	Inhibitor	Target kinase	working concentration	Supplier
1	SB20219	p38	10µM	CALBIOCHEM
2	PD98059	MEK1	50µM	CALBIOCHEM
3	LY294002	<b>PI3K</b>	50µM	CALBIOCHEM
4	Calphostin C	PKC	0.25µM	CALBIOCHEM
5	GF 109203X	РКС	0.1µM	CALBIOCHEM
6	Staurosporine	PKC	0.1µM	CALBIOCHEM
7	RO-3306	Cdk1	20µM	CALBIOCHEM
8	W-7	CaMKII	50µM	CALBIOCHEM
9	SB216763	<b>GSK3</b> β	10µM	Tocris bioscience
10	SB218078	Chk1	10µM	CALBIOCHEM
11	Chk2 inhibitor	Chk2	10µM	CALBIOCHEM
12	Rottlerin	РКС	5µM	CALBIOCHEM
13	Caffeine	ATM/ATR	5mM	Sigma

Supplementary Table S1. Protein kinase inhibitors used in this study.



# Supplementary Figure S1. Effects of protein kinase inhibitors on Mig-6 phosphorylation *in vivo*.

HEK293 cells were transfected with FLAG-Mig-6 and treated with the indicated kinase inhibitors for 3 h before harvesting. FLAG-Mig-6 was immunoprecipitated with anti-FLAG antibody from the cell lysates and p-Mig-6 was detected by the Phos-tag biotin system. Details of the kinase inhibitors are indicated in Supplementary Table 1.



# Supplementary Figure S2. EGF-promoted phosphorylation of Mig-6 is suppressed by Chk1 depletion.

**A.** MDA-MB-231 cells were transfected with another siRNA oligo, *Chk1*(04). Quiescence was induced by 16 h serum starvation, and then the cells were stimulated with 20 ng/ml EGF for 15 min. Cell lysates were separated by 6% Phos-tag SDS-PAGE or normal SDS-PAGE and analyzed by immunoblotting with the indicated antibodies. **B.** MDA-MB-231 cells were made quiescent by 16 h serum starvation, and then stimulated with 20 ng/ml EGF for 15 min. Cell lysates were incubated in 20  $\mu$ l 0.05 mM of Tris-HCl (PH 8.0), with or without 1 unit Calf Intestine Alkaline Phosphatase (CIAP) at 37°C for 30 min. The reaction was stopped by the addition of SDS sample buffer and then the proteins were separated by Phos-tag SDS-PAGE or normal SDS-PAGE followed by immunoblotting. **C.** MDA-MB-231 cells were made quiescent by 16 h serum starvation, pretreated with or without 10  $\mu$ M MG132 for 5 h, then stimulated with 20 ng/ml EGF for 15 min. Mig-6 was immunoprecipitated with anti-Mig-6 antibody and then immunoblotted with anti-ubiquitin antibody to detect ubiquitylation of Mig-6. (**\***: nonspecific

b a n d . )







### Supplementary Figure S3. Phosphorylation status of wild-type and mutant Mig-6.

**A**. *In vitro* phosphorylation assays of mutant recombinant Mig-6 proteins. Wild-type (WT) and the indicated mutant recombinant Mig-6 proteins (0.1 μg) were incubated in 20 μl of kinase buffer with 32P-labeled ATP and 0.1 μg of purified recombinant GST-Chk1 kinase at 30°C for 30 min. The reaction was stopped by the addition of SDS sample buffer and then the proteins were separated by SDS-PAGE. Phosphorylated Mig-6 was analyzed by 32P-autoradiography or immunoblotting (IB) with anti-phospho-serine antibody. **B**. Comparison of the phosphorylation status between wild-type and mutant Mig-6 in HEK293 cells. HEK293 cells were transfected with wild-type or the indicated mutant FLAG-Mig-6. After 48 h, cells were harvested, and the cell lysates were separated by 6% Phos-tag SDS-PAGE (upper panel) or normal SDS-PAGE and immunoblotted.

#### Supplementary Figure S4. Analysis of Mig-6 phosphorylation sites.

**A.** Identification of S251 phosphorylation of exogenous Mig-6 using MS. HEK293 cells were transfected with FLAG-Mig-6, quiescence was induced by 16 h serum starvation, then the cells were treated with 20 ng/ml EGF for 15 min. FLAG-Mig-6 was immunoprecipitated with anti-FLAG antibody from the lysate and was resolved by SDS-PAGE. After tryptic digestion of the FLAG-Mig-6 band, the phosphorylated peptides were analyzed by LC-MS. The spectrum of the charged ion (m/z 503.27) shows that S251 is phosphorylated in the indicated peptide. **B.** Identification of S251 phosphorylation of recombinant Mig-6 using MS. Recombinant Mig-6 protein (0.1 μg) was incubated in 20 μl of kinase buffer with <sup>32</sup>P-labeled ATP and 0.1 μg of purified recombinant GST-Chk1 kinase at 30°C for 30 min. The reaction was stopped by the addition of SDS sample buffer, then the proteins were separated by SDS-PAGE, followed by tryptic digestion and LC-MS. The spectrum of the charged ion (m/z 503.82) shows that S251 is phosphorylated in the indicated peptide in the indicated peptide. b ions, fragmentation ions containing the amino terminus of the peptide; y ions, fragmentation ions containing the carboxy terminus of the peptide.



### Supplementary Figure S5. Effect of Mig-6 S251 phosphorylation on EGF signaling.

A. Effect of Mig-6 S251 mutation on EGF signaling in HEK 293 cells. Wild type (WT), S251A, or S251E mutant FLAG-Mig-6 was expressed in HEK293 cells. After 16 h serum starvation, cells were stimulated with 20 ng/ml EGF, then harvested 15 min later. Cell lysates were separated using SDS-PAGE followed by immunoblotting with the indicated antibodies. B. Intensities of the EGF-induced phospho-tyrosine of EGFR (p-Y) in (A) (lane 2, lane 4, lane 6 vs. lane 8) were quantified by image analysis. The data from triplicate experiments were evaluated statistically and are shown graphically relative to the vector only (vec). C. Effect of S251 status on cell growth. Wild-type, S251A, or S251E mutant FLAG-Mig-6 was transfected into HEK293 cells. The cell growth was evaluated at the indicated times. Error bars indicate the standard deviation of three independent experiments. D. Effect of Mig-6 S251 mutation on EGF signaling in MDA-MB-231 cells. MDA-MB-231 cells were infected with retroviruses encoding Mig-6 wild type (WT), S251A, or S251E, and then transfected with an siRNA targeting Mig-6. After 16 h serum starvation, cells were stimulated with 20 ng/ml EGF for 15 min. Cell lysates were separated with SDS-PAGE followed by immunoblotting. The experiments were repeated 3 times and indicated in Fig 4A (Exp. 1) and the supplementary Fig S5D (Exp.2 and 3). The level of p-EGFR (Y1068) protein was measured and normalized to the level of  $\beta$ -actin. The triplicate data were statistically analyzed in Fig 4B.



## Supplementary Figure S6. Effect of EGFR-binding domain mutations in Mig-6 on EGF signaling.

**A.** Wild-type or S302/334/369A mutant FLAG-Mig-6 was co-expressed with EGFR in HEK293 cells. After 16 h serum starvation, cells were stimulated with 20 ng/ml EGF, then harvested at the indicated times. Cell lysates were separated by SDS-PAGE followed by immunoblotting. **B.** Effect of Mig-6 S302A mutation on EGF signaling. MDA-MB-231 cells were infected with retroviruses encoding wild type (WT), or S302A mutant Mig-6, and then transfected with an siRNA targeting *Mig-6*. After 16 h serum starvation, cells were stimulated with 20 ng/ml EGF for 15 min. Cell lysates were separated with SDS-PAGE followed by immunoblotting. **C.** The intensity of p-EGFR (Y1068) protein in (B) was quantified by image analysis. The data from triplicate experiments were evaluated statistically and are shown graphically. **D.** Effect of S302 status on cell growth. MDA-MB-231 cells were infected with retroviruses encoding wild type (WT), or S302A mutant Mig-6, and then treated with an siRNA for *Mig-6*. Cell proliferation assays were performed. The doubling times

r

а

indicated graphically.



## Supplementary Figure S7. EGF-promoted phosphorylation of Chk1 at S280.

**A–C.** EGF signaling promotes activation of PI3K/Akt and phosphorylation of Chk1 at S280. Quiescence was induced in HeLa cells by 16 h serum starvation, the cells were then treated with EGF and harvested at the indicated times. The whole cell lysates were subjected to phosphoiTRAQ (see Materials and Methods). Oscillations in the phosphorylation of EGFR (Y1172, Y1192), PRAS40 (T246), and Chk1 (S280, S301, S331) are indicated.

Exp. 2





Supplementary Figure S8. Modulation of EGFR phosphorylation in MDA-MB-231 cells depleted of Chk1.

MDA-MB-231 cells were transfected with an siRNA for human *Chk1*(01) or *Mig-6*(01), or a control siRNA (cont). Quiescence was induced by 16 h serum starvation, then the cells were stimulated with 20 ng/ml EGF for the indicated times. Cell lysates were separated by SDS-PAGE and subjected to immunoblotting with the indicated antibodies. The experiments were repeated 3 times and indicated in Fig 6A (Exp.1) and the supplementary Fig S8 (Exp.2 and 3). The levels of phosphorylated proteins were measured and normalized to the level of  $\beta$ -actin.



# Supplementary Figure S9. Modulation of EGFR phosphorylation in MDA-MB-231 cells depleted of Chk1.

**A.** MDA-MB-231 cells were transfected with an siRNA for human *Chk1*(01) or *Mig-6*(01), or a control siRNA (cont). Quiescence was induced by 16 h serum starvation, then the cells were stimulated with 20 ng/ml EGF for the indicated times. Cell lysates were separated by SDS-PAGE and subjected to immunoblotting with the indicated antibodies. **B**, **C**. Quantification of the levels of phosphorylated proteins in (A) were normalized to the level of  $\beta$ -actin.



# Supplementary Figure S10. Modulation of EGFR phosphorylation in MDA-MB-231 cells in early stage of EGF stimulation.

**A.** MDA-MB-231 cells were transfected with an siRNA for human *Chk1*(04) or *Mig-6*(05), or a control siRNA (cont), serum starved for 16 h, then stimulated with 20 ng/ml EGF for 5 min, 15 min, or 45 min. The cell lysates were analyzed by immunoblotting with the indicated antibodies. **B**, **C.** Quantification of the levels of phosphorylated proteins in (A) were normalized to the level of  $\beta$ -actin.



# Supplementary Figure S11. Phosphorylation of Mig-6 at S251 affects on binding of Mig-6 to 14-3-3.

**A.** HA-14-3-3  $\xi$ ,  $\beta$ , or  $\delta$ , were co-expressed with FLAG-Mig-6 in HEK293 cells. The cell lysates were subjected to immunoprecipitation with anti-FLAG antibody, followed by immunoblotting with anti-HA antibody. **B.** Depletion of Chk1 inhibited binding of Mig-6 to 14-3-3  $\xi$ . HA-14-3-3  $\xi$ ,  $\beta$ , or  $\delta$  were co-expressed with FLAG-Mig-6 in HEK293 cells. Then Chk1 was depleted by siRNA transfection. The cell lysates were immunoprecipitated with anti-FLAG antibody and immunoblotted with the indicated antibodies.