

1 (Supplemental Material)

2 Supplemental Material Figure legends

3

4 Fig. S1. The relative fluorescence of the linker region-deletion mutants. The red  
5 fluorescence of displayed mRFP1 was measured as described in the method. The deletion  
6 mutant (missing 24 ~ 948 region) showed only 33 % of the fluorescence of the other  
7 mutants having predicted linker regions. This result showed that the 785 ~ 948 region  
8 plays an important role for efficient display of heterologous proteins.

9

10 Fig. S2. The overview of construction of pATLIC vector (A). The passenger domain was  
11 deleted by PCR using the primers containing TEV protease recognition and SmaI  
12 restriction site sequences. Amplified PCR products were treated by T4 DNA polymerase  
13 without dNTPs for annealing of single-stranded ends. A typical ligation independent  
14 cloning (LIC) procedure for display of proteins of interest (POI) using pATLIC vector  
15 was shown in (B).

16

17 Fig. S3. The protease accessibility test for surface-displayed or periplasmic mRFP1. For  
18 periplasmic mRFP1, we expressed mRFP1 in pATLIC having an identical signal sequence  
19 but lacking the translocator domain by incorporating a stop codon followed by the  
20 mRFP1 sequence (pATLIC\*-mRFP1). We observed the change of relative fluorescence of  
21 whole-cells after trypsin digestion as described in the method. The periplasmic mRFP1  
22 was not susceptible to trypsin treatment but surface-displayed mRFP1 was degraded by  
23 trypsin. The fluorescence of untreated (□) and treated (■) cells are presented in the figure.

24

25 Fig. S4. (A) Optimal inducer concentration for the maximum surface display and its  
26 effect on the cell growth. Effects of inducer (L-(+)-arabinose) concentration on surface  
27 display of Aga16B. The amount of reducing sugars produced at the range of 0 ~ 0.2% of  
28 inducer. (B) Growth of host cells (*E. coli* BW25113) displaying various proteins was  
29 measured by OD<sub>600</sub>. *E. coli* BW25113 harboring pATLIC-*aga50A* (●), pATLIC-*aga16B*  
30 (◆), pATLIC-*aga86C* (▲), pATLIC-*aga50D* (■) and pATLIC-*aga86E* (X) were grown  
31 for 24 h at 16°C. An asterisk (\*) indicates the inducing time point with L-(+)-arabinose.

32

33

34 Fig. S5. The neoagarobiose hydrolase (NABH) activity in the culture supernatant of  
35 NABH displayed cells. Before NABH displayed cells were treated by TEV protease, the  
36 culture supernatant of the cells was concentrated, incubated in the reaction mixture and  
37 analyzed by TLC. Lane 1, L-(+)-arabinose (L-Ara); lane 2, D-galactose (D-Gal); lane 3,  
38 3,6-anhydro-L-galactose (L-AHG); lane 4, neoagarobiose (DP2); lane 5, the culture  
39 supernatant

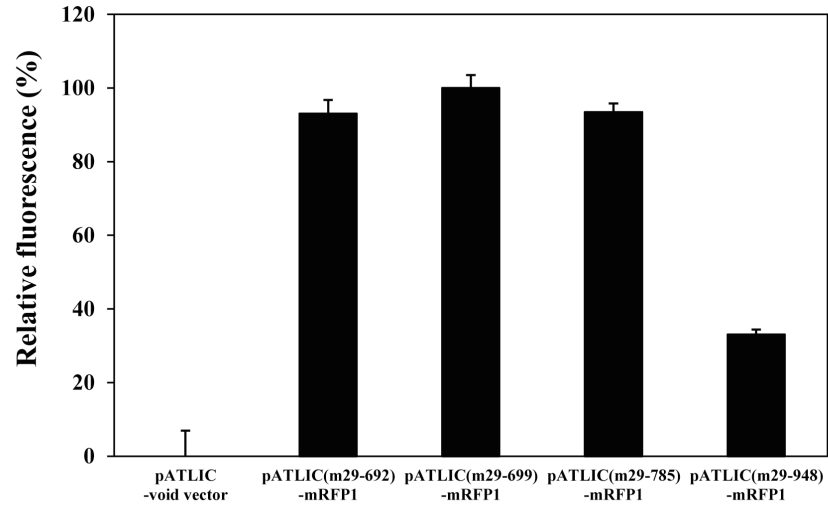
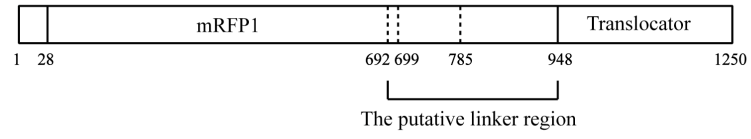
40

41 Fig. S6. The red fluorescence unit (RFU) curve of mRFP1. The correlation between the  
42 red fluorescence unit and amount of mRFP1 was calculated from the curve.

43

44

pATLIC<sub>(deletion mutants)</sub>-mRFP1 system

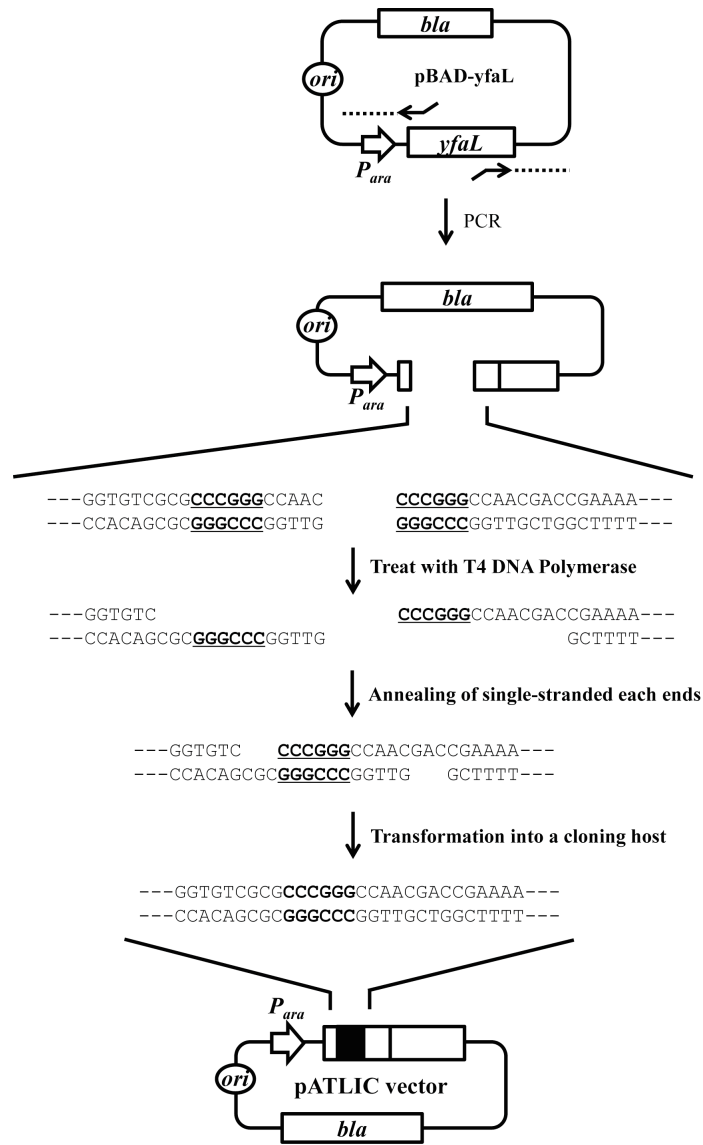


45

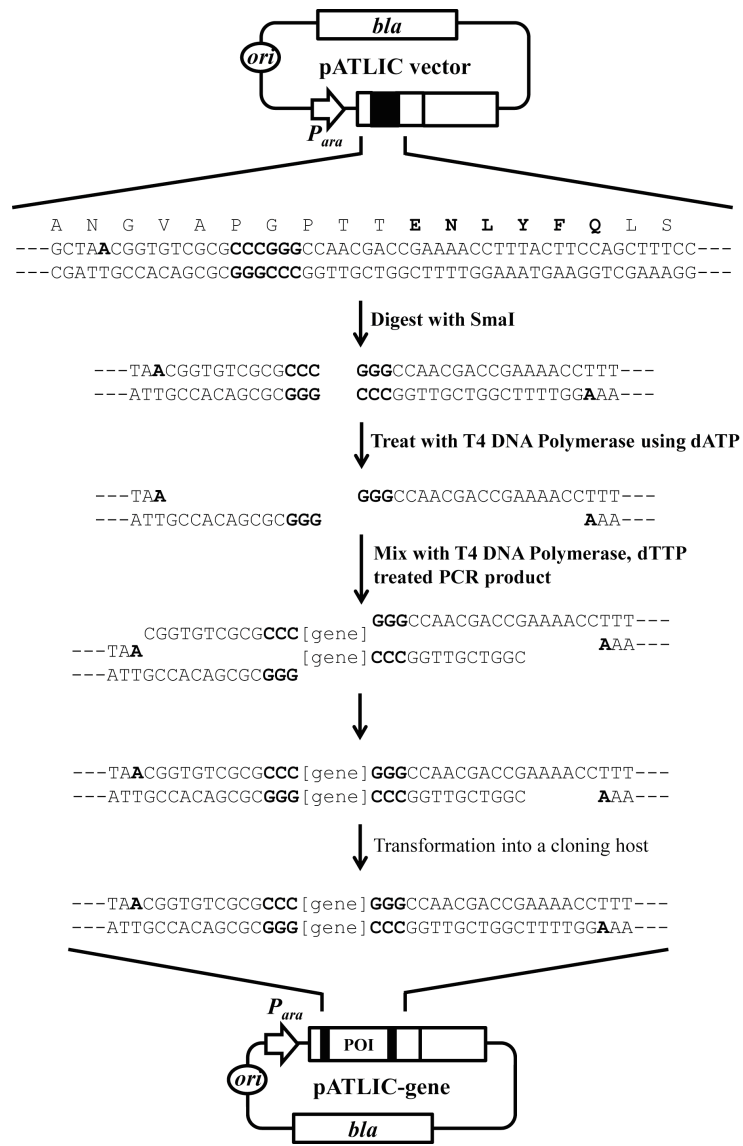
46

47

(Fig. S1)



(Fig. S2A)

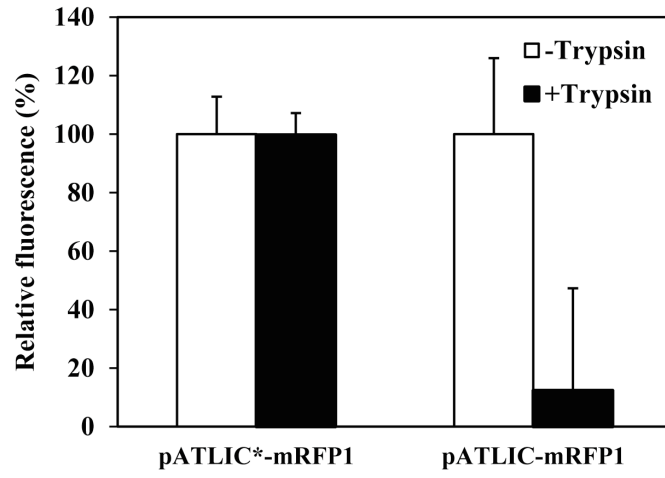


51

52

53

(Fig. S2B)

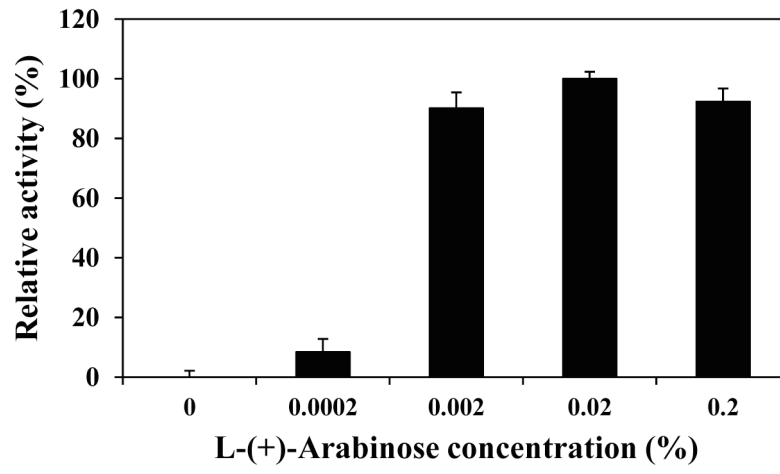


54

55

56

(Fig. S3)

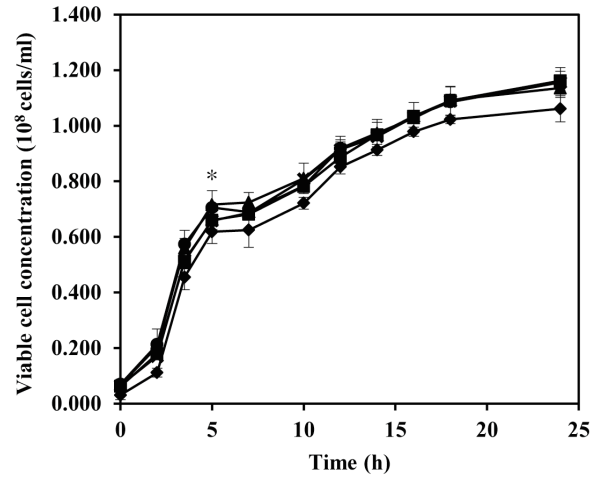


57

58

59

(Fig. S4A)



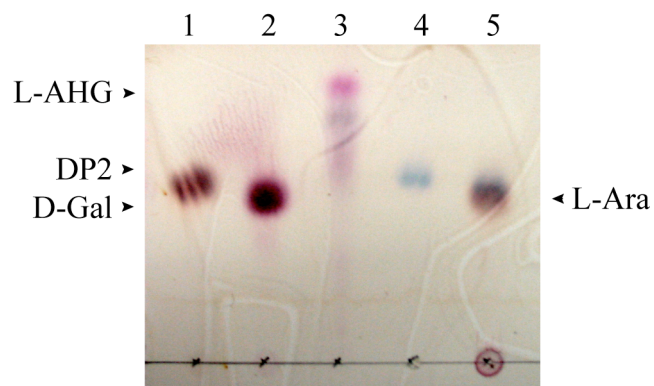
60

61

62

(Fig. S4B)



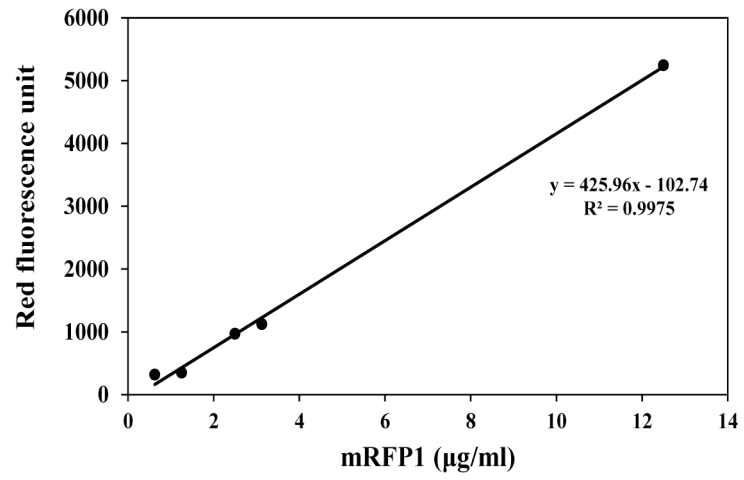


63

64

65

(Fig. S5)



66

67

(Fig. S6)