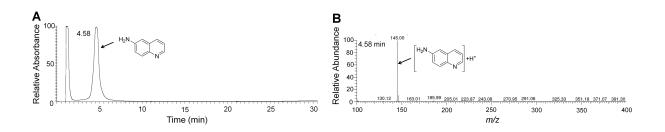
## Supporting information for: Hypoxia-selective, enzymatic conversion of 6-nitroquinoline into a fluorescent helicene: pyrido[3,2-*f*]quinolino[6,5-*c*]cinnoline 3-oxide

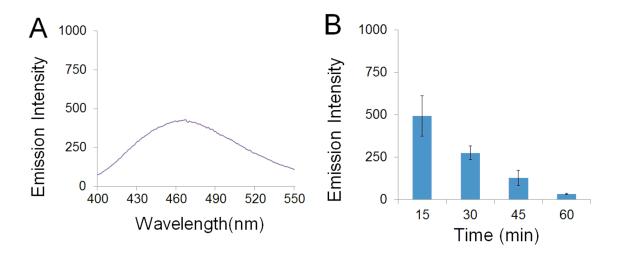
Anuruddha Rajapakse<sup>§</sup> and Kent S. Gates<sup>§,‡,\*</sup> <sup>§</sup>Department of Chemistry, University of Missouri, Columbia, MO 65211 <sup>‡</sup>Department of Biochemistry, University of Missouri, Columbia, MO 65211 E-mail: gatesk@missouri.edu, Ph: (573) 882-6763

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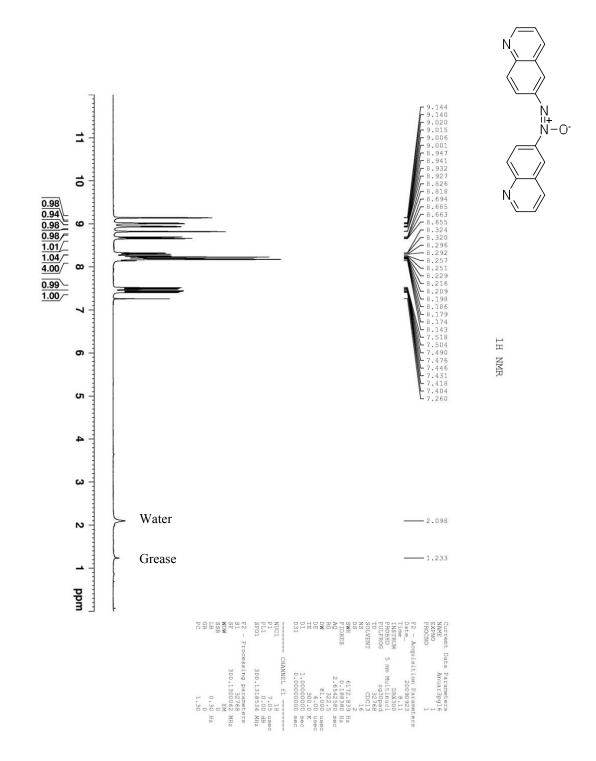
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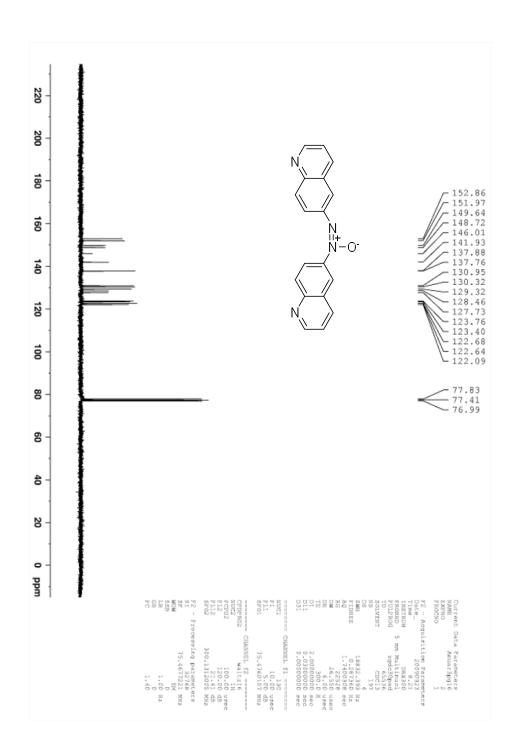
**Figure S1**. LC/MS analysis of 6-aminoquinoline (**2**). (A) UV chromatogram of **2**. (B) LC/MS analysis of the product eluting 4.6 min in the chromatogram.



**Figure S2**. Fluorescence spectra of NADPH and evidence that residual NADPH is consumed during reaction workup. (A) Fluorescence spectrum of NADPH (50  $\mu$ M,  $\lambda_{ex}$  307 nm, in sodium phosphate buffer, 12 mM, pH 7.4). Note that the shape of the fluorescence emission peak generated by NADPH is different from that for **4** shown in the manuscript. (B) Control reactions showing that NADPH fluorescence diminishes over the course of 1 h under our typical workup conditions. Reactions containing **1** (0.8 mM), NADPH:cytochrome P450 reductase (1.1 U/mL), and NADPH (3 mM) in sodium phosphate buffer, 10 mM, pH 7.4 ( $\lambda_{ex}$  307 nm,  $\lambda_{em}$  460 nm) were incubated for various times before fluorescence analysis. This control experiment provides additional evidence that fluorescence detected in the metabolism of **1** is *not* due to residual NADPH.



**Figure S3.** <sup>1</sup>H NMR of **3** (CDCl<sub>3</sub>, 300 MHz).



**Figure S4.** <sup>13</sup>C NMR of **3** (CDCl<sub>3</sub>, 125 MHz).

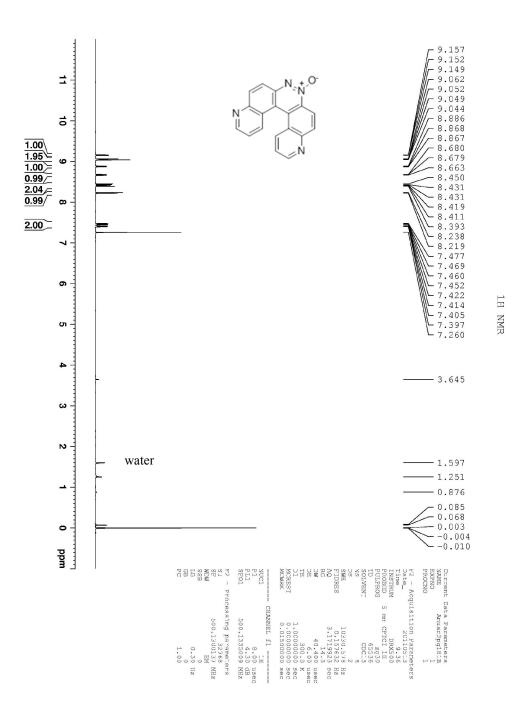
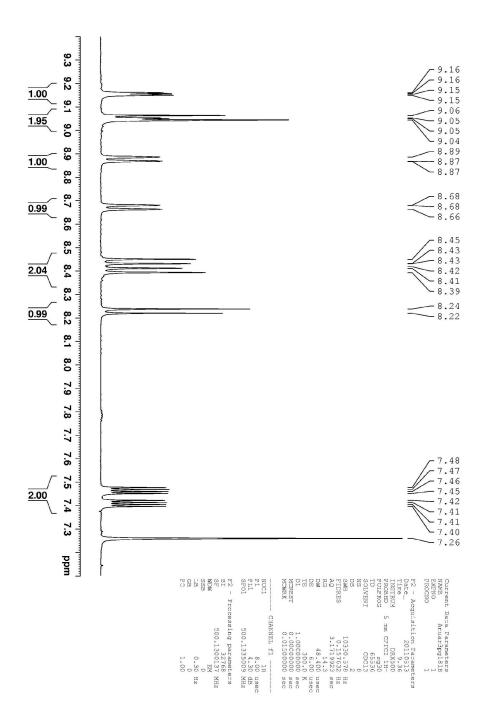
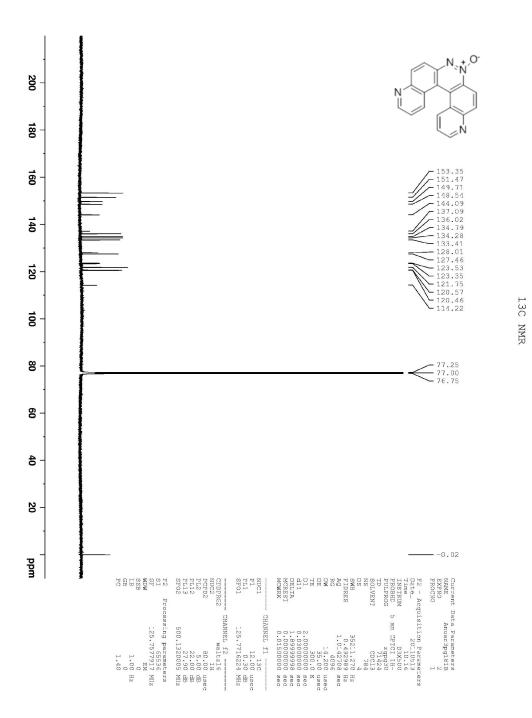


Figure S5. <sup>1</sup>H NMR of 4 (CDCl<sub>3</sub>, 500 MHz).



1H NMR

Figure S6. <sup>1</sup>H NMR of 4, aromatic region (CDCl<sub>3</sub>, 500 MHz).



**Figure S7**. <sup>13</sup>C NMR of **4** (CDCl<sub>3</sub>, 125 MHz).

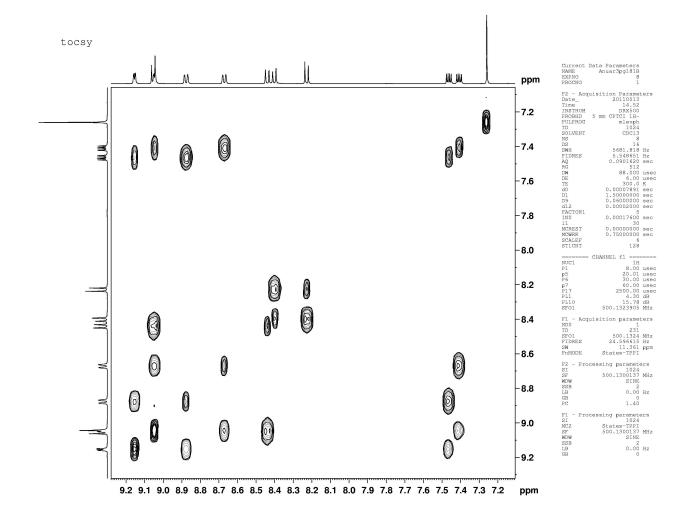
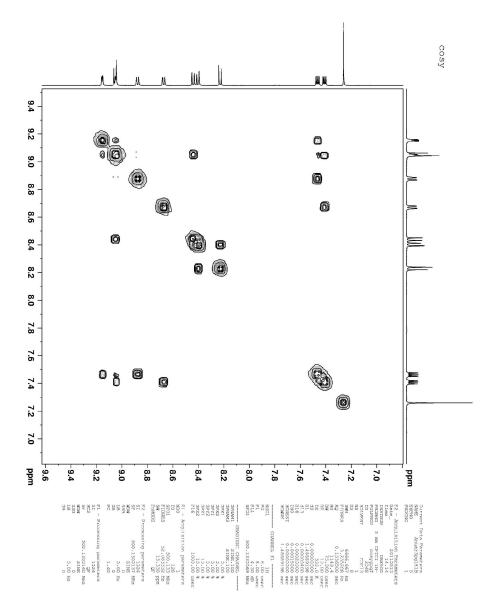


Figure S8. <sup>1</sup>H–<sup>1</sup>H TOCSY of 4 (CDCl<sub>3</sub>, 500 MHz).



**Figure S9**. <sup>1</sup>H–<sup>1</sup>H COSY of **4** (CDCl<sub>3</sub>, 500 MHz).

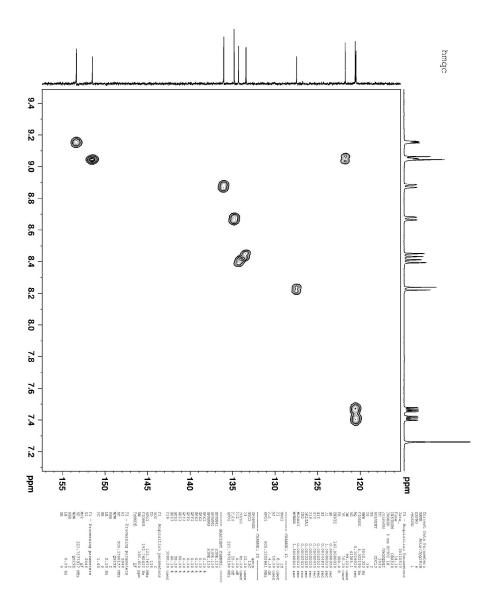


Figure S10.  $^{1}H-^{13}C$  HMQC of 4 (CDCl<sub>3</sub>).

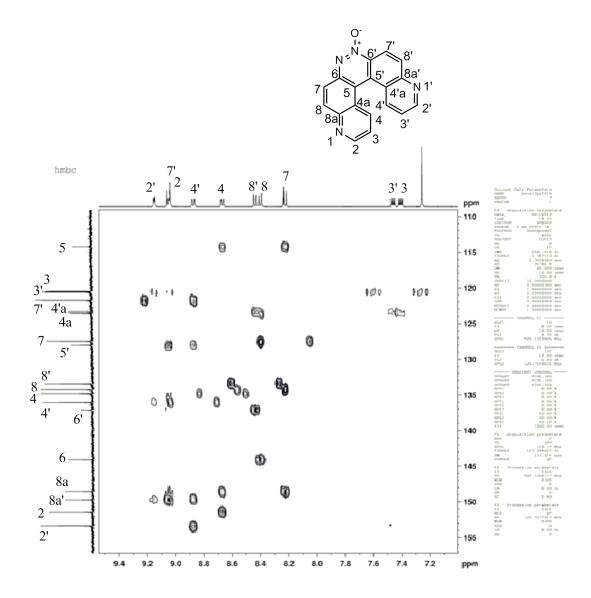


Figure S11.  $^{1}H^{-13}C$  HMBC of 4 (CDCl<sub>3</sub>).

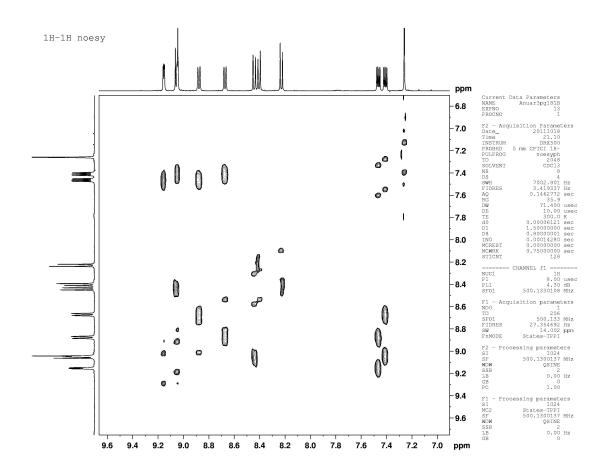
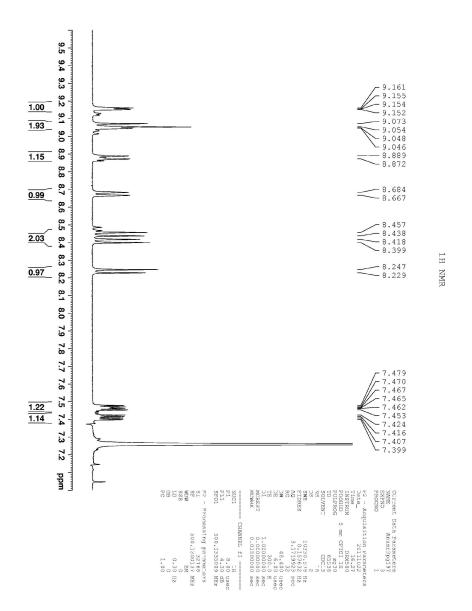
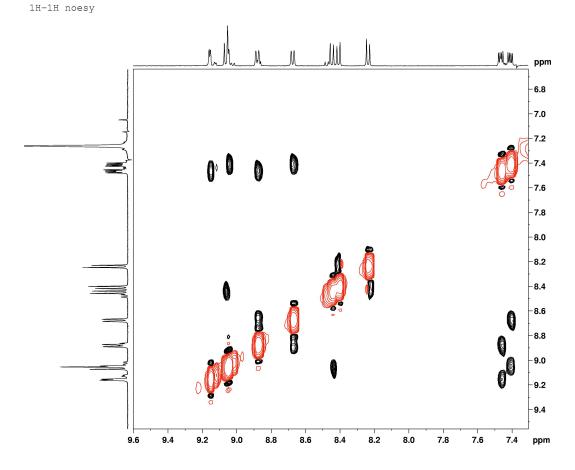


Figure S12. <sup>1</sup>H-<sup>1</sup>H NOESY of 4 (CDCl<sub>3</sub>, 500 MHz).



**Figure S13**. <sup>1</sup>H NMR of **4** (CDCl<sub>3</sub>, 500 MHz), obtained from NADPH and NADPH:cytochrome P450 reductase-mediated synthesis.



**Figure S14.** <sup>1</sup>H-<sup>1</sup>H NOESY of **4** (CDCl<sub>3</sub>, 500 MHz), obtained from NADPH and NADPH:cytochrome P450 reductase-mediated synthesis.