## Supporting Information

## García-Fandiño and Sansom 10.1073/pnas.1119326109

## SI Text

SI Methods Parametrization was based on that used in previous stu-SI Text<br>SI Text<br>SI Methods Parametrization was based on that used in previous stu-<br>dies (1): Carbon atoms were modeled as uncharged Lennard– Jones particles with  $\sigma$ cc = 3.400 Å and  $\epsilon$ cc = 0.086 kcal mol<sup>-1</sup>, corresponding to sp2 carbons in the AMBER99 force field. Let the carbon atoms were modeled as uncharged Lennard-<br>Jones particles with  $\sigma$ cc = 3.400 Å and  $\epsilon$ cc = 0.086 kcal mol<sup>-1</sup>,<br>corresponding to sp2 carbons in the AMBER99 force field.<br>Carbon–carbon bond lengths of 1.4 Å a were maintained by harmonic potentials with spring constants of 938 kcal mol<sup>-1</sup> Å<sup>-2</sup> and 126 kcal mol<sup>-1</sup> rad<sup>-2</sup>, respectively. In addition, a weak dihedral angle potential was applied to bonded were maintained by harmonic potentials with spring constants of 938 kcal mol<sup>-1</sup>  $\text{Å}^{-2}$  and 126 kcal mol<sup>-1</sup> rad<sup>-2</sup>, respectively. In addition, a weak dihedral angle potential was applied to bonded carbon atoms. The  $\sigma$ co = 3.2751 Å and  $\epsilon$ co = 0.114333 kcal mol<sup>-1</sup>. The parameters for the functional groups COO<sup>-</sup> and  $NH_3$ <sup>+</sup> were obtained from the corresponding residues in the Amber force field, Glu and Lys, respectively. The partial charges were modified to C(0.01360)-C  $(0.\dot{8}0540)$ -O(-0.9095) in the carboxylate group and C(-0.01430)-N(-0.3867)-H(0.457) in the amine group.

We used the water (SPC/E)/ion combination parameters published by Joung and Cheatham (2) implemented in AM-BER10 to prevent crystallization previously reported at high concentrations with other parameter sets (3). The GAFF force field was used for dioleoyl phosphatidyl choline lipids, employed also in previous studies (4) from which we obtained a preequilibrated membrane for subsequent replication. The bilayer was replicated three times in  $x$  and  $y$  directions [or only twice in the case of the potential of mean force (PMF) calculations], and after carbon nanotube (CNT) insertion, the complete system was solvated. Water in the hydrophobic region of the tails and also inside of the CNT was removed, so that in the first step of the simulation the channel was completely dry. The resultant system was ionized using different salt concentrations (NaCl 1 M, KCl 1 M, and  $CaCl<sub>2</sub> 0.5 M$ ). The initial size of the unit cell was equal to  $18.2 \times 17.4 \times 7.0$  nm<sup>3</sup> and contained 560 lipids and approximately 18,000 water molecules.

The equilibrium simulations were performed with the GRO-MACS 4.0 (5) molecular dynamics program. All the systems were

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partially optimized, thermalized, and equilibrated, followed by unrestrained simulations for at least 30 ns (time step of 2 fs) for each one of the systems studied. The constant pressure and temperature canonical ensemble was employed with the pressure of 1 bar controlled using a semii each one of the systems studied. The constant pressure and temperature canonical ensemble was employed with the pressure of stat (6), and the temperature of 300 K imposed by a Berendsen (7) thermostat. The LINCS (8) algorithm was employed to remove the bond vibrations. The Particle Mesh Ewald method (9) coupled to periodic boundary conditions was used to treat the long-range electrostatics using a direct-space cutoff of 1.0 nm and a grid spacing of 0.12 nm. Van der Waals interactions were<br>computed using periodic boundary conditions coupled to a sphe-<br>rical cutoff of 1.0 nm.<br>Poisson–Boltzmann calculations were carried out using the computed using periodic boundary conditions coupled to a spherical cutoff of 1.0 nm.<br>Poisson–Boltzmann calculations were carried out using the

software package APBS (10). These calculations were performed as described in ref. 11. The program HOLE (12) yielded the radius profile of the CNT system and also provided sample points along the pore axis at which to place the ion. The test ions were assigned a Born radius of  $0.1680$  nm (for Na<sup>+</sup>),  $0.2172$  nm (for K<sup>+</sup>), 0.1862 nm (for Ca<sup>2+</sup>), and 0.1937 nm (for Cl<sup>−</sup>) (13). The electrostatic binding energy of the ion was calculated at subsequent positions z as

$$
\Delta G_{\rm B}(z) = G_{\rm pore+ion}(z) - G_{\rm ion} - G_{\rm pore}.
$$
 [S1]

The temperature was 300 K and the dielectric constant  $\epsilon p$  of the pore was set to four, the ionic strength 0.15 M, and the dielectric constant for the solvent was 78.5. For each data point, the ion was placed at a sample point on the pore center line as identified by HOLE. The sample points were 0.25 Å apart along the z axis. The channel was contained in a coarse grid of dimensions 15.1 nm  $\times$  15.2 nm  $\times$  6.7 nm<sup>3</sup> and 289 points along the z axis were used.

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Fig. S1. Numbers of cations and chloride ions inside the nanopores during equilibrium molecular dynamics simulations.





Fig. S2. Radial distributions of cations (black), Cl- ions (red), and water (green).

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Fig. S3. Distributions of cations (red) and Cl<sup>-</sup> ions (black) along the long axis (z) of the nanopores. The selectivity filters are positioned at z ca. 0 nm.

AC<br>A



**Fig. S4.** Poisson–Boltzmann profiles for several ions for the pristine CNT (A) and those derivatized with 2 COO- C-2 (*B*), 4 COO- C-4 (C) 2 COO- plus 2 NH $_3^+$  C0 (*D*), and 2COO- plus  $NH_3^+$ /CH<sub>3</sub> C-1 groups at the central position of the pore.

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Fig. S5. PMFs for single ions as a function of position along the z axis of the pore of C-1. The bilayer extends from z ca. -1.5 to +1.5 nm. The selectivity filters are positioned at z ca. 0 nm.

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Fig. S6. Coordination of ions inside the CNT in the PMF calculations.



Fig. S7. Radial distributions of Na<sup>+</sup> ions (black), Cl<sup>-</sup> ions (red), and water (green) for ions and water inside the nanotube in the vicinity of the selectivity filter, i.e.,  $|z|$  < 0.5 nm.





Standard deviations are given in parentheses.

AC<br>A





The diffusion coefficients (D in units of 10<sup>−</sup><sup>9</sup> m<sup>2</sup>∕s) are given parallel to the nanopore axis (with standard deviations in parentheses). For comparison, values for diffusion in bulk solution are Na† 1.2, Cl− 1.8, K† 1.8, and Ca<sup>2+</sup> 0.53  $\times$  10<sup>-9</sup> m<sup>2</sup>/s (1).

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