

SUPPLEMENTAL TABLE LEGENDS

Table S1. Nucleosome calls. Start and stop positions of all called nucleosomes.

Table S2. Nucleosome metaset. Start and stop sites, dynamic class, and \log_2 normalized sequencing counts per nucleosome are listed for nucleosomes in the metaset.

Table S3. Nucleosome positions and changes for loading into an integrated genome browser.

Table S4. Msn2p-bound loci. Start and stop positions, dynamic class, and \log_2 average array signal over the identified peak.

Table S5. Gene expression data. \log_2 fold-change in expression is shown for coding and non-coding transcripts, whose start and stop positions and significance of change are also shown

Table S6. Venters comparison. The Bonferroni-corrected p-values are shown for the overlap in genes bound by the denoted factor at either 25C or 37C, and containing an upstream nucleosome from the denoted class. See text for details.

SUPPLEMENTAL FIGURE LEGENDS

Figure S1. Biological reproducibility of dynamic classes of nucleosome repositioning. Of the 9,509 nucleosomes scored as changing in occupancy in the MNase-seq experiment, 4,943 were measured with at least two observations of 1.5X change in an independent time course hybridized to tiled yeast genomic arrays from Nimblegen (unpublished data). Nucleosomes were organized according to the classification of nucleosomes shown in Figure 3, and the mean of each cluster was calculated for the MNase-seq experiment (blue) versus the biological replicate hybridized to tiled arrays (red). The trends in the data were highly reproducible in the biological replicates, although the MNase-seq experiment had a significantly wider dynamic range, as expected for sequencing data. Biological replicates of the 30 min time point that were

hybridized to microarrays were highlight reproducible ($R \sim 0.7$ for \log_2 changes in nucleosome occupancy). Adjacent timepoints in the MNase-seq experiment were also well correlated ($R \sim 0.72 - 0.81$).

Figure S2. Associations between dynamic classes of nucleosome loss and chromatin regulators. Genes harboring upstream nucleosomes repositioned with different dynamic patterns were compared to genes whose upstream regions were bound by chromatin regulators at 25C (left) or after a 25C-37C heat shock (right) by Venters *et al.* (67). Blue boxes in the diagram represent significant (Bonferroni-corrected) overlap between genes bound by different regulators (y-axis) and genes harboring upstream nucleosomes in different dynamic classes (x-axis). Enrichments that were significant both before and after heat shock (left) and those for which at least nucleosome class changed significantly in enrichment upon stress (right) are shown. Annotations list the bound protein and the complex(es) it participates with. ATP-dependent chromatin remodelers are highlighted in red, SAGA components in pink, TFII subunits in blue, and transcription factors in green. P-values for all comparisons (including nucleosome loss and gain classes) are available in Table S6.

Figure S1

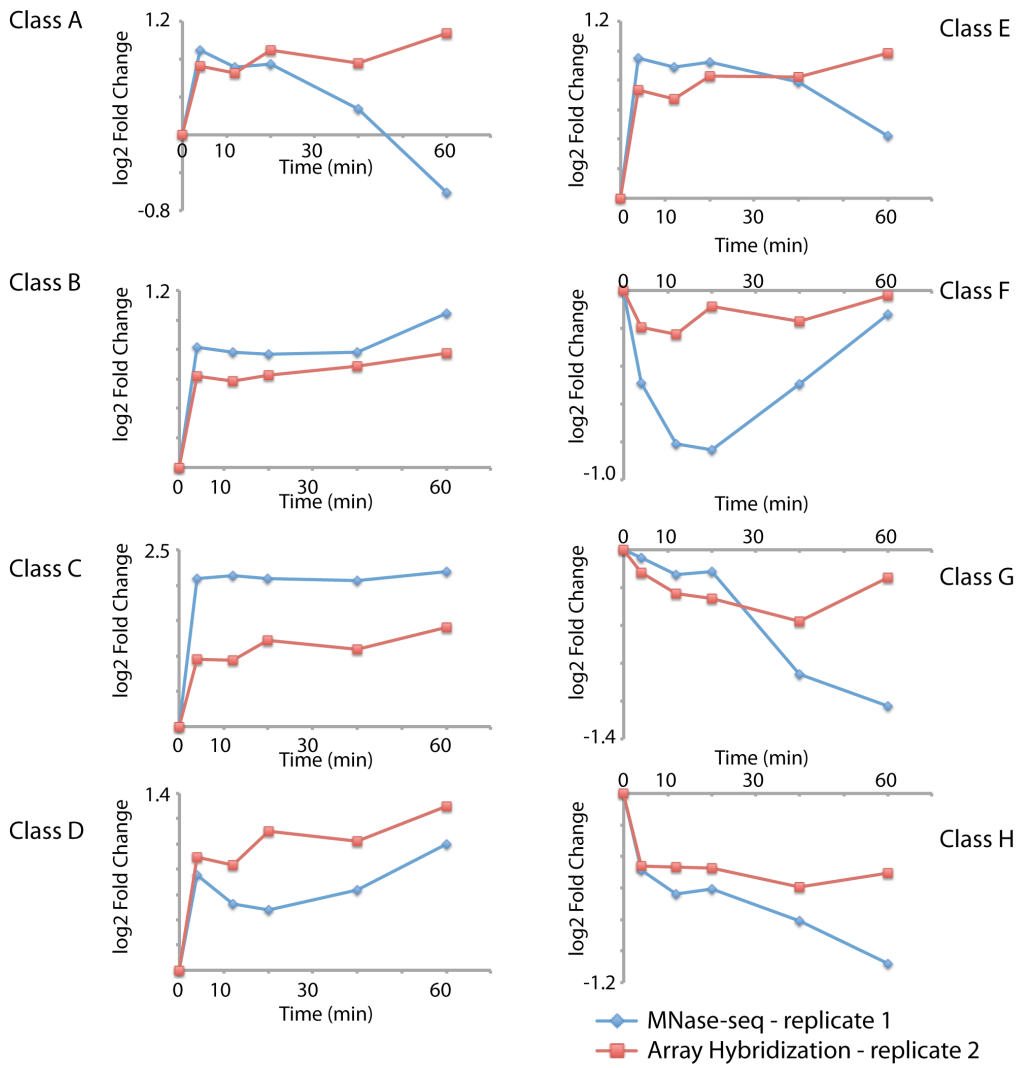
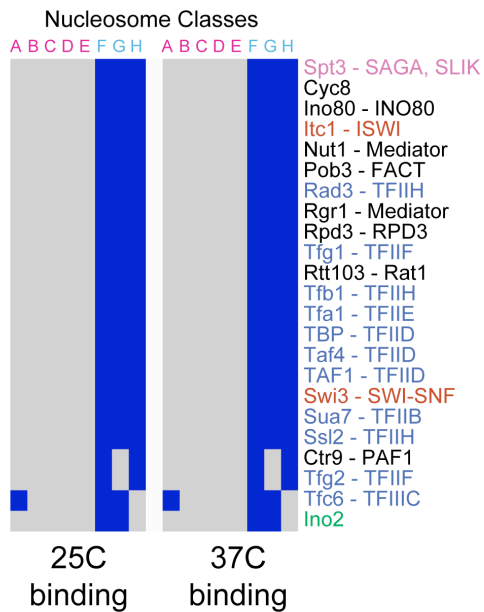


Figure S2

A



B

