

Supporting Information

Stereoselective Assembly of Complex Oligosaccharides Using Anomeric Sulfonium Ions as Glycosyl Donors

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1. Preparations and characterizations of prepared compounds

Reagents and General Procedures. Reagents were obtained from commercial sources and used as purchase. Dichloromethane (DCM) was freshly distilled using standard procedures. Other organic solvents were purchased anhydrous and used without further purification. Unless otherwise noted, all reactions were carried out at room temperature in oven-dried glassware with magnetic stirring. Molecular sieves were flame dried *in vacuo* prior to use. Organic solutions were concentrated under diminished pressure with bath temperature < 40 °C. Flash column chromatography was carried out on silica gel G60 (Silicycle, 60-200 µm, 60 Å). Thin-layer chromatography (TLC) was carried out on Silica gel 60 F₂₅₄ (EMD Chemicals Inc.) and detection was performed by UV absorption (254 nm) where applicable, and by spraying with 20% sulfuric acid in ethanol followed by charring at ~150 °C, or by spraying with a solution of (NH₄)₆Mo₇O₂₄H₂O (25 g/L) in 10% sulfuric acid in ethanol followed by charring at ~150 °C. Optical rotations were measured by ATAGO POLAX-2L with 1 ml (5 cm) micro observation tube at 589 nm. ¹H and ¹³C NMR spectra were recorded on a Varian Inova-300 (300/75 MHz), a Varian Inova-500 (500 MHz) and a Varian Inova-600 (600/150 MHz) spectrometer equipped with sun workstations. Multiplicities are quoted as singlet (s), doublet (d), doublet of doublets (dd), triplet (t), or multiplet (m). All NMR signals were assigned on the basis of ¹H NMR, ¹³C NMR, gCOSY, and gHSQC experiments. All chemical shifts are quoted on the δ-scale in parts per million (ppm). Signals marked with a superscript Roman numeral I were the reducing end, whereas II and III were the second and the third sugar from the reducing end, and IV is the branching sugar. Residual solvent signals were used as an internal reference. Mass spectra were recorded on an Applied Biosystems 5800 MALDI-TOF

mass spectrometer. The matrix used was 2,5-dihydroxy-benzoic acid (DHB). Reverse-Phase HPLC was performed on an Agilent 1200 series system equipped with an auto-sampler, fraction-collector, UV-detector, and eclipse XDB-C18 column (5 μ m, 4.6 \times 250 mm or 9.4 \times 250 mm).

2-(S)-Phenyl-(1,2-dideoxy- β -D-glucopyranoso)[1,2-*e*]-1,4-oxathiane (2). Compound **1** (11 g, 35 mmol) was dissolved in anhydrous CH₃CN (400 mL) and hexamethyldisiloxane (TMS₂O) (44 mL, 0.21 mol) and TMSOTf (6.6 mL, 37 mmol) were added. After 30 min, Et₃SiH (46 mL, 0.29 mol) was added and the reaction mixture was stirred for another 4 h before quenching by the addition of MeOH (50 mL) and Et₃N (10 mL). The solution was concentrated *in vacuo* and the resulting yellow oil was purified by flash chromatography over silica gel (toluene/acetone, 3/1 \rightarrow 1/1, v/v) to give **2** (7.7 g, 74%). Proton chemical shifts are identical to reported data.^[1] ¹H NMR (300 MHz, CDCl₃) δ 7.49 – 7.09 (m, 5H, ArH), 4.67 (dd, *J* = 10.6, 1.6 Hz, 1H, SCH₂CHPh), 4.35 (d, *J* = 8.2 Hz, 1H, H-1), 3.84 – 3.71 (m, 2H, H-6_{a,b}), 3.70 – 3.43 (m, 3H, H-2, H-3, H-4), 3.43 – 3.29 (m, 1H, H-5), 3.00 (dd, *J* = 13.9, 10.8 Hz, 2H, SCHHCHPh), 2.69 (d, *J* = 12.4 Hz, 1H, SCHHCHPh).

2-(S)-Phenyl-(4,6-O-benzylidene-1,2-dideoxy- β -D-glucopyranoso)[1,2-*e*]-1,4-oxathiane (3). Compound **2** (3.3 g, 11 mmol) was dissolved in DMF (60 mL) and benzaldehyde dimethyl acetal (2.5 mL, 17 mmol) and camphorsulfonic acid (40 mg, 0.17 mmol) were added. The reaction mixture was heated at 50 °C under reduced pressure (~15 mm Hg) for 16 h after which it was quenched by adding Et₃N (0.5 mL). The mixture was diluted with DCM (300 mL) and the organic solution was washed with water (2 \times 200 mL) and brine (150 mL). The organic phase was dried (MgSO₄), filtered and the

filtrate was concentrated *in vacuo*. The resulting yellow oil was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/5→1/2, v/v) to give **3** (3.7 g, 87 %). $R_f = 0.26$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{26}^d$ (deg cm³ g⁻¹ dm⁻¹) = +157.1 ($c = 0.7$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.64 – 7.27 (m, 10H, ArH), 5.56 (s, 1H, PhCH<), 4.73 (dd, $J = 10.7, 2.0$ Hz, 1H, SCH₂CHPh), 4.54 (d, $J = 8.9$ Hz, 1H, H-1), 4.37 (dd, $J = 10.3, 4.6$ Hz, 1H, H-6_a), 3.91 (t, $J = 8.8$ Hz, 1H, H-3), 3.85 – 3.56 (m, 4H, H-4, H-5, H-2, H-6_b), 3.09 (dd, $J = 14.1, 10.8$ Hz, 1H, SCHHCHPh), 2.77 (dd, $J = 14.0, 2.0$ Hz, 1H, SCHHCHPh); ¹³C NMR (75 MHz, CDCl₃): δ 140.20, 137.09, 129.49, 128.86, 128.60, 128.54, 126.56, 126.20, 102.28, 84.67, 81.15, 80.89, 77.69, 77.26, 76.84, 76.47, 72.34, 72.04, 68.64, 35.98; HR MALDI-TOF MS: m/z : calcd for C₂₁H₂₂O₅S [M+Na]⁺: 409.1086; found: 409.1097.

2-(S)-Phenyl-(3-O-acetyl-4,6-O-benzylidene-1,2-dideoxy- β -D-glucopyranoso)[1,2-*e*]-1,4-oxathiane (4). Compound **3** (1.35 g, 3.50 mmol) was dissolved in pyridine (10 mL) and acetic anhydride (5 mL) was added. After stirring for 16 h, the reaction mixture was diluted with DCM (120 mL) and washed with saturated NaHCO₃ (2 × 100 mL) and brine (90 mL). The organic phase was dried (MgSO₄), filtered and the filtrate was concentrated *in vacuo*. The resulting yellow oil was purified by flash chromatography over silica gel (EtOAc/Toluene, 1/16→1/8, v/v) to give **4** (1.47 g, 98%). $R_f = 0.53$ (EtOAc/Toluene, 1/8, v/v); $[\alpha]_{26}^d$ (deg cm³ g⁻¹ dm⁻¹) = +40.0 ($c = 0.5$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.53 – 7.16 (m, 10H, ArH), 5.53 (s, 1H, PhCH<), 5.42 (t, $J = 9.4$ Hz, 1H, H-3), 4.72 (dd, $J = 10.6, 1.9$ Hz, 1H, SCH₂CHPh), 4.62 (d, $J = 8.9$ Hz, 1H, H-1), 4.39 (dd, $J = 9.9, 4.1$ Hz, 1H, H-6_a), 3.87 – 3.65 (m, 4H, H-2, H-4, H-5, H-6_b), 3.02 (dd, $J = 14.0, 10.6$ Hz, 1H, SCHHCHPh), 2.84 (dd, $J = 14.0, 2.1$ Hz, 1H, SCHHCHPh), 2.04 (s, 3H); ¹³C NMR (75

MHz, CDCl₃): δ 170.29, 140.18, 137.03, 129.34, 128.72, 128.47, 128.09, 126.36, 125.52, 101.84, 82.67, 80.10, 79.38, 77.66, 77.24, 76.93, 76.82, 72.48, 71.80, 68.61, 36.10, 21.17; HR MALDI-TOF MS: m/z: calcd for C₂₃H₂₄O₆S [M+Na]⁺: 451.1192; found: 451.1201.

2-(S)-Phenyl-(3-O-acetyl-4,6-O-benzylidene-1,2-dideoxy- β -D-glucopyranoso)[1,2-e]-1,4-oxathiane (5). Levulinic acid (464 μ L, 4.56 mmol), *N,N'*-dicyclohexylcarbodiimide (DCC) (940 mg, 4.56 mmol) and 4-dimethylaminopyridine (DMAP) (44 mg, 0.36 mmol) were added to a stirred solution of **3** (440 mg, 1.14 mmol) in DCM (8 mL). After stirring for 16 h, the reaction mixture was filtered and the organic layer was concentrated *in vacuo*. The resulting oil was purified by flash chromatography over silica gel (toluene/acetone, 15/1 \rightarrow 5/1, v/v) to give **5** (436 mg, 79%). *R_f* = 0.5 (toluene/acetone, 6/1, v/v); $[\alpha]_{26}^d$ (deg cm³ g⁻¹ dm⁻¹) = +66.7 (*c* = 1.2 in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.52 – 7.16 (m, 10H, ArH), 5.53 (s, 1H, PhCH<), 5.38 (t, *J* = 9.4 Hz, 1H, H-3), 4.71 (dd, *J* = 10.5, 1.9 Hz, 1H, SCH₂CHPh), 4.61 (d, *J* = 8.9 Hz, 1H, H-1), 4.39 (dd, *J* = 10.0, 4.3 Hz, 1H, H-5), 3.87 – 3.61 (m, 4H, H-2, H-4, H-6_{a,b}), 3.02 (dd, *J* = 14.0, 10.6 Hz, 1H, SCHHCHPh), 2.83 (dd, *J* = 14.0, 2.1 Hz, 1H, SCHHCHPh), 2.71 – 2.45 (m, 4H, 2 \times CH₂ Lev), 1.99 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 206.24, 172.27, 140.12, 137.01, 129.29, 128.68, 128.44, 128.11, 126.37, 125.61, 101.78, 82.67, 80.13, 79.31, 77.67, 77.25, 76.86, 76.83, 72.47, 72.11, 68.59, 38.45, 36.00, 29.82, 28.39.; HR MALDI-TOF MS: m/z: calcd for C₂₁H₂₁O₄S [M+Na]⁺: 392.1059; found: 392.1051.

2-(S)-Phenyl-(3-O-allyloxycarbonyl-4,6-O-benzylidene-1,2-dideoxy- β -D-glucopyranoso)[1,2-e]-1,4-oxathiane (6). Allyl chloroformate (274 μ L, 2.56 mmol), tetramethylethylenediamine (TMEDA) (200 μ L, 1.22 mmol) and 4-dimethylaminopyridine (DMAP) (50 mg, 0.41 mmol) were added to a stirred solution of

3 (470 mg, 1.22 mmol) in DCM (10 mL). After stirring for 16 h, the reaction mixture was diluted with DCM (100 mL) and washed with saturated NaHCO₃ (2 × 100 mL), brine (90 mL). The organic phase was dried (MgSO₄), filtered and the filtrate was concentrated *in vacuo*. The resulting yellow oil was purified by flash chromatography over silica gel (toluene/acetone, 20/1→8/1, v/v) to give **6** (500 mg, 87%). *R_f* = 0.56 (toluene/acetone, 8/1, v/v); [α]₂₆^d (deg cm³ g⁻¹ dm⁻¹) = +53.8 (*c* = 1.3 in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.54 – 7.19 (m, 10H, ArH), 5.91 – 5.70 (m, 1H, CH alloc), 5.53 (s, 1H, PhCH<), 5.28 – 5.13 (m, 2H, CHH alloc, H-3), 5.08 (d, *J* = 10.5 Hz, 1H, CHH alloc), 4.72 (dd, *J* = 10.6, 1.8 Hz, 1H, SCH₂CHPh), 4.63 (d, *J* = 8.9 Hz, 1H, H-1), 4.57 (d, *J* = 5.7 Hz, 1H, CH₂ alloc), 4.40 (dd, *J* = 10.0, 4.1 Hz, 1H, H-6_a), 3.90 – 3.65 (m, 4H, H-2, H-4, H-5, H-6_b), 3.05 (dd, *J* = 14.0, 10.7 Hz, 1H, SCHHCHPh), 2.83 (dd, *J* = 14.0, 2.0 Hz, 1H, SCHHCHPh); ¹³C NMR (75 MHz, CDCl₃): δ 154.59, 140.05, 136.94, 131.41, 129.35, 128.62, 128.43, 128.13, 126.42, 125.70, 119.03, 101.89, 82.57, 80.18, 79.16, 77.66, 77.44, 77.24, 76.82, 76.73, 76.03, 72.35, 68.92, 68.56, 35.98; HR MALDI-TOF MS: *m/z*: calcd for C₂₅H₂₆O₇S [M+Na]⁺: 493.1297; found: 493.1288.

2-(S)-Phenyl-(3-O-benzyl-4,6-O-benzylidene-1,2-dideoxy-β-D-glucopyranoso)[1,2-*e*]-1,4-oxathiane (7). Benzyl bromide (246 μL, 2.08 mmol) and sodium hydride (124 mg, 3.12 mmol) were added to a stirred solution of **3** (400 mg, 1.04 mmol) in DMF (5 mL). After stirring for 16 h, the reaction mixture was quenched with MeOH (2 mL), diluted with DCM (100 mL) and washed with 1 M HCl solution (100 mL), saturated NaHCO₃ (100 mL), and brine (90 mL). The organic phase was dried (MgSO₄), filtered and the filtrate was concentrated *in vacuo*. The resulting yellow oil was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/4 → 1/2, v/v) to give **7** (390 mg, 79%).

$R_f = 0.28$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{26}^d$ (deg cm³ g⁻¹ dm⁻¹) = +54.5 ($c = 1.1$ in CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ 7.56 – 7.16 (m, 15H, ArH), 5.60 (s, 1H, PhCH<), 4.85 (s, 2H, CH₂Ph), 4.78 (d, $J = 10.6$ Hz, 1H, SCH₂CHPh), 4.53 (d, $J = 8.8$ Hz, 1H, H-1), 4.37 (dd, $J = 10.4, 4.8$ Hz, 1H, H-4), 3.88 – 3.72 (m, 4H, H-2, H-3, H-6_{a,b}), 3.61 (td, $J = 9.5, 4.9$ Hz, 1H, H-5), 3.06 (dd, $J = 13.9, 10.9$ Hz, 1H, SCHHCHPh), 2.81 (d, $J = 13.9$ Hz, 1H, SCHHCHPh); ¹³C NMR (75 MHz, CDCl₃): δ 140.55, 138.58, 137.42, 129.22, 128.68, 128.46, 128.40, 128.23, 127.76, 126.28, 125.91, 101.65, 84.99, 81.71, 80.40, 78.79, 77.66, 77.24, 76.99, 76.82, 74.83, 72.37, 68.68, 36.09; HR MALDI-TOF MS: m/z: calcd for C₂₈H₂₈O₅S [M+Na]⁺: 499.1555; found: 499.1562.

2-(S)-Phenyl-(3,4-di-O-acetyl-6-O-benzyl-1,2-dideoxy- β -D-glucopyranoso)[1,2-e]-1,4-oxathiane (9). A mixture of **4** (550 mg, 1.29 mmol) and activated molecular sieves (4Å) in DCM (5 mL) was stirred for 1 h under an atmosphere of argon. After cooling to -78 °C, triethylsilane (408 μ L, 2.58 mmol) and trifluoromethanesulfonic acid (171 μ L, 1.94 mmol) were added. After 1 h, the reaction was quenched by the addition of MeOH (1 mL) and Et₃N (0.5 mL). The mixture was diluted with DCM (10 mL), filtered and the filtrate was concentrated *in vacuo*. The residue was redissolved in pyridine (5 mL) and acetic anhydride (5 mL) was added. After stirring for 16 h, the solvent was removed *in vacuo* and the resulting residue was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/6 \rightarrow 1/2, v/v) to afford **9** (516 mg, 85%). $R_f = 0.37$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{26}^d$ (deg cm³ g⁻¹ dm⁻¹) = +141.7 ($c = 1.2$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.43 – 7.18 (m, 10H, ArH), 5.34 – 5.10 (m, 2H, H-3, H-4), 4.69 (dd, $J = 10.5, 1.8$ Hz, 1H, SCH₂CHPh), 4.58 – 4.44 (m, 3H, CH₂Ph, H-1), 3.86 – 3.66 (m, 2H, H-5, H-2), 3.62 – 3.50 (m, 2H, H-6_{a,b}), 2.97 (dd, $J = 14.0, 10.6$ Hz, 1H, SCHHCHPh), 2.80 (dd, J

= 14.0, 2.1 Hz, 1H, SCHHCHPh), 1.99 (s, 3H), 1.92 (s, 3H).; ^{13}C NMR (75 MHz, CDCl_3): δ 170.65, 169.83, 140.32, 137.84, 128.69, 128.61, 128.22, 128.07, 128.00, 125.57, 81.30, 79.75, 78.81, 77.71, 77.29, 76.86, 76.01, 73.84, 73.38, 69.62, 68.89, 35.86, 21.03, 20.89; HR MALDI-TOF MS: m/z: calcd for $\text{C}_{25}\text{H}_{28}\text{O}_7\text{S}$ $[\text{M}+\text{Na}]^+$: 495.1454; found: 495.1463.

2-(S)-Phenyl-(3-O-acetyl-4,6-di-O-benzyl-1,2-dideoxy- β -D-glucopyranoso)[1,2-e]-1,4-oxathiane (10). A mixture of **4** (430 mg, 1.00 mmol) and activated molecular sieves (4 Å) in DCM (5 mL) was stirred for 1 h under an atmosphere of argon. After cooling to -78 °C, triethylsilane (316 μL , 2.00 mmol) and trifluoromethanesulfonic acid (132 μL , 1.50 mmol) were added. After 1 h, the reaction was quenched by the addition of MeOH (1 mL) and Et_3N (0.5 mL). The mixture was diluted with DCM (10 mL), filtered and the filtrate was concentrated *in vacuo*. The resulting residue was loaded onto a small plug of silica gel and product fractions were collected and concentrated. The residue was redissolved in DMF (5 mL) followed by the addition of benzyl bromide (356 μL , 3.00 mmol) and Ag_2O (1.2 g, 5.2 mmol). After stirring for 16 h, the reaction mixture was filtered and the filtrate was concentrated *in vacuo*. The resulting residue was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/6 \rightarrow 1/3, v/v) to afford **10** (287 mg, 55%). R_f = 0.45 (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{26}^{\text{d}}$ (deg $\text{cm}^3 \text{g}^{-1} \text{dm}^{-1}$) = +66.7 (c = 0.3 in CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ 7.45 – 7.09 (m, 15H, ArH), 5.30 (dd, J = 11.0, 7.9 Hz, 1H, H-3), 4.72 – 4.50 (m, 5H, $2\times\text{CH}_2\text{Ph}$, SCH_2CHPh), 4.46 (d, J = 8.9 Hz, 1H, H-1), 3.83 (t, J = 9.5 Hz, 1H, H-4), 3.76 (d, J = 2.8 Hz, 2H, H-6_{a,b}), 3.72 – 3.55 (m, 2H, H-2, H-5), 2.95 (dd, J = 14.0, 10.5 Hz, 1H, SCHHCHPh), 2.79 (dd, J = 14.0, 2.1 Hz, 1H, SCHHCHPh) 1.93 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 170.34, 140.49, 138.13, 137.99, 128.65, 128.21, 128.10, 128.06, 127.98, 127.91, 125.49, 81.77, 80.59, 79.57, 77.66, 77.24,

76.81, 76.18, 75.93, 75.27, 74.84, 73.85, 68.59, 35.86, 21.21; HR MALDI-TOF MS: m/z: calcd for C₃₀H₃₂O₆S [M+Na]⁺: 543.1818; found: 543.1824.

2-(S)-Phenyl- $\{$ 3-O-acetyl-6-O-benzyl-4-O-(9-fluorenylmethyloxycarbonyl)-1,2-dideoxy- β -D-glucopyranoso $\}$ [1,2-*e*]-1,4-oxathiane (11). A mixture of **4** (1.05 g, 2.45 mmol) and activated molecular sieves (4 Å) in DCM (10 mL) was stirred for 1 h under an atmosphere of argon. After cooling to -78 °C, triethylsilane (0.78 mL, 4.90 mmol) and trifluoromethanesulfonic acid (323 μL, 3.68 mmol) were added. After 1 h, the reaction was quenched by the addition of MeOH (1 mL) and Et₃N (0.5 mL). The resulting mixture was diluted with DCM (10 mL), filtered and the filtrate was concentrated *in vacuo*. The resulting residue was redissolved in a mixture of pyridine and DCM (10 mL, 1/1, v/v) and FmocCl (0.95 g, 3.70 mmol) was added. After stirring for 16 h, the reaction mixture was diluted with DCM (50 mL), poured into an aqueous 1 M HCl solution (100 mL) and washed with H₂O (100 mL) and brine (100 mL). The organic layer dried (MgSO₄) and concentrated *in vacuo*. The resulting residue was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/6 → 1/3, v/v) to afford **11** (1.06 g, 66%). *R_f* = 0.47 (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{26}^d$ (deg cm³ g⁻¹ dm⁻¹) = +47.5 (*c* = 0.6 in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.88 – 7.08 (m, 18H), 5.40 (t, *J* = 9.5 Hz, 1H, H-3), 5.07 (t, *J* = 9.7 Hz, 1H, H-4), 4.70 (d, *J* = 9.3 Hz, 1H, SCH₂CHPh), 4.64 – 4.45 (m, 3H, H-1, CH₂Ph), 4.45 – 4.08 (m, 3H, CH Fmoc, CH₂ Fmoc), 3.98 – 3.83 (m, 1H, H-5), 3.83 – 3.61 (m, 3H, H-2, H-6_{a,b}), 2.98 (dd, *J* = 14.0, 10.7 Hz, 1H, SCHHCHPh), 2.81 (dd, *J* = 13.9, 1.7 Hz, 1H, SCHHCHPh), 1.95 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 171.85, 140.45, 137.87, 128.73, 128.70, 128.12, 128.07, 125.61, 81.28, 80.01, 79.73, 77.77, 77.35, 76.92, 76.03, 75.98, 74.02, 70.95, 70.06, 35.85, 21.29; HR MALDI-TOF MS: m/z: calcd for

$C_{38}H_{36}O_8S$ $[M+Na]^+$: 675.2029; found: 675.2017.

2-(S)-Phenyl-(3,6-di-O-acetyl-4-O-benzyl-1,2-dideoxy- β -D-glucopyranoso)[1,2-e]-1,4-oxathiane (13). A mixture of **4** (610 mg, 1.43 mmol) and activated molecular sieves (4Å) in DCM (8 mL) was stirred for 1 h under an atmosphere of argon. After cooling to -78 °C, triethylsilane (451 μ L, 2.86 mmol) and dichlorophenylborane (280 μ L, 2.15 mmol) were added. After 30 min, the reaction was quenched by the addition of MeOH (1 mL) and Et₃N (0.5 mL). The resulting mixture was diluted with DCM (10 mL), filtered and the filtrate was concentrated *in vacuo*. The residue was redissolved in pyridine (5 mL) and acetic anhydride (5 mL) was added. After stirring for 16 h, the mixture was concentrated *in vacuo* and the resulting residue was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/4 \rightarrow 1/3, v/v) to afford **13** (525 mg, 78%). R_f = 0.43 (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{26}^d$ (deg cm³ g⁻¹ dm⁻¹) = +80.0 (c = 0.3 in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.43 – 7.18 (m, 10H, ArH), 5.36 (t, J = 9.3 Hz, 1H), 4.73 – 4.53 (m, 3H, SCH₂CHPh, PHCH₂), 4.48 (d, J = 8.9 Hz, 1H, H-1), 4.38 (dd, J = 12.2, 1.8 Hz, 1H, H-6_a), 4.22 (dd, J = 12.1, 4.6 Hz, 1H, H-6_b), 3.81 – 3.54 (m, 3H, H-5, H-4, H-2), 2.95 (dd, J = 14.0, 10.5 Hz, 1H, SCHHCHPh), 2.80 (dd, J = 14.0, 2.1 Hz, 1H, SCHHCHPh), 2.08 (s, 3H), 1.98 (m, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.85, 170.20, 140.34, 137.44, 128.80, 128.69, 128.37, 128.27, 128.00, 125.46, 81.70, 79.62, 78.46, 77.70, 77.28, 76.86, 76.17, 75.80, 75.24, 74.91, 63.18, 35.80, 21.23, 21.12; HR MALDI-TOF MS: m/z : calcd for C₂₅H₂₈O₇S $[M+Na]^+$: 495.1454; found: 495.1447.

2-(S)-Phenyl-(4-O-acetyl-6-O-benzyl-3-O-levulinoyl-1,2-dideoxy- β -D-glucopyranoso)[1,2-e]-1,4-oxathiane (14). A mixture of **5** (300 mg, 0.62 mmol) and activated molecular sieves (4 Å) in DCM (5 mL) was stirred for 1 h under an atmosphere

of argon. After cooling to $-78\text{ }^{\circ}\text{C}$, triethylsilane (196 μL , 1.24 mmol) and trifluoromethanesulfonic acid (82 μL , 0.93 mmol) were added. After 1 h, the reaction was quenched by the addition of MeOH (1 mL) and Et₃N (0.5 mL). The resulting mixture was diluted with DCM (10 mL), filtered and the filtrate was concentrated *in vacuo*. The residue was redissolved in pyridine (5 mL) and acetic anhydride (5 mL) was added. After stirring for 16 h, the mixture was removed *in vacuo*. The resulting residue was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/4 \rightarrow 1/2, v/v) to afford **14** (259 mg, 79%). $R_f = 0.29$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{26}^{\text{d}}$ (deg cm³ g⁻¹ dm⁻¹) = +105.3 ($c = 1.9$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.43 – 7.18 (m, 10H, ArH), 5.23 (t, $J = 9.6$ Hz, 1H, H-3), 5.17 (t, $J = 9.6$ Hz, 1H, H-4), 4.67 (dd, $J = 10.5, 1.8$ Hz, 1H, SCH₂CHPh), 4.63 – 4.43 (m, 3H, CH₂Ph, H-1), 3.86 – 3.66 (m, 2H, H-2, H-5), 3.61 – 3.48 (m, 2H, H-6_{ab}), 2.96 (dd, $J = 14.0, 10.6$ Hz, 1H, SCHHCHPh), 2.79 (dd, $J = 14.0, 2.1$ Hz, 1H, SCHHCHPh), 2.76 – 2.31 (m, 4H, 2 \times CH₂ Lev), 2.07 (s, 3H), 1.99 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 206.19, 172.23, 170.06, 140.28, 137.86, 128.66, 128.60, 128.21, 128.06, 127.98, 125.60, 81.39, 79.74, 78.93, 77.69, 77.26, 76.84, 75.94, 73.85, 73.38, 69.20, 68.95, 38.10, 35.80, 29.80, 28.25, 20.92; HR MALDI-TOF MS: m/z : calcd for C₂₃H₂₅O₅S [M+Na]⁺: 436.1321; found: 436.1329.

2-(S)-Phenyl-(4-O-acetyl-3-O-allyloxycarbonyl-6-O-benzyl-1,2-dideoxy- β -D-glucopyranoso)[1,2-e]-1,4-oxathiane (15). A mixture of **6** (300 mg, 0.64 mmol) and activated molecular sieves (4 Å) in DCM (5 mL) was stirred for 1 h under an atmosphere of argon. After cooling to $-78\text{ }^{\circ}\text{C}$, triethylsilane (202 μL , 1.28 mmol) and trifluoromethanesulfonic acid (85 μL , 0.96 mmol) were added. After 1 h, the reaction was quenched by the addition of MeOH (1 mL) and Et₃N (0.5 mL), diluted by DCM (10 mL)

and filtered. The filtrate was concentrated *in vacuo*. The residue was redissolved in pyridine (5 mL) and acetic anhydride (5 mL) was added. After stirring for 16 h, the solvent was removed. The resulting residue was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/8 → 1/2, v/v) to afford **15** (302 mg, 92%). $R_f = 0.58$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +94.4 ($c = 3.6$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.42 – 7.17 (m, 10H, ArH), 5.79 (ddd, $J = 22.7, 10.8, 5.6$ Hz, 1H, CH alloc), 5.29 – 4.99 (m, 4H, H-3, H-4, CH₂ alloc), 4.69 (dd, $J = 10.5, 1.8$ Hz, 1H, SCH₂CHPh), 4.64 – 4.45 (m, 5H, CH₂Ph, CH₂ alloc, H-1), 3.89 – 3.71 (m, 2H, H-2, H-5), 3.61 – 3.51 (m, 2H, H-6_{a,b}), 2.99 (dd, $J = 14.0, 10.6$ Hz, 1H, SCHHCHPh), 2.80 (dd, $J = 14.0, 2.0$ Hz, 1H, SCHHCHPh), 1.93 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 169.69, 154.84, 140.19, 137.83, 131.45, 128.61, 128.60, 128.21, 128.10, 128.00, 125.77, 125.73, 118.89, 81.32, 79.79, 78.84, 77.70, 77.46, 77.28, 76.85, 75.80, 73.88, 69.45, 69.10, 68.84, 35.72, 20.90; HR MALDI-TOF MS: m/z : calcd for C₂₇H₃₀O₈S [M+Na]⁺: 537.1559; found: 537.1551.

2-(S)-Phenyl-(4,6-di-O-acetyl-3-O-benzyl-1,2-dideoxy-β-D-glucopyranoso)[1,2-e]-1,4-oxathiane (16). TsOH·H₂O (80 mg, 0.42 mmol) and EtSH (31 μL, 0.42 mmol) were added to a stirred solution of **7** (200 mg, 0.42 mmol) in DCM (5 mL). After stirring for 2 h, the solvent was removed *in vacuo* and the residue was redissolved in pyridine (5 mL) and acetic anhydride (5 mL). After stirring for 16 h, the reaction mixture was concentrated under reduced pressure. The resulting residue was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/5 → 1/4, v/v) to afford **16** (167 mg, 84%). $R_f = 0.21$ (EtOAc/hexanes, 1/4, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +80.0 ($c = 0.4$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.50 – 7.09 (m, 10H, ArH), 5.11 (t, $J = 9.7$ Hz,

1H, H-4), 4.80 (t, $J = 12.2$ Hz, 2H, *CHHPh*, *SCH₂CHPh*), 4.63 (d, $J = 12.0$ Hz, 1H, *CHHPh*), 4.44 (d, $J = 8.9$ Hz, 1H, H-1), 4.25 – 4.04 (m, 2H, H-6_{a,b}), 3.84 (t, $J = 9.0$ Hz, 1H, H-2), 3.75 – 3.55 (m, 2H, H-5, H-3), 3.05 (dd, $J = 13.9, 10.7$ Hz, 1H, *SCHHCHPh*), 2.81 (dd, $J = 13.9, 1.6$ Hz, 1H, *SCHHCHPh*), 2.08 (s, 3H), 1.96 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 171.00, 169.67, 140.51, 138.40, 128.74, 128.49, 128.28, 128.14, 127.87, 125.82, 84.59, 80.05, 79.98, 77.65, 77.55, 77.23, 76.80, 75.92, 74.87, 69.62, 62.68, 35.67, 21.03, 21.00; HR MALDI-TOF MS: m/z : calcd for C₂₅H₂₈O₇S [M+Na]⁺: 495.1454; found: 495.1463.

2-(*S*)-phenyl-(3,4-di-*O*-acetyl-6-benzyl-1,2-dideoxy-β-*D*-glucopyranoso)[1,2-*e*]-1,4-oxathiane (*R,S*)-*S*-oxide (17). Compound **17** (1.4 g, 92 %, *R/S* = 1.3/1) was prepared according to the general procedure for the preparation of sulfoxide donors starting from **9** (1.5 g, 3.2 mmol) and using *m*-CPBA (0.79 g, ≤77%, 3.5 mmol). **17S**: $R_f = 0.24$ (EtOAc/hexanes, 1/1, v/v); ¹H NMR (300 MHz, CDCl₃): δ 7.48 – 7.19 (m, 10H, *ArH*), 5.55 – 5.30 (m, 2H, H-3, *SCH₂CHPh*), 5.05 (t, $J = 9.8$ Hz, 1H, H-4), 4.59 – 4.45 (m, 3H, *CH₂Ph*, H-2), 4.14 (d, $J = 9.7$ Hz, 1H, H-1), 3.96 – 3.81 (m, 1H, H-5), 3.73 – 3.52 (m, 2H, H-6_{a,b}), 3.21 (dd, $J = 14.5, 1.6$ Hz, 1H, *SCH_{eq}HCHPh*), 2.71 (dd, $J = 14.4, 11.3$ Hz, 1H, *SCHH_{ax}CHPh*), 2.01 (s, 3H), 1.94 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.59, 169.72, 138.87, 137.72, 128.94, 128.65, 128.38, 128.22, 128.07, 125.70, 85.41, 79.27, 77.68, 77.26, 76.83, 73.98, 73.37, 69.40, 69.33, 69.16, 68.51, 53.01, 21.01, 20.82; HR MALDI-TOF MS: m/z : calcd for C₂₅H₂₈O₈S [M+Na]⁺: 511.1403; found: 511.1397. **17R**: $R_f = 0.18$ (EtOAc/hexanes, 1/1, v/v); ¹H NMR (300 MHz, CDCl₃): δ 7.48 – 7.19 (m, 10H, *ArH*), 5.47 – 5.20 (m, 2H, H-3, H-4), 4.70 (d, $J = 10.9$ Hz, 1H, *SCH₂CHPh*), 4.62 (d, $J = 12.0$ Hz, 1H, *CHHPh*), 4.48 (d, $J = 12.0$ Hz, 1H, *CHHPh*) 4.31 (d, $J = 10.0$ Hz, 1H, H-1), 3.84

– 3.54 (m, 5H, H-2, H-5, H-6_{a,b}, SCH_{eq}HCHPh), 3.05 (dd, $J = 12.7, 11.8$ Hz, 1H, SCHH_{ax}CHPh), 1.99 (s, 3H), 1.91 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.32, 169.60, 138.03, 137.69, 129.08, 128.94, 128.81, 128.62, 128.29, 128.21, 128.04, 125.69, 125.60, 95.26, 78.74, 77.65, 77.23, 76.81, 75.74, 75.45, 73.95, 73.47, 68.60, 67.92, 57.81, 20.96, 20.81; HR MALDI-TOF MS: m/z: calcd for C₂₅H₂₈O₈S [M+Na]⁺: 511.1403; found: 511.1404.

2-(S)-Phenyl-(3-O-acetyl-4,6-di-O-benzyl-1,2-dideoxy-β-D-glucopyranoso)[1,2-e]-1,4-oxathiane(R,S)-S-oxide (18). Compound **18** (0.17 g, 96 %, $R/S = 1.1/1$) was prepared according to the general procedure for the preparation of sulfoxide donors starting from **10** (0.17 g, 0.33 mmol) and using *m*-CPBA (0.82 g, ≤ 77%, 0.36 mmol). **18S**: $R_f = 0.24$ (EtOAc/hexanes, 1/1, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +48.0 ($c = 0.6$ in CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ 7.43 – 7.13 (m, 15H, ArH), 5.45 (t, $J = 9.4$ Hz, 1H, H-3), 5.35 (d, $J = 11.1$ Hz, 1H, SCH₂CHPh), 4.66 – 4.49 (m, 4H, 2×CH₂Ph), 4.42 (t, $J = 9.6$ Hz, 1H, H-2), 4.11 (d, $J = 9.6$ Hz, 1H, H-1), 3.80 – 3.64 (m, 4H, H-4, H-5, H-6_{a,b}), 3.20 (d, $J = 14.5$ Hz, 1H, SCH_{eq}HCHPh), 2.70 (dd, $J = 14.3, 11.4$ Hz, 1H, SCHH_{ax}CHPh), 1.96 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.11, 138.87, 137.81, 137.40, 128.69, 128.49, 128.43, 128.02, 128.00, 127.78, 125.38, 85.27, 80.83, 77.23, 77.01, 76.80, 75.47, 75.15, 74.81, 73.72, 69.53, 68.60, 68.00, 52.80, 20.98.; HR MALDI-TOF MS: m/z: calcd for C₃₀H₃₂O₇S [M+Na]⁺: 559.1767; found: 559.1779. **18R**: $R_f = 0.21$ (EtOAc/hexanes, 1/1, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +80.0 ($c = 0.5$ in CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ 7.45 – 7.12 (m, 15H, ArH), 5.41 (t, $J = 9.3$ Hz, 1H, H-3), 4.77 – 4.44 (m, 5H, 2×CH₂Ph, SCH₂CHPh), 4.28 (d, $J = 9.6$ Hz, 1H, H-1), 3.95 – 3.81 (m, 3H, H-4, H-6_{a,b}), 3.64 (dd, $J = 19.2, 11.4$ Hz, 2H, H-5, SCH_{eq}HCHPh), 3.52 (t, $J = 9.6$ Hz, 1H, H-2), 3.02

(t, $J = 12.2$ Hz, 1H, SCHH_{ax}CHPh), 1.90 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 169.72, 137.96, 137.59, 128.79, 128.76, 128.45, 128.44, 128.40, 128.29, 128.05, 128.01, 127.89, 127.86, 125.24, 125.21, 95.05, 80.39, 77.19, 76.98, 76.77, 75.73, 75.26, 74.92, 74.84, 74.73, 73.81, 67.65, 57.46, 20.85; HR MALDI-TOF MS: m/z: calcd for C₃₀H₃₂O₇S [M+Na]⁺: 559.1767; found: 559.1759.

2-(S)-Phenyl-{3-O-acetyl-6-O-benzyl-4-O-(9-fluorenylmethyloxycarbonyl)-1,2-dideoxy- β -D-glucopyranoso}[1,2-*e*]-1,4-oxathiane(*R,S*)-S-oxide (19). Compound **19** (1.9 g, 98 %, $R/S = 1/1$) was prepared according to the general procedure for the preparation of sulfoxide donors starting from **11** (1.9 g, 2.9 mmol) and using *m*-CPBA (712 mg, $\leq 77\%$, 3.2 mmol). **19S**: $R_f = 0.18$ (EtOAc/hexanes, 1/1, v/v); ¹H NMR (300 MHz, CDCl₃): δ 7.81 – 7.08 (m, 18H, ArH), 5.47 (t, $J = 9.5$ Hz, 1H, H-3), 5.30 (d, $J = 10.9$ Hz, 1H, SCH₂CHPh), 4.86 (t, $J = 9.7$ Hz, 1H, H-4), 4.60 – 4.38 (m, 3H, H-2, CH₂Ph), 4.36 – 4.02 (m, 4H, CH Fmoc, CH₂ Fmoc, H-1), 3.98 – 3.81 (m, 1H, H-5), 3.70 – 3.52 (m, 2H, H-6_{a,b}), 3.15 (d, $J = 14.1$ Hz, 1H, SCH_{eq}HCHPh), 2.64 (dd, $J = 14.4, 11.3$ Hz, 1H, SCHH_{ax}CHPh), 1.89 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.44, 154.34, 143.38, 143.24, 141.51, 141.49, 138.84, 137.77, 128.97, 128.63, 128.39, 128.24, 128.22, 128.05, 128.03, 127.52, 127.47, 125.69, 125.37, 125.31, 120.36, 120.33, 85.34, 78.88, 77.76, 77.33, 76.91, 74.00, 73.28, 73.11, 70.65, 69.42, 69.37, 68.50, 52.93, 46.77, 21.05; HR MALDI-TOF MS: m/z: calcd for C₃₈H₃₆O₉S [M+Na]⁺: 691.1978; found: 691.1971. **19R**: $R_f = 0.15$ (EtOAc/hexanes, 1/1, v/v); ¹H NMR (300 MHz, CDCl₃): δ 7.88 – 7.18 (m, 18H), 5.51 (t, $J = 9.4$ Hz, 1H, H-3), 5.16 (t, $J = 9.7$ Hz, 1H, H-4), 4.71 (d, $J = 11.0$ Hz, 1H, SCH₂CHPh), 4.55 (dd, $J = 28.1, 12.1$ Hz, 2H, CH₂Ph), 4.45 – 4.10 (m, 4H, H-1, CH Fmoc, CH₂ Fmoc), 3.96 – 3.86 (m, 1H, H-5), 3.86 – 3.55 (m, 4H, H-6_{a,b}, H-2,

SCH_{eq}HCHPh), 3.06 (dd, $J = 12.4, 12.0$ Hz, 1H, SCHH_{ax}CHPh), 1.94 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 169.89, 154.01, 143.15, 143.03, 141.27, 137.79, 137.52, 128.87, 128.59, 128.36, 127.99, 127.96, 127.84, 127.75, 127.29, 127.24, 125.34, 125.11, 120.10, 95.02, 78.25, 77.44, 77.22, 77.01, 76.59, 75.52, 75.29, 73.80, 72.93, 72.37, 70.45, 67.85, 57.63, 53.80, 46.51, 30.93, 29.71, 29.28, 20.74; HR MALDI-TOF MS: m/z: calcd for C₃₈H₃₆O₉S [M+Na]⁺: 691.1978; found: 691.1973.

2-(S)-phenyl-(3,6-di-O-acetyl-4-benzyl-2-dideoxy-β-D-glucopyranoso)[1,2-e]-1,4-oxathiane (R,S)-S-oxide (20). Compound **20** (245 mg, 93 %, $R/S = 1/1$) was prepared according to the general procedure for the preparation of sulfoxide donors starting from **13** (250 mg, 0.54 mmol) and using *m*-CPBA (133 mg, ≤ 77%, 0.60 mmol). **20S**: $R_f = 0.15$ (EtOAc/hexanes, 1/1, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +60.0 ($c = 0.5$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.45 – 7.20 (m, 10H, ArH), 5.49 (t, $J = 9.4$ Hz, 1H, H-3), 5.35 (d, $J = 10.5$ Hz, 1H, SCH₂CHPh), 4.65 (d, $J = 11.2$ Hz, 1H, CHHPh), 4.58 (d, $J = 11.2$ Hz, 1H, CHHPh), 4.47 – 4.37 (m, 2H, H-2, H-6_a), 4.21 (dd, $J = 12.2, 5.5$ Hz, 1H, H-6_b), 4.14 (d, $J = 9.7$ Hz, 1H, H-1), 3.82 (ddd, $J = 9.8, 5.5, 2.0$ Hz, 1H, H-5), 3.65 (t, $J = 9.5$ Hz, 1H, H-4), 3.22 (dd, $J = 14.5, 1.6$ Hz, 1H, SCH_{eq}HCHPh), 2.71 (dd, $J = 14.4, 11.3$ Hz, 1H, SCHH_{ax}CHPh), 2.07 (s, 3H), 2.00 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.82, 170.23, 138.94, 137.17, 128.95, 128.85, 128.48, 128.34, 128.28, 125.56, 85.36, 78.79, 77.67, 77.24, 76.82, 75.52, 75.36, 75.14, 69.67, 68.25, 63.12, 52.98, 21.21, 21.08; HR MALDI-TOF MS: m/z: calcd for C₂₅H₂₈O₈S [M+Na]⁺: 511.1403; found: 511.1408. **20R**: $R_f = 0.12$ (EtOAc/hexanes, 1/1, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +26.7 ($c = 0.4$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.52 – 7.10 (m, 10H, ArH), 5.48 (t, $J = 9.1$ Hz, 1H, H-3), 4.69 (d, $J = 11.3$ Hz, 1H, SCH₂CHPh), 4.60 (q, $J = 11.2$ Hz, 2H, CH₂Ph), 4.43 (dd, $J = 12.4, 1.9$ Hz,

1H, H-6_a), 4.31 (dd, $J = 13.0, 3.4$ Hz, 2H, H-6_b, H-1), 3.85 – 3.73 (m, 1H, H-5), 3.73 – 3.59 (m, 2H, H-4, SCH_{eq}HCHPh), 3.52 (t, $J = 9.7$ Hz, 1H, H-2), 3.12 – 2.96 (m, 1H, SCHH_{ax}CHPh), 2.08 (s, 3H), 1.97 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.73, 169.82, 138.08, 137.15, 129.08, 128.85, 128.72, 128.50, 128.29, 125.46, 94.99, 78.47, 77.67, 77.25, 76.82, 75.85, 75.62, 75.34, 75.11, 75.07, 62.67, 57.74, 21.13, 21.07; HR MALDI-TOF MS: m/z : calcd for C₂₅H₂₈O₈S [M+Na]⁺: 511.1403; found: 511.1411.

2-(S)-phenyl-(4-O-acetyl-6-benzyl-3-levulinoyl-1,2-dideoxy- β -D-glucopyranoso)[1,2-*e*]-1,4-oxathiane (R,S)-S-oxide (21). Compound **21** (0.41 g, 98 %, $R/S = 1.3/1$) was prepared according to the general procedure for the preparation of sulfoxide donors using *m*-CPBA (0.24 g, $\leq 77\%$, 1.1 mmol) and starting from **14** (0.4 g, 0.97 mmol). **21S**: $R_f = 0.19$ (EtOAc/hexanes, 1/1, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +87.5 ($c = 0.6$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.48 – 7.17 (m, 10H, ArH), 5.42 (t, $J = 9.6$ Hz, 1H, H-3), 5.36 (d, $J = 11.0$ Hz, 1H, SCH₂CHPh), 5.06 (t, $J = 9.8$ Hz, 1H, H-4), 4.60 – 4.41 (m, 3H, H-2, CH₂Ph), 4.14 (d, $J = 9.7$ Hz, 1H, H-1), 3.97 – 3.79 (m, 1H, H-5), 3.66 (dd, $J = 11.0, 6.0$ Hz, 1H, H-6_a), 3.57 (dd, $J = 11.0, 2.8$ Hz, 1H, H-6_b), 3.21 (d, $J = 13.5$ Hz, 1H, SCH_{eq}HCHPh), 2.83 – 2.36 (m, 5H, SCHH_{ax}CHPh, 2×CH₂ Lev), 2.08 (s, 3H), 2.01 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 206.04, 171.97, 169.81, 138.01, 137.72, 129.04, 128.77, 128.61, 128.28, 128.02, 125.63, 95.22, 78.85, 77.66, 77.24, 76.81, 75.68, 75.56, 73.96, 73.47, 68.19, 68.01, 57.73, 37.97, 29.79, 28.14, 20.86; HR MALDI-TOF MS: m/z : calcd for C₂₃H₂₅O₆S [M+Na]⁺: 452.1270; found: 452.1277. **21R**: $R_f = 0.18$ (EtOAc/hexanes, 1/1, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +56.0 ($c = 0.7$ in CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ 7.47 – 7.17 (m, 10H, ArH), 5.38 (t, $J = 9.4$ Hz, 1H, H-3), 5.29 (t, $J = 9.7$ Hz, 1H, H-4), 4.70 (d, $J = 11.2$ Hz, 1H, SCH₂CHPh), 4.61 (d, $J = 12.0$ Hz, 1H,

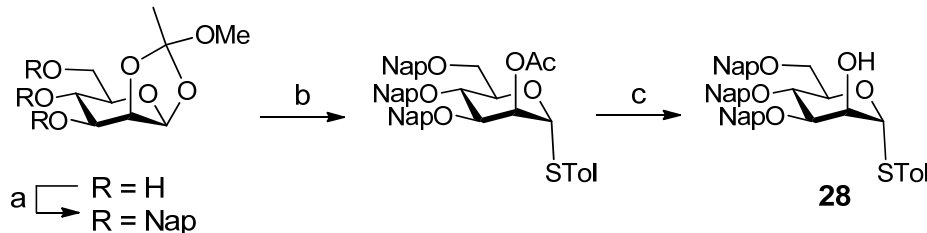
*CHHP*h), 4.49 (d, $J = 12.0$ Hz, 1H, *CHHP*h), 4.31 (d, $J = 10.0$ Hz, 1H, H-1), 3.83 – 3.75 (m, 1H, H-5), 3.72 (dd, $J = 11.1, 2.9$ Hz, 1H, H-6_a), 3.69 – 3.53 (m, 3H, H-6_b, H-2, *SCH*_{eq}*HCHPh*), 3.05 (dd, $J = 12.8, 11.8$ Hz, 1H, *SCHH*_{ax}*CHPh*), 2.78 – 2.37 (m, 4H, 2×*CH*₂ Lev), 2.08 (s, 3H), 1.98 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 206.12, 172.12, 170.01, 138.84, 137.74, 128.92, 128.64, 128.36, 128.21, 128.06, 125.71, 85.39, 79.41, 77.66, 77.44, 77.23, 76.81, 73.99, 73.35, 69.43, 68.71, 68.46, 52.98, 38.01, 29.79, 28.22, 20.87; HR MALDI-TOF MS: *m/z*: calcd for C₂₃H₂₅O₆S [M+Na]⁺: 452.1270; found: 452.1268.

2-(*S*)-phenyl-(3-allyloxycarbonyl-4-*O*-acetyl-6-*O*-benzyl-1,2-dideoxyl-β-*D*-glucopyranoso)[1,2-*e*]-1,4-oxathiane (*R,S*)-*S*-oxide (22). Compound **22** (0.42 g, 82 %, *R/S* = 1.5/1) was prepared according to the general procedure for the preparation of sulfoxide donors using *m*-CPBA (0.24 g, ≤ 77%, 0.97 mmol) and starting from **15** (0.50 g, 0.97 mmol). **22S**: $R_f = 0.33$ (EtOAc/hexanes, 1/1, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +78.0 ($c = 4.1$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.46 – 7.21 (m, 10H, *ArH*), 5.92 – 5.64 (m, 1H, CH alloc), 5.37 (d, $J = 10.0$ Hz, 1H, *SCH*₂*CHPh*), 5.30 – 5.17 (m, 2H, H-3, *CHH* alloc), 5.17 – 5.00 (m, 2H, H-4, *CHH* alloc), 4.63 – 4.48 (m, 5H, H-2, *CH*₂ alloc, *CH*₂*Ph*), 4.15 (d, $J = 9.7$ Hz, 1H, H-1), 3.97 – 3.81 (m, 1H, H-5), 3.73 – 3.47 (m, 2H, H-6_{a,b}), 3.21 (dd, $J = 14.5, 1.6$ Hz, 1H, *SCH*_{eq}*HCHPh*), 2.72 (dd, $J = 14.4, 11.3$ Hz, 1H, *SCHH*_{ax}*CHPh*), 1.95 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 169.60, 154.76, 138.75, 137.72, 131.35, 128.86, 128.65, 128.40, 128.22, 128.08, 125.86, 119.01, 85.23, 79.27, 77.73, 77.60, 77.31, 76.89, 73.99, 69.52, 69.40, 69.01, 68.95, 68.57, 52.87, 20.84.; HR MALDI-TOF MS: *m/z*: calcd for C₂₇H₃₀O₉S [M+Na]⁺: 553.1509; found: 553.1500. **22R**: $R_f = 0.31$ (EtOAc/hexanes, 1/1, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +81.3 ($c = 1.6$ in CHCl₃);

^1H NMR (300 MHz, CDCl_3): δ 7.49 – 7.19 (m, 10H, ArH), 5.90 – 5.68 (m, 1H, CH alloc), 5.42 – 5.05 (m, 4H, CH_2 alloc, H-3, H-4), 4.71 (d, $J = 11.0$ Hz, 1H, SCH_2CHPh), 4.66 – 4.43 (m, CH_2 alloc, CH_2Ph), 4.32 (d, $J = 10.0$ Hz, 1H, H-1), 3.87 – 3.49 (m, 4H, H-5, H-6, H-2, $\text{SCH}_{\text{eq}}\text{HCHPh}$), 3.07 (t, $J = 12.0$ Hz 1H, $\text{SCHH}_{\text{ax}}\text{CHPh}$), 1.93 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 169.45, 154.56, 137.93, 137.69, 131.29, 128.99, 128.83, 128.62, 128.28, 128.04, 125.76, 119.07, 95.07, 78.72, 77.69, 77.62, 77.26, 76.84, 75.81, 75.44, 73.97, 69.01, 68.48, 68.15, 57.71, 20.83; HR MALDI-TOF MS: m/z : calcd for $\text{C}_{27}\text{H}_{30}\text{O}_9\text{S}$ $[\text{M}+\text{Na}]^+$: 553.1509; found: 553.1514.

2-(S)-phenyl-(4,6-di-O-acetyl-3-O-benzyl-1,2-dideoxy- β -D-glucopyranoso)[1,2-*e*]-1,4-oxathiane (R,S)-S-oxide (23). Compound **23** (142 mg, 96 %, $R/S = 1.7/1$) was prepared as an inseparable mixture of diastereomers according to the general procedure for the preparation of sulfoxide donors using *m*-CPBA (75 mg, $\leq 77\%$, 0.33 mmol) and starting from **16** (140 mg, 0.30 mmol). $R_f = 0.27$ (EtOAc/hexanes, 2/1, v/v); ^1H NMR (600 MHz, CDCl_3): δ 7.51 – 7.12 (m, 20H, ArH), 5.43 (d, $J = 10.3$ Hz, 1H, $\text{SCH}_2\text{CHPh}^{\text{S}}$), 5.12 (t, $J = 12.0$ Hz, 1H, H-4^R), 5.08 (t, $J = 12.0$ Hz, 1H, H-4^S), 4.85 – 4.74 (m, 3H, $\text{SCH}_2\text{CHPh}^{\text{R}}$, $2 \times \text{CHHPh}$), 4.68 – 4.57 (m, 3H, H-2^S, $2 \times \text{CHHPh}$), 4.35 – 4.13 (m, 5H, H-1^R, $2 \times \text{H-6}_{\text{a,b}}$) 4.10 (d, $J = 9.7$ Hz, 1H, H-1^S), 3.85 – 3.62 (m, 6H, H-3^S, H-3^R, H-2^R, H-5^S, H-5^R, $\text{SCHHCHPh}^{\text{R}}$), 3.20 (dd, $J = 14.5, 1.5$ Hz, 1H, $\text{SCHHCHPh}^{\text{S}}$), 3.14 – 3.05 (m, 1H, $\text{SCHHCHPh}^{\text{R}}$), 2.79 (dd, $J = 14.4, 11.3$ Hz, 1H, $\text{SCHHCHPh}^{\text{S}}$), 2.13 – 2.02 ($2 \times \text{s}$, 6H), 1.99 – 1.86 ($2 \times \text{s}$, 6H); selected ^{13}C NMR (gHSQC, CDCl_3): δ 94.98 (C-1^R), 85.40 (C-1^S), 79.91, 78.33, 77.58, 75.90, 75.08, 74.42, 68.84, 68.41, 62.66, 61.84, 57.34, 52.51; HR MALDI-TOF MS: m/z : calcd for $\text{C}_{25}\text{H}_{28}\text{O}_8\text{S}$ $[\text{M}+\text{Na}]^+$: 511.1403; found: 511.1411.

Scheme S1. Preparation of 28



Reagents and conditions: (a) NaH, NapBr, DMF (95%); (b) HgBr (cat.), TolSH, DCE, 60 °C, 16 h (81%); (c) NaOMe, MeOH (quantitative).

***p*-toluene 3,4,6-tri-*O*-(2-naphthyl)- α -thiomannoside (**28**):** $R_f = 0.53$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +141.5 ($c = 1.6$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.91 – 6.89 (m, 25H, ArH), 5.58 (s, 1H, H-1), 5.00 (d, $J = 11.2$ Hz, 1H, NapCHH), 4.90 (d, $J = 11.7$ Hz, 1H, NapCHH), 4.84 (d, $J = 11.7$ Hz, 1H, NapCHH), 4.78 (d, $J = 12.2$ Hz, 1H, NapCHH), 4.67 (d, $J = 11.2$ Hz, 1H, NapCHH), 4.59 (d, $J = 12.2$ Hz, 1H, NapCHH), 4.37 (dd, $J = 9.1, 2.8$ Hz, 1H, H-5), 4.32 (s, 1H, H-2), 4.10 – 3.94 (m, 2H, H-3, H-4), 3.87 (dd, $J = 10.9, 4.5$ Hz, 1H, H-6_a), 3.75 (dd, $J = 10.8, 1.8$ Hz, 1H, H-6_b), 2.25 (s, 3H, Me); ¹³C NMR (75 MHz, CDCl₃) δ 137.90, 135.91, 135.87, 135.30, 133.49, 133.46, 133.43, 133.32, 133.21, 133.14, 132.44, 130.16, 130.04, 128.67, 128.32, 128.24, 128.21, 128.14, 127.95, 127.91, 127.84, 127.05, 126.91, 126.60, 126.47, 126.33, 126.27, 126.25, 126.19, 126.05, 126.03, 126.01, 87.96, 80.60, 77.68, 77.26, 76.83, 75.41, 74.84, 73.73, 72.51, 72.43, 70.11, 69.07, 21.29; HR MALDI-TOF MS: m/z : calcd for C₄₆H₄₂O₅S [M+Na]⁺: 729.2651; found: 729.2638.

Methyl **3,4-di-*O*-acetyl-6-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-2,3,4-tri-*O*-benzoyl- α -**

D-glucopyranoside (29). Compound **29** was prepared according to the general glycosylation procedure using glycosyl donor **17** (45 mg, 0.09 mmol) and glycosyl acceptor **24** (39 mg, 0.08 mmol). Purification by LH20 size exclusion chromatography afforded compound **29** (80 mg, 91%). $R_f = 0.45$ (acetone/toluene, 1/9, v/v); $[\alpha]_{27}^d$ (deg $\text{cm}^3 \text{g}^{-1} \text{dm}^{-1}$) = +86.7 ($c = 3.0$ in CHCl_3); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 8.10 – 6.98 (m, 25H, *ArH*), 6.18 (t, $J = 9$ Hz, 1H, H-3^I), 6.04 (s, 2H, *ArH*), 5.51 (d, $J = 3.3$ Hz, 1H, H-1^{II}), 5.44 (t, $J = 9$ Hz, 1H, H-4^I), 5.38 (t, $J = 9$ Hz, 1H, H-3^{II}), 5.30 – 5.16 (m, 2H, H-1^I, H-2^I), 4.99 (t, $J = 9.7$ Hz, 1H, H-4^{II}), 4.66 – 4.36 (m, 4H, CH_2Ph , H-5^I, SCH_2CHPh), 4.30 – 4.08 (m, 2H, H-5^{II}), 4.03 – 3.91 (m, 2H, H-6^I_{a,b}), 3.81 (s, 3H, OMe), 3.74 (s, 6H, 2 × OMe), 3.66 – 3.37 (m, 6H, OMe, H-2^{II}, H-6^{II}_{a,b}), 2.99 – 2.71 (m, 2H, SCH_2CHPh), 1.83 (s, 3H), 1.40 (s, 3H); $^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 170.26, 169.79, 165.89, 165.67, 165.54, 161.68, 161.60, 142.02, 137.88, 133.29, 133.14, 132.97, 129.95, 129.79, 129.64, 129.34, 129.15, 128.74, 128.39, 128.32, 128.22, 128.15, 127.94, 127.61, 127.41, 126.20, 101.82, 97.36, 96.50, 90.89, 84.33, 78.82, 77.44, 77.22, 77.02, 76.60, 73.39, 72.42, 72.04, 70.67, 69.86, 69.47, 68.83, 68.30, 67.98, 67.08, 55.89, 55.47, 55.34, 43.33, 20.68, 20.32; HR MALDI-TOF MS: m/z : calcd for $\text{C}_{62}\text{H}_{64}\text{O}_{19}\text{S}$ $[\text{M}+\text{Na}]^+$: 1167.3660; found: 1167.3655.

Methyl 3,4-di-O-acetyl-6-O-benzyl-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1→4)-2,3-di-O-acetyl-O-benzyl- α -D-glucopyranoside (30). Compound **30** was prepared according to the general procedure using glycosyl donor **17** (50 mg, 0.10 mmol) and glycosyl acceptor **25** (31 mg, 0.083 mmol). Purification by LH20 size exclusion chromatography afforded compound **30** (52 mg, 62%). $R_f = 0.34$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{27}^d$ (deg $\text{cm}^3 \text{g}^{-1} \text{dm}^{-1}$) = +75.0 ($c = 1.6$ in CHCl_3); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.55 – 6.87 (m, 15H, *ArH*), 6.12 (s,

2H, ArH), 5.77 (d, $J = 3.2$ Hz, 1H, H-1^{II}), 5.72 (t, $J = 9.2$ Hz, 1H, H-3^I), 5.23 (t, $J = 9.8$ Hz, 1H, H-3^{II}), 5.04 – 4.79 (m, 3H, H-1^I, H-2^I, H-4^{II}), 4.61 (s, 2H, CH₂Ph), 4.48 (d, $J = 11.8$ Hz, 1H, CHHPh), 4.33 (d, $J = 11.8$ Hz, 1H, CHHPh), 4.27 – 4.12 (m, 2H, H-4^I, SCH₂CHPh), 4.12 – 3.90 (m, 3H, H-5^I, H-5^{II}, H-6^I_a), 3.84 (s, 3H, OMe), 3.79 (s, 7H, 2 × OMe, H-6^I_b), 3.66 (dd, $J = 10.2, 3.2$ Hz, 1H, H-2^{II}), 3.41 (s, 3H, OMe), 3.39 – 3.29 (m, 2H, H-6^{II}_a, H-6^{II}_b), 3.02 (dd, $J = 13.9, 3.8$ Hz, 1H, SCHHCHPh), 2.77 (dd, $J = 14.0, 8.7$ Hz, 1H, SCHHCHPh), 2.08 (s, 3H), 2.06 (s, 3H), 1.84 (s, 3H), 1.26 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.70, 170.30, 170.16, 170.08, 162.11, 162.01, 142.30, 138.46, 138.07, 128.53, 128.50, 128.39, 128.22, 128.06, 127.83, 127.64, 127.53, 127.49, 126.22, 100.99, 96.97, 96.90, 91.16, 84.30, 79.90, 77.70, 77.48, 77.27, 76.85, 74.29, 73.61, 73.58, 72.33, 72.16, 71.17, 69.97, 69.72, 69.25, 68.94, 68.43, 56.21, 55.59, 55.49, 42.99, 21.67, 21.11, 20.94, 20.31; HR MALDI-TOF MS: m/z: calcd for C₅₂H₆₂O₁₈S [M+Na]⁺: 1029.3555; found: 1029.3541.

N^α-(9-Fluorenylmethyloxycarbonyl)-O-{3,4-di-O-acetyl-6-O-benzyl-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]-α-D-glucopyranosyl}-L-threonine benzyl ester (31). Compound **31** was prepared according to the general glycosylation procedure using glycosyl donor **17** (35 mg, 0.070 mmol) and glycosyl acceptor **26** (26 mg, 0.060 mmol). Purification by LH20 size exclusion chromatography afforded compound **31** (57 mg, 89%). $R_f = 0.23$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +53.3 ($c = 1.9$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.82 – 6.97 (m, 23H, ArH), 6.25 (d, $J = 7.7$ Hz, 1H, NHFmoc), 6.15 (s, 2H, ArH), 5.70 (d, $J = 3.5$ Hz, 1H, H-1), 5.34 (t, $J = 9.8$ Hz, 1H, H-3), 5.27 (s, 2H, CH₂Ph), 4.95 (t, $J = 9.8$ Hz, 1H, H-4), 4.58 – 4.17 (m, 8H, CH₂Ph, 2 × CH^{Thr}, CH₂^{Fmoc}, CH^{Fmoc}, SCH₂CHPh), 4.07 (d, $J = 10.1$ Hz, 1H, H-5), 3.85 –

3.81 (2s, 10H, 3 × OMe, H-2), 3.64 – 3.39 (m, 2H, H-6_a, H-6_b), 3.09 (dd, $J = 14.0, 2.6$ Hz, 1H, SCHHCHPh), 2.86 (dd, $J = 14.0, 9.6$ Hz, 1H, SCHHCHPh), 1.87 (s, 3H), 1.40 (d, $J = 6.4$ Hz, 3H, CH₃^{Thr}), 1.18 (d, $J = 13.2$ Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.25, 170.07, 169.87, 161.95, 161.79, 156.78, 144.05, 143.91, 142.38, 141.19, 141.15, 137.84, 135.49, 128.50, 128.41, 128.31, 128.23, 127.80, 127.65, 127.55, 127.18, 125.66, 125.41, 125.33, 119.81, 100.97, 98.93, 91.01, 84.72, 80.40, 77.44, 77.22, 77.02, 76.59, 75.93, 73.45, 72.67, 69.59, 69.08, 68.27, 67.61, 67.41, 59.44, 55.96, 55.37, 47.09, 43.20, 30.13, 20.69, 19.93, 19.35; HR MALDI-TOF MS: m/z: calcd for C₆₀H₆₃NO₁₅S [M+Na]⁺: 1092.3816; found: 1092.3808.

***p*-Methylphenyl 3,4-di-*O*-acetyl-6-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 4)- 2,3-di-*O*-acetyl-6-*O*-benzyl-1-thio- β -D-glucopyranoside (32).** Compound **32** was prepared according to the general glycosylation procedure using glycosyl donor **17** (45 mg, 0.090 mmol) and glycosyl acceptor **27** (35 mg, 0.080 mmol). Purification by LH20 size exclusion chromatography afforded compound **32** (63 mg, 75%). $R_f = 0.25$ (acetone/toluene, 1/9, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +68.6 ($c = 0.9$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.51 – 6.96 (m, 19H, ArH), 6.12 (s, 2H, ArH), 5.64 (d, $J = 3.2$ Hz, 1H, H-1^{II}), 5.43 (t, $J = 9.0$ Hz, 1H, H-3^I), 5.18 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 4.92 (t, $J = 9$ Hz, 2H, H-2^I, H-4^{II}), 4.68 (d, $J = 10.0$ Hz, 1H, H-1^I), 4.56 (s, 2H, CH₂Ph), 4.45 (d, $J = 11.9$ Hz, 1H, CHHPh), 4.30 (d, $J = 11.9$ Hz, 1H, CHHPh), 4.19 (dd, $J = 8.3, 4.2$ Hz, 1H, SCH₂CHPh), 4.10 (t, $J = 9.4$ Hz, 1H, H-4^I), 4.03 – 3.87 (m, 2H, H-5^I, H-5^{II}), 3.82 (s, 3H, OMe), 3.78 (s, 6H, 2 × OMe), 3.74 – 3.56 (m, 3H, H-2^{II}, H-6^I_a, H-6^I_b), 3.32 (dd, $J = 10.7, 2.4$ Hz, 1H, H-6^{II}_a), 3.25 (dd, $J = 10.7, 3.4$ Hz, 1H, H-6^{II}_b), 2.99 (dd, $J = 13.9, 4.1$ Hz, 1H, SCHHCHPh), 2.82 (dd, $J =$

13.9, 8.3 Hz, 1H, SCHHCHPh), 2.31 (s, 3H, CH₃), 2.09 (s, 3H), 2.04 (s, 3H), 1.82 (s, 3H), 1.32 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.16, 169.99, 169.76, 169.67, 161.91, 161.78, 141.96, 138.42, 138.31, 137.81, 133.57, 129.65, 128.27, 128.23, 128.13, 128.02, 127.86, 127.60, 127.44, 127.39, 127.31, 126.10, 116.18, 100.98, 96.79, 91.03, 85.36, 83.75, 79.16, 79.11, 77.44, 77.22, 77.02, 76.59, 75.26, 73.39, 73.34, 73.18, 72.05, 71.12, 69.34, 69.06, 68.93, 68.05, 56.02, 55.37, 42.76, 30.15, 21.40, 21.16, 20.89, 20.67, 20.14.; HR MALDI-TOF MS: m/z: calcd for C₅₈H₆₆O₁₇S₂ [M+Na]⁺: 1121.3639; found: 1121.3630.

***p*-Methylphenyl 3,4-di-*O*-acetyl-6-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 2)-3,4,6-tri-*O*-(2-ethylnaphthyl)-1-thio- α -D-mannopyranoside (33).** Compound **33** was prepared according to the general glycosylation procedure using glycosyl donor **17** (56 mg, 0.11 mmol) and glycosyl acceptor **28** (68 mg, 0.10 mmol). Purification by LH20 size exclusion chromatography afforded compound **33** (93 mg, 72%). $R_f = 0.23$ (EtOAc/hexanes, 1/3, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +100 ($c = 0.5$ in CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.09 – 6.83 (m, 35H, ArH), 6.05 (s, 2H, ArH), 5.88 (d, $J = 3.5$ Hz, 1H, H-1^{II}), 5.83 (s, 1H, H-1^I), 5.47 (t, $J = 9.6$ Hz, 1H, H-3^{II}), 5.22 – 5.06 (m, 2H, CHHNap, CHHNap), 4.96 (t, $J = 9.8$ Hz, 1H, H-4^{II}), 4.82 (d, $J = 11.8$ Hz, 1H, CHHNap), 4.72 (dd, $J = 16.6, 11.8$ Hz, 2H, CHHNap, CHHNap), 4.67 – 4.55 (m, 2H, H-2^I, CHHNap), 4.47 (d, $J = 12.2$ Hz, 1H, CHHPh), 4.42 – 4.26 (m, 4H, CHHPh, SCH₂CHPh^{II}, H-5^I, H-5^{II}), 4.23 (t, $J = 9.2$ Hz, 1H, H-4^I), 4.12 (d, $J = 8.6$ Hz, 1H, H-3^I), 3.91 – 3.64 (m, 12H, H-2^{II}, H-6^I_a, H-6^I_b, 3 × OMe), 3.48 – 3.33 (m, 2H, H-6^{II}_a, H-6^{II}_b), 3.01 (dd, $J = 13.8, 4.0$ Hz, 1H, SCHHCHPh^{II}), 2.76 (dd, $J = 13.7, 8.7$ Hz, 1H, SCHHCHPh^{II}), 2.23 (s, 3H),

1.69 (s, 3H), 1.31 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 170.44, 170.17, 161.97, 161.88, 142.31, 138.19, 137.63, 136.29, 136.26, 136.20, 133.59, 133.49, 133.20, 133.16, 133.10, 132.77, 132.37, 130.94, 130.01, 129.95, 128.54, 128.48, 128.36, 128.28, 128.23, 128.21, 128.13, 128.09, 128.04, 127.87, 127.76, 127.74, 127.45, 126.80, 126.77, 126.59, 126.44, 126.36, 126.20, 126.13, 126.05, 125.98, 125.89, 125.86, 125.82, 101.69, 99.23, 91.20, 87.52, 84.36, 79.77, 79.44, 77.68, 77.50, 77.25, 76.83, 75.33, 73.38, 72.92, 72.88, 71.24, 69.81, 69.58, 69.03, 68.22, 56.17, 55.55, 43.45, 30.35, 21.27, 20.81, 20.38; HR MALDI-TOF MS: m/z : calcd for $\text{C}_{80}\text{H}_{80}\text{O}_{15}\text{S}_2$ $[\text{M}+\text{Na}]^+$: 1367.4836; found: 1367.4829.

3,4,6-tri-*O*-acetyl-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose (38).

Compound **38** was prepared according to the general glycosylation procedure using glycosyl donor **34** (55 mg, 0.13 mmol) and glycosyl acceptor **36** (27 mg, 0.10 mmol). Purification by LH20 size exclusion chromatography afforded compound **38** (71 mg, 80%). $R_f = 0.14$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{27}^{\text{D}}$ (deg $\text{cm}^3 \text{g}^{-1} \text{dm}^{-1}$) = +36.4 ($c = 2.2$ in CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ 7.41 – 7.01 (m, 5H, ArH), 6.14 (s, 2H, ArH), 5.58 – 5.47 (m, 2H, H-1^I, H-1^{II}), 5.33 (t, $J = 9.6$ Hz, 1H, H-3^{II}), 4.94 – 4.76 (m, 1H, H-4^{II}), 4.64 (dd, $J = 7.9, 2.3$ Hz, 1H, H-3^I), 4.45 – 3.94 (m, 8H, H-4^I, H-2^I, SCH_2CHPh , H-5^I, H-5^{II}, H-6_{ab}^{II}, H-6_a^I), 3.94 – 3.74 (m, 10H, 3 \times OMe, H-6_b^I), 3.62 (dd, $J = 10.0, 3.6$ Hz, 1H, H-2^{II}), 3.01 (dd, $J = 13.8, 4.4$ Hz, 1H, SCHHCHPh), 2.90 (dd, $J = 13.8, 8.2$ Hz, 1H, SCHHCHPh), 2.05 (s, 3H), 1.95 (s, 3H), 1.60 (s, 3H), 1.45 (s, 3H), 1.38 (s, 3H), 1.35 (s, 6H); ^{13}C NMR (75 MHz, CDCl_3): δ 170.94, 170.31, 170.13, 161.96, 161.89, 142.09, 128.34, 127.67, 126.49, 109.48, 108.92, 101.77, 98.63, 96.51, 91.21, 84.38, 79.21, 77.68, 77.25, 76.83, 72.28, 71.33, 70.91, 70.88, 69.15, 68.98, 67.38, 67.22, 62.46, 56.16, 55.60,

42.91, 26.41, 26.24, 25.25, 24.71, 20.98, 20.88, 20.44; HR MALDI-TOF MS: m/z : calcd for $C_{46}H_{58}O_{16}S$ $[M+Na]^+$: 873.2979; found: 873.2991.

Methyl 3,4,6-tri-*O*-acetyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-2,3,4-tri-*O*-benzoyl- α -D-glucopyranoside (39).

Compound **39** was prepared according to the general glycosylation procedure using glycosyl donor **34** (50 mg, 0.11 mmol) and glycosyl acceptor **24** (48 mg, 0.095 mmol). Purification by LH20 size exclusion chromatography afforded compound **39** (62 mg, 59%). $R_f = 0.12$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{27}^d$ (deg $cm^3 g^{-1} dm^{-1}$) = +50.0 ($c = 1.0$ in $CHCl_3$); 1H NMR (500 MHz, $CDCl_3$): δ 8.09 – 6.97 (m, 20H), 6.19 (t, $J = 9.3$ Hz, 1H, H-2^I), 6.07 (s, 2H, ArH), 5.56 (d, $J = 3.1$ Hz, 1H, H-1^{II}), 5.45 (t, $J = 9.9$ Hz, 1H, H-3^I), 5.40 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 5.32 – 5.20 (m, 2H, H-1^I, H-2^I), 4.83 (t, $J = 9.7$ Hz, 1H, H-4^{II}), 4.57 – 4.46 (m, 1H, H-5^I), 4.36 – 4.16 (m, 3H, SCH_2CHPh , H-5^{II}, H-6_a^{II}), 4.08 (d, $J = 12.0$ Hz, 1H, H-6_b^{II}), 4.01 – 3.90 (m, 2H, H-6_{a,b}^I), 3.84 (s, 3H, OMe), 3.77 (s, 6H, 2 \times OMe), 3.64 – 3.51 (m, 4H, OMe, H-2^{II}), 2.96 – 2.79 (m, 2H, SCH_2CHPh), 2.09 (s, 3H), 1.95 (s, 3H), 1.38 (s, 3H); ^{13}C NMR (150 MHz, $CDCl_3$): δ 184.56, 184.53, 184.51, 184.50, 184.48, 175.63, 175.04, 174.88, 170.81, 170.62, 170.51, 166.61, 166.58, 146.93, 138.25, 138.14, 137.94, 134.89, 134.74, 134.58, 134.22, 134.05, 133.61, 133.34, 133.20, 133.17, 133.14, 132.41, 131.07, 109.95, 106.66, 102.28, 101.97, 101.47, 95.84, 89.59, 84.03, 82.95, 82.94, 82.16, 81.95, 81.74, 77.28, 76.64, 75.53, 74.76, 73.97, 73.74, 72.21, 72.17, 67.23, 60.81, 60.43, 60.31, 48.31, 34.64, 25.74, 25.60, 25.17; HR MALDI-TOF MS: m/z : calcd for $C_{57}H_{60}O_{20}S$ $[M+Na]^+$: 1119.3296; found: 1119.3288.

Methyl 3,4,6-tri-*O*-acetyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- α -D-glucopyranoside (40).

Compound **40** was prepared according to the general glycosylation procedure using glycosyl donor **34** (68 mg, 0.15 mmol) and glycosyl acceptor **37** (60 mg, 0.13 mmol). Purification by LH20 size exclusion chromatography afforded compound **40** (65 mg, 48%). $R_f = 0.32$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +35.0 ($c = 0.3$ in CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.47 – 6.80 (m, 20H, ArH), 6.19 – 6.05 (m, 3H, ArH, H-1^{II}), 5.35 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 5.06 (s, 2H, CH₂Ph), 4.80 (t, $J = 9.8$ Hz, 1H, H-4^{II}), 4.70 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.64 (d, $J = 3.5$ Hz, 1H, H-1^I), 4.63 – 4.54 (m, 3H, CH₂Ph, CHHPh), 4.29 – 4.08 (m, 4H, H-3^I, H-4^I, H-5^{II}, SCHHCHPh), 4.03 (dd, $J = 10.7, 3.5$ Hz, 1H, H-6^I_a), 3.98 (dd, $J = 12.2, 3.3$ Hz, 2H, H-5^I, H-6^{II}_a), 3.90 – 3.71 (m, 11H, 3×OMe, H-6^I_b, H-6^{II}_b), 3.67 – 3.53 (m, 2H, H-2^I, H-2^{II}), 3.40 (s, 3H, OMe), 2.90 (dd, $J = 13.9, 4.0$ Hz, 1H, SCHHCHPh), 2.77 (dd, $J = 13.9, 8.7$ Hz, 1H, SCHHCHPh), 1.96 (s, 3H), 1.91 (s, 3H), 1.24 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.38, 170.13, 169.79, 161.89, 161.77, 142.08, 139.55, 138.21, 138.16, 128.40, 128.27, 128.12, 128.10, 127.84, 127.40, 127.23, 126.86, 126.74, 126.08, 101.15, 97.77, 95.91, 90.98, 84.39, 80.60, 80.10, 79.78, 77.68, 77.23, 77.02, 76.80, 74.18, 73.87, 73.47, 73.27, 71.91, 69.54, 69.21, 69.12, 67.55, 62.04, 55.85, 55.35, 42.77, 20.67, 20.64, 19.97; HR MALDI-TOF MS: m/z : calcd for C₅₇H₆₆O₁₇S [M+Na]⁺: 1077.3918; found: 1077.3926.

3,6-di-O-acetyl-4-O-benzyl-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1→6)-1,2:3,4-di-O-isopropylidene- α -D-galactopyranose (41). Compound **41** was prepared according to the general glycosylation procedure using glycosyl donor **20** (60 mg, 0.12 mmol) and glycosyl acceptor **36** (27 mg, 0.10 mmol). Purification by LH20 size exclusion chromatography afforded compound **41** (79 mg, 84%). $R_f = 0.25$ (EtOAc/hexanes, 1/2, v/v); ¹H NMR (300 MHz, CDCl₃): δ 7.34 – 7.06

3.62 – 3.54 (m, 1H, H-2^I), 3.40 (dd, $J = 10.1, 3.4$ Hz, 1H, H-2^{II}), 3.38 (s, 3H, OMe), 3.36 – 3.31 (m, 1H, H-4^{II}), 2.94 (dd, $J = 13.7, 7.4$ Hz, 1H, SCHHCHPh), 2.87 (dd, $J = 13.6, 5.8$ Hz, 1H, SCHHCHPh), 1.95 (s, 3H), 1.41 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 170.38, 169.74, 161.78, 161.60, 141.50, 139.59, 138.19, 138.13, 137.54, 128.44, 128.38, 128.17, 128.13, 128.09, 128.06, 128.01, 127.99, 127.93, 127.85, 127.81, 127.76, 127.43, 127.30, 127.28, 126.86, 126.80, 126.68, 101.35, 97.77, 95.86, 90.83, 83.63, 80.83, 80.34, 79.03, 77.23, 77.02, 76.80, 76.42, 74.06, 74.04, 73.31, 73.29, 73.15, 69.56, 69.03, 68.69, 62.87, 55.81, 55.31, 55.23, 42.10, 20.82, 20.56; HR MALDI-TOF MS: m/z : calcd for C₆₂H₇₀O₁₆S [M+Na]⁺: 1125.4282; found: 1125.4275.

3,4-di-*O*-acetyl-6-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose (43). Compound **43** was prepared according to the general glycosylation procedure using glycosyl donor **17** (70 mg, 0.14 mmol) and glycosyl acceptor **36** (31 mg, 0.12 mmol). Purification by LH20 size exclusion chromatography afforded compound **43** (101 mg, 94%). $R_f = 0.33$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{27}^d$ (deg cm³ g⁻¹ dm⁻¹) = +62.5 ($c = 1.6$ in CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.47 – 7.01 (m, 10H, ArH), 6.21 – 6.01 (m, 2H, ArH), 5.52 (d, $J = 4.7$ Hz, 1H, H-1^I), 5.45 (d, $J = 3$ Hz, 1H, H-2^{II}), 5.31 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 4.98 (t, $J = 9.7$ Hz, 1H, H-4^{II}), 4.63 (dd, $J = 7.9, 2.2$ Hz, 1H, H-3^I), 4.57 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.45 – 4.38 (m, 2H, CHHPh, H-4^I), 4.31 (dd, $J = 4.9, 2.3$ Hz, 1H, H-2^I), 4.27 (dd, $J = 7.9, 4.9$ Hz, 1H, SCH₂CHPh), 4.19 – 4.00 (m, 2H, H-5^I, H-5^{II}), 3.99 – 3.86 (m, 2H, H-6^{II}_a, H-6^{II}_b), 3.83 (s, 3H, OMe), 3.81 (s, 6H, 2 \times OMe), 3.63 (dd, $J = 10.1, 3.5$ Hz, 1H, H-2^{II}), 3.56 (dd, $J = 10.7, 2.5$ Hz, 1H, H-6^I_a), 3.46 (dd, $J = 10.8, 3.6$ Hz, 1H, H-6^I_b), 3.02 (dd, $J = 13.7, 4.8$ Hz, 1H, SCHHCHPh), 2.91 (dd, $J = 13.8, 7.9$ Hz, 1H,

SCHHCHPh), 1.84 (s, 3H), 1.60 (s, 3H), 1.48 – 1.21 (4s, 12H); ^{13}C NMR (75 MHz, CDCl_3): δ 170.29, 169.83, 161.73, 161.62, 141.85, 137.93, 128.27, 128.05, 127.91, 127.56, 127.40, 126.37, 116.19, 109.20, 108.67, 101.53, 98.34, 96.30, 90.94, 83.76, 78.64, 77.45, 77.23, 77.03, 76.60, 73.38, 72.39, 70.97, 70.80, 70.62, 69.52, 68.22, 68.08, 67.99, 66.83, 55.96, 55.36, 42.47, 30.16, 26.24, 26.05, 25.04, 24.53, 20.70, 20.31; HR MALDI-TOF MS: m/z : calcd for $\text{C}_{46}\text{H}_{58}\text{O}_{16}\text{S}$ $[\text{M}+\text{Na}]^+$: 921.3343; found: 921.3351.

Methyl 3,4-di-O-acetyl-6-O-benzyl-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-glucopyranoside (44). Compound **44** was prepared according to the general glycosylation procedure using glycosyl donor **17** (45 mg, 0.09 mmol) and glycosyl acceptor **37** (36 mg, 0.08 mmol). Purification by LH20 size exclusion chromatography afforded compound **44** (57 mg, 67%). $R_f = 0.43$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^{\text{d}}$ (deg $\text{cm}^3 \text{g}^{-1} \text{dm}^{-1}$) = +50.0 ($c = 0.8$ in CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ 7.53 – 6.78 (m, 25H, ArH), 6.07 (s, 3H, ArH, H-1^{II}), 5.33 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 5.05 (s, 2H, CH_2Ph), 4.96 (t, $J = 9.8$ Hz, 1H, H-4^{II}), 4.78 – 4.49 (m, 5H, $2 \times \text{CH}_2\text{Ph}$, H-1^I), 4.41 – 4.32 (m, 2H, CH_2Ph), 4.22 – 4.11 (m, 3H, SCH_2CHPh , H-3^I, H-4^I), 4.10 – 3.85 (m, 3H, H-5^I, H-5^{II}, H-6^I_a), 3.80 (s, 1H, OMe), 3.76 – 3.69 (m, 7H, $2 \times \text{OMe}$, H-6^I_b), 3.64 (m, 2H, H-2^I, H-2^{II}), 3.39 (s, 3H, OMe), 3.28 (d, $J = 10.6$ Hz, 1H, H-6^{II}_a), 3.12 (d, $J = 10.9$ Hz, 1H, H-6^{II}_b), 2.89 (dd, $J = 13.9, 4.3$ Hz, 1H, SCHHCHPh), 2.78 (dd, $J = 13.9, 8.2$ Hz, 1H, SCHHCHPh), 1.81 (s, 3H), 1.27 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3): δ 170.52, 169.94, 162.08, 161.91, 142.24, 139.73, 138.55, 138.40, 138.23, 128.62, 128.43, 128.40, 128.35, 128.31, 128.28, 128.03, 127.70, 127.49, 127.43, 127.36, 127.09, 126.37, 101.28, 97.97, 96.08, 91.09, 84.18, 80.95, 80.49, 79.57, 77.66, 77.43, 77.23, 76.81, 74.26, 73.91, 73.52,

72.46, 69.81, 69.69, 69.51, 68.95, 68.15, 56.10, 55.53, 42.84, 30.38, 20.91, 20.31; HR MALDI-TOF MS: m/z : calcd for $C_{62}H_{70}O_{16}S$ $[M+Na]^+$: 1125.4282; found: 1125.4292.

3-*O*-allyloxycarbonyl-4-*O*-acetyl-6-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose (45). Compound **45** was prepared according to the general glycosylation procedure using glycosyl donor **22** (76 mg, 0.14 mmol) and glycosyl acceptor **36** (31 mg, 0.12 mmol). Purification by LH20 size exclusion chromatography afforded compound **45** (76 mg, 67%). $R_f = 0.3$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +70.0 ($c = 0.3$ in CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ 7.43 – 7.06 (m, 10H, ArH), 6.10 (s, 2H, ArH), 5.77 – 5.61 (m, 1H, CH alloc), 5.52 (d, $J = 5.0$ Hz, 1H, H-1^I), 5.46 (d, $J = 3.5$ Hz, 1H, H-1^{II}), 5.26 – 5.09 (m, 2H, CH₂ alloc), 5.04 (t, $J = 9.7$ Hz, 1H, H-4^{II}), 4.64 (dd, $J = 7.9, 2.2$ Hz, 1H, H-3^I), 4.54 (d, $J = 11.8$ Hz, 1H, CHHPh), 4.50 – 4.39 (m, 2H, CHHPh, H-4^I), 4.36 – 4.29 (m, 2H, H-2^I, SCH₂CH), 4.27 (dd, $J = 13.1, 5.8$ Hz, 1H, CHH alloc), 4.09 (t, $J = 6.6$ Hz, 1H, H-5^I), 4.08 – 3.99 (m, 2H, H-5^{II}, CHH alloc), 3.98 – 3.85 (m, 2H, H-6^I_a, H-6^I_b), 3.83 – 3.80 (2s, 9H, 3 \times OMe), 3.73 (dd, $J = 9.9, 3.5$ Hz, 1H, H-2^{II}), 3.57 (dd, $J = 10.8, 2.7$ Hz, 1H, H-6^{II}_a), 3.47 (dd, $J = 10.8, 3.8$ Hz, 1H, H-6^{II}_b), 3.02 (dd, $J = 13.9, 4.3$ Hz, 1H, SCHHCH), 2.85 (dd, $J = 13.9, 8.2$ Hz, 1H, SCHHCH), 1.86 (s, 3H), 1.60 (s, 3H), 1.45 (s, 3H), 1.36 (s, 3H), 1.35 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 169.61, 161.74, 161.66, 154.09, 141.63, 137.94, 131.61, 128.26, 127.95, 127.92, 127.54, 127.21, 126.23, 118.19, 109.20, 108.66, 101.31, 98.48, 96.30, 95.09, 90.95, 83.31, 78.35, 78.09, 78.07, 77.22, 77.01, 76.80, 73.40, 70.92, 70.77, 70.61, 69.36, 68.30, 68.13, 67.82, 66.75, 55.93, 55.36, 42.75, 26.24, 26.05, 25.03, 24.56, 20.70; HR MALDI-TOF MS: m/z : calcd for $C_{48}H_{60}O_{17}S$ $[M+Na]^+$: 963.3449; found:

963.3456.

Methyl 3-*O*-allyloxycarbonyl-4-*O*-acetyl-6-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-2,3,4-tri-*O*-benzoyl- α -D-glucopyranoside (46). Compound **46** was prepared according to the general glycosylation procedure using glycosyl donor **22** (50 mg, 0.094 mmol) and glycosyl acceptor **24** (40 mg, 0.079 mmol). Purification by LH20 size exclusion chromatography afforded compound **46** (73 mg, 78%). $R_f = 0.24$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg $\text{cm}^3 \text{g}^{-1} \text{dm}^{-1}$) = +60.0 ($c = 0.3$ in CHCl_3); $^1\text{H NMR}$ (500 MHz, CDCl_3): δ 8.07 – 6.97 (m, 25H, ArH), 6.18 (t, $J = 9.4$ Hz, 1H, H-3^I), 6.04 (s, 2H, ArH), 5.77 – 5.60 (m, 1H, CH alloc), 5.53 – 5.42 (m, 2H, H-1^{II}, H-4^I), 5.31 – 5.22 (m, 2H, H-1^I, H-2^I), 5.22 – 5.10 (m, 3H, H-3^{II}, CH₂ alloc), 5.04 (dd, $J = 18.4, 8.9$ Hz, 1H, H-4^{II}), 4.59 – 4.39 (m, 3H, CH₂Ph, H-5^I), 4.31 – 4.23 (m, 2H, SCH₂CHPh, CHH alloc), 4.19 – 4.11 (m, 1H, H-5^{II}), 4.07 (dd, $J = 13.2, 5.5$ Hz, 1H, CHH alloc), 3.98 (dd, $J = 10.6, 8.3$ Hz, 1H, H-6_a^I), 3.91 (dd, $J = 10.6, 1.7$ Hz, 1H, H-6_b^I), 3.82 (s, 3H, OMe), 3.77 – 3.65 (m, 7H, 2 \times OMe, H-2^{II}), 3.61 – 3.51 (m, 4H, OMe, H-6_a^{II}), 3.45 (dd, $J = 10.8, 3.7$ Hz, 1H, H-6_b^{II}), 2.87 (ddd, $J = 22.8, 14.1, 6.2$ Hz, 2H, SCH₂CHPh), 1.85 (s, 3H); $^{13}\text{C NMR}$ (150 MHz, CDCl_3): δ 179.57, 179.54, 179.51, 169.54, 165.85, 165.67, 165.48, 161.65, 161.60, 154.03, 141.67, 137.87, 133.28, 133.17, 132.97, 131.56, 129.93, 129.80, 129.63, 129.29, 129.12, 128.74, 128.37, 128.28, 128.24, 128.21, 127.99, 127.90, 127.56, 127.18, 126.03, 118.15, 97.36, 96.49, 90.87, 83.84, 78.47, 77.97, 77.20, 76.98, 76.77, 73.37, 72.37, 70.62, 69.77, 69.29, 68.75, 68.33, 68.15, 68.11, 66.93, 55.86, 55.43, 55.32, 43.43, 20.66; HR MALDI-TOF MS: m/z : calcd for $\text{C}_{64}\text{H}_{66}\text{O}_{20}\text{S}$ $[\text{M}+\text{Na}]^+$: 1209.3766; found: 1209.3752.

4-*O*-acetyl-3-*O*-levulinoyl-6-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-

trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-1,2:3,4-di-O-isopropylidene- α -D-galactopyranose (47). Compound **47** was prepared according to the general glycosylation procedure using glycosyl donor **21** (40 mg, 0.074 mmol) and glycosyl acceptor **36** (16 mg, 0.062 mmol). Purification by LH20 size exclusion chromatography afforded compound **47** (51 mg, 85%). $R_f = 0.14$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_D^{26}$ (deg cm³ g⁻¹ dm⁻¹) = +27.5 ($c = 0.5$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.38 – 7.00 (m, 10H, ArH), 6.10 (s, 2H, ArH), 5.52 (d, $J = 5.0$ Hz, 1H, H-1^I), 5.45 (d, $J = 3.4$ Hz, 1H, H-1^{II}), 5.34 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 4.98 (t, $J = 9.8$ Hz, 1H, H-4^{II}), 4.62 (dd, $J = 7.9, 2.2$ Hz, 1H, H-3^I), 4.59 – 4.36 (m, 3H, CH₂Ph, H-4^I), 4.31 (dd, $J = 4.9, 2.3$ Hz, 1H, H-2^I), 4.25 (dd, $J = 7.7, 4.8$ Hz, 1H, SCH₂CHPh), 4.17 – 3.97 (m, 2H, H-5^I, H-5^{II}), 3.97 – 3.72 (m, 11H, H-6^I_a, H-6^I_b, 3 \times OMe), 3.63 (dd, $J = 10.0, 3.5$ Hz, 1H, H-2^{II}), 3.59 – 3.36 (m, 2H, H-6^{II}_a, H-6^{II}_b), 3.02 (dd, $J = 13.7, 4.9$ Hz, 1H, SCHHCHPh), 2.90 (dd, $J = 13.7, 7.8$ Hz, 1H, SCHHCHPh), 2.44 – 2.08 (m, 3H, 3 \times CHH Lev), 2.06 (s, 3H), 1.88 (s, 3H), 1.68 – 1.62 (m, 1H, CHH Lev), 1.59 (s, 3H), 1.45 (s, 3H), 1.34 (s, 6H); ¹³C NMR (75 MHz, CDCl₃): δ 206.39, 172.00, 170.15, 161.94, 161.86, 142.07, 138.15, 128.48, 128.22, 128.15, 127.87, 127.77, 127.52, 126.66, 109.41, 108.87, 101.61, 98.58, 96.51, 91.16, 83.84, 78.85, 77.66, 77.44, 77.23, 76.81, 73.62, 72.57, 71.15, 71.00, 70.83, 69.43, 68.57, 68.37, 68.08, 66.96, 56.18, 55.58, 42.60, 37.83, 29.94, 27.75, 26.46, 26.26, 25.24, 24.76, 20.92; HR MALDI-TOF MS: m/z : calcd for C₄₉H₆₂O₁₇S [M+Na]⁺: 977.3605; found: 977.3597.

Methyl 4-O-acetyl-6-O-benzyl-3-O-levulinoyl-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-glucopyranoside (48). Compound **48** was prepared according to the general

glycosylation procedure using glycosyl donor **21** (50 mg, 0.093 mmol) and glycosyl acceptor **37** (36 mg, 0.078 mmol). Purification by LH20 size exclusion chromatography afforded compound **48** (57 mg, 64%). $R_f = 0.11$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_D^{29}$ (deg $\text{cm}^3 \text{g}^{-1} \text{dm}^{-1}$) = +40.0 ($c = 0.8$ in CHCl_3); ^1H NMR (500 MHz, CDCl_3): δ 7.46 – 6.83 (m, 20H, *ArH*), 6.07 (s, 3H, *ArH*, H-1^{II}), 5.38 (t, $J = 9.8$ Hz, 1H, H-3^{II}), 5.04 (s, 2H, CH_2Ph), 4.95 (dd, $J = 19.0, 9.3$ Hz, 1H, H-4^{II}), 4.70 (d, $J = 12.0$ Hz, 1H, *CHHPh*), 4.63 (d, $J = 3.5$ Hz, 1H, H-1^I), 4.60 – 4.52 (m, 3H, *CHHPh*, CH_2Ph), 4.38 (d, $J = 11.9$ Hz, 1H, *CHHPh*), 4.27 (d, $J = 11.9$ Hz, 1H, *CHHPh*), 4.19 – 4.10 (m, 3H, H-3^I, H-4^I, SCH_2CHPh), 4.07 – 3.90 (m, 3H, H-5^I, H-5^{II}, H-6_a^{II}), 3.85 – 3.68 (m, 10H, 3×OMe, H-6_b^{II}), 3.65 – 3.56 (m, 1H, H-2^I), 3.39 (s, 3H, OMe), 3.29 (dd, $J = 10.7, 2.5$ Hz, 1H, H-6_a^{II}), 3.13 (dd, $J = 10.8, 3.4$ Hz, 1H, H-6_b^{II}), 2.87 (dd, $J = 13.9, 4.6$ Hz, 1H, *SCHHCHPh*), 2.78 (dd, $J = 13.9, 8.2$ Hz, 1H, *SCHHCHPh*), 2.42 – 2.30 (m, 1H, *CHH Lev*), 2.21 – 2.09 (m, 1H, *CHH Lev*), 2.06 (s, 3H), 2.04 – 1.95 (m, 1H, *CHH Lev*), 1.87 (s, 3H), 1.41 (dt, $J = 17.4, 6.1$ Hz, 1H, *CHH Lev*); ^{13}C NMR (75 MHz, CDCl_3): δ 206.36, 172.06, 170.05, 162.07, 161.93, 142.22, 139.74, 138.55, 138.42, 138.24, 128.62, 128.43, 128.39, 128.35, 128.29, 128.23, 128.05, 127.70, 127.47, 127.37, 127.31, 127.16, 127.07, 126.49, 101.20, 97.99, 96.27, 91.10, 84.05, 80.87, 80.70, 79.55, 77.67, 77.45, 77.24, 76.82, 74.35, 74.00, 73.52, 72.35, 69.76, 69.48, 69.11, 68.28, 56.10, 55.52, 42.75, 37.83, 29.95, 27.54, 20.92; HR MALDI-TOF MS: m/z : calcd for $\text{C}_{65}\text{H}_{74}\text{O}_{17}\text{S}$ $[\text{M}+\text{Na}]^+$: 1181.4545; found: 1181.4556.

4,6-di-O-acetyl-3-O-benzyl-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-1,2:3,4-di-O-isopropylidene- α -D-galactopyranose (49). Compound **49** was prepared according to the general glycosylation procedure using glycosyl donor **23** (60 mg, 0.12 mmol) and glycosyl acceptor **36** (26 mg, 0.10 mmol).

Purification by LH20 size exclusion chromatography afforded compound **49** (88 mg, 98%). $R_f = 0.19$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +13.5 ($c = 0.3$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.25 – 7.05 (m, 10H, ArH), 6.09 (s, 2H, ArH), 5.54 (d, $J = 5.0$ Hz, 1H, H-1^I), 5.21 (d, $J = 3.4$ Hz, 1H, H-1^{II}), 4.89 (t, $J = 9.9$ Hz, 1H, H-4^{II}), 4.67 (dd, $J = 7.9, 2.3$ Hz, 1H, H-3^I), 4.60 – 4.54 (m, 2H, CHHPh, SCH₂CHPh), 4.48 – 4.41 (m, 2H, CHHPh, H-4^I), 4.34 (dd, $J = 5.0, 2.4$ Hz, 1H, H-2^I), 4.22 (dd, $J = 12.3, 4.4$ Hz, 1H, H-6^{II}_a), 4.10 (br t, $J = 5.8$ Hz, 1H, H-5^I), 4.05 – 3.71 (m, 14H, H-5^{II}, H-6^{II}_b, H-6^I_a, H-6^I_b, H-3^{II}, 3 × OMe), 3.58 (dd, $J = 9.5, 3.5$ Hz, 1H, H-2^{II}), 3.12 (dd, $J = 13.3, 7.0$ Hz, 1H, SCHHCHPh), 3.01 (dd, $J = 13.3, 5.6$ Hz, 1H, SCHHCHPh), 2.04 (s, 3H), 1.86 (s, 3H), 1.59 (s, 3H), 1.47 (s, 3H), 1.39 (s, 3H), 1.35 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 171.08, 169.87, 161.83, 161.79, 141.09, 138.99, 128.36, 128.30, 127.91, 127.85, 127.45, 127.16, 109.53, 108.83, 102.16, 98.24, 96.55, 91.20, 81.73, 79.05, 78.58, 77.66, 77.44, 77.24, 76.81, 75.17, 71.36, 70.92, 70.88, 70.04, 68.28, 67.76, 67.09, 62.61, 56.17, 55.57, 42.37, 30.37, 26.40, 26.31, 25.24, 24.84, 21.02, 20.99; HR MALDI-TOF MS: m/z : calcd for C₄₆H₅₈O₁₆S [M+Na]⁺: 921.3344; found: 921.3352.

Methyl **4,6-di-O-acetyl-3-O-benzyl-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]-D-glucopyranosyl-(1→4)-2,3,6-tri-O-benzyl-D-glucopyranose (50)**. Compound **50** was prepared according to the general glycosylation procedure using glycosyl donor **23** (30 mg, 0.074 mmol) and glycosyl acceptor **37** (24 mg, 0.063 mmol). Purification by LH20 size exclusion chromatography afforded compound **50** (25 mg, 46%, $\alpha/\beta=7/1$). **50 α** : $R_f = 0.23$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +20.0 ($c = 0.5$ in CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.50 – 6.92 (m, 25H, ArH), 6.06 (s, 2H, ArH), 5.86 (d, $J = 3.4$ Hz, 1H, H-1^{II}), 5.09 (d, $J = 11.9$ Hz, 1H,

CHHPh), 5.02 (d, $J = 13.5$ Hz, 1H, *CHHPh*), 4.83 (t, $J = 10.0$ Hz, 1H, H-4^{II}), 4.69 (d, $J = 12.0$ Hz, 1H, *CHHPh*), 4.65 (d, $J = 3.5$ Hz, 1H, H-1^I), 4.62 – 4.52 (m, 3H, *CHHPh*, *CH₂Ph*), 4.48 (t, $J = 6.4$ Hz, 1H, *SCH₂CHPh*), 4.42 (d, $J = 11.6$ Hz, 1H, *CHHPh*), 4.34 (d, $J = 11.6$ Hz, 1H, *CHHPh*), 4.17 (t, $J = 9.1$ Hz, 1H, H-3^I), 4.04 (t, $J = 9.2$ Hz, 1H, H-4^I), 3.99 (dd, $J = 12.3, 4.1$ Hz, 1H, H-6^I_a), 3.96 – 3.83 (m, 3H, H-5^I, H-5^{II}, H-6^{II}_a), 3.83 – 3.77 (m, 4H, OMe, H-6^I_b), 3.77 – 3.67 (m, 7H, 2×OMe, H-6^{II}_b), 3.61 (dd, $J = 9.4, 3.5$ Hz, 1H, H-2^I), 3.50 (dd, $J = 9.6, 3.4$ Hz, 1H, H-2^{II}), 3.40 (s, 3H, OMe), 3.06 (dd, $J = 13.3, 6.5$ Hz, 1H, *SCHHCHPh*), 2.87 (dd, $J = 13.3, 6.4$ Hz, 1H, *SCHHCHPh*), 1.98 (s, 3H), 1.83 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 170.62, 169.48, 161.58, 161.55, 140.85, 139.13, 138.55, 138.03, 138.01, 128.59, 128.40, 128.31, 128.25, 128.12, 128.06, 127.99, 127.87, 127.53, 127.45, 127.32, 127.21, 127.09, 126.93, 126.83, 101.36, 97.72, 95.95, 90.88, 81.80, 81.24, 80.36, 78.62, 78.14, 78.09, 77.21, 77.00, 76.78, 74.76, 74.06, 73.40, 73.24, 72.79, 70.02, 69.54, 69.12, 68.10, 62.33, 55.80, 55.27, 55.24, 41.56, 20.77, 20.74.; HR MALDI-TOF MS: m/z: calcd for C₆₂H₇₀O₁₆S [M+Na]⁺: 1125.4283; found: 1125.4276.

The β anomer was purified by reversed phase HPLC on an analytical C-18 column using a gradient of 40→100% acetonitrile in H₂O over 40 min. **50β**: ¹H NMR (500 MHz, CDCl₃): δ 7.46 – 7.08 (m, 25H, *ArH*), 6.06 (s, 2H, *ArH*), 4.99 (t, $J = 6.7$ Hz, 1H, *SCH₂CHPh*), 4.90 (d, $J = 11.2$ Hz, 1H, *CHHPh*), 4.84 – 4.68 (m, 3H, H-4^{II}, 2×*CHHPh*), 4.68 – 4.54 (m, 4H, H-1^I, 2×*CHHPh*, *CHHPh*), 4.54 – 4.40 (m, 2H, 2×*CHHPh*), 4.32 (d, $J = 7.2$ Hz, 1H, H-1^{II}), 4.22 (d, $J = 8.2$ Hz, 1H, H-6^I_a), 4.07 – 3.96 (m, 2H, H-4^I, H-6^{II}_a), 3.89 – 3.81 (m, 3H, H-6^{II}_b, H-3^I, H-5^I), 3.78 (s, 9H, 3×OMe), 3.71 (d, $J = 11.0$ Hz, 1H, H-6^I_b), 3.51 (dd, $J = 9.6, 3.8$ Hz, 1H, H-2^I), 3.42 (s, 3H, OMe), 3.30 (dd, $J = 13.2, 6.3$ Hz, 1H, *SCHHCHPh*), 3.26 – 3.15 (m, 2H, H-2^{II}, H-3^{II}), 3.11 (d, $J = 8.5$ Hz, 1H, H-5^{II}), 2.82

(dd, $J = 13.2, 6.8$ Hz, 1H, SCHHCHPh), 1.92 (s, 3H), 1.85 (s, 3H); selected ^{13}C NMR (gHSQC, CDCl_3): δ 102.01 (C-1^H), 98.90 (C-1^I), 82.61, 82.16, 81.94, 81.66, 80.63, 78.75, 78.02, 75.76, 75.26, 73.72, 70.31, 69.87, 68.27, 67.85, 62.24, 55.92, 55.64, 41.89; HR MALDI-TOF MS: m/z : calcd for $\text{C}_{62}\text{H}_{70}\text{O}_{16}\text{S}$ $[\text{M}+\text{Na}]^+$: 1125.4283; found: 1125.4272.

3-*O*-acetyl-4,6-di-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose (51). Compound **51** was prepared according to the general procedure using glycosyl donor **18** (40 mg, 0.075 mmol) and glycosyl acceptor **36** (16 mg, 0.062 mmol). Purification by LH20 size exclusion chromatography afforded compound **51** (56 mg, 95 %). $R_f = 0.34$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^{\text{d}}$ (deg $\text{cm}^3 \text{g}^{-1} \text{dm}^{-1}$) = +21.5 ($c = 0.5$ in CHCl_3); ^1H NMR (600 MHz, CDCl_3): δ 7.42 – 7.01 (m, 15H, ArH), 6.06 (s, 2H, ArH), 5.50 (d, $J = 5.0$ Hz, 1H, H-1^I), 5.39 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 5.32 (d, $J = 3.5$ Hz, 1H, H-1^{II}), 4.61 (dd, $J = 7.4, 2.9$ Hz, 2H, H-3^I, CHHPh), 4.46 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.39 (s, 3H, CH_2Ph , H-4^I), 4.29 (dd, $J = 5.0, 2.3$ Hz, 1H, H-2^I), 4.25 (t, $J = 6.6$ Hz, 1H, SCH_2CHPh), 4.10 – 4.00 (m, 1H, H-6^a^I), 3.97 – 3.92 (m, 1H, H-5^{II}), 3.89 (dd, $J = 9.9, 7.2$ Hz, 1H, H-6^b^I), 3.86 – 3.71 (m, 11H, 3 \times OMe, H-5^I, H-6^a^{II}), 3.66 (dd, $J = 10.8, 1.8$ Hz, 1H, H-6^b^{II}), 3.57 (t, $J = 9.7$ Hz, 1H, H-4^{II}), 3.45 (dd, $J = 10.0, 3.5$ Hz, 1H, H-2^{II}), 3.02 (d, $J = 6.6$ Hz, 2H, SCH_2CHPh), 1.55 (s, 3H), 1.44 (2s, 6H), 1.34 (s, 3H), 1.32 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3): δ 179.61, 179.58, 179.57, 169.89, 161.60, 161.47, 141.44, 138.04, 137.98, 128.32, 128.25, 127.95, 127.90, 127.75, 127.66, 127.54, 127.45, 126.79, 109.09, 108.63, 101.58, 98.11, 96.24, 90.81, 83.16, 78.39, 78.03, 78.02, 77.20, 76.99, 76.78, 76.23, 73.72, 73.44, 73.18, 70.79, 70.77, 70.53, 69.66, 68.32, 67.64, 66.57, 55.91, 55.32, 41.63, 26.21, 26.00, 24.96, 24.45, 20.70; HR MALDI-TOF MS: m/z : calcd for

C₅₁H₆₂O₁₅S [M+Na]⁺: 969.3707; found: 969.3718.

Methyl **3-O-acetyl-4,6-di-O-benzyl-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]-D-glucopyranosyl-(1→4)- 2,3,6-tri-O-benzyl-D-glucopyranoside (52)**. Compound **52** was prepared according to the general glycosylation procedure using glycosyl donor **18** (50 mg, 0.093 mmol) and glycosyl acceptor **37** (36 mg, 0.078 mmol). Purification by LH20 size exclusion chromatography afforded compound **52** (59 mg, 66%, $\alpha/\beta=10/1$). **52a**: $R_f = 0.39$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +48.0 ($c = 0.6$ in CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ 7.48 – 6.90 (m, 30H, ArH), 6.03 (s, 2H, ArH), 5.95 (d, $J = 3.4$ Hz, 1H, H-1^{II}), 5.39 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 5.03 (d, $J = 11.8$ Hz, 1H, CHHPh), 4.94 (d, $J = 11.8$ Hz, 1H, CHHPh), 4.71 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.63 (d, $J = 3.5$ Hz, 1H, H-1^I), 4.58 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.50 (q, $J = 12.2$ Hz, 2H, CH₂Ph), 4.43 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.36 (d, $J = 11.2$ Hz, 1H, CHHPh), 4.31 (d, $J = 11.2$ Hz, 1H, CHHPh), 4.27 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.21 (t, $J = 6.7$ Hz, 1H, SCH₂CHPh), 4.15 – 4.08 (m, 2H, H-4^I, H-3^I), 3.96 (dd, $J = 10.7, 3.4$ Hz, 1H, H-6^{aI}), 3.94 – 3.90 (m, 1H, H-5^I), 3.81 (d, $J = 10.0$ Hz, 1H, H-5^{II}), 3.76 (s, 3H, OMe), 3.71 (s, 7H, 2×OMe, H-6^{bI}), 3.60 (dd, $J = 9.2, 3.5$ Hz, 1H, H-2^I), 3.52 (t, $J = 9.6$ Hz, 1H, H-4^{II}), 3.43 (dd, $J = 10.2, 3.4$ Hz, 1H, H-2^{II}), 3.37 (s, 3H, OMe), 3.33 (d, $J = 1.9$ Hz, 2H, H-6^{a, bII}), 1.40 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.09, 161.98, 161.78, 161.66, 141.61, 139.65, 138.56, 138.44, 138.30, 128.79, 128.63, 128.52, 128.46, 128.37, 128.35, 128.33, 128.17, 128.06, 127.93, 127.85, 127.75, 127.64, 127.38, 127.36, 127.28, 127.14, 127.00, 101.59, 97.96, 95.91, 91.04, 83.32, 81.17, 80.89, 78.80, 77.68, 77.46, 77.26, 76.83, 76.54, 74.36, 73.54, 73.51, 73.35, 72.86, 70.63, 69.70, 69.37, 68.32, 56.23, 56.09, 55.48, 55.43, 42.10, 20.85; HR MALDI-TOF MS: m/z: calcd for

$C_{67}H_{74}O_{15}S$ $[M+Na]^+$: 1173.4646; found:1173.4638. The β anomer was purified by reversed phase HPLC on an analytical C-18 column using a gradient of 50 \rightarrow 100% acetonitrile in H_2O over 40 min. **52B**: 1H NMR (600 MHz, $CDCl_3$): δ 7.48 – 7.03 (m, 30H, ArH), 6.01 (s, 2H, ArH), 5.00 (d, $J = 11.4$ Hz, 1H, CHHPh), 4.96 (t, $J = 9.4$ Hz, 1H, H-3^{II}), 4.79 (d, $J = 12.2$ Hz, 1H, CHHPh), 4.77 – 4.74 (m, 1H, SCH_2CHPh), 4.71 (d, $J = 11.4$ Hz, 1H, CHHPh), 4.66 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.63 – 4.58 (m, 2H, H-1^I, CHHPh), 4.49 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.45 – 4.33 (m, 5H, $2\times CH_2Ph$, H-1^{II}), 4.09 – 4.02 (m, 1H, H-4^I), 3.96 (dd, $J = 11.0, 2.6$ Hz, 1H, H-6_a^I), 3.87 (t, $J = 9.4$ Hz, 1H, H-3^I), 3.80 – 3.73 (m, 10H, H-5^I, $3\times OMe$), 3.63 (dd, $J = 11.2, 1.7$ Hz, 1H, H-6_a^{II}), 3.61 (dd, $J = 11.0, 1.7$ Hz, 1H, H-6_b^I), 3.50 – 3.43 (m, 2H, H-2^I, H-6_b^{II}), 3.43 – 3.35 (m, 4H, H-4^{II}, OMe), 3.21 – 3.09 (m, 3H, H-5^{II}, H-2^{II}, SCHHCHPh), 2.93 (dd, $J = 13.3, 8.6$ Hz, 1H, SCHHCHPh), 1.70 (s, 3H); ^{13}C NMR (gHSQC, 150 MHz, $CDCl_3$): δ ; HR MALDI-TOF MS: m/z : calcd for $C_{67}H_{74}O_{15}S$ $[M+Na]^+$: 1173.4646; found:1173.4641.

3,4,6-tri-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 6)-1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose (53).

Compound **53** was prepared according to the general glycosylation procedure using glycosyl donor **35** (50 mg, 0.086 mmol) and glycosyl acceptor **36** (18 mg, 0.070 mmol). Purification by LH20 size exclusion chromatography afforded compound **53** (68 mg, 98%). $R_f = 0.39$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg $cm^3 g^{-1} dm^{-1}$) = +6.7 ($c = 1.5$ in $CHCl_3$); 1H NMR (300 MHz, $CDCl_3$): δ 7.49 – 6.98 (m, 20H, ArH), 6.06 (s, 2H, ArH), 5.53 (d, $J = 5.0$ Hz, 1H, H-1^I), 5.10 (d, $J = 3.5$ Hz, 1H, H-1^{II}), 4.86 – 4.50 (m, 7H, CH_2Ph , CHHPh, CHHPh, SCH_2CH , H-4^I, H-3^I), 4.49 – 4.37 (m, 2H, CHHPh, CHHPh), 4.32 (dd, $J = 5.0, 2.3$ Hz, 1H, H-2^I), 4.18 – 3.98 (m, 1H, H-5^I), 3.98 – 3.68 (m, 15H, $3\times OMe$, H-3^{II},

H-5^{II}, H-6^I_a, H-6^I_b, H-6^{II}_a, H-6^{II}_b), 3.62 (dd, $J = 10.6, 1.9$ Hz, 1H, H-4^{II}), 3.57 – 3.44 (m, 1H, H-2^{II}) 3.19 (dd, $J = 13.1, 6.4$ Hz, 1H, SCHHCH), 3.02 (dd, $J = 13.1, 6.5$ Hz, 1H, SCHHCH), 1.56 (s, 3H), 1.47 (s, 3H), 1.41 (s, 3H), 1.33 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 161.76, 161.64, 140.94, 139.37, 138.71, 138.25, 128.53, 128.48, 128.30, 128.15, 128.02, 127.88, 127.82, 127.73, 127.44, 127.40, 116.41, 109.39, 108.81, 102.33, 98.07, 96.54, 91.16, 81.79, 80.92, 78.21, 77.80, 77.68, 77.46, 77.25, 76.83, 75.49, 75.08, 73.65, 71.10, 71.01, 70.84, 70.37, 68.72, 67.27, 66.48, 56.15, 55.53, 41.84, 30.38, 26.46, 26.35, 25.21, 24.91; HR MALDI-TOF MS: m/z : calcd for C₅₆H₆₆O₁₄S [M+Na]⁺: 1017.4071; found: 1017.4063.

Methyl 3,4,6-tri-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]-*D*-glucopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-*D*-glucopyranoside (54).

Compound **54** was prepared according to the general glycosylation procedure using glycosyl donor **35** (50 mg, 0.086 mmol) and glycosyl acceptor **37** (33 mg, 0.071 mmol). Purification by LH20 size exclusion chromatography afforded compound **54** (49 mg, 57%, $\alpha/\beta=1.5/1$). **54a**: $R_f = 0.43$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +21.4 ($c = 1.4$ in CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ 7.45 – 7.00 (m, 35H, ArH), 6.01 (s, 2H, ArH), 5.81 (d, $J = 3.3$ Hz, 1H, H-1^{II}), 5.09 (d, $J = 11.8$ Hz, 1H, CHHPh), 4.98 (d, $J = 11.8$ Hz, 1H, CHHPh), 4.75 – 4.55 (m, 8H, H-1^I, CHHPh, CH₂Ph×3), 4.53 – 4.47 (m, 2H, SCH₂CHPh, CHHPh), 4.45 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.36 (d, $J = 11.0$ Hz, 1H, CHHPh), 4.26 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.15 (t, $J = 9.1$ Hz, 1H, H-3^I), 4.07 (t, $J = 9.2$ Hz, 1H, H-4^I), 3.92 – 3.64 (m, 14H, H-5^I, H-3^{II}, H-5^{II}, H-6_{a,b}^I, 3×OMe), 3.61 (dd, $J = 9.4, 3.5$ Hz, 1H, H-2^I), 3.51 – 3.46 (m, 1H, H-4^{II}), 3.46 – 3.40 (m, 1H, H-6_a^{II}), 3.40 – 3.33 (m, 5H, OMe, H-2^{II}, H-6_b^{II}), 3.18 (dd, $J = 13.0, 5.5$ Hz, 1H, SCHHCHPh),

2.83 (dd, $J = 13.0, 7.6$ Hz, 1H, SCHHCHPh); ^{13}C NMR (150 MHz, CDCl_3): δ 161.57, 161.43, 140.61, 139.15, 139.07, 138.64, 138.21, 138.14, 138.04, 128.41, 128.33, 128.28, 128.26, 128.17, 128.15, 128.09, 128.04, 128.01, 127.91, 127.87, 127.71, 127.57, 127.53, 127.49, 127.43, 127.27, 127.25, 127.16, 127.10, 127.02, 101.53, 97.74, 95.70, 90.93, 90.87, 81.77, 81.44, 80.89, 80.52, 77.69, 77.45, 77.26, 77.05, 76.84, 75.07, 74.77, 74.17, 73.37, 73.26, 73.07, 71.43, 70.82, 69.53, 69.03, 68.33, 56.00, 55.81, 55.21, 55.15, 41.15; HR MALDI-TOF MS: m/z : calcd for $\text{C}_{72}\text{H}_{78}\text{O}_{14}\text{S}$ $[\text{M}+\text{Na}]^+$: 1221.5010; found: 1221.5001. The β anomer was purified by reversed phase HPLC on an analytical C-18 column using a gradient of 50 \rightarrow 100% acetonitrile in H_2O over 40 min. **54 β** : $R_f = 0.35$ (EtOAc/hexanes, 1/2, v/v); ^1H NMR (500 MHz, CDCl_3): δ 7.42 – 7.03 (m, 30H, ArH), 6.05 (s, 2H, ArH), 5.04 (t, $J = 6.7$ Hz, 1H, SCH_2CHPh), 4.93 (d, $J = 11.2$ Hz, 1H, CHHPh), 4.79 (d, $J = 12.4$ Hz, 2H, CHHPh, CHHPh), 4.73 – 4.57 (m, 6H, H-1^I, $2\times\text{CH}_2\text{Ph}$, CHHPh), 4.52 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.49 – 4.42 (m, 2H, CHHPh, CHHPh), 4.37 (d, $J = 7.8$ Hz, 1H, H-1^{II}), 4.33 (d, $J = 12.3$ Hz, 1H, CHHPh), 4.12 (d, $J = 8.7$ Hz, 1H, H-6^{aI}), 4.06 (t, $J = 9.7$ Hz, 1H), 3.90 – 3.82 (m, 2H, H-3^I, H-5^I), 3.77 (s, 6H, $2\times\text{OMe}$), 3.75 (s, 3H, OMe), 3.71 – 3.62 (m, 2H, H-6^{bI}, H-6^{aII}), 3.51 – 3.39 (m, 5H, H-6^{bII}, H-2^I, OMe), 3.38 – 3.27 (m, 3H, H-3^{II}, H-4^{II}, SCHHCH), 3.25 – 3.16 (m, 2H, H-2^{II}, H-5^{II}), 2.82 (dd, $J = 13.3, 7.1$ Hz, 1H, SCHHCHPh).; ^{13}C NMR (150 MHz, CDCl_3): δ 179.72, 179.71, 179.71, 179.69, 179.69, 179.68, 179.67, 179.65, 179.61, 179.60, 161.36, 161.32, 140.27, 139.51, 138.92, 138.66, 138.45, 138.31, 137.84, 128.47, 128.30, 128.24, 128.17, 128.14, 128.05, 127.96, 127.90, 127.88, 127.74, 127.70, 127.68, 127.52, 127.49, 127.32, 127.19, 127.02, 102.85, 102.05, 98.76, 97.05, 97.05, 90.92, 84.68, 84.22, 82.04, 80.81, 78.67, 78.13, 78.09, 78.08, 77.20, 76.99, 76.77, 75.47, 75.42, 75.38, 75.22, 74.64,

73.73, 73.41, 73.34, 70.12, 68.93, 68.20, 55.98, 55.69, 55.29, 41.82; HR MALDI-TOF MS: m/z: calcd for C₇₂H₇₈O₁₄S [M+Na]⁺: 1221.5010; found: 1221.4998.

4-S-(2,3,5-trimethoxyphenyl)-2-(S)-phenyl-(3,4,6-tri-O-benzoyl-1,2-dideoxy-β-D-glucopyranoso)[1,2-e]-1,4-oxathianium triflate (56). Compound **56** was prepared according to the general procedure as described in the paper. Selected ¹H NMR (500 MHz, gHSQC, CDCl₃): δ 5.99 (H-3), 5.81 (d, *J* = 10 Hz, H-1), 5.74 (H-4), 4.88 (SCH₂CHPh), 4.50 (H-6_{a,b}), 4.37 (H-2), 4.30 (H-5), 4.35 (SCHHCHPh), 4.19 (SCHHCHPh); ¹³C NMR (125 MHz, gHSQC, CDCl₃): δ 81.60 (C-1), 78.68 (C-2), 78.25 (C-5), 76.00 (SCH₂CHPh), 72.79 (C-3), 68.93 (C-4), 62.55 (C-6), 45.44 (SCH₂CHPh).

4-S-(2,3,5-trimethoxyphenyl)-2-(S)-phenyl-(3,4,6-tri-O-acetyl-1,2-dideoxy-β-D-glucopyranoso)[1,2-e]-1,4-oxathianium triflate (57). Compound **57** was prepared according to the general procedure as described in the paper. Selected ¹H NMR (500 MHz, gHSQC, CDCl₃): δ 5.66 (d, *J* = 10 Hz, H-1), 5.47 (H-3), 5.22 (H-4), 5.07 (SCH₂CHPh), 4.26 (SCHHCHPh), 4.17 (H-6_{a,b}), 4.13 (SCHHCHPh, H-2), 3.92 (H-5); ¹³C NMR (125 MHz, gHSQC, CDCl₃): δ 81.54 (C-1), 78.23 (C-2), 77.59 (C-5), 76.64 (SCH₂CHPh), 72.72 (C-3), 68.06 (C-4), 62.55 (C-6), 45.60 (SCH₂CHPh).

4-S-(2,3,5-trimethoxyphenyl)-2-(S)-phenyl-(3,4,6-tri-O-benzyl-1,2-dideoxy-β-D-glucopyranoso)[1,2-e]-1,4-oxathianium triflate (58). Compound **58** was prepared according to the general procedure as described in the paper. Selected ¹H NMR (500 MHz, gHSQC, CDCl₃): δ 5.61 (d, *J* = 10 Hz, 1H, H-1), 5.24 (d, *J* = 15 Hz, 1H, SCH₂CHPh), 4.92 – 4.78 (m, 3H, PhCH₂, PhCHH), 4.57 (d, *J* = 15 Hz, 1H, SCH₂CHPh), 4.49 – 4.36 (m, 3H, PhCH₂, SCHHCHPh), 3.99 – 3.97 (m, 3H, H-2, H-3, SCHHCHPh), 3.80 (H-4), 3.68 (H-6_{a,b}), 3.66 (H-5); ¹³C NMR (125 MHz, gHSQC, CDCl₃): δ 82.53 (C-2,

C-3), 81.70 (C-1), 81.39 (C-5), 77.82 (SCH₂CHPh), 76.39 (C-4, PhCH₂<), 75.53 (PhCH₂<), 73.96 (PhCH₂<), 67.82 (C-6), 44.69 (SCH₂CHPh).

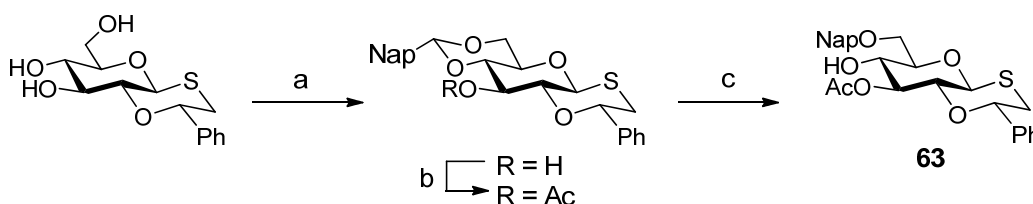
4-S-methyl-2-(S)-phenyl-(3,4,6-tri-O-benzoyl-1,2-dideoxy-β-D-glucopyranoso)[1,2-e]-1,4-oxathianium triflate (59). Compound **59** was prepared according to the general procedure as described in the paper. Selected ¹H NMR (500 MHz, CDCl₃): δ 8.13 – 7.16 (ArH, 20H), 5.99 (t, *J* = 9 Hz, 1H, H-3), 4.81 – 4.74 (m, 2H, H-1, H-4), 5.06 (d, *J* = 10 Hz, 1H, SCH₂CHPh), 4.69 – 4.65 (m, 1H, H-6_a), 4.56 – 4.50 (m, 2H, H-5, H-6_b), 4.30 – 4.27 (m, 2H, H-2, SCHHCHPh), 3.72 (t, *J* = 12 Hz, 1H, SCHHCHPh); ¹³C NMR (125 MHz, gHSQC, CDCl₃): δ 83.70 (C-1), 78.26 (C-2, C-5), 76.84 (SCH₂CHPh), 72.92 (C-3), 68.78 (C-4), 62.35 (C-6), 46.12 (SCH₂CHPh).

4-S-methyl-2-(S)-phenyl-(3,4,6-tri-O-acetyl-1,2-dideoxy-β-D-glucopyranoso)[1,2-e]-1,4-oxathianium triflate (60). Compound **60** was prepared according to the general procedure as described in the paper. Selected ¹H NMR (500 MHz, CDCl₃): δ 5.47 (H-1), 5.42 (H-3), 5.15 (H-4), 4.97 (SCH₂CHPh), 4.25 (H-6_a), 4.16 (SCHHCHPh), 4.14 (H-6_b), 4.12 (H-5), 3.99 (H-2), 3.63 (SCHHCHPh); ¹³C NMR (125 MHz, gHSQC, CDCl₃): δ 83.35 (C-1), 78.05 (C-5, C-2), 76.47 (SCH₂CHPh), 72.35 (C-3), 68.03 (C-4), 62.53 (C-6), 45.85 (SCH₂CHPh).

4-S-methyl-2-(S)-phenyl-(3,4,6-tri-O-benzyl-1,2-dideoxy-β-D-glucopyranoso)[1,2-e]-1,4-oxathianium triflate (61). Compound **61** was prepared according to the general procedure as described in the paper. Selected ¹H NMR (600 MHz, CDCl₃): δ 7.42 – 7.21 (ArH, 20H), 5.31 (d, *J* = 12 Hz, 1H, H-1), 4.89 (d, *J* = 12 Hz, 1H, SCH₂CHPh), 4.83 – 4.78 (m, 2H, PhCHH, PhCHH), 4.70 (d, *J* = 12 Hz, 1H, PhCHH), 4.55 – 4.42 (m, 3H, PhCH₂, PhCHH), 4.07 (d, *J* = 12 Hz, 1H, SCHHCHPh), 3.94 – 3.68 (m, 7H, H-2, H-3, H-4, H-5,

H-6_{a,b}, SCHHCHPh); ¹³C NMR (150 MHz, gHSQC, CDCl₃): δ 83.73 (C-1), 82.70 (C-3), 80.91 (C-2), 80.44 (C-4), 76.50 (SCH₂CHPh), 76.03 (C-5), 75.91 (PhCH₂<), 75.55 (PhCH₂<), 73.53 (PhCH₂<), 67.93 (C-6), 44.94 (SCH₂CHPh).

Scheme S2. Preparation of 63



Reagents and conditions: (a) NapCH(OMe)₂, DMF, TsOH·H₂O, reduced pressure (78%); (b) Ac₂O, Pyridine; (c) Et₃SiH, TfOH, -78 °C (83%, 2 steps).

2-(S)-Phenyl-(3-O-acetyl-6-O-(2-naphthyl)-1,2-dideoxy-β-D-glucopyranoso)[1,2-e]-

1,4-oxathiane (63): $R_f = 0.25$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +60.0 ($c = 1.0$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.94 – 7.12 (m, 12H, ArH), 5.11 (t, $J = 9.3$ Hz, 1H, H-3), 4.76 (2H, NapCH₂), 4.67 (dd, $J = 10.5, 1.7$ Hz, 1H, SCH₂CHPh), 4.47 (d, $J = 8.9$ Hz, 1H, H-1), 3.88 – 3.74 (m, 3H, H-4, H-6_{a,b}), 3.74 – 3.60 (m, 2H, H-2, H-5), 2.95 (dd, $J = 14.0, 10.6$ Hz, 1H, SCHHCHPh), 2.77 (dd, $J = 14.0, 2.0$ Hz, 1H, SCHHCHPh), 2.06 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 171.90, 140.44, 135.29, 133.47, 133.29, 128.69, 128.56, 128.16, 128.06, 127.94, 126.95, 126.42, 126.24, 125.91, 125.61, 81.24, 80.01, 79.74, 77.71, 77.29, 76.87, 76.08, 76.01, 74.10, 71.03, 70.06, 35.86, 21.27; HR MALDI-TOF MS: m/z : calcd for C₂₇H₂₈O₆S [M+Na]⁺: 503.1504; found: 503.1521.

2-(S)-Phenyl-(3,4-di-O-acetyl-6-O-benzyl-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 4)-3-O-acetyl-6-O-(2-methylnaphthyl)-1,2-dideoxy- β -D-glucopyranoso)[1,2-*e*]-1,4-oxathiane (64).

Compound **64** was prepared according to the general procedure using glycosyl donor **17** (53 mg, 0.11 mmol) and glycosyl acceptor **63** (43 mg, 0.091 mmol). Purification by LH20 size exclusion chromatography afforded compound **64** (69 mg, 68 %). $R_f = 0.27$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_D^{29}$ (deg cm³ g⁻¹ dm⁻¹) = +75.0 ($c = 1.6$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.94 – 6.94 (m, 22H, ArH), 6.08 (s, 2H, ArH), 5.82 (d, $J = 3.3$ Hz, 1H, H-1^{II}), 5.52 (t, $J = 9.5$ Hz, 1H, H-3^I), 5.25 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 4.92 (t, $J = 9.5$ Hz, 1H, H-4^{II}), 4.77 – 4.69 (m, 3H, SCH₂CHPh^{II}, CH₂Nap), 4.54 (d, $J = 9.0$ Hz, 1H, H-1^I), 4.45 – 4.30 (m, 2H, H-4^I, CHHPh), 4.30 – 4.18 (m, 2H, SCH₂CHPh^I, CHHPh), 4.11 (dd, $J = 11.5, 3.8$ Hz, 1H, H-6^I_a), 4.06 – 3.91 (m, 2H, H-6^I_b, H-5^{II}), 3.84 (m, 1H, H-5^I), 3.81 (s, 3H, OMe), 3.77 (s, 6H, 2 \times OMe), 3.71 – 3.67 (m, 2H, H-2^I, H-2^{II}), 3.35 (dd, $J = 10.6, 2.6$ Hz, 1H, H-6^{II}_a), 3.26 (dd, $J = 10.7, 3.7$ Hz, 1H, H-6^{II}_b), 3.07 – 2.67 (m, 4H, SCH₂CHPh^I, SCH₂CHPh^{II}), 2.05 (s, 3H), 1.83 (s, 3H), 1.32 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.46, 170.23, 170.04, 162.13, 161.96, 142.19, 140.53, 138.03, 136.09, 133.50, 133.11, 128.66, 128.48, 128.37, 128.18, 128.17, 128.03, 127.96, 127.85, 127.80, 127.53, 126.33, 126.30, 126.15, 125.89, 125.86, 125.45, 101.43, 97.11, 91.17, 84.20, 82.15, 80.71, 79.61, 79.47, 77.67, 77.25, 76.82, 75.64, 74.15, 73.78, 73.50, 72.35, 69.70, 69.29, 69.19, 68.47, 56.21, 55.57, 43.33, 35.76, 21.80, 20.91, 20.39; HR MALDI-TOF MS: m/z : calcd for C₆₁H₆₆O₁₆S₂ [M+Na]⁺: 1141.3690; found: 1141.3697.

2-(S)-Phenyl 3-O-acetyl-6-O-benzyl-4-O(9-fluorenylmethyloxycarbonyl)-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 4)-

3-*O*-acetyl-6-*O*-(2-methylnaphthyl)-1,2-dideoxy- β -D-glucopyranoso)[1,2-*e*]-1,4-oxathiane (65). Compound **65** was prepared according to the general glycosylation procedure using glycosyl donor **19** (67 mg, 0.09 mmol) and glycosyl acceptor **63** (40 mg, 0.08 mmol). Purification by LH20 size exclusion chromatography afforded compound **65** (60 mg, 56%). $R_f = 0.33$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +56.0 ($c = 2.5$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.86 – 6.97 (m, 30H, ArH), 6.08 (s, 2H, ArH), 5.86 (d, $J = 3.1$ Hz, 1H, H-1^{II}), 5.55 (t, $J = 9.5$ Hz, 1H, H-3^I), 5.41 (t, $J = 9.7$ Hz, 1H, H-3^{II}), 4.94 – 4.64 (m, 4H, SCH₂CHPh^I, CH₂Nap, H-4^{II}), 4.55 (d, $J = 8.9$ Hz, 1H, H-1^I), 4.48 – 4.19 (m, 5H, SCH₂CHPh^{II}, H-4^I, CH₂Ph, CHH Fmoc), 4.18 – 4.05 (m, 4H, H-5^{II}, CH Fmoc, CHH Fmoc, H-6^{aI}), 3.97 (d, $J = 11.1$ Hz, 1H, H-6^{bI}), 3.92 – 3.61 (m, 12H, H-5^I, H-2^I, H-2^{II}, 3×OMe), 3.51 – 3.29 (m, 2H, H-6^{a, bII}), 3.05 – 2.75 (m, 4H, SCH₂CHPh^I, SCH₂CHPh^{II}), 2.07 (s, 3H), 1.27 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.19, 169.79, 161.91, 161.73, 154.28, 143.28, 143.18, 141.95, 141.20, 141.16, 140.30, 137.83, 135.86, 133.27, 132.90, 128.43, 128.20, 128.15, 127.98, 127.85, 127.65, 127.62, 127.51, 127.29, 127.25, 127.21, 126.16, 126.08, 125.89, 125.70, 125.64, 125.25, 125.21, 125.15, 119.96, 101.13, 96.82, 90.93, 84.09, 81.98, 80.41, 79.38, 77.45, 77.02, 76.60, 75.37, 73.95, 73.66, 73.58, 73.32, 71.69, 70.18, 68.90, 68.85, 68.38, 55.97, 55.33, 46.54, 43.11, 35.51, 21.58, 20.16; HR MALDI-TOF MS: m/z : calcd for C₇₄H₇₄O₁₇S₂ [M+Na]⁺: 1321.4265; found: 1321.4659.

2-(*S*)-Phenyl 3,6-di-*O*-acetyl-4-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1→4)-3-*O*-acetyl-6-*O*-(2-methylnaphthyl)-1,2-dideoxy- β -D-glucopyranoso)[1,2-*e*]-1,4-oxathiane (66).

Compound **66** was prepared according to the general glycosylation procedure using

glycosyl donor **20** (40 mg, 0.082 mmol) and glycosyl acceptor **63** (33 mg, 0.069 mmol). Purification by LH20 size exclusion chromatography afforded compound **66** (42 mg, 55%). $R_f = 0.25$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +36.4 ($c = 1.1$ in CHCl₃); ¹H NMR (600 MHz, CDCl₃): δ 7.92 – 7.03 (m, 22H, ArH), 6.09 (s, 2H, ArH), 5.71 (d, $J = 3.3$ Hz, 1H, H-1^{II}), 5.47 (t, $J = 9.5$ Hz, 1H, H-3^I), 5.39 (t, $J = 9.5$ Hz, 1H, H-3^{II}), 4.76 (s, 2H, CH₂Nap), 4.69 (d, $J = 9.3$ Hz, 1H, SCH₂CHPh^{II}), 4.51 (d, $J = 8.9$ Hz, 1H, H-1^I), 4.46 (d, $J = 11.1$ Hz, 1H, CHHPh), 4.39 (d, $J = 11.1$ Hz, 1H, CHHPh), 4.34 – 4.20 (m, 2H, H-4I, SCH₂CHPh^I), 4.19 – 4.09 (m, 2H, H-6^{II}_{a,b}), 4.09 – 4.01 (m, 2H, H-5^{II}, H-6^I_a), 3.96 (d, $J = 10.3$ Hz, 1H, H-6^I_b), 3.87 – 3.75 (m, 10H, 3×OMe, H-5^I), 3.66 (t, $J = 9.3$ Hz, 1H, H-2^I), 3.46 (dd, $J = 10.0, 3.3$ Hz, 1H, H-2^{II}), 3.33 (t, $J = 9.5$ Hz, 1H, H-4^{II}), 3.06 – 2.89 (m, 3H, SCH₂CHPh^I, SCHHCHPh^{II}), 2.82 (dd, $J = 13.9, 1.8$ Hz, 1H, SCHHCHPh^{II}), 2.08 (s, 3H), 1.96 (s, 3H), 1.43 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.62, 170.46, 169.72, 162.13, 161.92, 141.85, 140.56, 137.71, 135.97, 133.49, 133.11, 128.65, 128.40, 128.21, 128.17, 128.11, 127.87, 127.84, 127.76, 126.78, 126.40, 126.10, 125.89, 125.85, 125.49, 101.85, 96.72, 91.20, 84.11, 82.03, 80.82, 79.57, 79.41, 77.68, 77.46, 77.26, 76.84, 76.53, 75.76, 74.55, 73.93, 73.76, 73.18, 69.30, 68.99, 63.25, 56.22, 55.61, 43.12, 35.80, 21.66, 21.03, 20.83; HR MALDI-TOF MS: m/z : calcd for C₆₁H₆₆O₁₆S₂ [M+Na]⁺: 1141.3690; found: 1141.3685.

2-(S)-Phenyl 2,3-di-O-acetyl-6-O-benzyl-4-O- (9-fluorenylmethoxycarbonyl)- α -D-glucopyranosyl-(1→4)-3-O-acetyl-6-O-(2-methylnaphthyl)-1,2-dideoxy- β -D-glucopyranoso)[1,2-e]-1,4-oxathiane (67). C-2 auxiliary of **65** was removed using general procedure. Then the residue was redissolved in pyridine and an equal volume of acetic anhydride was added. After stirring for 16 h, the solvents were removed and the

product was purified by silica column chromatography to afford **67** (89%, 2 steps). $R_f = 0.35$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{26}^d$ (deg cm³ g⁻¹ dm⁻¹) = +23.5 ($c = 0.8$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.99 – 7.00 (m, 25H, ArH), 5.57 (d, $J = 3.9$ Hz, 1H, H-3^{II}), 5.52 (t, $J = 9.9$ Hz, 1H, H-3^{II}), 5.39 (t, $J = 9.4$ Hz, 1H, H-3^I), 5.04 (t, $J = 9.9$ Hz, 1H, H-4^{II}), 4.93 – 4.75 (m, 2H, H-2^{II}, CHHNap), 4.75 – 4.61 (m, 2H, CHHNap, SCH₂CHPh), 4.51 (d, $J = 8.9$ Hz, 1H, H-1^I), 4.41 – 3.95 (m, 8H, H-4^I, CH₂ Fmoc, CH Fmoc, CH₂Ph, H-5^{II}, H-6^I_a), 3.86 (d, $J = 10.9$ Hz, 1H, H-6^I_b), 3.75 (d, $J = 9.5$ Hz, 1H, H-5^I), 3.61 (t, $J = 9.3$ Hz, 1H, H-2^I), 3.40 – 3.14 (m, 2H, H-6^{II}_a, H-6^{II}_b), 2.96 (dd, $J = 14.0, 10.5$ Hz, 1H, SCHHCHPh), 2.82 (dd, $J = 13.9, 2.0$ Hz, 1H, SCHHCHPh), 2.07 (s, 3H), 1.98 (s, 3H), 1.95 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.81, 170.57, 170.15, 154.24, 143.47, 143.34, 141.47, 140.28, 137.77, 135.79, 133.49, 133.24, 128.70, 128.39, 128.37, 128.21, 128.17, 127.93, 127.84, 127.74, 127.51, 127.46, 126.77, 126.35, 126.15, 126.12, 125.36, 125.32, 120.30, 120.28, 95.25, 81.96, 80.30, 79.74, 77.67, 77.45, 77.25, 76.83, 75.95, 75.83, 74.24, 73.56, 73.07, 71.11, 70.58, 70.53, 69.75, 69.00, 68.76, 67.81, 60.62, 46.77, 35.73, 21.31, 20.94, 20.86; HR MALDI-TOF MS: m/z : calcd for C₅₉H₅₈O₁₅S [M+Na]⁺: 1061.3394; found: 1061.3387.

3-azidopropyl 2,3-di-O-acetyl-6-O-benzyl-4-O-(9-fluorenylmethoxycarbonyl)- α -D-glucopyranosyl-(1 \rightarrow 4)-3-O-acetyl-6-O-(2-methylnaphthyl)-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranoside (70). Compound **67** was oxidized to give sulfoxide donor **68** according to the general procedure. Compound **70** was prepared according to the general glycosylation procedure using glycosyl donor **68** (45 mg, 0.04 mmol) and glycosyl acceptor **69** (22 mg, 0.22 mmol). Purification by LH20 size exclusion chromatography afforded compound **70** (48 mg, 86%). $R_f = 0.26$

(EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +80.0 ($c = 0.8$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.92 – 6.97 (m, 25H, ArH), 6.11 (s, 2H, ArH), 5.55 – 5.35 (m, 3H, H-1^I, H-3^I, H-3^{II}), 5.29 (d, $J = 3.8$ Hz, 1H, H-1^{II}), 4.97 (t, $J = 9.9$ Hz, 1H, H-4^{II}), 4.81 (dd, $J = 10.5, 3.8$ Hz, 1H, H-2^{II}), 4.72 (q, $J = 12.2$ Hz, 2H, CH₂Nap), 4.44 – 4.11 (m, 5H, SCH₂CHPh, CH₂Ph, CHH Fmoc, CH Fmoc), 4.07 – 3.90 (m, 4H, CHH Fmoc, H-5^I, H-4^I, H-5^{II}), 3.83 (s, 10H, 3×OMe, H-6^I_a), 3.76 – 3.57 (m, 3H, H-6^I_b, CH₂ linker), 3.57 – 3.40 (m, 3H, H-2^I, CH₂ linker), 3.27 (d, $J = 2.8$ Hz, 2H, H-6^{II}_a, H-6^{II}_b), 2.94 (qd, $J = 13.8, 6.3$ Hz, 2H, SCH₂CHPh), 2.02 (s, 3H), 2.00 – 1.95 (m, 2H, CH₂ linker), 1.92 (s, 3H) 1.44 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 170.59, 169.82, 161.85, 161.77, 154.03, 143.23, 143.14, 141.46, 141.23, 137.56, 135.61, 133.24, 132.94, 128.22, 128.13, 128.07, 127.91, 127.88, 127.71, 127.54, 127.47, 127.26, 127.21, 126.69, 126.14, 126.11, 125.89, 125.56, 125.13, 120.04, 101.72, 97.24, 94.86, 90.96, 83.93, 78.97, 77.44, 77.22, 77.01, 76.59, 73.76, 73.65, 73.32, 73.07, 71.83, 70.26, 70.19, 69.67, 69.50, 68.86, 68.69, 67.92, 65.31, 56.02, 55.37, 48.53, 46.53, 42.74, 29.00, 20.73, 20.69, 20.67; HR MALDI-TOF MS: m/z : calcd for C₇₁H₇₅N₃O₁₉S [M+Na]⁺: 1328.4613; found: 1328.4622.

3-azidopropyl 2,3-di-O-acetyl-6-O-benzyl-4-O-(9-fluorenylmethoxycarbonyl)- α -D-glucopyranosyl-(1 \rightarrow 4)-3-O-acetyl-6-O-(2-methylnaphthyl)-2-O-[(1S)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranoside (71). The C-2 auxiliary of **70** was removed using the general procedure. The residue was redissolved in pyridine and an equal volume of acetic anhydride was added. After stirring for 16 h, the solvent was removed and the product was purified by silica column chromatography to afford **71** (98%, 2 steps). $R_f = 0.44$ (EtOAc/hexanes, 1/2, v/v); $[\alpha]_{29}^d$ (deg cm³ g⁻¹ dm⁻¹) = +64.7 ($c = 1.7$ in CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.94 – 6.98 (m, 20H, ArH), 5.65 – 5.39

(m, 3H, H-1^I, H-3^I, H-3^{II}), 5.09 – 4.99 (m, 2H, H-1^{II}, H-4^{II}), 4.91 – 4.80 (m, 2H, H-2^I, H-2^{II}), 4.74 (dd, $J = 28.2, 9.5$ Hz, 2H, CH₂Nap), 4.43 – 4.11 (m, 5H, CHH Fmoc, CH Fmoc, CH₂Ph, H-4^I), 4.08 – 3.88 (m, 3H, CHH Fmoc, H-5^I, H-5^{II}), 3.88 – 3.67 (m, 3H, H-6^I_a, CH₂ linker), 3.58 – 3.37 (m, 3H, H-6^I_b, CH₂ linker), 3.27 (s, 2H, H-6^{II}_a, H-6^{II}_b), 2.06 (s, 3H), 2.04 (s, 3H), 2.02 (s, 3H), 1.95 (s, 3H), 1.93 – 1.86 (m, 2H, CH₂ linker); ¹³C NMR (75 MHz, CDCl₃): δ 170.56, 170.33, 170.01, 169.90, 154.01, 143.23, 143.10, 141.25, 137.55, 135.48, 133.25, 133.00, 128.16, 127.94, 127.71, 127.57, 127.51, 127.27, 127.22, 126.34, 126.15, 125.93, 125.70, 125.11, 120.07, 95.86, 95.15, 77.43, 77.21, 77.01, 76.59, 73.90, 73.32, 72.89, 72.75, 71.59, 71.52, 70.44, 70.29, 69.87, 69.46, 68.83, 68.50, 67.73, 65.03, 48.14, 46.54, 28.75, 21.01, 20.70, 20.66, 20.65; HR MALDI-TOF MS: m/z: calcd for C₅₆H₅₉N₃O₁₇ [M+Na]⁺: 1068.3742; found: 1068.3738.

3-azidopropyl 2,3-di-O-acetyl-6-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-O-acetyl-6-O-(2-methylnaphthyl)- α -D-glucopyranoside (72). *N*-methyl-2-pyrrolidone (NMP) (0.8 mL) was added to a stirred solution of **71** (130 mg, 0.022 mmol) in DCM (4 mL). After stirring for 1 h, the reaction mixture was diluted with DCM (20 mL) and washed H₂O (2 \times 20 mL) and brine (20 mL). The organic phase was dried (MgSO₄), filtered and the filtrate was concentrated *in vacuo*. The residue was purified by flash chromatography over silica gel (EtOAc/hexanes, 1/4 \rightarrow 1/2, v/v) to give **72** (93 mg, 91%). $R_f = 0.24$ (EtOAc/hexanes, 1/2, v/v); ¹H NMR (300 MHz, CDCl₃): δ 7.93 – 7.06 (m, 12H, ArH), 5.68 – 5.43 (m, 1H, H-3^{II}), 5.38 (d, $J = 3.9$ Hz, 1H, H-1^I), 5.28 – 5.09 (m, 1H, H-3^I), 5.01 (d, $J = 3.7$ Hz, 1H, H-1^{II}), 4.93 – 4.63 (m, 4H, H-2^I, H-2^{II}, CH₂Nap), 4.28 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.22 – 4.06 (m, 2H, H-4^{II}, CHHPh), 3.92 (d, $J = 9.2$ Hz, 2H, H-5^I, H-6^I_a), 3.87 – 3.59 (m, 4H, H-6^I_b, H-4^I, H-5^I, CHH linker), 3.57 – 3.36 (m, 4H, CHH

linker, CH₂ linker, H-6^{II}_a), 3.32 (d, *J* = 10.1 Hz, 1H, H-6^{II}_b), 2.63 (s, 1H, OH), 2.08 (s, 3H), 2.07 (s, 3H), 2.03 (s, 3H), 2.00 (s, 2H), 1.97 – 1.81 (m, 2H, CH₂ linker); ¹³C NMR (75 MHz, CDCl₃): δ 171.41, 170.97, 170.54, 170.20, 137.72, 135.73, 133.46, 133.19, 128.60, 128.40, 128.13, 127.98, 127.93, 127.78, 126.39, 126.15, 125.75, 96.06, 95.48, 77.66, 77.43, 77.23, 76.81, 73.84, 73.73, 73.09, 72.33, 71.76, 71.63, 70.95, 70.65, 70.44, 69.96, 69.57, 68.70, 65.22, 48.36, 28.95, 21.22, 21.12, 20.92, 20.88; HR MALDI-TOF MS: *m/z*: calcd for C₄₁H₄₉N₃O₁₅ [M+Na]⁺: 846.3061; found: 846.3077.

3-azidopropyl **3,4-di-*O*-acetyl-6-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-*O*-acetyl-6-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-*O*-acetyl-6-*O*-(2-methylnaphthyl)- α -D-glucopyranoside (73).** Compound **73** was prepared according to the general glycosylation procedure using glycosyl donor **17** (20 mg, 0.041 mmol) and glycosyl acceptor **72** (28 mg, 0.034 mmol). Purification by LH20 size exclusion chromatography afforded compound **73** (32 mg, 65%). *R_f* = 0.51 (acetone/toluene, 1/4, v/v); ¹H NMR (300 MHz, CDCl₃): δ 7.99 – 6.96 (m, 22H, ArH), 6.10 (s, 2H, ArH), 5.69 (d, *J* = 3.4 Hz, 1H, H-1^{III}), 5.64 (t, *J* = 10.1 Hz, 1H, H-3^I), 5.55 (t, *J* = 9.6 Hz, 1H, H-3^{II}), 5.42 (d, *J* = 3.9 Hz, 1H, H-1^I), 5.25 (t, *J* = 9.8 Hz, 1H, H-3^{III}), 5.00 (d, *J* = 3.7 Hz, 1H, H-1^{II}), 4.93 (t, *J* = 9.8 Hz, 1H, H-4^{III}), 4.81 (dd, *J* = 10.1, 3.7 Hz, 2H, H-2^I, H-2^{II}), 4.76 – 4.63 (m, 2H, CH₂Nap), 4.39 (d, *J* = 11.9 Hz, 1H, CHHPh), 4.31 – 4.13 (m, 6H, CHHPh, CH₂Ph SCH₂CHPh, H-4^I, H-4^{II}), 4.06 – 3.85 (m, 5H, H-5^I, H-5^{II}, H-5^{III}, H-6^{III}_a, H-6^{III}_b), 3.85 – 3.63 (m, 12H, 3×OMe, H-2^{III}, CHH linker, H-6^I_a), 3.53 – 3.34 (m, 4H, CH₂ linker, CHH linker, H-6^I_b), 3.27 (d, *J* = 10.8 Hz, 1H, H-6^{II}_a), 3.15 (dd, *J* = 10.6, 3.3 Hz, 1H, H-6^{II}_b), 3.00 (dd, *J* = 13.8, 4.0 Hz, 1H, SCHHCHPh), 2.86 (dd, *J* = 13.6, 8.6 Hz, 1H, SCHHCHPh), 2.07 (s,

3H), 2.06 (s, 3H), 2.04 (s, 3H), 2.03 (s, 3H), 1.92 – 1.80 (m, 5H, CH₂ linker, CH₃), 1.40 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ171.02, 170.57, 170.23, 169.94, 162.07, 161.93, 142.11, 138.30, 137.96, 135.89, 133.51, 133.21, 128.46, 128.35, 128.28, 128.21, 128.12, 127.91, 127.83, 127.49, 127.31, 126.43, 126.33, 126.20, 125.92, 101.54, 97.21, 96.10, 95.43, 91.17, 83.52, 78.82, 77.65, 77.43, 77.23, 76.81, 73.85, 73.54, 73.25, 73.05, 72.75, 72.44, 71.83, 71.66, 71.49, 69.99, 69.22, 68.76, 68.18, 65.19, 56.16, 55.56, 48.37, 43.04, 28.96, 21.73, 21.25, 20.90, 20.50; HR MALDI-TOF MS: m/z: calcd for C₇₈H₈₇N₃O₂₅S [M+Na]⁺: 1484.5247; found: 1484.5251.

3-azidopropyl 2,3,4-tri-O-acetyl-6-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-O-acetyl-6-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-O-acetyl-6-O-(2-methylnaphthyl)- α -D-glucopyranoside (74). The C-2 auxiliary of **73** was removed using general procedure as described. The residue was redissolved in pyridine and equal volume of acetic anhydride was added. After stirring for 16 h, the solvents were removed and the product was purified by silica column chromatography to afford **74** (76%, 2 steps). R_f = 0.57 (EtOAc/hexanes, 1/1, v/v); ¹H NMR (300 MHz, CDCl₃): δ7.95 – 7.03 (m, 17H, ArH), 5.54 (t, *J* = 9.6 Hz, 1H, H-3^I), 5.49 – 5.29 (m, 4H, H-1^{II}, H-1^{III}, H-3^{II}, H-3^{III}), 5.15 (t, *J* = 9.9 Hz, 1H, H-4^{III}), 5.01 (d, *J* = 3.7 Hz, 1H, H-1^I), 4.88 – 4.64 (m, 5H, H-2^{III}, H-2^{II}, H-2^I, CH₂Nap), 4.42 (d, *J* = 12.1 Hz, 1H, CHHPh), 4.23 – 3.88 (m, 7H, CH₂Ph, CHHPh, H-6_a^I, H-5^I, H-4^{II}, H-4^I), 3.88 – 3.64 (m, 4H, CHH linker, H-6_b^I, H-5^{III}, H-5^{II}), 3.64 – 3.37 (m, 4H, H-6_a^{II}, CH₂ linker, CHH linker), 3.28 (d, *J* = 10.8 Hz, 1H, H-6_b^{II}), 3.17 – 3.07 (m, 2H, H-6_{a,b}^{III}), 2.09 – 1.97 (4×s, 18H), 1.96 – 1.80 (m, 5H, CH₂ linker overlap); ¹³C NMR (75 MHz, CDCl₃): δ170.91, 170.74, 170.56, 170.46, 170.18, 170.15, 169.52, 138.17, 137.75, 135.83, 133.51, 133.24, 128.48, 128.42, 128.36, 128.20,

128.17, 127.92, 127.88, 127.67, 127.39, 126.54, 126.30, 126.07, 125.97, 96.12, 95.46, 95.12, 77.66, 77.44, 77.24, 76.81, 74.12, 73.54, 72.85, 72.49, 72.13, 71.75, 71.27, 70.99, 70.81, 70.53, 70.11, 69.15, 69.00, 68.79, 68.48, 67.47, 65.22, 48.35, 28.97, 21.21, 20.96, 20.86, 20.83; HR MALDI-TOF MS: m/z: calcd for C₆₀H₇₁N₃O₂₃ [M+Na]⁺: 1224.4376; found: 1224.4370.

3-azidopropyl 2,3,4-tri-O-acetyl-6-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-O-acetyl-6-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-O-acetyl- α -D-glucopyranoside (75). 2,3-Dichloro-5,6-dicyano-*p*-benzoquinone (DDQ) (9.4 mg, 0.066 mmol) was added to a stirred solution of **74** (27 mg, 0.022 mmol) in DCM (2 mL) and H₂O (0.2 mL). After stirring for 2 h, the reaction mixture was diluted with DCM (20 mL) and washed with saturated NaHCO₃ (2 \times 20 mL), brine (20 mL). The organic phase was dried (MgSO₄), filtered and the filtrate was concentrated *in vacuo*. The residue was purified by flash chromatography over silica gel (EtOAc/hexanes, 2/1 \rightarrow 1/1, v/v) to give **74** (22 mg, 92%). R_f = 0.37 (EtOAc/hexanes, 1/1, v/v); ¹H NMR (300 MHz, CDCl₃): δ 7.41 – 7.16 (m, 10H), 5.55 (t, *J* = 9.6 Hz, 1H, H-3^{II}), 5.46 – 5.25 (m, 3H, H-1^I, H-1^{III}, H-3^{III}), 5.08 (t, *J* = 9.8 Hz, 1H, H-4^{III}), 4.97 (d, *J* = 3.7 Hz, 1H, H-1^{II}), 4.83 (dd, *J* = 10.5, 4.0 Hz, 1H, H-2^{III}), 4.80 – 4.69 (m, 2H, H-2^I, H-2^{II}), 4.55 (s, 2H, CH₂Ph), 4.50 (d, *J* = 12.0 Hz, 1H, CHHPh), 4.27 (d, *J* = 12.0 Hz, 1H, CHHPh), 4.01 – 3.88 (m, 3H, CH₂ linker, H-4^I), 3.88 – 3.65 (m, 5H, H-5^{II}, H-5^{III}, H-6^I_a, CH₂ linker), 3.58 – 3.38 (m, 3H, H-6^I_b, H-6^{III}_a, H-6^{III}_b), 3.33 – 3.19 (m, 2H, H-6^{II}_a, H-6^{II}_b), 2.72 (dd, *J* = 7.7, 4.9 Hz, 1H, OH), 2.04 – 1.9 (6s, 23H, 7 \times Me, CH₂ linker overlap); ¹³C NMR (75 MHz, CDCl₃): δ 170.95, 170.73, 170.50, 170.32, 170.14, 170.08, 169.57, 137.70, 137.47, 128.69, 128.58, 128.17, 128.14, 128.07, 128.02, 96.20, 95.76, 95.60, 77.65, 77.43, 77.23, 76.80, 74.04, 73.72,

73.11, 72.46, 72.35, 71.86, 71.58, 70.84, 70.76, 70.33, 70.23, 70.04, 69.59, 69.02, 68.92, 67.77, 65.20, 60.84, 48.35, 28.98, 21.22, 21.16, 20.90, 20.86, 20.84, 20.80; HR MALDI-TOF MS: m/z : calcd for $C_{49}H_{63}N_3O_{23}$ $[M+Na]^+$: 1084.3750; found: 1084.3759.

3-azidopropyl 2,3,4-tri-*O*-acetyl-6-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-*O*-acetyl-6-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-*O*-acetyl-6-*O*-{3,6-di-*O*-acetyl-4-*O*-benzyl-2-*O*-[(1*S*)-phenyl-2-(2,3,5-trimethoxyphenylsulfanyl)-ethyl]- α -D-glucopyranosyl}- α -D-glucopyranoside (76).

Compound **76** was prepared according to the general glycosylation procedure using glycosyl donor **20** (35 mg, 0.072 mmol) and glycosyl acceptor **75** (25 mg, 0.024 mmol). Purification by LH20 size exclusion chromatography afforded compound **76** (28 mg, 70%). $R_f = 0.19$ (EtOAc/hexanes, 1/1, v/v); $[\alpha]_D^{29}$ (deg $cm^3 g^{-1} dm^{-1}$) = +80.0 ($c = 1.0$ in $CHCl_3$); 1H NMR (600 MHz, $CDCl_3$) δ 7.41 – 7.05 (m, 15H, *ArH*), 6.14 (s, 2H, *ArH*), 5.77 (d, $J = 3.4$ Hz, 1H, H-1^{IV}), 5.58 – 5.40 (m, 4H, H-3^{II}, H-3^I, H-3^{IV}, H-1^{III}), 5.37 – 5.26 (m, 2H, H-1^I, H-3^{III}), 5.17 (t, $J = 9.9$ Hz, 1H, H-4^{III}), 4.98 (d, $J = 3.8$ Hz, 1H, H-1^{II}), 4.87 – 4.77 (m, 3H, H-2^I, H-2^{II}, H-2^{III}), 4.64 (d, $J = 11.9$ Hz, 1H, *CHHPh*), 4.48 – 4.37 (m, 2H, *CHHPh*, *CHHPh*), 4.34 – 4.16 (m, 5H, SCH_2CHPh , H-4^{II}, H-4^I, CH_2Ph), 4.09 – 3.94 (m, 7H, H-5^I, H-5^{II}, H-5^{IV}, *CHHPh*, H-6^I_a, H-6^{IV}_{a,b}), 3.94 – 3.78 (m, 13H, 3 \times OMe, H-5^{III}, H-6^I_b, *CHH* linker, H-6^{II}_a), 3.68 (dd, $J = 10.2, 3.4$ Hz, 1H, H-2^{IV}), 3.54 – 3.34 (m, 5H, CH_2 linker, *CHH* linker, H-6^{II}_b, H-4^{IV}), 3.22 – 3.05 (m, 3H, $SCHHCHPh$, H-6^{III}_{a,b}), 2.82 (dd, $J = 14.1, 8.8$ Hz, 2H, $SCHHCHPh$), 2.12 – 1.85 (m, 20H, 6 \times OAc, CH_2 linker), 1.76 (s, 3H), 1.26 (s, 3H); ^{13}C NMR (150 MHz, $CDCl_3$): δ 179.62, 179.60, 179.58, 179.57, 179.55, 170.67, 170.55, 170.45, 170.05, 170.01, 169.91, 169.47, 169.37, 161.79, 161.57, 142.26, 138.62, 137.64, 137.48, 128.33, 128.19, 128.13, 128.11, 127.95, 127.90, 127.82, 127.53,

127.46, 127.27, 127.18, 125.93, 101.25, 98.11, 95.89, 95.54, 95.06, 90.89, 84.21, 80.22, 77.99, 77.21, 76.99, 76.78, 76.52, 74.00, 73.71, 73.64, 73.54, 73.19, 72.56, 72.53, 71.53, 71.16, 71.08, 71.04, 70.25, 70.22, 70.09, 68.87, 68.81, 68.49, 68.43, 66.83, 65.05, 65.00, 63.02, 55.87, 55.36, 48.22, 42.89, 28.70, 20.99, 20.98, 20.84, 20.70, 20.67, 20.64, 20.60, 20.52, 20.31. HR MALDI-TOF MS: m/z: calcd for C₈₃H₁₀₁N₃O₃₃S [M+Na]⁺: 1722.5936; found: 1722.5947.

3-azidopropyl 2,3,4-tri-O-acetyl-6-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-O-acetyl-6-O-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)- α -D-glucopyranosyl-(1 \rightarrow 4)-2,3-di-O-acetyl-6-O-(3,4-di-O-acetyl-6-O-benzyl-D-glucopyranosyl)- α -D-glucopyranoside (78).

Compound **77** was prepared according to the general glycosylation procedure using glycosyl donor **17** (19 mg, 0.039 mmol) and glycosyl acceptor **75** (21 mg, 0.020 mmol). Purification by LH20 size exclusion chromatography afforded compound **77** (25 mg, 73%, $\alpha/\beta=8/1$). The resulting tetrasaccharide was subjected to general C-2 auxiliary removal condition to give **78** (17 mg, 85%). Isomers were separated by silica gel chromatography. **78a**: R_f = 0.14 (EtOAc/hexanes, 1/1, v/v); ¹H NMR (600 MHz, CDCl₃): δ 7.41 – 7.14 (m, 15H, ArH), 5.50 (t, *J* = 9.5 Hz, 1H, H-3^{II}), 5.45 (d, *J* = 4.0 Hz, 1H, H-1^{III}), 5.40 (t, *J* = 9.6 Hz, 1H, H-3^I), 5.35 (d, *J* = 4.0 Hz, 1H, H-1^I), 5.31 (t, *J* = 9.6 Hz, 1H, H-3^{III}), 5.25 (t, *J* = 9.7 Hz, 1H, H-3^{IV}), 5.19 – 5.08 (m, 3H, H-1^{IV}, H-4^{III}, H-4^{IV}), 4.96 (d, *J* = 3.8 Hz, 1H, H-1^{II}), 4.83 (dd, *J* = 10.5, 4.0 Hz, 1H, H-2^{III}), 4.76 – 4.70 (m, 2H, H-2^I, H-2^{II}), 4.60 (d, *J* = 11.9 Hz, 1H, CHHPh), 4.57 (d, *J* = 12.1 Hz, 1H, CHHPh), 4.51 – 4.40 (m, 3H, CHHPh, CHHPh, CHHPh), 4.16 – 4.08 (m, 2H, CHHPh, H-4^I), 4.04 – 4.00 (m, 1H, H-5^{IV}), 4.00 – 3.80 (m, 8H, H-5^I, H-5^{II}, H-4^{II}, H-5^{III}, CH₂ linker, H-6^I_a, H-6^{II}_a), 3.76 – 3.67 (m, 2H, H-2^{IV}, H-6^I_b), 3.57 – 3.42 (m, 5H, H-6^{IV}_{a,b}, CH₂ linker, H-6^{II}_b), 3.26 – 3.05

(m, 2H, H-6^{III}_{a,b}), 2.60 (d, $J = 10.7$ Hz, 1H, OH), 2.06 – 1.99 (6s, 21H), 1.97 – 1.90 (m, 2H, CH₂ linker), 1.89 (s, 3H), 1.83 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 171.00, 170.95, 170.52, 170.29, 170.14, 169.86, 169.80, 169.54, 169.30, 138.04, 137.68, 137.50, 128.33, 128.31, 128.30, 128.26, 128.25, 127.97, 127.95, 127.88, 127.87, 127.67, 127.60, 127.41, 127.40, 98.10, 95.75, 95.53, 95.04, 77.20, 76.99, 76.77, 73.92, 73.69, 73.45, 73.44, 73.32, 72.14, 72.08, 71.36, 71.12, 71.10, 71.01, 70.87, 70.22, 69.87, 69.68, 69.03, 68.93, 68.63, 68.59, 68.07, 67.12, 65.11, 48.11, 28.66, 20.94, 20.90, 20.69, 20.64, 20.58; HR MALDI-TOF MS: m/z: calcd for C₆₆H₈₃N₃O₃₀ [M+Na]⁺: 1420.4959; found: 1420.4950. **78 β** : R_f = 0.12 (EtOAc/hexanes, 1/1, v/v); ¹H NMR (600 MHz, CDCl₃): δ 7.40 – 7.16 (m, 15H, ArH), 5.50 (t, $J = 9.0$, 1H, H-3^I), 5.44 – 5.37 (m, 3H, H-1^{II}, H-1^{III}, H-3^{II}), 5.40 (t, $J = 9.6$, 1H, H-3^{III}), 5.14 (t, $J = 9.3$ Hz, 1H, H-4^{III}), 5.10 (t, $J = 9.5$ Hz, 1H, H-3^{IV}), 4.99 – 4.91 (m, 2H, H-1^I, H-4^{IV}), 4.84 (dd, $J = 10.5, 4.0$ Hz, 1H, H-2^{III}), 4.80 (dd, $J = 10.2, 3.8$ Hz, 1H, H-2^I), 4.65 (dd, $J = 10.2, 3.8$ Hz, 1H, H-2^{II}), 4.59 – 4.45 (m, 5H, 2×CH₂Ph, CHHPh), 4.42 (d, $J = 7.9$ Hz, 1H, H-1^{IV}), 4.23 (d, $J = 12.0$ Hz, 1H, CHHPh), 4.12 (dd, $J = 11.5, 1.9$ Hz, 1H, H-6^{aI}), 4.07 – 3.96 (m, 4H, H-6^{bI}, H-4^{II}, H-5^{II}, H-4^I), 3.93 – 3.85 (m, 2H, H-5^I, H-5^{III}), 3.83 – 3.75 (m, 2H, CHH linker, H-6^{aII}), 3.72 (d, $J = 10.9$ Hz, 1H, H-6^{bII}), 3.67 – 3.60 (m, 1H, H-5^{IV}), 3.60 – 3.50 (m, 2H, H-2^{IV}, H-6^{aIV}), 3.47 – 3.38 (m, 3H, CH₂ linker, CHH linker), 3.37 (d, $J = 3.8$ Hz, 1H, H-6^{bIV}), 3.31 – 3.22 (m, 2H, H-6^{aIII}), 2.07 – 1.99 (7×s, 21H), 1.91 – 1.86 (2×s, 8H, CH₂ linker overlap); selected ¹³C NMR (150 MHz, gHSQC, CDCl₃): δ 102.46 (C-1^{IV}), 95.90 (C-1^I), 95.02 (C-1^{II}, C-1^{III}), 74.80 (C-3^{IV}), 73.42 (C-4^I), 73.32 (C-5^{IV}), 72.00 (C-3^{II}), 71.71 (C-3^I, C-2^{II}), 71.24 (C-4^{II}), 71.05 (C-2^I), 70.10 (C-2^{III}), 69.62 (C-3^{III}), 69.43 (C-4^{IV}, C-5^{II}, C-5^{III}), 69.96 (C-2^{IV}), 68.77 (C-4^{III}), 65.26 (CH₂), 48.46 (CH₂), 21.41 (CH₂); HR MALDI-TOF MS: m/z: calcd

for C₆₆H₈₃N₃O₃₀ [M+Na]⁺: 1420.4959; found: 1420.4962.

3-aminopropyl- α -D-glucopyranosyl-(1 \rightarrow 4)- α -D-glucopyranosyl-(1 \rightarrow 4)- α -D-glucopyranosyl-(1 \rightarrow 4)-6-O-(β -D-glucopyranosyl)- α -D-glucopyranoside (79). Freshly prepared NaOMe in a methanolic solution (0.2 mL, 1.5 M) was added to a stirred solution of **78a** (8 mg, 5.7 μ mol) in methanol (1.5 mL). The reaction mixture was stirred for 2 h and then neutralized by the addition of Dowex® 50W X8-200 H⁺ resin. The resin was removed by filtration and the filtrate was concentrated *in vacuo*. The residue was dissolved in a mixture of *t*BuOH (4 mL), H₂O (0.1 mL), and AcOH (0.1 mL) and a catalytic amount of 20 wt% Pd(OH)₂/C was added. The reaction mixture was purged with H₂ gas for 2 min followed by stirring for 6 h under and atmosphere of H₂. The progress of the reaction was monitored by MALDI-TOF mass spectrometer. Upon completion, the reaction mixture was purged with Ar gas followed by filtration through a plug of Celite. The filtrate was concentrated *in vacuo* to afford **79** (3.2 mg, 83% over two steps). ¹H NMR (600 MHz, D₂O): δ 5.25 (d, *J* = 3.9 Hz, 1H), 5.19 (d, *J* = 3.9 Hz, 1H), 4.82 (d, *J* = 3.6 Hz, 1H), 4.78 (d, *J* = 3.8 Hz, 1H), 3.87 – 3.20 (m, 26H), 3.11 – 2.94 (m, 2H), 1.91 – 1.81 (m, 2H); selected ¹³C NMR (150 MHz, gHSQC, CDCl₃): δ 99.80 (C-1), 98.44 (C-1), 98.02 (C-1), 78.14, 76.79, 73.66, 72.81, 72.39, 71.96 (C-2), 71.46 (C-2), 71.03, 69.51, 66.21 (CH₂, linker), 60.46 (C-6), 37.70 (CH₂, linker), 26.70 (CH₂, linker). HR MALDI-TOF MS: *m/z*: calcd for C₂₇H₄₉NO₂₁ [M+Na]⁺: 746.2695; found: 746.2683.

2. NMR analysis of sulfonium ion 55

Sulfonium ions were prepared according to general procedures. As an example, detailed analysis for 4-*S*-(2,3,5-trimethoxyphenyl)-2-(*S*)-phenyl-(3,4-di-acetyl-6-*O*-benzyl-1,2-

dideoxy- β -D-glucopyranoso)[1,2-*e*]-1,4-oxathianium triflate (**55**) was discussed here.

A mixture of *R/S* sulfoxide **17** (5.0 mg, 10 μ mol), 1,3,5-trimethoxybenzene (2.6 mg, 16 μ mol), 2,6-di-*tert*-butyl-4-methylpyridine (4.2 mg, 21 μ mol) and activated molecular sieves (4 \AA , pellets) in CDCl_3 (1 mL) was shaken for 30 min under an atmosphere of argon at room temperature. Then 0.5 mL of the solution was taken out and injected into a 5 mm NMR tube and sealed. After cooling to 0 $^\circ\text{C}$, trifluoromethanesulfonic anhydride stock solution (25 μL , 0.23 M in CDCl_3) was added. After 3 min, NMR spectra of the reaction mixture were recorded (^1H , gCOSY, gHSQC and HMBC were recorded at 25 $^\circ\text{C}$, **Fig. S1**); ^1H NMR (500 MHz, CDCl_3) δ 7.63 – 7.09 (m, *ArH*), 6.31 – 6.18 (m, 2H, *ArH*), 5.63 (d, $J = 9.7$ Hz, 1H, H-1), 5.45 (t, $J = 9.4$ Hz, 1H, H-3), 5.30 (d, $J = 10.7$ Hz, 1H, SCH_2CHPh), 5.22 (t, $J = 9.7$ Hz, 1H, H-4), 4.47 (d, $J = 11.8$ Hz, 1H, *CHHPh*), 4.38 (d, $J = 11.8$ Hz, 1H, *CHHPh*), 4.28 (t, $J = 11.6$ Hz, 1H, $\text{SCH}_{\text{ax}}\text{HCHPh}$), 4.14 – 3.96 (m, 8H, $\text{SCHH}_{\text{eq}}\text{CHPh}$, 2 \times OMe, H-2), 3.95 – 3.82 (m, 4H, OMe, H-5), 3.67 – 3.46 (m, 2H, H-6_{a,b}), 2.02 (s, 3H), 1.94 (s, 3H); selected ^{13}C NMR (200 MHz, gHSQC, CDCl_3): δ 81.50 (C-1), 79.38 (C-5), 78.02 (C-2), 76.74 (SCH_2CHPh), 73.62 (PhCH_2), 72.02 (C-3), 67.94 (C-4), 67.62 (C-6), 44.33 (SCH_2CHPh). The ^1H NMR spectrum showed that two sets of peaks corresponding to the mixture of *R/S* sulfoxide converted into a single set of peaks. Specifically, the anomeric proton (H-1) signal of *R*-sulfoxide ($\delta = 4.31$, d, $J = 10$ Hz) and *S*-sulfoxide ($\delta = 4.14$, d, $J = 9.7$ Hz) shifted downfield ($\delta = 5.63$, d, $J = 9.7$ Hz). The retaining of β anomeric configuration and the shift of H-1 chemical shift indicated that the sulfoxide donors had completely transformed to a new intermediate within 3 min after activation. The rest of the proton signals were assigned from 2D spectra. The HMBC spectrum indicated the presence of three-bond coupling between C-1 and H-8_{eq}, which

confirmed the formation of *trans*-decalin sulfonium ion as the reactive intermediate.

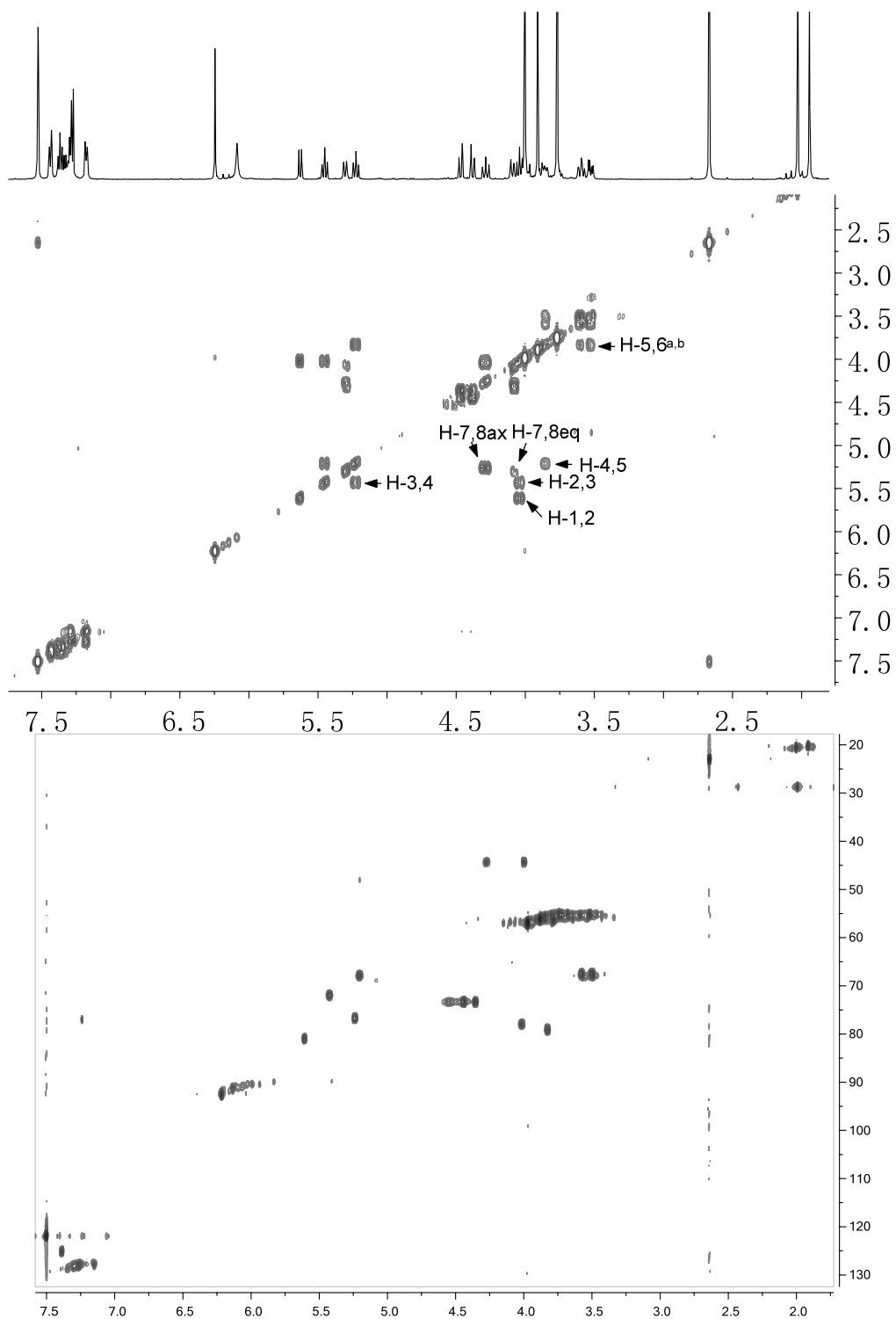


Figure S1. ¹H, gCOSY, gHSQC spectra of sulfonium ion **55**.

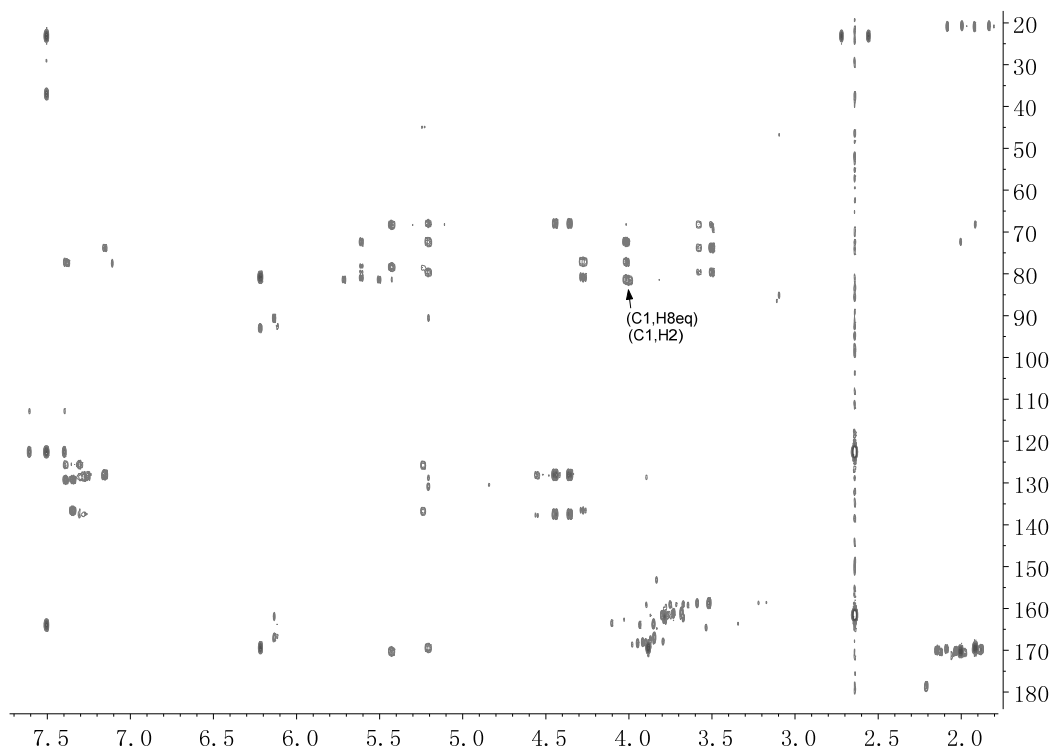


Figure S1 (cont'd). HMBC spectrum of sulfonium ion **55**.

Since sulfonium ion was proposed to be the reactive intermediate and responsible for the high stereoselectivity in glycosylations, its stability therefore is of great interest for the purpose of optimizing reaction conditions. So, the above obtained sulfonium ion **55** was continually monitored by NMR at room temperature (**Fig. S2**). Surprisingly, **55** was stable at this temperature for at least 9 h. During this period of time, the intensity of characteristic sugar peaks remained unchanged while aromatic signal ($\delta = 6.08$) corresponding to trimethoxybenzene decreased. Subsequent heating to 45 °C in 5 min resulted prompt decomposition of the sulfonium ion. The temperature sensitivity of the sulfonium ion has also been observed when the glycosylation reaction was heated and resulted in low yield and poor stereoselectivity.

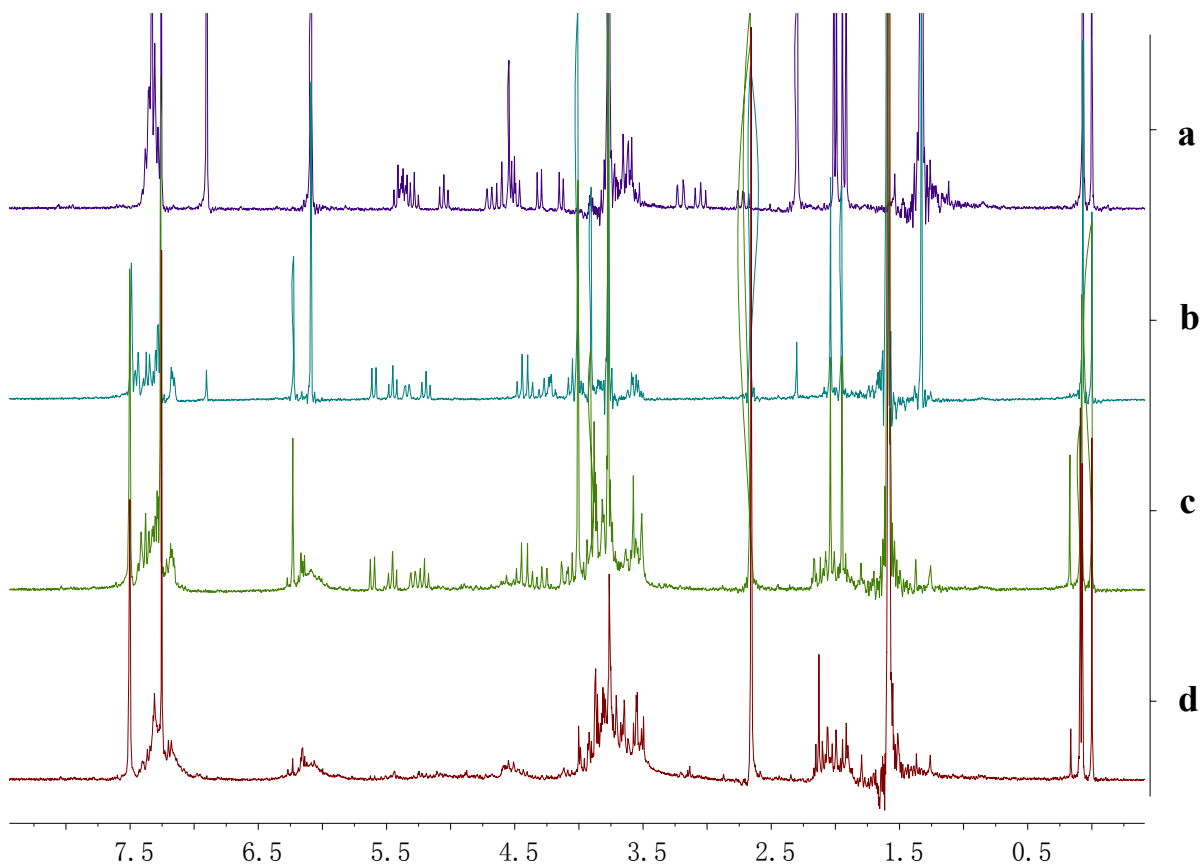


Figure S2. Thermostability of sulfonium ion **55**. a) ^1H spectrum of a mixture of *R/S* sulfoxide with 1,3,5-trimethoxybenzene and DTBMP; b) ^1H spectrum of the activated reaction mixture after 3 min at 25 °C; c) ^1H spectrum of the activated reaction mixture after 9 h at 25 °C; d) ^1H spectrum recorded after raising to 45 °C in 5 min.

3. Assignment and conformational analysis of diastereomeric sulfonium ions

Various criteria have been applied to the assignment of diastereomeric sulfur substituents in a six-member ring system^[2]. Usually employed methods include the comparison of the midpoint of chemical shifts for δ_{H8eq} and δ_{H8ax} or the geminal coupling constant for the AB quartet of H8eq and H8ax (**Fig. S3**). Because the substituents on heteroatoms or

elsewhere on the ring alter, the chemical shift criteria can be overridden. But coupling constant criterion has been proven to hold true regardless of the substitution in the ring^[3]. Assignments have also been made for similar oxathiane sulfonium ion systems^[4-6]. Generally, the equatorial diastereomer with axial lone pair has a smaller geminal coupling constant (~12 Hz) than that of the axial diastereomer with equatorial lone pair (~15 Hz). This trend has also been tested and proven to be effective for similar systems such as sulfoxide donors **17-23**, where both diastereomers are available. Therefore, similar rule was applied to the assignment of distereomeric sulfonium ions **55-61** (**Table S1**).

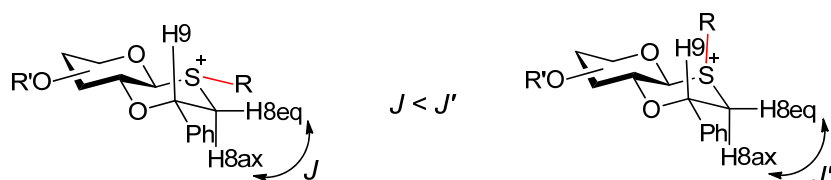


Figure S3. Schematic presentation for the interpretation of stereochemistry at sulfur atom using geminal coupling constant of H8eq and H8ax.

Table S1. List of δ_{H8eq} , δ_{H8ax} and $J_{eq,ax}$

Entry	Sulfonium Ion ^[a]	δ_{H8eq} (ppm) doublet	δ_{H8ax} (ppm) triplet	$J_{eq,ax}$ (Hz)	
1		4.19	4.35	10.0	
2		4.13	4.26	12.5	
3		3.97	4.36	11.8	δ_{Heq} < δ_{Hax}
4		3.98	4.28	11.6	
5		4.27	3.72	11.5	
6		4.16	3.63	12.0	δ_{Heq} > δ_{Hax}
7		4.07	3.88	10.5	
8 ^[6]		4.32	3.66	11.0	

Once the stereochemistry at sulfur atom was identified, we could make sure we were comparing sulfonium ions with the same stereochemistry. Then the assignment of the rest of the proton signals can be taken out. Due to the large coupling of H8ax-H9 and small coupling of H8eq-H9, they are showing characteristic splittings that give a triplet and a doublet respectively. Therefore, both H8ax and H8eq can be easily assigned on ^1H spectra or gHSQC spectra if there are overlaps of peaks in 1D experiment.

As shown in **Table S1**, there is a clear trend that the relative proton chemical shifts (H8ax vs. H8eq) of trimethoxybenzene substituted sulfonium ions reversed when compared to methyl or the phenyl substituted sulfonium ions. This large relative chemical shift change should not be introduced by merely the inductive effect from the changing of sulfur substitutions, since H8 is three bonds away from the substitution and there is no dramatic change of the electronegativity of substitutions. However, one obvious difference between phenyl and trimethoxybenzene is their steric effect, which may change the orientation of aromatic substitutions and therefore change the orientation of shielding/deshielding environment ^[7] (**Figure S4**). So, the different relative chemical shifts of H8ax and H8eq may indicate different orientations of sulfur substitutions in space, which may contribute to the slight difference when comparing phenyl and trimethoxybenzene substituted systems in regard of stereoselectivity in glycosylations.

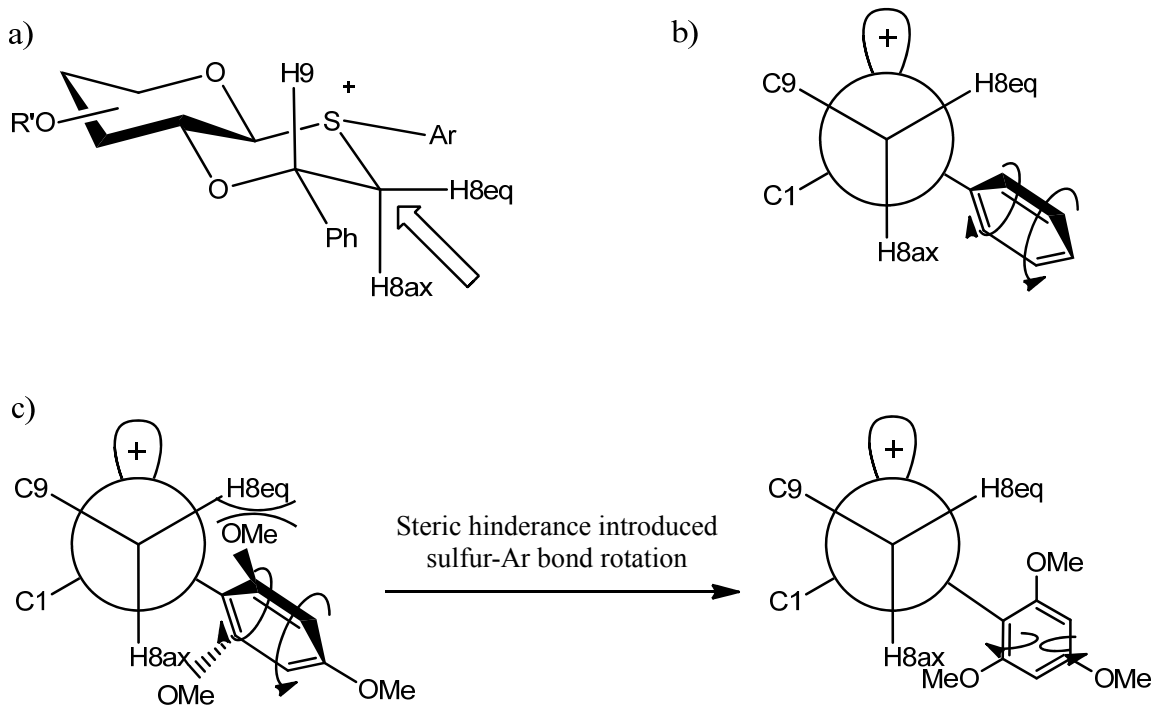
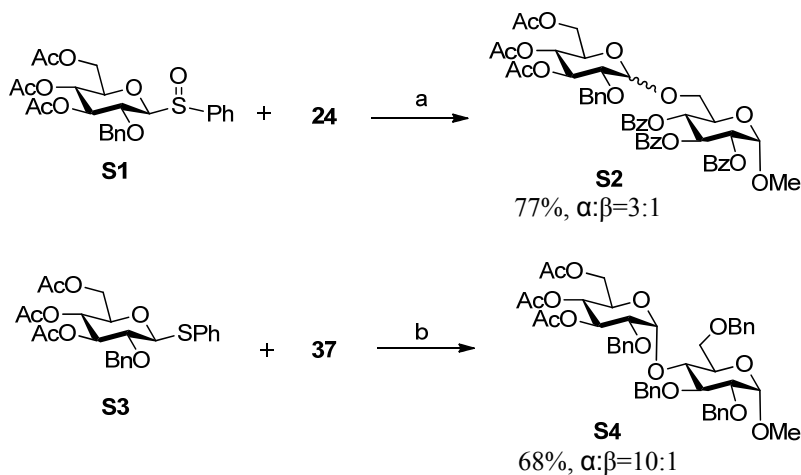


Figure S4. Schematic presentation of Sulfur-Ar bond rotation and shielding/deshielding changes for H8eq and H8ax in phenyl and trimethoxybenzene substituted sulfonium ions. a) Sight angle for the Newman projections; b) Proposed orientation of phenyl group, H8ax is located in the shielded space and H8eq is located in the deshielded space; c) Due to the steric hinderance, the favored coordination for trimethoxybenzene changed making the shielded space rotated toward H8eq.

4. Control glycosylations using donors without C-2 auxiliary

Scheme S2. Glycosylations using donors without C-2 participation



Reagents and conditions: a) TMSOTf, TEP, -60 °C to 0 °C, 2 h^[8]; b) NIS, TfOH, 0 °C.

Methyl 3,4,6-tri-O-acetyl-2-O-benzyl-D-glucopyranosyl-(1→6)- 2,3,4-tri-O-benzoyl- α -D-glucopyranoside (S2). A mixture of glycosyl donor **S1**^[5] (60 mg, 0.12 mmol), acceptor **24** (30 mg, 0.059 mmol) and activated molecular sieves (4 Å) in DCM (3 mL) was stirred for 60 min under an atmosphere of argon at room temperature. After cooling to -60 °C, triethyl phosphite (TEP)^[9] (32 μ L, 0.20 mmol) and TMSOTf (22 μ L, 0.10 mmol) were added and the reaction mixture was allowed to warm to -30 °C over a period of 30 min. TLC showed donor was not fully consumed. Another 22 μ L TMSOTf was added at -30 °C. Then the reaction mixture was allowed to warm to 0 °C over 2 h. After diluting with DCM (10 mL), aqueous saturated NaHCO₃ (10 mL) was added and the organic phase was dried (MgSO₄), filtered and the filtrate was concentrated *in vacuo*. The residue was purified by sephadex® LH20 size exclusion chromatography (DCM/MeOH, 1/1) to afford the pure disaccharide **S2** (40 mg, 77%, $\alpha:\beta=3:1$). $R_f = 0.17$ (EtOAc/hexanes,

1/2, v/v); **S2a**: ^1H NMR (500 MHz, CDCl_3): δ 8.08 – 7.05 (m, 20H, ArH), 6.17 (t, $J = 9.4$ Hz, 1H, H-3^I), 5.53 – 5.38 (m, 2H, H-5^{II}, H-4^I), 5.24– 5.21 (m, 2H, H-1^I, H-2^I), 4.95 (t, $J = 9.4$ Hz, 1H, H-4^{II}), 4.74 (d, $J = 3.5$ Hz, 1H, H-1^{II}), 4.65 (d, $J = 12.5$ Hz, 1H, CHHPh), 4.55 (d, $J = 12.5$ Hz, 1H, CHHPh), 4.43 – 4.29 (m, 1H, H-5^I), 4.19 (q, $J = 4.2$ Hz, 2H, H-5^{II}, H-6a^{II}), 4.02 (dd, $J = 14.2, 4.3$ Hz, 1H, H-6b^{II}), 3.83 (dd, $J = 10.7, 7.6$ Hz, 1H, H-6a^I), 3.60 – 3.46 (m, 5H, H-2^{II}, H-6b^I, OMe), 2.07 (s, 3H), 2.01 (s, 3H), 1.99 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3): δ 184.54, 175.60, 175.00, 174.83, 170.77, 170.67, 170.31, 142.70, 138.50, 138.30, 138.03, 134.87, 134.84, 134.60, 134.12, 133.97, 133.72, 133.43, 133.41, 133.35, 133.21, 132.95, 132.76, 101.60, 101.50, 82.94, 82.16, 81.95, 81.83, 81.74, 78.10, 77.12, 76.63, 75.33, 74.58, 73.55, 73.49, 72.29, 71.69, 66.94, 60.54, 25.79, 25.70, 25.63; HR MALDI-TOF MS: m/z: calcd for $\text{C}_{47}\text{H}_{48}\text{O}_{17}$ $[\text{M}+\text{Na}]^+$: 907.2789; found: 907.2799. The β anomer was purified by reversed phase HPLC on an analytical C-18 column using a gradient of 50→100% acetonitrile in H_2O over 40 min. **S2B**: ^1H NMR (500 MHz, CDCl_3): δ 8.04 – 7.21 (m, 20H, ArH), 6.16 (t, $J = 9.8$ Hz, 1H, H-3^I), 5.45 (t, $J = 9.9$ Hz, 1H, H-4^I), 5.29 – 5.20 (m, 2H, H-2^I, H-1^I), 5.14 (t, $J = 9.5$ Hz, 1H, H-3^{II}), 4.97 – 4.88 (m, 2H, H-4^{II}, CHHPh), 4.59 (dd, $J = 15.2, 9.8$ Hz, 2H, CHHPh, H-1^{II}), 4.37 (t, $J = 7.9$ Hz, 1H, H-5^I), 4.23 (dd, $J = 12.2, 5.2$ Hz, 1H, H-6a^{II}), 4.05 (t, $J = 13.3$ Hz, 2H, H-6b^{II}, H-6a^I), 3.86 (dd, $J = 11.2, 7.5$ Hz, 1H, H-6b^I), 3.73 – 3.59 (m, 1H, H-5^{II}), 3.42 (s, 4H, H-2^{II}, OMe), 2.00 (s, 3H), 1.99 (s, 3H), 1.90 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3): δ 184.48, 184.45, 184.43, 184.42, 175.60, 175.05, 174.64, 170.75, 170.69, 170.39, 142.93, 138.48, 138.32, 138.04, 134.85, 134.79, 134.57, 134.10, 133.94, 133.90, 133.75, 133.56, 133.42, 133.35, 133.31, 133.21, 132.93, 132.71, 118.04, 108.77, 101.95, 101.83, 83.60, 82.96, 82.94, 82.94, 82.16, 81.95, 81.73, 79.33, 78.66, 76.94, 76.50, 75.26, 74.75, 74.01, 73.97,

73.57, 67.06, 60.59, 34.64, 25.63, 25.58; HR MALDI-TOF MS: m/z : calcd for $C_{47}H_{48}O_{17}$ $[M+Na]^+$: 907.2789; found: 907.2781.

Methyl 3,4,6-tri-*O*-acetyl-2-*O*-benzyl- α -D-glucopyranosyl-(1 \rightarrow 4)- 2,3,6-tri-*O*-benzyl- α -D-glucopyranoside (S4). A mixture of glycosyl donor **S3**^[5] (60 mg, 0.12 mmol), acceptor **37** (48 mg, 0.10 mmol) and activated molecular sieves (4 Å) in DCM (2 mL) was stirred for 60 min under an atmosphere of argon at room temperature. After cooling to 0 °C, NIS (42 mg, 0.31 mmol) was added followed by the addition of TfOH (4 μ L, 0.056 mmol). The reaction mixture was stirred for 2 h at 0 °C before quenching by the addition of pyridine (0.1 mL) and diluting with DCM (10 mL). The reaction mixture was filtered, and the filtrate was washed with 10% $Na_2S_2O_3$ (20 mL) and brine (50 mL). The organic phase was dried ($MgSO_4$), filtered and the filtrate was concentrated under reduced pressure. The residue was purified by sephadex® LH20 size exclusion chromatography (DCM/MeOH, 1/1) to afford the pure disaccharide **S4** (73 mg, 68%). R_f = 0.16 (EtOAc/hexanes, 1/2, v/v); 1H NMR (600 MHz, $CDCl_3$): δ 7.41 – 7.01 (m, 20H, ArH), 5.69 (d, J = 3.7 Hz, 1H, H-1^{II}), 5.41 (t, J = 9.7 Hz, 1H, H-3^{II}), 5.03 (d, J = 11.8 Hz, 1H, CHHPh), 4.93 (t, J = 9.7 Hz, 1H, H-4^{II}), 4.75 (d, J = 11.8 Hz, 1H, CHHPh), 4.68 (d, J = 12.1 Hz, 1H, CHHPh), 4.65 – 4.53 (m, 4H, H-1^I, CHHPh, CH_2 Ph), 4.44 (d, J = 12.2 Hz, 1H, CHHPh), 4.33 (d, J = 12.2 Hz, 1H, CHHPh), 4.12 – 4.04 (m, 2H, H-3^I, H-6^a^{II}), 4.04 – 3.97 (m, 2H, H-5^{II}, H-4^I), 3.92 – 3.84 (m, 2H, H-5^I, H-6^a^I), 3.80 (dd, J = 12.4, 2.1 Hz, 1H, H-6^b^{II}), 3.67 (dd, J = 10.9, 1.6 Hz, 1H, H-6^b^{II}), 3.58 (dd, J = 9.4, 3.5 Hz, 1H, H-2^I), 3.47 (dd, J = 10.1, 3.7 Hz, 1H, H-2^{II}), 3.39 (s, 3H, OMe), 2.00 (s, 3H), 1.99 (s, 3H), 1.92 (s, 3H); ^{13}C NMR (150 MHz, $CDCl_3$): δ 170.51, 170.02, 169.66, 139.05, 137.92,

137.88, 137.50, 128.56, 128.50, 128.43, 128.37, 128.33, 128.31, 128.20, 128.14, 127.93, 127.76, 127.63, 127.52, 127.39, 127.05, 126.58, 97.70, 96.39, 81.59, 80.13, 77.31, 77.09, 76.88, 76.60, 74.27, 73.47, 73.44, 73.26, 72.93, 71.72, 69.47, 68.93, 68.48, 67.72, 61.81, 55.23, 20.78, 20.72, 20.68; HR MALDI-TOF MS: m/z: calcd for C₄₇H₅₄O₁₄ [M+Na]⁺: 865.3412; found: 865.3403.

5. References and notes

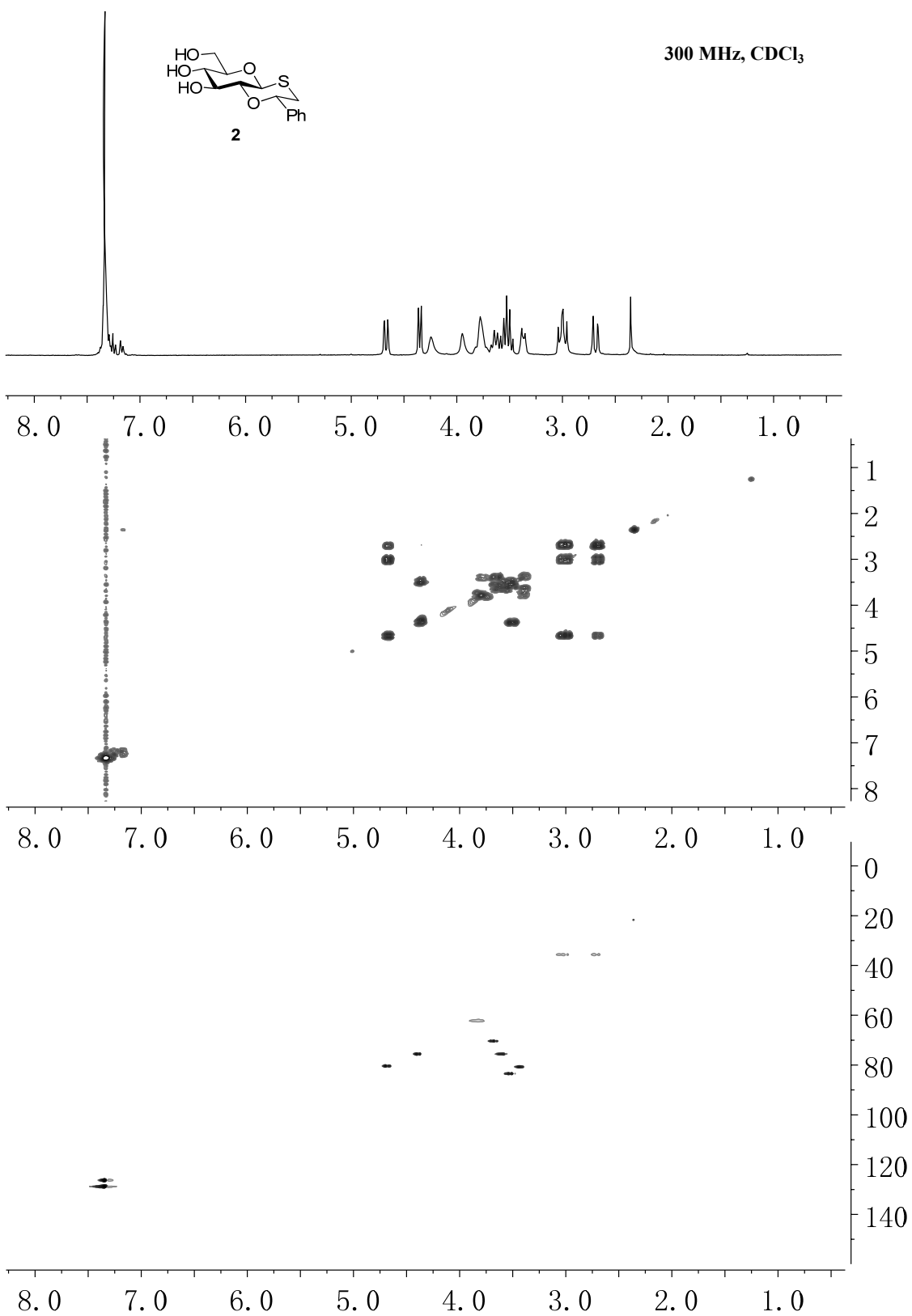
1. Fascione, M. A.; Adshead, S. J.; Stalford, S. A.; Kilner, C. A.; Leach, A. G.; Turnbull, W. B. *Chem. Commun.* **2009**, 5841-5843.
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5. Mydock, L. K.; Kamat, M. N.; Demchenko, A. V. *Org. Lett.* **2011**, *13*, 2928–2931.
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9. Note: Initially, Tf₂O/DTBMP was employed as activator but the use of this reagent did not provide the desired coupling product probably due to sulfenylation as a major side reaction. Lewis acid alone was also examined as glycosylation promoters but the product yields were low probably due to the *in-situ* dimerization of generated sulfenic acid.

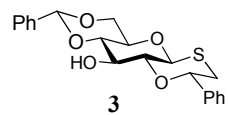
However, a combination of TMSOTf and TEP gave good yields of product. Probably, TEP functions as an effective acid scavenger and quencher of ionic species generated by Lewis acid activated phenylsulfenyl trimethylsilyl ester.

For more information, please see:

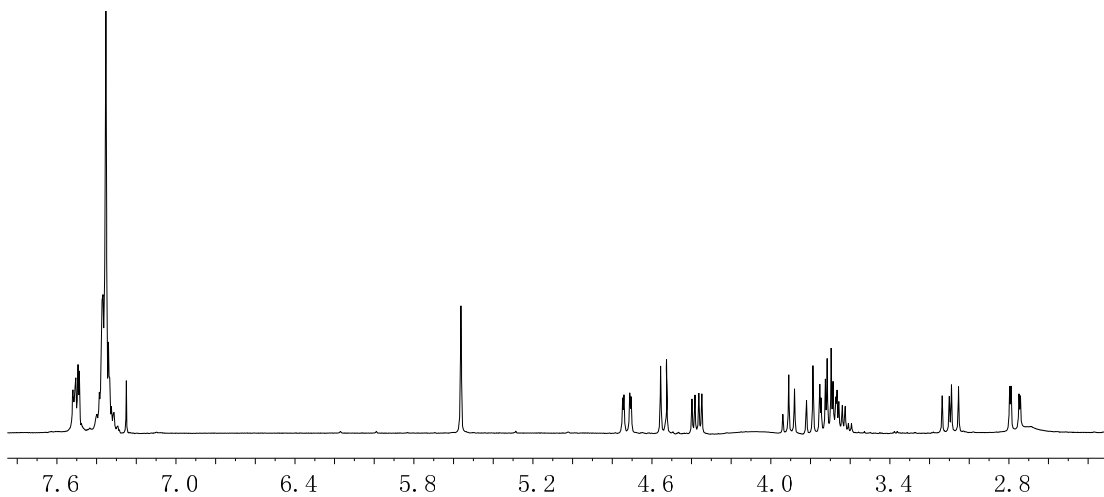
- a) Ito, Y.; Ogawa, T. *Tetrahedron Lett.* **1987**, 28, 4701-4704.
- b) Alonso, I.; Khier, N.; Martin-Lomas, M. *Tetrahedron Lett.* **1996**, 37, 1477-1480.
- c) Gildersleeve, J.; Smith, A.; Sakurai, K.; Raghavan, S.; Kahne, D. *J. Am. Chem. Soc.* **1999**, 121, 6167– 6175.

6. Copies of ^1H and ^{13}C NMR spectra

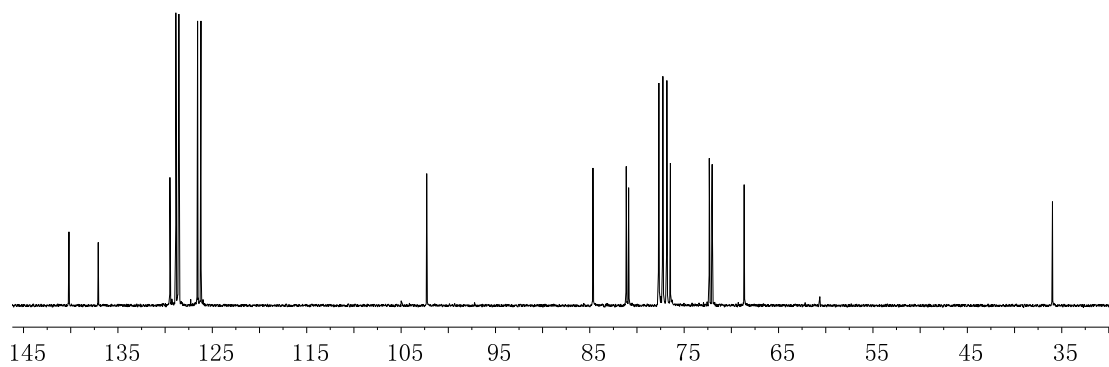


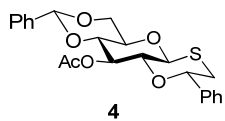


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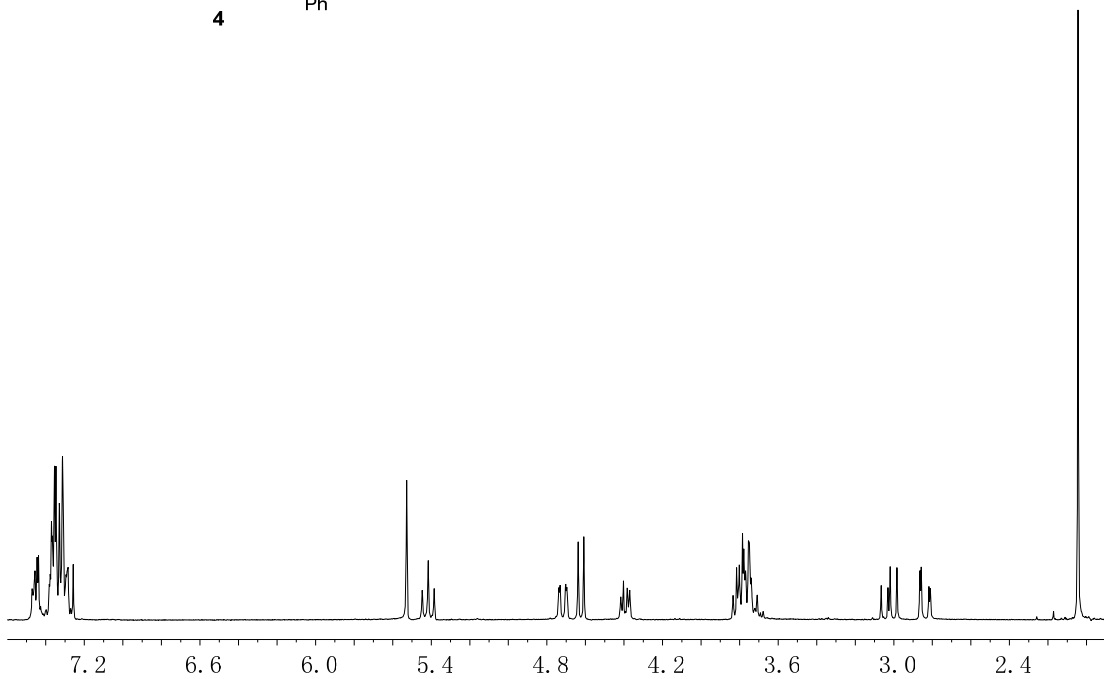


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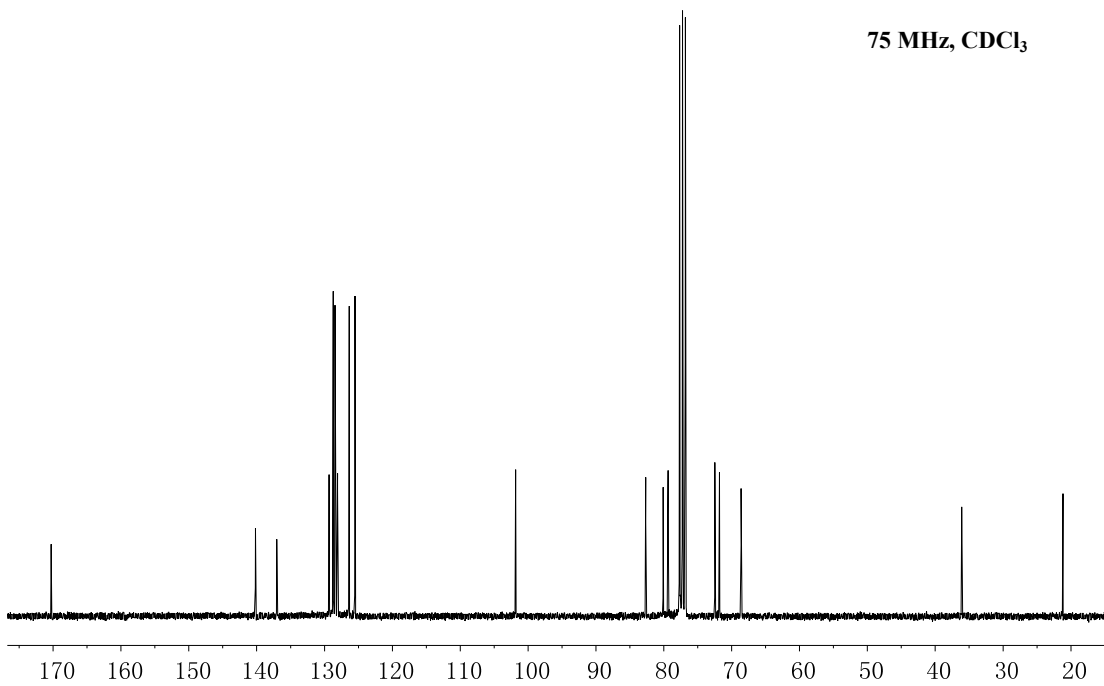


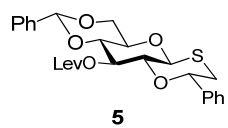


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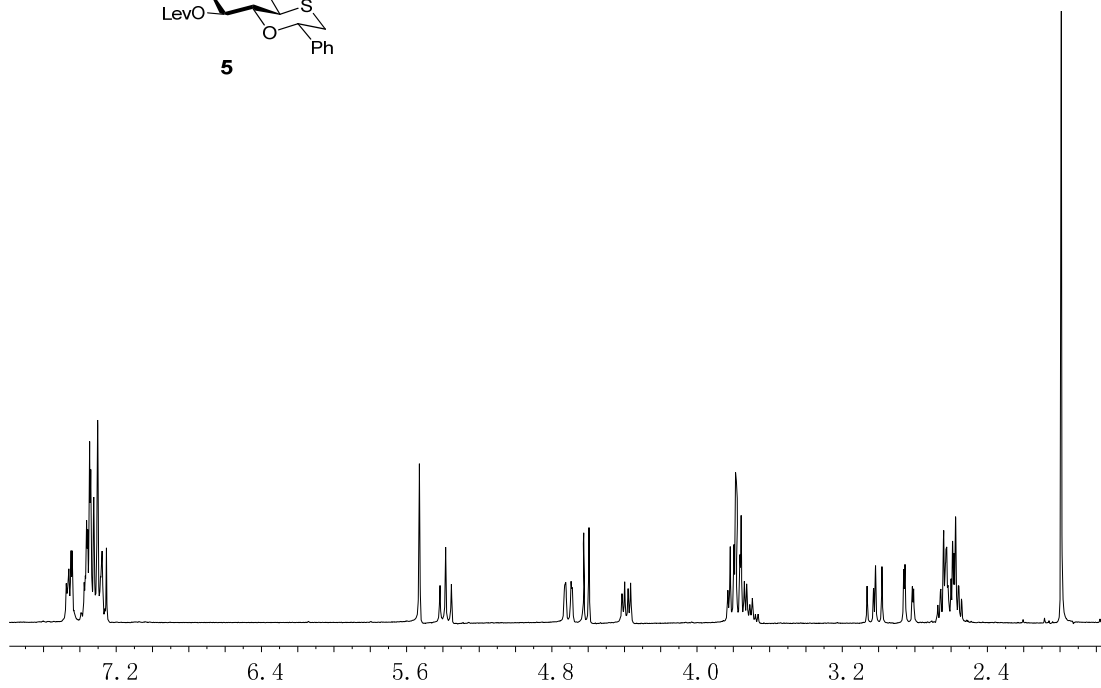


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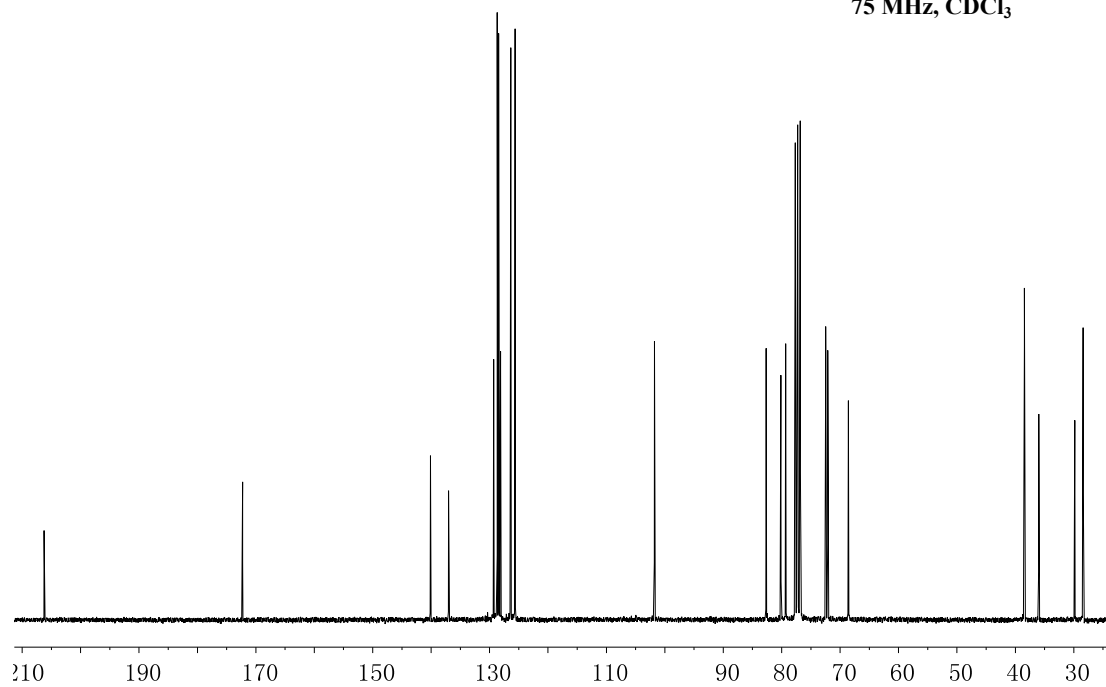


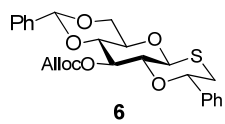


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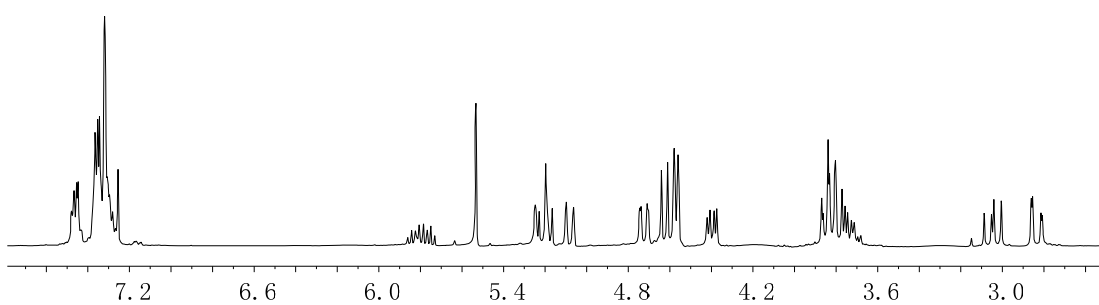


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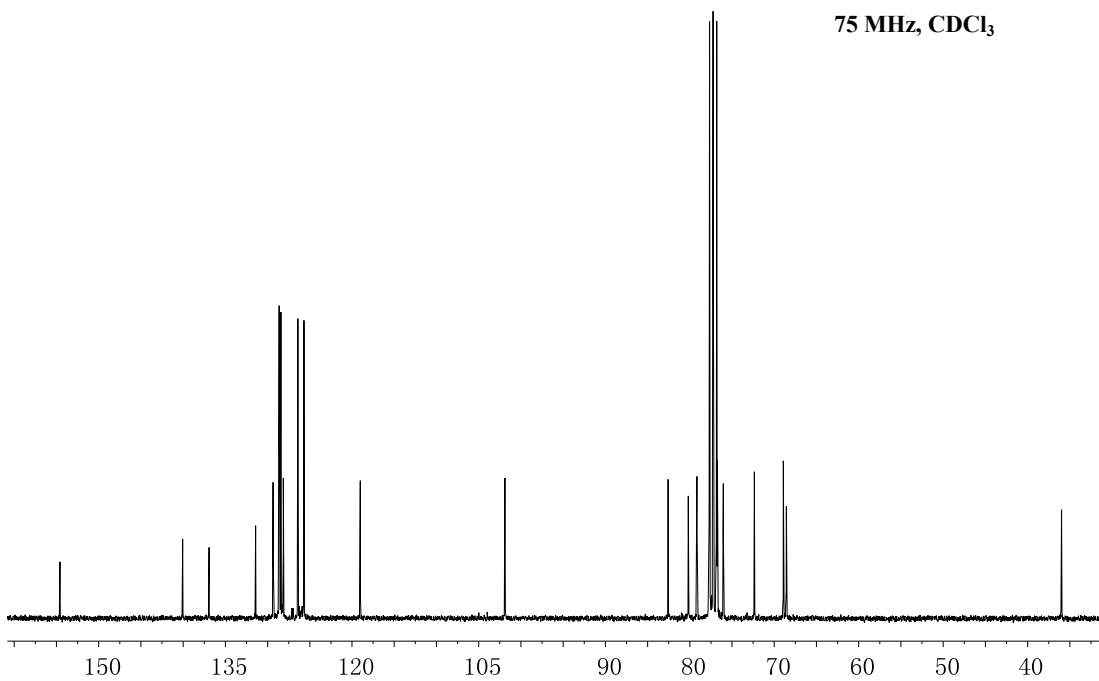


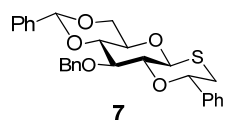


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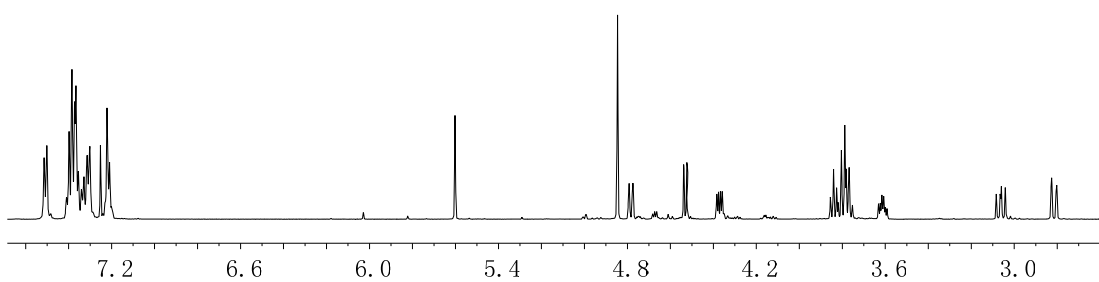


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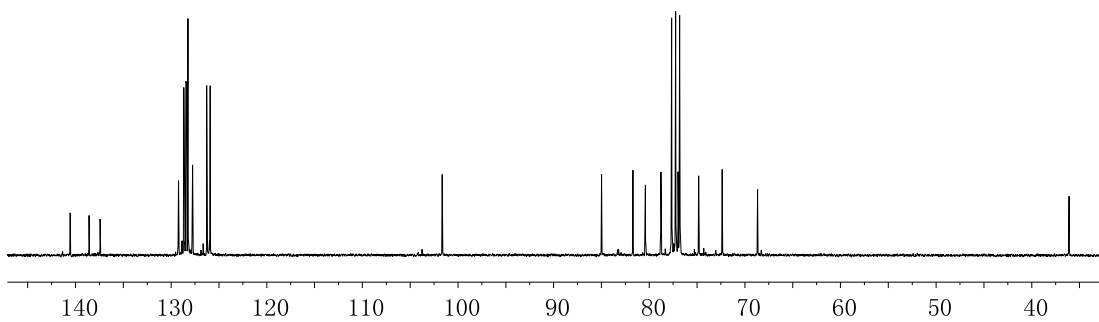


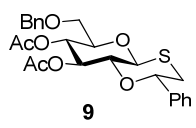


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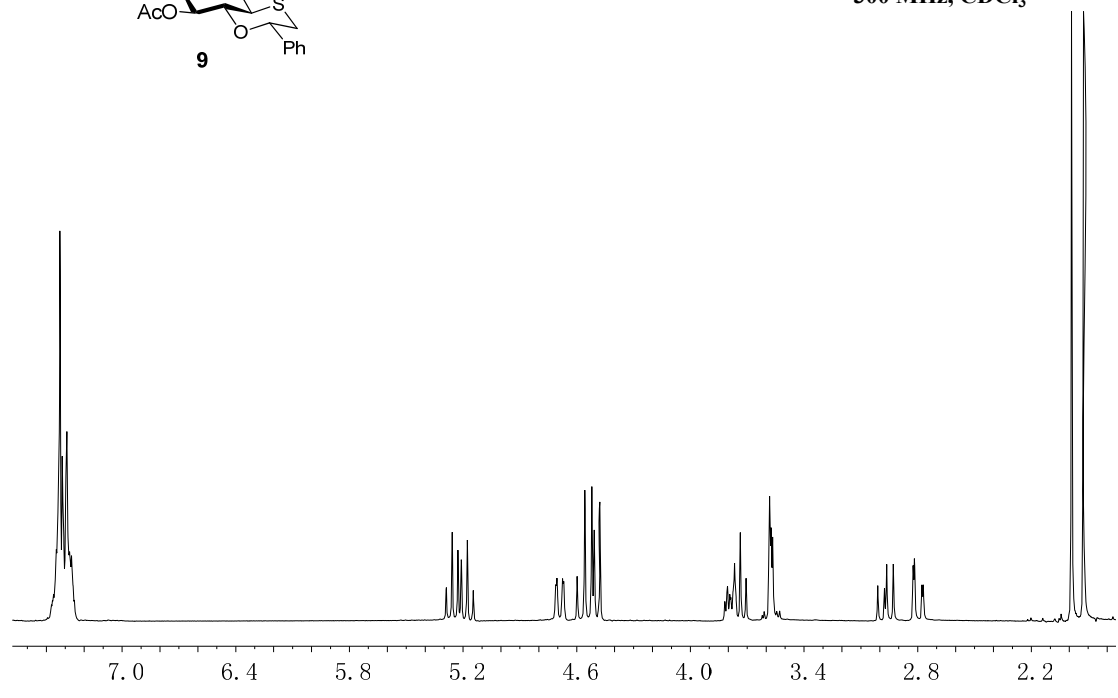


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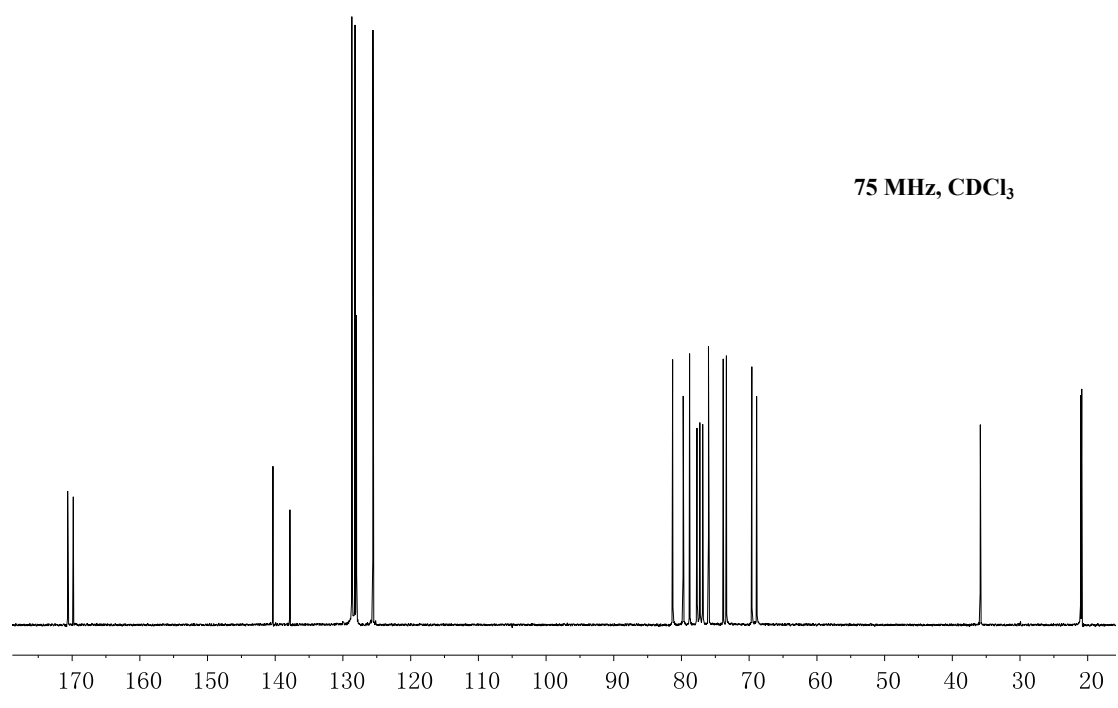


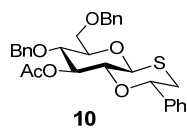


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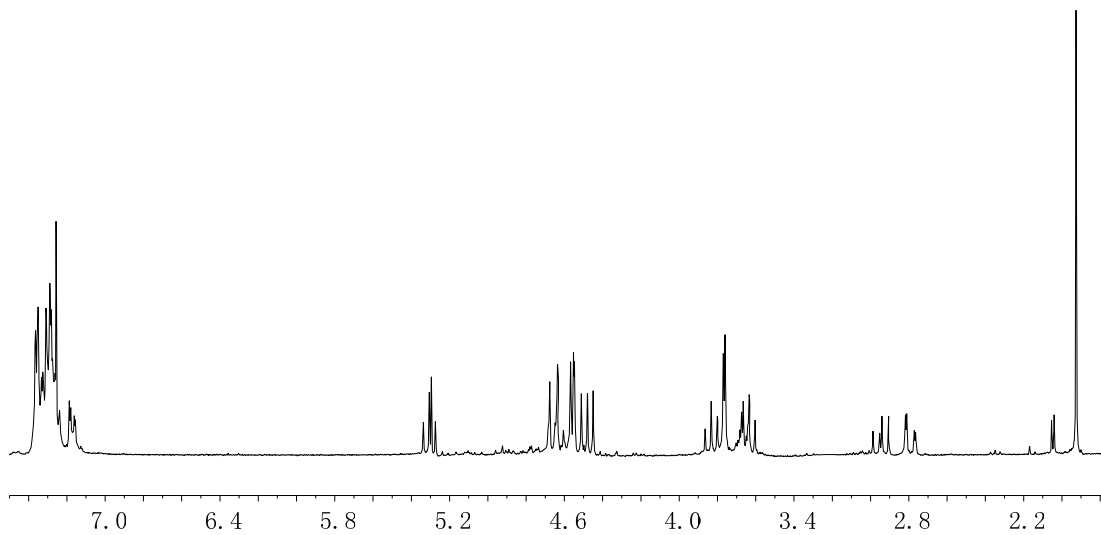


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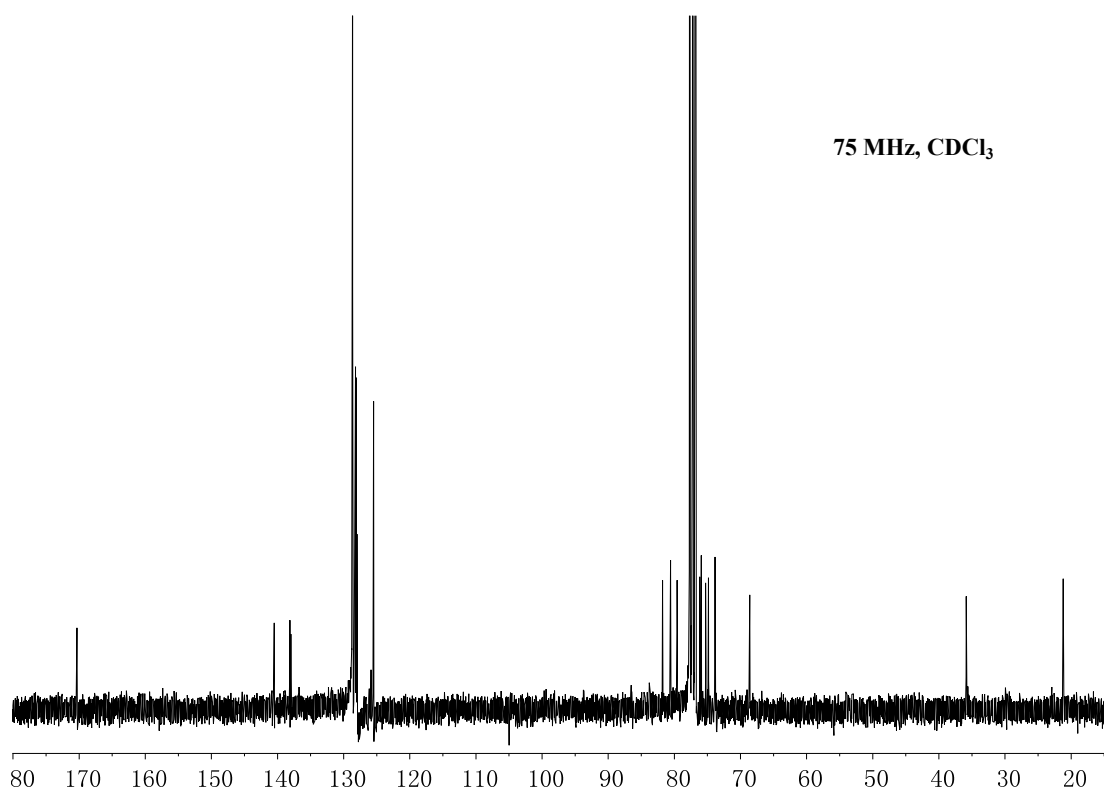


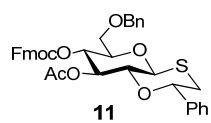


300 MHz, CDCl₃

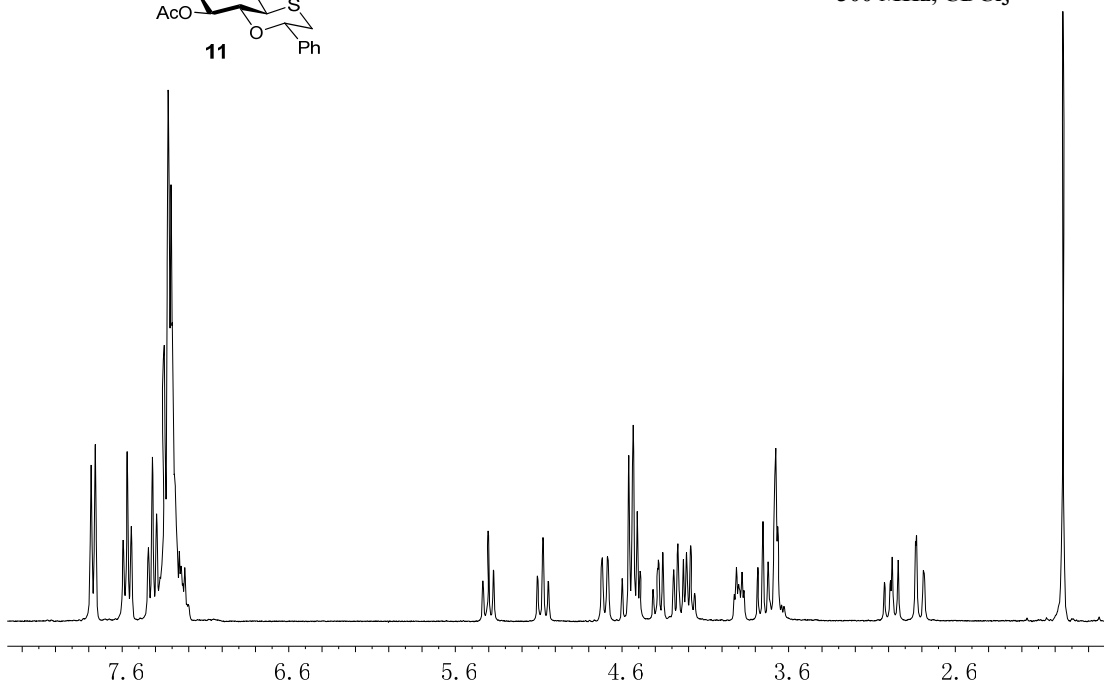


75 MHz, CDCl₃

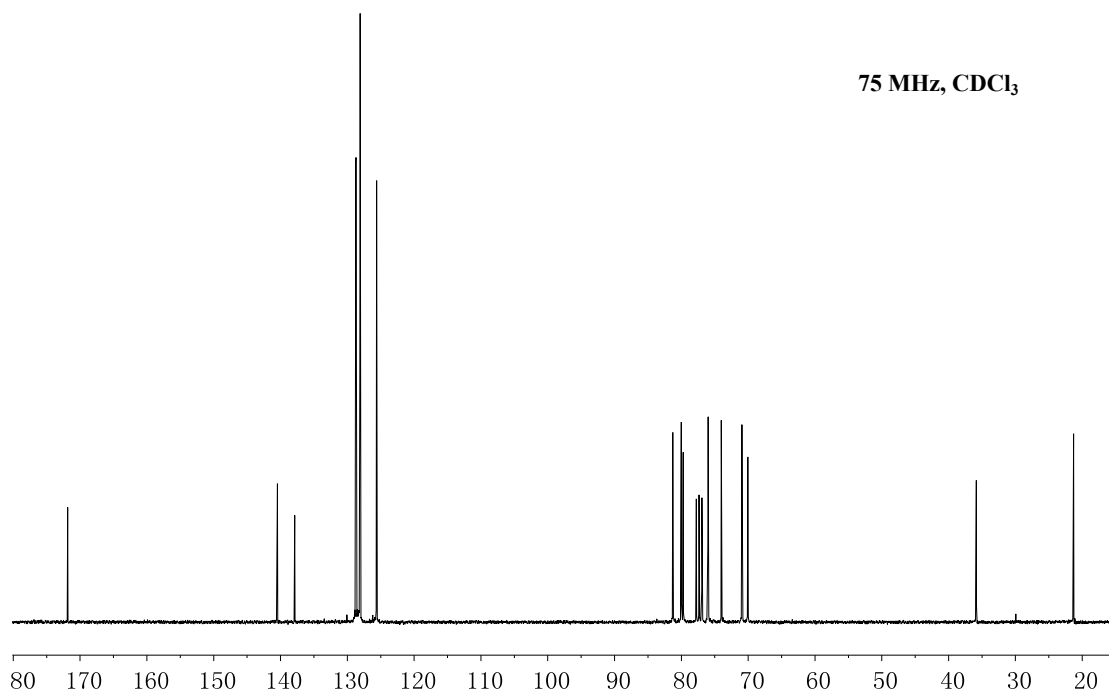


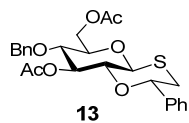


300 MHz, CDCl₃

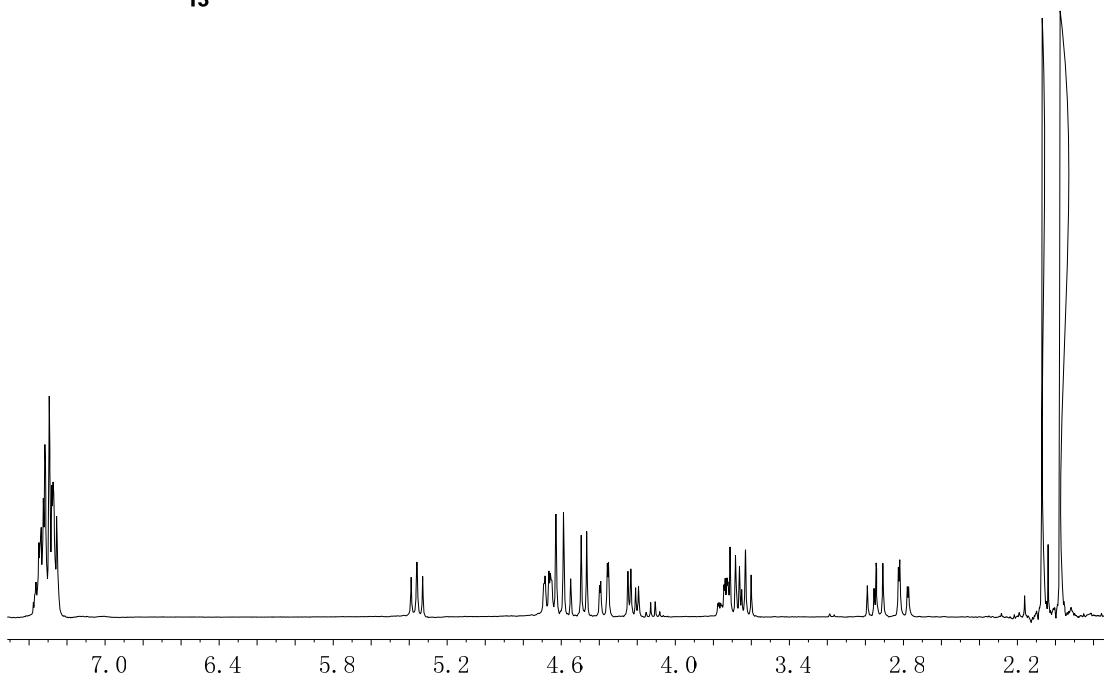


75 MHz, CDCl₃

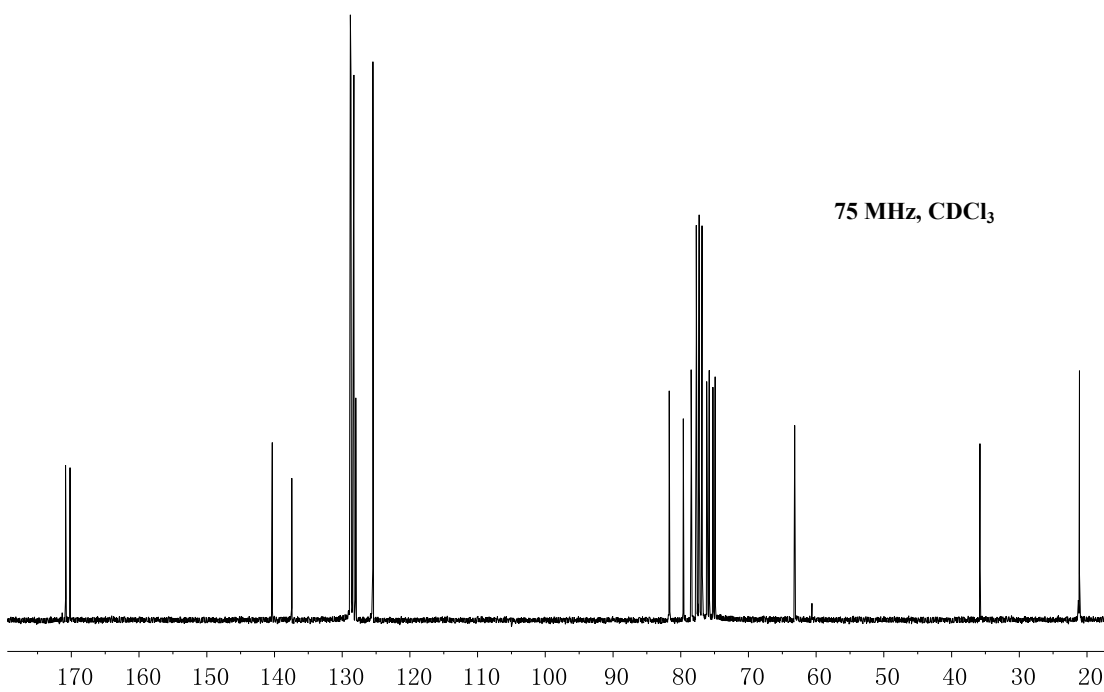


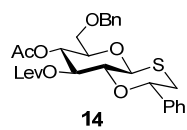


300 MHz, CDCl₃

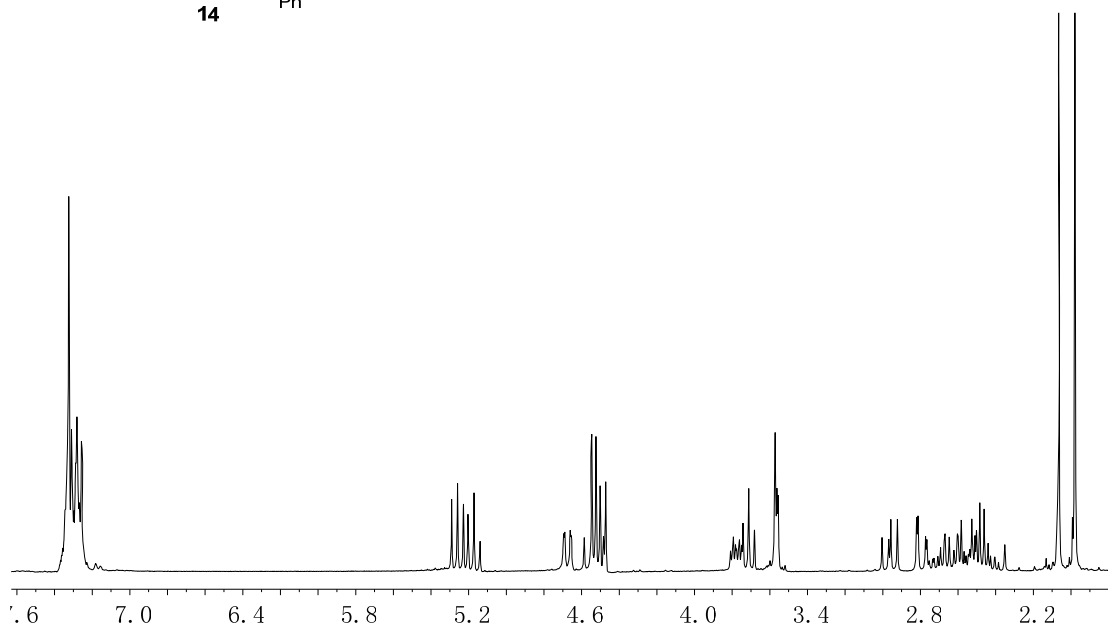


75 MHz, CDCl₃

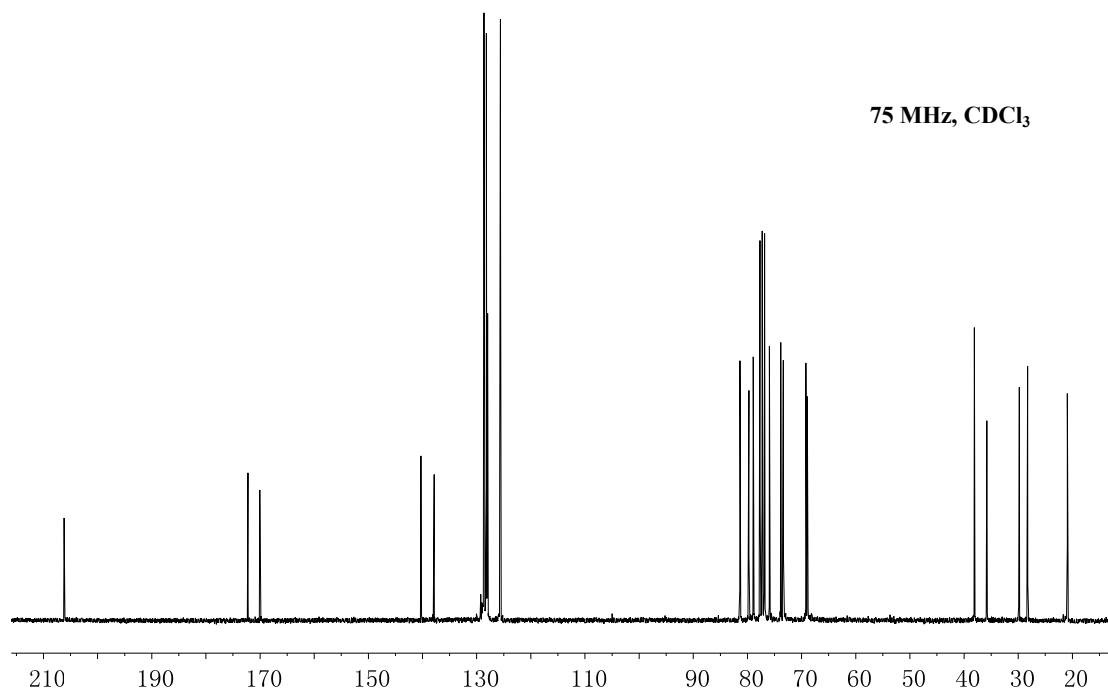


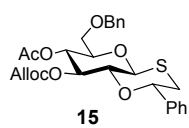


300 MHz, CDCl₃

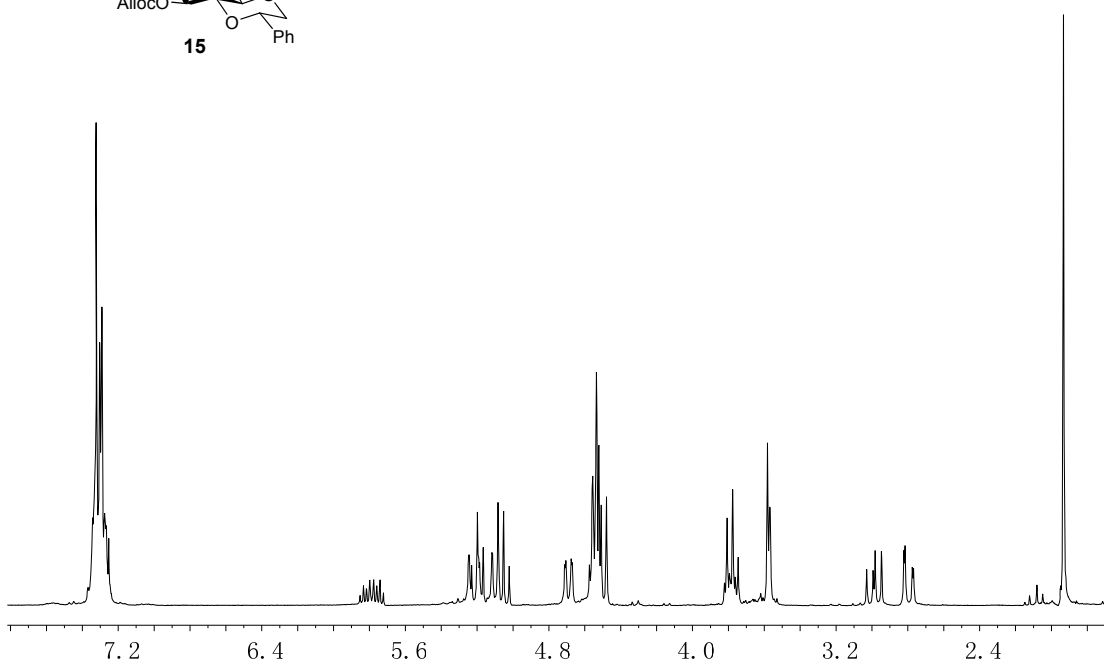


75 MHz, CDCl₃

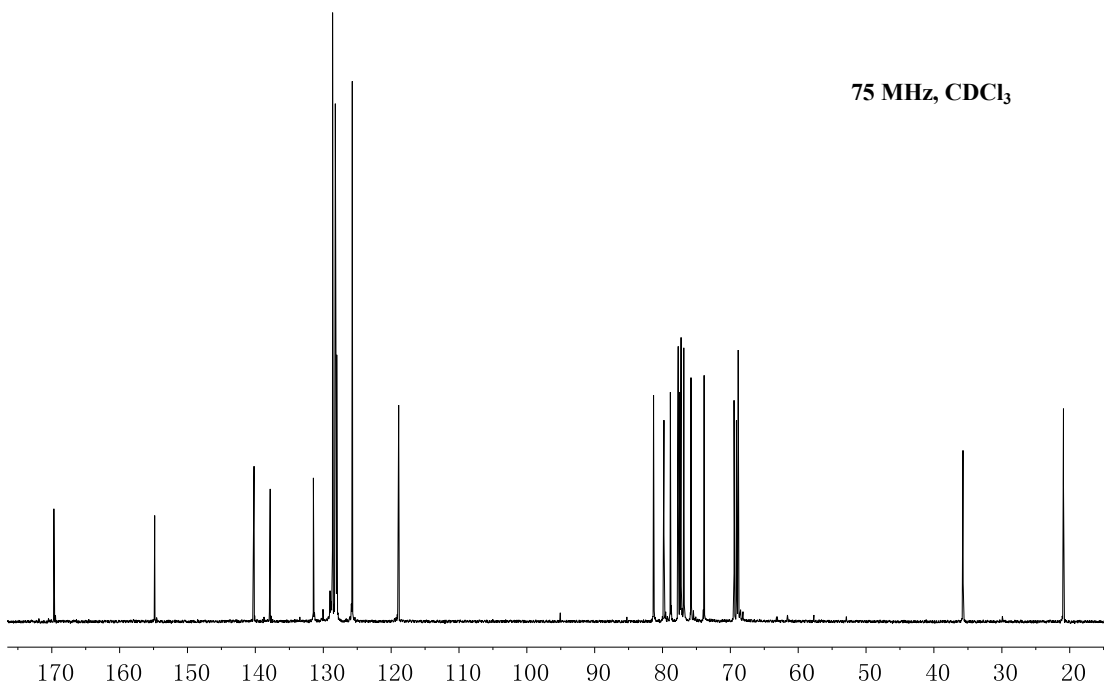


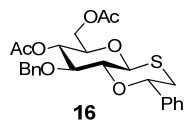


300 MHz, CDCl₃

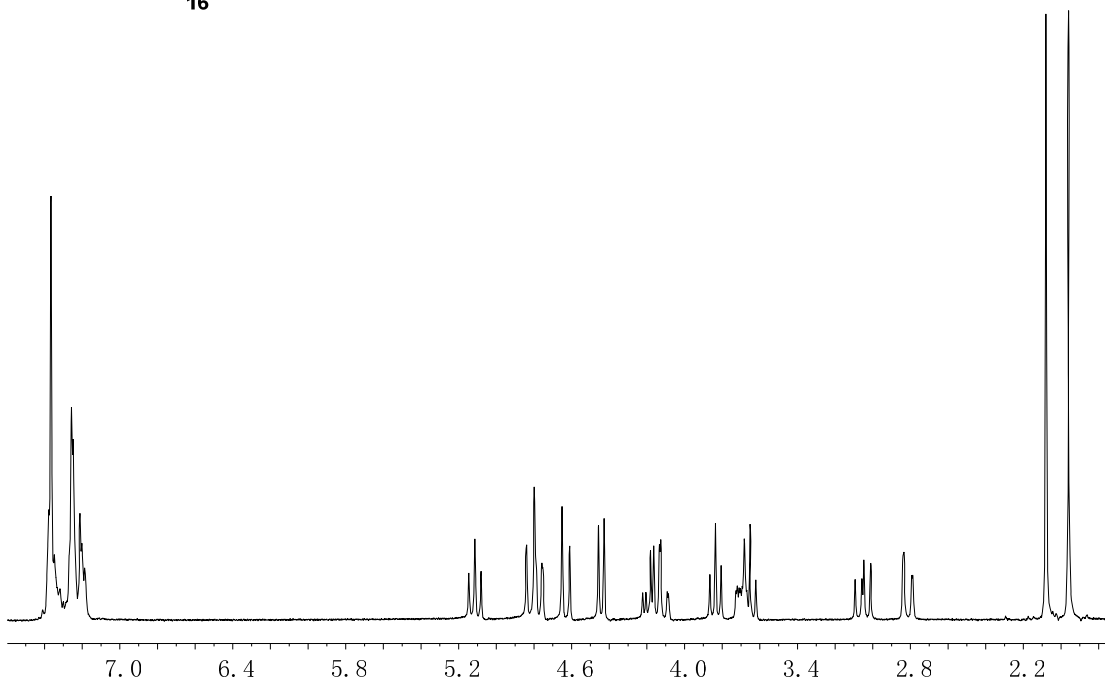


75 MHz, CDCl₃

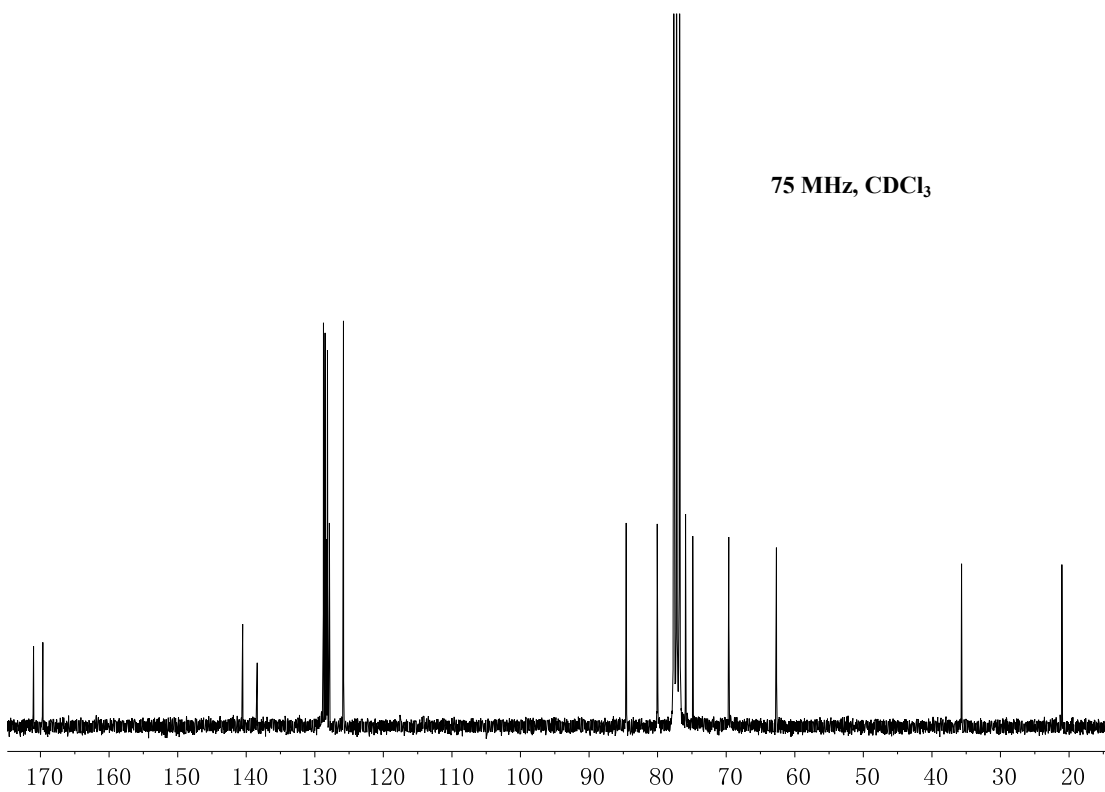


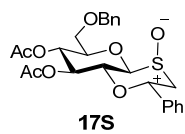


300 MHz, CDCl₃

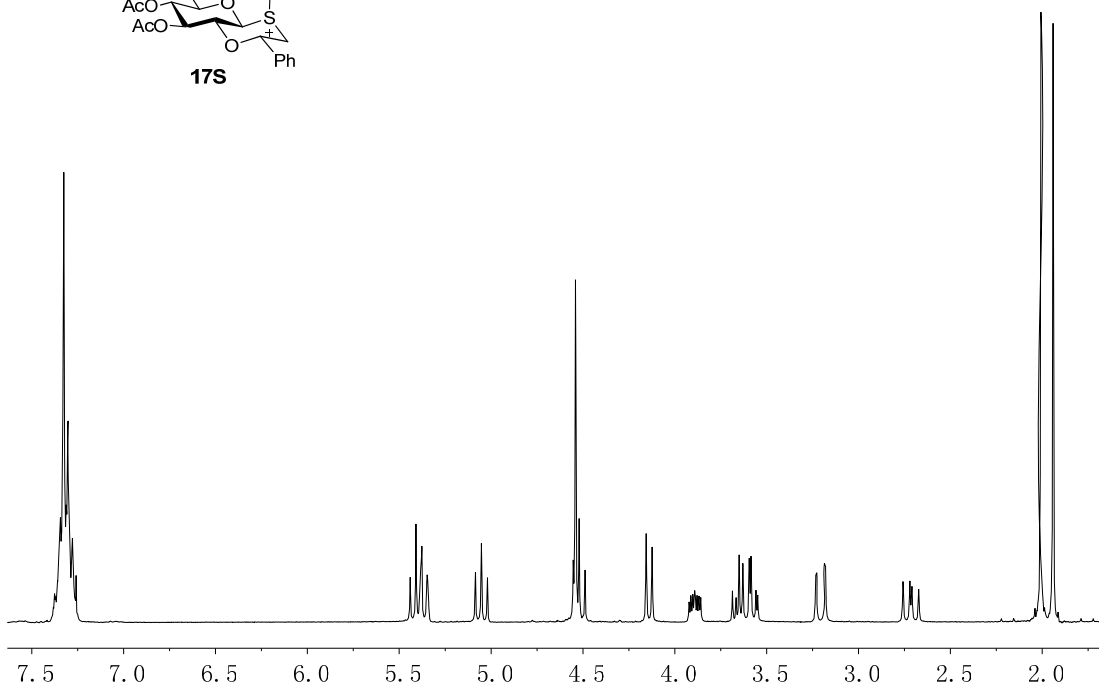


75 MHz, CDCl₃

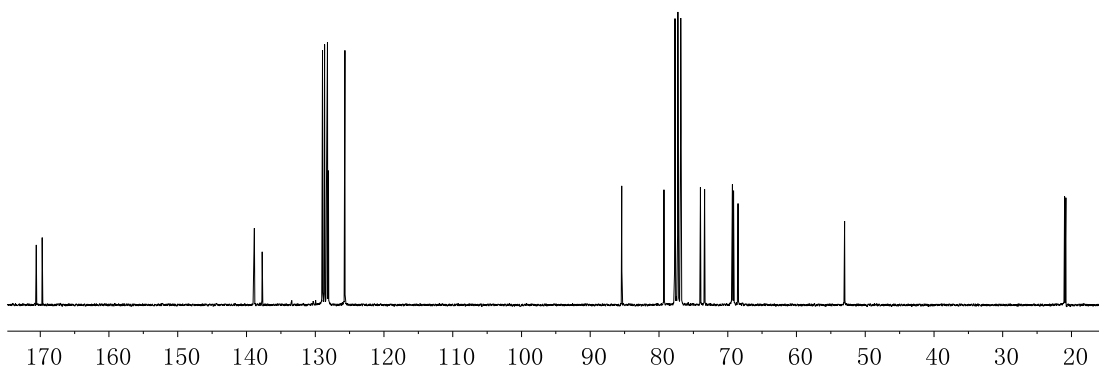


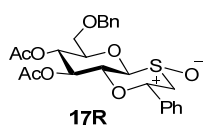


300 MHz, CDCl₃

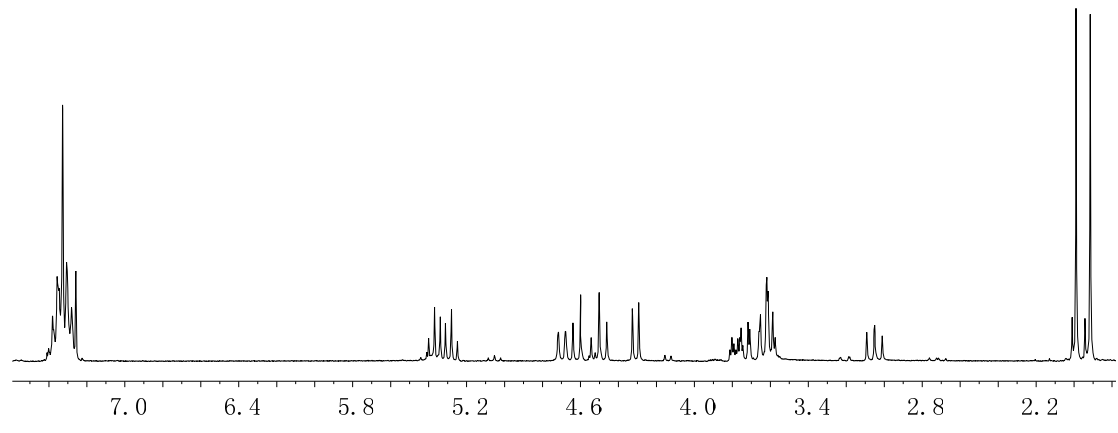


75 MHz, CDCl₃

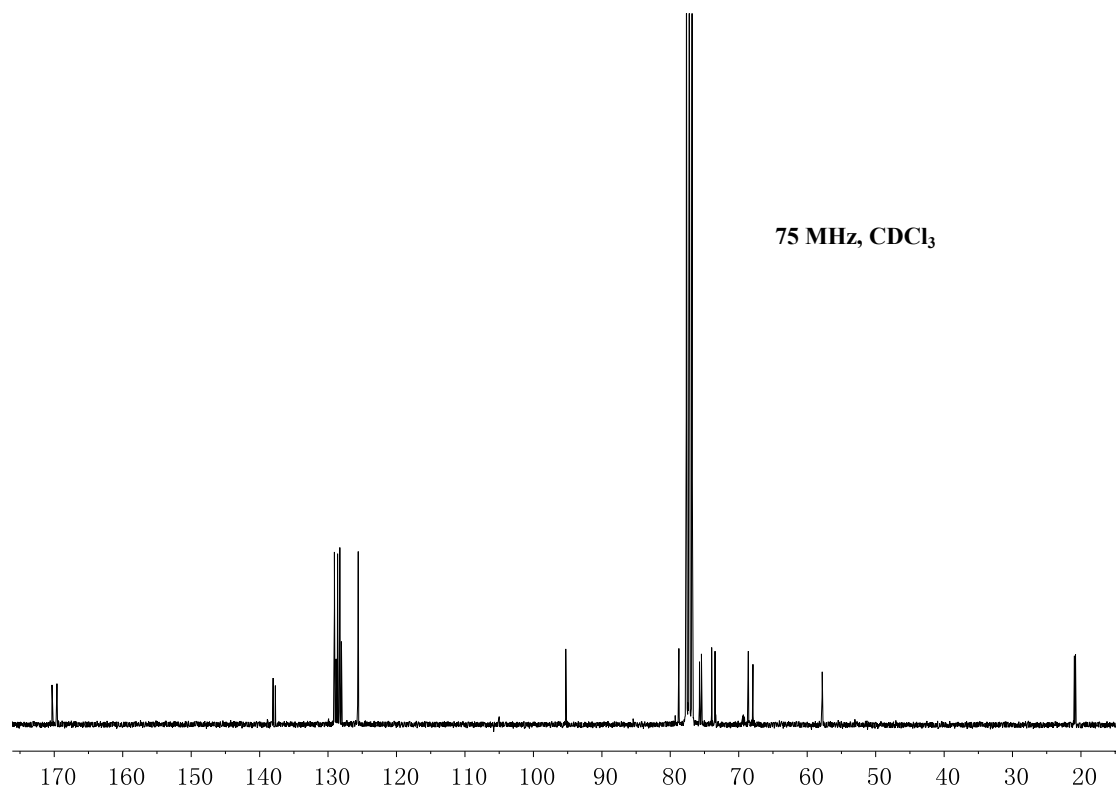


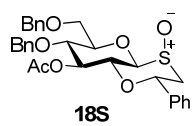


300 MHz, CDCl₃

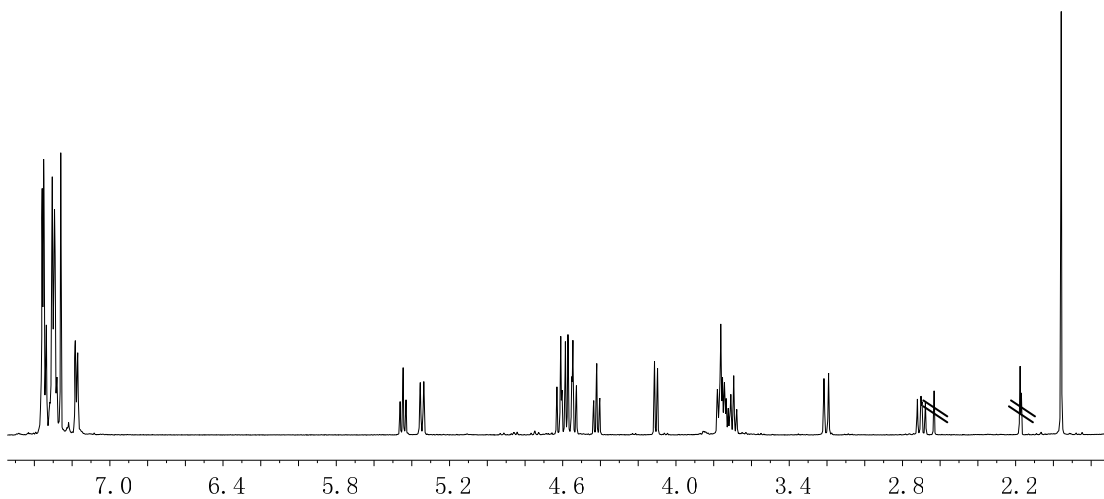


75 MHz, CDCl₃

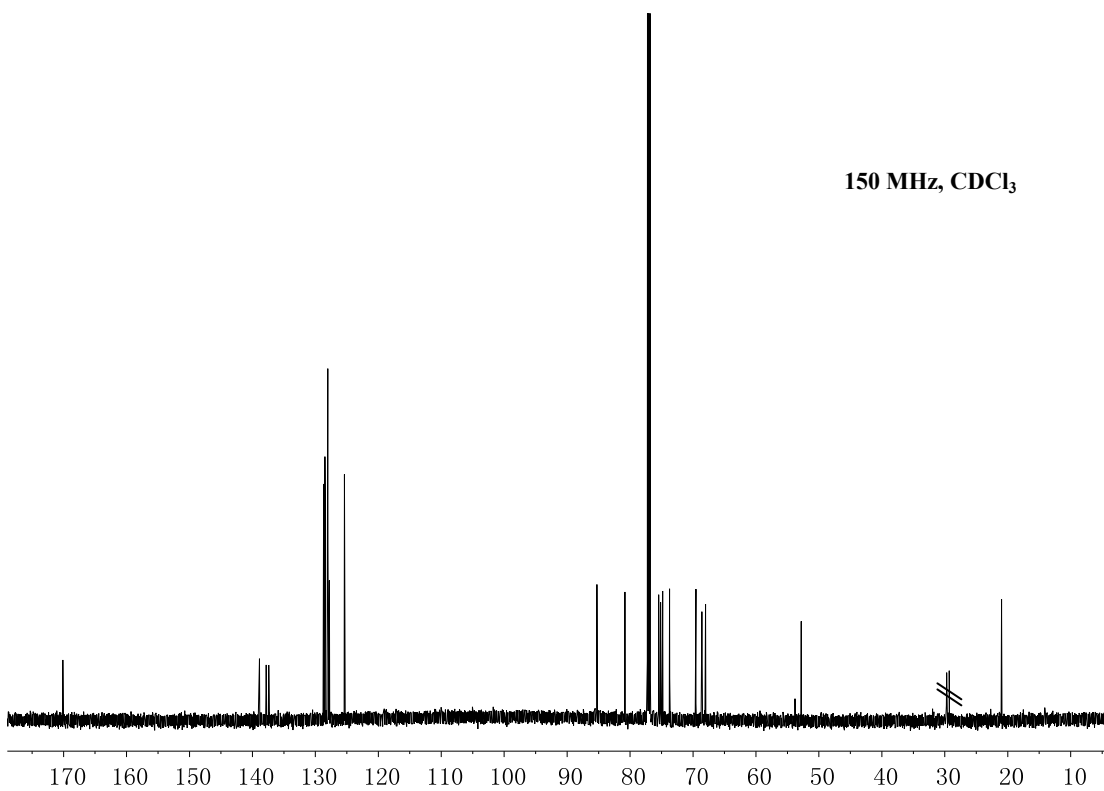


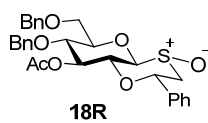


600 MHz, CDCl₃

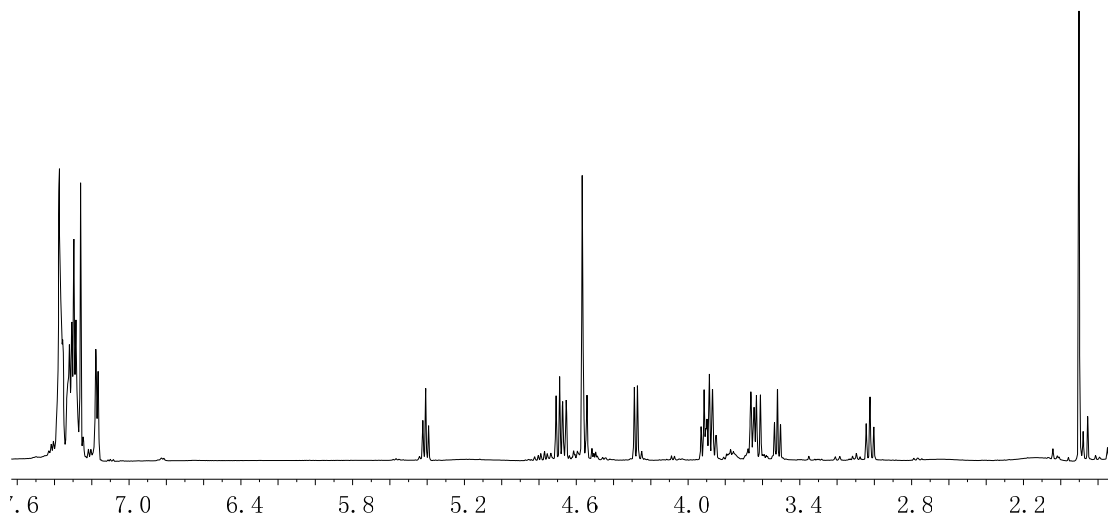


150 MHz, CDCl₃

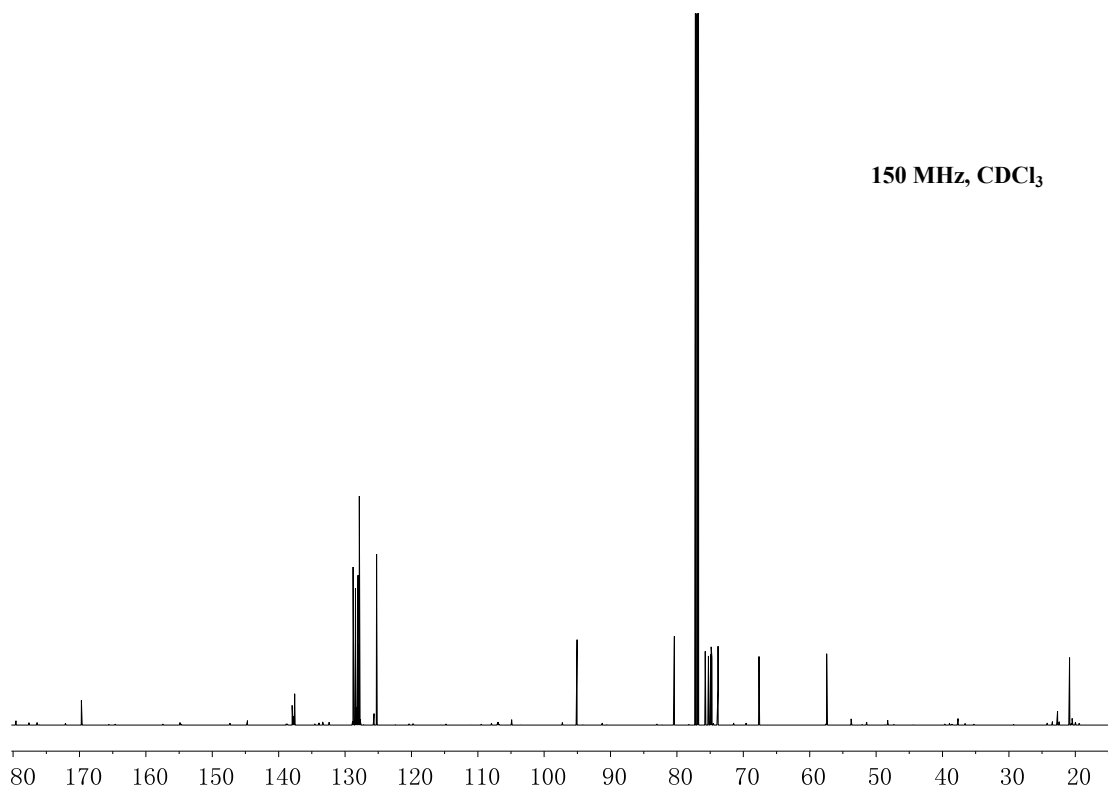


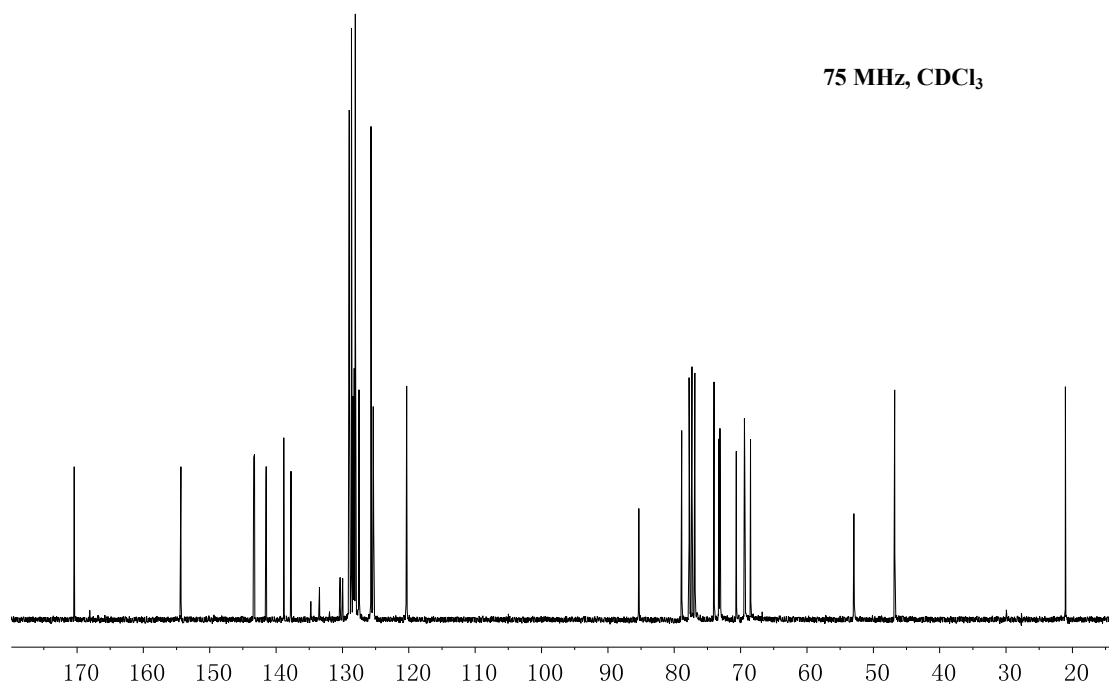
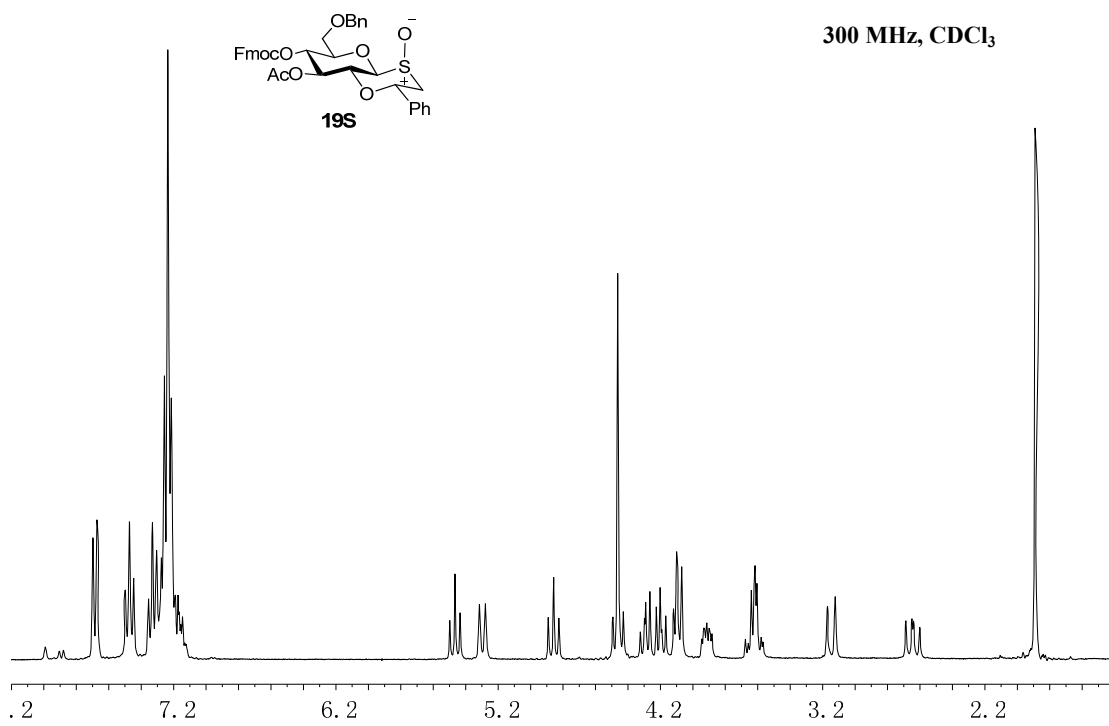


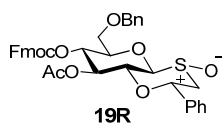
600 MHz, CDCl₃



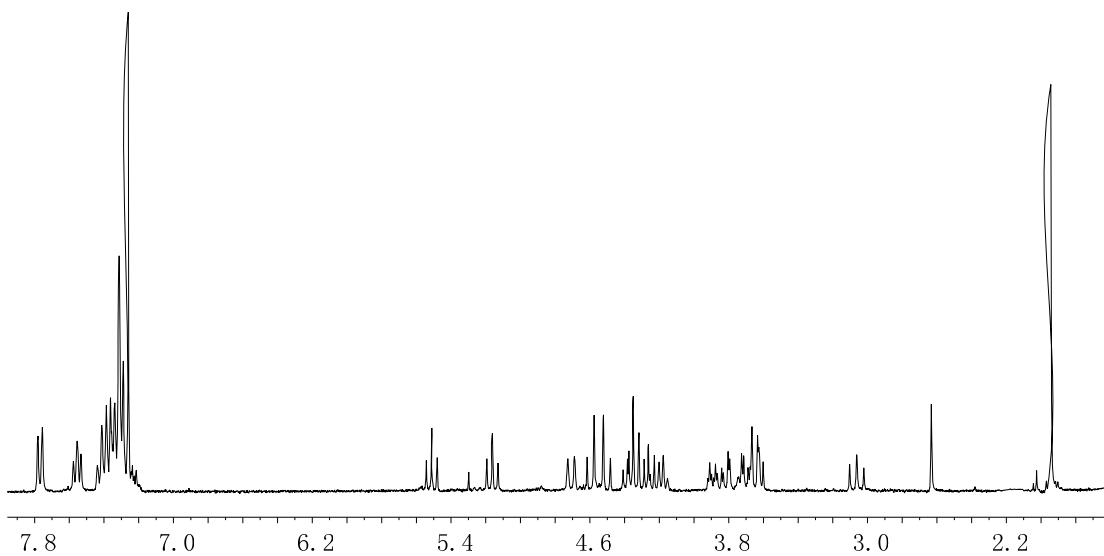
150 MHz, CDCl₃



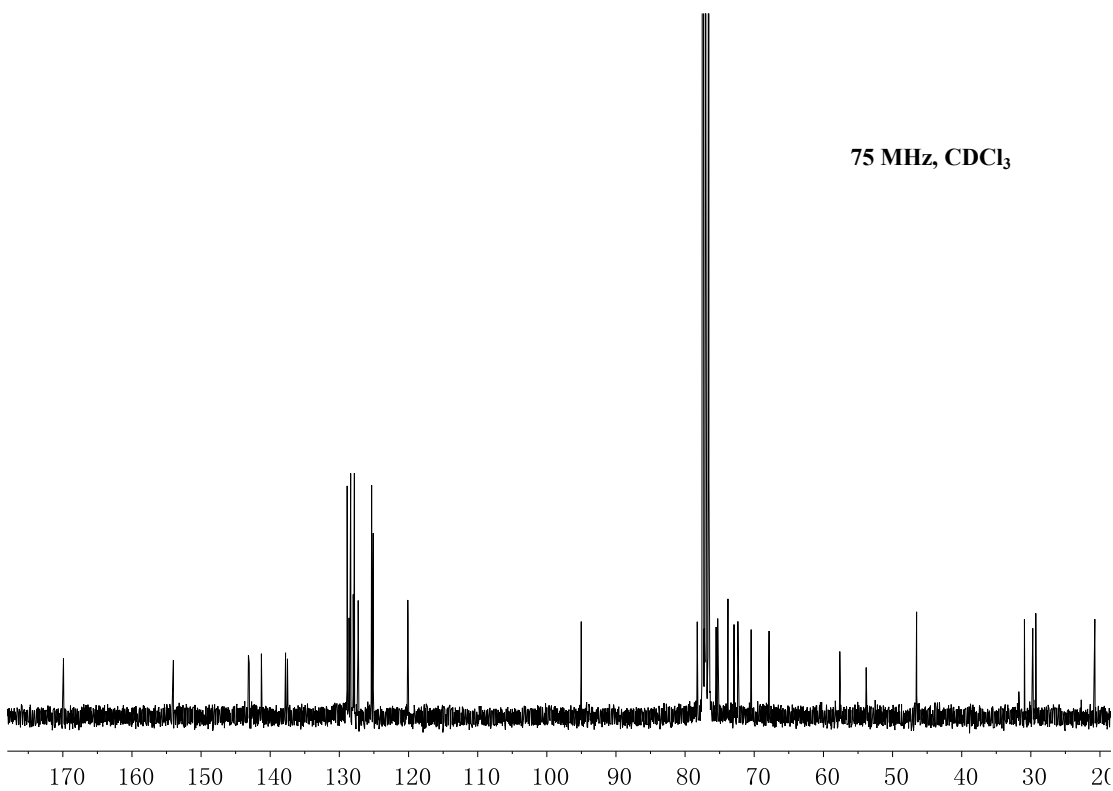


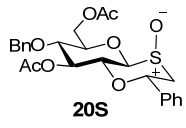


300 MHz, CDCl₃

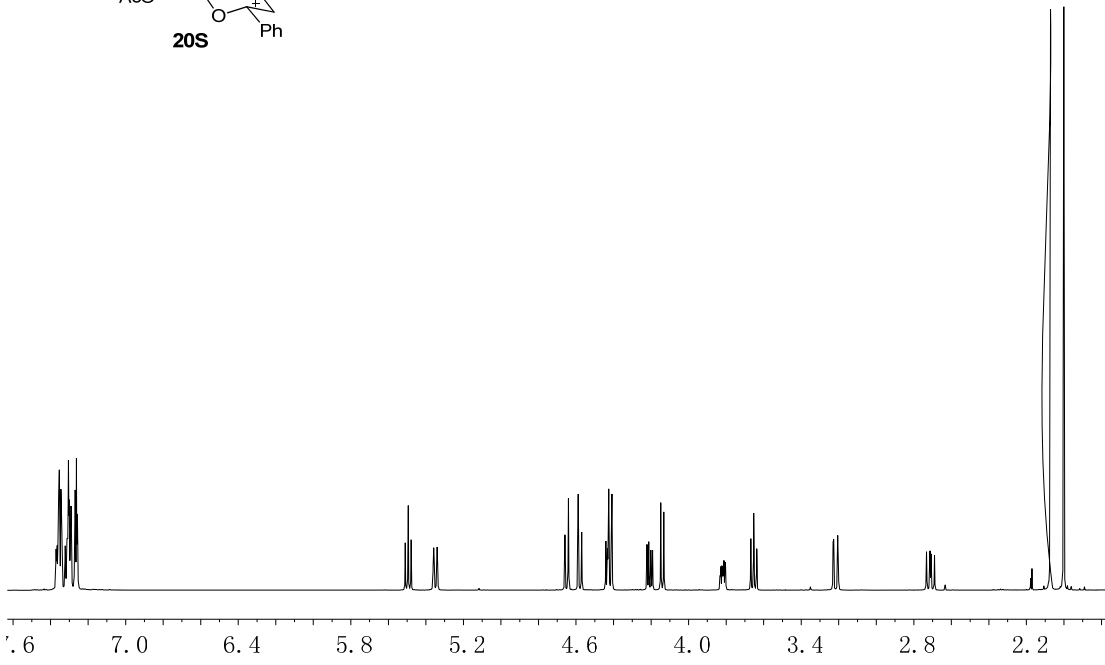


75 MHz, CDCl₃

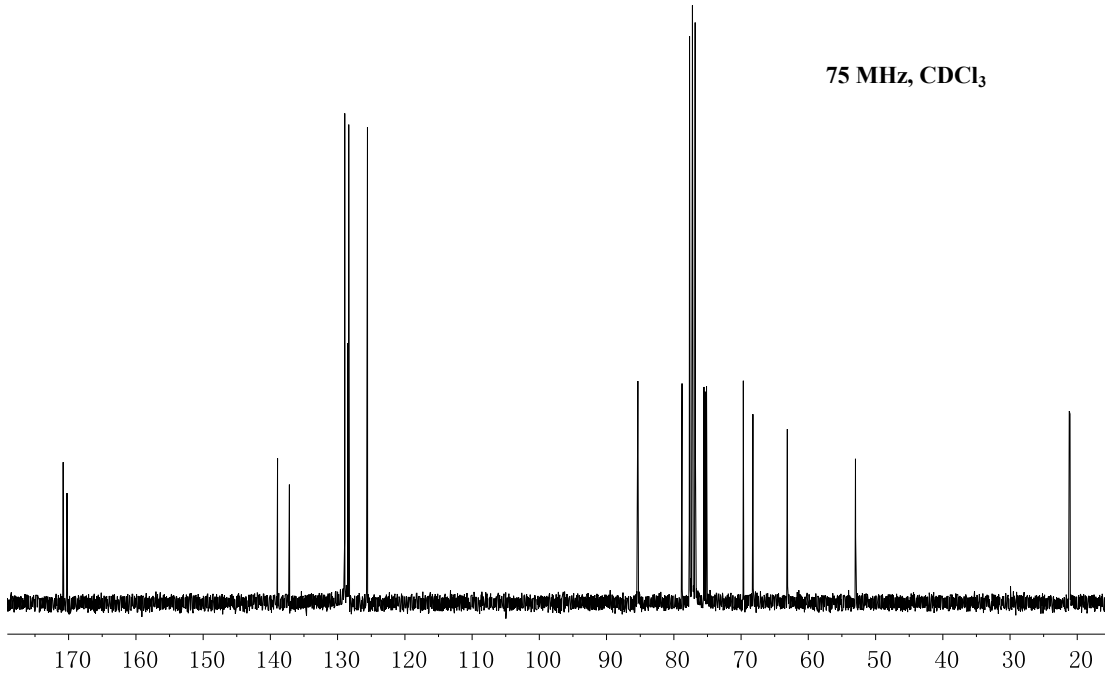


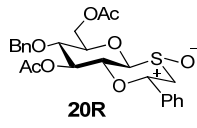


600 MHz, CDCl₃

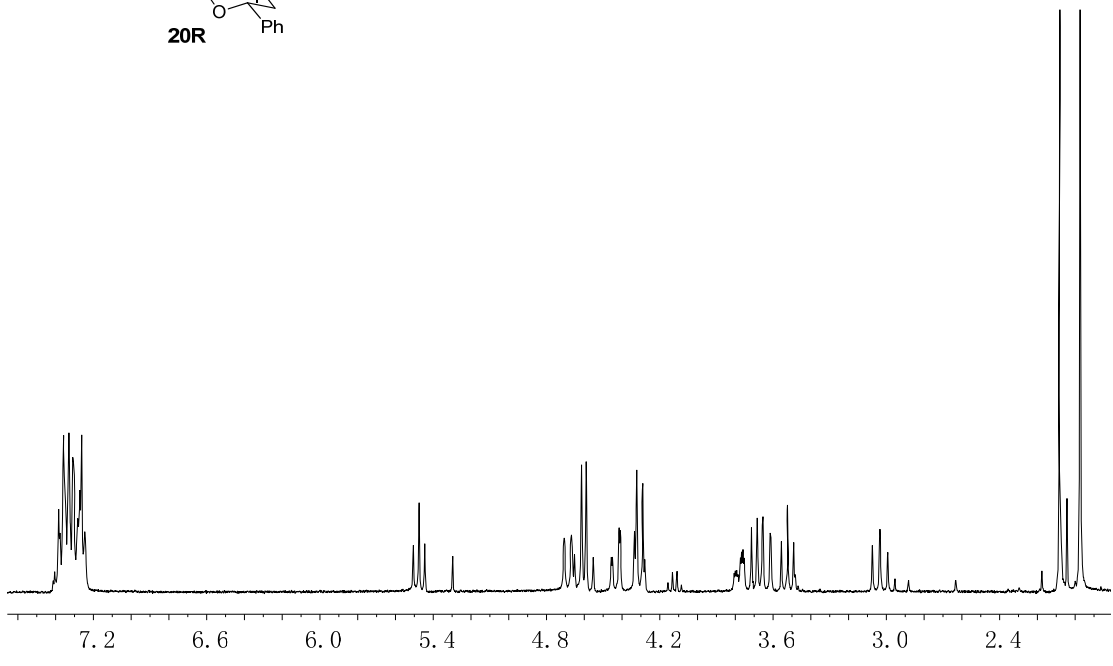


75 MHz, CDCl₃

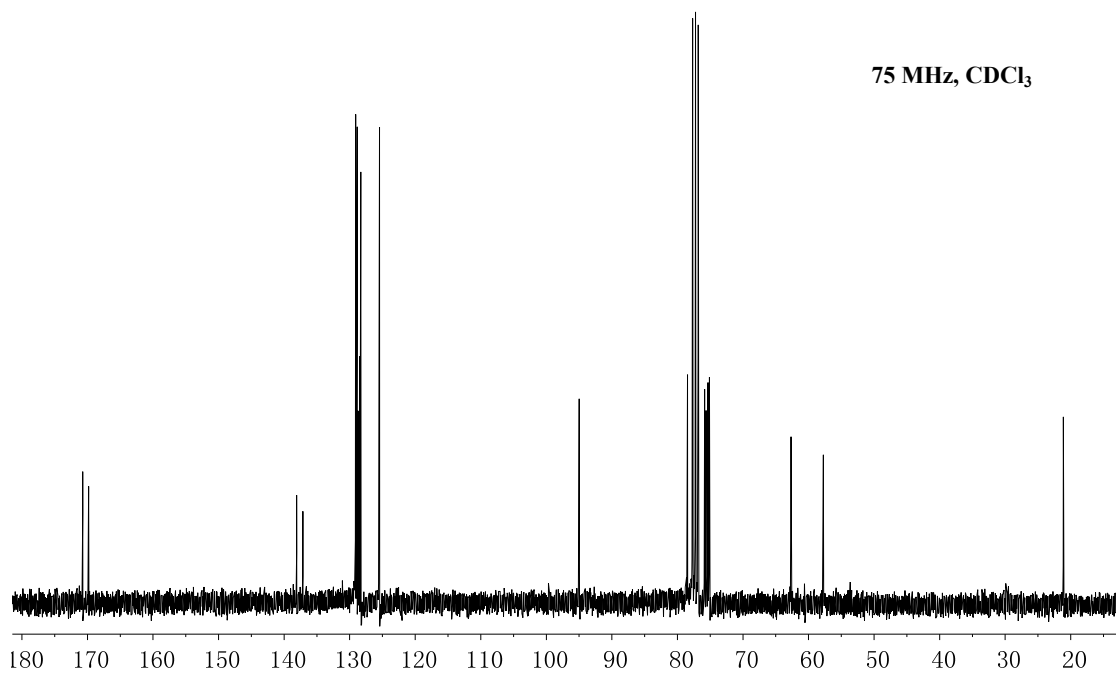


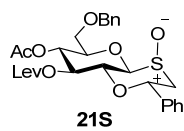


300 MHz, CDCl₃

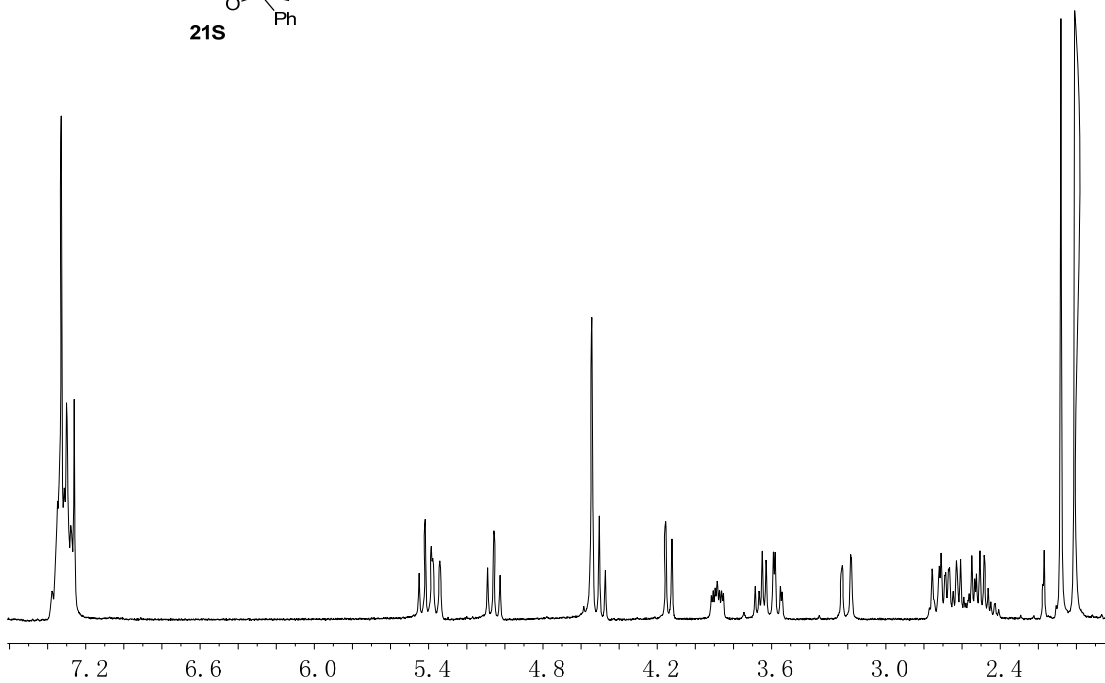


75 MHz, CDCl₃

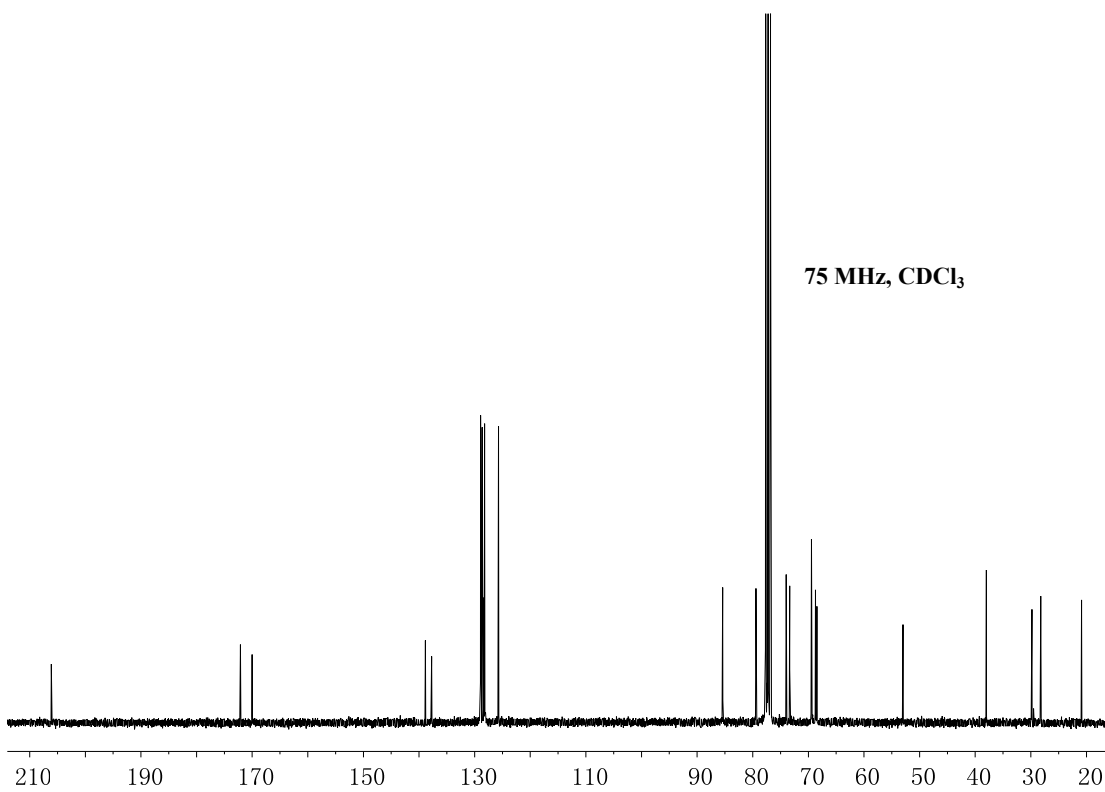


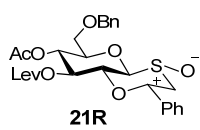


300 MHz, CDCl₃

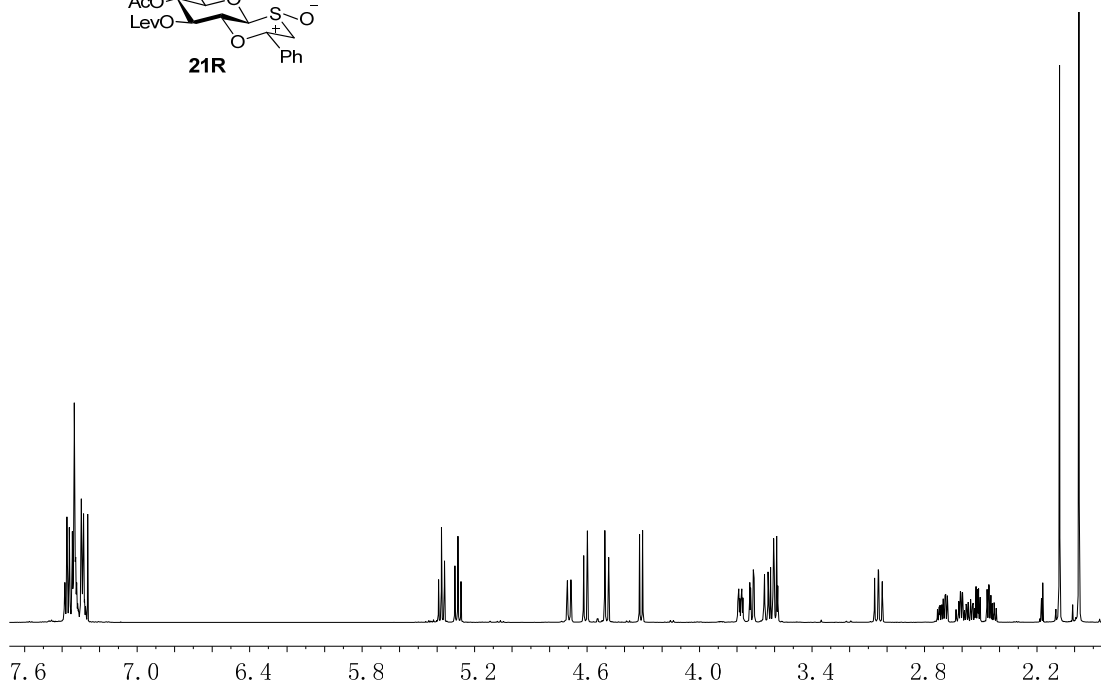


75 MHz, CDCl₃

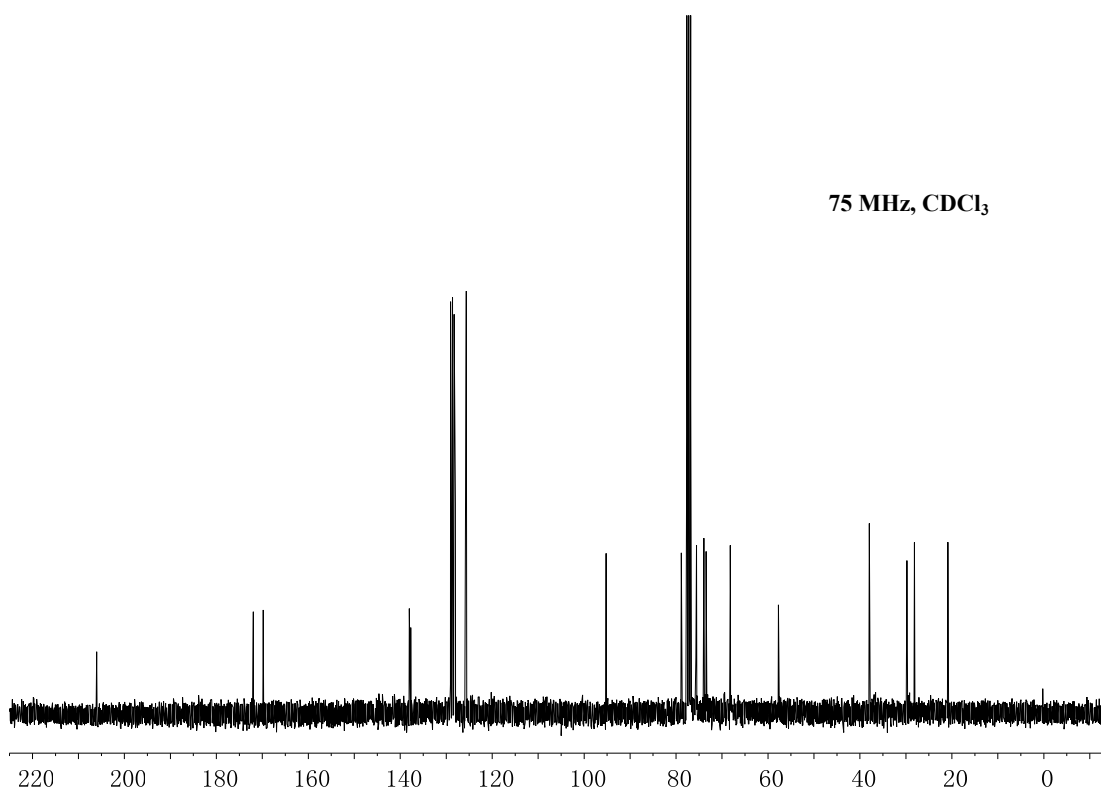


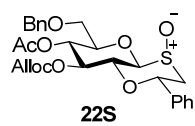


600 MHz, CDCl₃

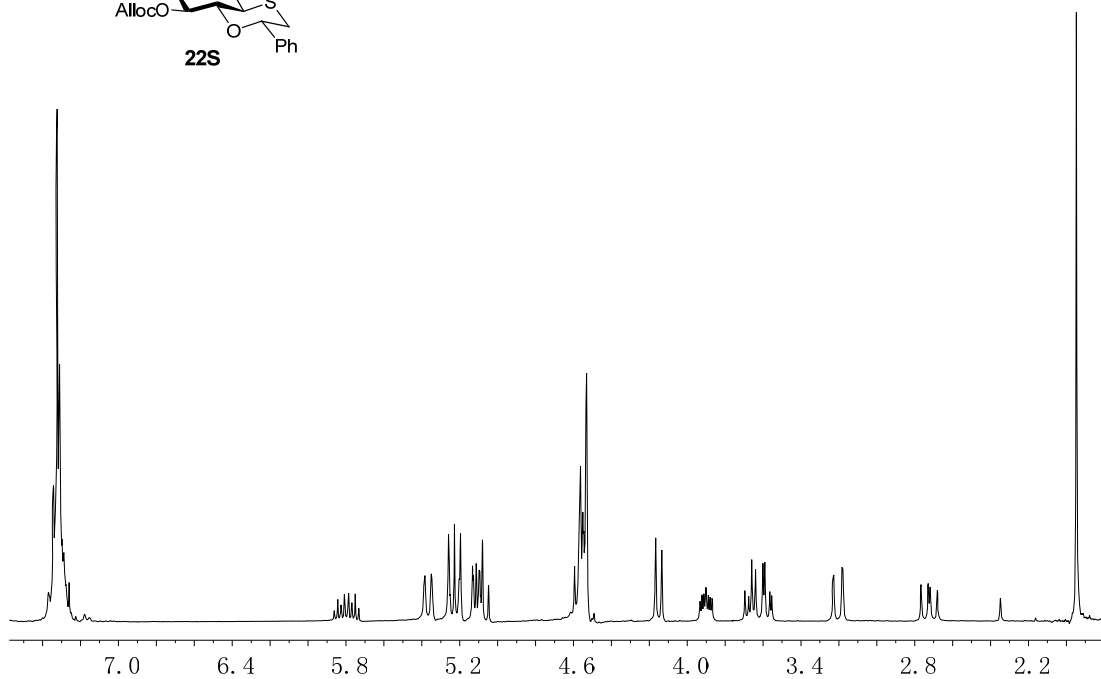


75 MHz, CDCl₃

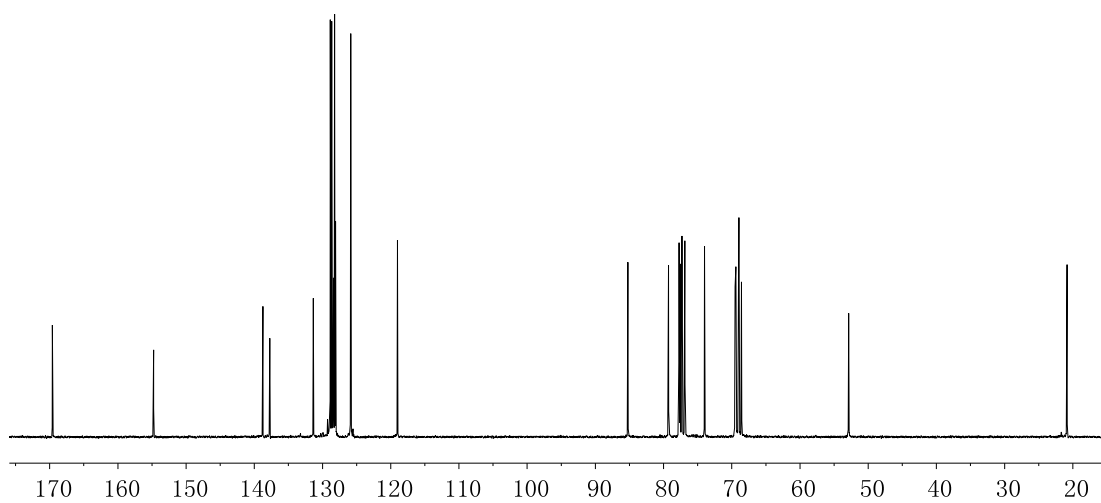


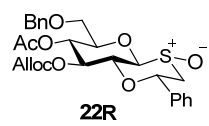


300 MHz, CDCl₃

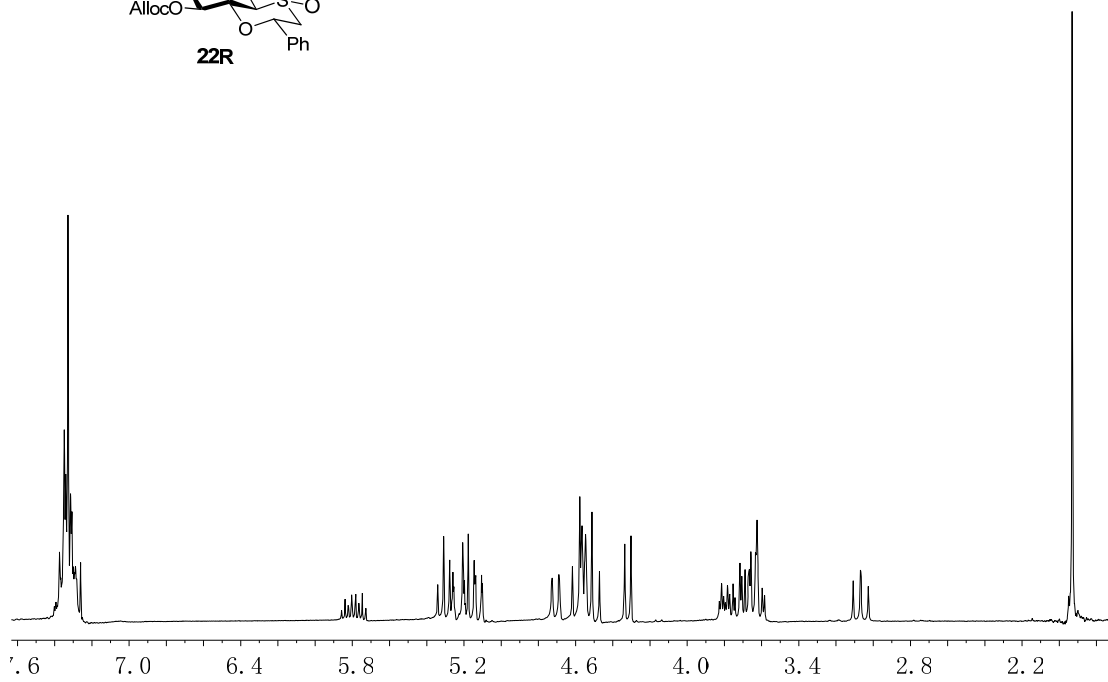


75 MHz, CDCl₃

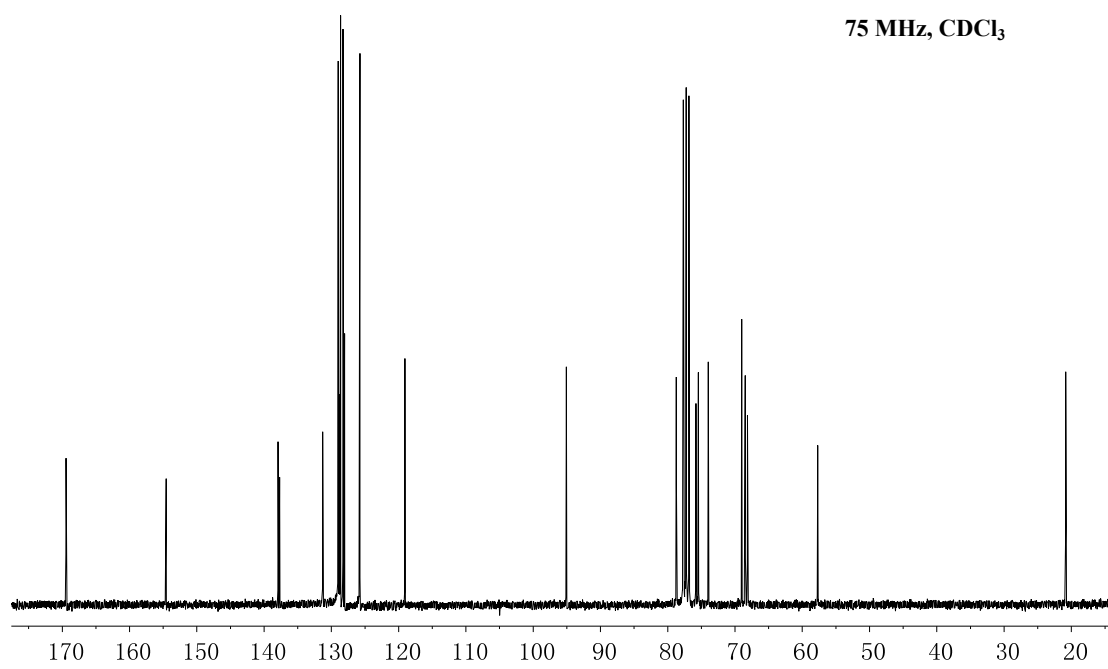


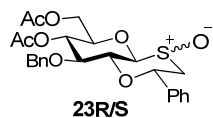


300 MHz, CDCl₃

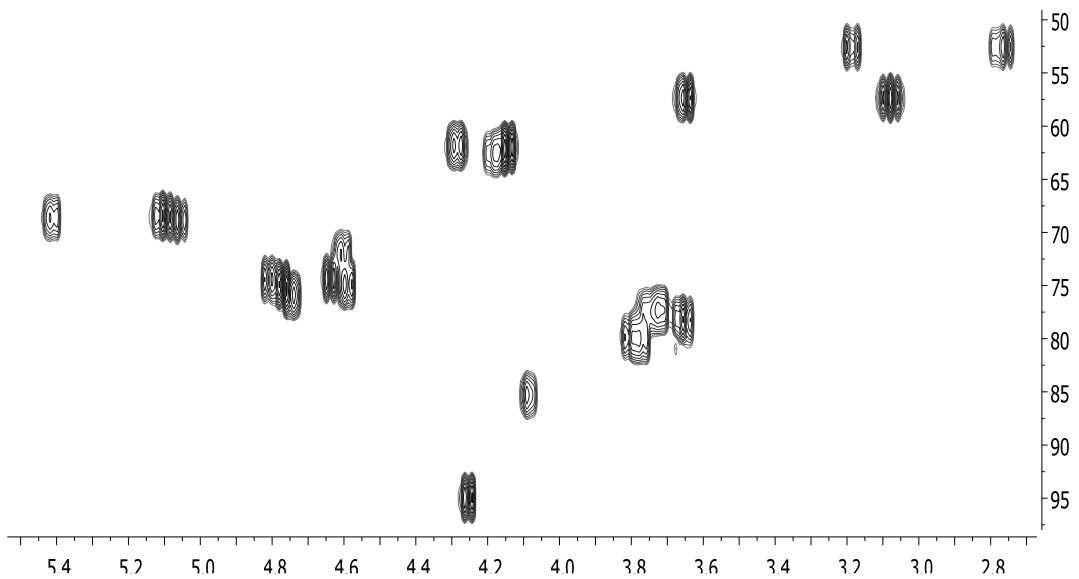
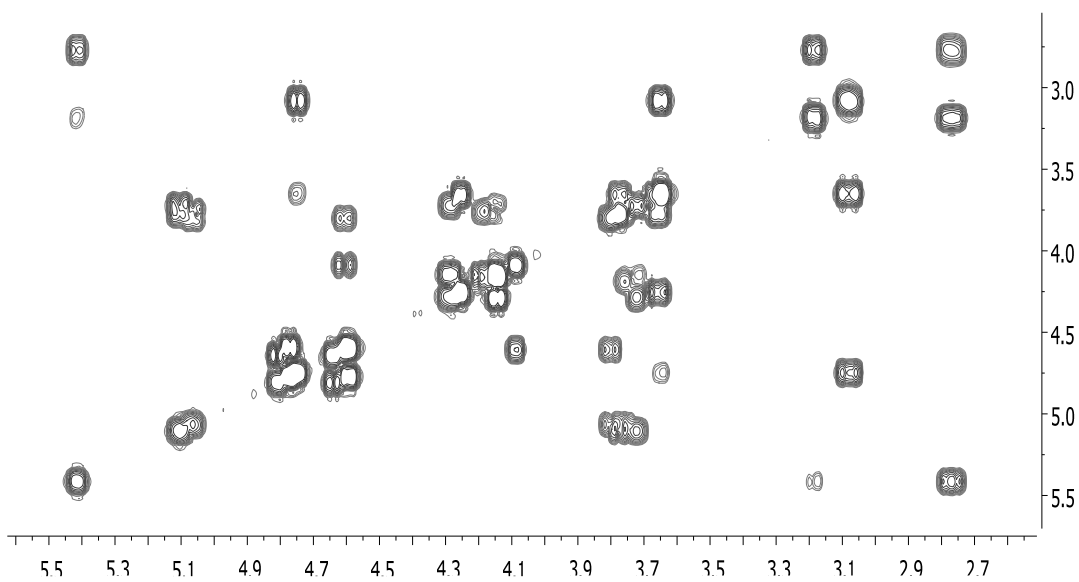
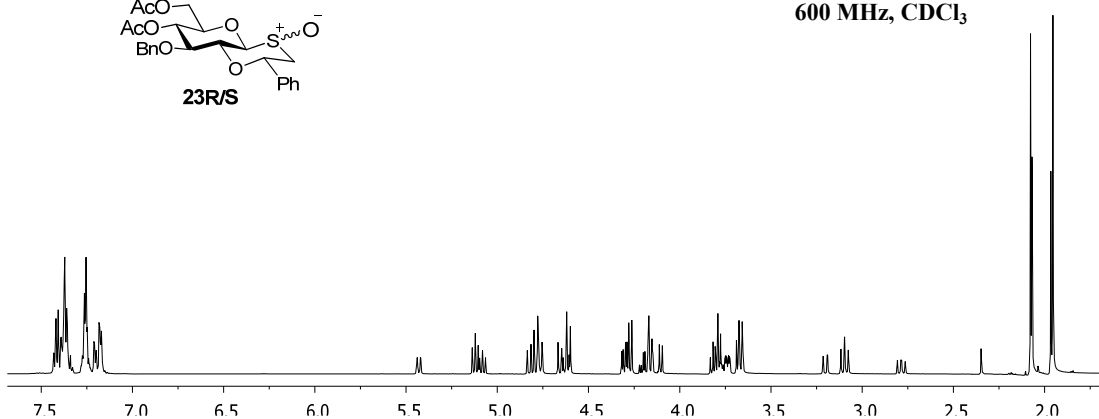


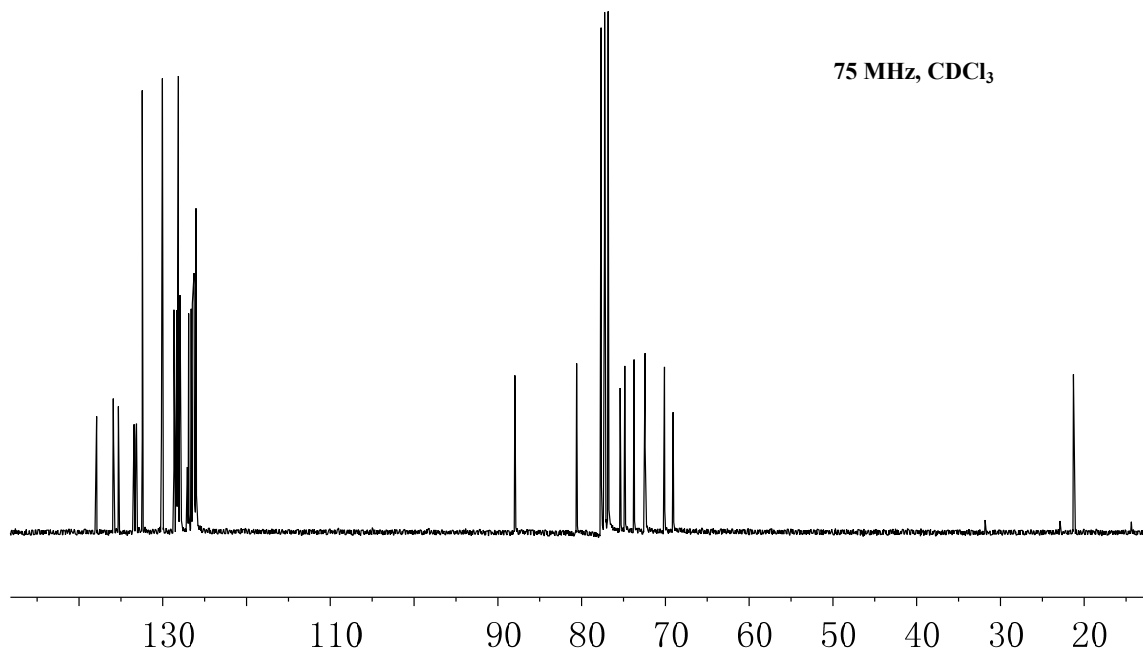
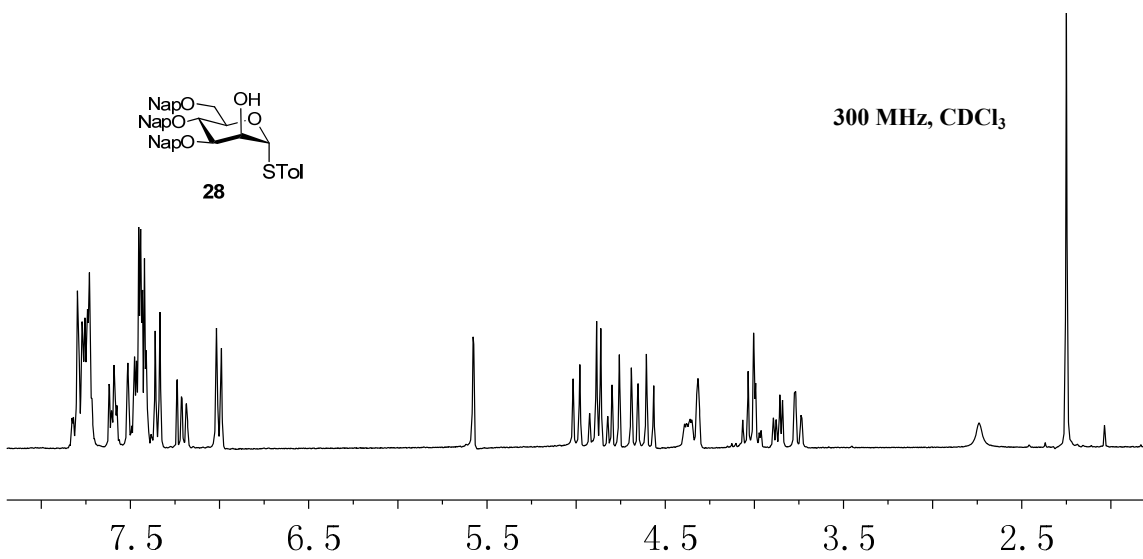
75 MHz, CDCl₃

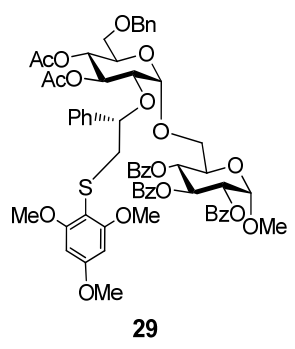




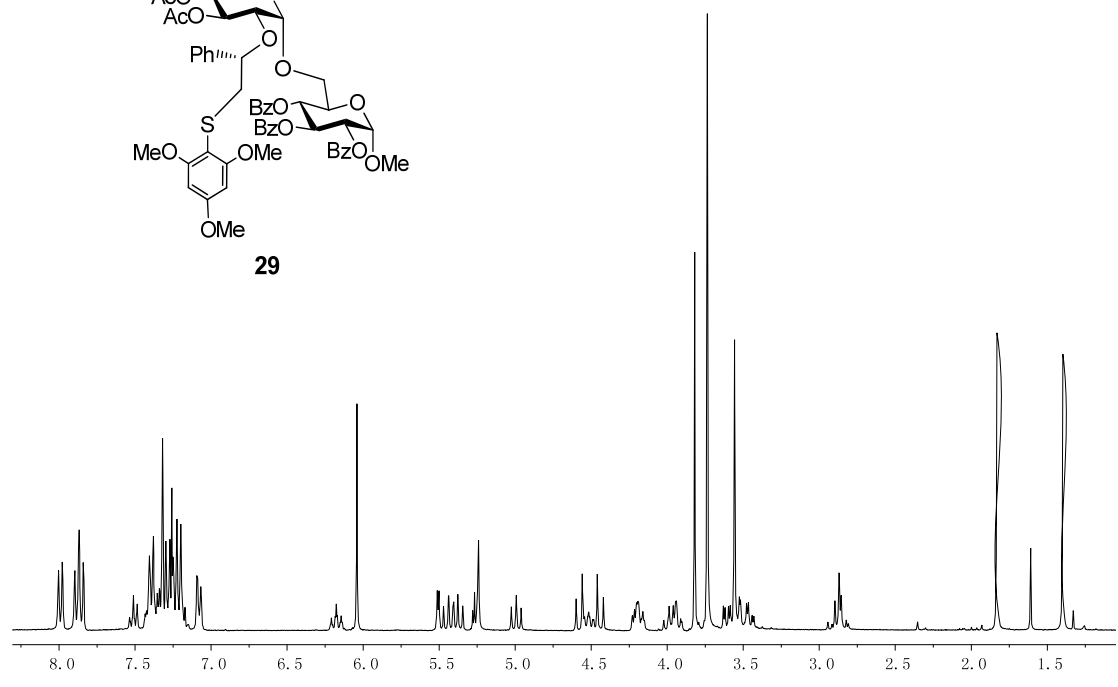
600 MHz, CDCl₃



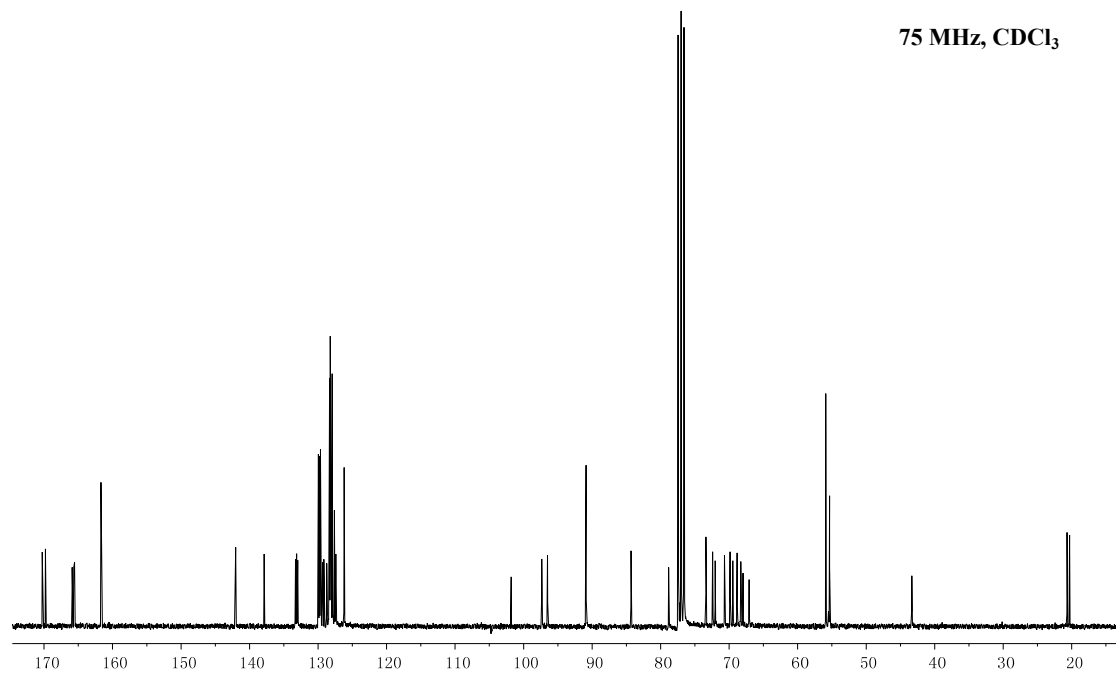


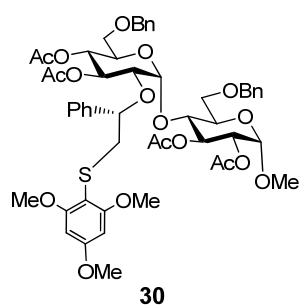


300 MHz, CDCl₃

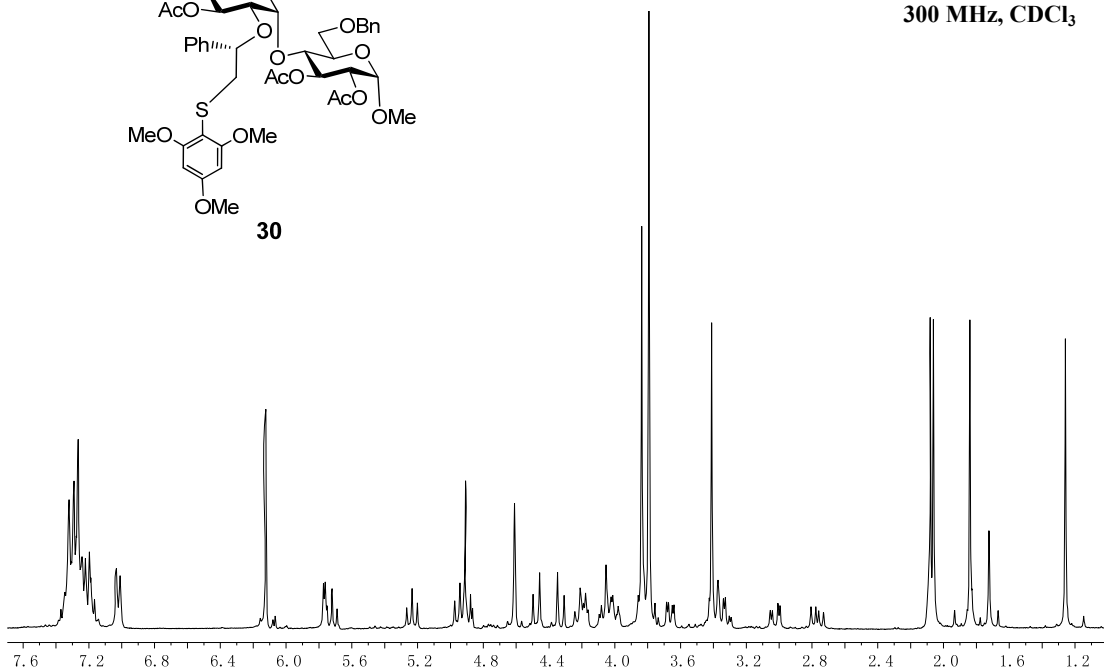


75 MHz, CDCl₃

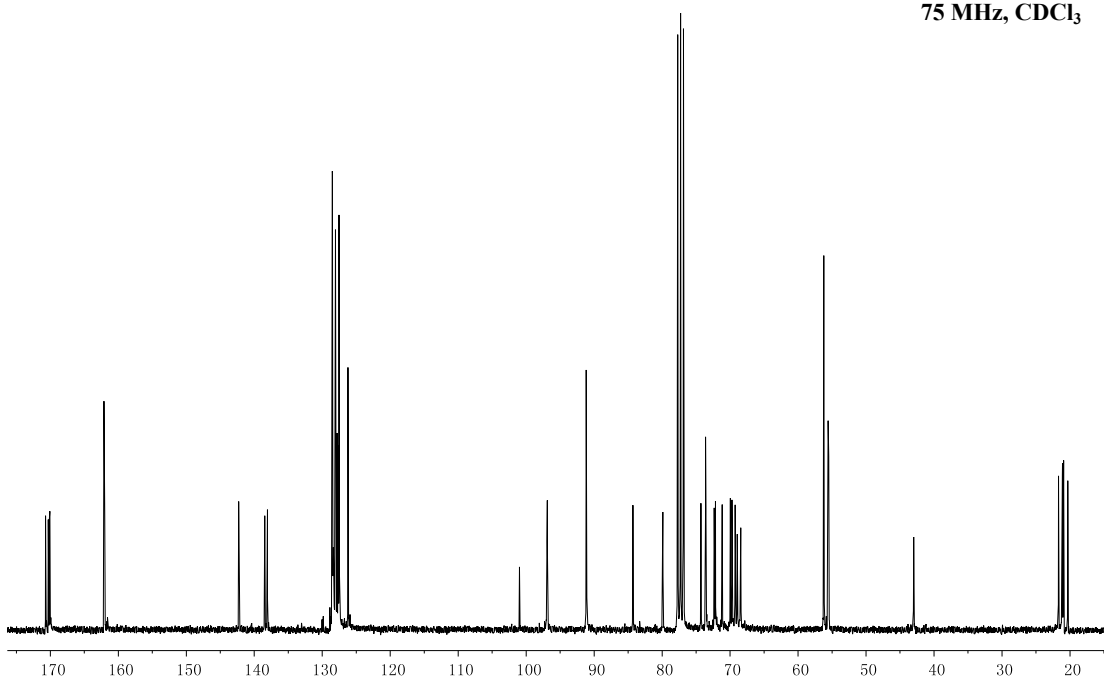


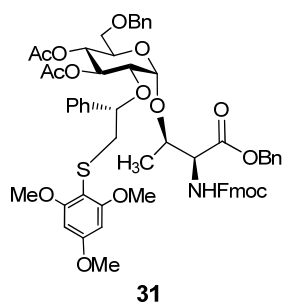


300 MHz, CDCl₃

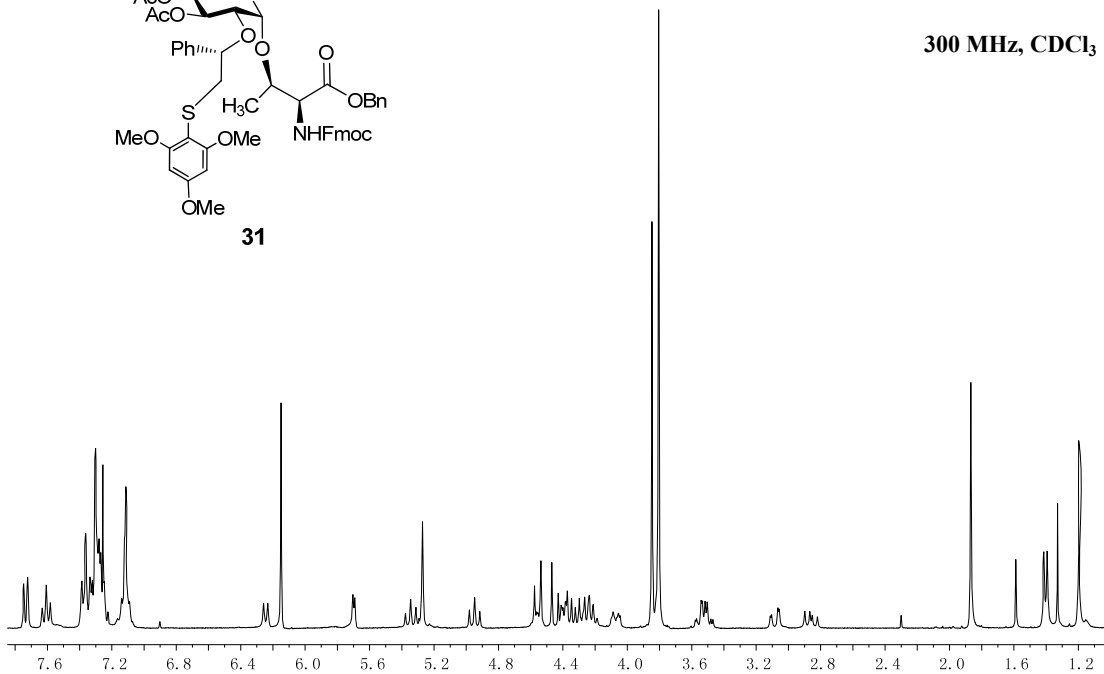


75 MHz, CDCl₃

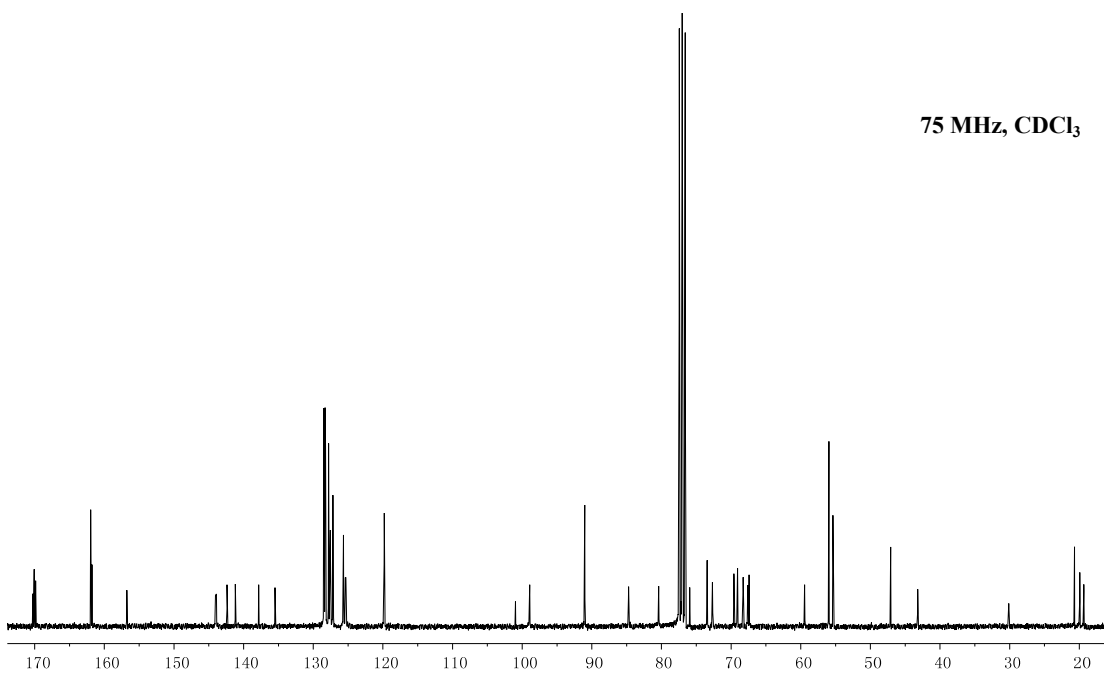


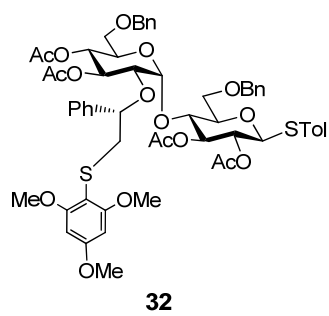


300 MHz, CDCl₃

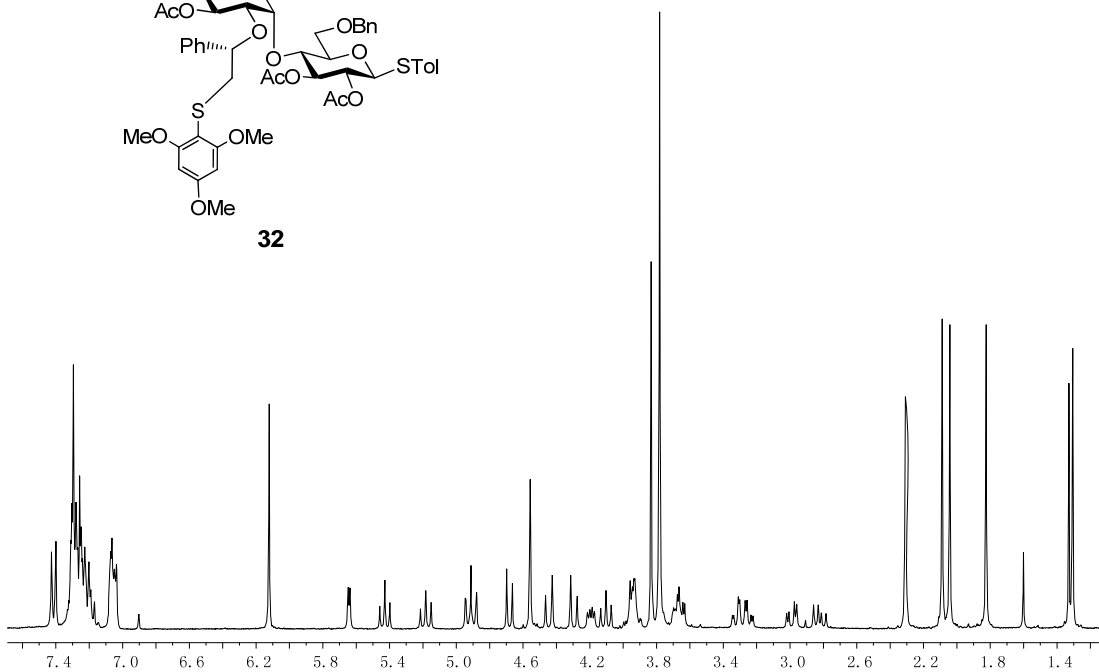


75 MHz, CDCl₃

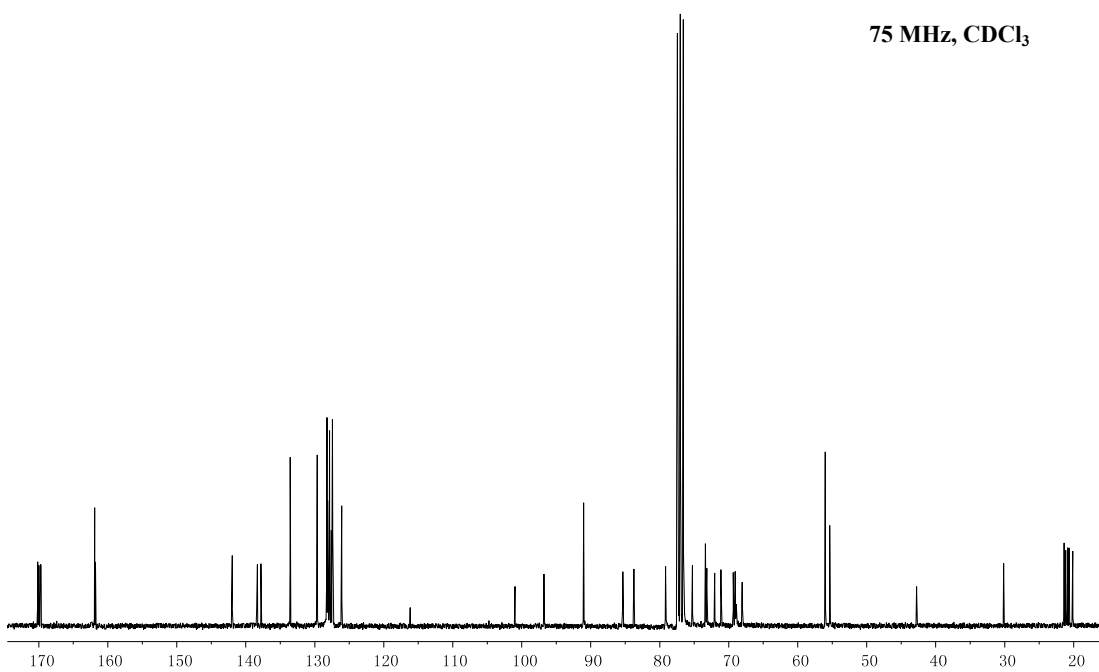


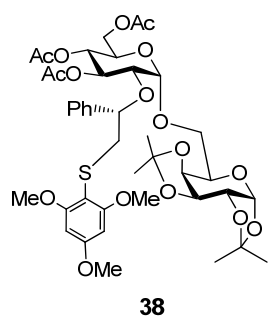


300 MHz, CDCl₃

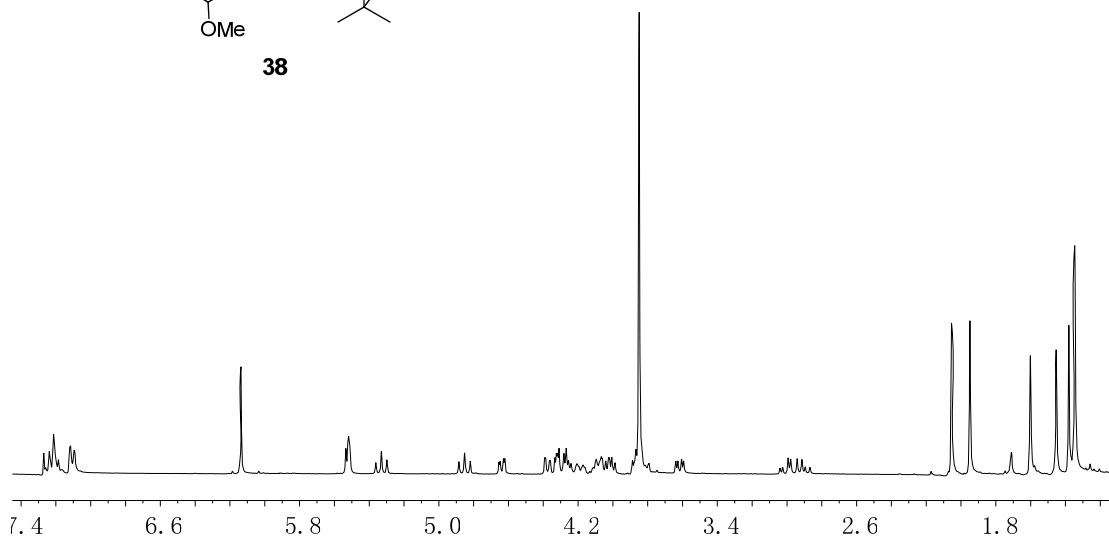


75 MHz, CDCl₃

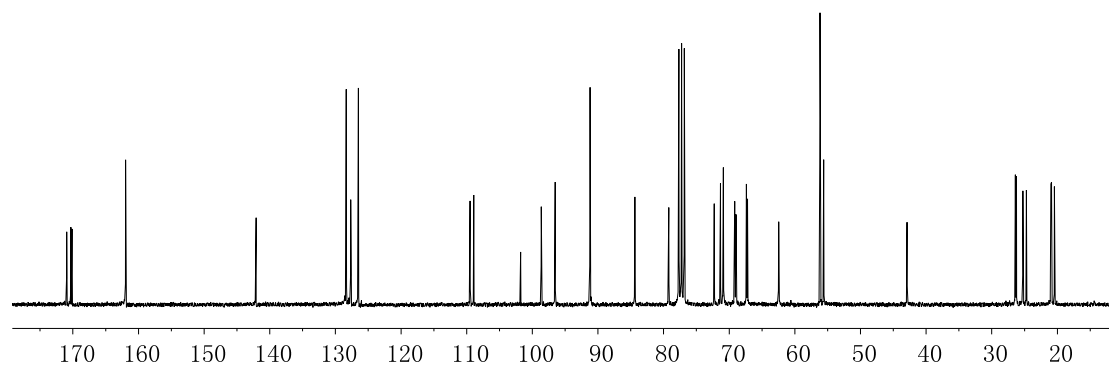


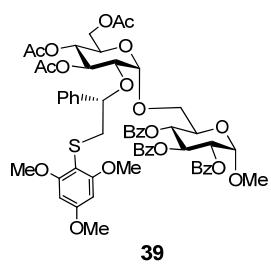


300 MHz, CDCl₃

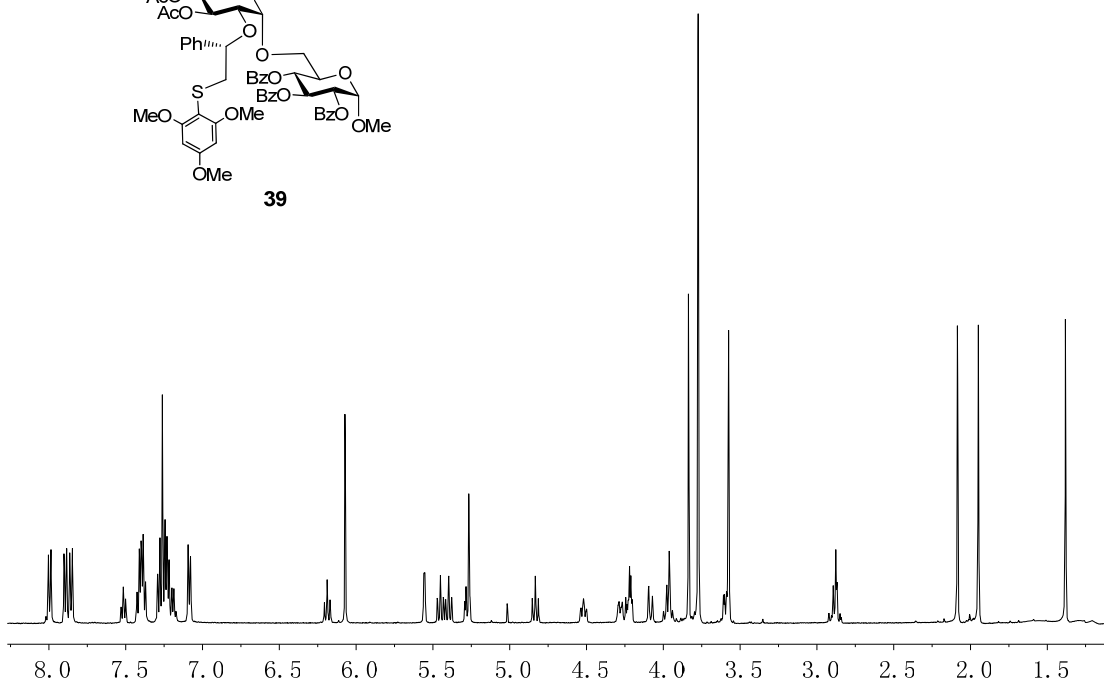


75 MHz, CDCl₃

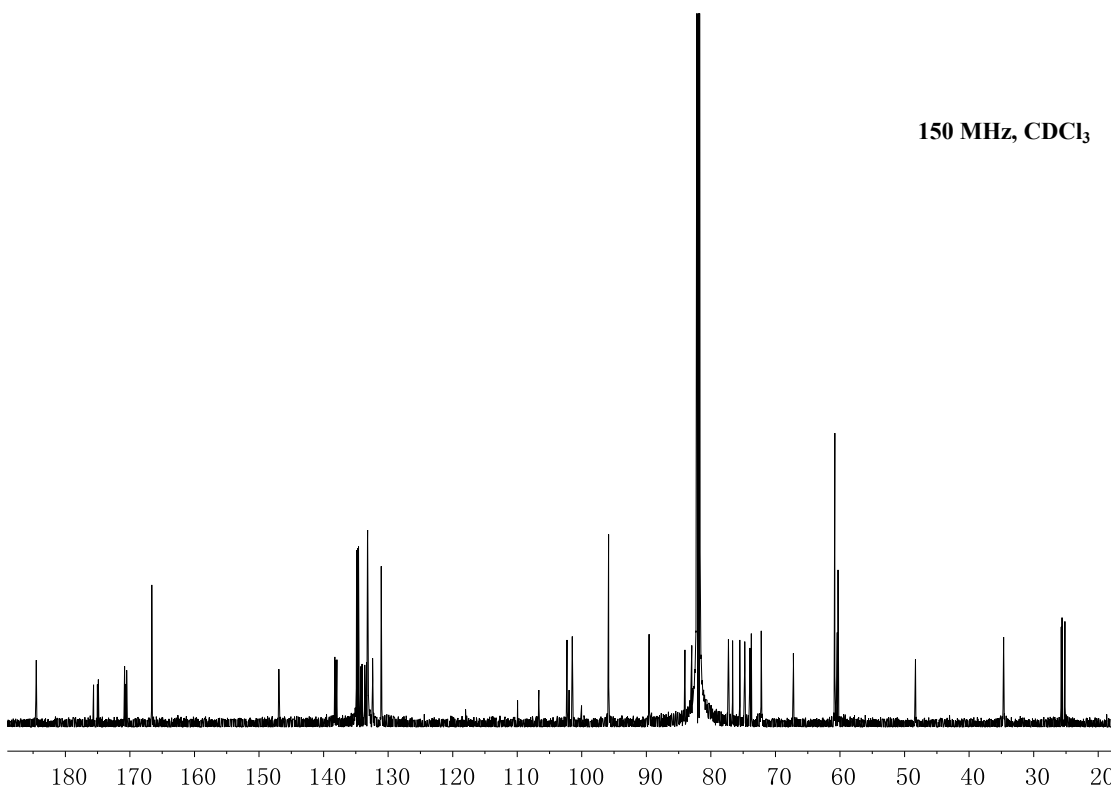


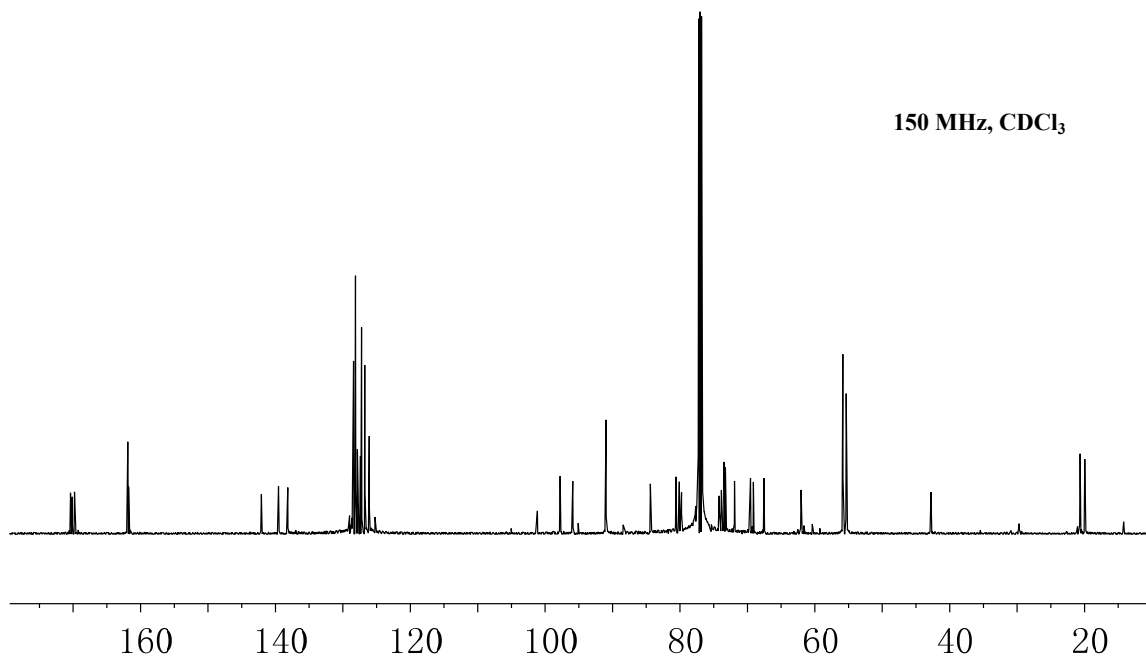
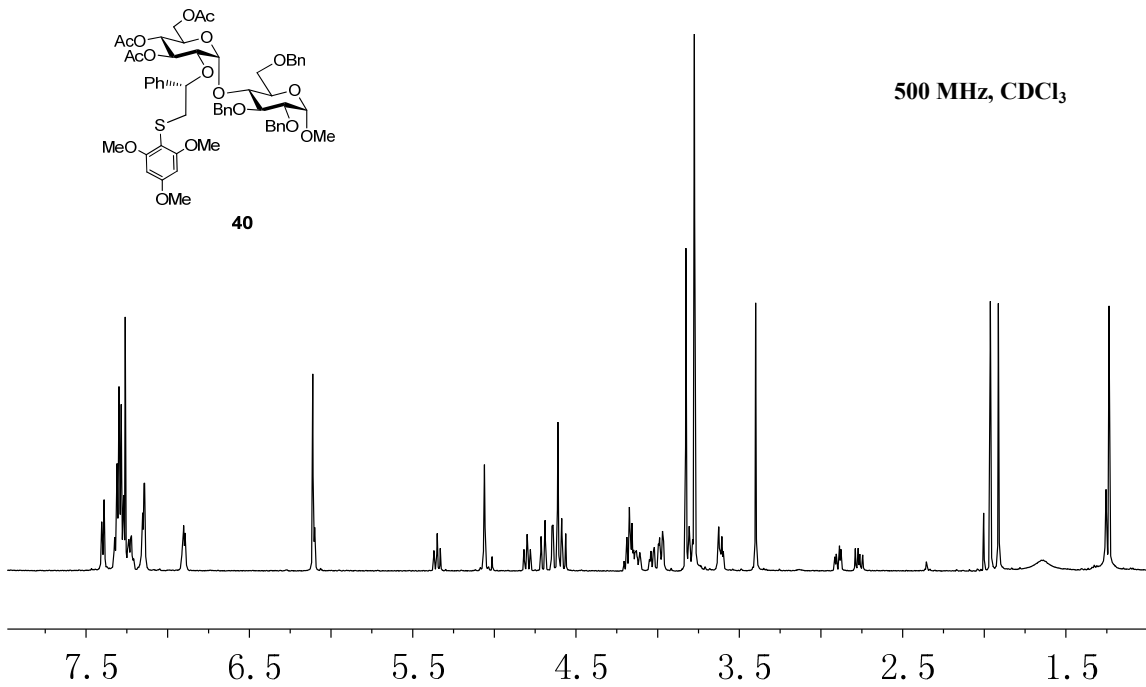


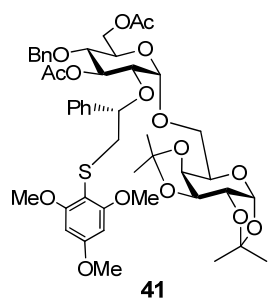
600 MHz, CDCl₃



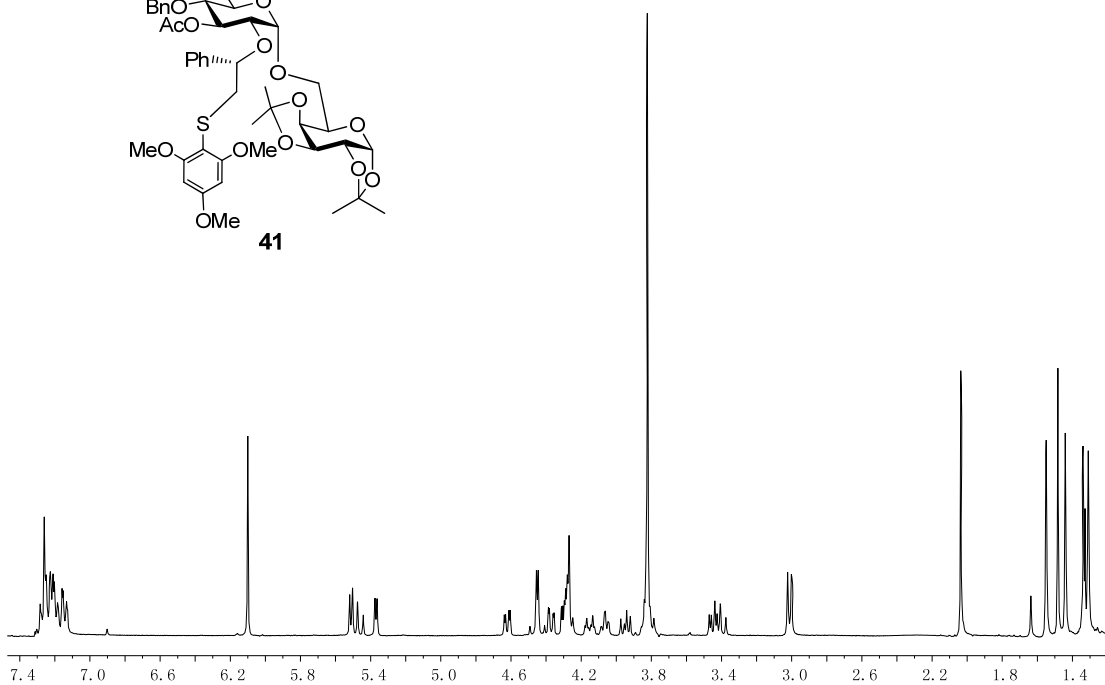
150 MHz, CDCl₃



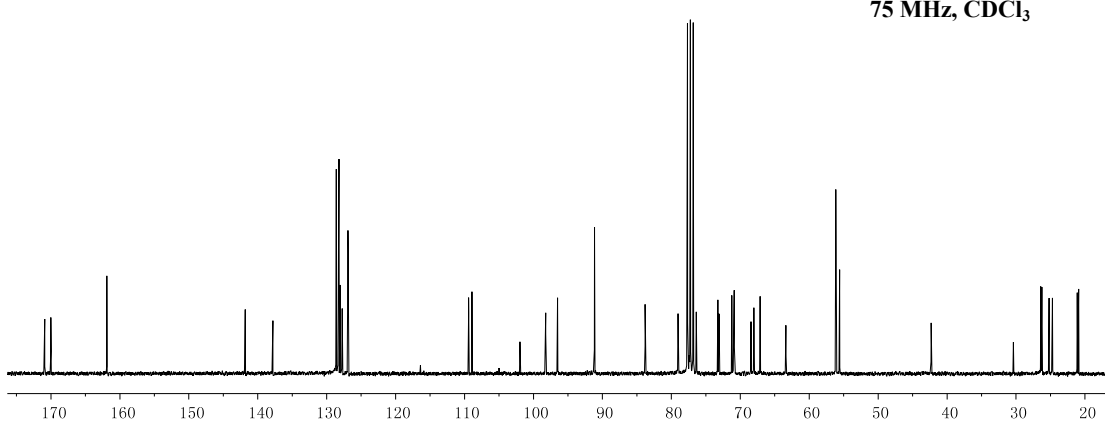


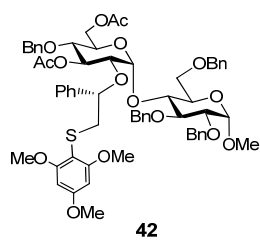


300 MHz, CDCl₃

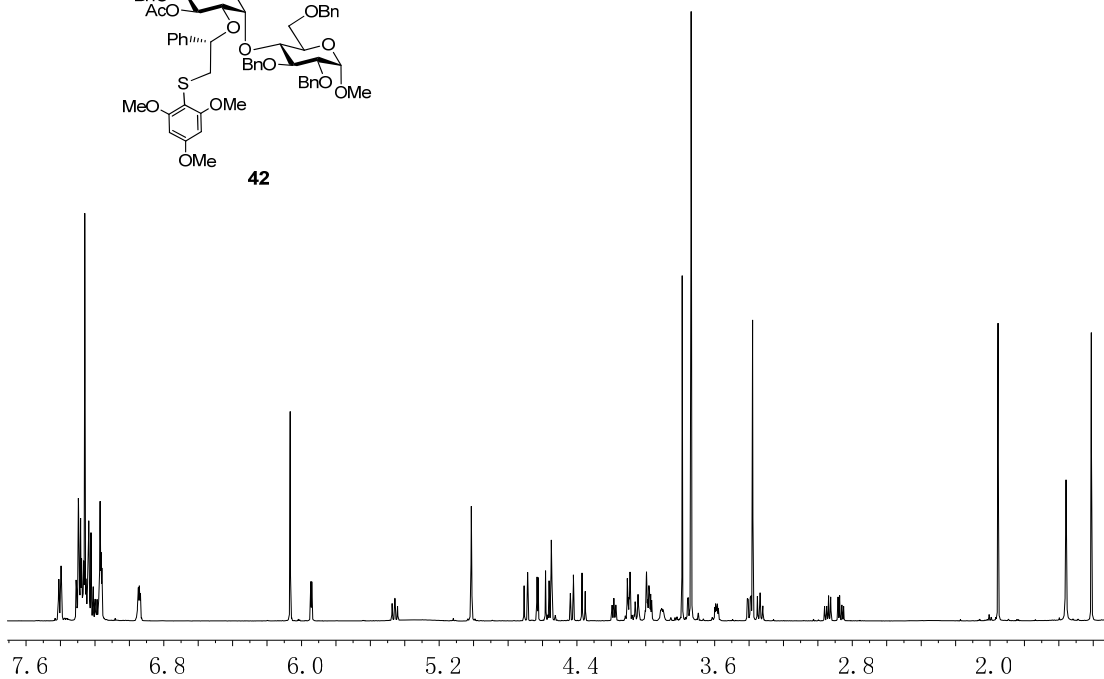


75 MHz, CDCl₃

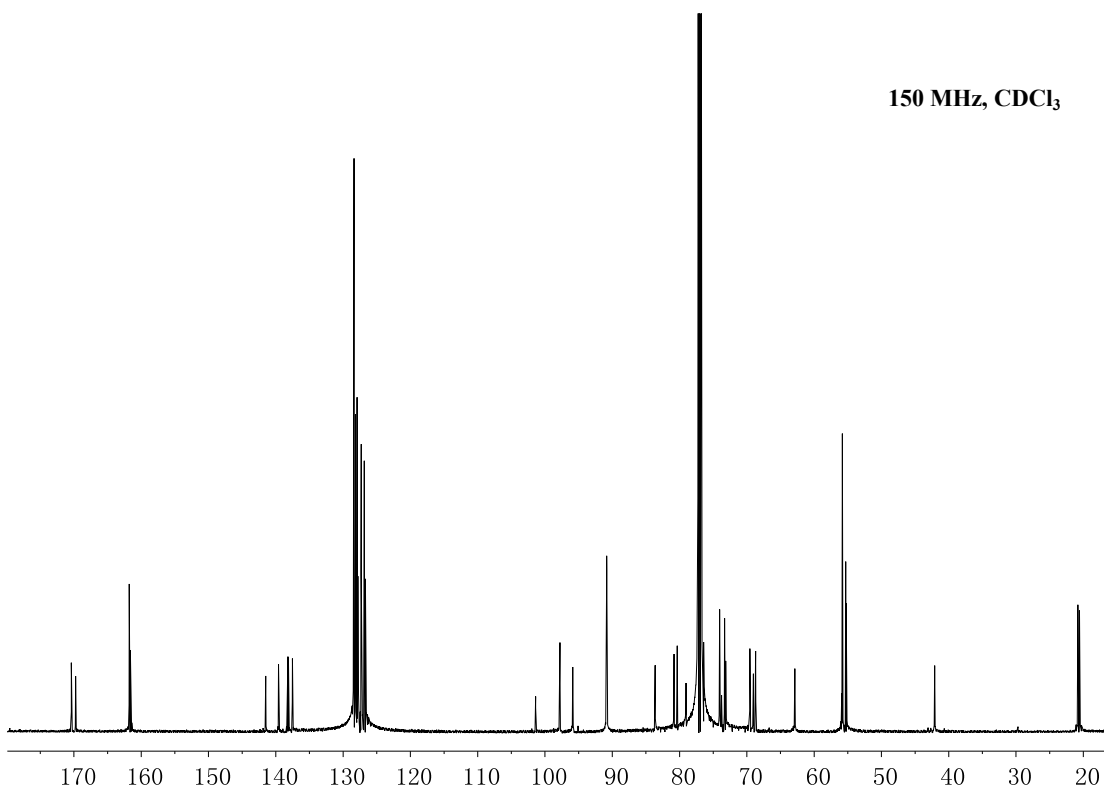


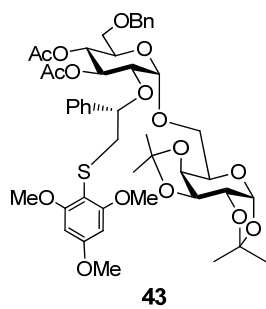


600 MHz, CDCl₃

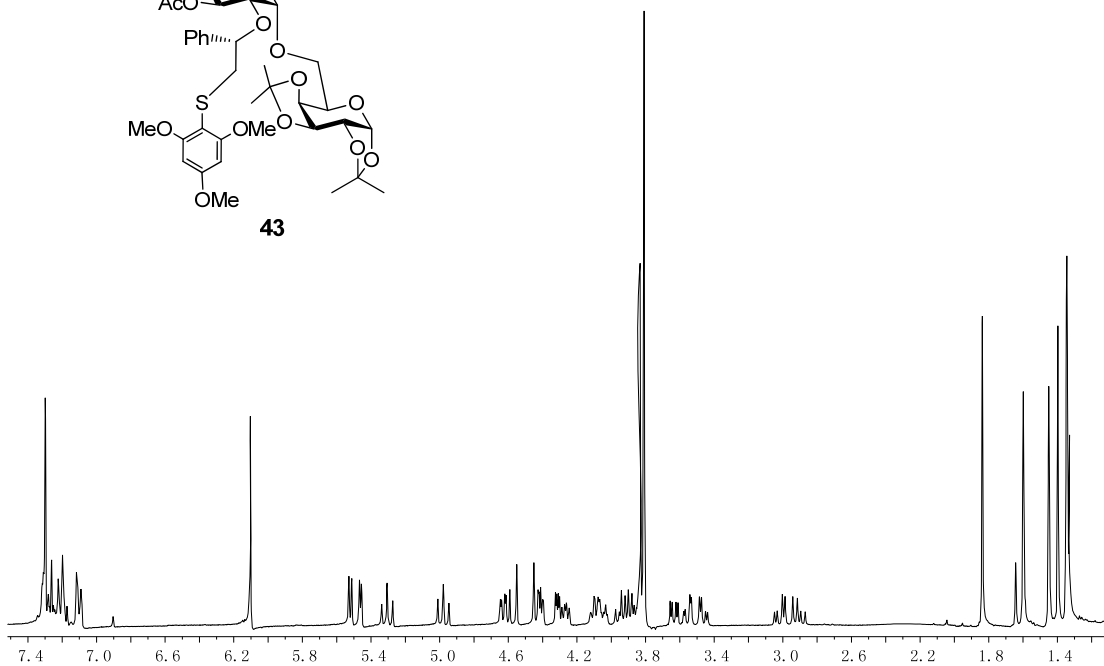


150 MHz, CDCl₃

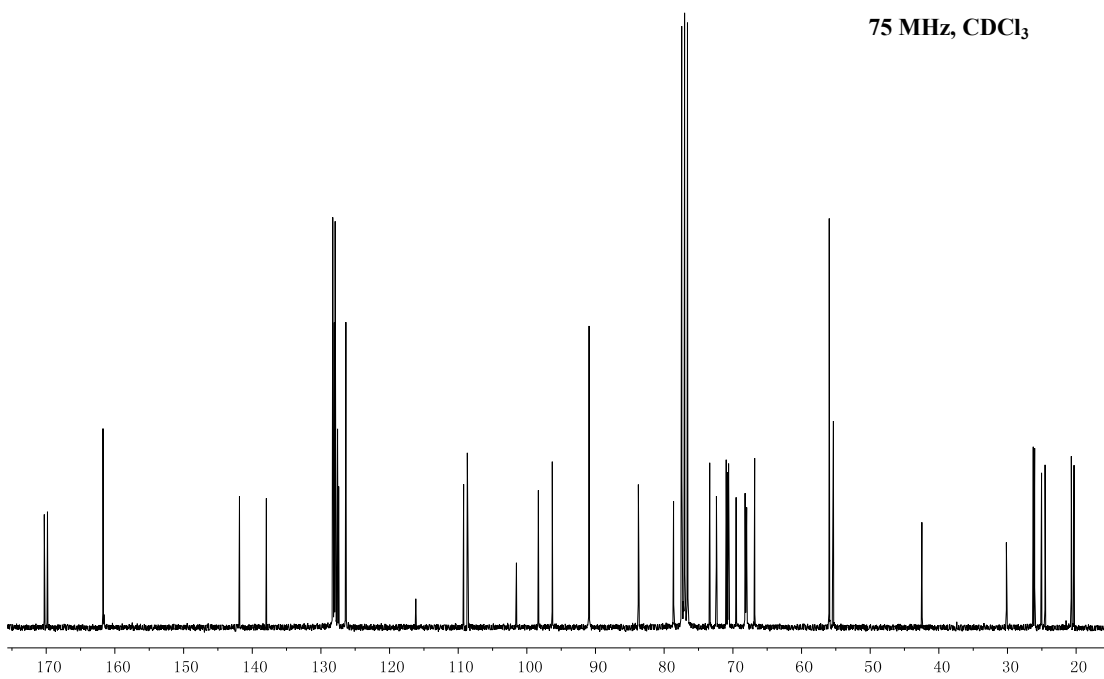


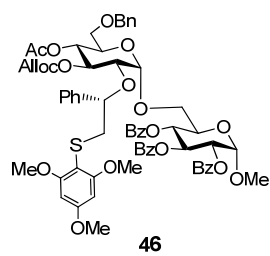


300 MHz, CDCl₃

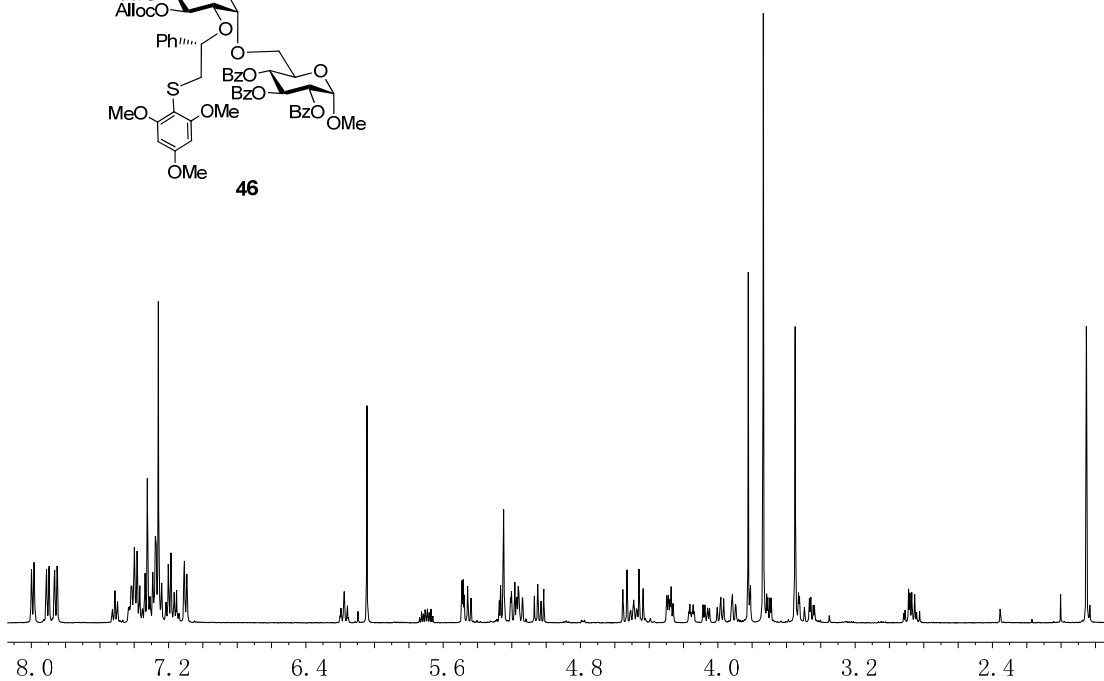


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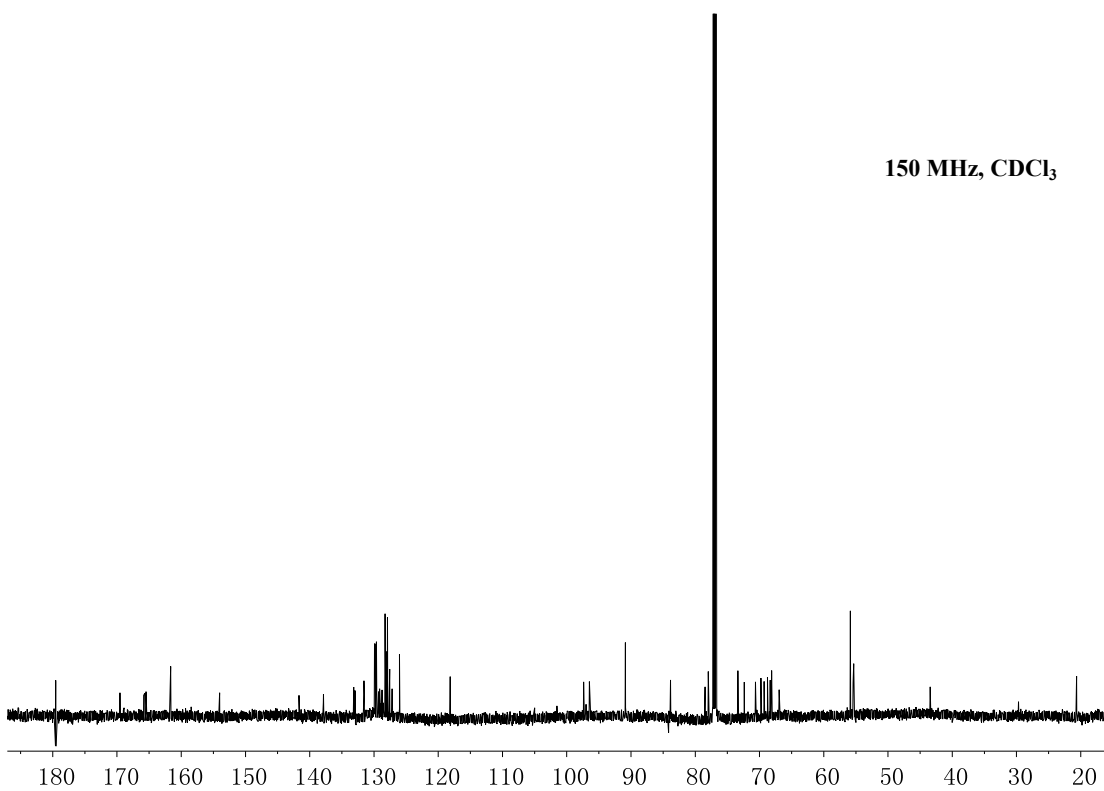


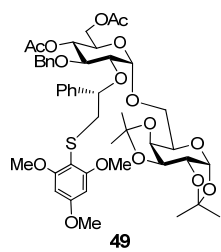


500 MHz, CDCl₃

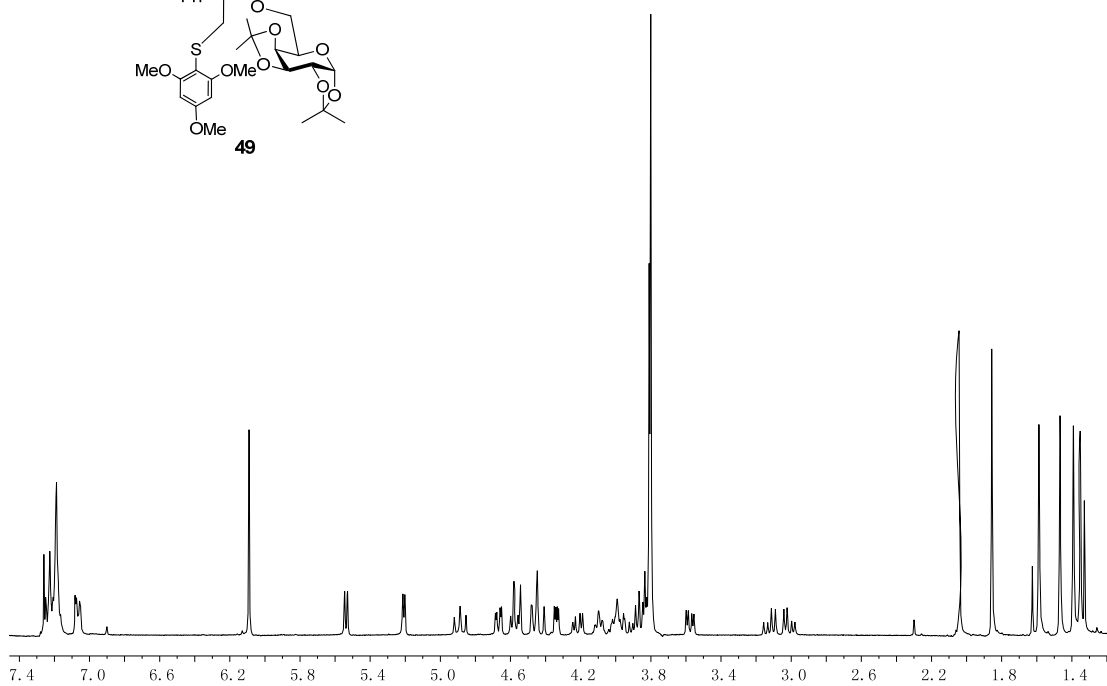


150 MHz, CDCl₃

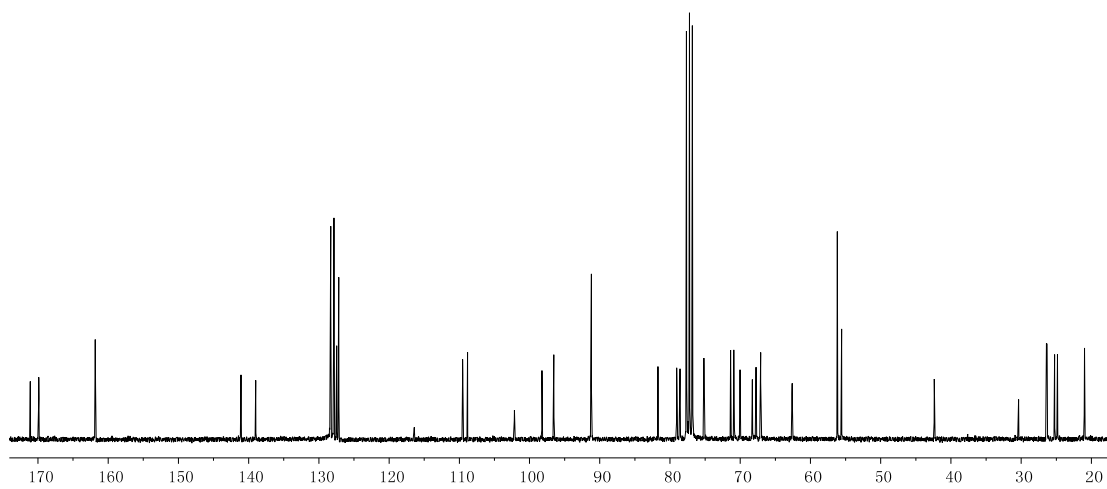


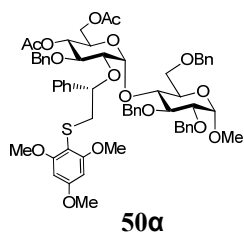


300 MHz, CDCl₃

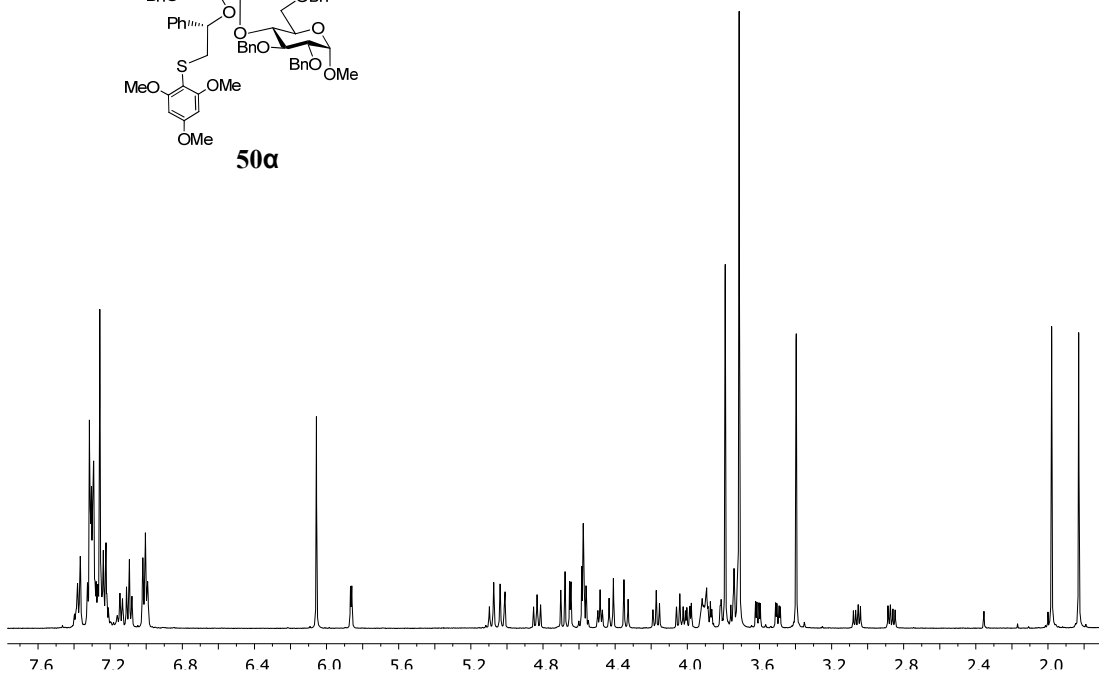


75 MHz, CDCl₃

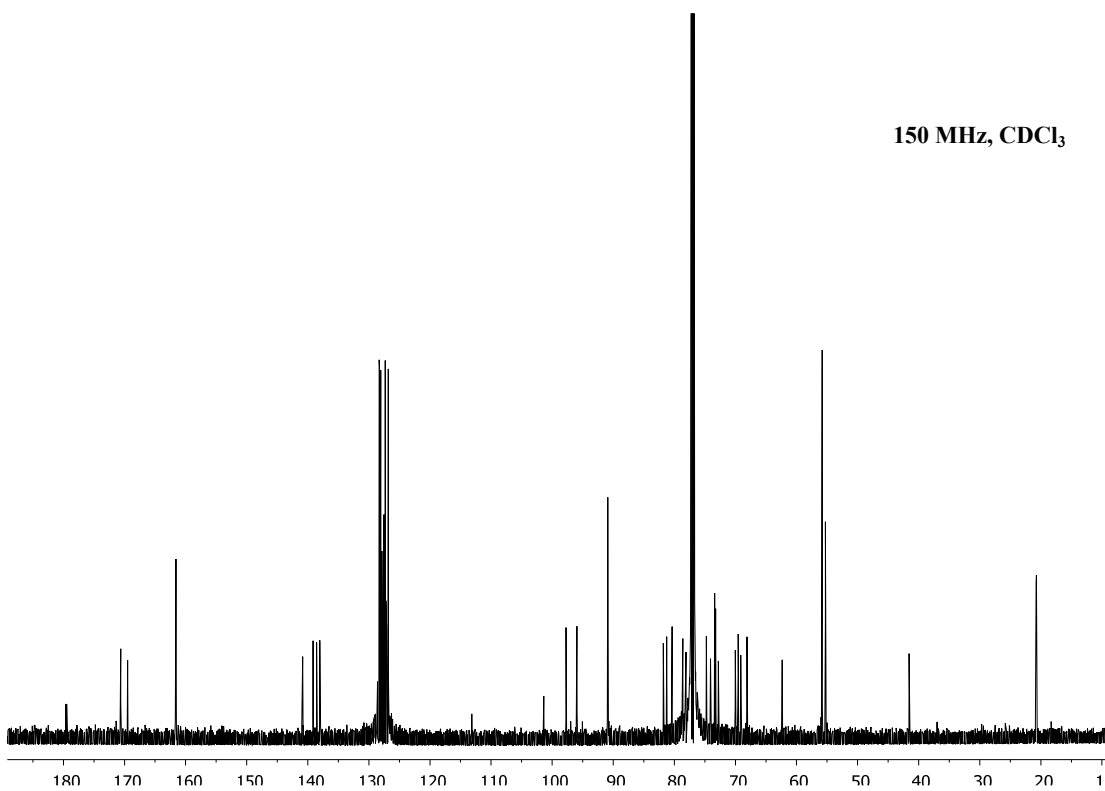


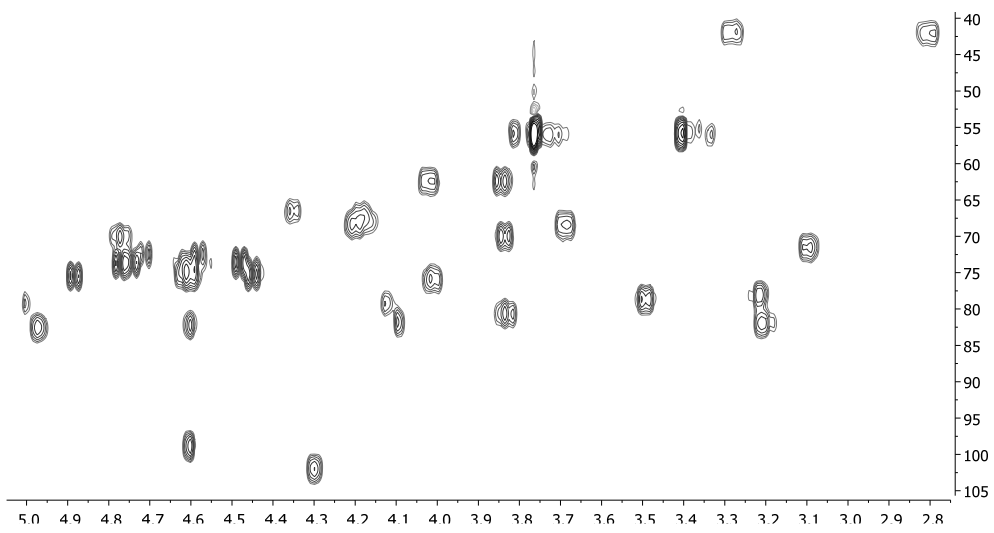
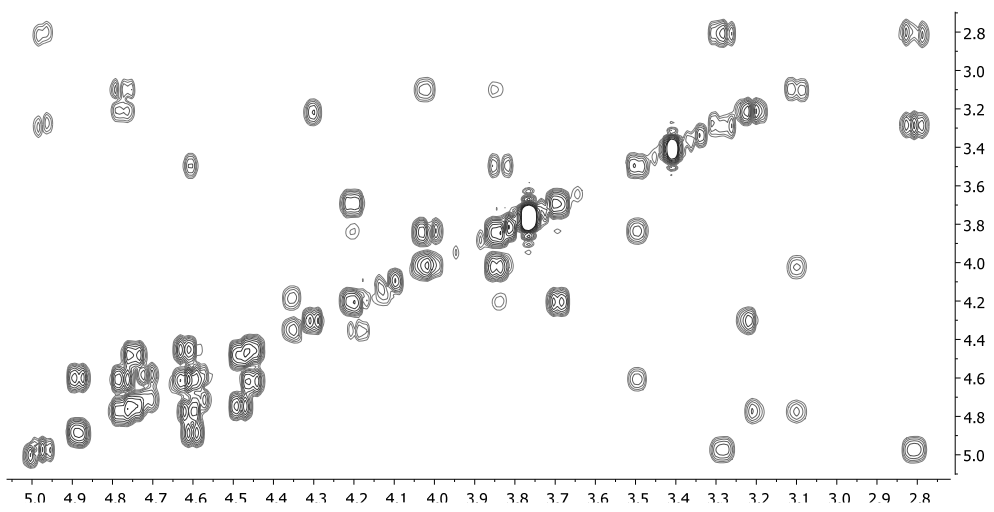
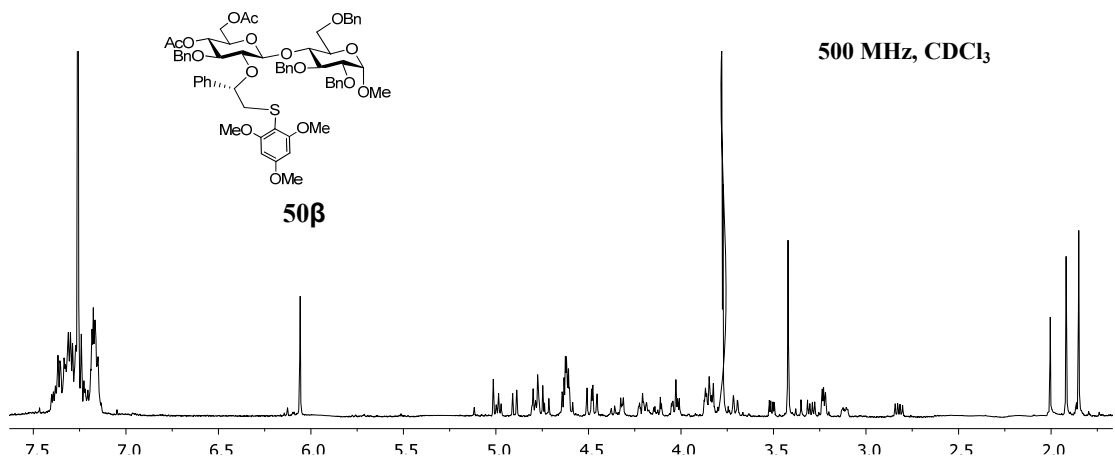


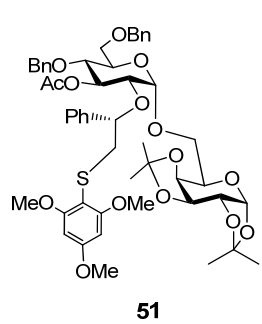
500 MHz, CDCl₃



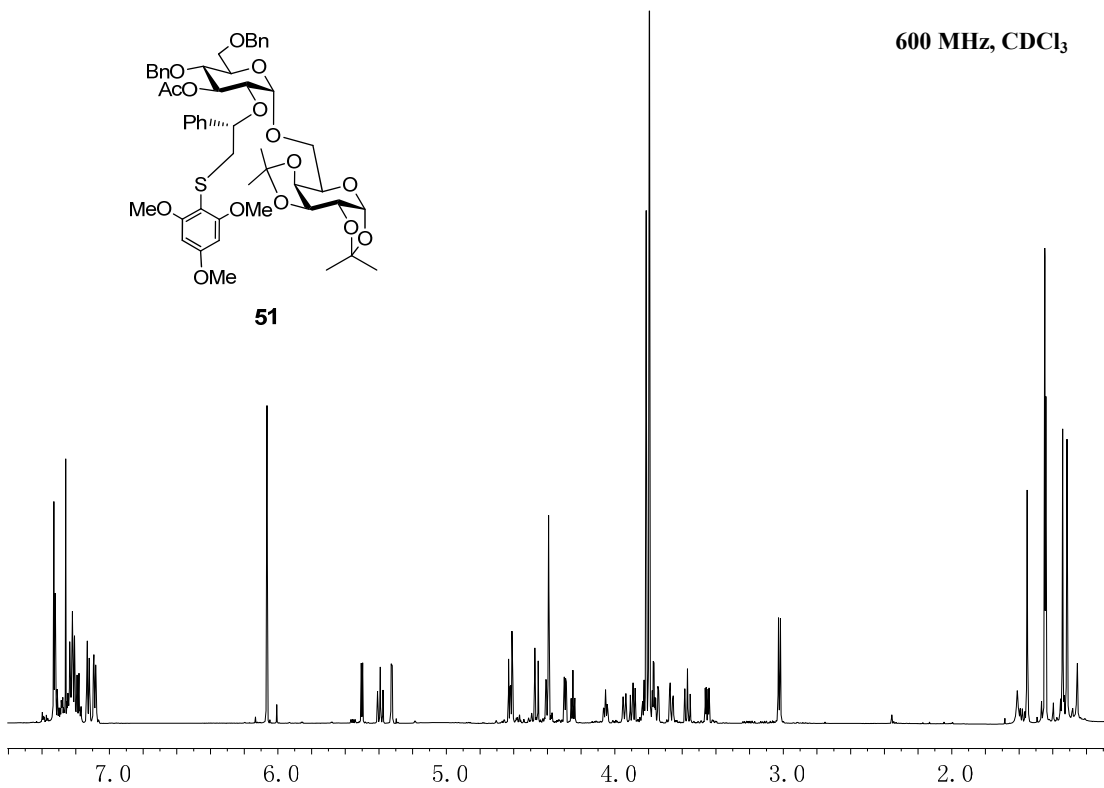
150 MHz, CDCl₃



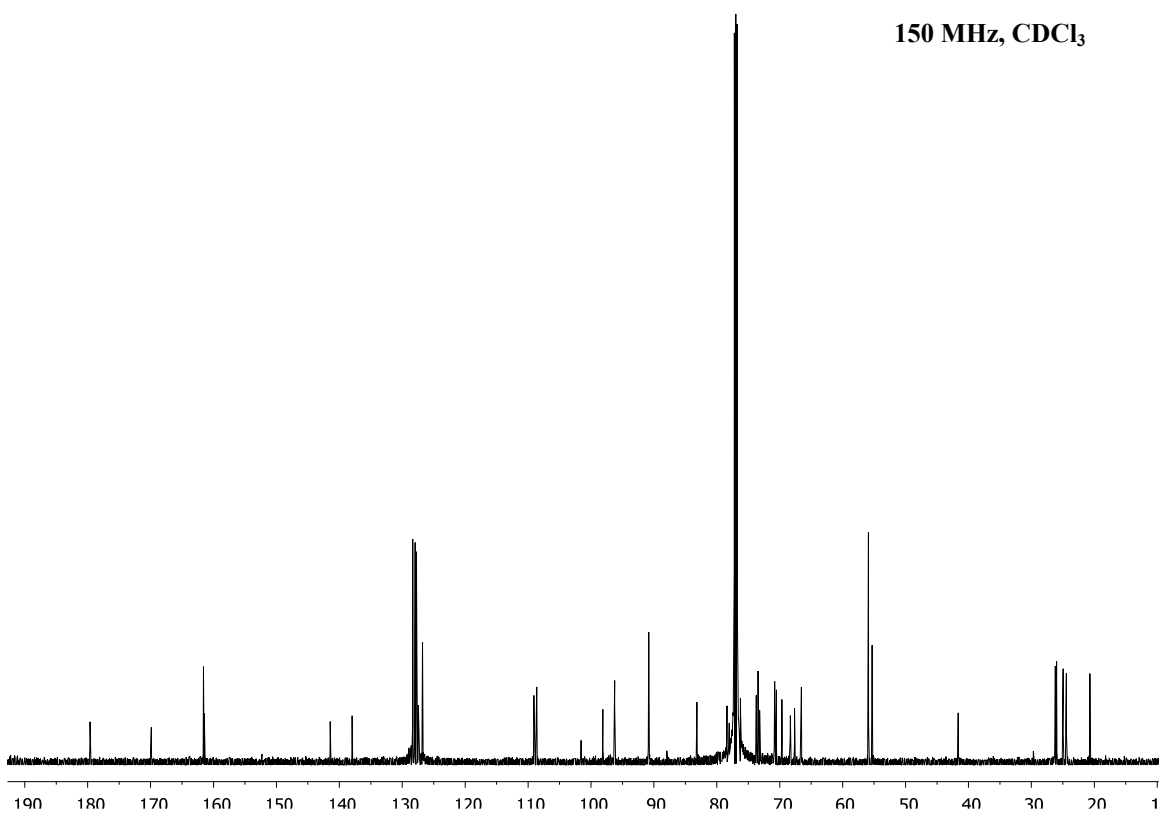


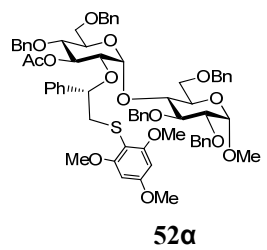


600 MHz, CDCl₃

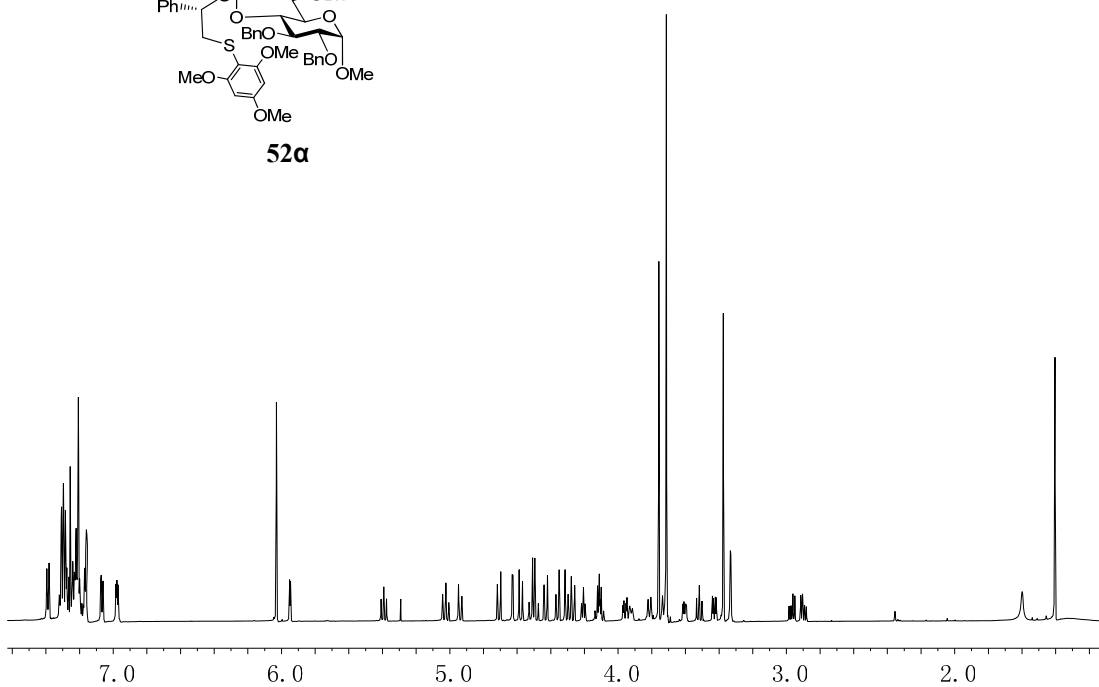


150 MHz, CDCl₃

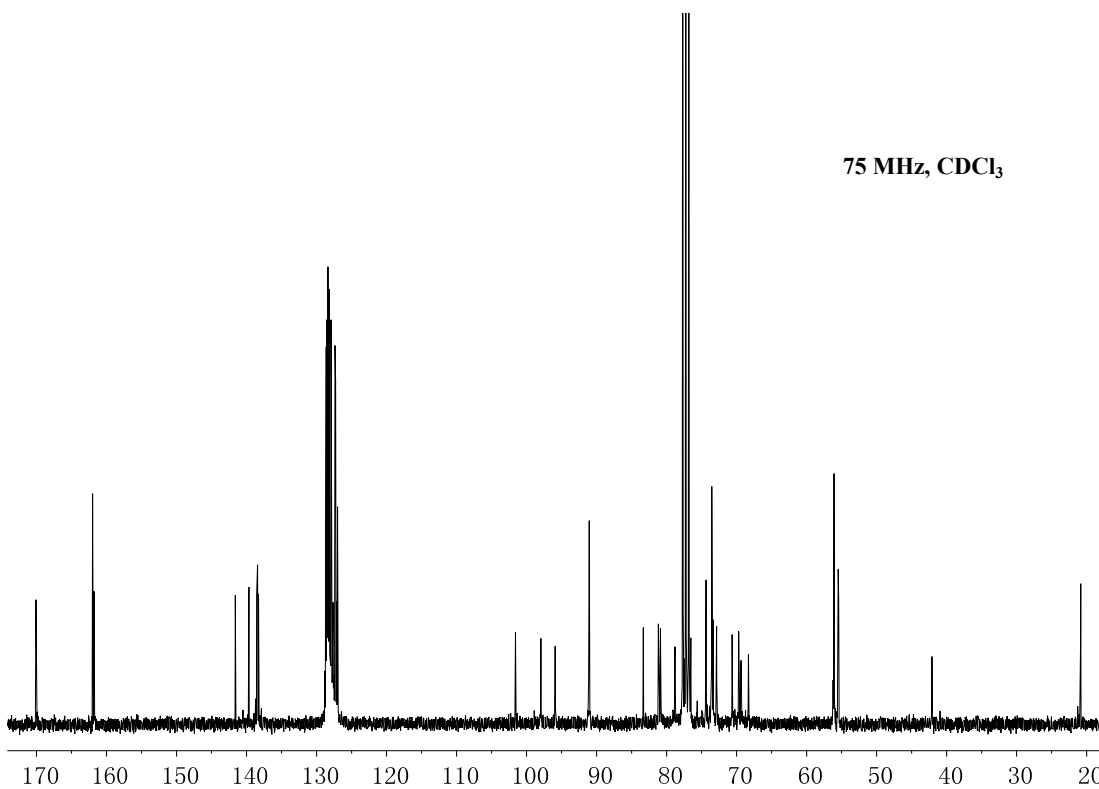


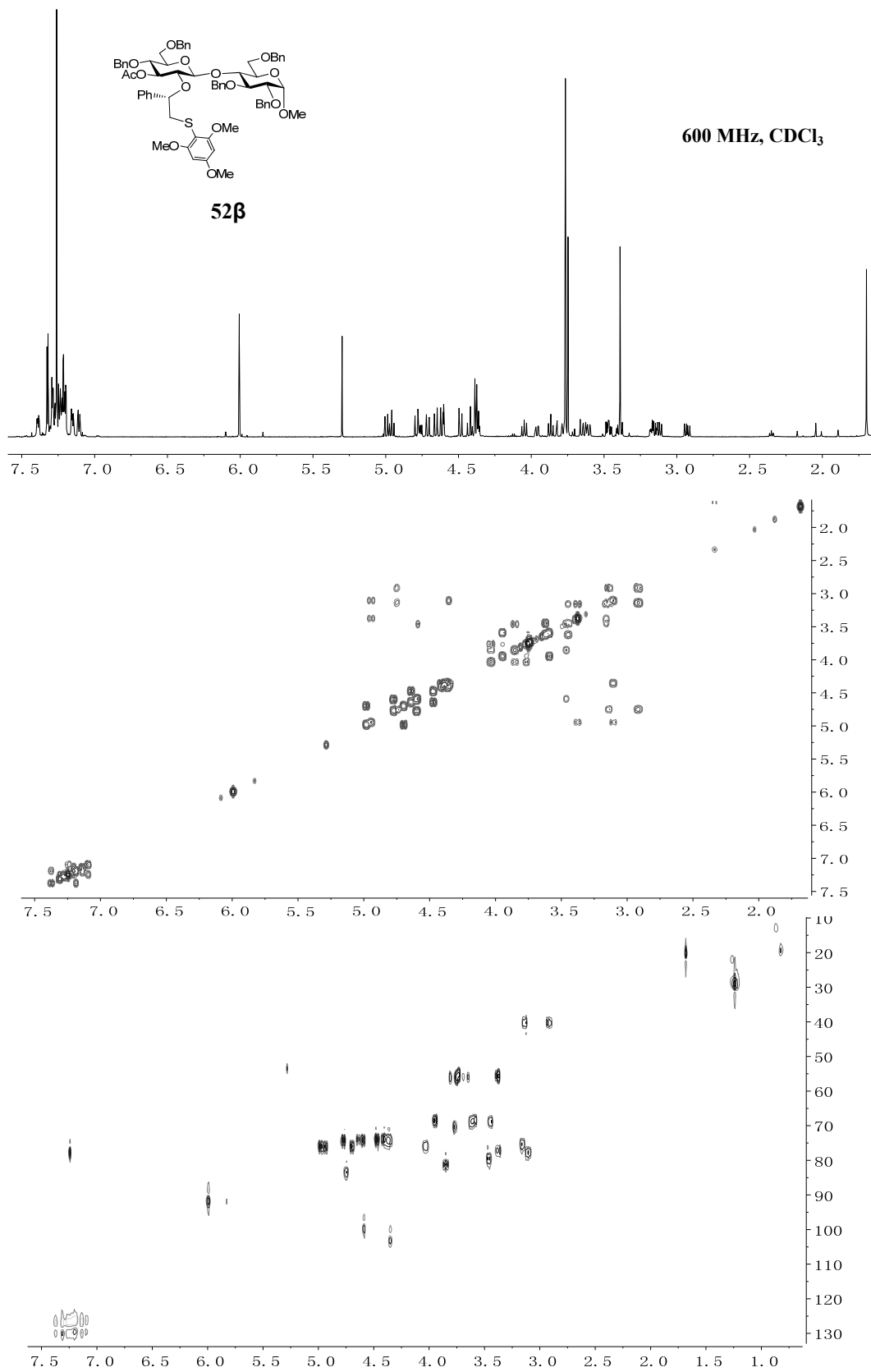


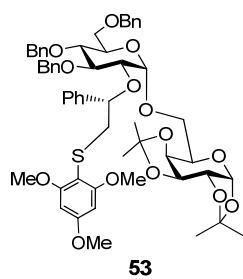
600 MHz, CDCl₃



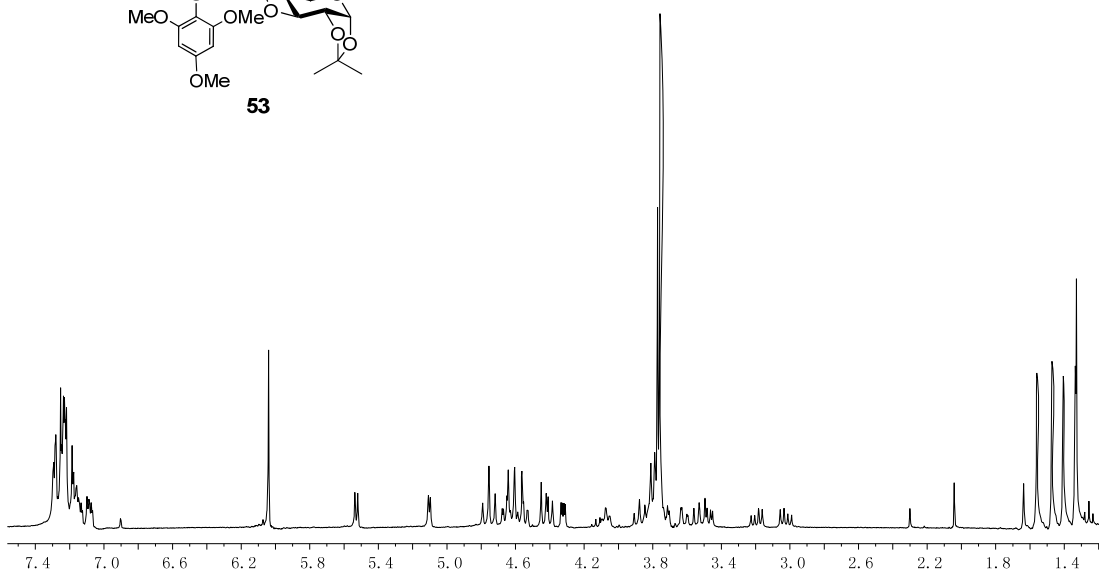
75 MHz, CDCl₃



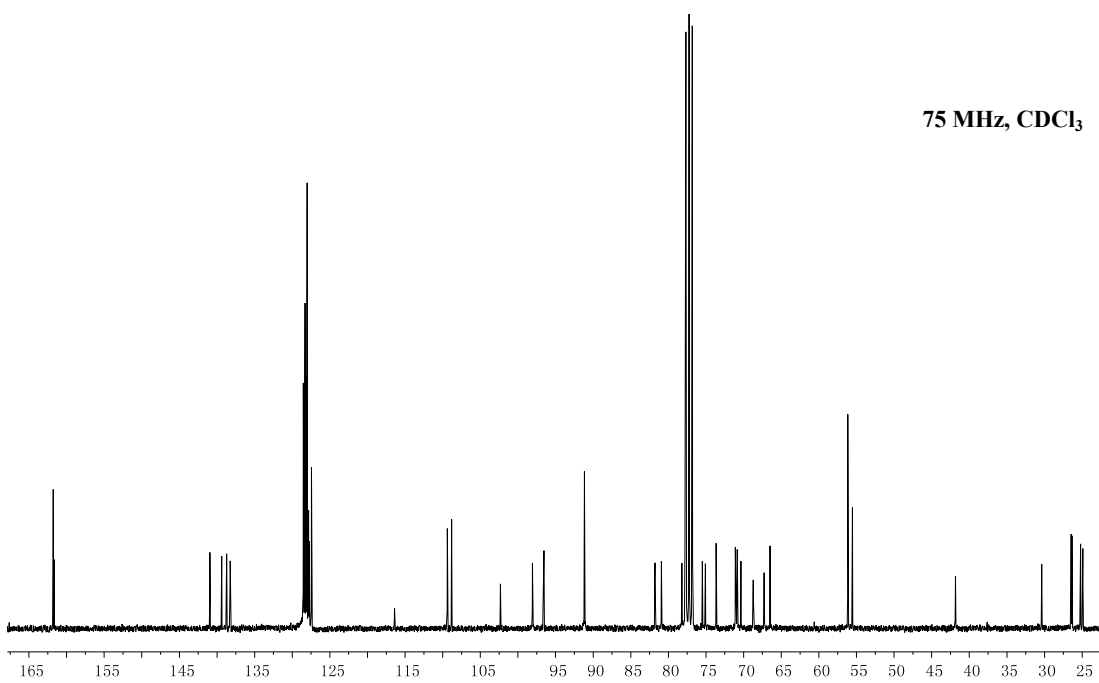


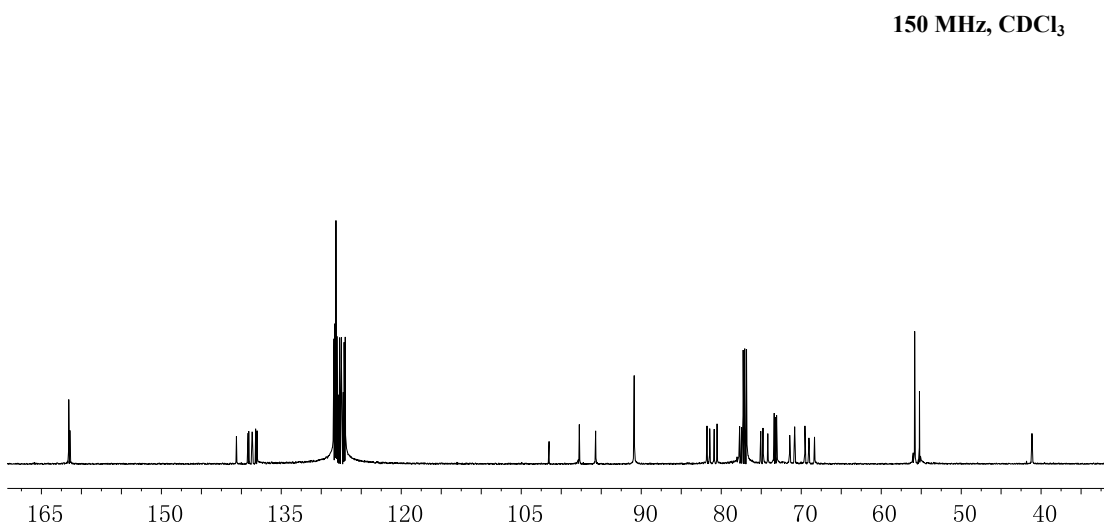
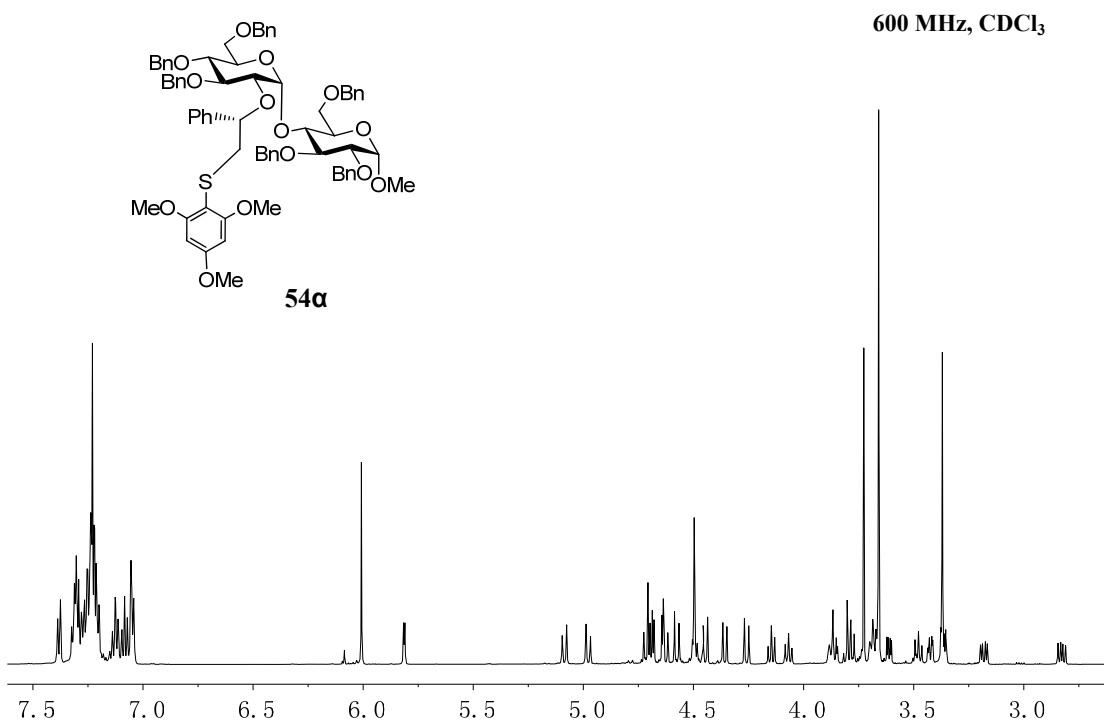


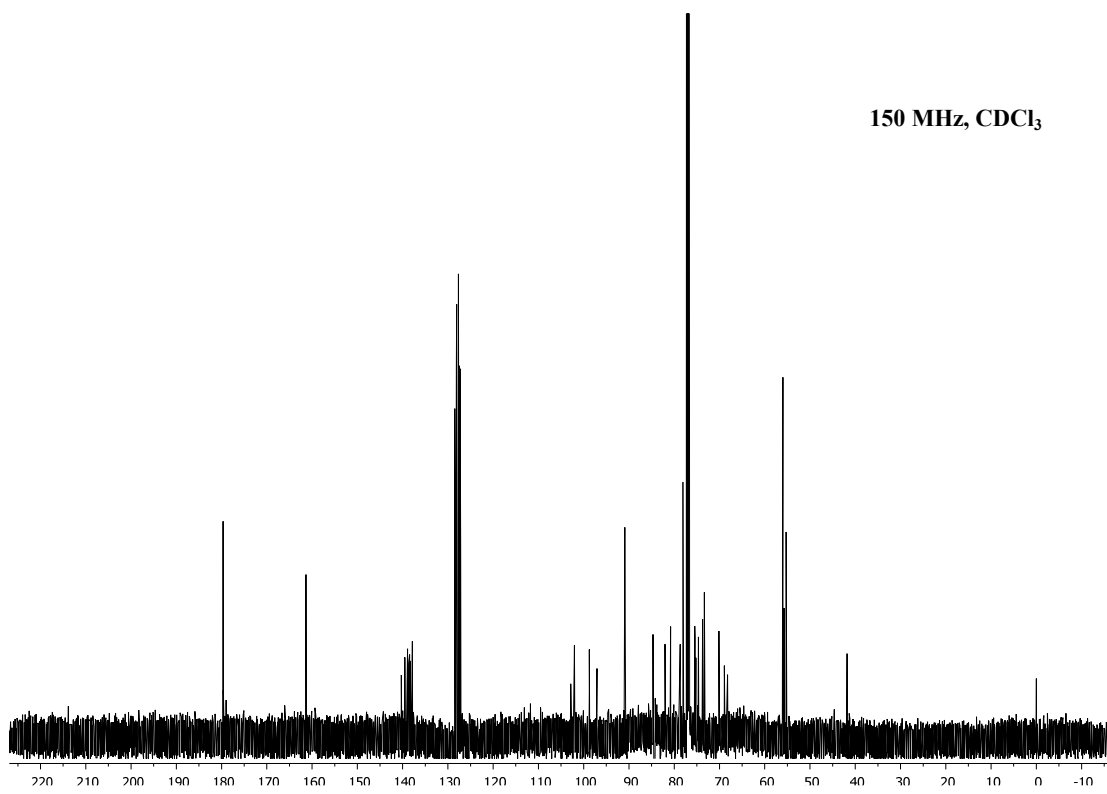
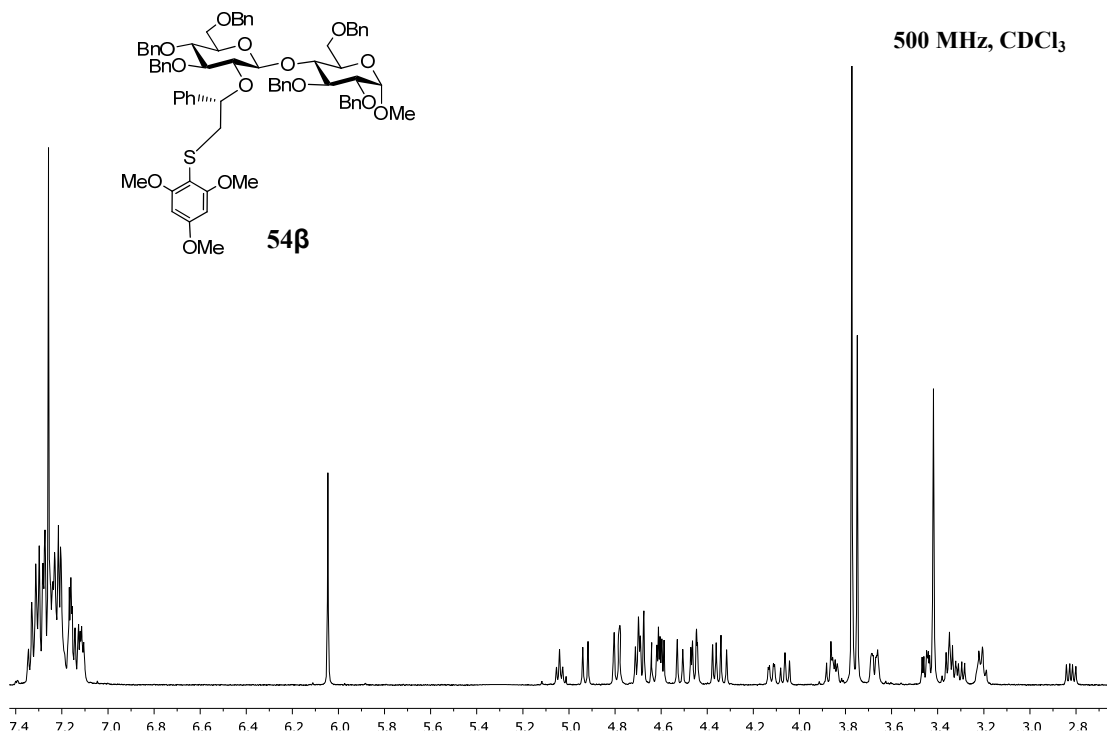
300 MHz, CDCl₃

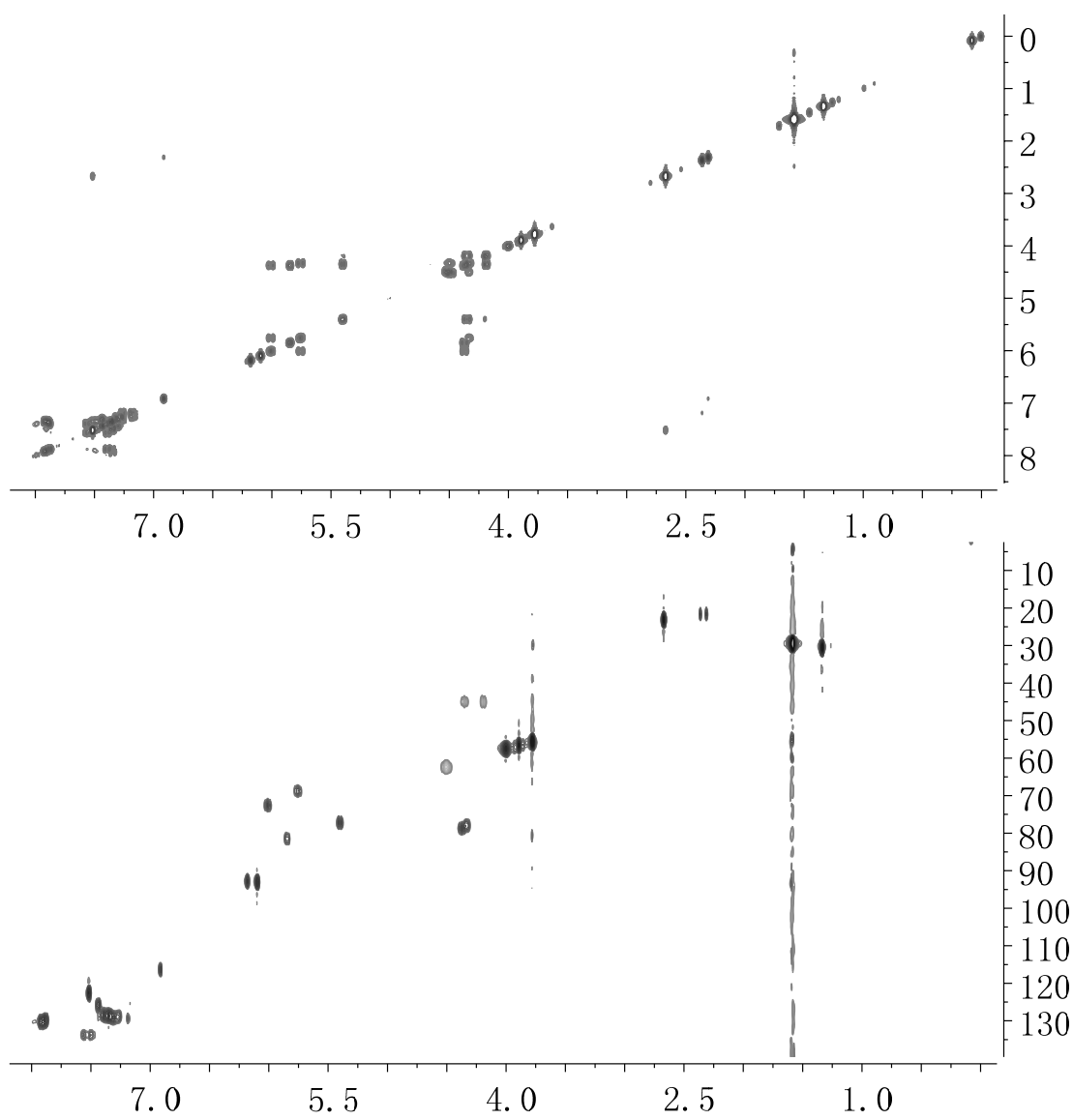
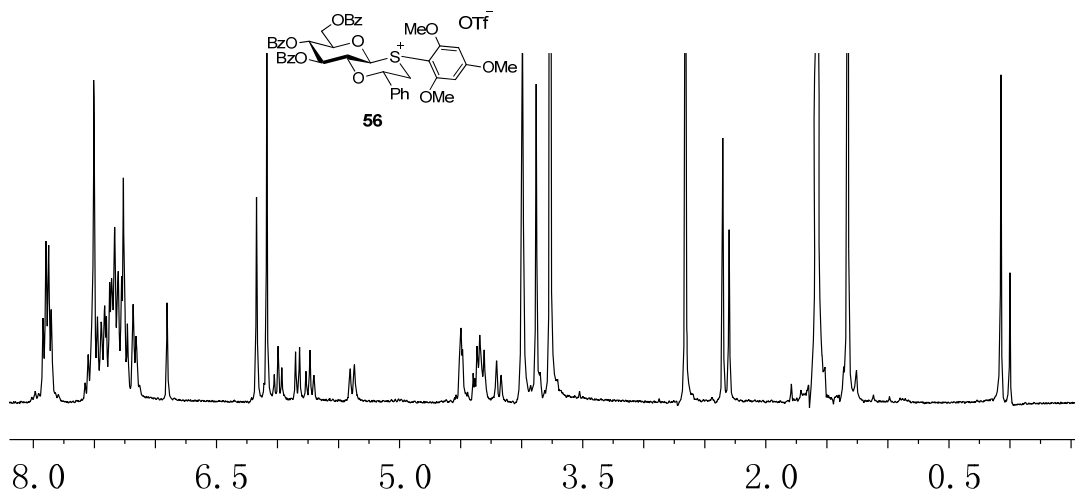


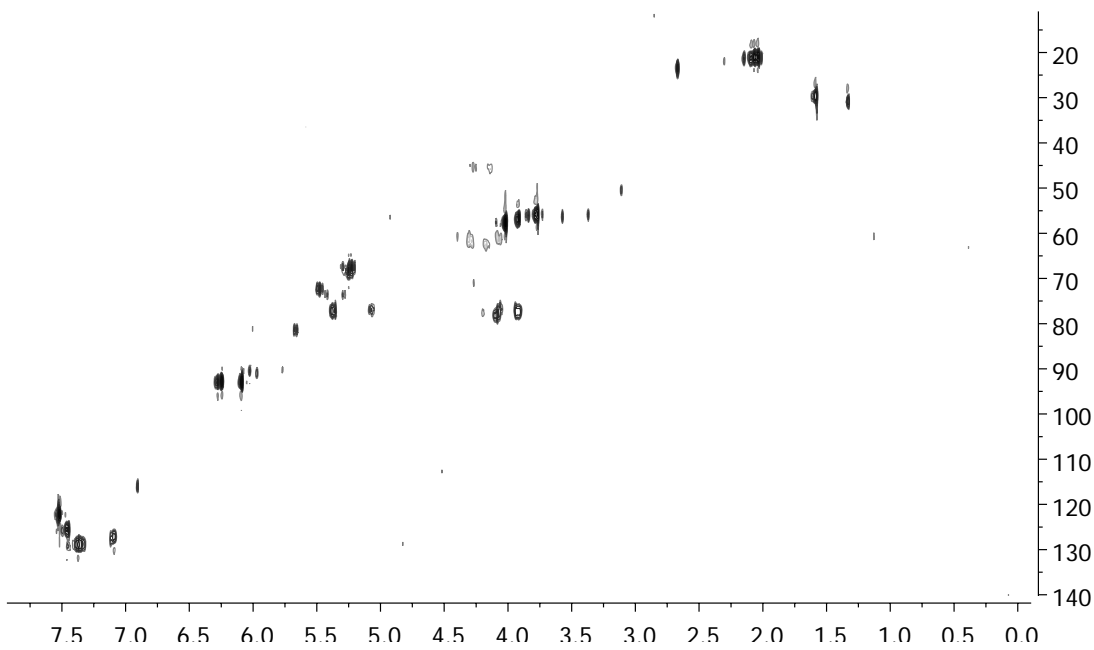
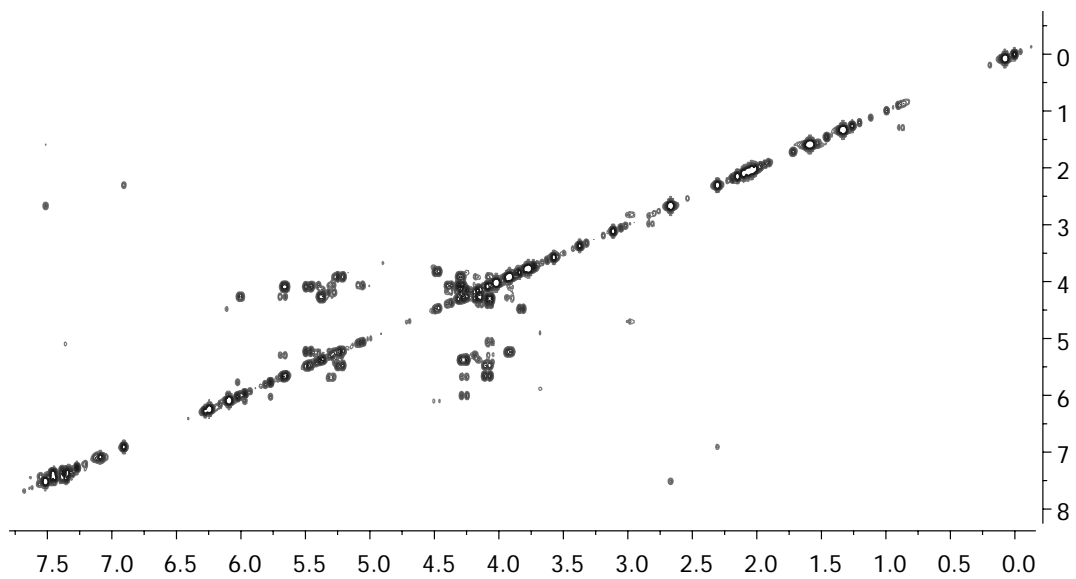
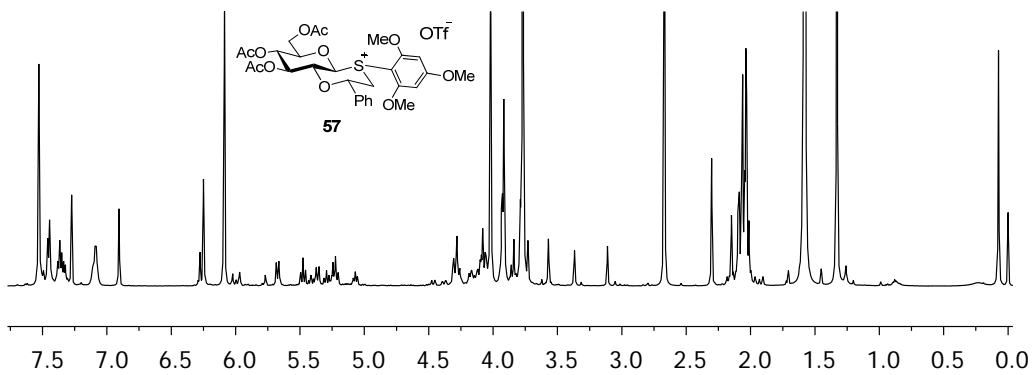
75 MHz, CDCl₃

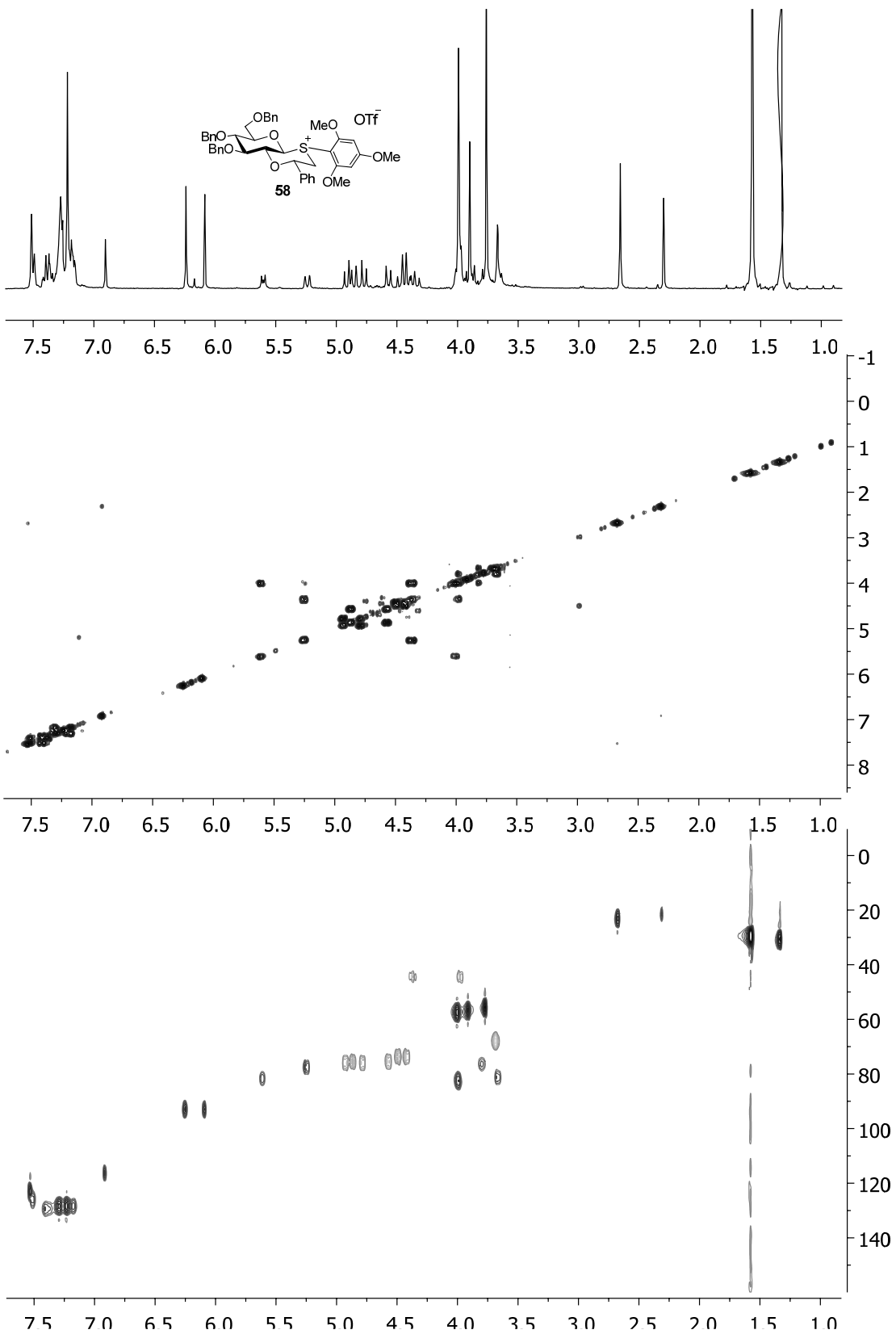


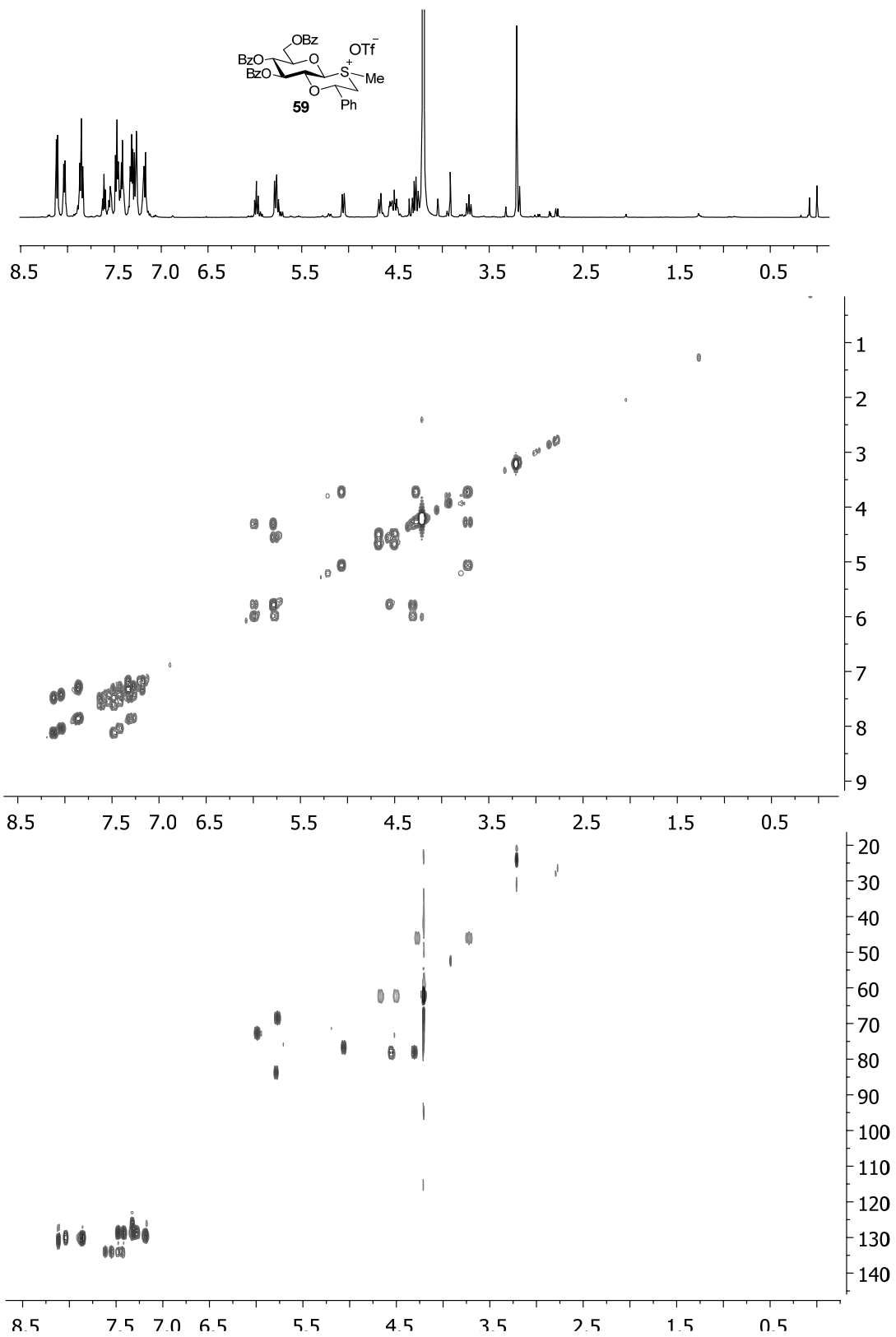


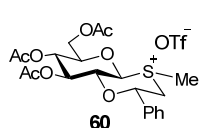




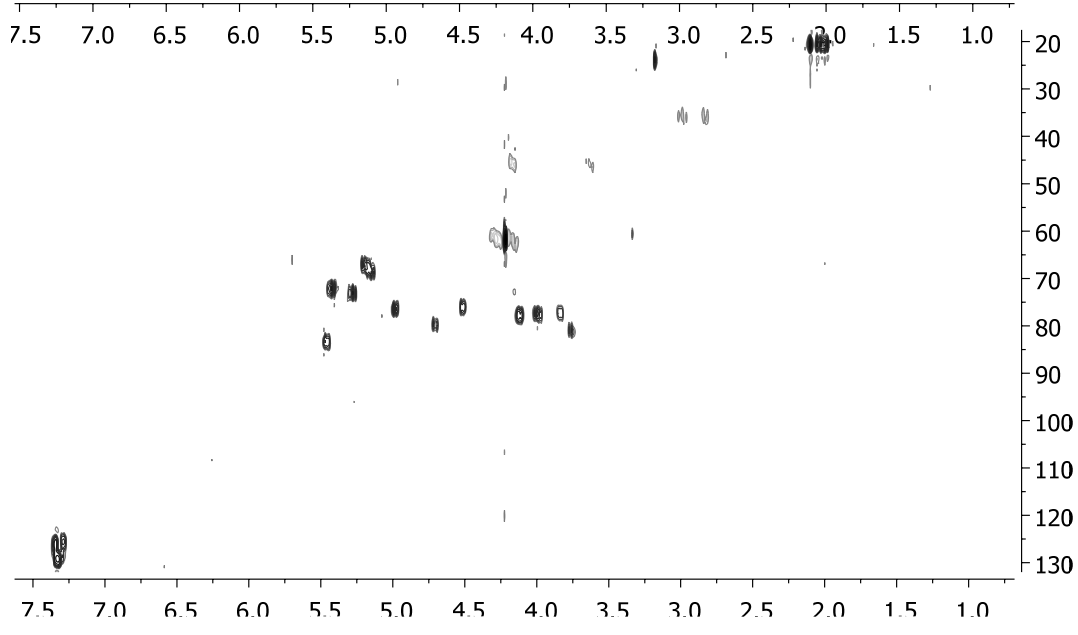
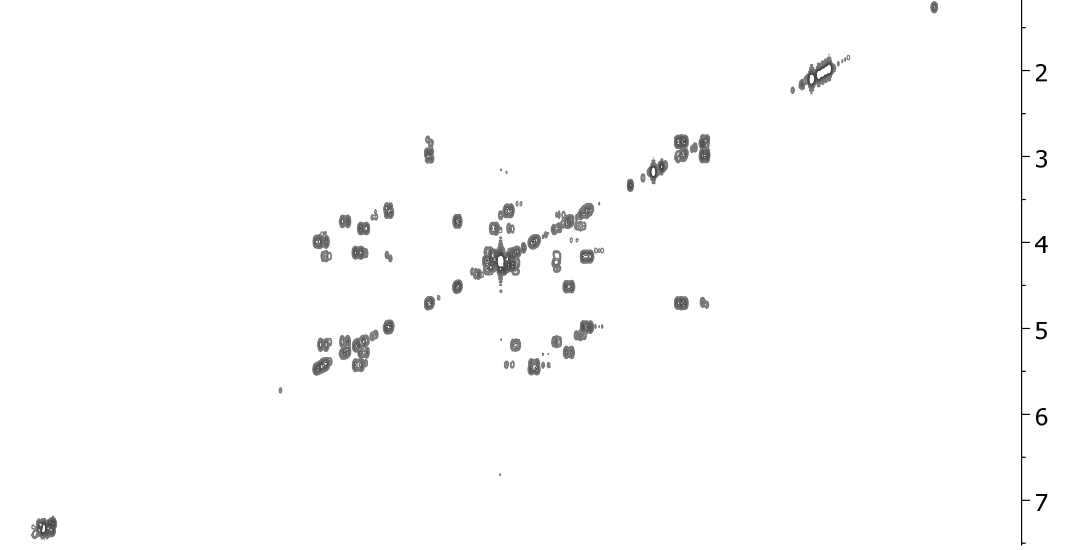
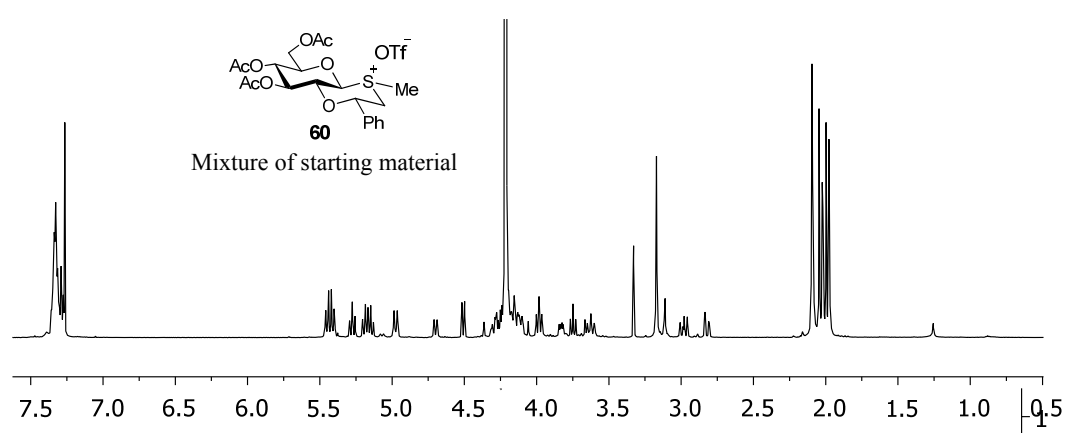


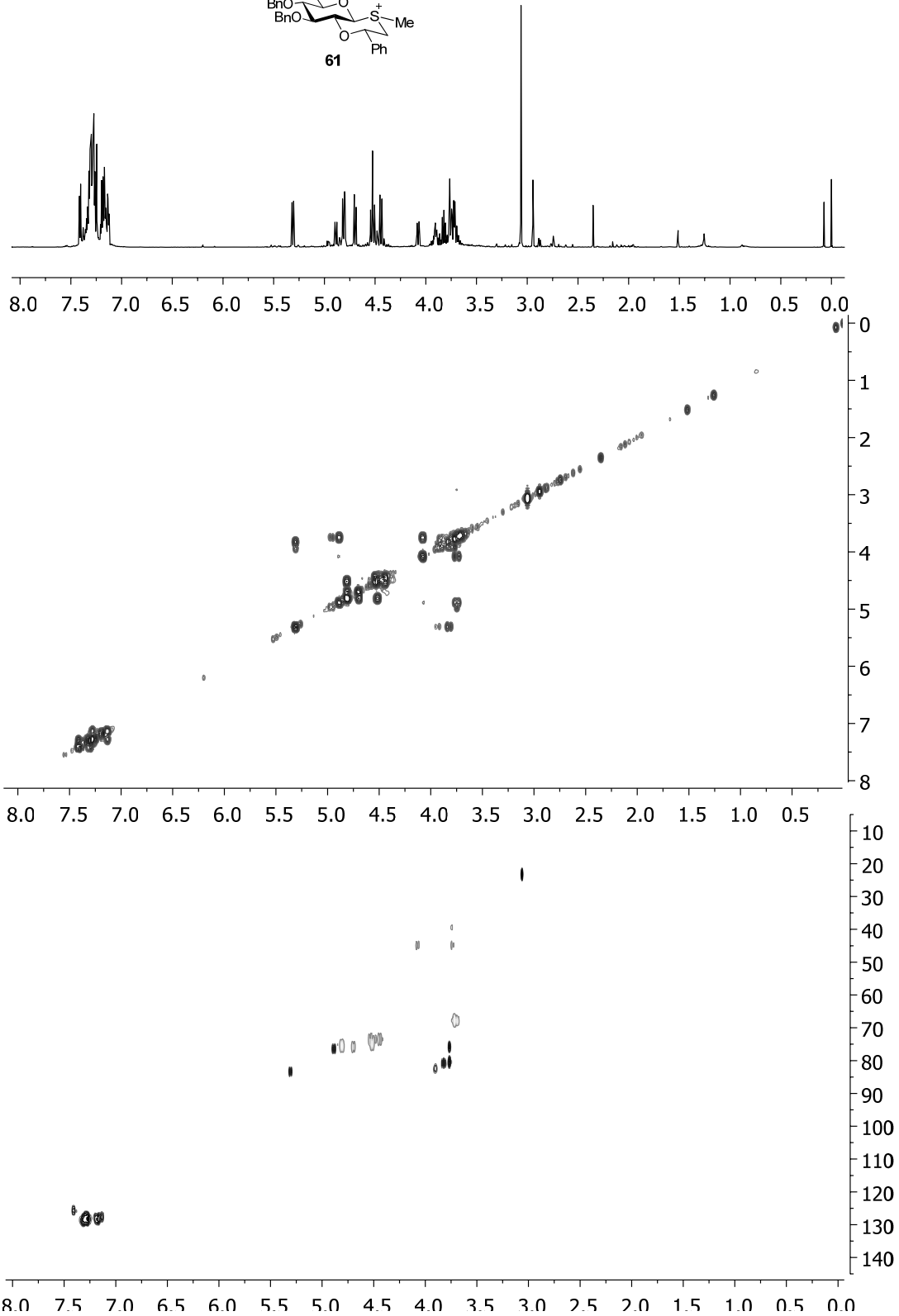
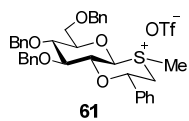


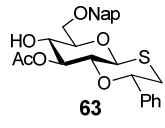




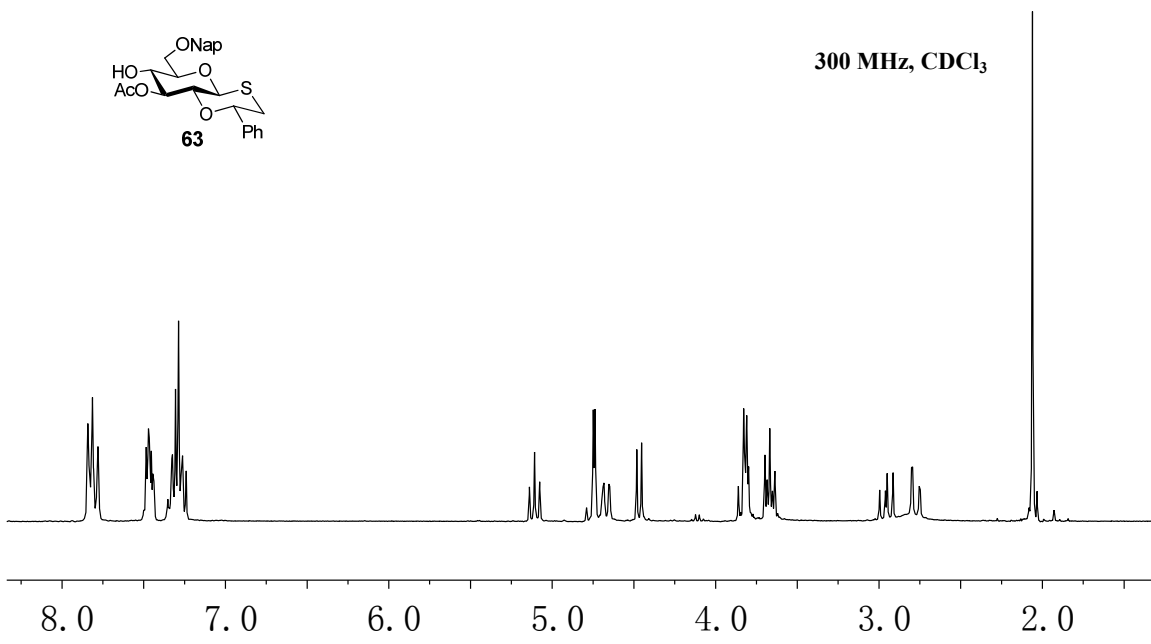
Mixture of starting material



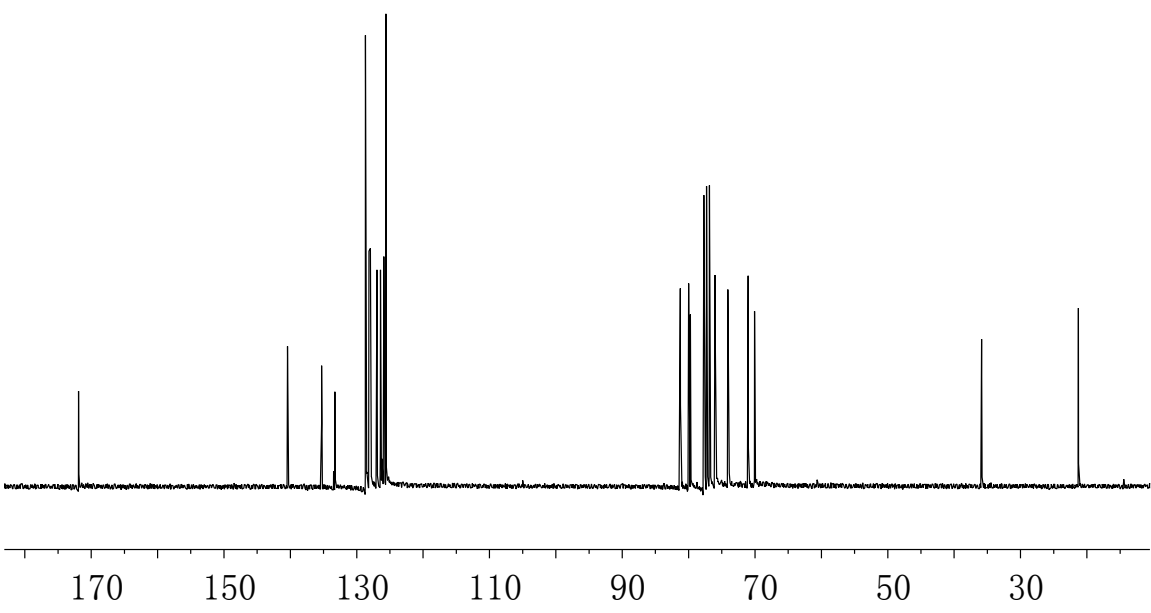


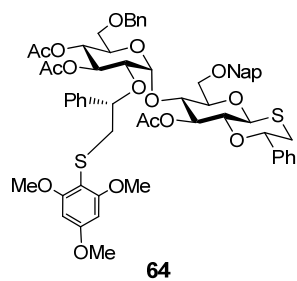


300 MHz, CDCl₃

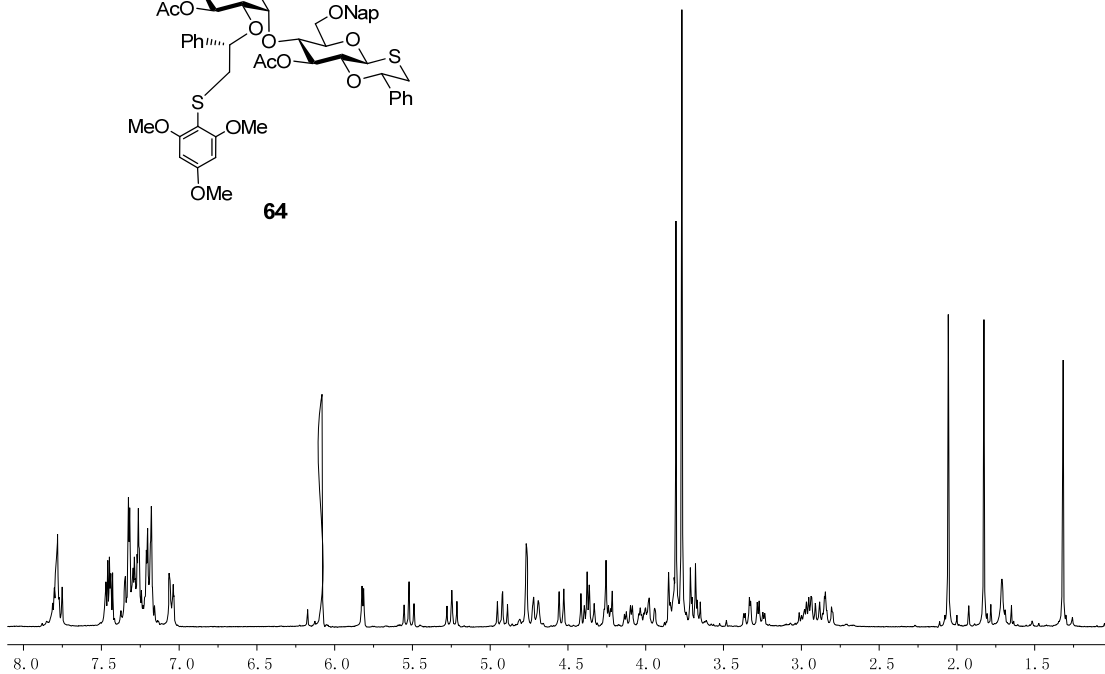


75 MHz, CDCl₃

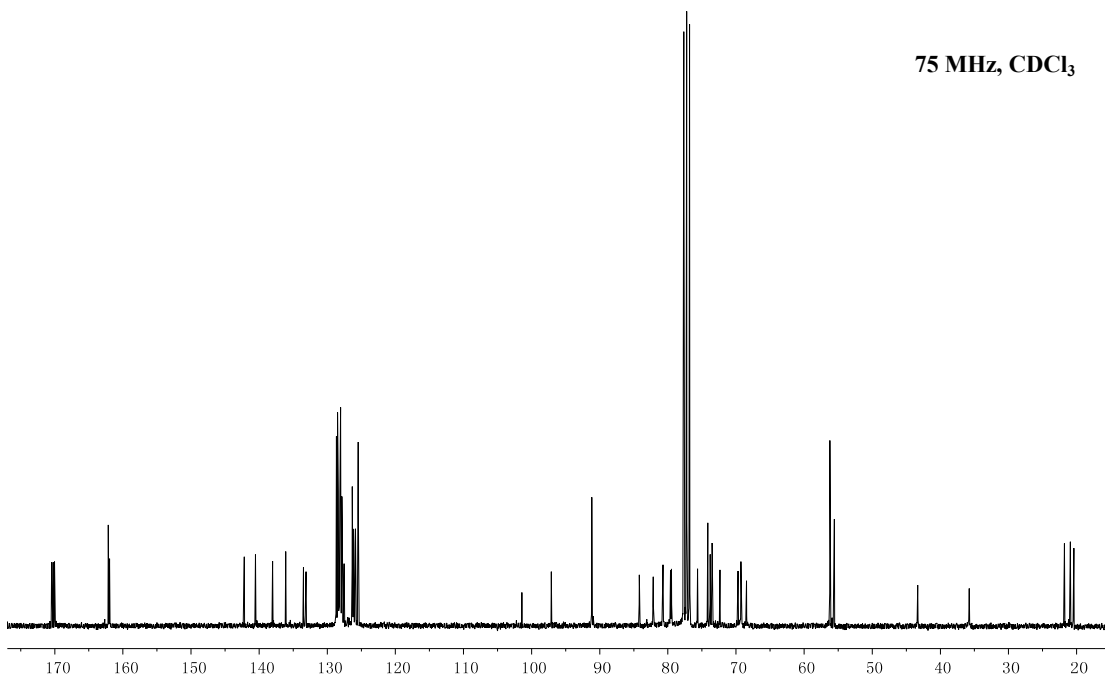


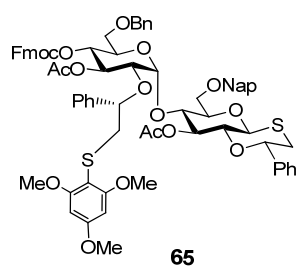


300 MHz, CDCl₃

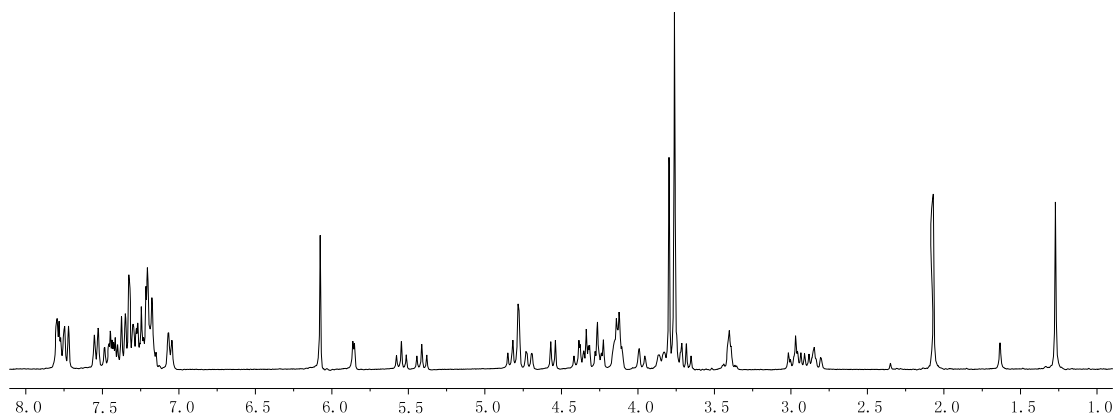


75 MHz, CDCl₃

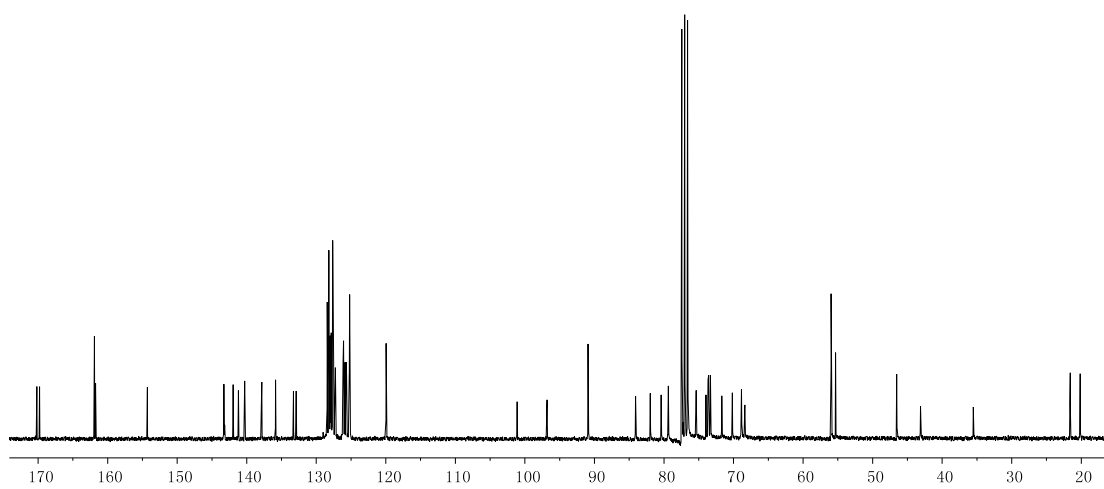


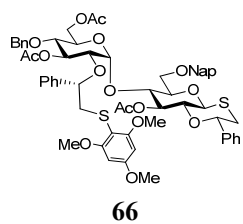


300 MHz, CDCl₃

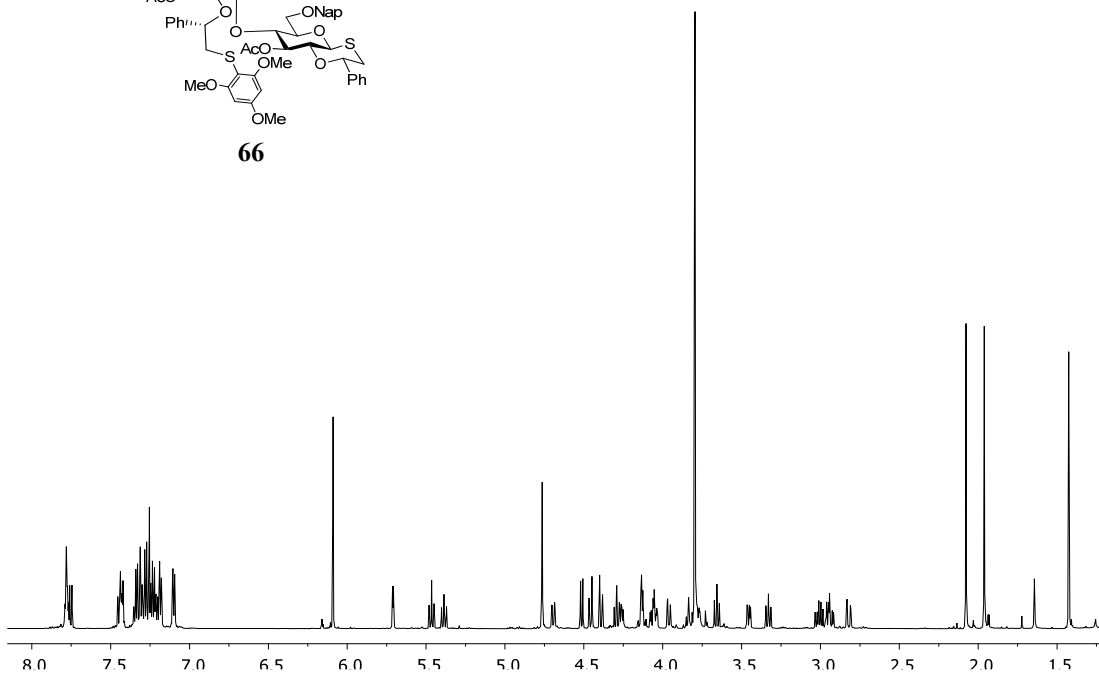


75 MHz, CDCl₃

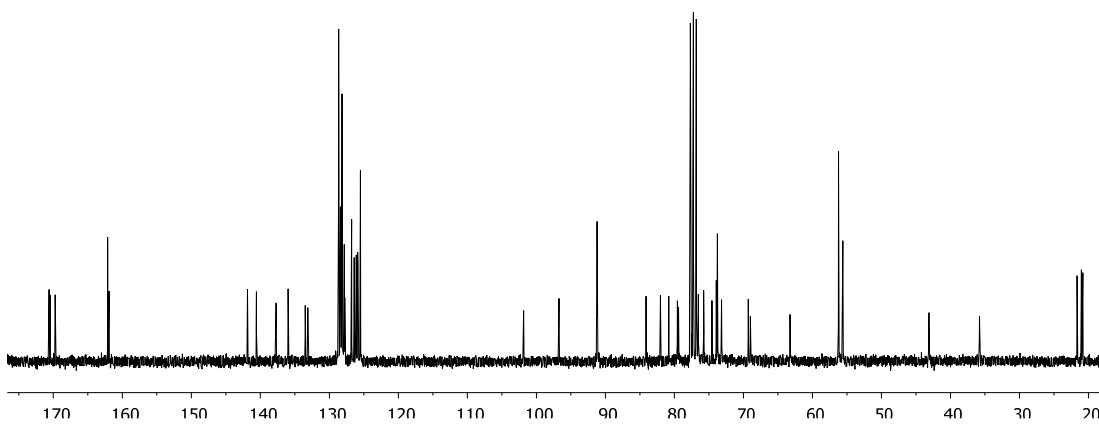


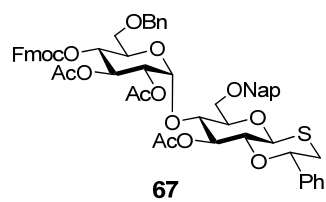


600 MHz, CDCl₃

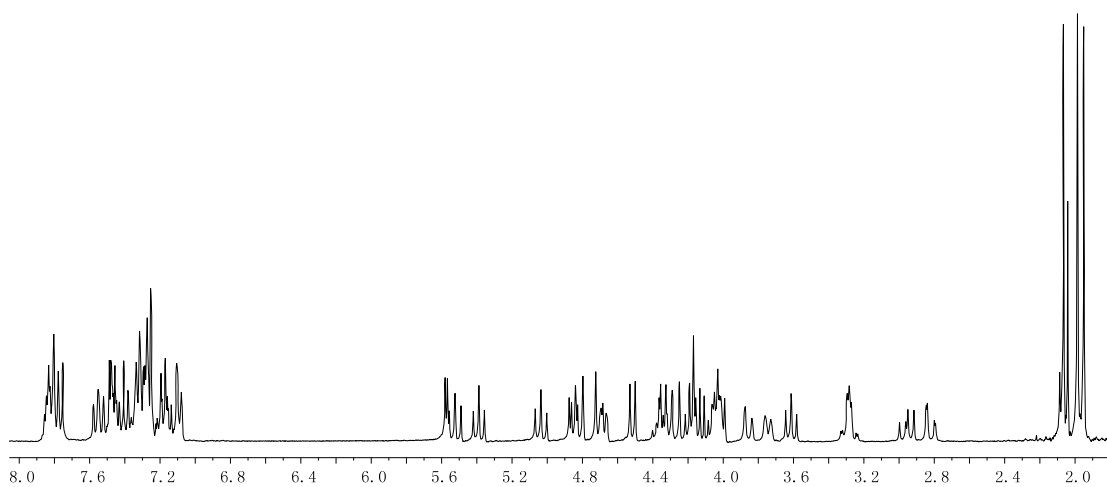


75 MHz, CDCl₃

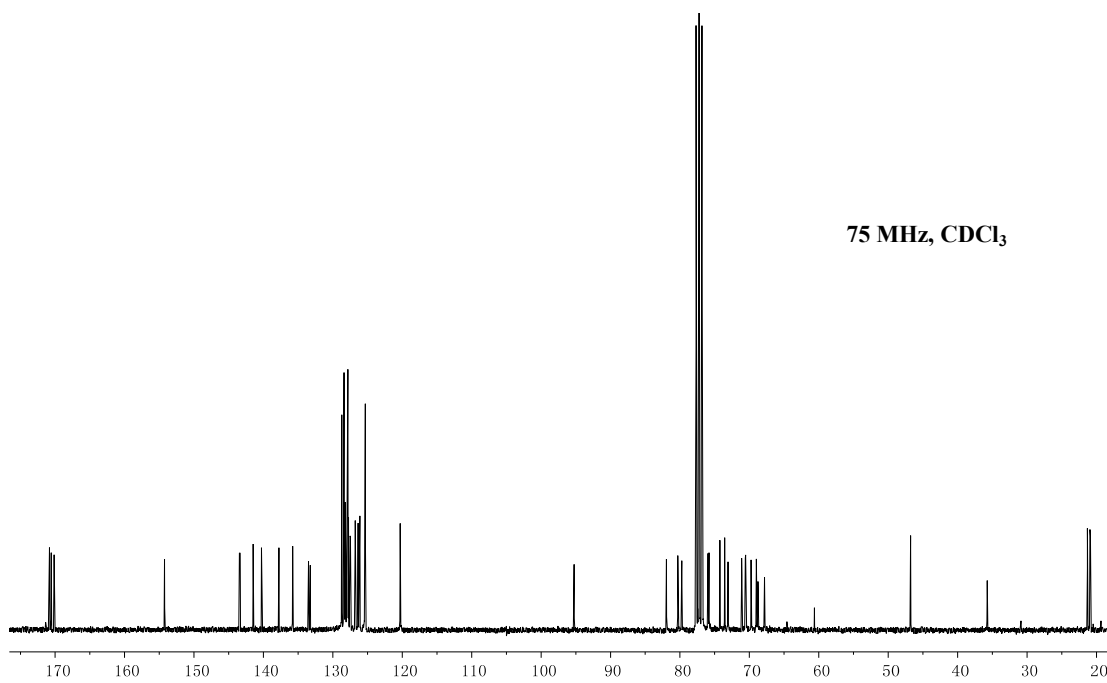


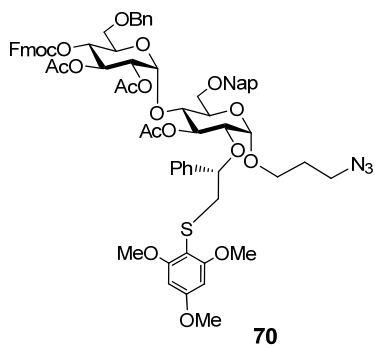


300 MHz, CDCl₃

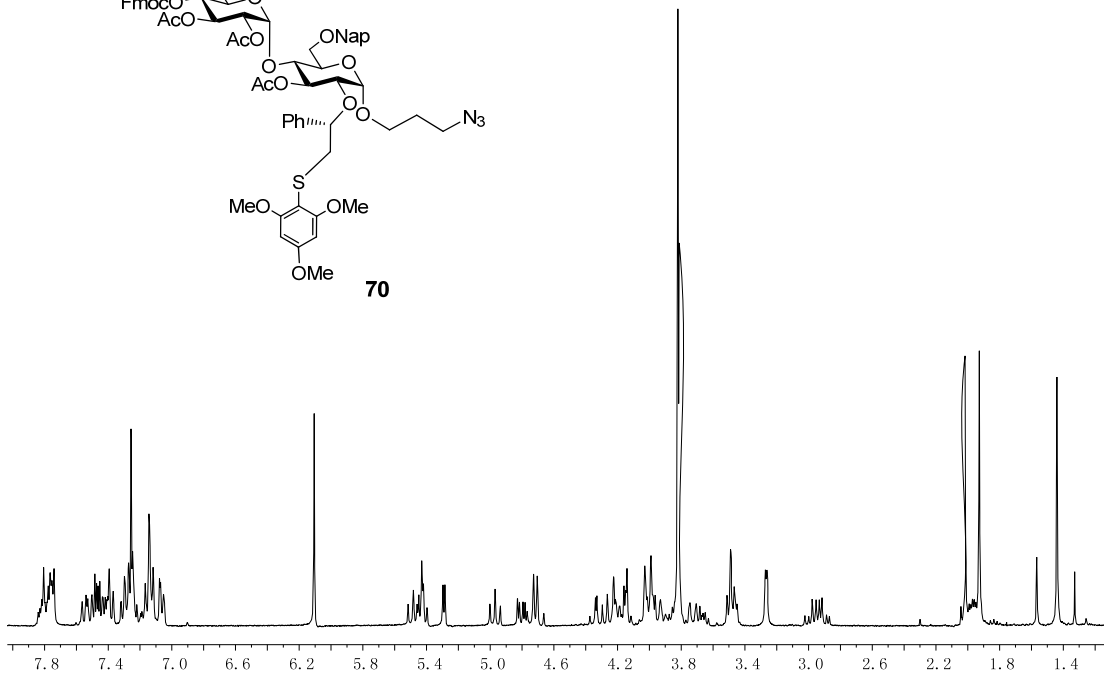


75 MHz, CDCl₃

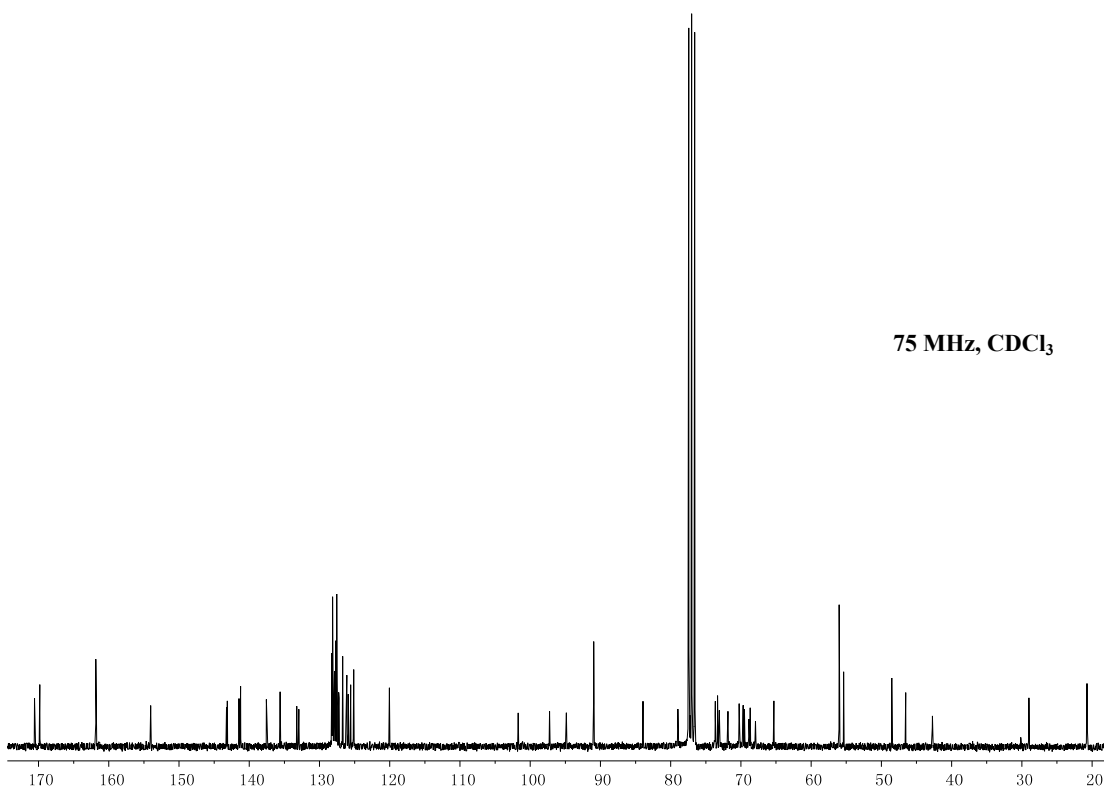


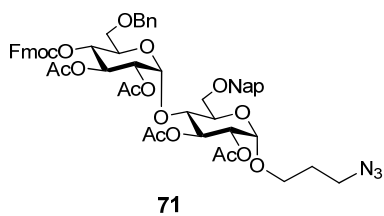


300 MHz, CDCl₃

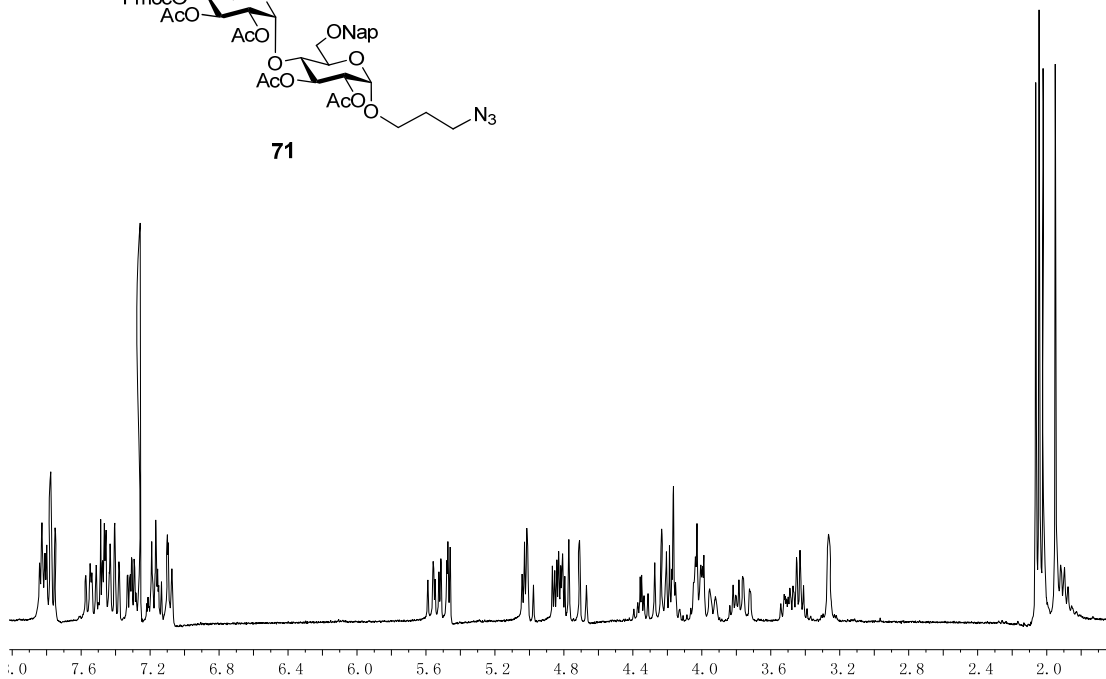


75 MHz, CDCl₃

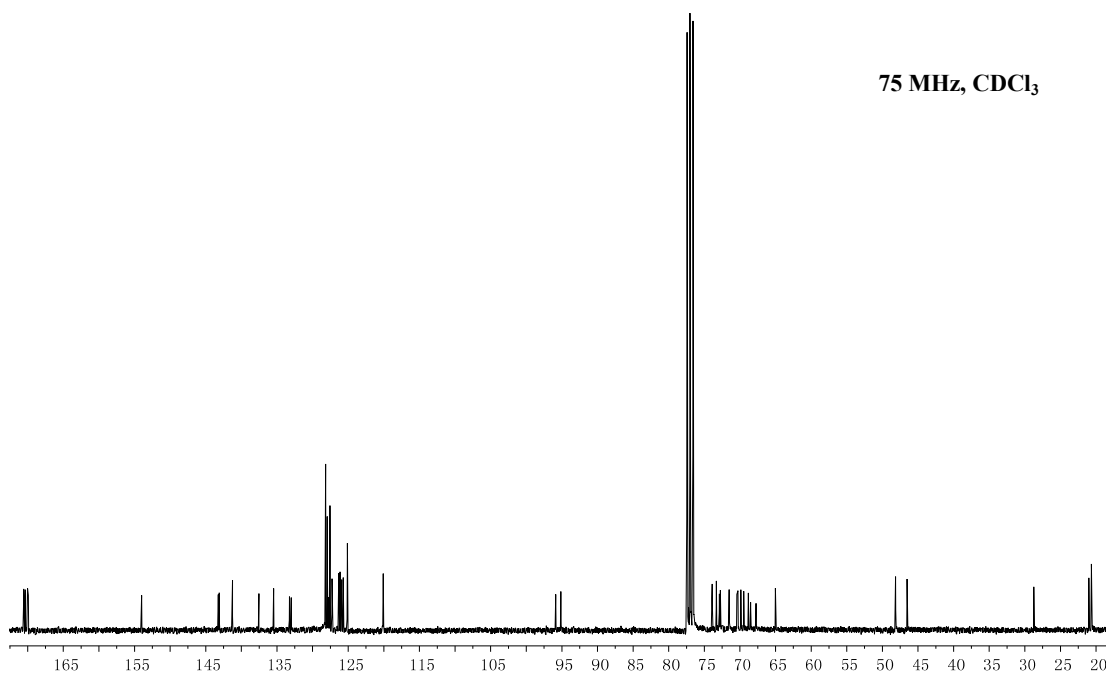


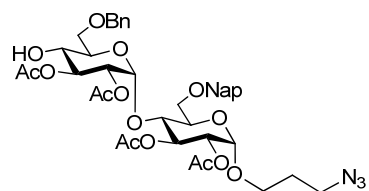


300 MHz, CDCl₃



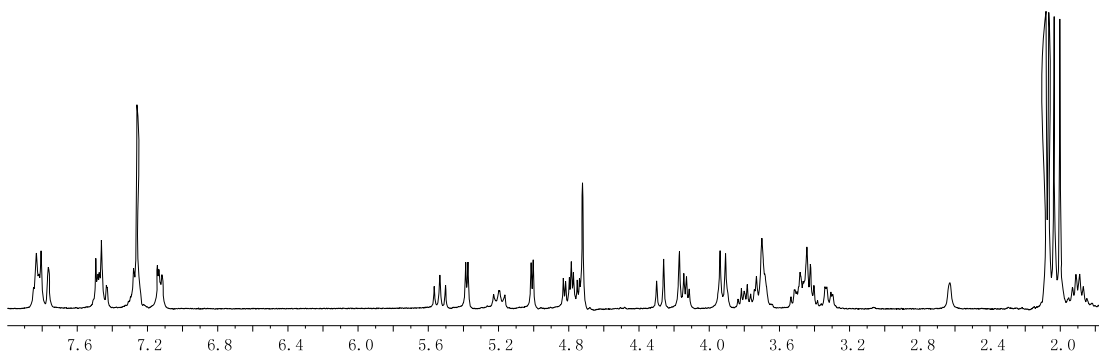
75 MHz, CDCl₃



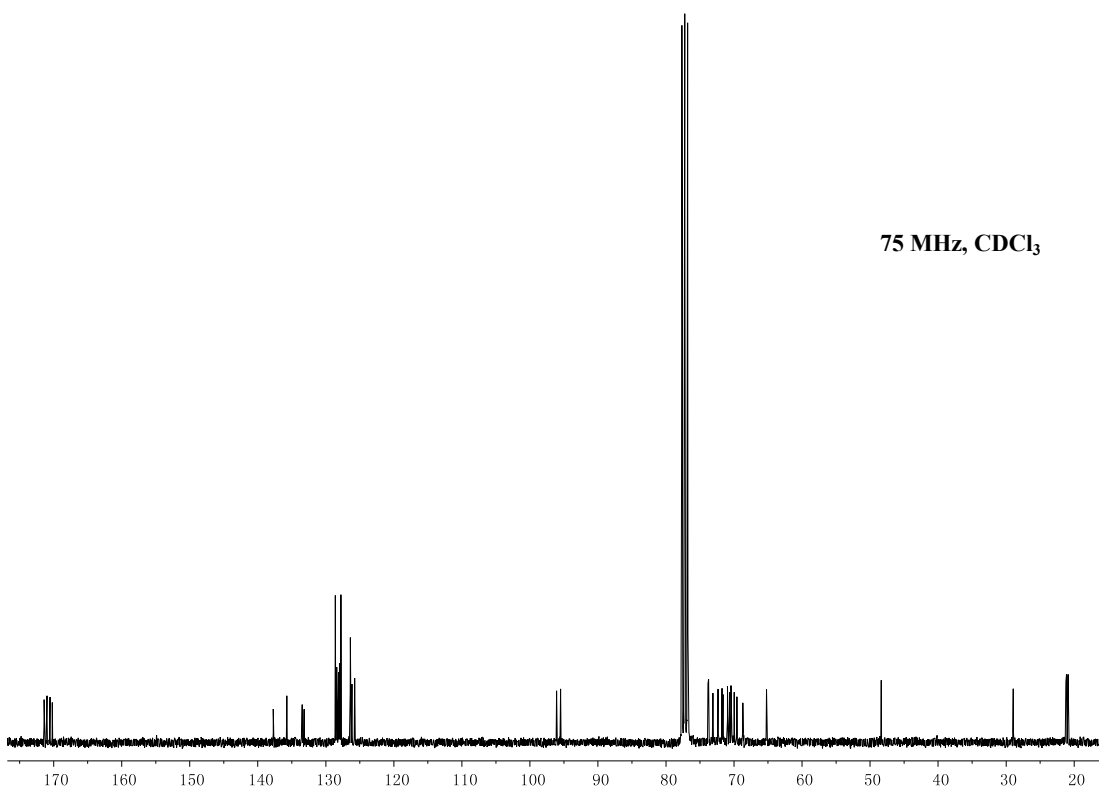


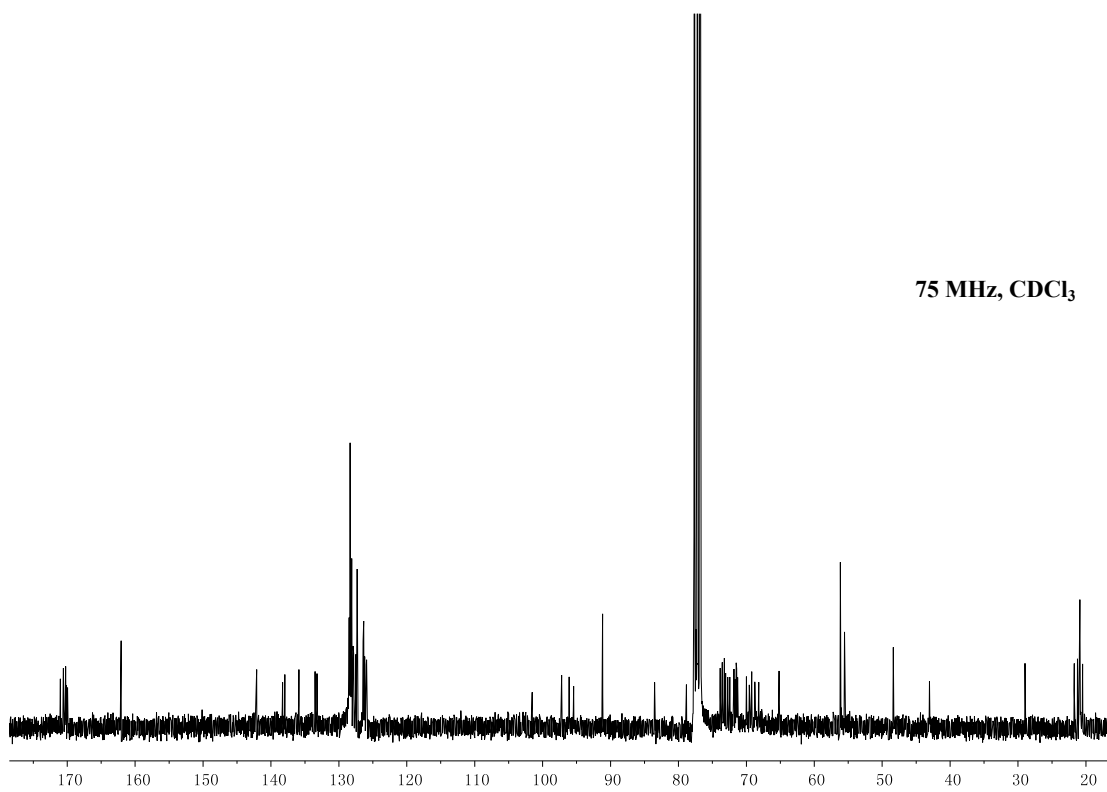
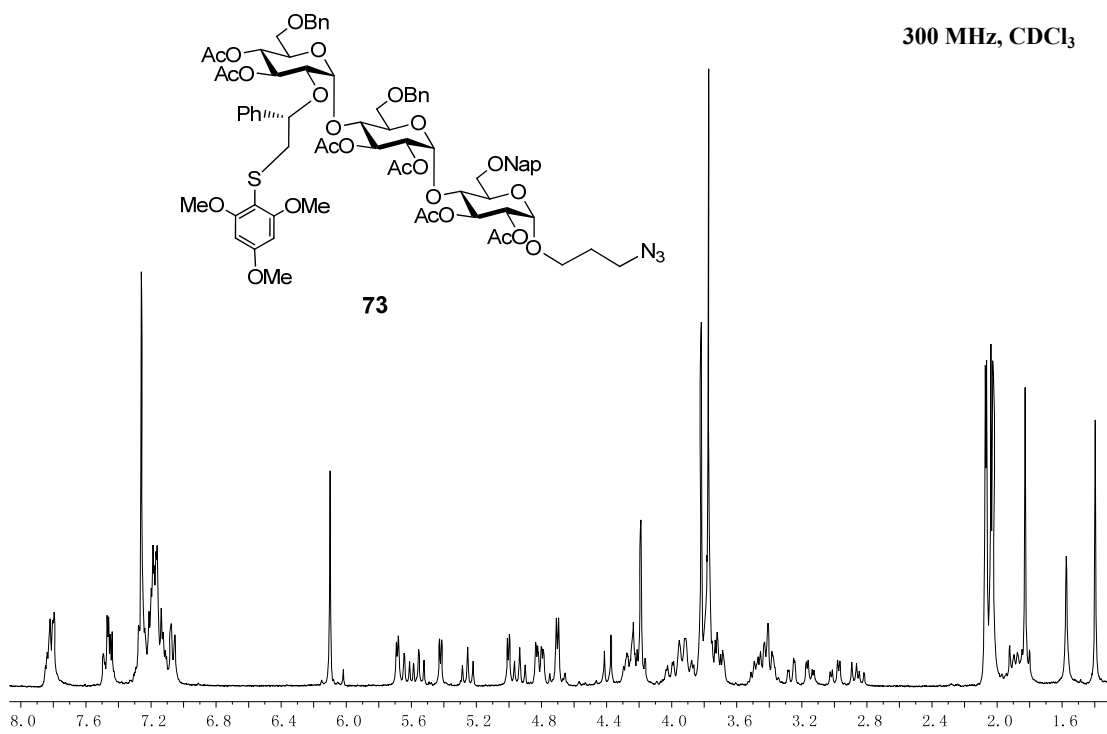
72

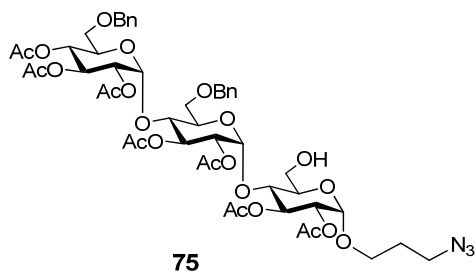
300 MHz, CDCl₃



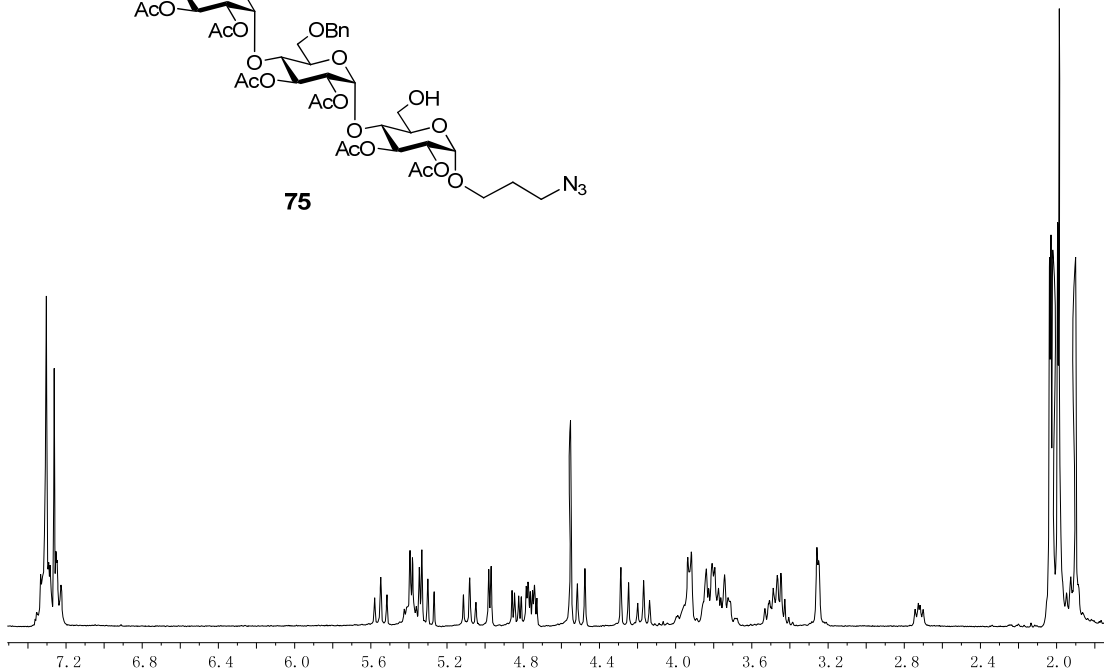
75 MHz, CDCl₃



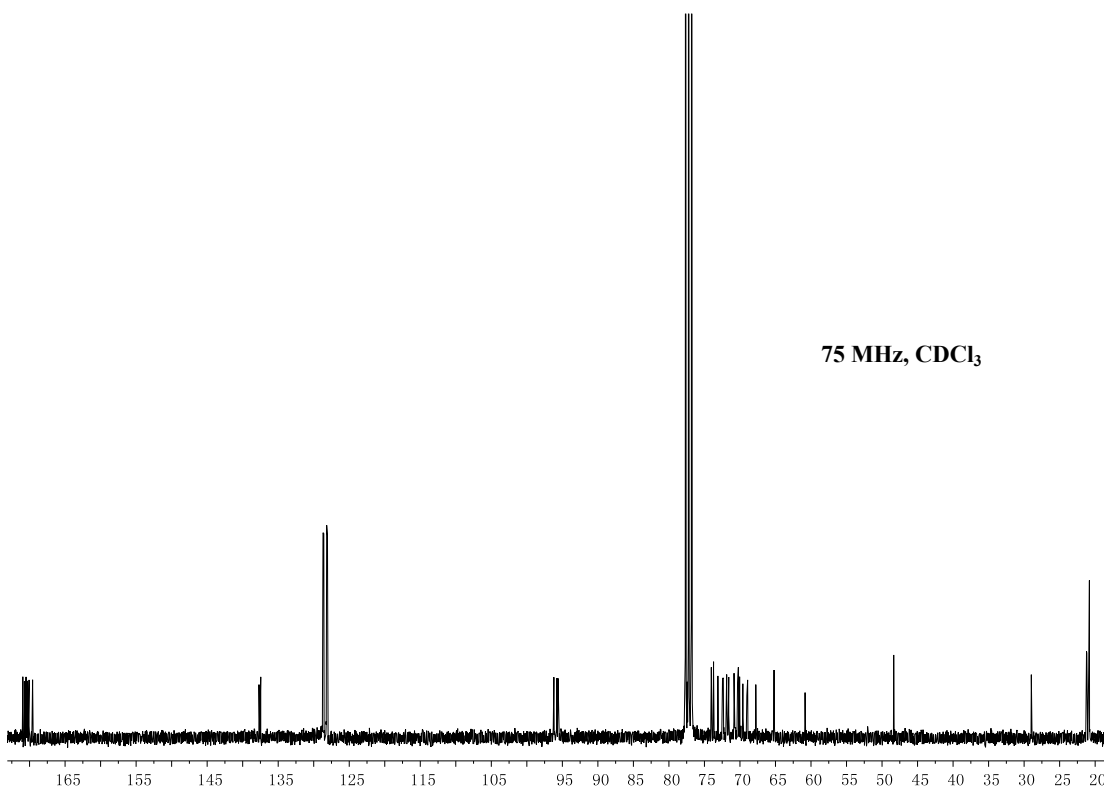


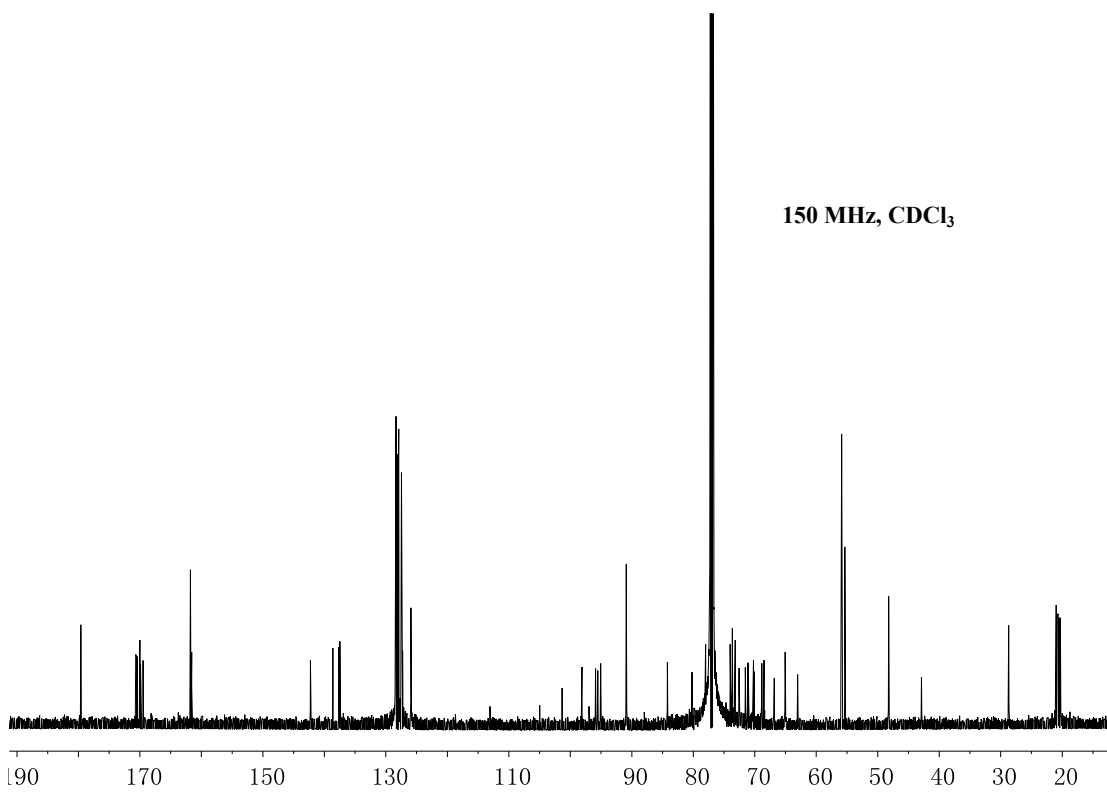
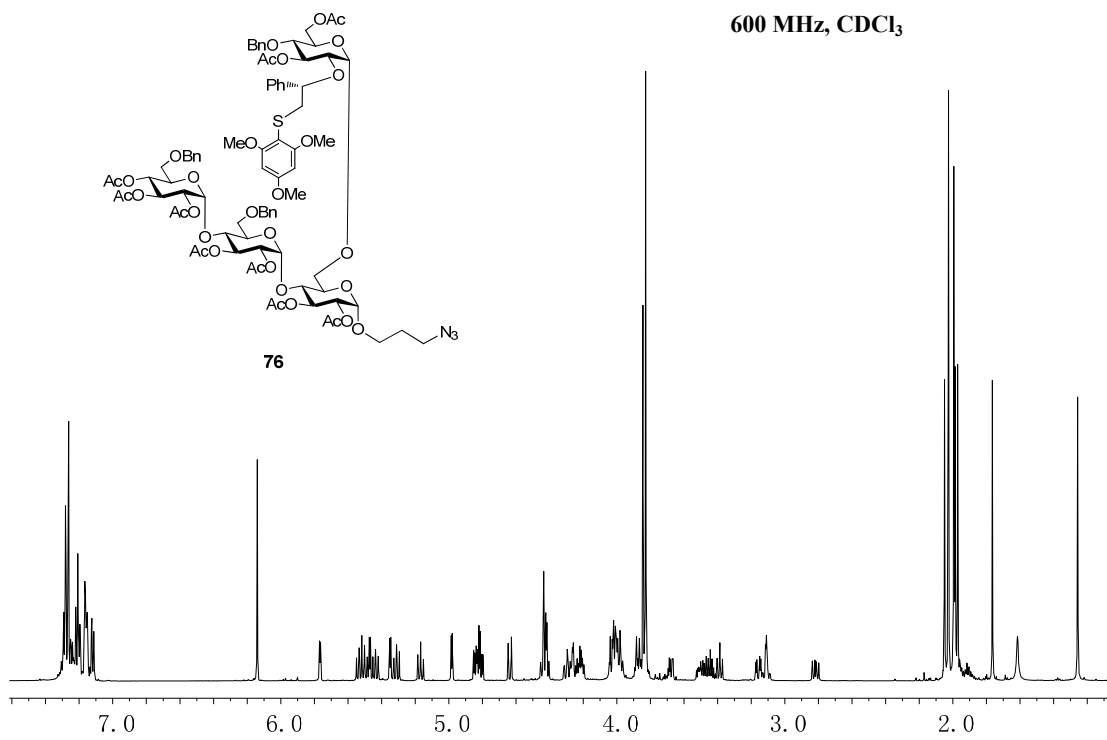


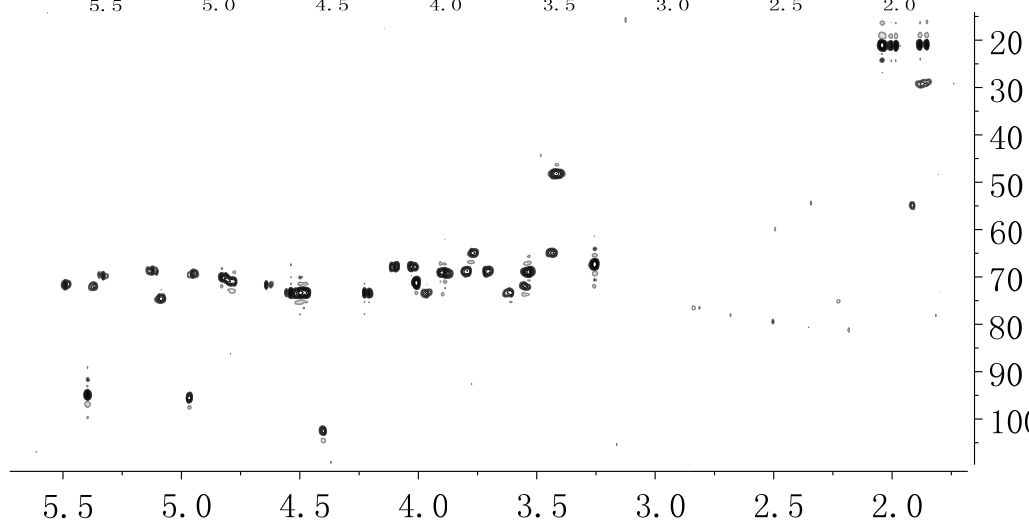
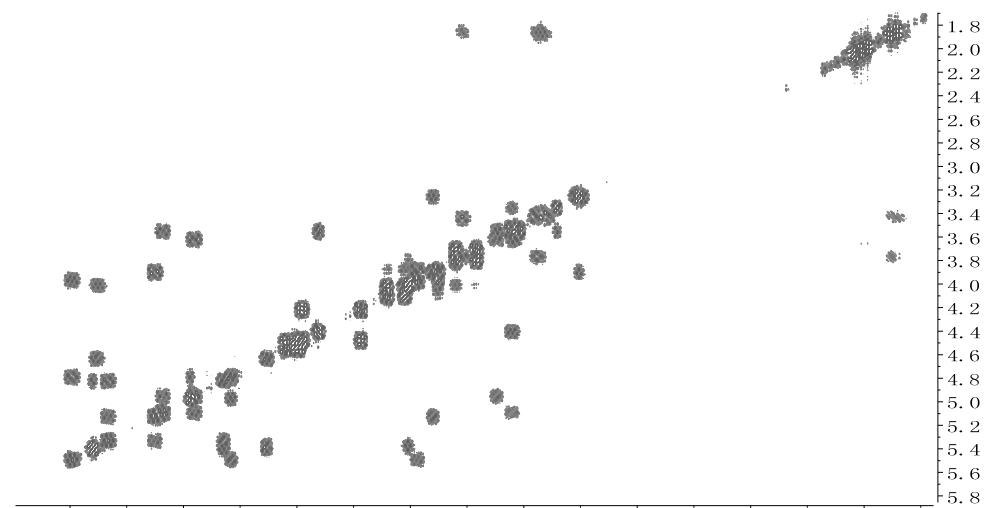
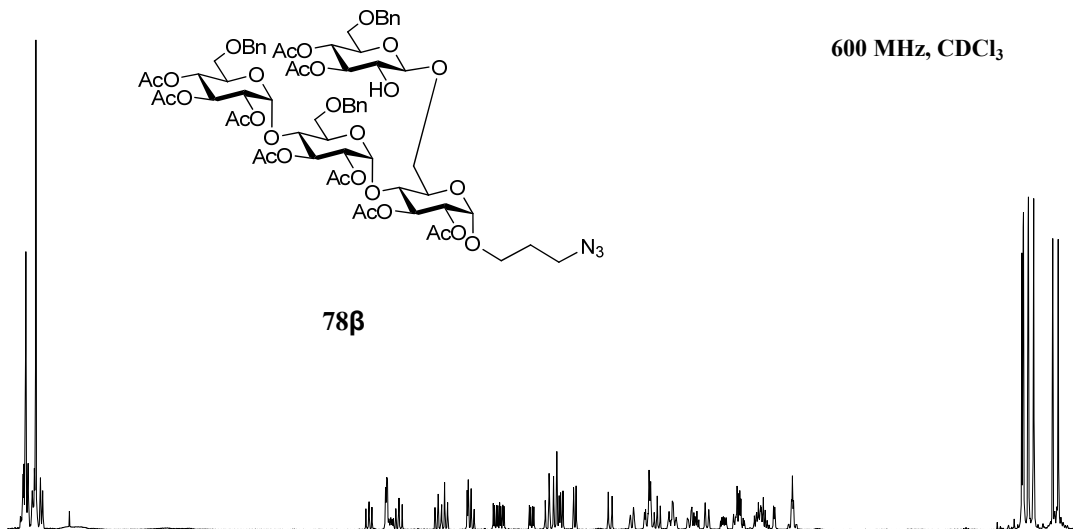
300 MHz, CDCl₃

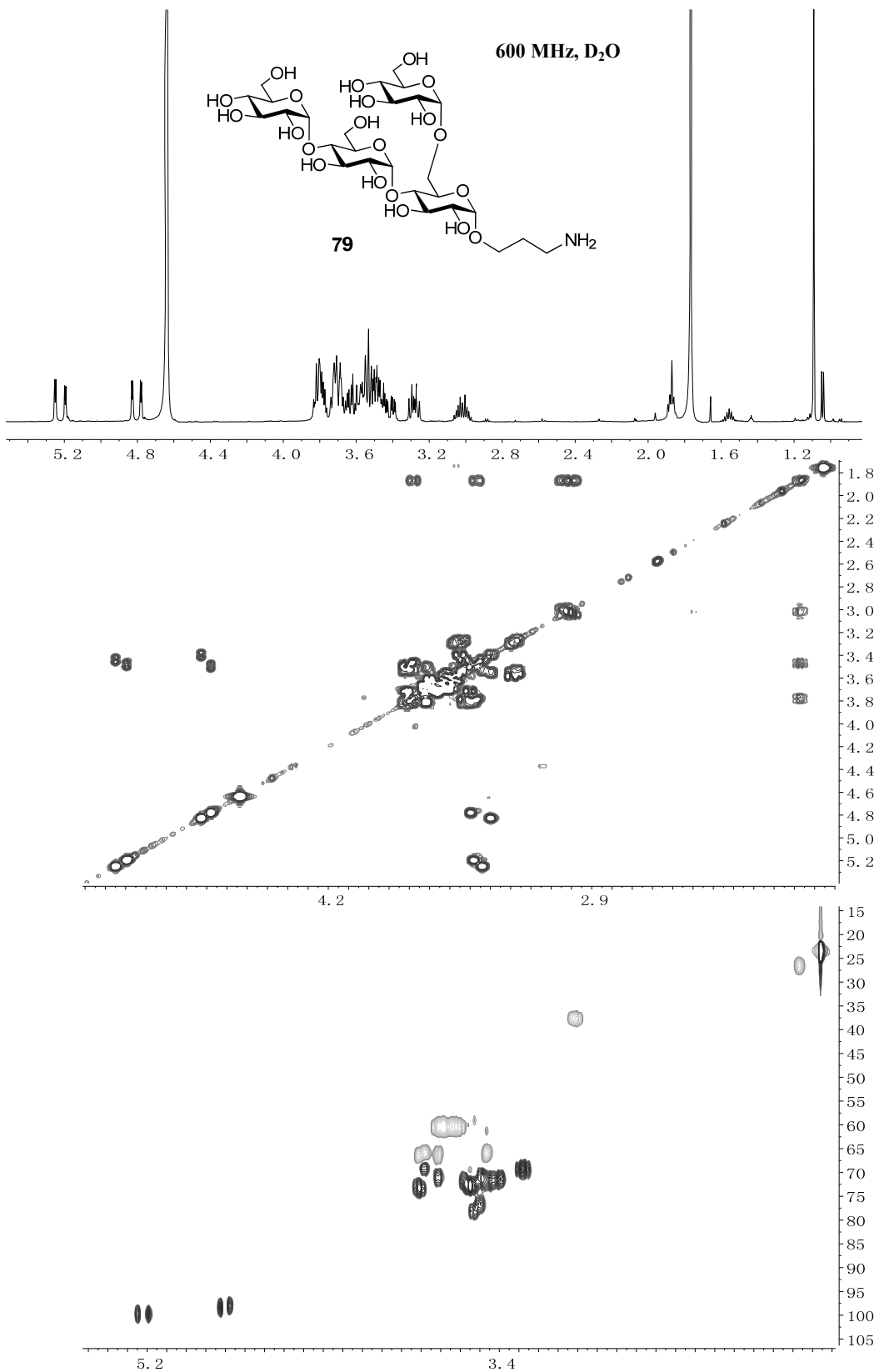


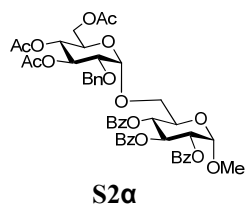
75 MHz, CDCl₃



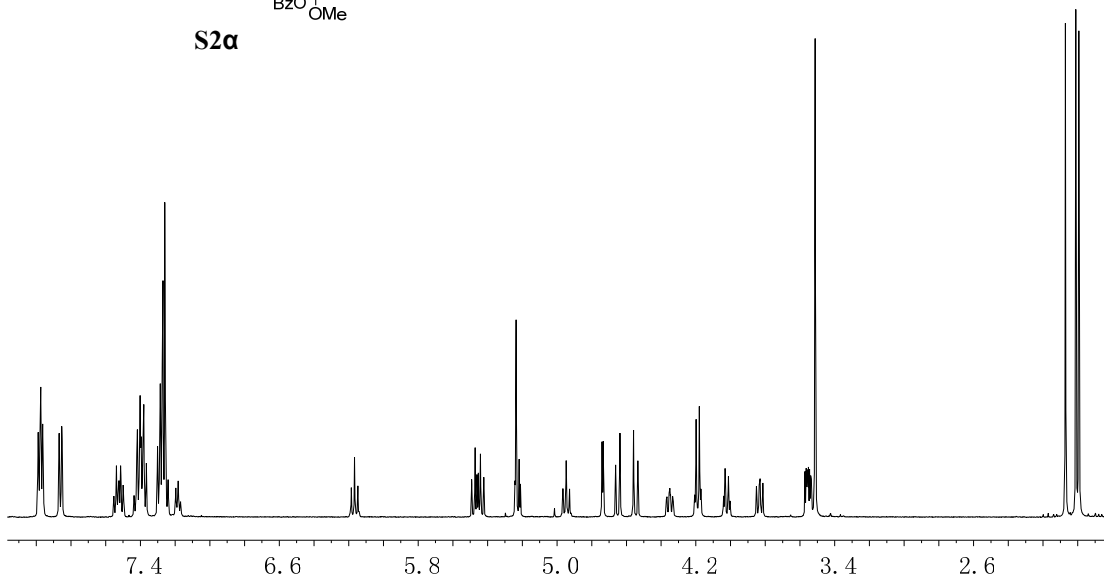




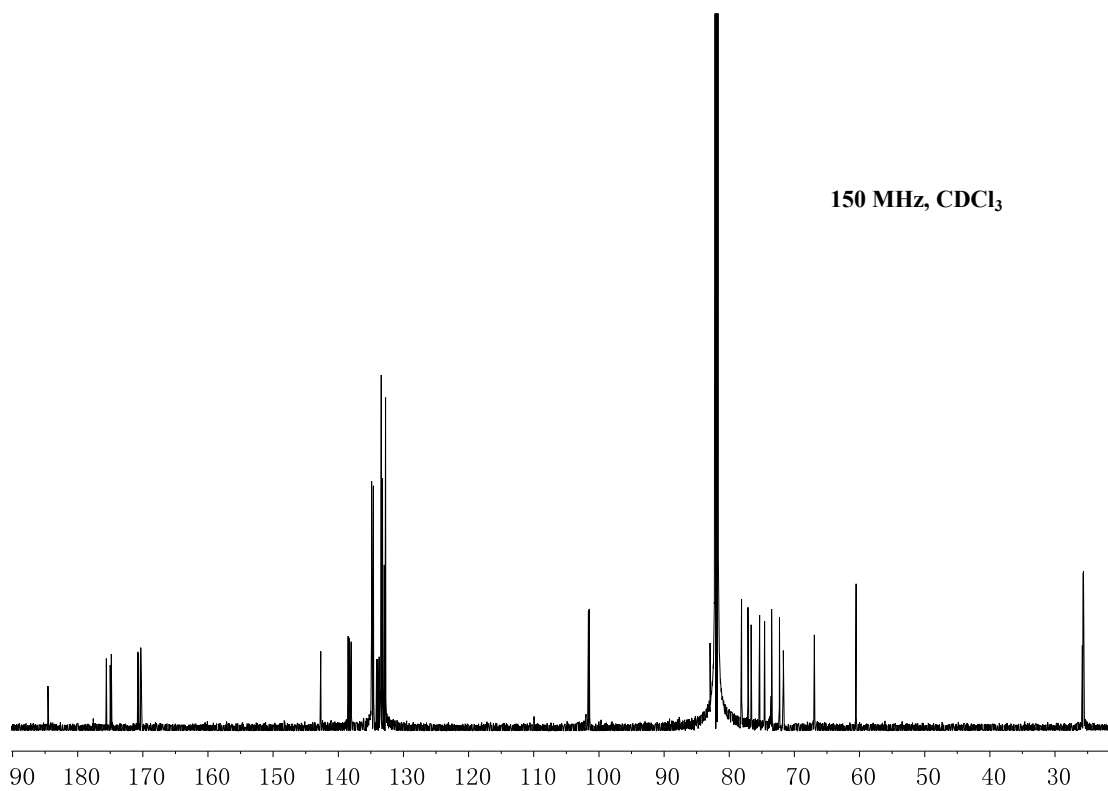


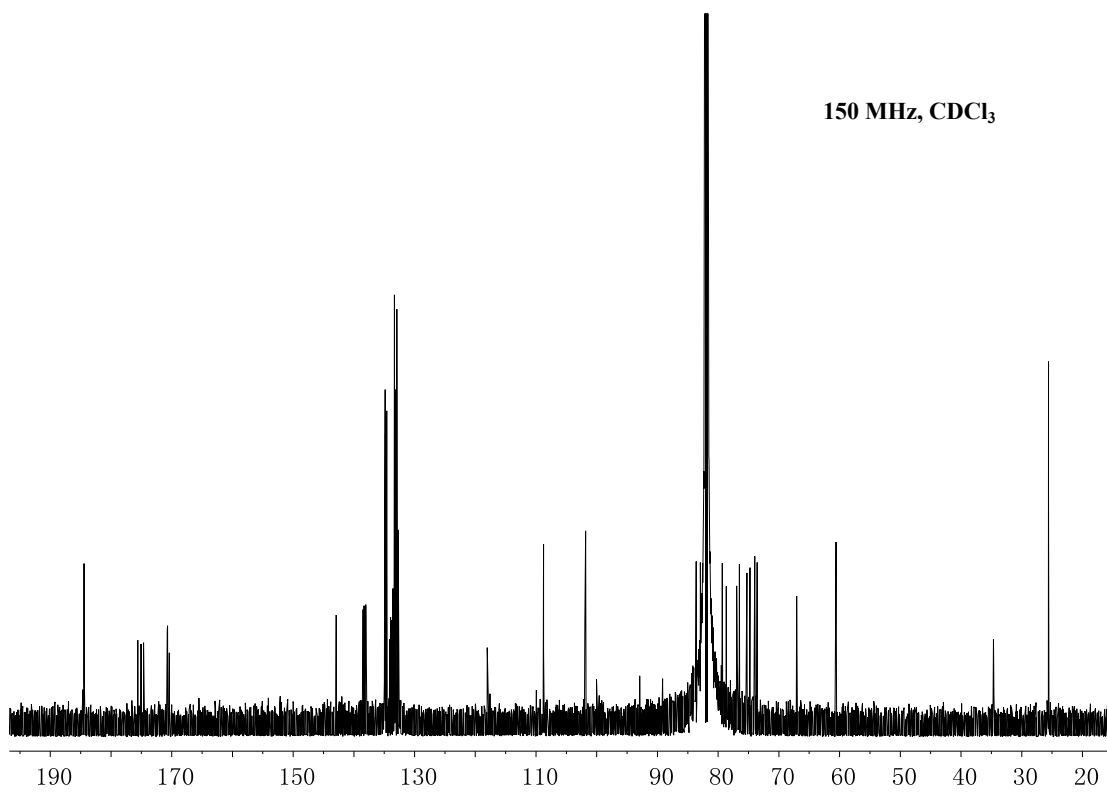
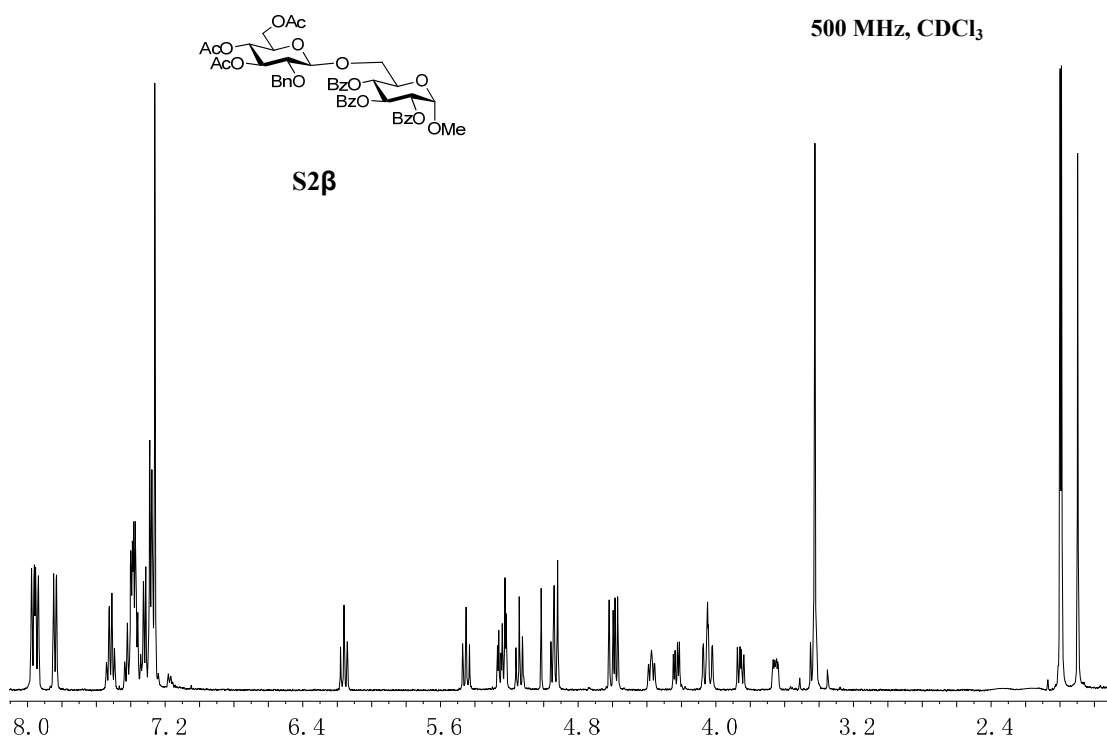


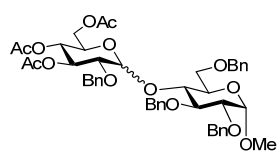
500 MHz, CDCl₃



150 MHz, CDCl₃

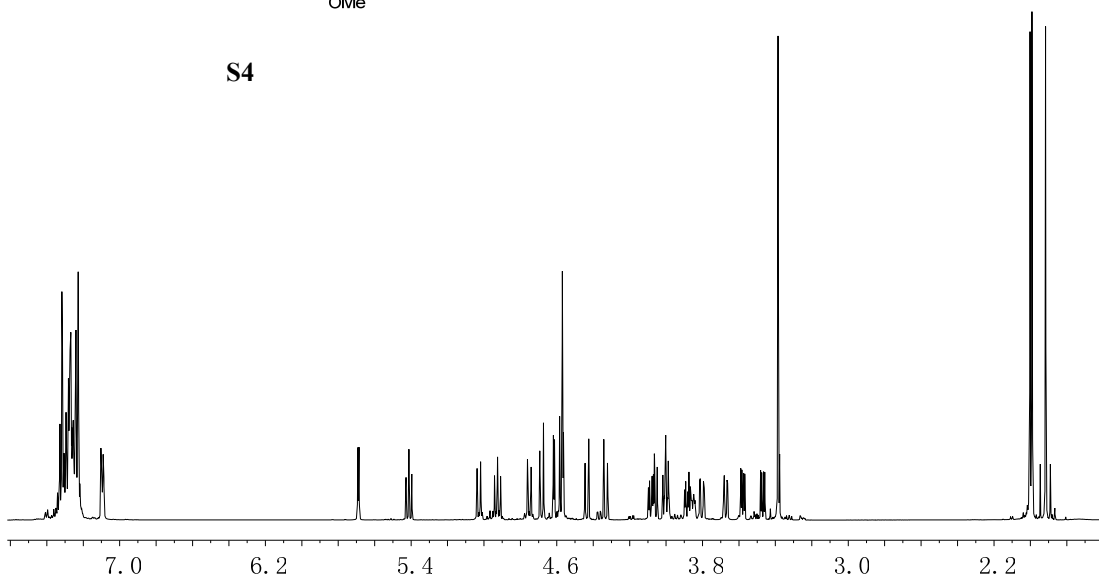






600 MHz, CDCl₃

S4



150 MHz, CDCl₃

