Supplementary information, Table S2 The list of culture methods for the cell samples used in the study

Sample Name	Culture Medium	Culture Condition and Passage Method
Human embryonic stem cells		
BG01	StemPro® hESC SFM[1]	Matrigel, enzymatic passage[8]
BG02	StemPro® hESC SFM[1]	Matrigel, enzymatic passage[8]
BG03	StemPro® hESC SFM[1]	Matrigel, enzymatic passage[8]
WA01	hESC medium[2]	Radiation-inactivated MEF feeder, enzymatic passage[8]
WA07	EctoBias medium[5,6]	Matrigel[6]
WA09 P31	hESC medium[7]	Mitomycin C-treated SNL feeder, enzymatic passage[9]
WA01 P35	hESC medium[7]	Mitomycin C-treated SNL feeder, enzymatic passage[9]
WA09P90_MEF/MAN	hESC medium[2]	Radiation-inactivated MEF feeder, manual passage
WA09P90_MG/MAN	StemPro® hESC SFM[1]	Matrigel, manual passage
WA09P90_MEF/ACC	hESC medium[2]	Radiation-inactivated MEF feeder, enzymatic passage[8]
WA09P42_MG/ACC	StemPro® hESC SFM[1]	Matrigel, enzymatic passage[8]
WA09P90_MG/ACC	StemPro® hESC SFM[1]	Matrigel, enzymatic passage [8]
Induced pluripotent stem cells from Human Dermal Fibroblasts (HDF)		
201B2 P24	hESC medium[7]	Mitomycin C-treated SNL feeder, enzymatic passage[9]
201B6 P24	hESC medium[7]	Mitomycin C-treated SNL feeder, enzymatic passage[9]
201B7 P24	hESC medium[7]	Mitomycin C-treated SNL feeder, enzymatic passage[9]
201B7 P78	hESC medium[7]	Mitomycin C-treated SNL feeder, enzymatic passage[9]
253G1 P24	hESC medium[7]	Mitomycin C-treated SNL feeder, enzymatic passage[9]
253G4 P24	hESC medium[7]	Mitomycin C-treated SNL feeder, enzymatic passage[9]
iPS2.HDF-ALS275	hESC medium[2]	Radiation-inactivated MEF feeder, manual passage
iPS8.HDF-ALS284	hESC medium[2]	Radiation-inactivated MEF feeder, manual passage
iPS5.HDF-ALS285	hESC medium[2]	Radiation-inactivated MEF feeder, manual passage
iPS11.HDF	StemPro® hESC SFM[1]	Matrigel, enzymatic passage[8]
iPS7.HDF	StemPro® hESC SFM[1]	Matrigel, enzymatic passage[8]
iPS1.HDF	StemPro® hESC SFM[1]	Matrigel, enzymatic passage[8]
Induced pluripotent stem cells from Human Epidermal Melanocytes (HEM)		
iPS1.HEMd	hESC medium[2]	Radiation-inactivated MEF feeder, manual passage
iPS2.HEMd	hESC medium[2]	Radiation-inactivated MEF feeder, manual passage
Cells used for reprogramming		
aHDF	DMEM with 10% FBS[3]	enzymatic passage[10]
HDF51	DMEM with 10% FBS	enzymatic passage[10]
HDF-ALS275	DMEM with 10% FBS	enzymatic passage[10]
HDF-ALS284	DMEM with 10% FBS	enzymatic passage[10]
HEMd	MelM[4]	enzymatic passage[10]
Other cell types		
SKMEL2	RPMI1640 with 10% FBS[4]	enzymatic passage[10]
MALME3M	IMDM with 10% FBS[4]	enzymatic passage[10]
SKMEL5	RPMI1640 with 10% FBS[4]	enzymatic passage[10]
UACC257	RPMI1640 with 10% FBS[4]	enzymatic passage[10]
WA07P34MNPD29	MN differentiation medium[6]	Laminin[6]
HREpic	EpiCM[4]	Poly-D-Lysine, enzymatic passage[10]
HSkMC	SkMCM[4]	Poly-D-Lysine, enzymatic passage[10]
MEF	DMEM with 10% FBS	enzymatic passage[10]

1. StemPro® hESC serum-free medium (SFM): DMEM/F12 plus GlutaMax™-I media (Invitrogen, Carlsbad, CA) containing 12ng/ml bFGF (Stemgent, Cambridge, MA), 1.8% BSA (Invitrogen, Carlsbad, CA), 100μM β-mercaptoethanol (Invitrogen, Carlsbad, CA) and 1x StemPro® hESC supplement (Invitrogen, Carlsbad, CA)
2. hESC medium: MDEM/F12 media with L-glutamine (Invitrogen, Carlsbad, CA) containing 20% KnockOut™ Serum Replacement (Invitrogen, Carlsbad, CA), 100μM non-essential amino acids (Invitrogen, Carlsbad, CA), 100μM β-mercaptoethanol (Invitrogen, Carlsbad, CA), 12ng/ml bFGF (Stemgent, Cambridge, MA)
3. This medium contains 50 units/ml penicillin and 50 μg/ml streptomycin (Invitrogen, Carlsbad, CA).
4. This medium contains 100 units/ml penicillin and 100 μg/ml streptomycin (Invitrogen, Carlsbad, CA).
5. This medium contains 20 ng/ml bFGF (Millipore, Billerica, MA).
6. Ref. 18
7. MDEM/F12 media with L-glutamine (Invitrogen, Carlsbad, CA) containing 20% KnockOut™ Serum Replacement (Invitrogen, Carlsbad, CA), 100μM non-essential amino acids (Invitrogen, Carlsbad, CA), 100μM β-mercaptoethanol (Invitrogen, Carlsbad, CA), 4ng/ml bFGF (Wako, Osaka, Japan), 50 units/ml penicillin and 50 μg/ml streptomycin (Invitrogen, Carlsbad, CA)
8. StemPro® Accutase® Cell Dissociation Agent was used as the enzyme source.
9. CTK solution containing 0.25% trypsin, 100μg/ml collagenase IV, 1mM calcium chloride and 20% KnockOut™ Serum Replacement was used as the enzyme source.
10. Solution containing 0.05% Trypsin and 0.02% EDTA(Invitrogen, Carlsbad, CA) was used as the enzyme source.