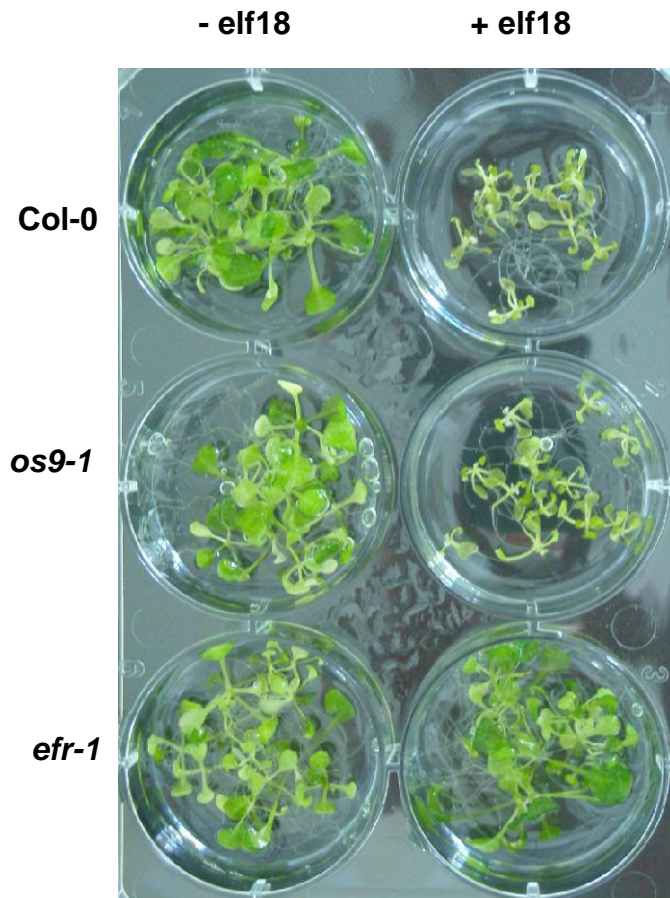


Fig. S9. *os9-1* seedlings are sensitive to elf18. A seedling growth inhibition assay in the absence (- elf18) or presence (+ elf18) of elf18 peptide is shown. The elf18 peptide blocks seedling growth of *os9-1* similar to wild-type seedlings (Col-0), while the mutant *efr-1* (Zipfel et al., 2006) that served as a control is insensitive to elf18. Seven-day-old seedlings were incubated for one week in 1x MS liquid medium containing 1% sucrose in the presence of 50 nM elf18 peptide.

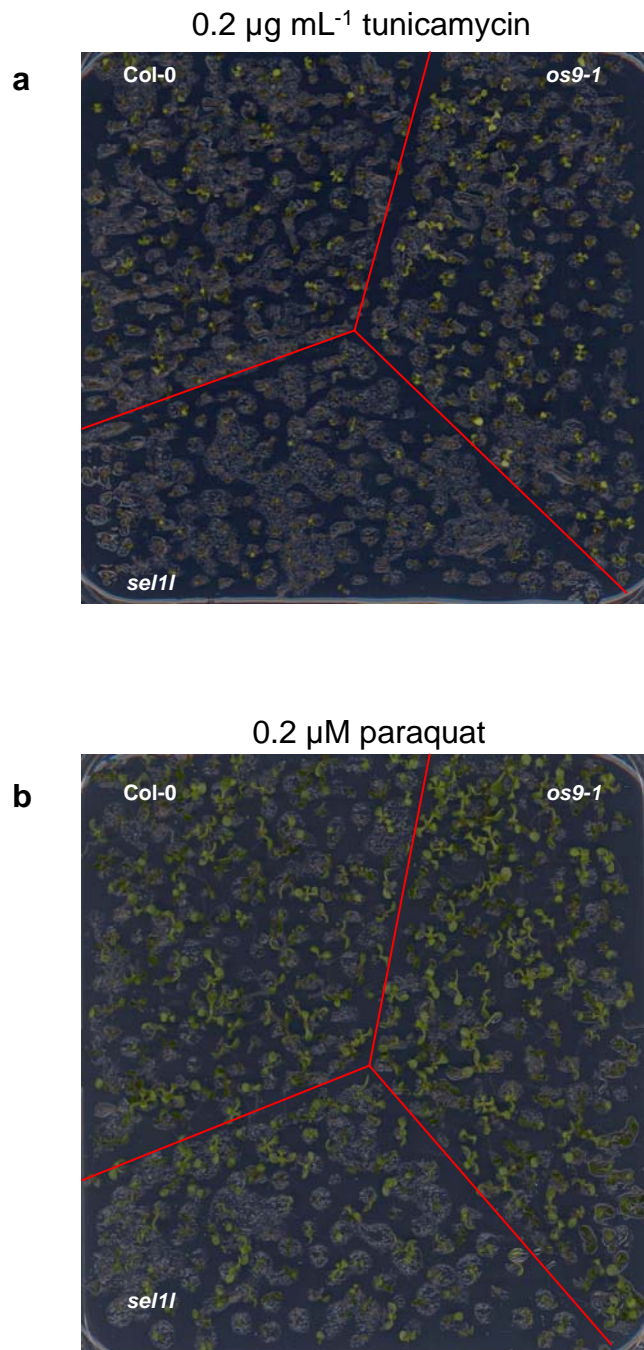


Fig. S10. Effect of tunicamycin and paraquat on *os9-1* seedlings. Seeds were germinated on $\frac{1}{2}\times$ MS medium containing 1.5% sucrose **(a)** in the presence of $0.2 \mu\text{g mL}^{-1}$ tunicamycin or **(b)** in the presence of $0.2 \mu\text{M}$ paraquat. *sel1l* was used as a control (Liu et al., 2011). Seedlings were photographed after 11 days of treatment.