

**Table S1. Primers used for cloning of *PAA1* and *PAA2***

<b>AGI</b>	<b>Gene</b>	<b>Sequence (5' to 3')</b>	<b>Sense</b>
At4g33520	<i>PAA1</i> <i>NcoI</i>	catgccatgGAGAGTGGTGATTCCAAGTCAAAACTGGG	Forward
	<i>PAA1</i> <i>BamHI</i>	cgcgatccCTCGCGCCACTCTCTTTAAGCG	Reverse
At5g21930	<i>PAA2</i> <i>NcoI</i>	catgccatgGAATCTTCAATCGAATCTGTGAAATCCATT	Forward
	<i>PAA2</i> <i>BamHI</i>	cgcgatccACGGTTCCTGCTCTTAACAAGCAA	Reverse
<b>qRT-PCR primers</b>			
<b>AGI</b>	<b>Gene</b>	<b>Sequence (5' to 3')</b>	<b>Sense</b>
At5g08290	Mitosis protein <i>YLS8</i>	T TACTGTTTCGGTTGTTCTCCATTT C ACTGAATCATGTTCTGAAGCAAGT	Forward Reverse
At2g28390	<i>SAND</i> family	A AACTCTATGCAGCATTGATCCACT T GATTGCATATCTTTATCGCCATC	Forward Reverse
At5g21930	<i>PAA2</i>	A AATCTAGCGTGGGCAATTGCG C TCCAAAATAGATGTGGAGCCG	Forward Reverse
At4g33520	<i>PAA1</i>	A ACGACAAAATGTCAAACCGG C CCCTCTCAAGACCAAGAGC	Forward Reverse
At1g12520	<i>CCS</i>	A AGATCAAACCTGGCACAGAGCC G C TCTCCCGATAAGGTCTGC	Forward Reverse
<b>miRNA Stem-loop qRT-PCR primers</b>			
<b>AGI</b>	<b>Gene</b>	<b>Sequence (5' to 3')</b>	<b>Sense</b>
At2g47015	<i>miR408</i> RT	GTCGTATCCAGTGCAGGGTCCGAGGTATTTCGACTGGATACGACGCCAGG	Reverse
	<i>miR408</i> SL	G TGTGATGCACTGCCTCTTC	Reverse
At3g63375	<i>miR167</i> RT	GTCGTATCCAGTGCAGGGTCCGAGGTATTTCGACTGGATACGACTAGATC	Reverse
	<i>miR167</i> SL	T C GCGTGAAGCTGCCAGCAT	Reverse
-	miRUniversal	C CAGTGCAGGGTCCGAGG	Forward
<b><i>miR408</i> mutant line verification</b>			
<b>AGI</b>	<b>Gene</b>	<b>Sequence (5' to 3')</b>	<b>Sense</b>
At2g47015	<i>miR408F</i>	A GAGGGTTAGGAATACAGTTGG	Forward
	<i>miR408R</i>	C ATTTGTTTCACCCACTACGC	Reverse
T-DNA	LBb1.3	A TTTTGCCGATTCGGAAC	Forward

**Table S2. Cellular Cu content**

Cu content was measured for protoplasts and chloroplasts of wild-type (Col-0), *paa1-3*, and *ccs* plants grown *in vitro* for 4 weeks in the presence of 0.05  $\mu$ M or 5  $\mu$ M CuSO<sub>4</sub>. Values for protoplasts (cells) and chloroplasts of each line and treatment are given in ng Cu/ mg chlorophyll and represent averages of three measurements  $\pm$  SD. The ratio represents the fraction of cellular Cu found in the chloroplasts.

CuSO <sub>4</sub>	Col-0		<i>paa1-3</i>		<i>ccs</i>	
	0.05 $\mu$ M	5 $\mu$ M	0.05 $\mu$ M	5 $\mu$ M	0.05 $\mu$ M	5 $\mu$ M
Protoplast	<b>333 <math>\pm</math> 70</b>	<b>500 <math>\pm</math> 98</b>	<b>385 <math>\pm</math> 68</b>	<b>580 <math>\pm</math> 162</b>	<b>312 <math>\pm</math> 63</b>	<b>523 <math>\pm</math> 145</b>
Chloroplast	<b>234 <math>\pm</math> 38</b>	<b>440 <math>\pm</math> 163</b>	<b>137 <math>\pm</math> 41</b>	<b>247 <math>\pm</math> 75</b>	<b>265 <math>\pm</math> 8</b>	<b>382 <math>\pm</math> 174</b>
Ratio	<b>0.7</b>	<b>0.88</b>	<b>0.36</b>	<b>0.43</b>	<b>0.85</b>	<b>0.73</b>

## SUPPLEMENTARY FIGURES

### FIGURE S1. PAA1 and PAA2 antibody specificity

(a,b) Immunoblot analysis of PAA2, PAA1 and cFBPase in rosette leaves of wild-type (Col-0), *paa2-1* and *paa1-1* seedlings grown *in vitro* for 18 days on agar solidified half-strength MS. *paa1-1* and *paa2-1* were used as negative controls for the respective antigens. \*indicates an unspecific band.

### FIGURE S2. Root-length of metal treated wild-type (Col-0) plants

Root length of 11 days old Col-0 seedlings grown vertically in the absence (Ø) or presence of different metals. Results represent the average  $\pm$  SD ( $n \geq 13$  seedlings per treatment).

### FIGURE S3. *miR408* knock-out line characterization

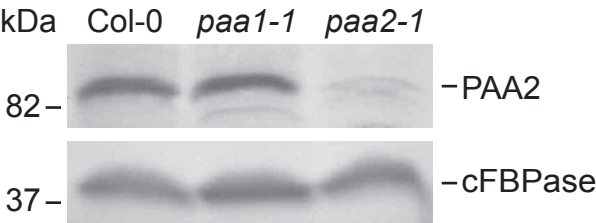
(a) Map of the putative *miR408* targeting site within the *PAA2* coding sequence. The schematic drawing furthermore indicates the location of the T-DNA insertion within the *miR408* coding sequence. The primer binding sites for verification of the T-DNA insert are indicated. Exons are represented by black boxes and introns by black lines. (b) The presence of the T-DNA insert was analyzed by PCR using the primers listed in Table S1. (c) Expression levels of mature *miR408* measured via stem-loop qRT-PCR in the wild-type (Col-0) and *miR408* knock out background. Plants were grown in the presence of 0.05  $\mu$ M CuSO<sub>4</sub> (left, black bars) or 5  $\mu$ M CuSO<sub>4</sub> (right, grey bars). Results represent the average ( $\pm$  SD) of three biological replicates including two technical replicates each. n.d. means not detectable.

### FIGURE S4. Intactness of isolated protoplasts and chloroplasts

Immunoblot analysis of isolated protoplasts (P) and chloroplasts (C) from Col-0, *paa1-3* and *ccs* plants, grown in the presence of the indicated CuSO<sub>4</sub> concentrations for 4 weeks. Protein extract, equivalent to 1.7  $\mu$ g chlorophyll was loaded into each lane. Blots were probed for PC and CCS using specific antibodies. cFBPase serves as a cytosolic marker and ferredoxin (FDX) as stromal marker.

Figure S1

**A**



**B**

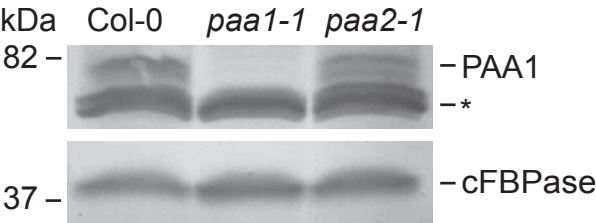


Figure S2

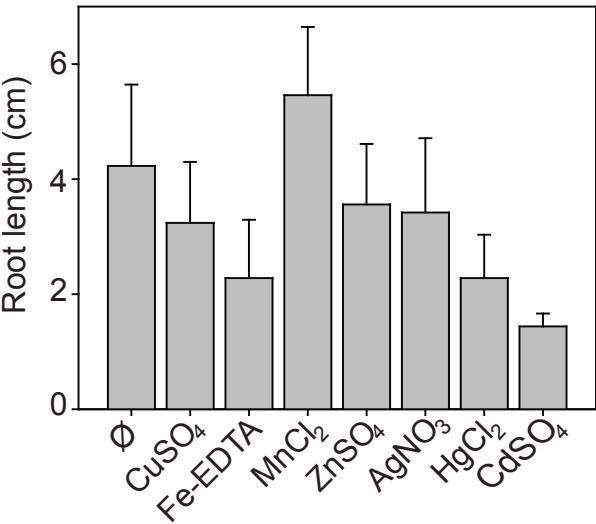
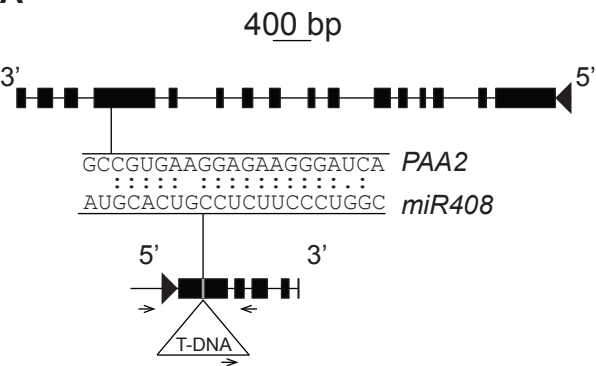
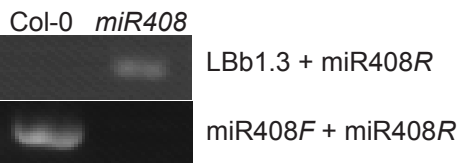


Figure S3

**A**



**B**



**C**

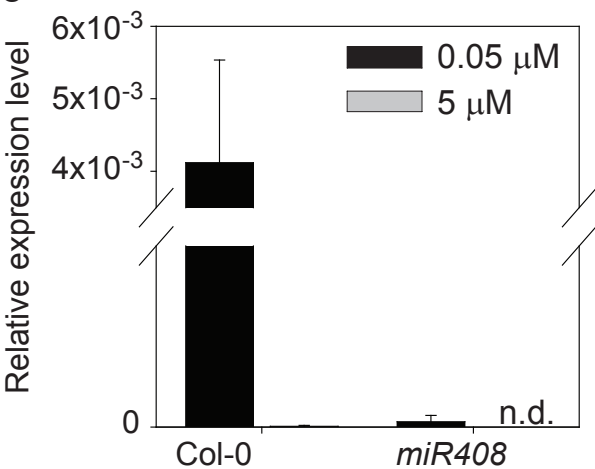


Figure S4

