

Supplementary material

Table S1. PCR primers used in this study.

| Primer | Sequence | Gene | Target organisms | Reference |
|-------------|------------------------------|-------------------------------|-----------------------|------------|
| 27f | 5'- AGAGTTTGATCCTGGCTCAG-3' | 16S rRNA | Bacteria | (3) |
| 1494r | 5'- CTACGGCTACCTTGTTACGA -3' | 16S rRNA | Universal | (3) |
| 341f* | 5'- CCTACGGGAGGCAGCAG-3' | 16S rRNA | Bacteria | (4) |
| 907r | 5'- CCGTCAATTCMTTGTGAGTTT-3' | 16S rRNA | Bacteria | (3) |
| Rick653f | 5'- GAGTGTAGTAGGGGATGATG -3' | 16S rRNA | <i>Rickettsia</i> | This study |
| Rick982r | 5'- CCACCATGTCAAGGGTTGGT -3' | 16S rRNA | <i>Rickettsia</i> | This study |
| Cox sp434f | 5'- CCTTTTGAGCGTTGACGTTA-3' | 16S rRNA | <i>Coxiela sp.</i> | This study |
| Cox sp1004r | 5'- CCAAAGGCACCAAGTCATTT -3' | 16S rRNA | <i>Coxiela sp.</i> | This study |
| Acari412f | 5'- CGGGACTCTTTTGAGGCC-3' | 18S rRNA | Acari | This study |
| Acari990r | 5'- ATCCTCCCAGTGTCCG-3' | 18S rRNA | Acari | This study |
| T1BF | 5'-AAACTAGGATTAGATACCCT-3' | 12S rRNA | Arthropoda | (1) |
| T2AR | 5'-AATGAGAGCGACGGGCGATGT-3' | 12S rRNA | Arthropoda | (1) |
| SNR | 5'-AATTGACATCCTATTTCAAA-3' | 12S rRNA | <i>Rh. sanguineus</i> | This study |
| 17kdf-1f | 5'- ATGAGTAAAGACGGTAACCT-3' | Gene 'D' 17kd surface antigen | <i>Rickettsia</i> | (5) |
| 17kd-1390r | 5'- CTTGCTTTTCAGCAATATCAC-3' | Gene 'D' 17kd surface antigen | <i>Rickettsia</i> | (5) |
| OmpA-70f | 5'- ATGGCGAATATTTCTCCAAAA-3' | Outer membrane protein A | <i>Rickettsia</i> | (2) |
| OmpA701r | 5'- GTTCCGTTAATGGCAGCATCT-3' | Outer membrane protein A | <i>Rickettsia</i> | (2) |

* For PCR-DGGE analyses a 5' GC rich tail was added to the primer: CGCCCGCCGCGCCCGCGCCCGTCCCGCCGCCCCGCCC (4).

Table S2. PCR and qPCR conditions for the different primer pairs used in this study.

| Primer pair | Use | Initial denaturing | No. of Cycles | Denaturing | Annealing | Extension | Final extension |
|---------------------|----------|--------------------|---------------|-------------|-------------|--------------|------------------|
| 27f-14942r | PCR | 95°C, 5 min | 35 | 95°C,30 sec | 60°C,30 sec | 72°C, 60 sec | 72°C, 5 min |
| 341f-907r | PCR | 95°C, 5 min | 35 | 95°C,30 sec | 58°C,30 sec | 72°C, 30 sec | 72°C, 5 min |
| T1B-T2A | PCR | 94°C, 5 min | 35 | 94°C,30 sec | 47°C,30 sec | 72°C, 60 sec | 72°C, 5 min |
| T1B-SNR | PCR | 94°C, 5 min | 35 | 94°C,30 sec | 54°C,30 sec | 72°C, 60 sec | 72°C, 5 min |
| ompA-70f-701r | PCR | 95°C, 5 min | 35 | 95°C,30 sec | 58°C,30 sec | 72°C, 30 sec | 72°C, 5 min |
| 17kd-1f-1390r | PCR | 95°C, 5 min | 35 | 95°C,30 sec | 60°C,30 sec | 72°C, 60 sec | 72°C, 5 min |
| 341f (GC)-907r | PCR-DGGE | 95°C, 5 min | 35 | 95°C,30 sec | 58°C,30 sec | 72°C, 30 sec | 72°C, 5 min |
| Rick653f-Rick982r | qPCR | 95°C, 15 min | 40 | 95°C,30 sec | 60°C,30 sec | 72°C, 30 sec | Melting protocol |
| Acari412f-Acari990r | qPCR | 95°C, 15 min | 40 | 95°C,30 sec | 60°C,30 sec | 72°C, 30 sec | Melting protocol |
| Cox sp-434f-1004r | qPCR | 95°C, 15 min | 40 | 95°C,30 sec | 60°C,30 sec | 72°C, 30 sec | Melting protocol |

Table S3: The average standard curves slopes, correlation coefficients and amplification efficiencies of the various qPCR assays used in this study

| Assay | Slope | R ² | Efficiency |
|---------------------|-------|----------------|------------|
| <i>Coxiella</i> sp. | -3.33 | 0.99 | 99.74 |
| <i>Rickettsia</i> | -3.37 | 0.98 | 99.11 |
| Tick | -3.37 | 0.99 | 99.08 |

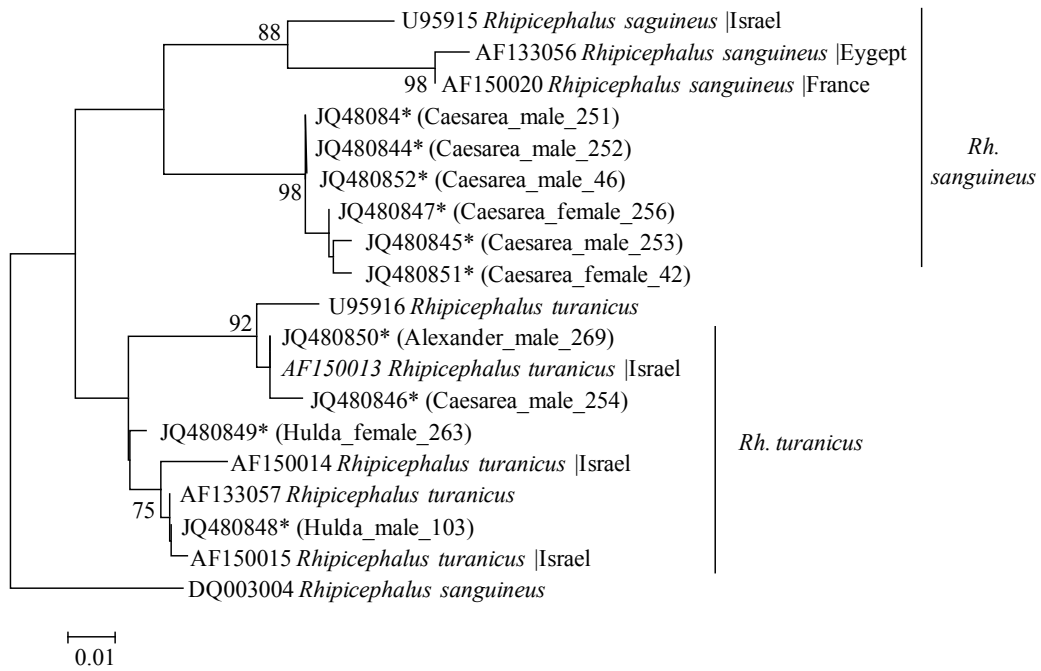


Figure S1. Phylogenetic tree based on Ticks 12S mitochondrial rRNA gene fragments. Maximum-likelihood tree based on Kimura 2-parameter model was constructed using MEGA software (version 5.05). Bootstrap analyses with 1,000 re-sampling were performed to test the robustness of the branching. Bootstrap values higher than 75% are presented. Sequences obtained in the present work are designated by asterisk. The place of collection the sex and the tick number are shown in parentheses. The bar indicates 0.01 substitutions per nucleotide per site.

References

1. **Beati L, Keirans JE.** 2001. Analysis of the systematic relationships among ticks of the genera *Rhipicephalus* and *Boophilus* (Acari : Ixodidae) based on mitochondrial 12S ribosomal DNA gene sequences and morphological characters. *J. Parasitol.* **87**:32-48

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3. **Lane DJ.** 1991. 16S/23S rRNA sequencing, p. 115-175. *In* Goodfelloe M, Stackebrandt E (ed), *Nucleic acid techniques in bacterial systematics.* John Wiley and Sons, Chichester, UK.
4. **Muyzer G, de Waal EC, UitterlindenAG.** 1993. Profiling of complex microbial populations by denaturing gradient gel electrophoresis analysis of polymerase chain reaction-amplified genes coding for 16S rRNA. *Appl. Environ. Microbiol.* **59**:695-700.
5. **Sekeyova Z, Roux V, Raoult D.** 2001. Phylogeny of *Rickettsia* spp. inferred by comparing sequences of 'gene D', which encodes an intracytoplasmic protein. *Int. J. Syst. Evol. Microbiol.* **51**:1353-1360.