

Supporting Information

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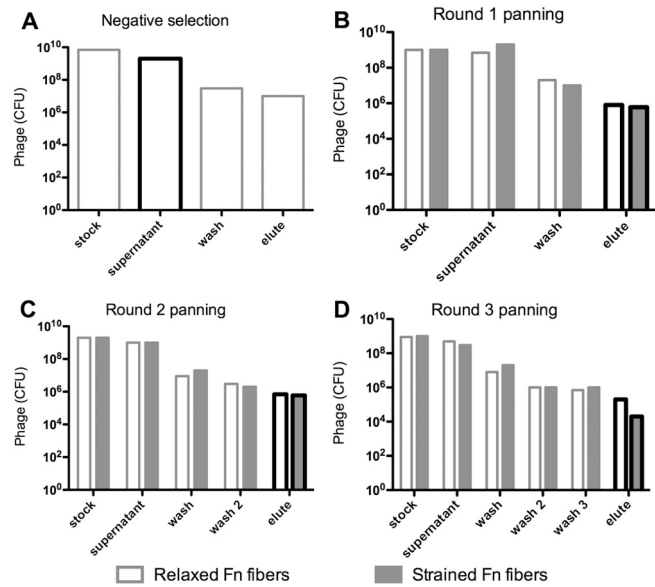


Fig. S1. Phage titers of successive panning rounds show emergence of a strongly bound phage population. (A) Negative selection round to deplete library of nonspecific binders to surface. (B–D) Three successive rounds of panning, with stringency of binding controlled by number of wash incubations.

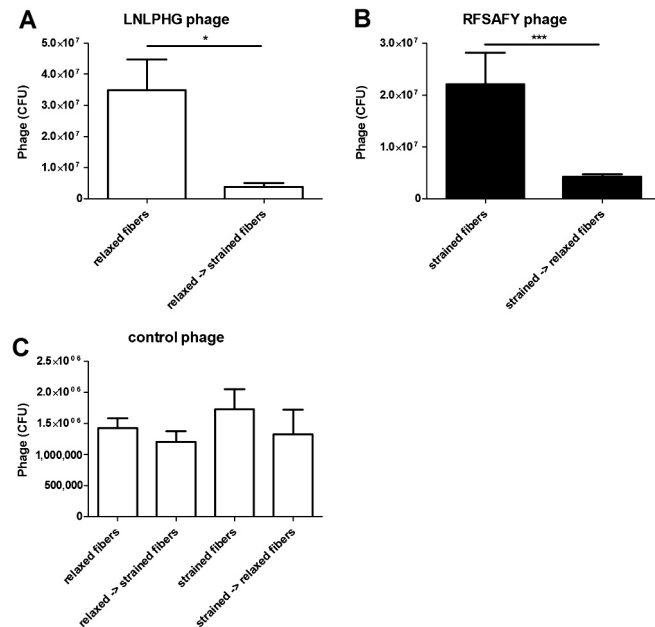


Fig. S2. Reversible binding of phage probes on fibronectin (Fn) fibers. (A) LNLPHG phage (1×10^{12} cfu) was incubated on relaxed Fn fibers ($\lambda = 0.93$) for 30 min, fibers were then strained to $\lambda = 2.6$ and phages were recovered and quantitated by titers. (B) RFSAFY phage (1×10^{12} cfu) was incubated on strained Fn fibers ($\lambda = 2.6$) for 30 min, fibers were then relaxed to $\lambda = 1$ and phages were recovered and quantitated by titers. (C) Control phage (1×10^{12} cfu) incubated on Fn fibers, and recovered after strain application (*, $p < 0.05$, ***, $p < 0.001$).

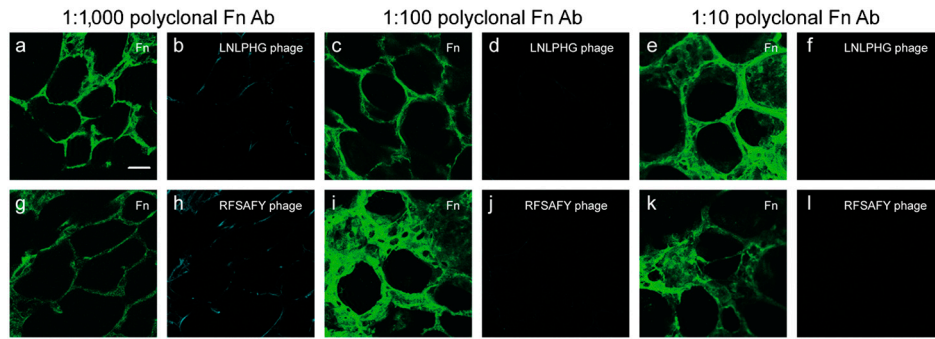


Fig. 53. Competitive binding of polyclonal anti-fibronectin (Fn) antibody displaces binding of phage probes on tissue lung slices. A polyclonal anti-Fn antibody (1 mg/mL) was titrated in along with the LNLPHG phage and RFSAFY phage at 1:1,000 (A, B, G, H), 1:100 (C, D, I, J), and 1:10 (E, F, K, L) dilutions. Images acquired with 63× oil immersion objective. (Scale bar: 20 μm.)

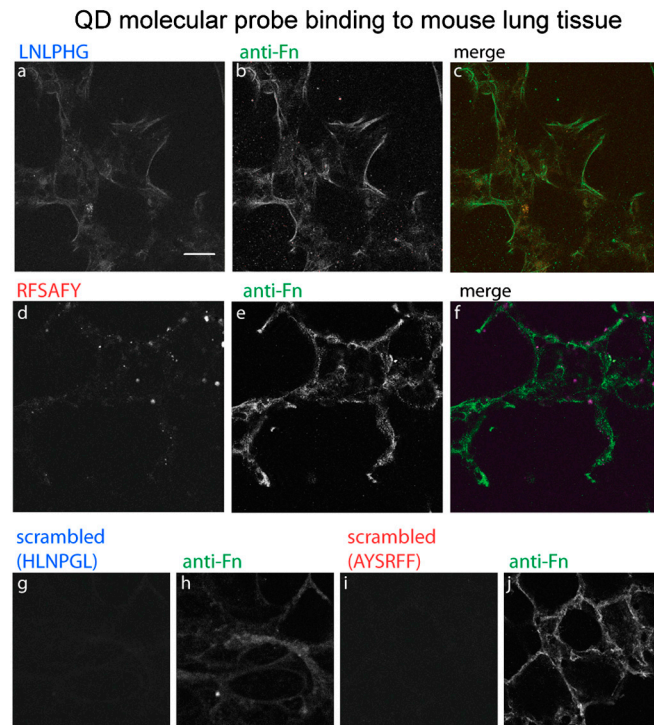


Fig. 54. Staining of labeled peptides on prepared living lung slices. Peptides corresponding displayed peptides on phage-based probes were generated by solid phase synthesis and contain a C-terminal cysteine for chemical ligation (peptides: LNLPHGGGC, RFSAFYGGC, HLNPLGGC, AYSRFFGGC). Peptides were conjugated to Qdot (QD) and purified per manufacturer's instructions (Molecular Probes). (A–C) Staining of LNLPHG probe and costaining with polyclonal anti-fibronectin (Fn) antibody. (D–F) Staining of RFSAFY probe and costaining with polyclonal anti-Fn antibody. (G and H) Staining of scrambled LNLPHG probe. (I and J) Staining of scrambled RFSAFY probe. All images acquired with 63× oil immersion objective. (Scale bar: 20 μm.)

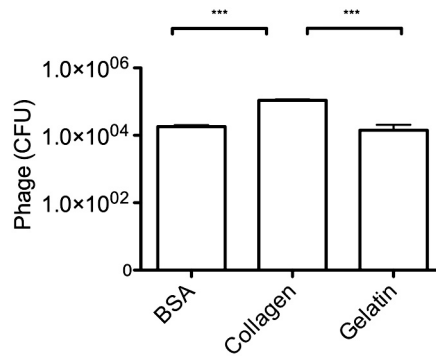


Fig. S7. Phage clones after round three of panning do not bind to control surfaces. Phage particles 1×10^{11} were incubated on polydimethylsiloxane surfaces coated with BSA (10 mg/mL), collagen I (10 mg/mL), or gelatin (10 mg/mL) for 1 h. $N > 3$ for all samples, statistics were performed using a one-way ANOVA with Bonferroni post test correction (***, $p < 0.001$).

Table S1. List of clones identified after panning with fuse5 6-mer library and frequency each clone was identified

Phage clone	Sequence	Identification freq.
<i>Relaxed</i>		
Fuse5-c15	SRWYRI	5
Fuse5-c20	LNLPHG	3
Fuse5-c12	ARERFY	2
Fuse5-c11	GSNSKY	1
<i>Strained</i>		
Fuse5-s05	RFSAFY	2
fuse5-39	EVIRQR	1
fuse5-03	GHVTRN	1
fuse5-11	GIYLLP	1
fuse5-15	GKPHGW	1
fuse5-25	IGGVSR	1
fuse5-21	KPSGLY	1
fuse5-24	KPTTVD	1
fuse5-35	LTRKSD	1
fuse5-07	LVGVLS	1
fuse5-33	PESSIG	1
fuse5-23	PPNDVH	1
fuse5-05	PRNLRQ	1
fuse5-22	QAPFR	1
fuse5-27	RTVKYW	1
fuse5-14	SSHWFT	1
fuse5-31	SSSRTQ	1
fuse5-17	SSTYGH	1
fuse5-02	TGRRID	1
fuse5-20	TLAKAH	1
fuse5-32	TRSGGK	1
fuse5-13	TVLVGQ	1
fuse5-10	VRGVTL	1
fuse5-01	VTRLLH	1
fuse5-29	WRAKLV	1
fuse5-04	WRVLYA	1
fuse5-36	WSYILS	1
fuse5-41	YFFVDT	1
fuse5-30	YIQDYN	1
fuse5-38	YTHLAK	1

Clones were sequenced after three rounds of panning. Bolded clones were produced and assayed for binding to Fn fibers.