

Supplemental Data

Table S1. ^{15}N and ^1H temperature coefficients of backbone amide sides in [$U\text{-}^{15}\text{N}$] synthetic elastin.

Peak No.	Temperature coefficient (ppb/K)	
	^{15}N	^1H
1	-32 +/- 7	-5.6 +/- 2.0
2	-7 +/- 5	-2.3 +/- 2.3
3	-20 +/- 4	-5.1 +/- 3.0
4	-35 +/- 7	-4.7 +/- 3.2
5	-32 +/- 7	-5.0 +/- 2.6
6	-40 +/- 21	-5.3 +/- 3.4
7	-14 +/- 3	-4.2 +/- 2.8

Table S2. Three-residue sequences and their occurrences in tropoelastin.

R ₋₁ -Gly-R ₊₁				R ₋₁ -Ala-R ₊₁				R ₋₁ -Val-R ₊₁					
PGV	26	AGV	9	FGL	3	AAA	38	VAP	6	GVG	31	PVP	1
PGG	12	AGL	8	FGA	2	AAK	21	VAA	3	GVP	17	PVV	1
PGA	11	AGA	5	FGG	1	AAG	6	VAG	2	GVA	11	<u>PVX</u>	<u>2</u>
PGF	4	AGI	4	FGP	1	AAQ	2	VAD	1	GVL	5		
PGL	3	AGG	1	FGV	1	AAV	2	<u>VAX</u>	<u>12</u>	GVV	3		
PGI	2	AGK	1	<u>FGX</u>	<u>8</u>	AAR	1			GVK	2		
PGK	1	AGP	1			AAZ	1	LAA	1	GVF	1		
<u>PGX</u>	<u>59</u>	AGQ	1	TGA	1	<u>AAZ</u>	<u>71</u>	LAG	1	GVS	1		
		AGY	1	TGK	1			LAP	1	GVY	1		
GGV	14	<u>AGX</u>	<u>31</u>	TGT	1	GAG	17	<u>LAX</u>	<u>3</u>	<u>GVX</u>	<u>72</u>		
GGL	6			TGV	1	GAA	7						
GGA	5	LGG	9	<u>TGX</u>	<u>4</u>	GAL	5	PAA	4	AVP	3		
GGI	4	LGV	6			GAF	2	PAV	1	AVT	1		
GGP	2	LGA	5	IGG	1	GAI	2	<u>PAX</u>	<u>5</u>	AVV	1		
GGF	1	LGK	1	IGP	1	GAR	2			<u>AVX</u>	<u>5</u>		
GGG	1	LGP	1	<u>IGX</u>	<u>2</u>	GAV	2	EAA	1				
GGK	1	LGY	1			GAC	1	EAQ	1	VVG	2		
GGY	1	<u>LGX</u>	<u>23</u>			<u>GAX</u>	<u>38</u>	<u>EAX</u>	<u>2</u>	VVP	2		
<u>GGX</u>	<u>35</u>									VVS	1		
		YGA	4			KAA	12	QAA	2	<u>VVX</u>	<u>5</u>		
VGW	13	YGV	2			KAG	2						
VGG	9	YGL	1			KAP	2	SAA	1	LVP	3		
VGA	5	YGP	1			KAC	1	SAG	1	LVG	1		
VGI	2	YGY	1			KAQ	1	<u>SAX</u>	<u>2</u>	<u>LVX</u>	<u>4</u>		
VGL	2	<u>YGX</u>	<u>9</u>			<u>KAX</u>	<u>18</u>						
VGP	2							DAA	1	KVA	1		
VGT	2							IAG	1	KVP	1		
<u>VGX</u>	<u>35</u>							RAA	1	<u>KVX</u>	<u>2</u>		
								FAG	1				

Figure S1. Pulse sequence for the “semi-constant time” 2D ^1H - ^{15}N rINEPT-HETCOR experiment (Ref. S1). ^1H chemical shift evolves *via* incrementation of t_{1a} , and the spectral width in the ^1H indirect dimension was 2400 Hz. The t_{1b} delay was defined as $t_{1b} = t_{1a} - 2\tau$. The delay τ was set to 1.6 ms. The phase of the first ^1H 90° pulse was incremented via the States method (Ref. S2). The phase cycling scheme was $\phi 1: [+x]_8 [-x]_8$; $\phi 2: [+x -x]$; $\phi 3: [+y -y]$; $\phi 4: [+x]_4 [+y]_4 [-x]_4 [-y]_4$; $\phi 5: [+x -x]_2 [+y -y]_2$; $\phi \text{rec}: [+x]_2 [-x]_2 [+y]_2 [-y]_2$ (Ref. S3).

Figure S2. Spin rate dependence of ^{15}N rINEPT (A) and ^1H MAS (B) spectra. Dependence of spectral intensities on ν_r for ^{15}N rINEPT spectra is shown in (C).

Figure S3. Detailed illustration of the calculated contour plot, using Ref. S4 (denatured peptide measurements). (A) The black lines point to the calculated ^{15}N and ^1H chemical shifts for the indicated 3-aa sequences. A Gaussian is centered at each set of chemical shift values and is weighted by the relative number of occurrences of the 3-aa sequence in tropoelastin (given in parentheses). “Peaks” result from overlap of chemical shifts with little or no differences and/or high occurrences of 3-aa sequences in tropoelastin. (B) Color contours only. Numbers in parentheses give the total number of occurrences for these sequences in tropoelastin.

Figure S4. Comparison of calculated and experimental (5 °C) chemical shifts, using the semi-empirical methods of Refs. S4 (denatured peptide) and S5 (compiled database information). Comparisons are made with the random coil (S4A-S4D), β -strand (S4E-S4F), and the α -helix (S4G-S4H). The method using the denatured peptide (ref. S4) is used for S4A-S4B, whereas the database method (ref. S5) is used for the rest. The experimental spectra are illustrated with the grey contours. Color contours denote the calculated ^{15}N and ^1H chemical shifts for the Gly (G), Ala (A), and Val (V), with assigned amino acid in **bold**. An “X” indicates 2 or more residues in the 3rd position of the 3-aa sequence. The predicted “peaks” were plotted as Gaussians that are centered at the (predicted) shifts and weighted by the number of occurrences in the sequence of tropoelastin.

References

- S1. Helmus, J. J., Surewicz, K., Surewicz, W. K., and Jaroniec, C. P. (2010) *J. Am. Chem. Soc.* **132**, 2393-2403
- S2. States, D. J., Haberkorn, R. A., and Ruben, D. J. (1982) *J. Magn. Reson.* **48**, 286-292
- S3. Elena, B., Lesage, A., Steuernagel, S., Bockmann, A., and Emsley, L. (2005) *J. Am. Chem. Soc.* **127**, 17296-17302
- S4. Schwarzinger, S., Kroon, G. J. A., Foss, T. R., Chung, J., Wright, P. E., and Dyson, H. J. (2001) *J. Am. Chem. Soc.* **123**, 2970-2978
- S5. Wang, Y. J., and Jardetzky, O. (2002) *J. Am. Chem. Soc.* **124**, 14075-14084

Figure S1

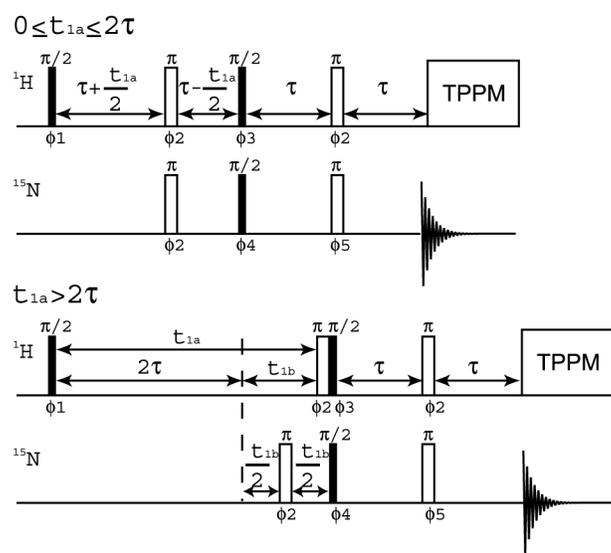
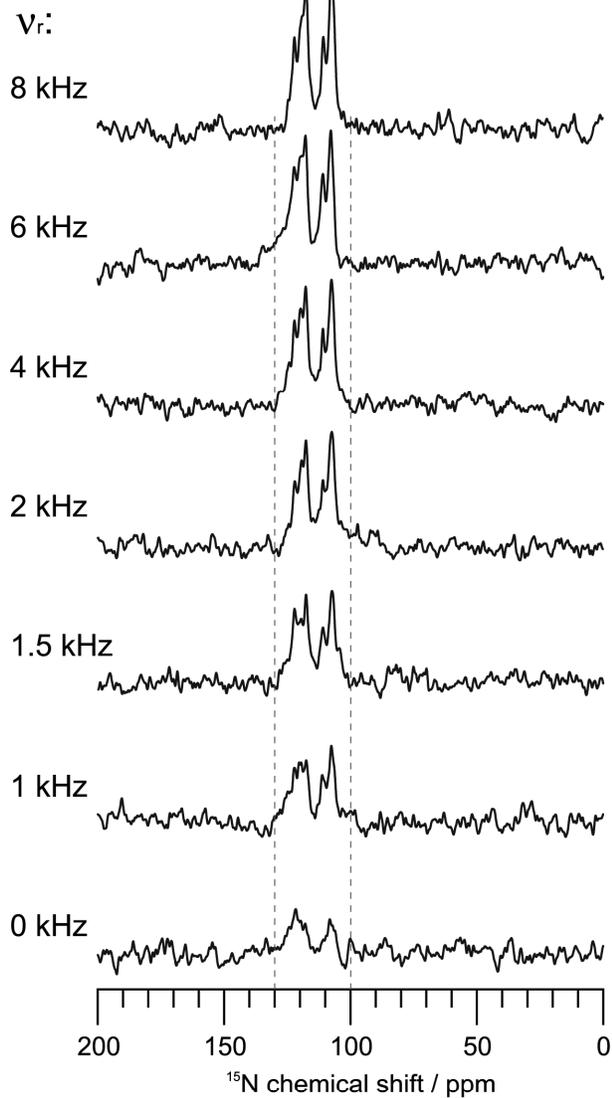
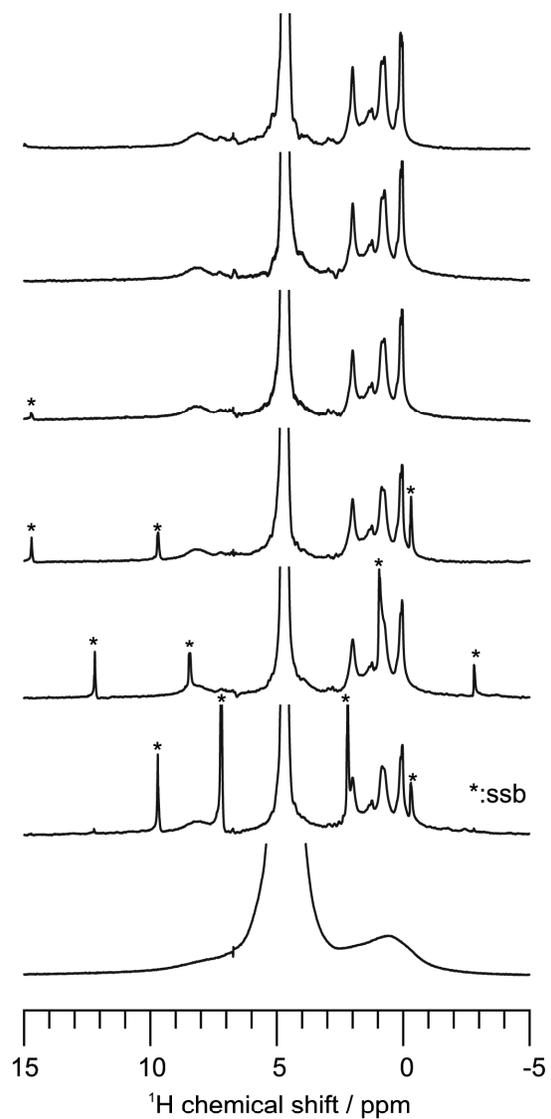


Figure S2

A) ^{15}N rINEPT



B) ^1H MAS



C)

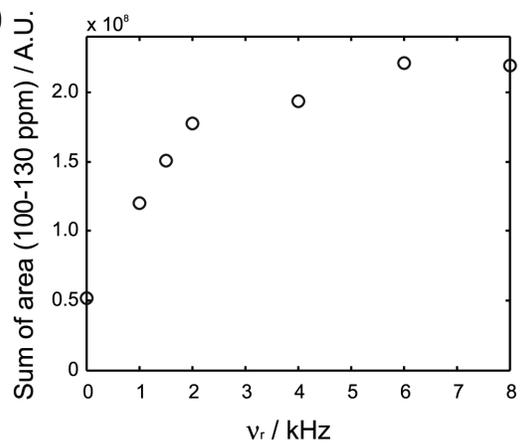


Figure S3

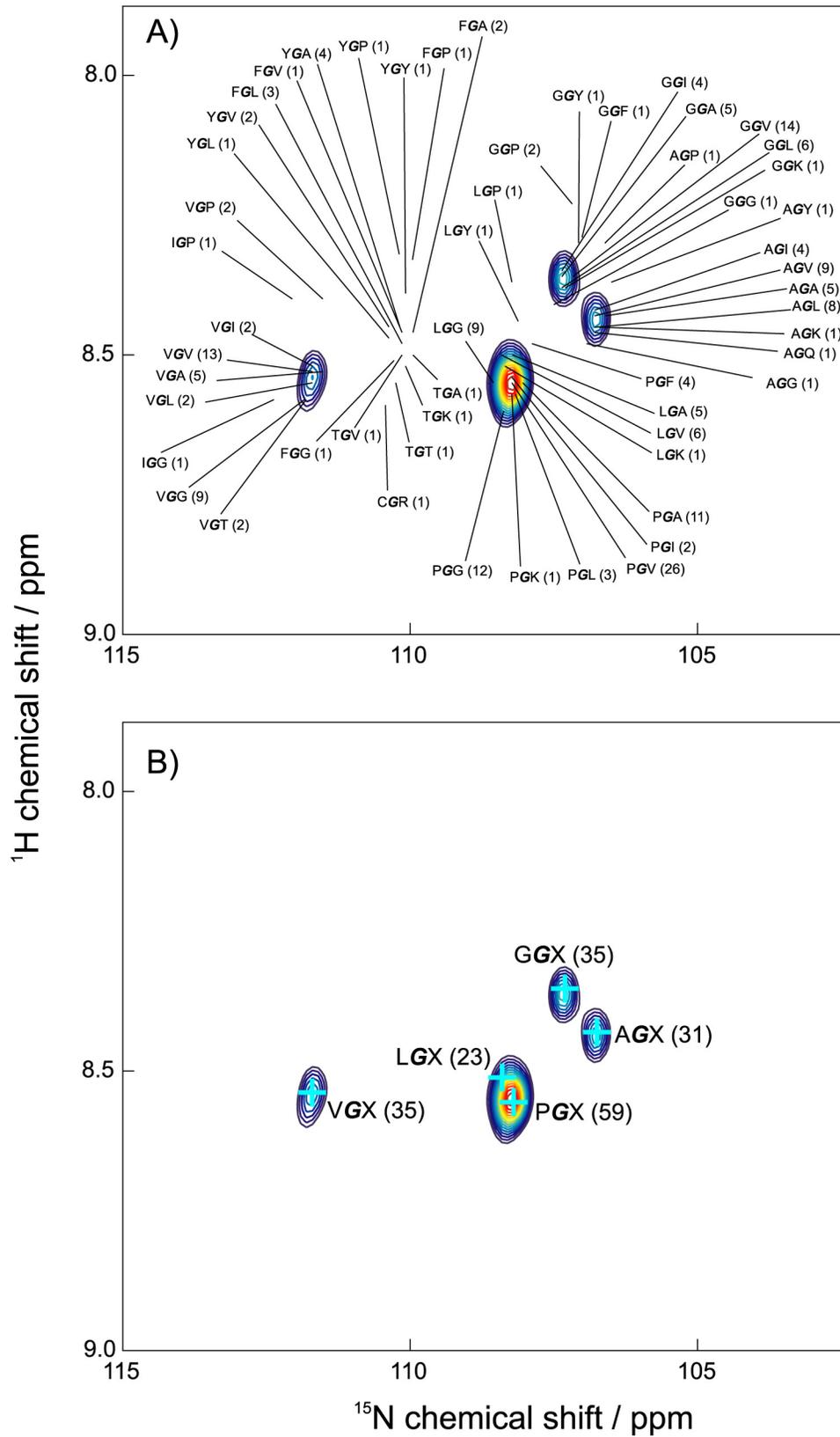


Figure S4

