

SUPPLEMENTARY FIGURES

Fig.S1. Skp2 Regulates ER α Levels and Stability in Breast Cancer Cells

A) Quantitative analysis of Skp2 protein in ER α -negative and positive cell-lines. B) Quantitative analysis of ER α protein in MCF-7 cells subjected to siRNA mediated Skp2 depletion. C) Quantitative analysis of ER protein in MCF-7 cells subjected to adenovirus mediated Skp2 over-expression. D) Quantitative analysis of ER α protein in MCF-7 cells subjected to adenovirus mediated Skp2 over-expression followed by Cycloheximide treatment for indicated times.

Fig.S2. Cul-7 and Rbx-1 are Essential for Skp2 Mediated Proteasomal Degradation of ER α in Breast Cancer Cells

A) Western analysis of ER α protein in MCF-7 cells subjected to siRNA mediated knockdown of indicated Cullin proteins and Rbx-1, in the absence or presence of ligand. B) Quantitative analysis of ER α protein in MCF-7 cells subjected to siRNA mediated depletion of Cul4B, Cul-7 or Rbx-1. C) Quantitative analysis of ER α protein in MCF-7 cells subjected to siRNA mediated depletion of various Cullin protein isoforms or Rbx-1 followed by adenovirus mediated Skp2 over-expression.

Fig.S3. Phosphorylation of ER α at Serine-294 is Required for its Ubiquitination by Skp2 and Impacts Ability of Skp2 to Alter ER α Protein Levels and Gene Regulation

A) Quantitative analysis of ER α protein in Cos-1 cells subjected to dose-dependent overexpression of Skp2-WT or S294A phospho-mutant. B) Western analysis of extracts from MDA-MB-468 cells transfected with WT or S294A mutant ER α in the presence of control or Skp2 adenovirus and treated with cycloheximide (50 μ g/ml) for the indicated times. C) HEC-1 cells were transfected with 2ERE-pS2-Luciferase expression plasmid along with wild type or S294A ER α and β -galactosidase in the presence or absence of Skp2 adenovirus, treated for 24h with either Veh or E2 (10nM), and monitored for luciferase activity. D) Silver stained gel for mass-spectrometry analysis of ER α upon Skp2 or p38MAPK over-expression in MCF-7 breast cancer cells. E) Quantitative analysis of ER α protein in MCF-7 cells subjected to adenovirus mediated Skp2 over-expression in the presence or absence of p38MAPK inhibitor. F) Quantitative analysis of p38MAPK protein in ER α -negative and positive cell lines.

Fig.S4. ER α Knockdown Blocks the Transcriptional Regulation of ER-Target Genes by Skp2 or p38MAPK, Skp2 and phospho-p38MAPK are Recruited to Target Genes, and Flow Cytometry Reveals Enhancement of S-Phase Entry by Skp2 and p38MAPK

Real-time PCR to monitor expression of ER α target genes, A) TFF1 and B) GREB1 in MCF-7 cells subjected to siRNA against ER α for 24h followed by infection with control, Skp2, p38MAPK or ER α adenovirus also for 24h and treatment with Veh (0.1% EtOH), E2 (10nM) or TOT (1 μ M) for 4h before harvest. The Western blot shows the extent of ER α protein knockdown with siRNA treatment. C) and D) ChIP assays: MCF-7 cells treated for 45 min, 4h or 24h with Veh (0.1% EtOH), E2 (1nM) or TOT (1 μ M), and then washed with cold PBS (pH 7.2) were harvested for ChIP assays using anti-ER α , Skp2, phospho-p38MAPK or IgG negative control antibody. Data are represented as recruitment index (Specific antibody signal/IgG signal ratio) and genes analyzed are p21 and p27. E) and F) ChIP re-ChIP analysis: MCF-7 cells treated with Veh (0.1% EtOH) or E2 (10nM) for 4h, were subjected to ChIP assays as described in Materials and Methods, using anti-Skp2, p38MAPK or IgG negative control antibody. Immunoprecipitated material from ChIP pull-downs were recovered with 10mM dithiothreitol in IP buffer at 37°C for 30 min, diluted and subjected to a second round of immunoprecipitation using specific ER α antibody. Data are represented as recruitment index (Specific antibody signal/ IgG signal ratio) and genes analyzed are TFF1 and GREB1. G) MCF-7 cells were processed and subjected to Propidium Iodide staining followed by Flow-cytometry analysis using BD-FACS-Canto. The graphs show relative number of cells in each phase of the cell cycle which is directly proportional to the surface area of the respective peak of any cell cycle stage. H) MCF-7 cells were synchronized at different stages of the cell cycle as described in Materials and Methods. Propidium Iodide staining followed by Flow-cytometry analysis was performed using BD-FACS-Canto and the relative number of cells in each phase of the cell cycle is shown.

Fig. S5: Impact of loss of ER α in mouse mammary gland or MCF-7 cells on Skp2 and p38MAPK protein levels.

A) Western analysis in mammary tissue protein extracts from WT of ER α knockout (ERKO) mice. Each lane shows extract from a different mouse. B) Western analysis on whole cell extracts from MCF-7 cells transfected with siGL-3 or siRNA against ER α .

Fig.S6. Skp2-S64A Mutant Defective in ER α Interaction/Ubiquitination has No Impact on ER α Target Gene Expression or Cell Proliferation, whereas Constitutively Active Phosphomimic Skp2-S64E Enhances Gene Expression and Cell Proliferation

A) Cos-1 cells transfected with pCMV-Skp2-WT, pCMV-Skp2-S64A (phosphorylation defective mutant) or pCMV-Skp2-S64E (constitutively active phospho-mimic) along with ER α

and HA-Ubiquitin (HA-Ub) or alone and treated with MG-132 for 5h before harvest, 24h after transfection, were subjected to ubiquitination assay for ER α . B) Hec-1 cells transfected with pCMV-2ERE-pS2-Luciferase, ER α and Renilla Luciferase (internal control to assess transfection efficiency) along with pCMV-Skp2-WT, Skp2-S64A or Skp2-S64E were treated for 24h with either Veh (0.1% EtOH) or E2 (10nM), 6h post transfection and thereafter were harvested and monitored for luciferase activity. C) Proliferation of MCF-7 cells with over-expressed Skp2-WT, Skp2-S64A or Skp2-S64E mutant followed by treatment with Veh (0.1% EtOH) or 10nM E2 and analysis of cell density using WST reagent.

Fig. S7: Assessment of Changes in ER α (ESR1) mRNA after Knockdown of Skp2 or p38MAPK Inhibition in MDA-MB-468 and MDA-MB-453 Cells.

Real-time q-PCR analysis of ER α mRNA in A) MDA-MB-468 or B) MDA-MB-453 cells transfected with scramble (control) or Skp2 siRNA, or treated or not with p38MAPK inhibitor for 24h followed by control vehicle, estradiol (E2, 10nM) or tamoxifen (TOT, 1 μ M) treatment for 24h.

Fig. S8: Effect of knockdown of Skp2 and p38MAPK on expression of pS2 and GREB1 mRNA in MCF-7 cells.

A) Real-time q-PCR analysis of pS2/TFF1 mRNA or B) GREB1 mRNA in MCF-7 cells transfected with scramble (control), Skp2, or p38MAPK siRNA for 24h followed by 24h treatment with control vehicle or estradiol (E2, 10nM).

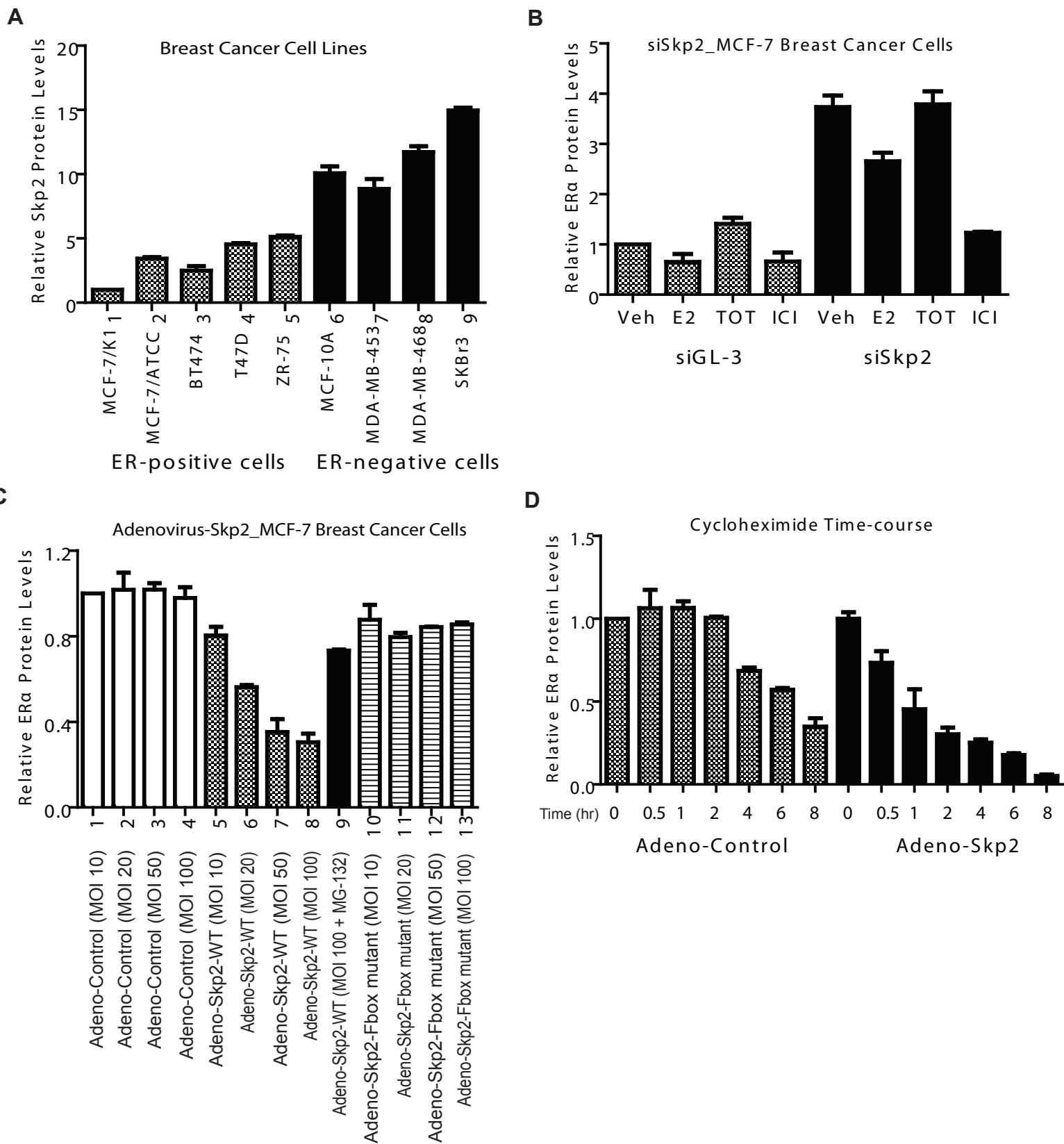


Fig.S1

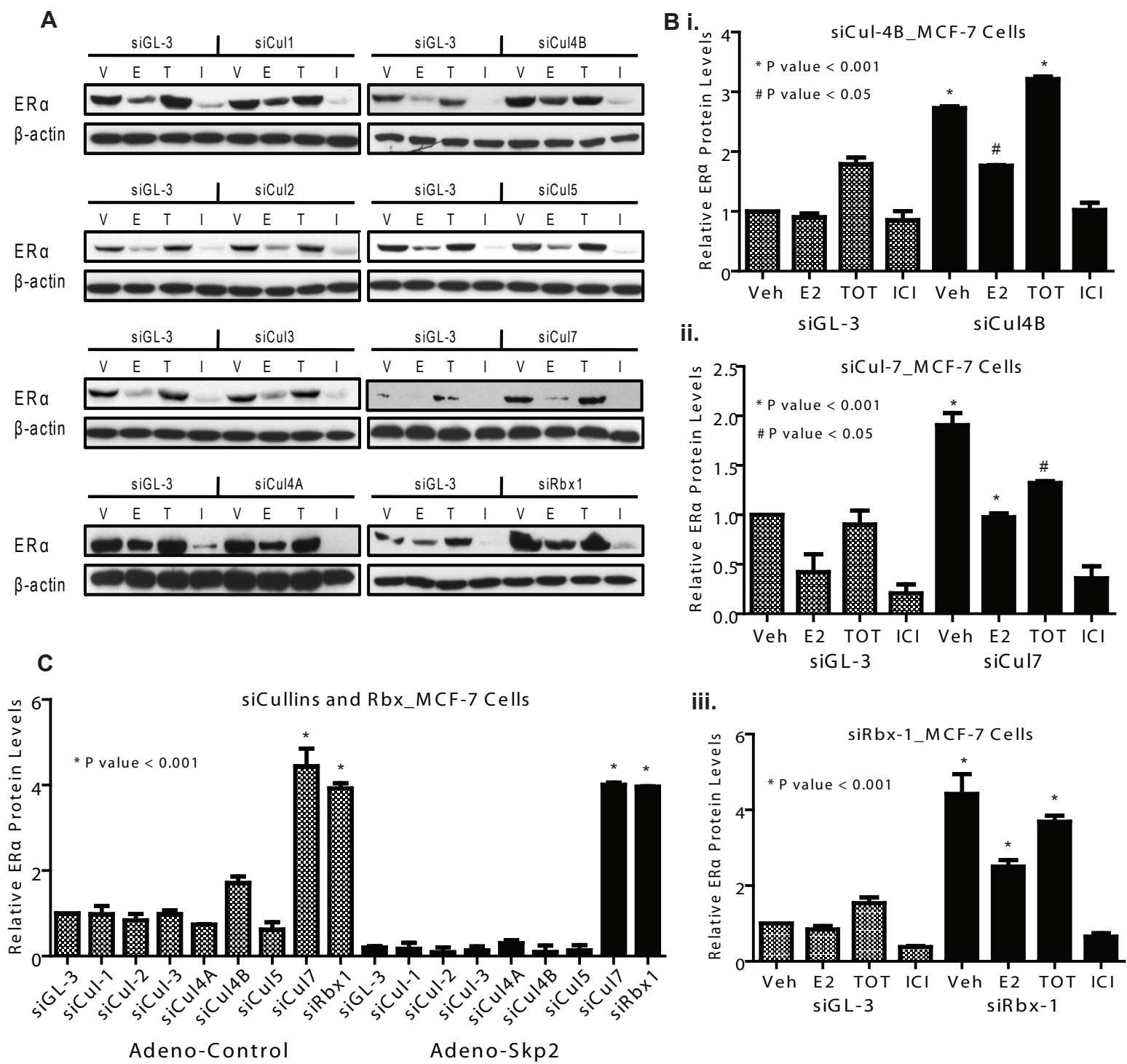


Fig.S2

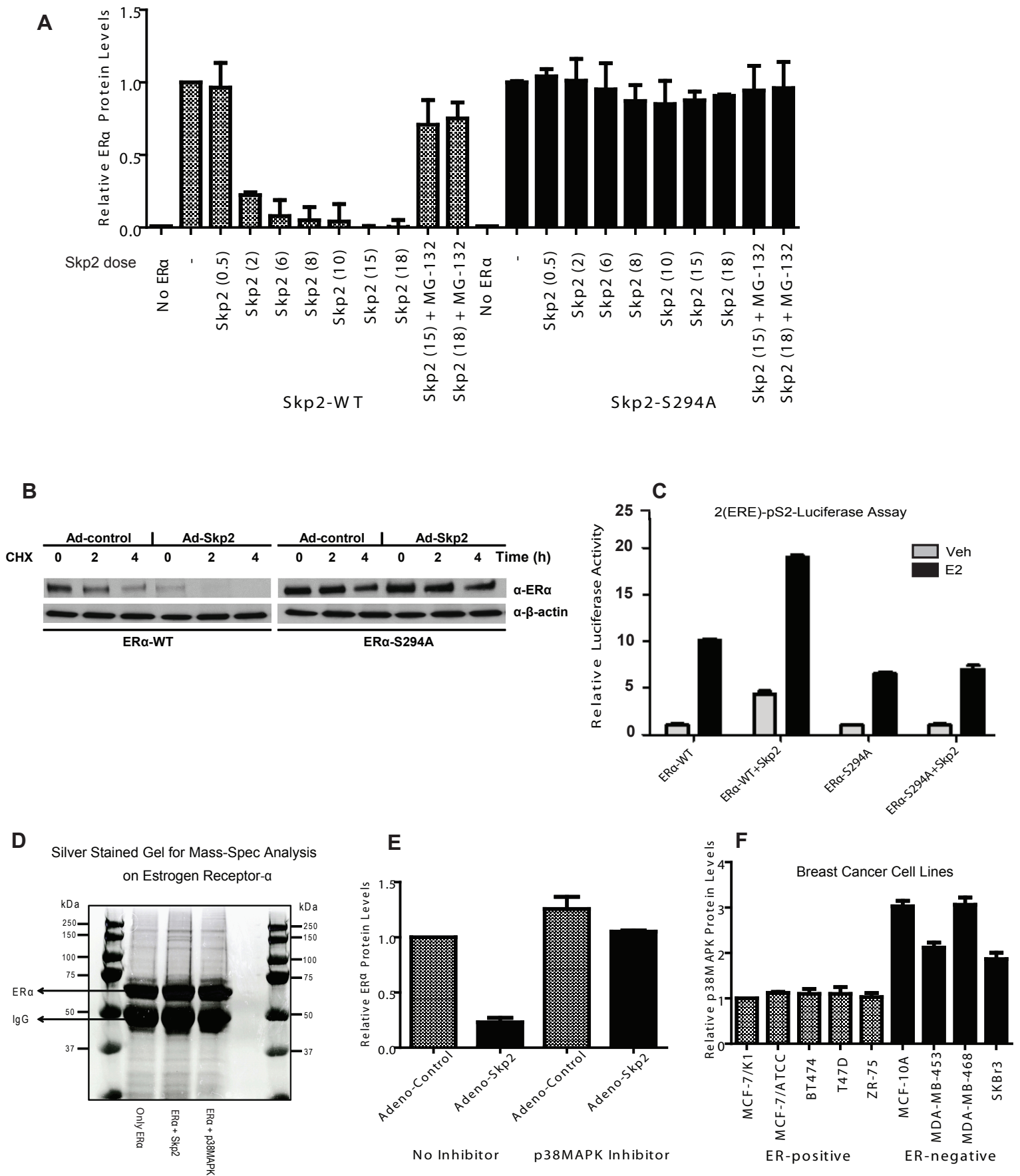


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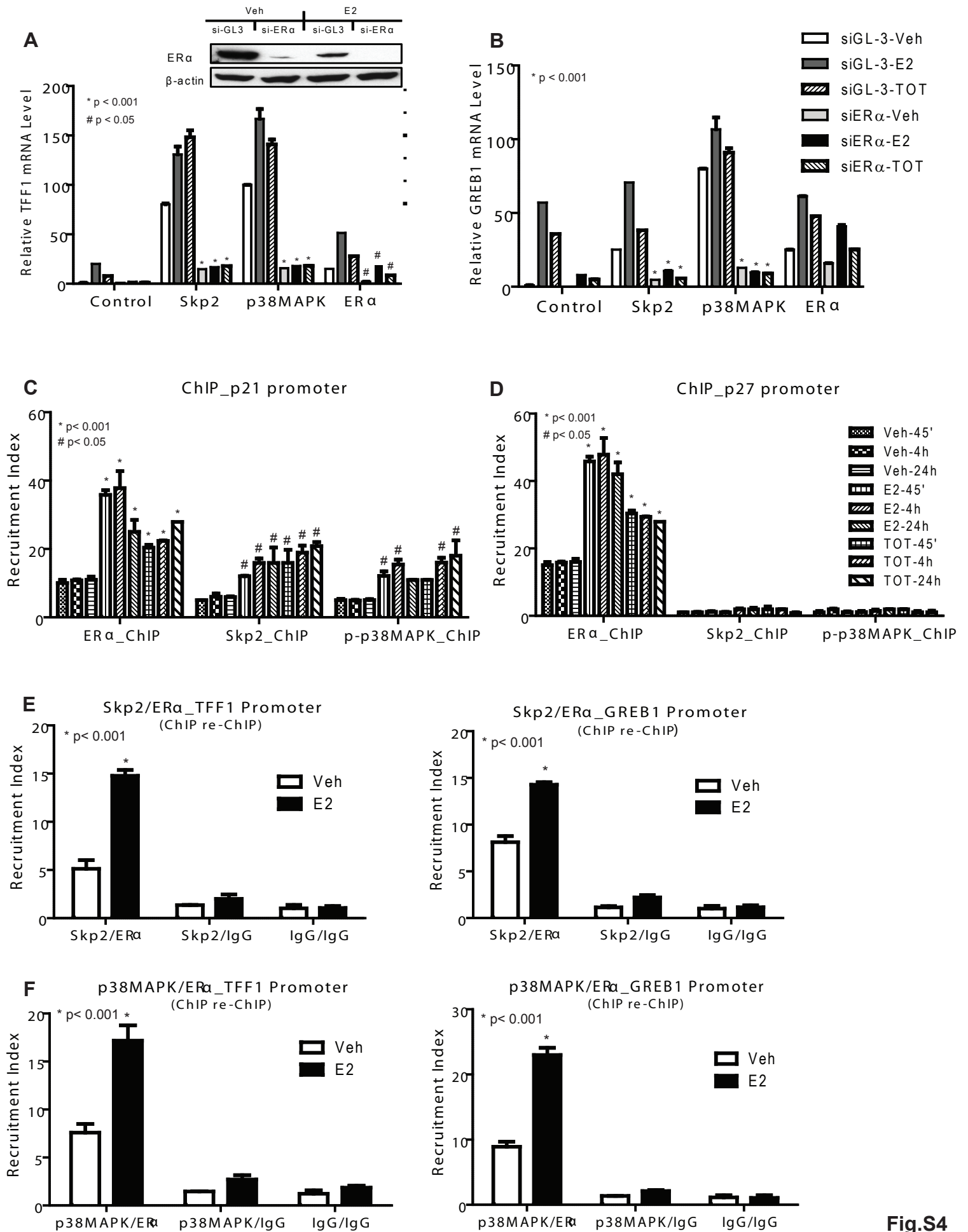
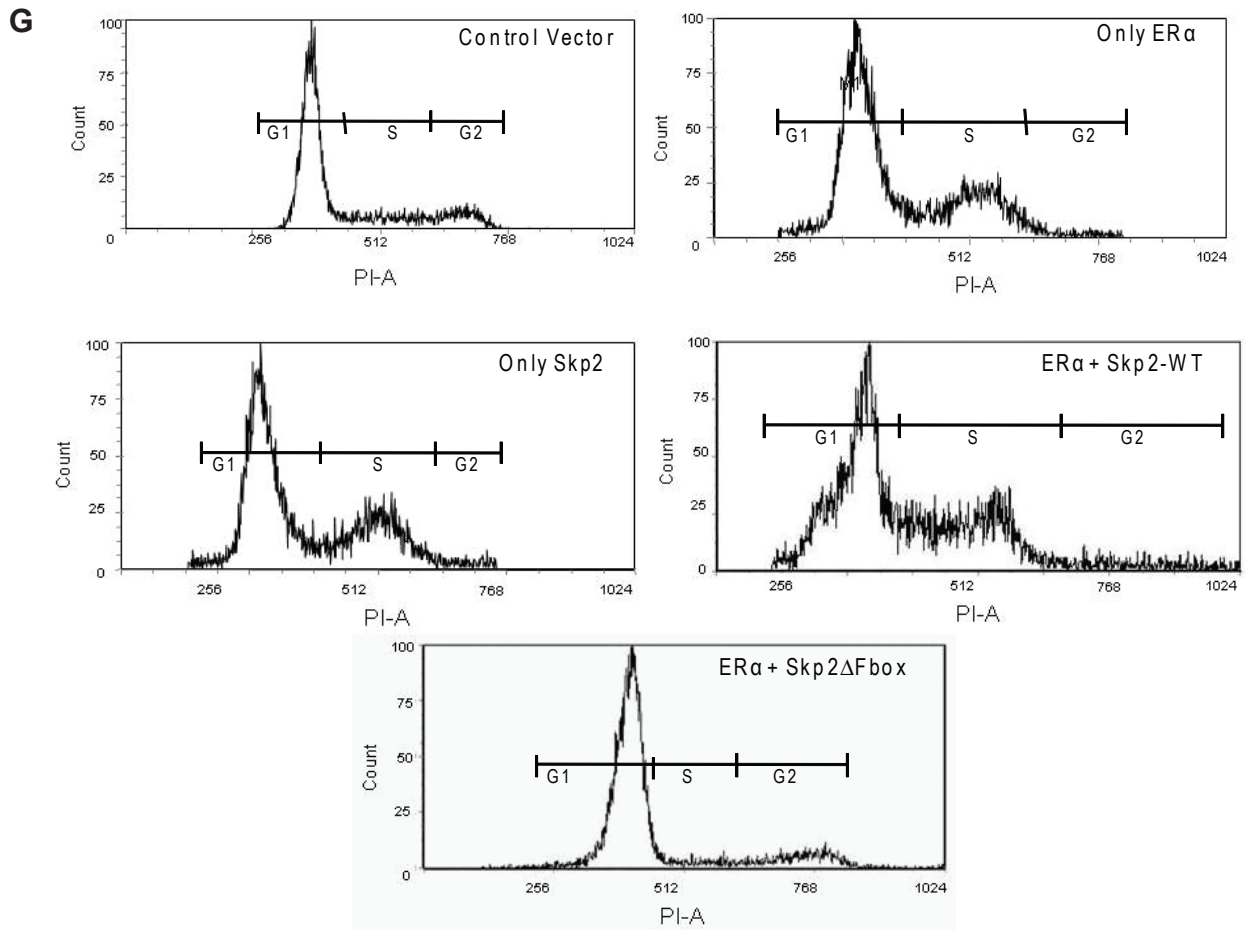


Fig.S4



Flow-cytometry analysis: S-phase Entry of MCF-7 Breast Cancer Cells

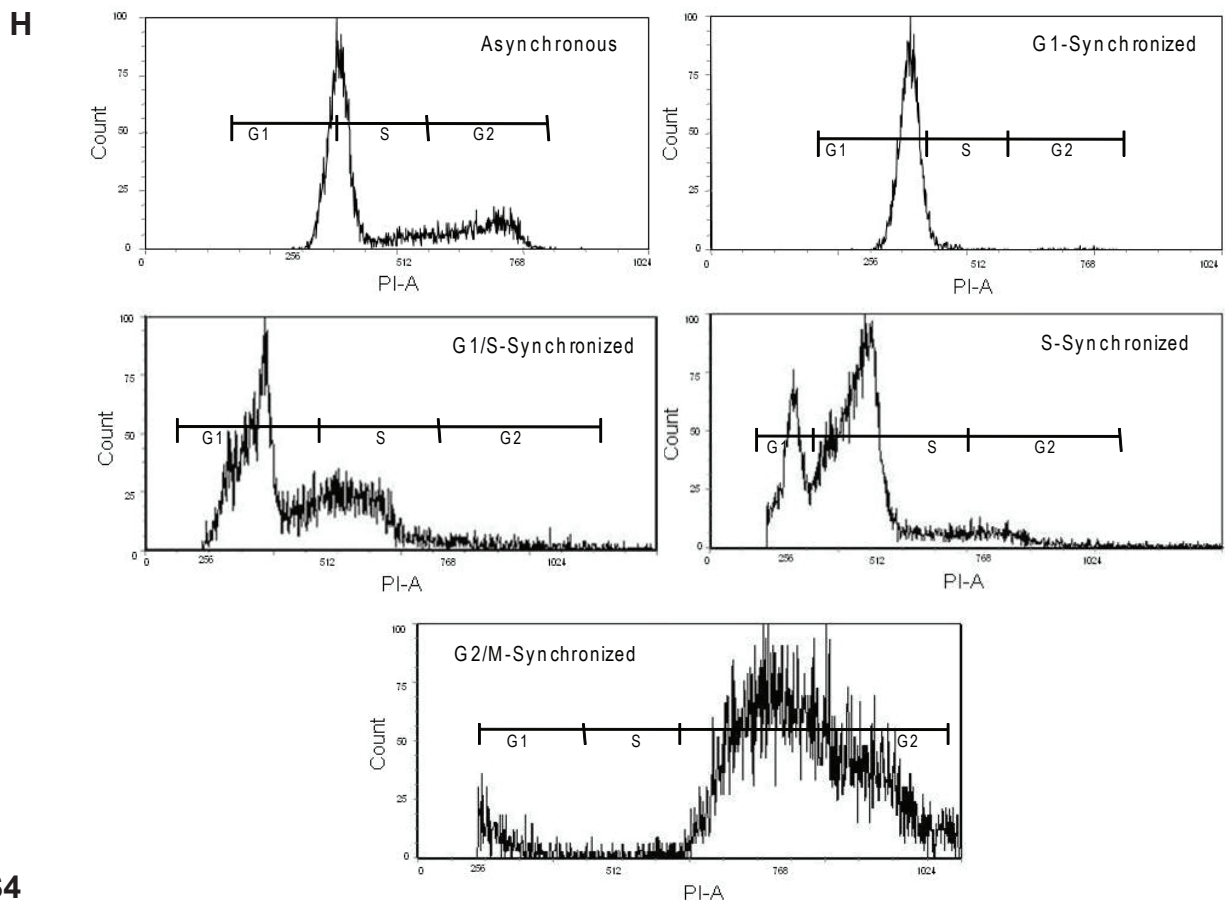


Fig.S4

Flow-cytometry analysis : Synchronization of MCF-7 Breast Cancer Cells

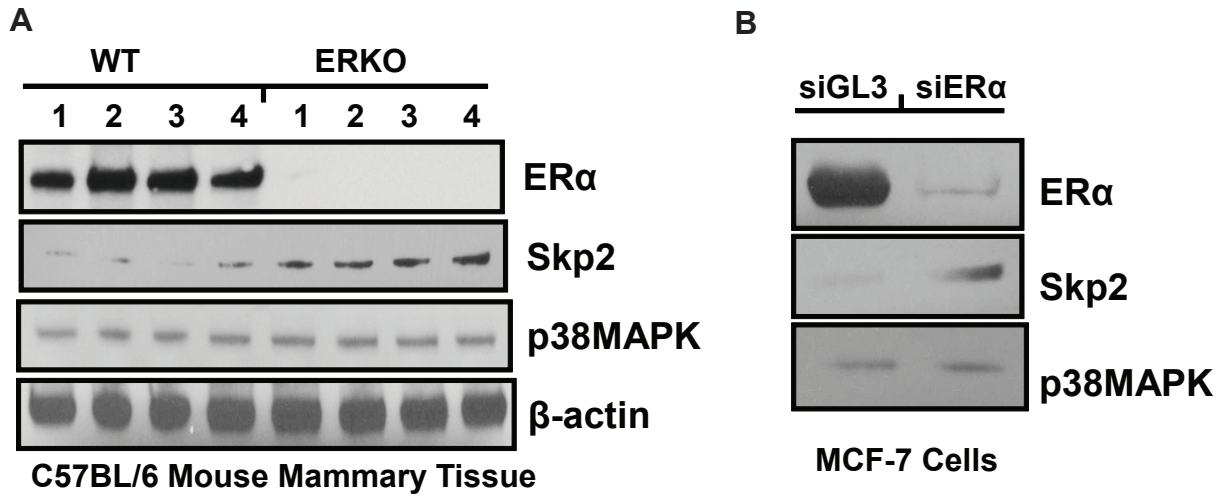


Fig.S5

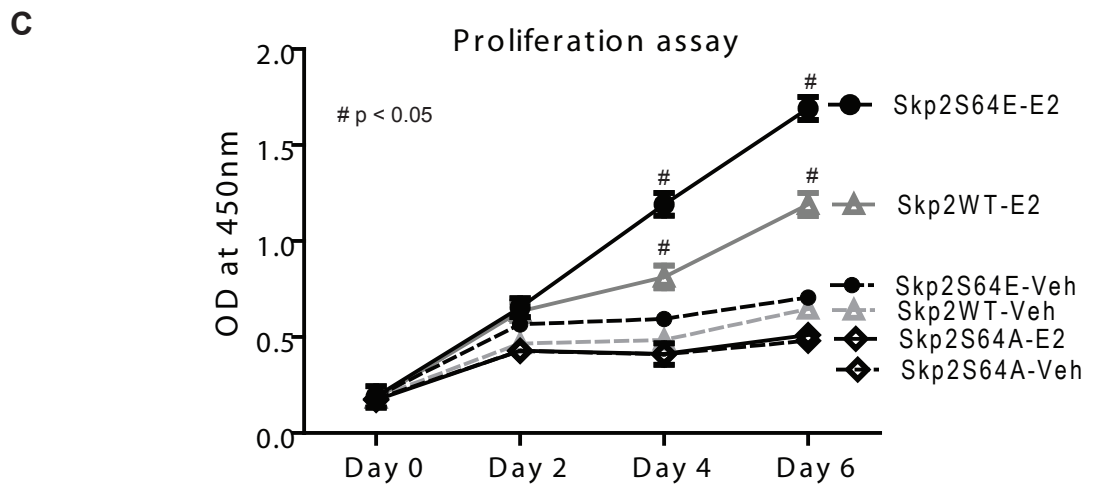
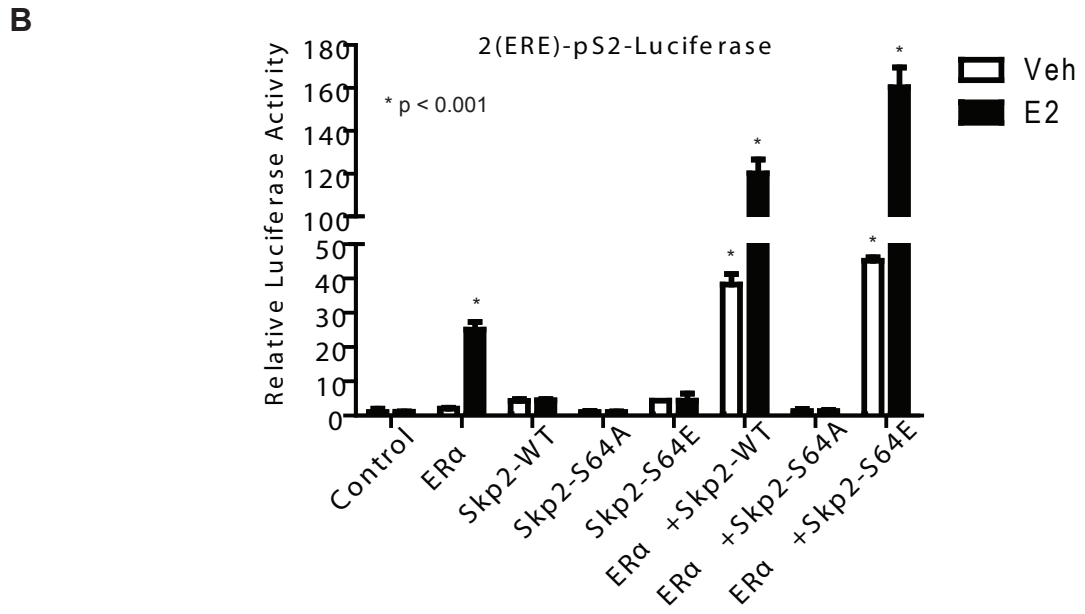
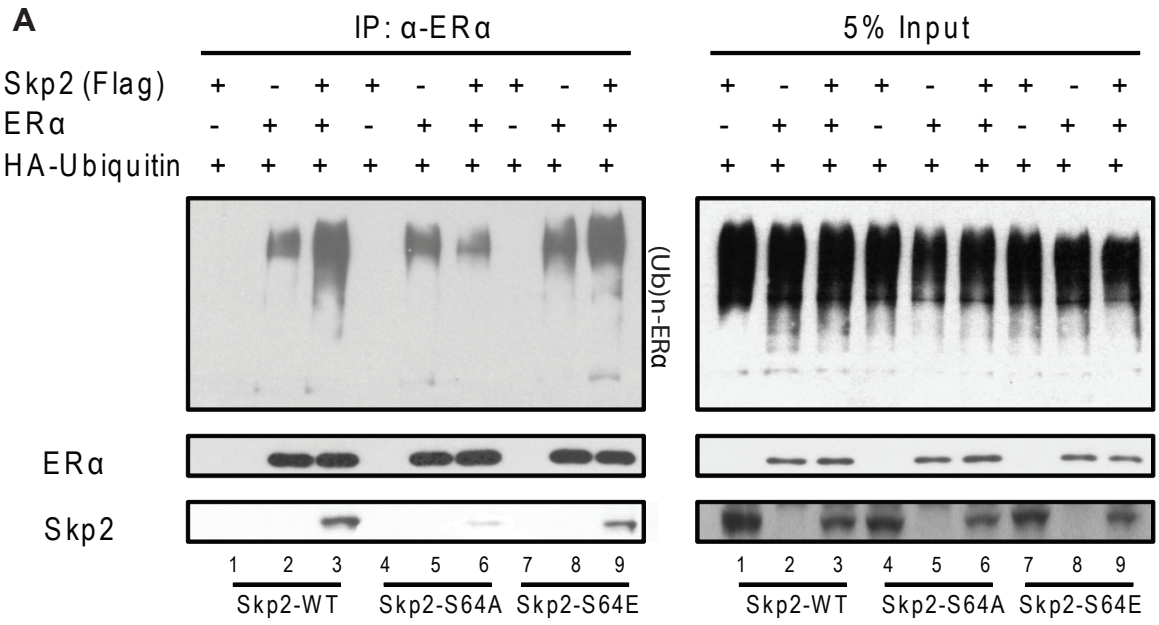


Fig.S6

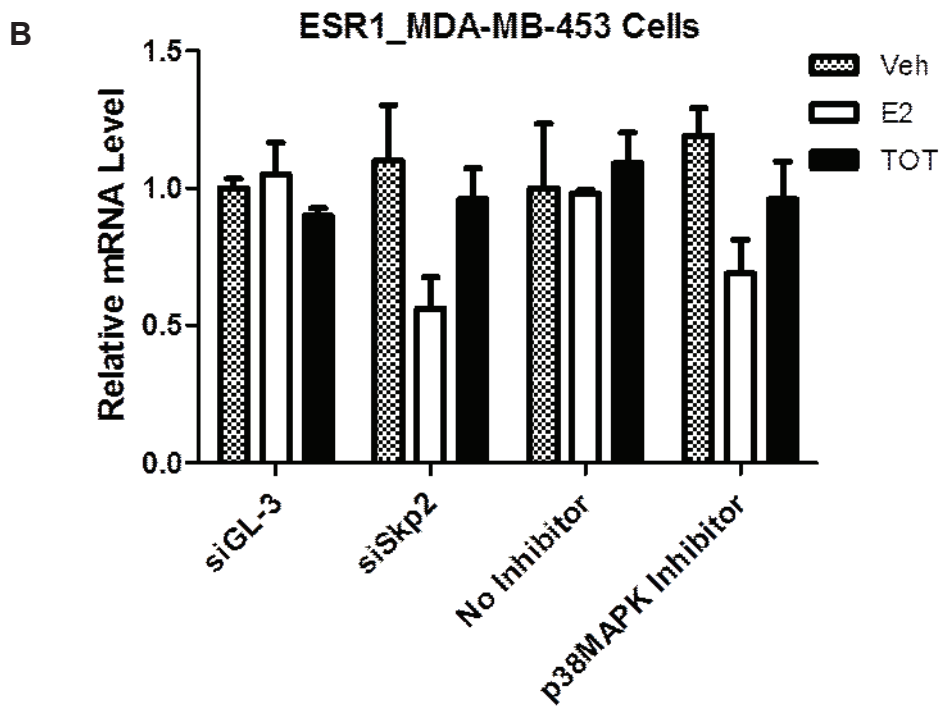
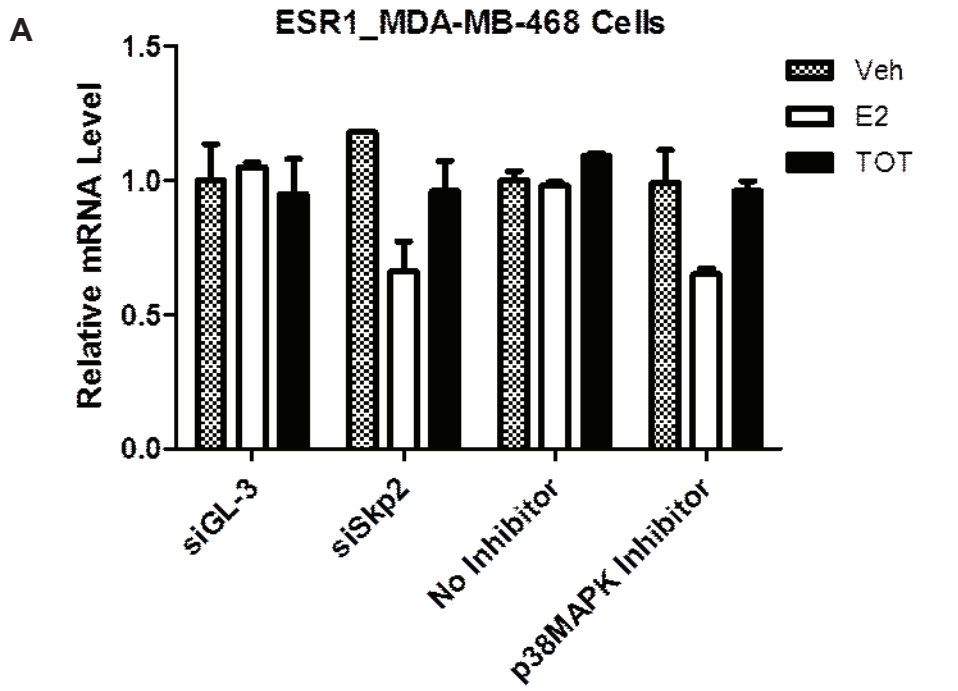


Fig.S7

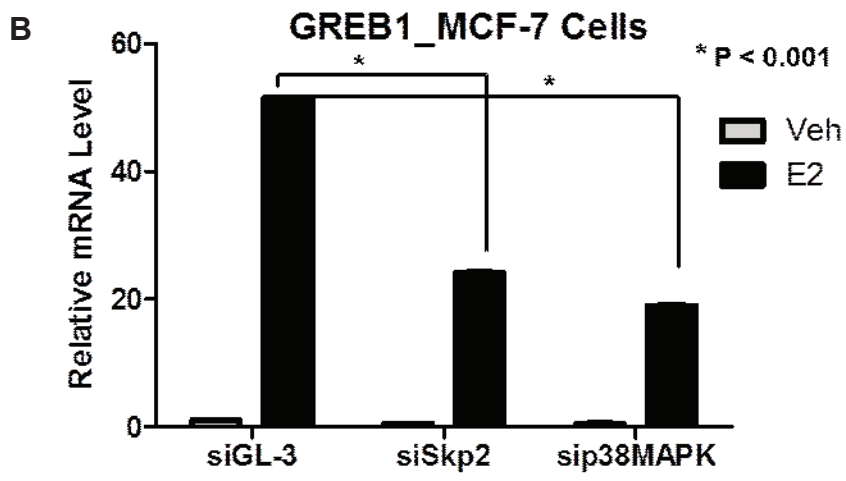
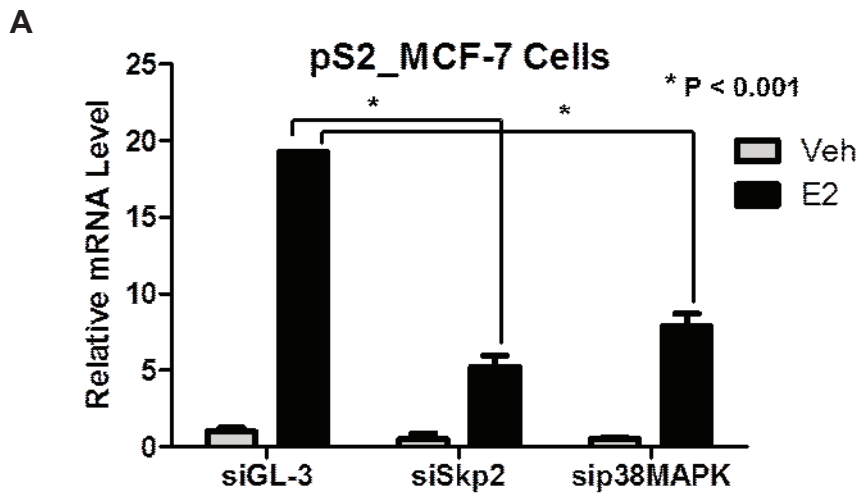


Fig.S8