

Supplementary Data (Calvo et al., 2012)

Supplementary Table S1. Strains used in this study

Strain	Genotype	Origin
972	<i>h⁻</i>	(39)
NG25	<i>h⁺ caf4/trr1::ura4⁺ ura4-D18</i>	(40)
EHH14	<i>h⁻ his2 ura4 pap1::ura4⁺ leu1-32 nmt::GFP-pap1::leu1⁺</i>	(41)
EHH14.C523D	<i>h⁻ his2 ura4 pap1::ura4-D18 leu1-32 nmt::GFP pap1C523D::leu1⁺</i>	(41)
AV19	<i>h⁺ trr1::ura4⁺ ura4-D18 ade6-704 leu1-32 nmt::GFP-pap1::leu1⁺</i>	This work
<i>caf1⁺::ura4⁺</i>	<i>h⁺ caf1/hba1::ura4⁺ ura4-D18 ade6-704 leu1-32</i>	(42)
EA33	<i>h⁺ caf1::ura4⁺ ura4-D18 ade6-704 leu1-32 nmt::GFP-pap1::leu1⁺</i>	(41)
IC1	<i>h⁻ pap1::ura4⁺ ura4-D18 leu1-32</i>	This work
IC2	<i>h⁻ pap1 ura4-D18 leu1-32</i>	This work
IC2.C523D	<i>h⁻ pap1.C523D ura4-D18 leu1-32</i>	This work
MS46	<i>h⁺ atf1::kanMX6 ura4-D18 leu1-32</i>	This work
MC16	<i>h⁻ prr1::kanMX6 ura4-D18 leu1-32</i>	This work
MC18	<i>h⁻ pap1::ura4⁺ ura4-D18 prr1::kanMX6 leu1-32</i>	This work
MC40	<i>h⁻ prr1-HA::kanMX6 ura4-D18 leu1-32</i>	This work
IC64	<i>h⁻ prr1-HA::kanMX6 pap1::natMX6 ura4-D18 leu1-32</i>	This work
MC41	<i>h⁻ prr1-GFP::kanMX6 ura4-D18 leu1-32</i>	This work
MC42	<i>h⁻ pap1-GFP::kanMX6 ura4-D18 leu1-32</i>	This work
IC81	<i>h⁻ prr1::kanMX6 leu1-32 nmt::GFP-pap1::leu1⁺ ura4-D18</i>	This work
IC83	<i>h⁺ pap1::ura4⁺ prr1-GFP::kanMX6 his2 ura4-D18 leu1-32</i>	This work
PG3	<i>h⁺ prr1-HA::kanMX6 pap1.C523D::natMX6 ura4-D18 leu1-32</i>	This work
PG7	<i>h⁻ caf4::ura4⁺ prr1-HA::kanMX6 leu1-32</i>	This work
IC70	<i>h⁻ pap1-HA::kanMX6 ura4-D18 leu1-32</i>	This work
IC102	<i>h⁻ trr1::ura4⁺ prr1-GFP::natMX6 leu1-32</i>	This work
PG20	<i>h⁻ hba1::ura4⁺ prr1-GFP::natMX6 ade6-704 leu1-32</i>	This work
IC97	<i>h⁻ pap1.C523D::ura4⁺ prr1-GFP::natMX6 leu1-32</i>	This work
PG21	<i>h⁻ prr1::kanMX6 pap1::natMX6 ura4-29 leu1-32</i>	This work
PG15	<i>h⁻ pap1::ura4⁺ atf1::natMX6 leu1-32</i>	This work
EP137	<i>h⁻ mpr1::natMX6 ura4-D18 leu1-32</i>	This work
IC86	<i>h⁻ mpr1::natMX6 prr1-HA::kanMX6 ura4-D18 leu1-32</i>	This work

Supplementary references

39. Leupold, U. (1970) Genetical methods for *Schizosaccharomyces pombe*. *Methods Cell Physiol.*, **4**, 169-177.
40. Calvo, I.A., Gabrielli, N., Iglesias-Baena, I., Garcia-Santamarina, S., Hoe, K.L., Kim, D.U., Sanso, M., Zuin, A., Perez, P., Ayte, J. *et al.* (2009) Genome-wide screen of genes required for caffeine tolerance in fission yeast. *PLoS One*, **4**, e6619.
41. Castillo, E.A., Ayte, J., Chiva, C., Moldon, A., Carrascal, M., Abian, J., Jones, N. and Hidalgo, E. (2002) Diethylmaleate activates the transcription factor Pap1 by covalent modification of critical cysteine residues. *Mol.Microbiol.*, **45**, 243-254.
42. Benko, Z., Sipiczki, M. and Carr, A.M. (1998) Cloning of *caf1+*, *caf2+* and *caf4+* from *Schizosaccharomyces pombe*: their involvement in multidrug resistance, UV and pH sensitivity. *Mol.Gen.Genet.*, **260**, 434-443.

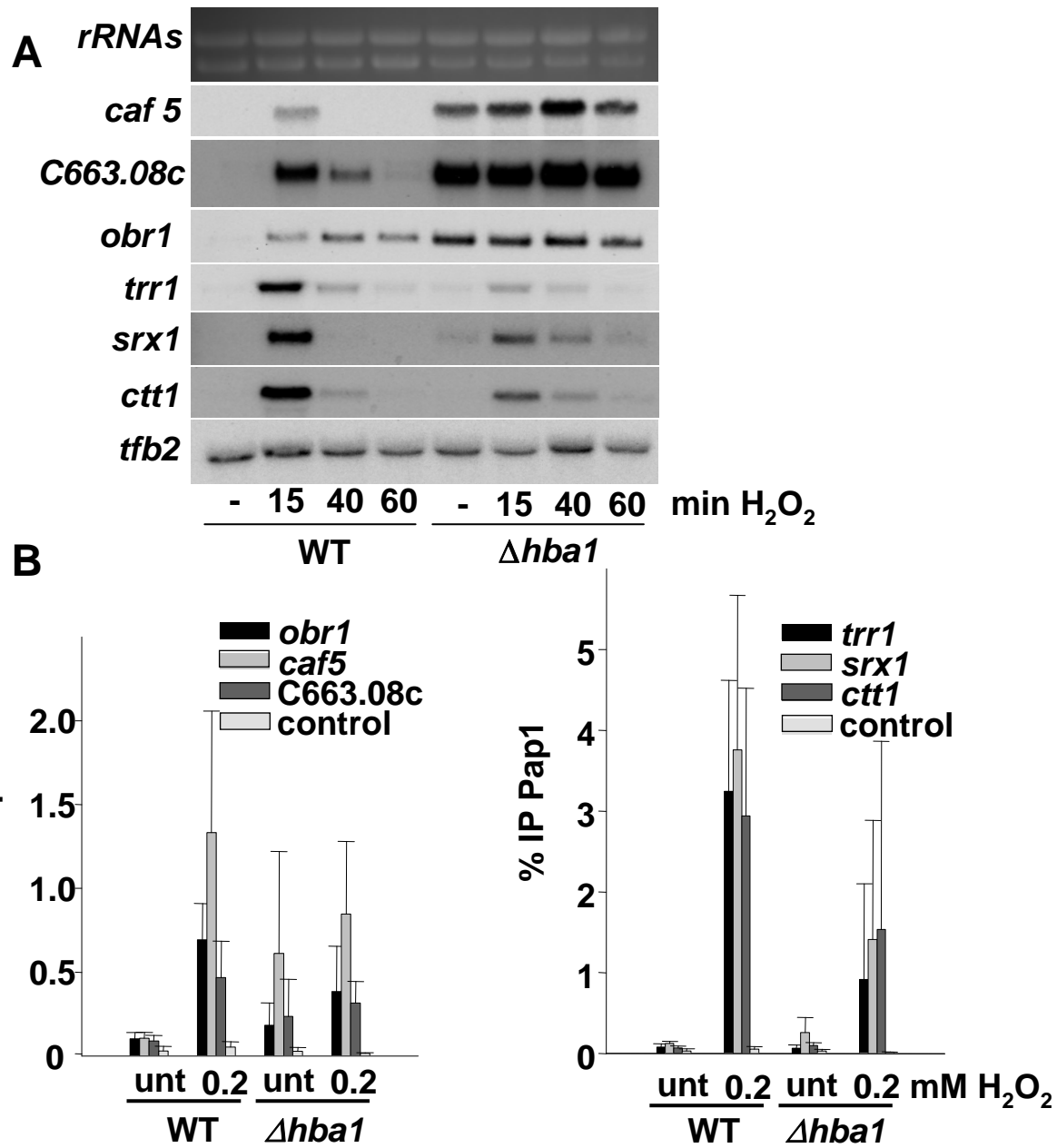
SUPPLEMENTARY FIGURE LEGENDS – Calvo *et al.* 2012

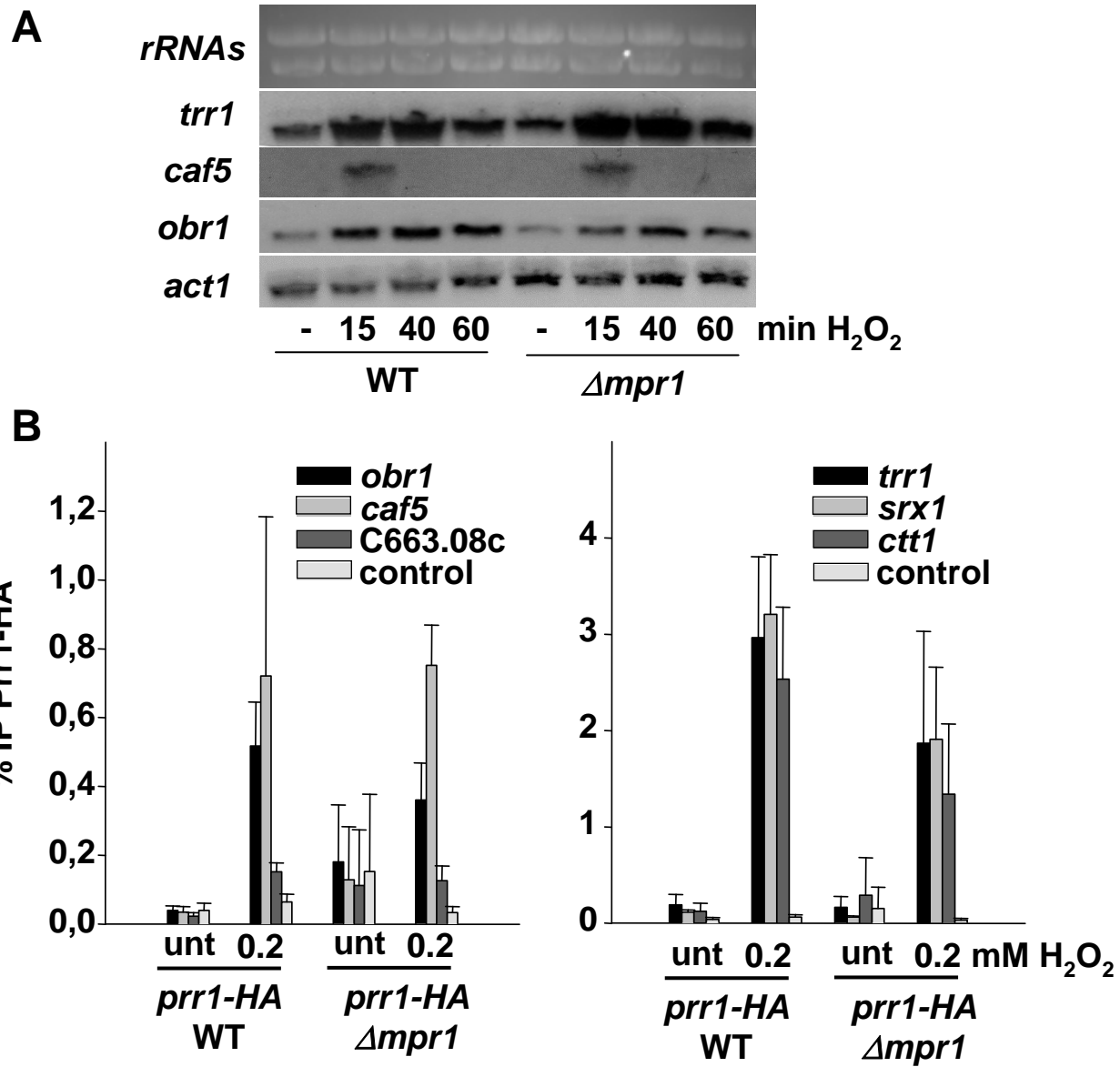
Supplementary Figure S1. Constitutively reduced/nuclear Pap1 can only bind and activate a subset of all Pap1-dependent genes. **(A)** Stress-dependent transcriptional analysis of wild-type and mutant strains. Total RNA from strains IC2 (WT) and $\Delta hba1$ (*caf1::ura4⁺*) was obtained from cultures treated or not with 0.2 mM H₂O₂ for the indicated times and analyzed by Northern blot as described in Figure 2A. **(B)** Binding of reduced/nuclear Pap1 to DNA *in vivo*. Cultures of strains such as those used in A were treated with 0.2 mM H₂O₂ for 5 min. CHIP analysis was performed as described in Figure 2B.

Supplementary Figure S2. Regulation of Prr1 by Mpr1 is not necessary for induction of Pap1-dependent genes. **(A)** Total RNA from strains IC2 (WT) and EP137 ($\Delta mpr1$) was obtained from cultures treated with 0.2 mM H₂O₂ for the indicated times, and analyzed by Northern blot as described in Figure 2A. **(B)** Binding of Prr1-HA to DNA *in vivo* in $\Delta mpr1$ strain. Cultures of strains MC40 (*prr1-HA*) and IC86 ($\Delta mpr1$ *prr1-HA*) were treated or not with 0.2 mM H₂O₂ for 5 min. CHIP of Prr1-HA using anti-HA antibody was performed as described in Figure 2B.

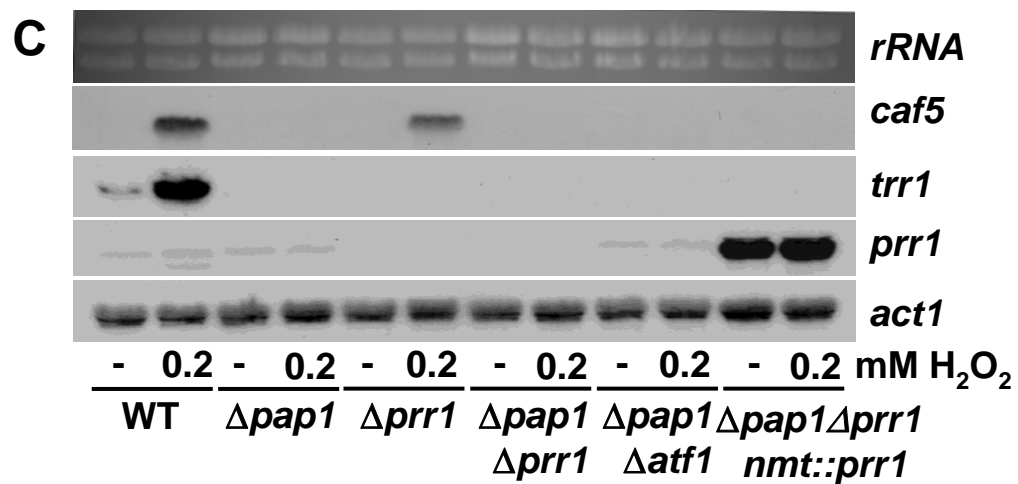
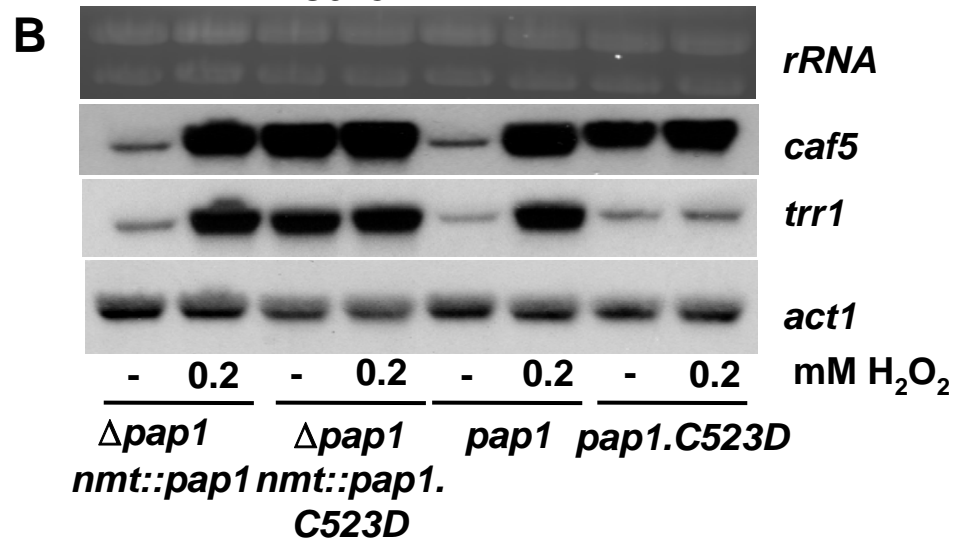
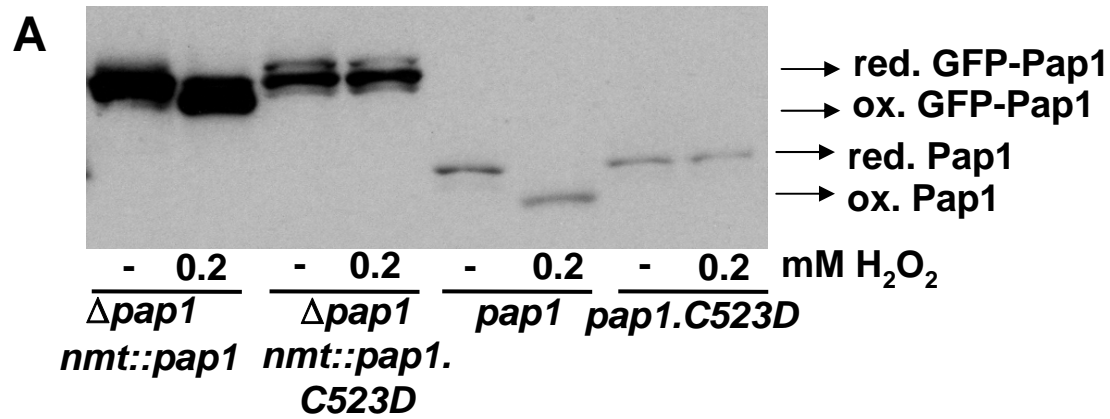
Supplementary Figure S3. Over-expression of reduced/nuclear Pap1 triggers constitutive transcription of not only *caf5* but also *trr1*. **(A)** The redox state and relative protein levels of Pap1 of extracts from strains EHH14 ($\Delta pap1$ *nm1::pap1*; the expression of GFP-Pap1 is under the control of the *nm141* promoter), EHH14.C523D ($\Delta pap1$ *nm1::pap1.C523D*), IC2 (WT) and IC2.C523D (*pap1.C523D*), before (-) or after 5 min of 0.2 mM H₂O₂ stress, was analyzed by Western blot using polyclonal anti-Pap1 antibodies. **(B)** Northern blot analysis of antioxidant Pap1-dependent gene (*trr1*) and drug tolerance gene (*caf5*) expression. Total RNA from strains as in Figure S3A was obtained from cultures treated or not (-) with 0.2 mM H₂O₂ for 15 min and analyzed by Northern blot as described in Figure 2A. **(C)** Prr1 over-expression does not trigger induction of Pap1-dependent genes. Total RNA from strains IC2 (WT), IC1 ($\Delta pap1$), MC16 ($\Delta prr1$), PG21 ($\Delta pap1$ $\Delta prr1$), PG15 ($\Delta pap1$ $\Delta atf1$) and PG21 with p397.41 ($\Delta pap1$ $\Delta prr1$ with *pnmt41::prr1*), treated or not (-) with 0.2 mM H₂O₂ for 15 min, was obtained and analyzed by Northern blot as described in Figure 2A.

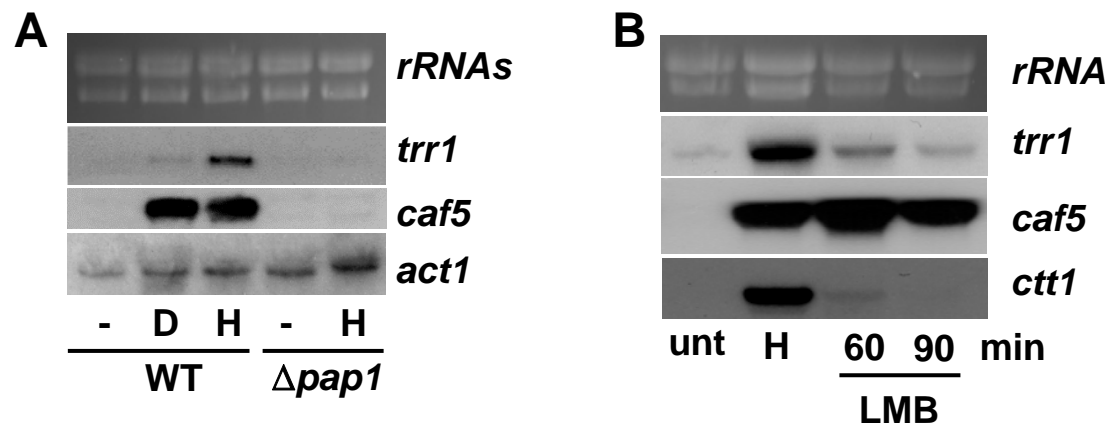
Supplementary Figure S4. Diethylmaleate and Leptomycin B do not induce the antioxidant response. **(A)** Total RNA from strains IC2 (WT) and IC1 ($\Delta pap1$) was obtained from cultures treated with 0.2 mM H₂O₂ for 15 min (H) and 4 mM diethylmaleate for 30 min (D) and analyzed by Northern blot as described in Figure 2A. **(B)** RNA from wild-type strain IC2 was obtained from cultures treated with 0.2 mM H₂O₂ for 15 min (H) or 100 μ M of Leptomycin B (LMB) for 60 and 90 min and analyzed by Northern blot as described in Figure 2A.





Calvo et al. Supplementary Figure S2





Calvo et al. Supplementary Figure S4