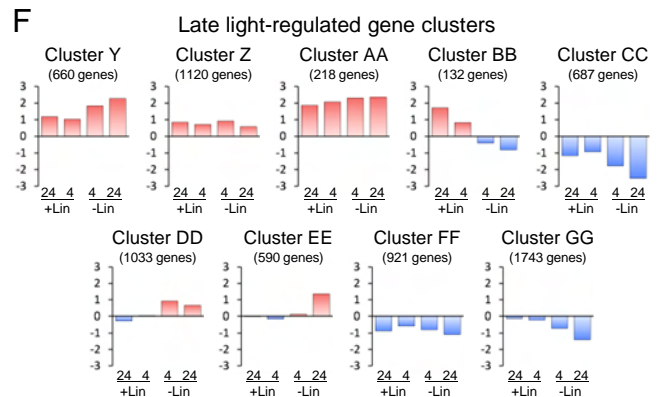
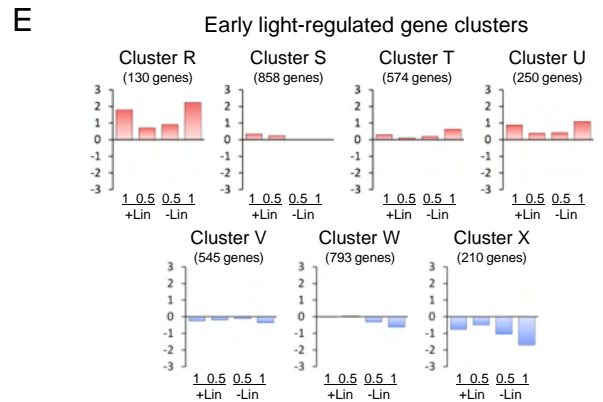
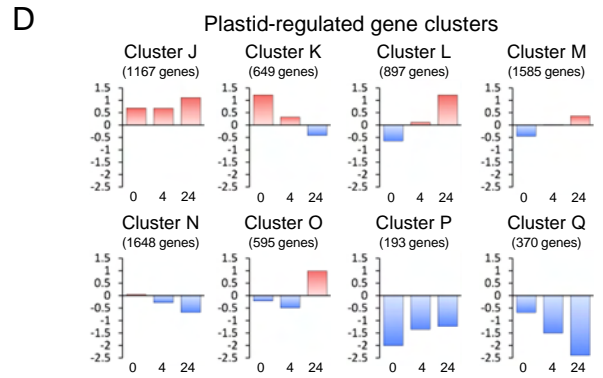
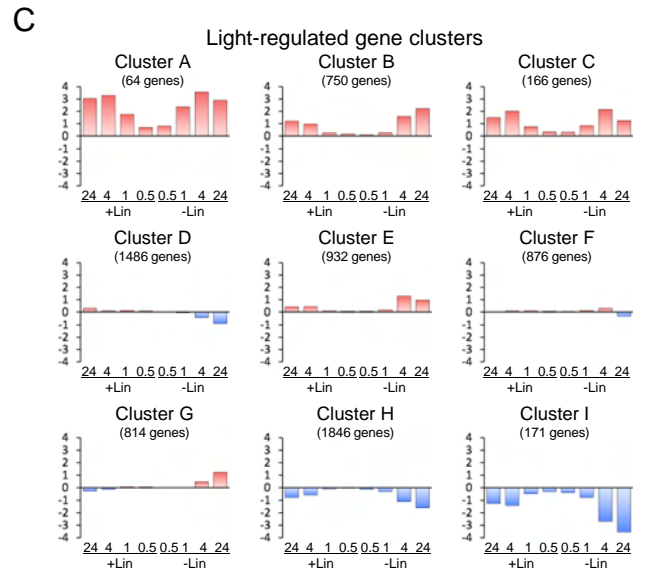
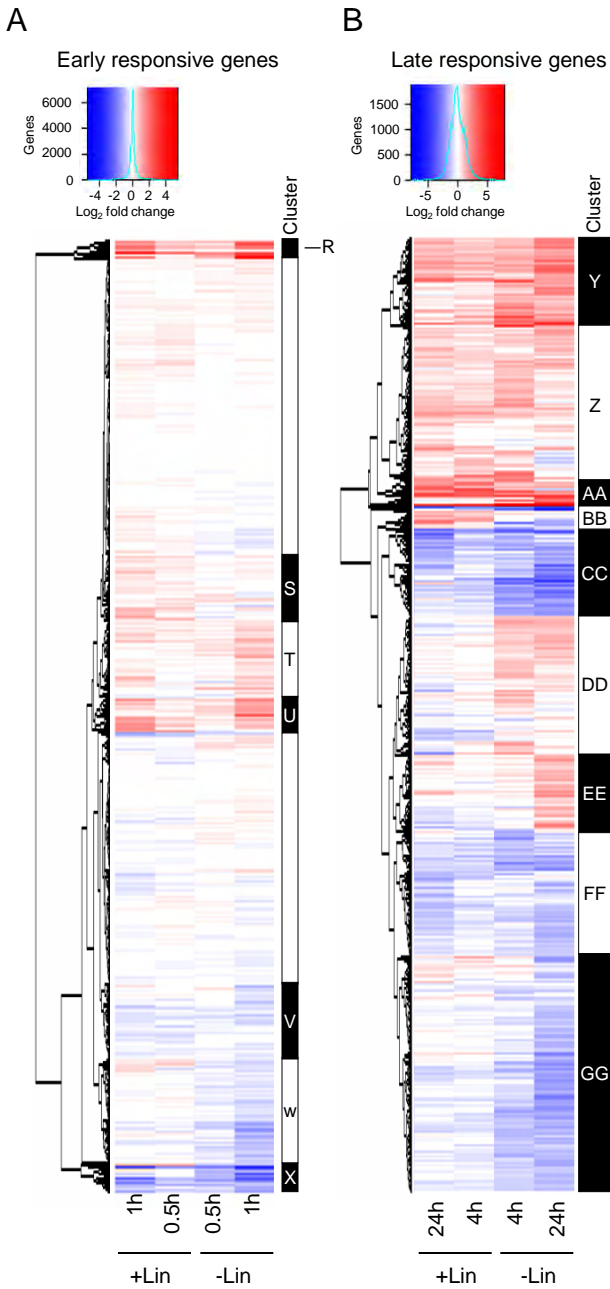


**Supplemental Figure S1.** Expression of *Lhcb1* and *RbcS* following a fluence-rate shift.

**A,** Expression of *Lhcb1* and *RbcS* in lincomycin-treated seedlings after a fluence-rate shift. Seedlings were grown in the presence of lincomycin (+Lin) or in the absence of lincomycin (-Lin) as described in Fig. 1. RNA was extracted and both *Lhcb1* and *RbcS* mRNA levels were quantified with RNA blot hybridizations that utilized 2.5  $\mu\text{g}$  of total RNA. Four biological replicates from each time point were quantified and normalized to total RNA stained with methylene blue. Numbers below each lane indicate the amount of hybridized RNA as a percentage of hybridized RNA in untreated seedlings after 24 h in 60  $\mu\text{mol m}^{-2} \text{s}^{-1}$  BR light.

**B,** Expression of *Lhcb1* and *RbcS* in seedlings not treated with lincomycin following a fluence-rate shift. Seedlings were grown in the absence of lincomycin (-Lin) and the levels of *Lhcb1* mRNA were quantified as in A.



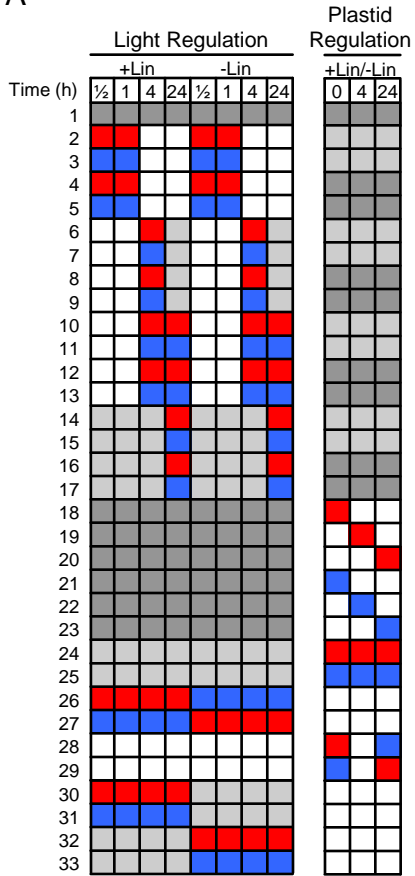
**Supplemental Figure S2.** Agglomerative clustering of early time points (i.e., 0.5 h and 1 h) and late time points (i.e., 4 h and 24 h).

A, Agglomerative hierarchical clustering of the 7104 active gene sets based on their regulation by light at 0.5 h and 1 h in lincomycin-treated and untreated seedlings. Up-regulated expression is indicated with red and down-regulated expression is indicated with blue. Color intensity is proportional to the degree of regulation.

B, Agglomerative hierarchical clustering of the 7104 active gene sets based on their regulation by the light at 4 h and 24 h in lincomycin-treated and untreated seedlings. Data are colored as in A.

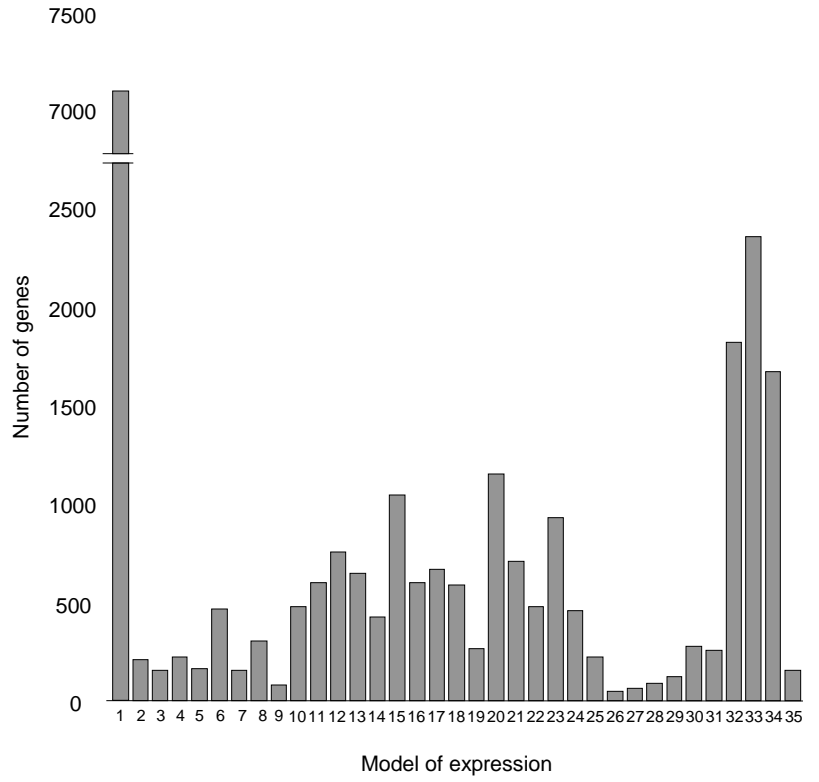
C-F, The average gene expression level ( $\log_2$ ) and number of genes represented by each of the clusters from Fig. 2E (C), Fig. 2F (D), Supplemental Fig. S2A (E), and Supplemental Fig. S2B (F).

A



■ Gene is induced  
■ Gene is repressed  
■ Gene is induced or repressed  
■ No significant change in gene expression  
■ Data was not considered

B

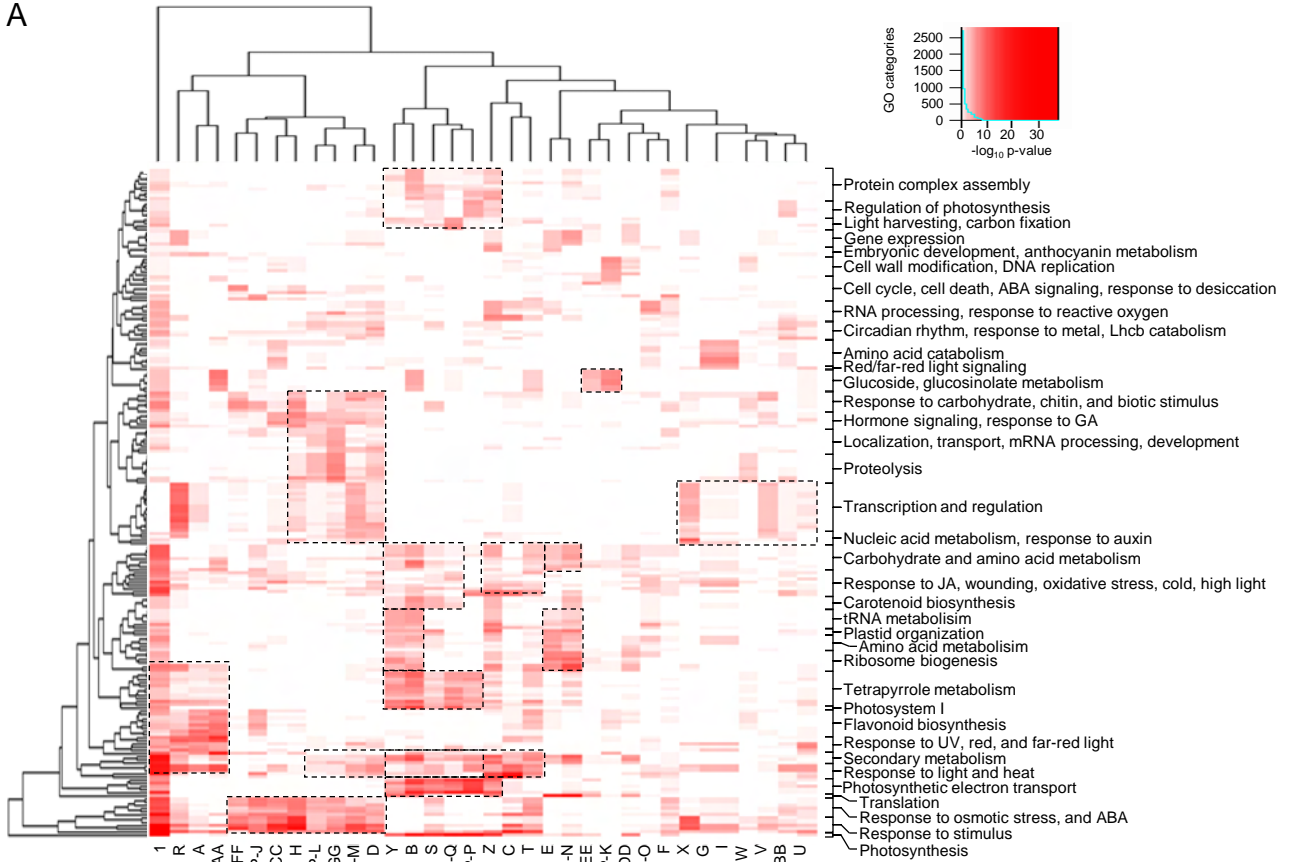


**Supplemental Figure S3.** Distribution of user-defined expression patterns among light- and plastid-regulated genes.

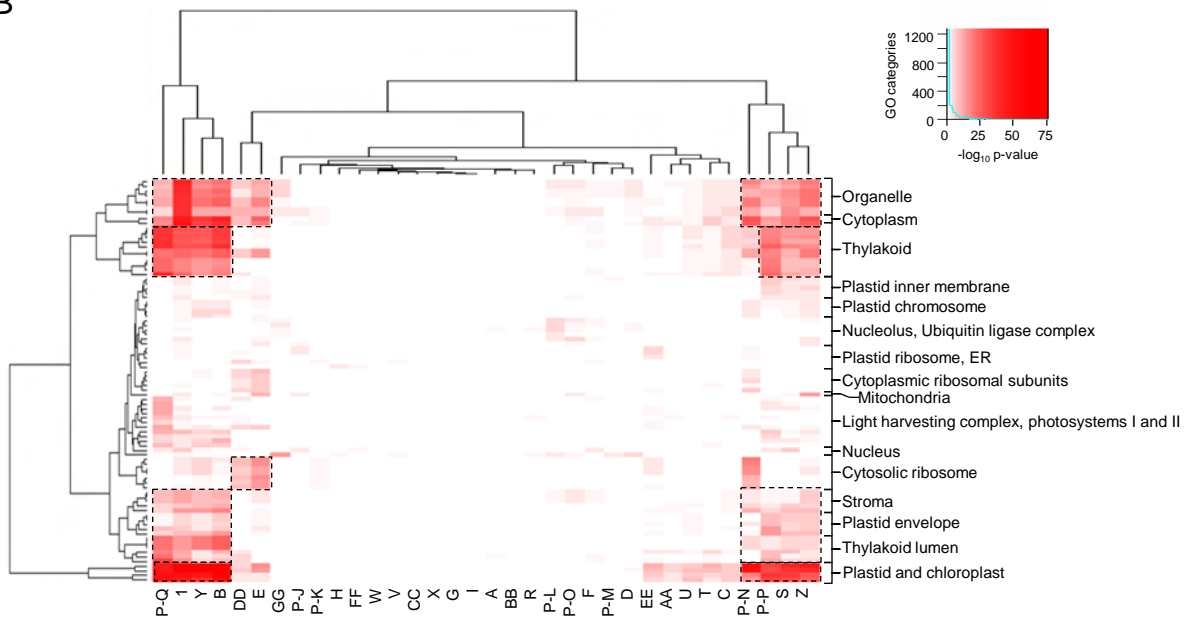
A, Expression patterns of light- and plastid-regulated genes. We defined 33 models of expression that arise from the interactions between light and plastid signaling. Gene expression was described in four possible ways: induced (red); repressed (blue); induced or repressed (dark gray); no significant change (light gray); data were not considered (white). Genes were considered induced (red) or repressed (blue) if the change in expression was greater than twofold ( $p \leq 0.01$ ). The expression patterns were: pattern 1, genes are induced or repressed; patterns 2-5, early-light-responsive genes (i.e., genes are regulated by light only at 0.5 h and 1 h); patterns 6-9, transiently responsive genes (i.e., genes that are regulated by light only at 4 h); patterns 10-13, light induction or repression at 4 h is maintained at 24 h in lincomycin-treated and/or untreated seedlings; patterns 14-17, late-responsive genes (i.e., genes that are regulated by light only at 24 h); patterns 18-25, plastid-responsive genes (i.e., genes that are regulated in lincomycin-treated seedlings at 0 h, 4 h, or at 24 h); patterns 26-27, genes that are oppositely regulated by light at the same time point in lincomycin-treated and untreated seedlings; patterns 28-29, genes that are oppositely regulated in lincomycin-treated and untreated seedlings 24 h after the BR-fluence-rate shift; patterns 30-31, genes that are either induced or repressed by light in lincomycin-treated but not in untreated seedlings; patterns 32-33 genes that are induced or repressed by light in untreated seedlings but not in lincomycin-treated seedlings.

B, Number of genes in each of the expression models described in A. The 33 expression patterns from A and two additional patterns, 34 and 35, are shown. Pattern 34 (positive correlation) describes genes that exhibit a similar response to the BR-fluence-rate shift regardless of whether seedlings are treated with lincomycin. For these genes, the correlation coefficient between the expression patterns in lincomycin-treated and untreated seedlings is greater than 0.95. Pattern 35 (negative correlation) describes genes that have an opposite response to the BR-fluence-rate shift in lincomycin-treated and untreated seedlings. For these genes, the correlation coefficient between the expression pattern in lincomycin-treated and untreated seedlings is less than -0.95.

A



B

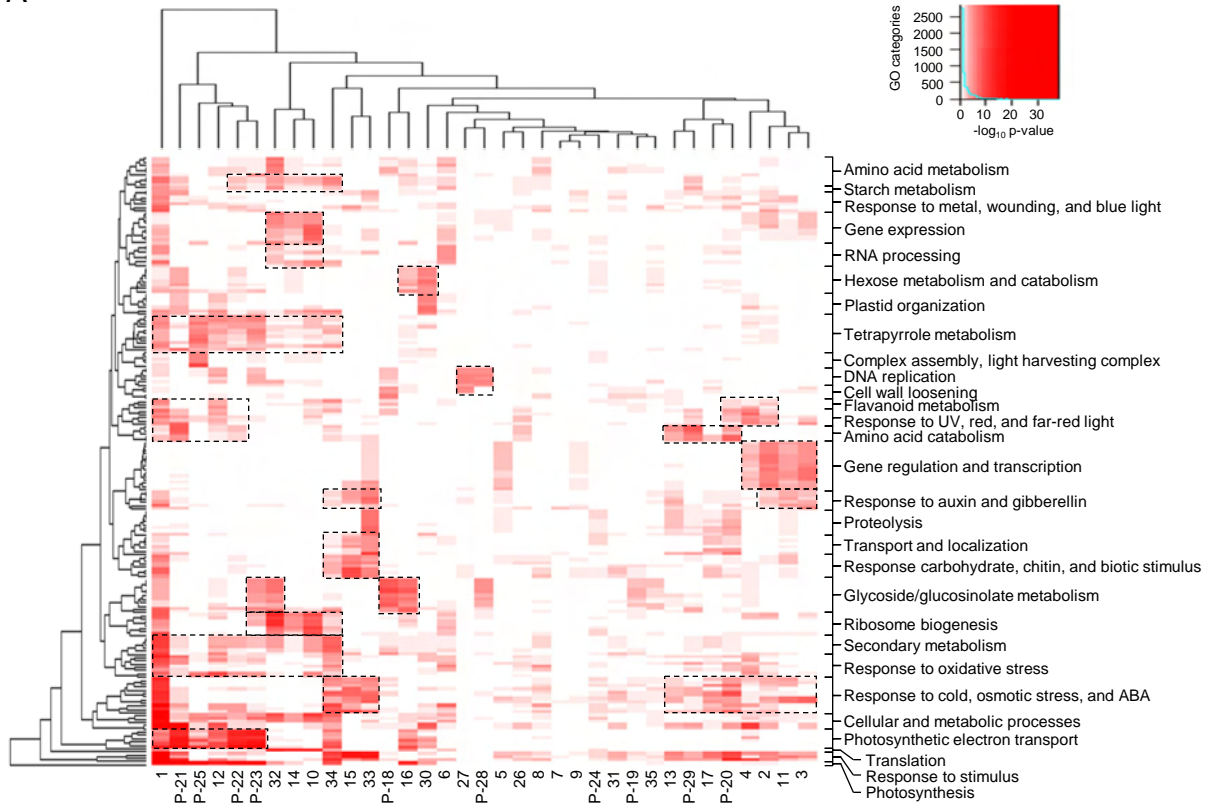


**Supplemental Figure S4.** Agglomerative hierarchical clustering of significantly enriched GO terms with expression clusters.

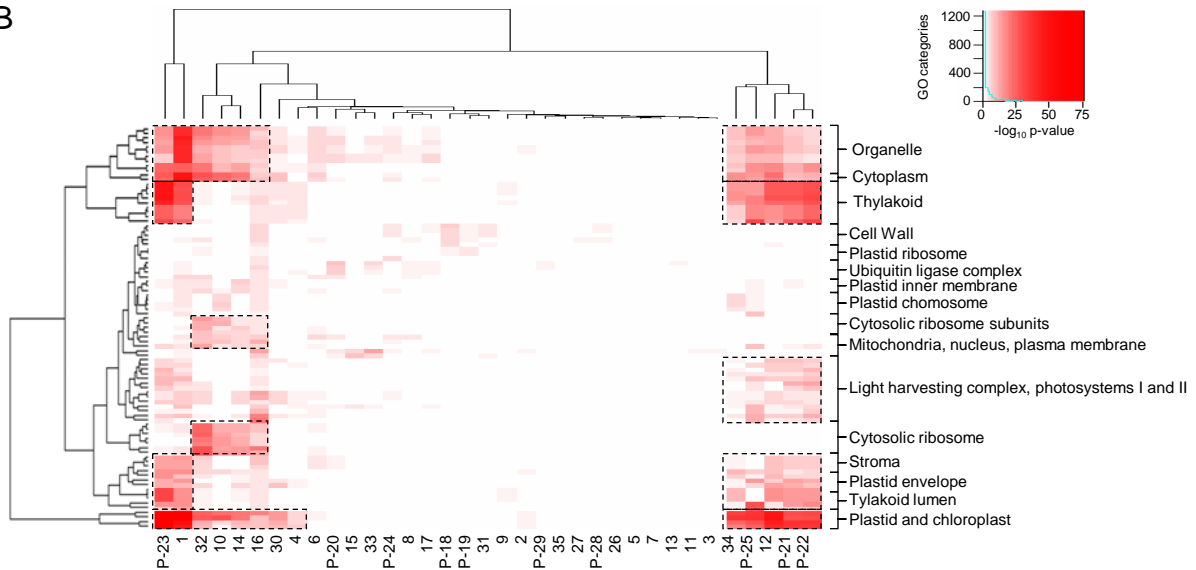
A, Agglomerative hierarchical clustering of GO terms defined as a biological process with expression clusters. We analyzed 205 GO terms and judged them to be enriched in at least one of the expression clusters if  $p \leq 0.01$  after a Westfall-Young correction. GO terms were included if they were enriched in at least one expression pattern and  $p \leq 0.01$ . The negative  $\log_{10}$  of the uncorrected p-value for each pattern was clustered. Clusters of GO terms are shown as a dendrogram at the left of the cluster matrix. Summary GO terms are shown at the right of the cluster matrix. These summary terms contain a representative group of genes from multiple GO categories. For example, tetrapyrrole metabolism is a summary term that contains a representative group of genes from the following GO categories: tetrapyrrole metabolic process, 0033013; porphyrin metabolic process, 0006778; chlorophyll metabolic process, 0015994; cofactor metabolic process, 0051186; tetrapyrrole biosynthetic process, 0033014; porphyrin biosynthetic process, 0006779; cofactor biosynthetic process, 0051188; chlorophyll biosynthetic process, 0015995. Expression clusters from Fig. 2 and Supplemental Fig. S2 are shown as a dendrogram at the top of the cluster matrix. Names of expression clusters from Fig. 2 and Supplemental Fig. S2 that correspond to each branch of the dendrogram are indicated at the bottom of the cluster matrix. The letter P next to the cluster letter indicates regulation by plastid dysfunction. No letter P next to the cluster letter indicates regulation by the BR-fluence-rate shift. The branch of the dendrogram labeled 1 contains all of the significantly regulated genes in the dataset. The intensity of the red color is proportional to the degree of enrichment. Co-regulated GO terms are indicated with dashed boxes.

B, Agglomerative hierarchical clustering of GO terms defined as cellular components with expression clusters. Eighty-eight GO terms defined as cellular components were enriched as described in A. The labeling of this heat map is as described for A.

A



B



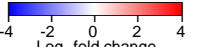


**Supplemental Figure S5.** Clustering of significantly enriched GO terms with user-defined expression patterns.

A, Agglomerative hierarchical clustering of GO terms defined as biological processes with expression patterns. One hundred ninety-eight active GO terms assigned as a biological process were considered enriched if  $p \leq 0.01$  after a Westfall-Young correction of at least one of the expression clusters. GO terms were included if they were enriched in at least one expression pattern and  $p \leq 0.01$ . The negative  $\log_{10}$  of the uncorrected p-values for each pattern was clustered. These one hundred ninety-eight GO clusters are shown as a dendrogram at the left of the cluster matrix. Summary terms that represent a cluster of GO terms are shown at the right of the cluster matrix and are as described in Supplemental Fig. S4A. User-defined expression patterns from Supplemental Fig. S3 are shown as a dendrogram at the top of the cluster matrix. User-defined expression pattern numbers from Supplemental Fig. S3 that correspond to each branch of the dendrogram are indicated at the bottom of the cluster matrix. The letter P next to the cluster number indicates regulation by the lincomycin treatment. No letter P next to the cluster letter indicates regulation by the BR-fluence-rate shift. The intensity of the red color is proportional to the degree of enrichment. Clusters of patterns are indicated with dashed boxes.

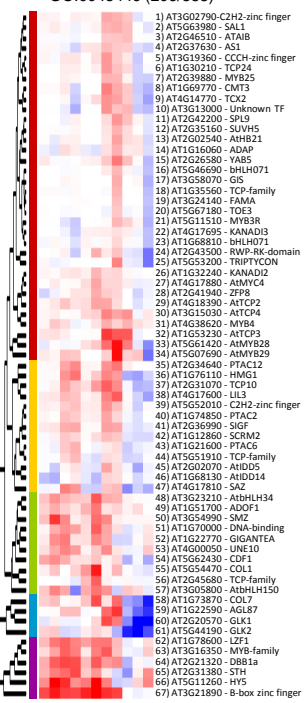
B, Agglomerative hierarchical clustering of GO terms defined as cellular components with expression patterns. Eighty-seven GO terms assigned as cellular components were enriched as described in A. Labeling of this heat map is as described in A.





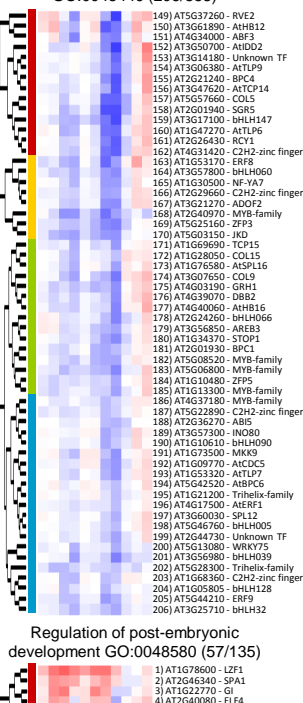
### Regulation of transcription cluster I

GO:0045449 (206/558)



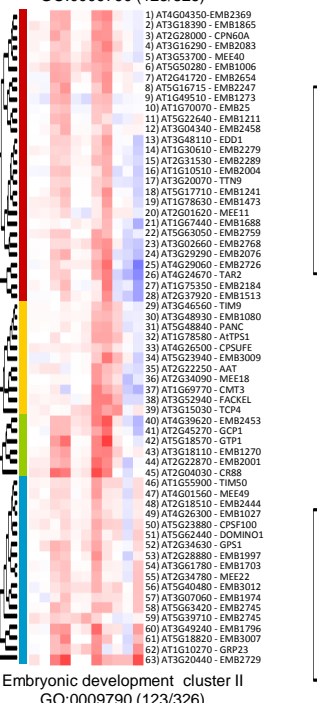
### Regulation of transcription cluster III

GO:0045449 (206/558)

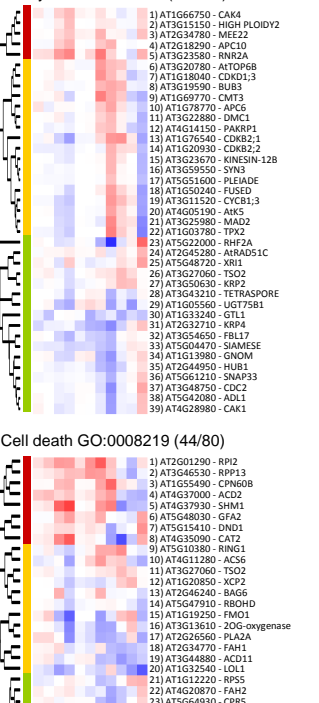


### Embryonic development cluster I

GO:0009790 (123/326)

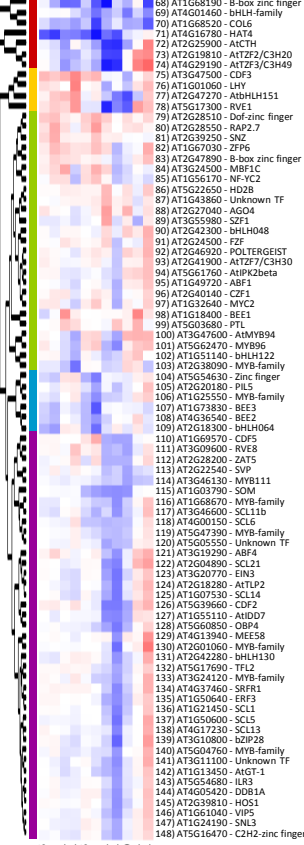


### Cell cycle GO:0007049 (39/123)



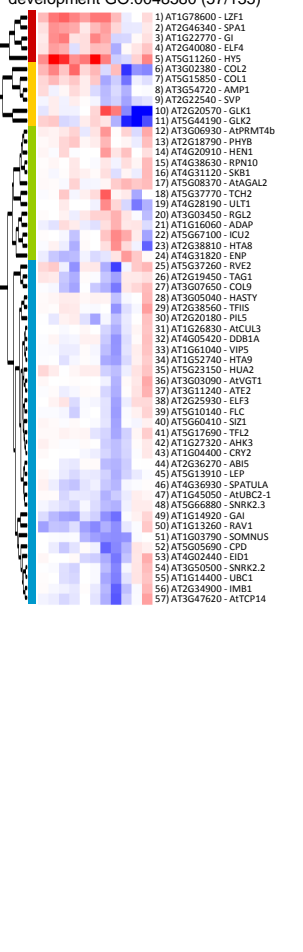
### Regulation of transcription cluster II

GO:0045449 (206/558)



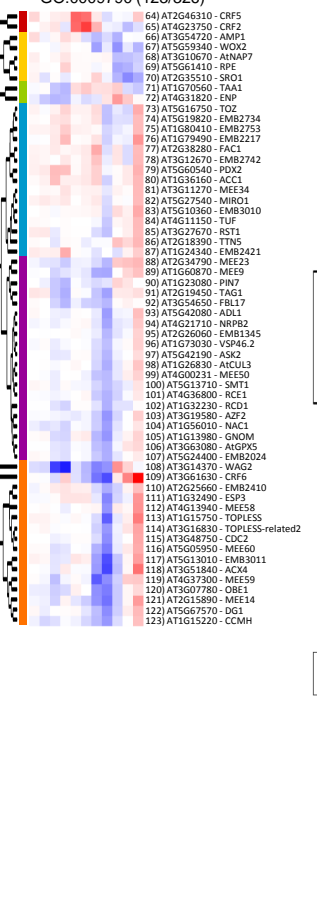
### Regulation of post-embryonic development

GO:0048530 (57/135)

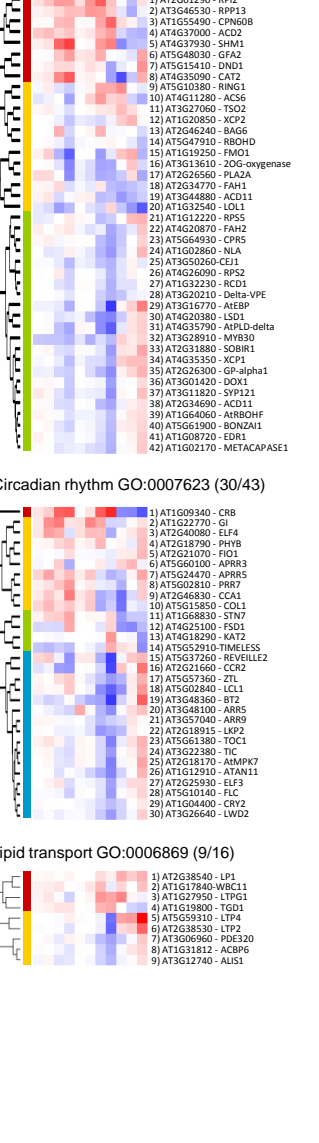


### Embryonic development cluster II

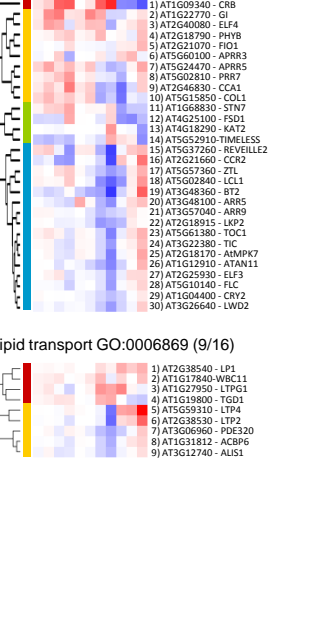
GO:0009790 (123/326)



### Cell death GO:0008219 (44/80)



### Circadian rhythm GO:0007623 (30/43)

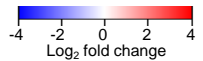


### Lipid transport GO:0006869 (9/16)



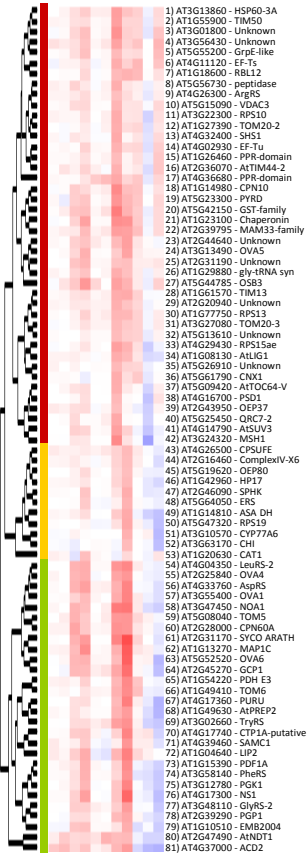
+Lin -Lin +Lin -Lin



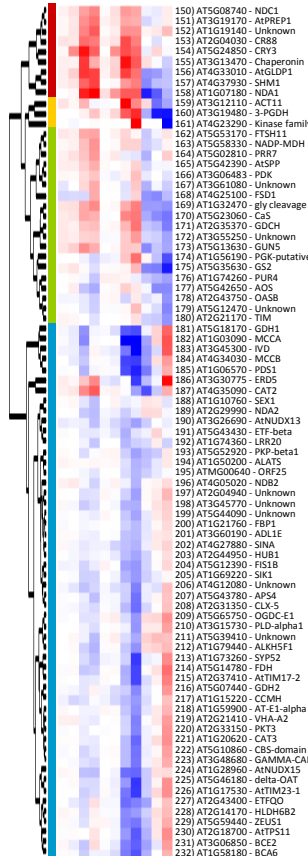


C

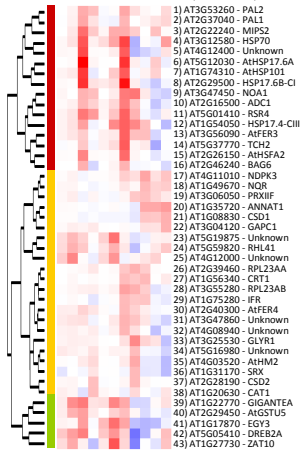
Mitochondrion GO:0005739 cluster I (232/551)



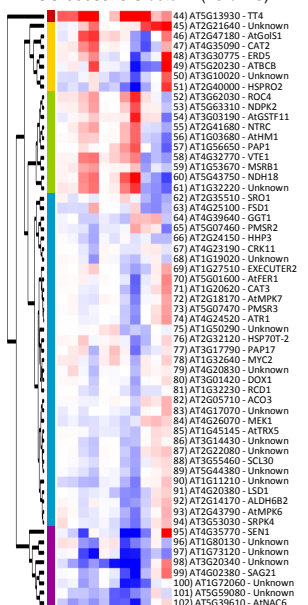
Mitochondrion GO:0005739 cluster III (232/551)



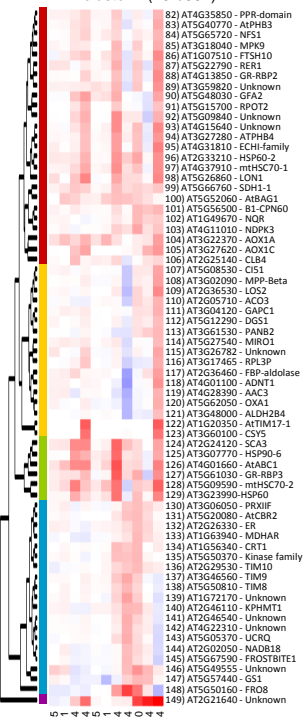
Response to oxidative stress GO:0006979 Cluster I (102/173)



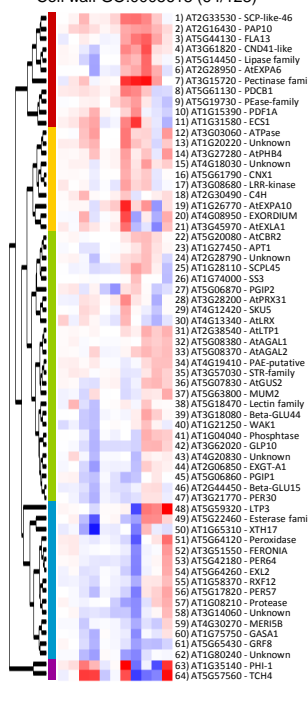
Response to oxidative stress GO:0006979 Cluster II (102/173)



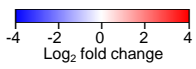
Mitochondrion GO:0005739 cluster II (232/551)



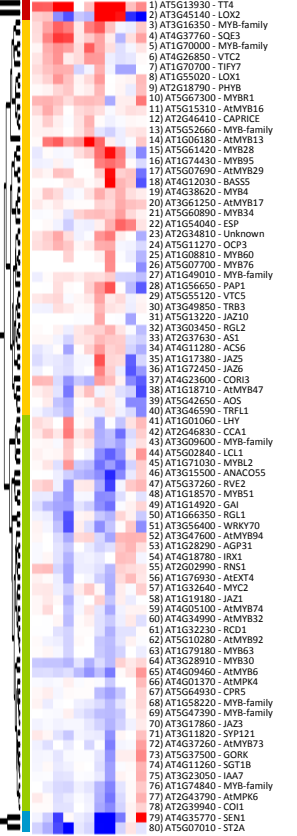
Cell wall GO:0005618 (64/123)



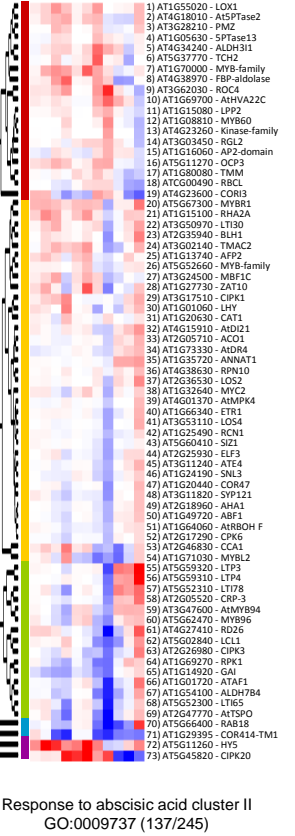
+Lin -Lin -Lin



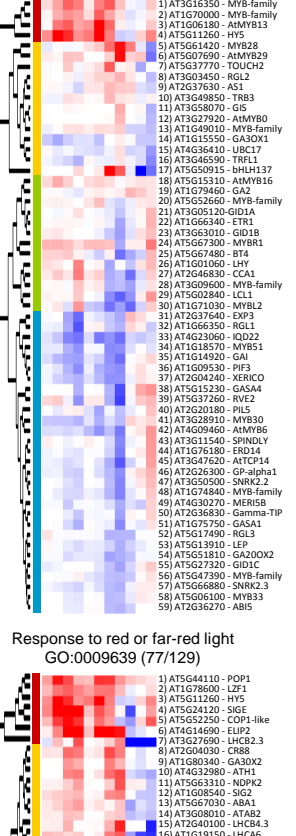
Response to jasmonic acid stimulus GO:0009753 (80/140)



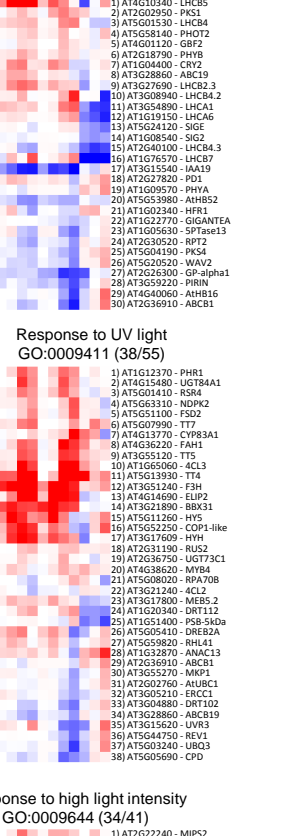
Response to abscisic acid cluster I GO:0009737 (137/245)



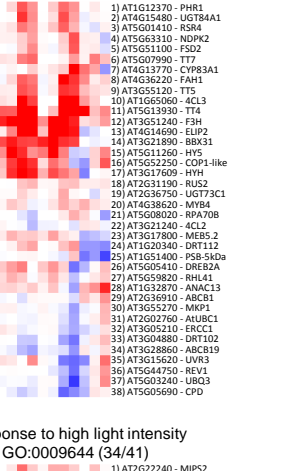
Response to gibberellin stimulus GO:0009739 (59/99)



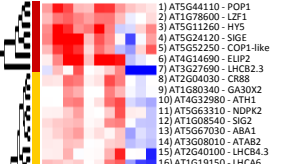
Response to blue light GO:0009637 (30/44)



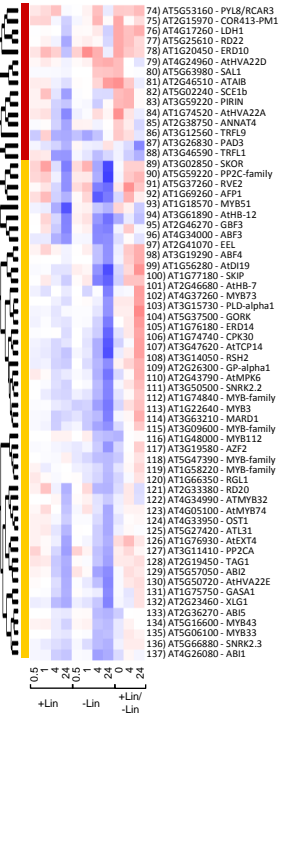
Response to UV light GO:0009411 (38/55)



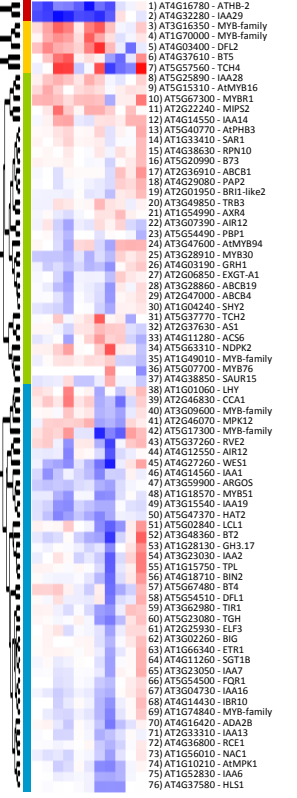
Response to red or far-red light GO:0009639 (77/129)



Response to abscisic acid cluster II GO:0009737 (137/245)



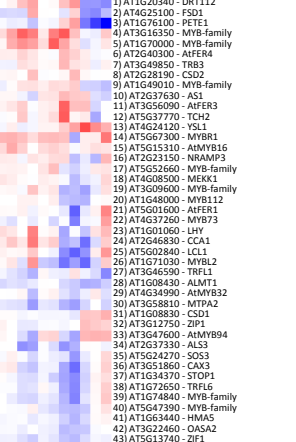
Response to auxin stimulus GO:0009733 (76/169)



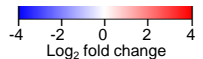
Response to high light intensity GO:0009644 (34/41)



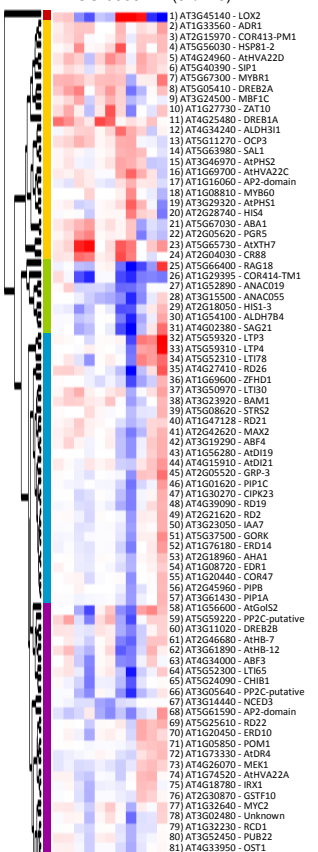
Response to metal ion GO:0010038 (44/70)



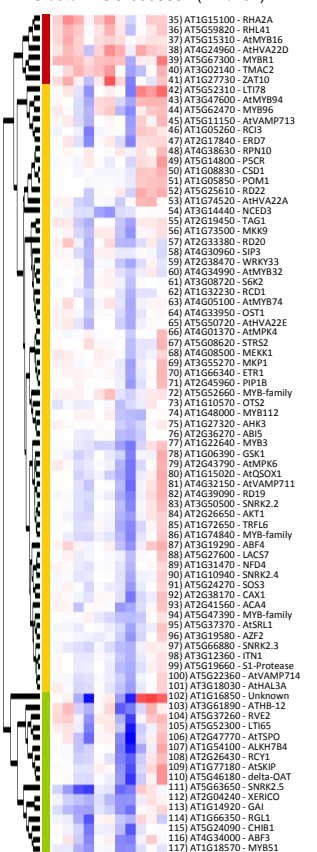
4 3 2 1 0 1 2 3 4  
+Lin -Lin -Lin



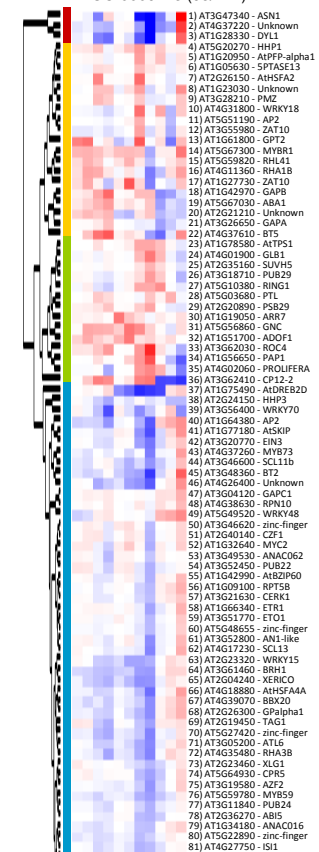
Response to water deprivation  
GO:0009414 (97/145)



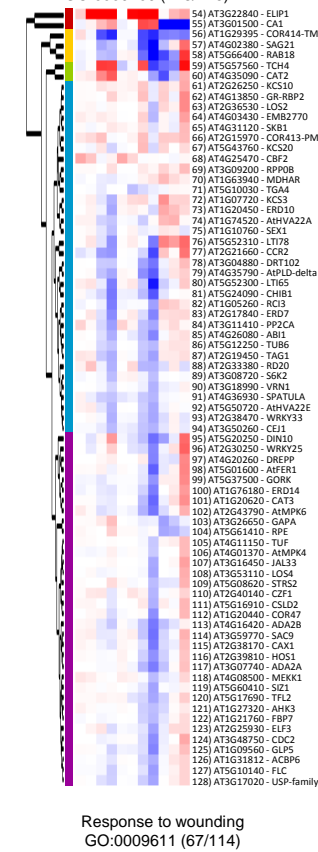
Response to salt stress  
Cluster II GO:0009651 (117/202)



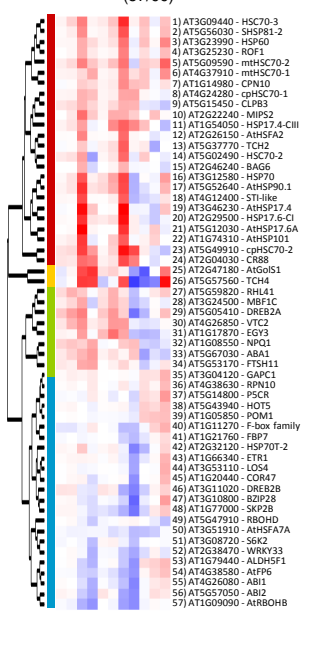
Response to carbohydrate stimulus  
GO:0009743 (93/174)



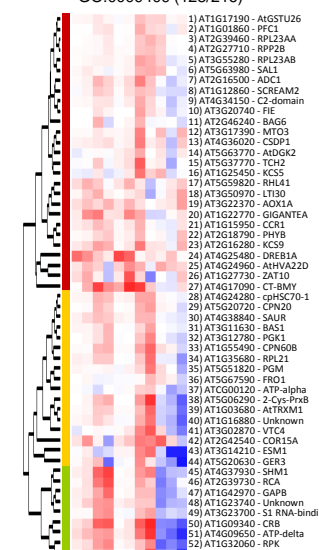
Response to cold cluster II  
GO:0009409 (128/216)



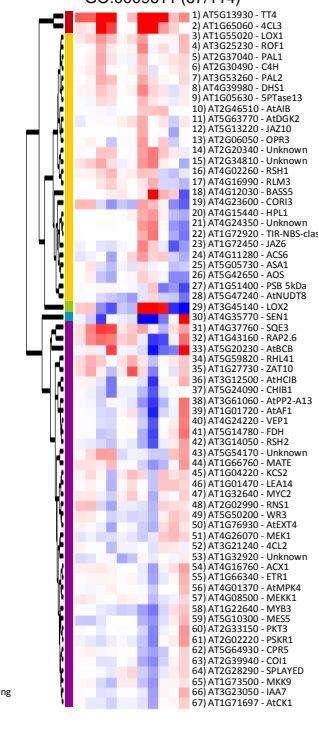
Response to heat GO:0009408 (57/99)



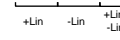
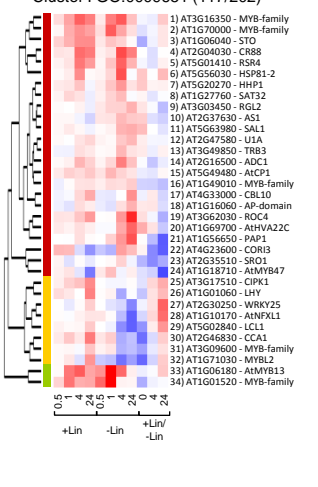
Response to cold cluster I  
GO:0009409 (128/216)



Response to wounding  
GO:0009611 (67/114)



Response to salt stress  
Cluster I GO:0009651 (117/202)







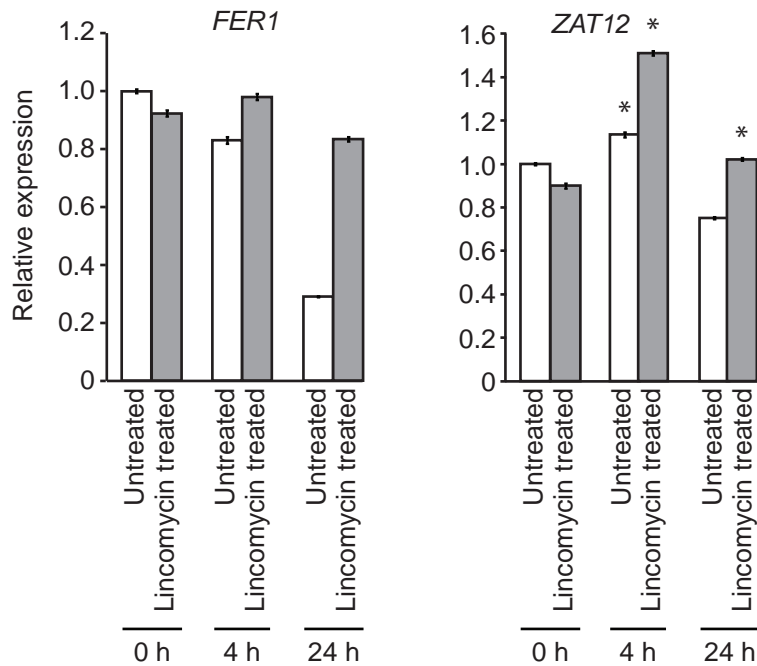
**Supplemental Figure S6.** Agglomerative hierarchical clustering of significantly regulated genes.

A, Agglomerative hierarchical clustering of significantly regulated genes annotated as contributing to 24 biological processes by their expression profiles. The criteria for significant regulation are described in Fig. 2. Enriched GO terms were identified using the criteria described in Supplemental Fig. S4A. The number of significantly regulated genes from a GO term/total number of genes in each GO term is indicated in parentheses next to each GO term number code. As indicated by the key at the bottom of the leftmost column, squares from the left to the right indicate light-regulated expression at 0.5 h, 1 h, 4 h, and 24 h following the BR-fluence-rate shift in lincomycin-treated seedlings; light-regulated expression at 0.5 h, 1 h, 4 h, and 24 h following the BR-fluence-rate shift in untreated seedlings; and plastid-regulated expression at 0 h, 4 h, and 24 following the fluence-rate shift. Up-regulated expression is indicated with red and down-regulated expression is indicated with blue. Color intensity is proportional to the degree of regulation.

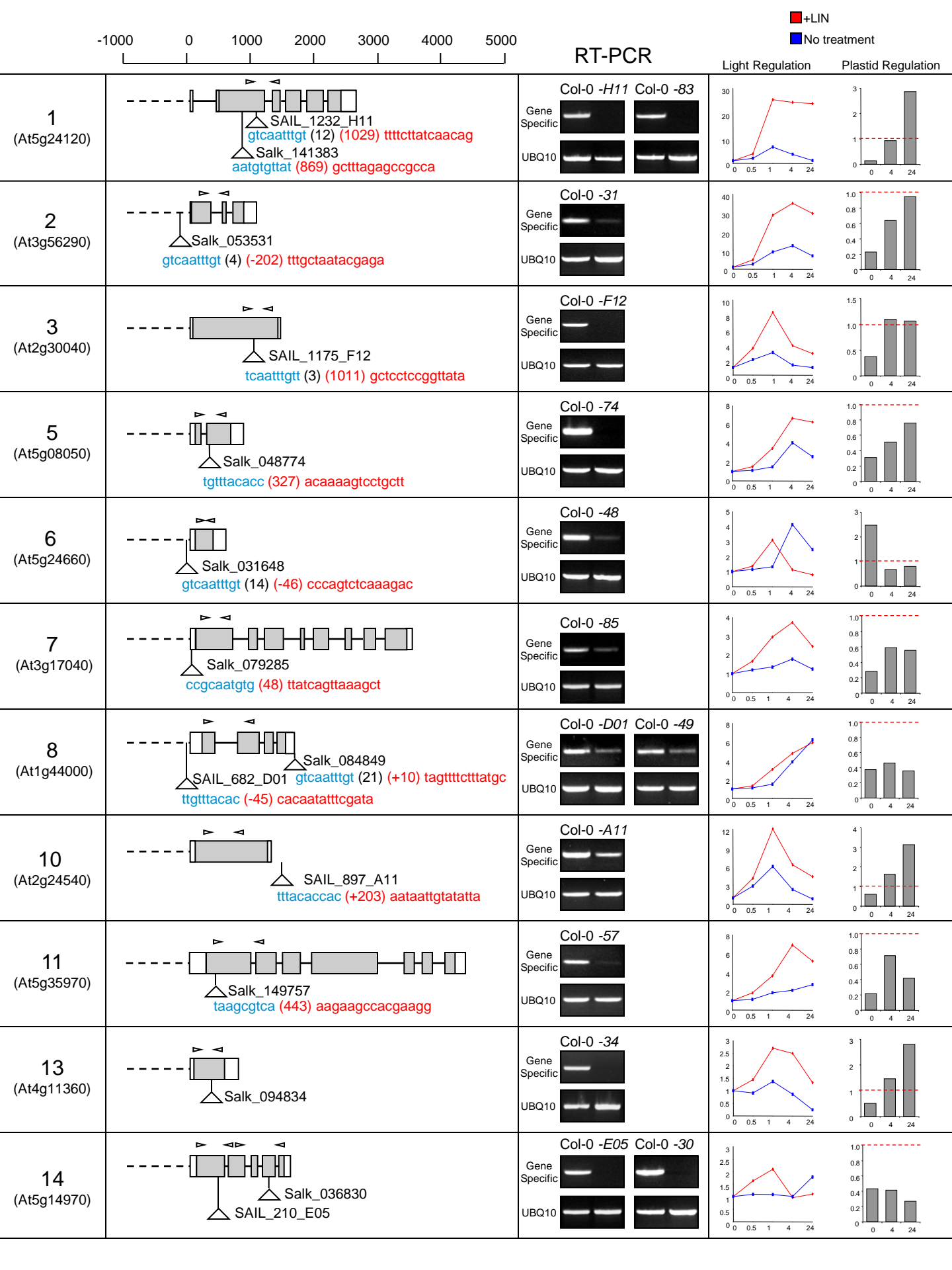
B, Agglomerative hierarchical clustering of significantly regulated genes annotated as contributing to the 14 cell components by their expression profiles. Genes and GO terms were clustered, arranged, and labeled as described in A.

C, Agglomerative hierarchical clustering of significantly regulated genes annotated as contributing to 16 biological responses to stimulus by their expression profiles. Genes and GO terms were clustered, arranged and labeled as described in A.

D, Agglomerative hierarchical clustering of significantly regulated ROS-responsive genes by their expression profiles. Genes whose expression is induced fivefold by ROS were previously described (Gadjev et al., 2006) and were obtained from the AtGenExpress repository. The confidence that each ROS-responsive gene list is enriched in our overall data set is indicated with p-values. Groups of ROS-responsive genes were clustered, arranged, and labeled as described in A.



**Supplemental Figure S7.** *FER1* and *ZAT12* expression in lincomycin-treated and untreated seedlings. The expression of *FER1* and *ZAT12* at 0 h, 4 h, and 24 h relative to the BR-fluence-rate shift was quantified with qRT-PCR. The expression of a particular gene in lincomycin-treated seedlings (gray bars) and untreated seedlings (white bars) is normalized to the expression of that same gene in untreated seedlings at 0 h. Error bars indicate standard deviation. \*, indicates a statistically significant increase relative to untreated seedlings at 0 h ( $P \leq 0.0001$  to 0.003).



-1000 0 1000 2000 3000 4000 5000

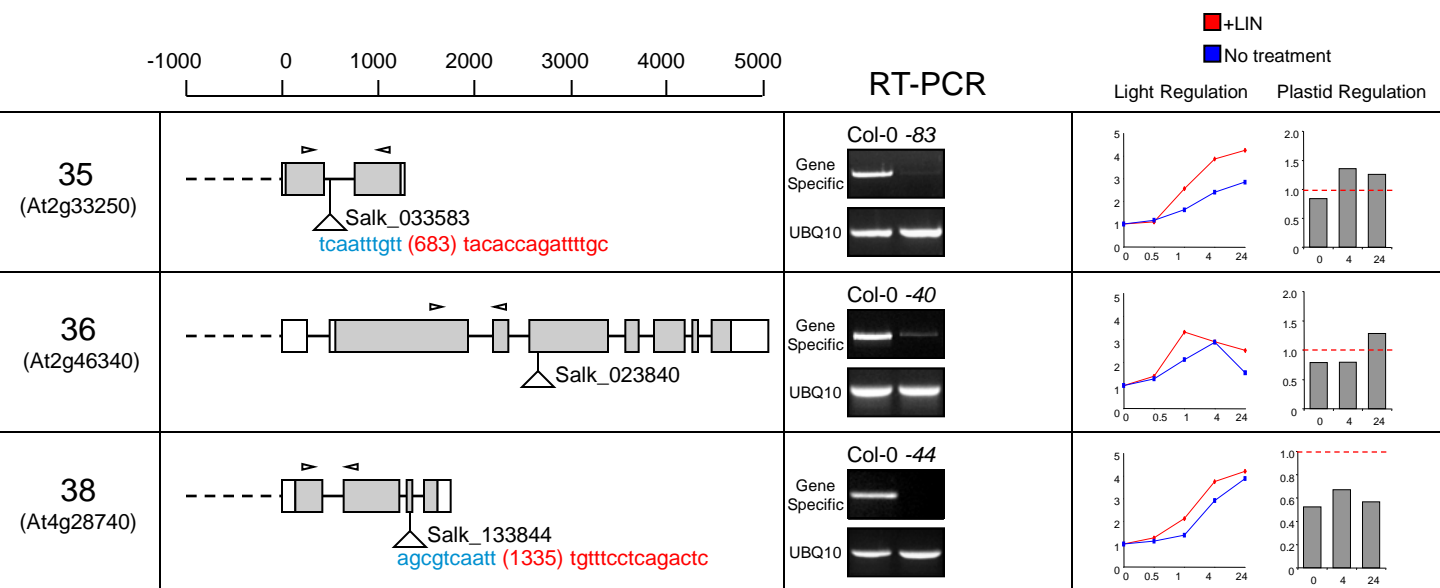
■ +LIN

■ No treatment

RT-PCR

Light Regulation

Plastid Regulation



**Supplemental Figure S8.** T-DNA insertion alleles of genes whose expression is more highly induced in lincomycin-treated than in untreated seedlings by 1 h following the BR-fluence-rate shift. The leftmost column contains the AGI code and an arbitrary number for each gene. The adjacent column contains the gene models based on information from The Institute for Genomic Research (TIGR). A scale in base pairs (bp) is indicated above this column. White boxes represent 5'- and 3'-untranslated regions (UTRs). Gray boxes represent exons. Dashed lines represent promoter sequences. Solid lines represent introns. The position of each T-DNA insertion is indicated to the left of the respective Salk or SAIL accession code. The sequence of the insertion site is indicated below each Salk or SAIL number and is color coded. The blue bases indicate the T-DNA sequence flanking the insertion site. Black numbers in parentheses indicate the number of bases that match neither the T-DNA nor the genomic sequence. The red bases indicate the genomic sequence flanking the T-DNA insertion. Red numbers in parentheses indicate the distance of the T-DNA insertion from the first base of the 5'-UTR sequence in bp. Negative red numbers indicate the number of bp upstream of the 5'-UTR sequence, and positive red numbers indicate the number of bp downstream of the 5'-UTR sequence. The third column from the left shows the RNA phenotype caused by each T-DNA insertion allele. RNA phenotypes were scored by extracting total RNA from 7-d-old seedlings and using RT-PCR to detect transcripts from genes that contain T-DNA insertions. The location of each gene-specific primer pair is represented as converging arrows above each gene model in the second column from the left. The last two numbers of Salk accession code or the last 3 digits of the SAIL accession code are used to name the T-DNA alleles. For example, the T-DNA alleles of gene number 1 (At5g24120) are SAIL\_1232\_H11 and Salk\_141383. In this figure, these alleles are named -H11 and -83, respectively. RT-PCR detection of *UBQ10* transcripts was used to test whether equal amounts of total RNA from both T-DNA mutants and wild type were used in each experiment. The fourth column from the left contains graphs that indicate the light- and plastid-regulated expression of each gene following the BR-fluence-rate shift. Expression levels are derived from microarray analysis. The leftmost graph indicates the expression level following the BR-fluence-rate shift. Relative transcript levels are indicated on the ordinate. Time in h following the BR-fluence-rate shift is indicated on the abscissa. Transcript levels in lincomycin-treated seedlings (+Lin) are indicated in red. Transcript levels in untreated seedlings (no treatment) are indicated in blue. The rightmost graph is a histogram that quantifies the plastid regulation of each gene. Plastid-regulated gene expression is defined as a ratio of expression in lincomycin-treated to expression in untreated seedlings at 0 h, 4 h, or 24 h following the fluence-rate shift. The dashed-red line indicates no effect of the lincomycin treatment on gene expression.

-1000 0 1000 2000 3000 4000 5000

RT-PCR

■ +LIN  
■ No treatment

Light Plastid Regulation

<p>47 (At5g43260)</p>	<p>Salk_072798 agcgtcaatg (247) tgttatcgatggtcc SALK_108550</p>	<p>Col-0 -98 Col-0 -50</p>	
<p>48 (At1g13670)</p>	<p>Salk_009943 gtcaagtgt (12) (-48) atcggtaactgaac</p>	<p>Col-0 -43</p>	
<p>51 (At1g22770)</p>	<p>gi-1</p>	<p>N/A</p>	
<p>52 (At5g24850)</p>	<p>Salk_122430</p>	<p>Col-0 -30</p>	
<p>53 (At2g46420)</p>	<p>Salk_029841 Salk_029850 agcgtcaatt (3176) tgttctattgatca (3317) ctctaagcgt (3317) caattaatctaagtc</p>	<p>Col-0 -50 Col-0 -41</p>	
<p>54 (At1g66330)</p>	<p>Salk_117745 gcgtcaattt (7) (367) taggtgttcccaat</p>	<p>Col-0 -45</p>	
<p>55 (At4g26150)</p>	<p>Salk_003995 ctgcaatttg (932) ttaccagtagaga</p>	<p>Col-0 -95</p>	
<p>57 (At3g02910)</p>	<p>Salk_145852</p>	<p>Col-0 -52</p>	
<p>58 (At1g51090)</p>	<p>SAIL_596_G09 SAIL_1150_A03 aattgtttac (196) (534) ggtttgggtggctca gtcaatttg (10) (-1053) tcaaggatcacgatg</p>	<p>Col-0 -G09 Col-0 -A03</p>	
<p>59 (At5g01820)</p>	<p>SALK_009699 Salk_149471 aatatactctg (9) (913) aaggagataaccgg tcaatttgtt (713) ttctcgtcgaggagg</p>	<p>Col-0 -71 Col-0 -99</p>	
<p>60 (At5g64170)</p>	<p>Salk_093925 Salk_071946 gcaatgtgtt (3929) attttgcttagccaa (4000) ttatgctgtgagat ccgcaatgtg (4000) ttatgctgtgagat</p>	<p>Col-0 -25 Col-0 -46</p>	

-1000 0 1000 2000 3000 4000 5000

RT-PCR

■ +LIN  
■ No treatment

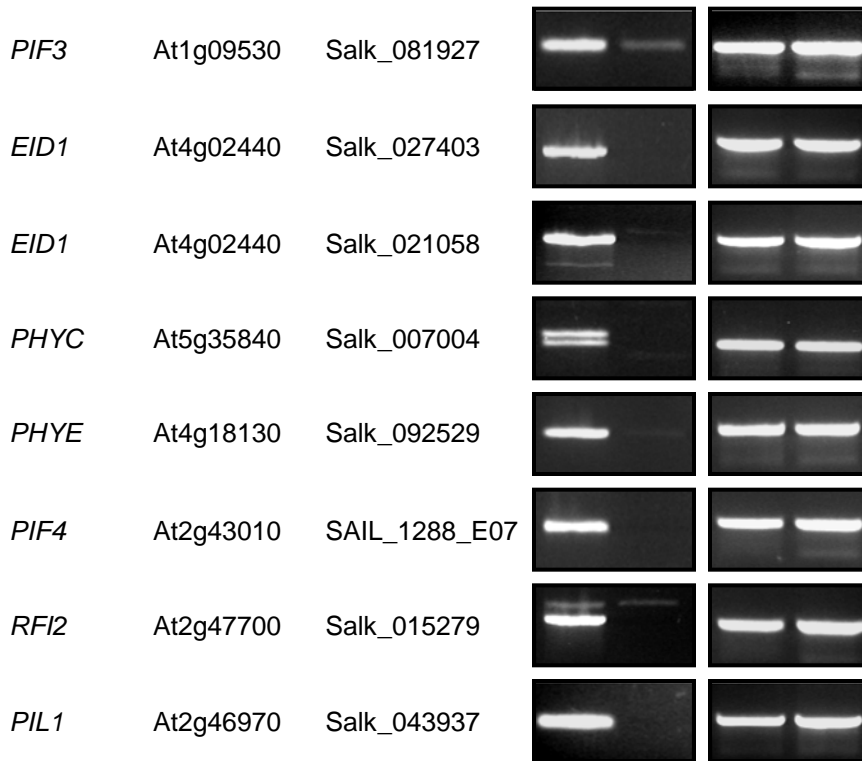
Light Plastid Regulation

<p>63 (At5g05410)</p>	<p>SAIL_365_F10 ttgtcacac (3) (1722) ggcccataaagcttc</p>	<p>Col-0 -F10</p> <p>Gene Specific UBQ10</p>	
<p>64 (At2g21320)</p>	<p>Salk_061956 gcgcaattt (5) (1436) tatctttacaggaa</p>	<p>Col-0 -56</p> <p>Gene Specific UBQ10</p>	
<p>65 (At1g78600)</p>	<p>SAIL_368_F08 aattgttta (126) (42) gtaggagtgtctaa</p>	<p>Col-0 -F08</p> <p>Gene Specific UBQ10</p>	
<p>66 (At5g67370)</p>	<p>Salk_023292 cgccaatttg (541) tccaagaacagttaa</p>	<p>Col-0 -92</p> <p>Gene Specific UBQ10</p>	
<p>67 (At5g59220)</p>	<p>Salk_142672 SAIL_520_H12 ccacaataata (111) (902) cttatgatcggagga</p>	<p>Col-0 -72 Col-0 -H12</p> <p>Gene Specific UBQ10</p>	
<p>68 (At2g28550)</p>	<p>Salk_069677 ttaagtgtc (1) (1699) gcaatcaacactaat</p>	<p>Col-0 -77</p> <p>Gene Specific UBQ10</p>	
<p>69 (At2g40080)</p>	<p>SAIL_424_B07 ttgtttacac (1) (362) agcttaatgaaccgg</p>	<p>Col-0 -B07</p> <p>Gene Specific UBQ10</p>	
<p>70 (At5g23730)</p>	<p>Salk_015765 ttaagtgtc (78) (-84) ccctcagtaaataat</p>	<p>Col-0 -65</p> <p>Gene Specific UBQ10</p>	
<p>71 (At4g28290)</p>	<p>Salk_086732 ttacaccac (53) aataaaataaaatct</p>	<p>Col-0 -32</p> <p>Gene Specific UBQ10</p>	
<p>72 (At5g58380)</p>	<p>Salk_111320 ttacaccac (1356) caatctgatggagat</p>	<p>Col-0 -20</p> <p>Gene Specific UBQ10</p>	
<p>74 (At3g17800)</p>	<p>Salk_073038 ccacaatata (1685) tcttgatcttatgt</p>	<p>Col-0 -38</p> <p>Gene Specific UBQ10</p>	

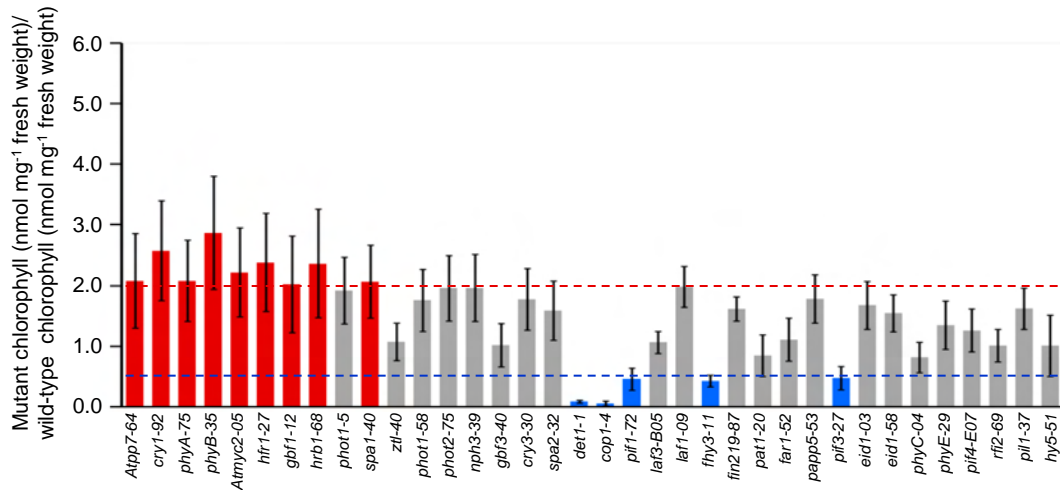
**Supplemental Figure S9.** T-DNA insertion alleles of genes whose expression is similarly induced in lincomycin-treated and untreated seedlings by 1 h following the BR-fluence-rate shift. The leftmost column contains the AGI code and an arbitrary number for each gene. The adjacent column contains the gene models based on information from The Institute for Genomic Research (TIGR). The gene models are labeled as described in Supplemental Fig. S8. *gi-1* (a.k.a., mutant 51) is not a T-DNA insertion allele. *gi-1* has a 5-bp deletion that introduces a premature stop codon (indicated with an arrow) and deletes 171 amino acid residues from the carboxy-terminus of the GIGANTEA protein (Park et al., 1999). The third column from the left shows the RNA phenotype caused by each T-DNA insertion allele. RNA phenotypes were scored using RT-PCR as described in Supplemental Fig. S8. The fourth column from the left contains graphs that indicate the light- and plastid-regulated expression of each gene following the BR-fluence-rate shift. The graphs are labeled as described in Supplemental Fig. S8.



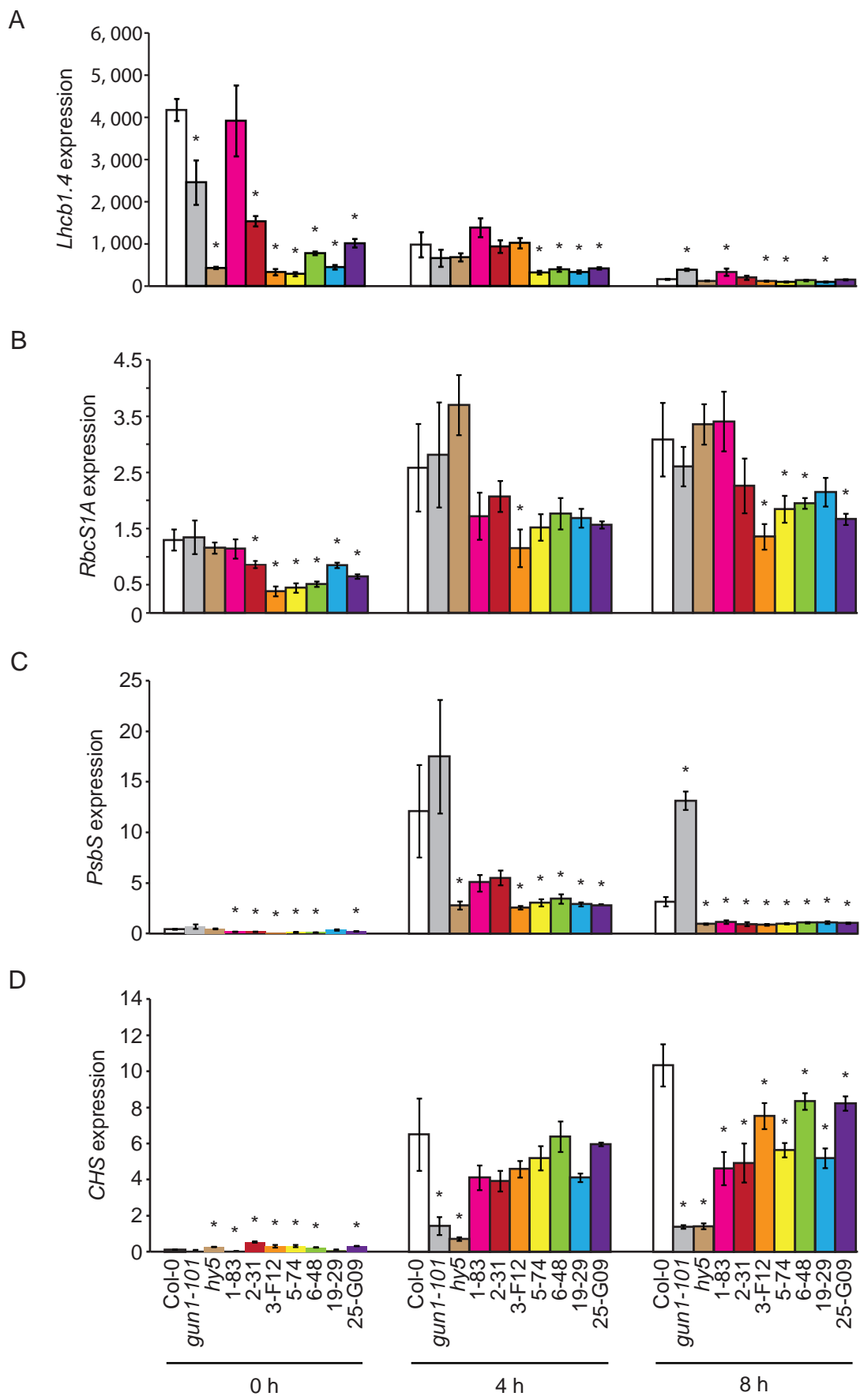
Gene name	AGI code	T-DNA allele	Gene Specific		<i>Ubg10</i>	
			T-DNA		T-DNA	
			Col-0	line	Col-0	line
<i>AtPP7</i>	At5g63870	Salk_089764				
<i>AtMYC2</i>	At1g32640	Salk_017005				
<i>HFR1</i>	At1g02340	Salk_037727				
<i>GBF1</i>	At4g36730	Salk_096612				
<i>HRB1</i>	At5g49230	Salk_087868				
<i>SPA1</i>	At2g46340	Salk_023840				
<i>ZTL</i>	At5g57360	Salk_012440				
<i>GBF3</i>	At2g46270	Salk_082840				
<i>SPA2</i>	At4g11110	Salk_009832				
<i>PIF1</i>	At2g20180	Salk_131872				
<i>LAF3</i>	At3g55850	SAIL_573_B05				
<i>LAF1</i>	At4g25560	Salk_079609				
<i>FHY3</i>	At3g22170	Salk_002711				
<i>FIN219</i>	At2g46370	Salk_075487				
<i>PAT1</i>	At5g48150	Salk_064220				
<i>FAR1</i>	At4g15090	Salk_031652				
<i>PAPP5</i>	At2g42810	Salk_021153				



**Supplemental Figure S10.** Characterization of T-DNA insertion alleles of genes that encode light-signaling factors. RNA phenotypes were scored using RT-PCR to detect transcripts from genes that contain T-DNA insertions. RT-PCR detection of transcripts from *UBQ10* was used to test whether equal amounts of total RNA from both T-DNA mutants and wild type were used in each experiment. These genes were at least previously implicated in light signaling: *AtPP7* (Møller et al., 2003); *AtMYC2/ZBF1* (Yadav et al., 2005); *HFR1* (Fairchild et al., 2000); *GBF1* (Schindler et al., 1992); *HRB1* (Kang et al., 2005); *SPA1* (Hoecker et al., 1998); *ZTL* (Kim et al., 2007); *GBF3* (Schindler et al., 1992); *SPA2* (Laubinger et al., 2004); *PIF1* (Huq et al., 2004); *LAF3* (Ballesteros et al., 2001); *LAF1* (Møller et al., 2001); *FHY3* (Wang and Deng, 2002); *FIN219* (Hsieh et al., 2000); *PAT1* (Bolle et al., 2000); *FAR1* (Hudson et al., 1999); *PAPP5* (Ryu et al., 2005); *PIF3* (Ni et al., 1998); *EID1* (Dieterle et al., 2001); *PHYC* (Monte et al., 2003); *PHYE* (Devlin et al., 1998); *PIF4* (Huq and Quail, 2002); *RFI2* (Chen and Ni, 2006); *PIL1* (Salter et al., 2003).



**Supplemental Figure S11.** Chlorophyll phenotypes of light signaling mutants. De-etiolation was performed and relative chlorophyll levels were quantified for the indicated light-signaling mutants as described in Fig. 7. These alleles were previously generated using either EMS or T-DNA insertional mutagenesis. Names for T-DNA insertion alleles were created from the gene name and the last two numbers of the Salk accession code or the last three digits of the SAIL accession code. For example, the T-DNA insertion allele of *AtPP7* that is derived from Salk\_089764 is named *Atpp7-64*. *cry1-92*, *phyA-75*, *phyB-35*, *phot1-58*, *phot2-75*, *nph3-39*, *cry3-30*, and *hy5-51* were previously characterized (Ruckle et al., 2007). The remaining alleles derived from T-DNA insertional mutagenesis are described in Supplemental Fig. S10. The alleles derived from EMS mutagenesis were *cop1-4* (Deng et al., 1992; Deng and Quail, 1992), *det1-1* (Chory et al., 1989), and *phot1-5*, which was formerly known as *nph1-5* (Liscum and Briggs, 1995; Huala et al., 1997). Error bars represent 95% confidence intervals.



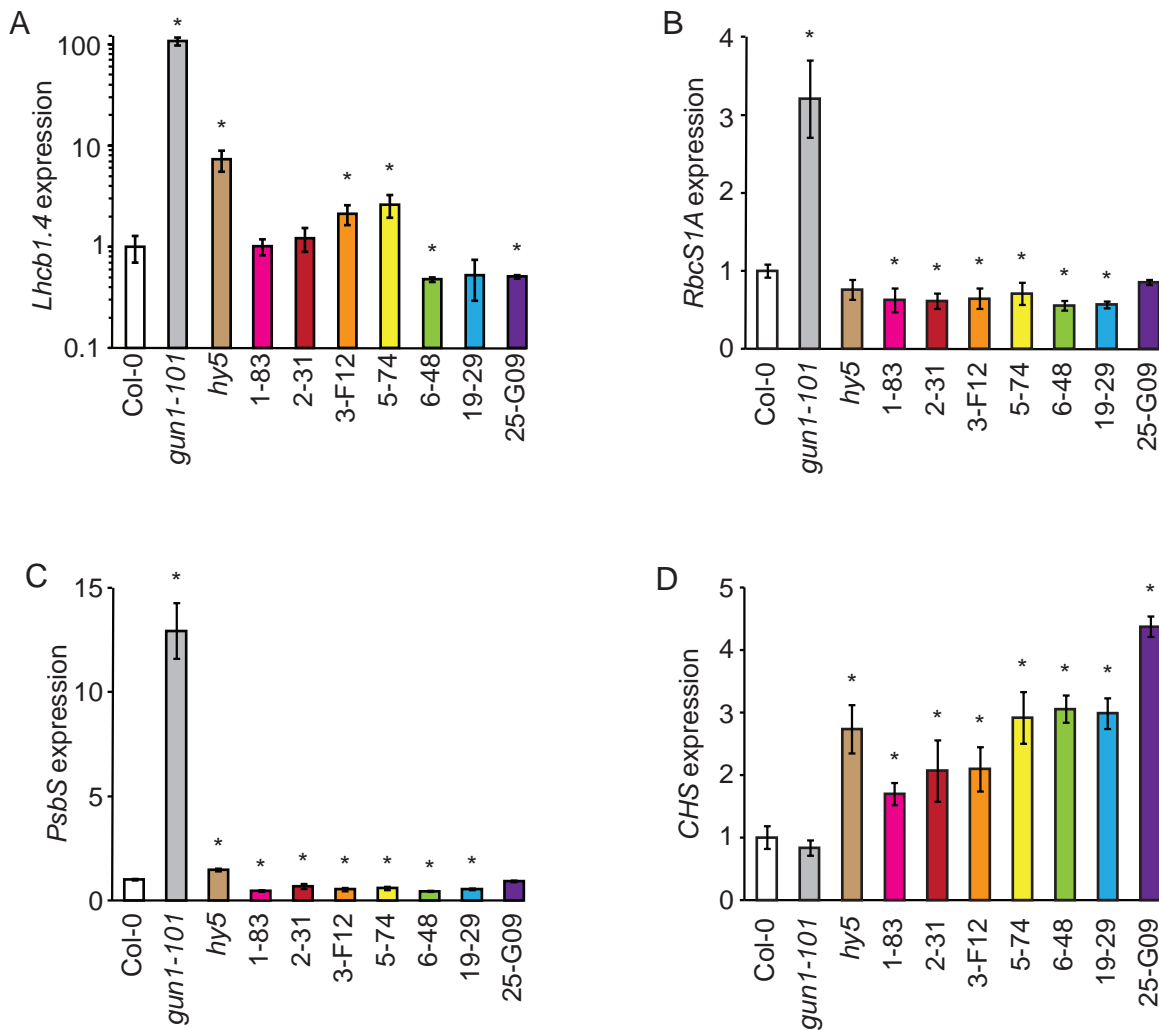
**Supplemental Figure S12.** *Lhcb1.4*, *RbcS1A*, *PsbS* and *CHS* expression in lincomycin-treated *end* mutants after an increase in fluence rate.

A, *Lhcb1.4* expression in particular *end* mutants after an increase in fluence rate. Wild type (Col-0) and the indicated mutants were grown on media that contained 0.5 mM lincomycin for 6 d in  $0.5 \mu\text{mol m}^{-2} \text{s}^{-1}$  BR light, and then transferred to  $60 \mu\text{mol m}^{-2} \text{s}^{-1}$  BR light. Seedlings were collected immediately before the fluence-rate shift (0 h), and at 4 h and 8 h following the fluence rate shift. Transcript levels were quantified by qRT-PCR. The order of the lines from left to right: wild type (Col-0, white bars), *gun1-101* (gray bars), *hy5* (brown bars), 1-83 (pink bars), 2-31 (red bars), 3-F12 (orange bars), 5-74 (yellow bars), 6-48 (green bars), 19-29 (blue bars), and 25-G09 (purple bars). Three biological replicates were analyzed for wild type (Col-0) and each mutant in each condition. Error bars indicate standard deviation. \* indicates a statistically significant increase ( $P=0.0002$  and  $0.02$ ) or statistically significant decrease ( $P<0.0001$  to  $0.03$ ) in the expression of *Lhcb1.4* in a particular mutant relative to wild type.

B, *RbcS1A* expression in particular *end* mutants after an increase in fluence rate. Analysis of *RbcS1A* expression was as described for *Lhcb1.4* expression in A. \* indicates a statistically significant decrease ( $P=0.002$  to  $0.04$ ) in the expression of *RbcS1A* in a particular mutant relative to wild type.

C, *PsbS* expression in particular *end* mutants after an increase in fluence rate. Analysis of *PsbS* expression was as described for *Lhcb1.4* expression in A. \* indicates a statistically significant increase ( $P<0.0001$ ) or a statistically significant decrease ( $P=0.001$  to  $0.03$ ) in the expression of *PsbS* in a particular mutant relative to wild type.

D, *CHS* expression in particular *end* mutants after an increase in fluence rate. Analysis of *CHS* expression was as described for *Lhcb1.4* expression in A. \* indicates a statistically significant increase ( $P=0.0001$  to  $0.007$ ) or a statistically significant decrease ( $P=0.0002$  to  $0.01$ ) in the expression of *CHS* in a particular mutant relative to wild type.



**Supplemental Figure S13.** *Lhcb1.4*, *RbcS1A*, *PsbS* and *CHS* expression in particular lincomycin-treated *end* mutants in continuous 60  $\mu\text{mol m}^{-2} \text{s}^{-1}$  BR light.

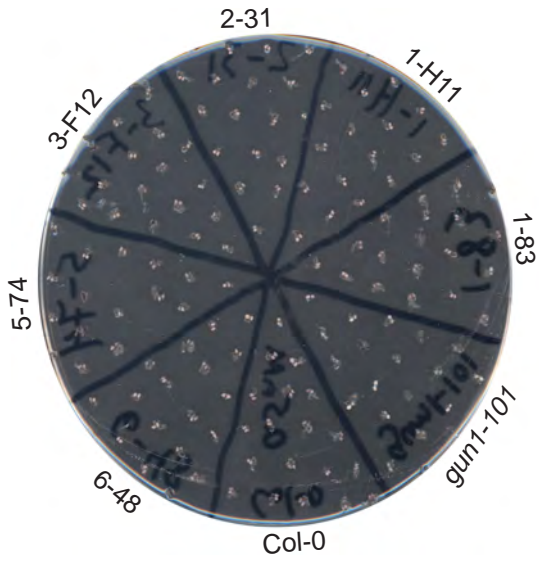
A, *Lhcb1.4* expression in particular *end* mutants grown in continuous 60  $\mu\text{mol m}^{-2} \text{s}^{-1}$  BR light. Wild type (Col-0) and the indicated mutants were grown on media that contained 0.5 mM lincomycin for 6 d in continuous 60  $\mu\text{mol m}^{-2} \text{s}^{-1}$  BR light. Transcript levels were quantified by qRT-PCR. Three biological replicates were analyzed for wild type (Col-0) and each mutant. Error bars indicate standard deviation. \* indicates a statistically significant increase ( $P < 0.0001$  to 0.003) or a statistically significant decrease ( $P = 0.04$ ) in the expression of *Lhcb1.4* in a particular mutant relative to wild type.

B, *RbcS1A* expression in particular *end* mutants. Analysis of *RbcS1A* expression was as described in A. \* indicates a statistically significant increase ( $P = 0.002$ ) or statistically significant decrease (0.001 to 0.04) in the expression of *RbcS1A* in a particular mutant relative to wild type.

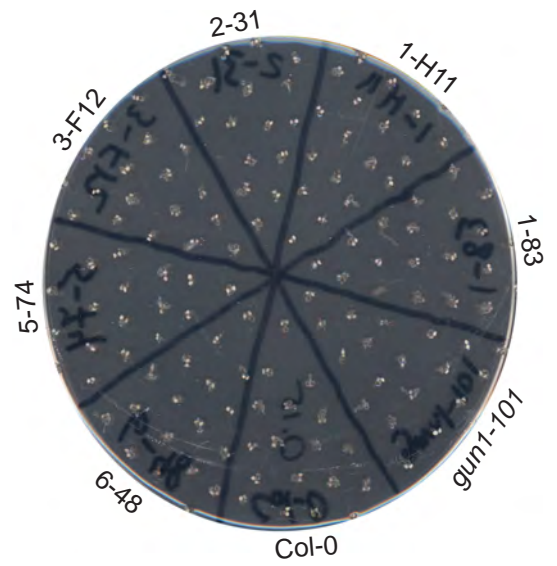
C, *PsbS* expression in particular *end* mutants. Analysis of *PsbS* expression was as described in A. \* indicates a statistically significant increase ( $P = 0.0001$  and 0.0004) or a statistically significant decrease ( $P < 0.0001$  to 0.01) in the expression of *PsbS* in a particular mutant relative to wild type.

D, *CHS* expression in indicated *end* mutants. Analysis of *CHS* expression was as described in A. \* indicates a statistically significant increase in the expression of *CHS* in a particular mutant relative to wild type ( $P < 0.0001$  to 0.02).

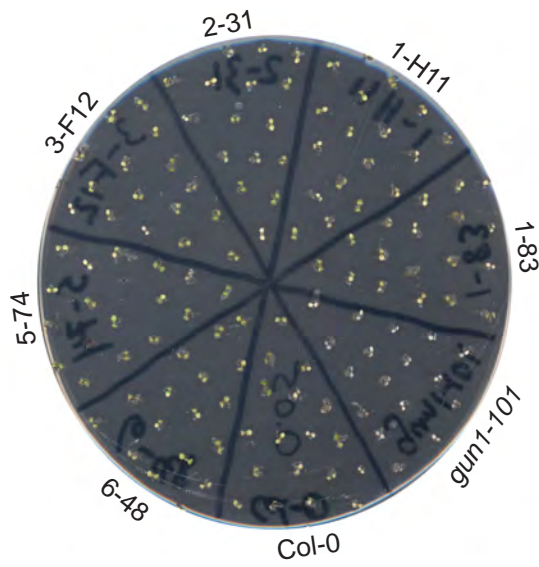
0.5 mM lincomycin



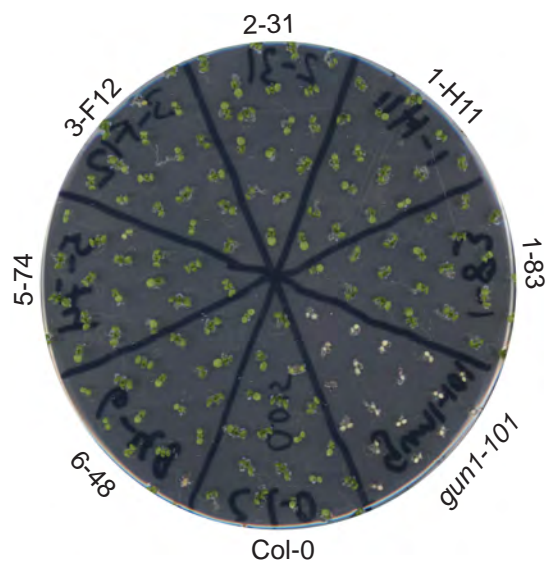
0.15 mM lincomycin



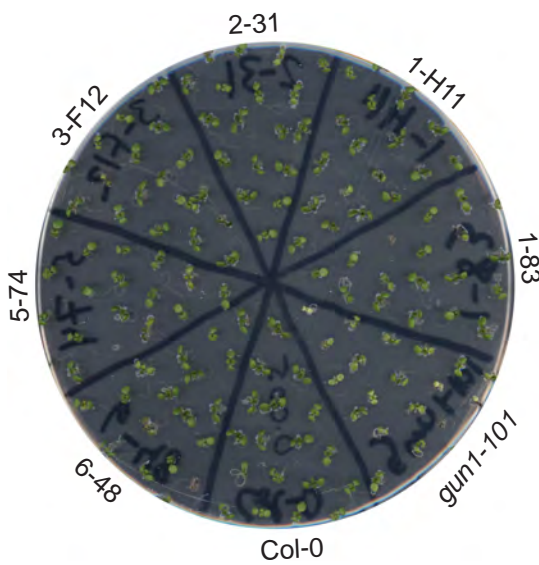
0.05 mM lincomycin



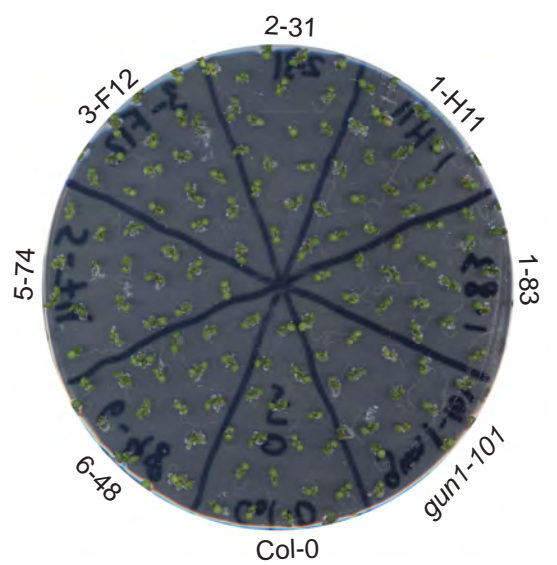
0.015 mM lincomycin



0.005 mM lincomycin



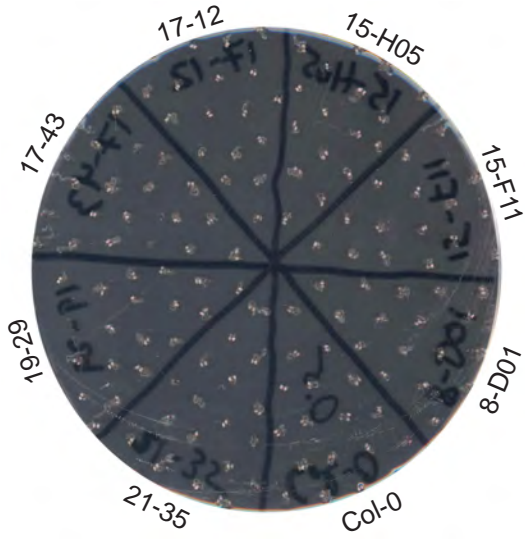
No lincomycin



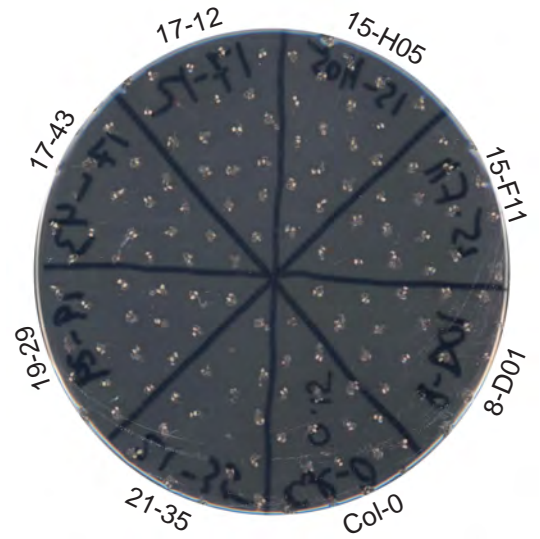
**Supplemental Figure S14.** *gun1-101*, 1-83, 1-H11, 2-31, 3-F12, 5-74, and 6-48 treated with various concentrations of lincomycin. *gun1-101* and the indicated *end* mutants were grown for 8 d in continuous white light at a fluence rate of  $100 \mu\text{mol m}^{-2} \text{s}^{-1}$  on media containing the indicated concentrations of lincomycin.



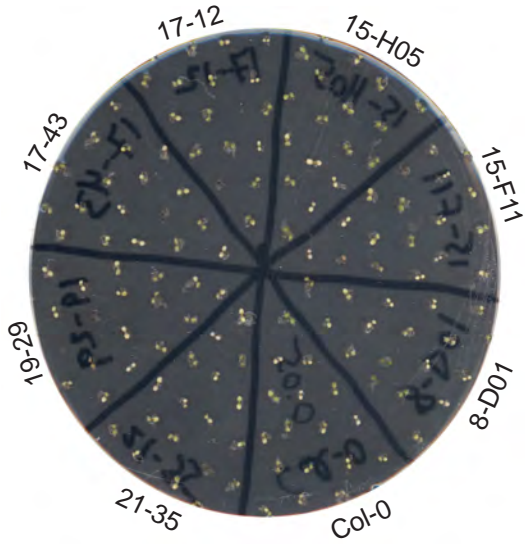
0.5 mM lincomycin



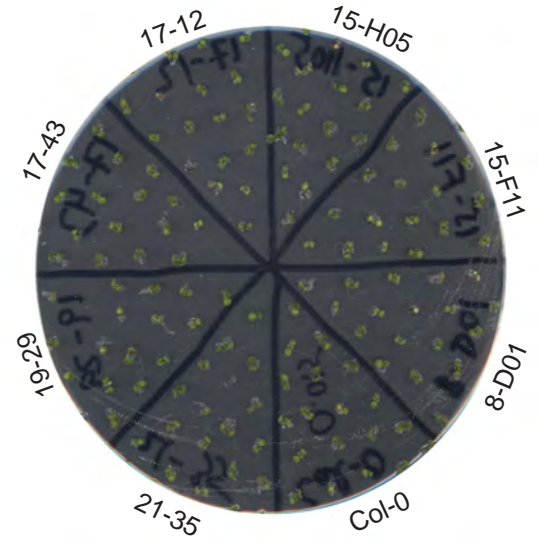
0.15 mM lincomycin



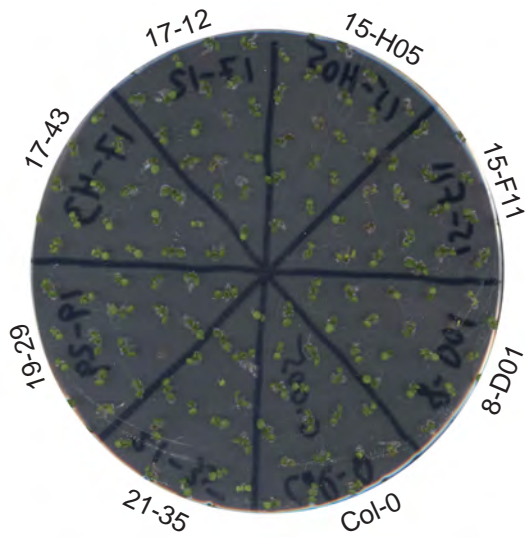
0.05 mM lincomycin



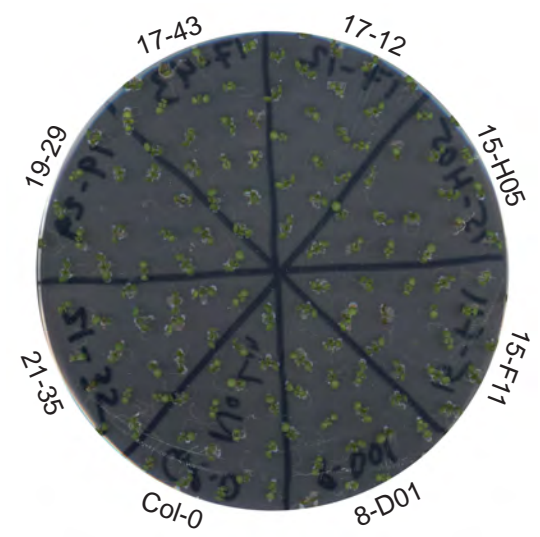
0.015 mM lincomycin



0.005 mM lincomycin

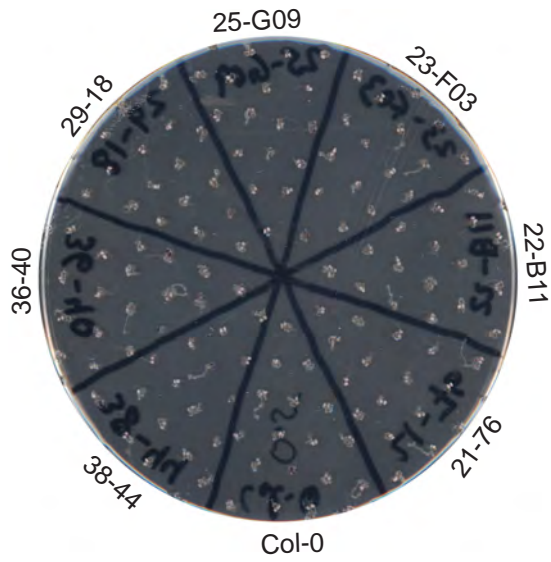


No lincomycin

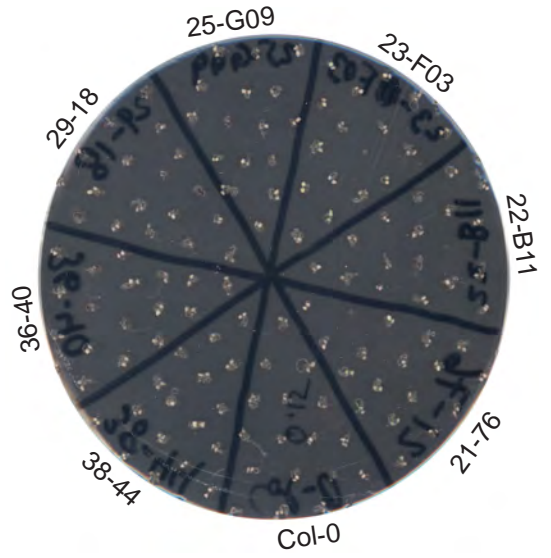


**Supplemental Figure S15.** 8-D01, 15-F11, 15-H05, 17-12, 17-43, 19-29, and 21-35 treated with various concentrations of lincomycin. The indicated *end* mutants were grown for 8 d in continuous white light at a fluence rate of  $100 \mu\text{mol m}^{-2} \text{s}^{-1}$  on media containing the indicated concentrations of lincomycin.

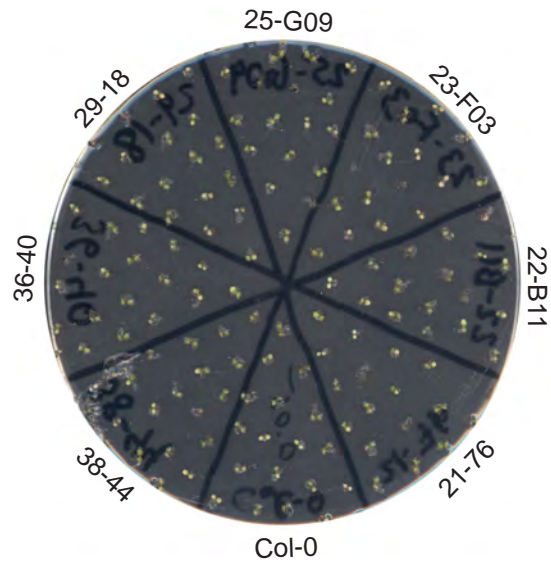
0.5 mM lincomycin



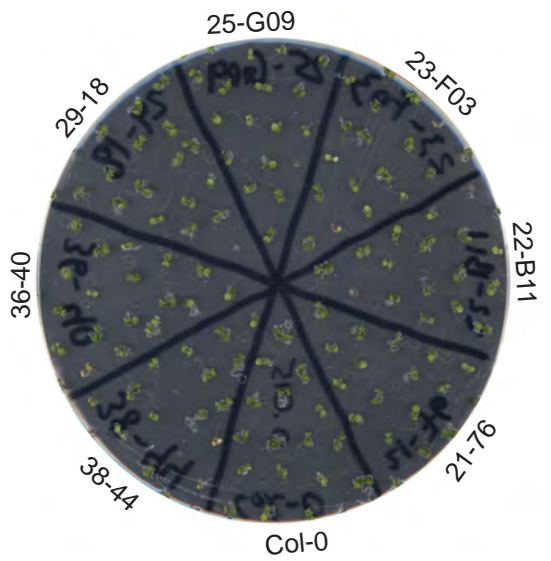
0.15 mM lincomycin



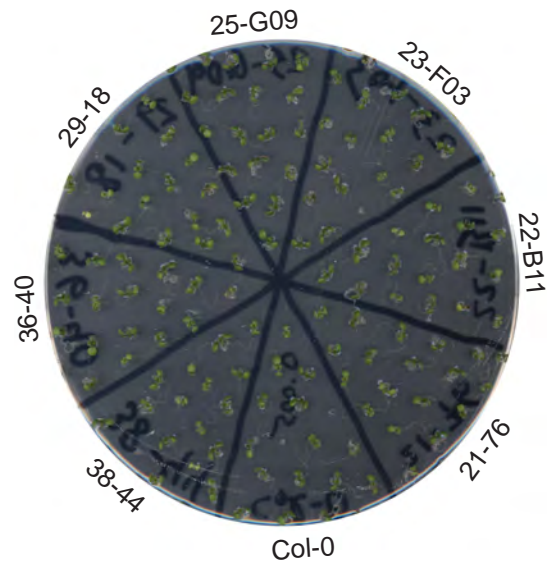
0.05 mM lincomycin



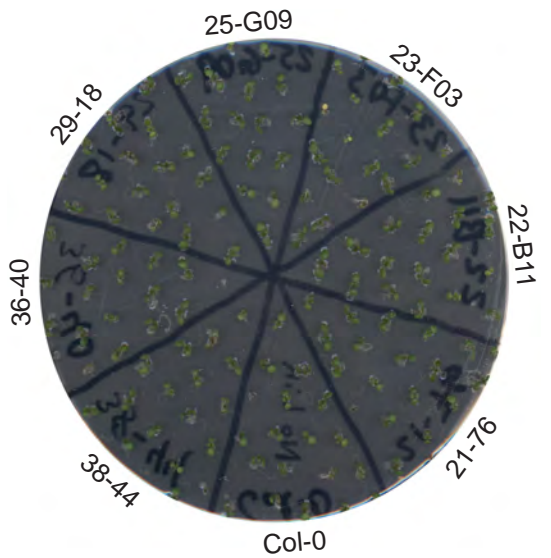
0.015 mM lincomycin



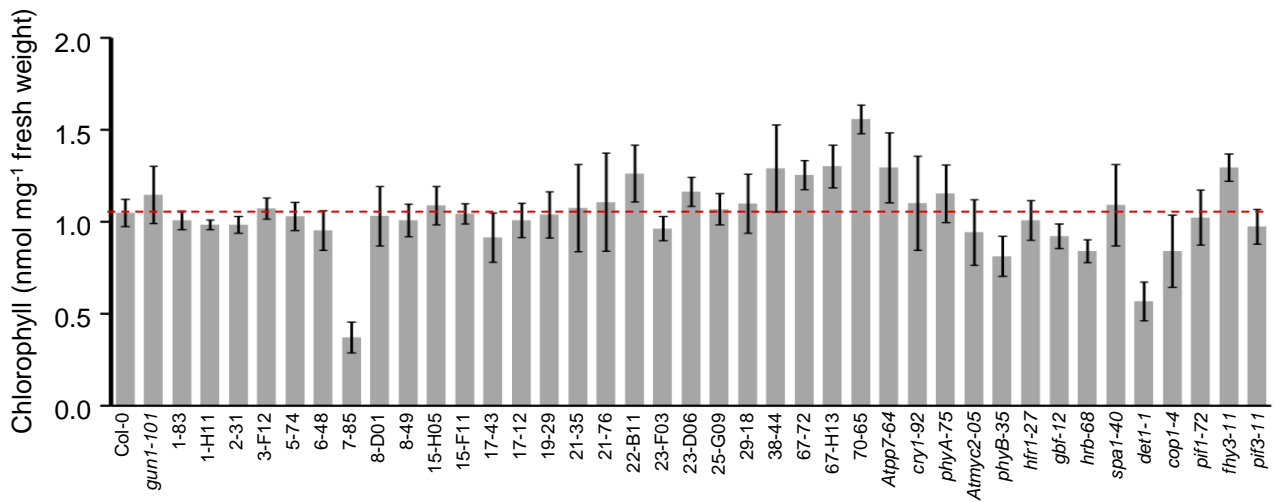
0.005 mM lincomycin



No lincomycin



**Supplemental Figure S16.** 21-76, 22-B11, 23-F03, 25-G09, 29-18, 36-40, and 38-44 treated with various concentrations of lincomycin. The indicated *end* mutants were grown for 8 d in continuous white light at a fluence rate of  $100 \mu\text{mol m}^{-2} \text{s}^{-1}$  on media containing the indicated concentrations of lincomycin.



**Supplemental Figure S17.** Chlorophyll levels in *end* mutants and light signaling mutants grown in continuous white light. Mutants that accumulated significantly more or less chlorophyll than wild type (Col-0) during de-etiolation experiments (Figs. 7A and B; Figs. 8A and B; Supplemental Fig. S11) were grown for 7 d in continuous 125  $\mu\text{mol m}^{-2} \text{s}^{-1}$  broad spectrum white light. Chlorophyll was extracted and quantified from four biological replicates for wild type and each mutant. The mean level of chlorophyll in wild type is indicated with a red-dashed line. Error bars indicate 95% confidence intervals.

**Supplemental Table S1.** Significance of enriched biological process and cellular component GO terms in particular expression patterns

	GO Term	GO ID	Enrichment in all genes	Expressed genes / Total genes in GO term	Light pattern of expression with the highest enrichment		Light cluster of expression with the highest enrichment		Plastid pattern of expression with the highest enrichment		Plastid cluster of expression with the highest enrichment		
					p-value	Pattern number	p-value	Cluster letter	p-value	Pattern number	p-value	Cluster letter	p-value
<b>Plastid</b>													
1	P	Photosynthesis	GO:0015979	8x10 <sup>-29</sup>	74/90	12	4x10 <sup>-15</sup>	B	4x10 <sup>-21</sup>	25	3x10 <sup>-34</sup>	Q	2x10 <sup>-35</sup>
2	C	Thylakoid lumen	GO:0031977	5x10 <sup>-17</sup>	45/48	12	2x10 <sup>-17</sup>	B	1x10 <sup>-27</sup>	23	4x10 <sup>-33</sup>	Q	2x10 <sup>-23</sup>
3	C	Photosystem II	GO:0009523	8x10 <sup>-14</sup>	14/15	12	8x10 <sup>-5</sup>	B	1x10 <sup>-6</sup>	25	1x10 <sup>-27</sup>	Q	3x10 <sup>-23</sup>
4	C	Thylakoid membrane	GO:0042651	9x10 <sup>-20</sup>	48/64	12	8x10 <sup>-17</sup>	B	4x10 <sup>-20</sup>	23	3x10 <sup>-26</sup>	Q	3x10 <sup>-26</sup>
5	C	Plastid stroma	GO:0009532	1x10 <sup>-16</sup>	80/117	34	2x10 <sup>-11</sup>	B	5x10 <sup>-21</sup>	23	7x10 <sup>-17</sup>	N	7x10 <sup>-13</sup>
6	C	Plastoglobule	GO:0010287	3x10 <sup>-15</sup>	49/56	12	6x10 <sup>-8</sup>	B	4x10 <sup>-13</sup>	21	1x10 <sup>-13</sup>	Q	5x10 <sup>-20</sup>
7	C	Photosystem I	GO:0009522	1x10 <sup>-5</sup>	8/9	32	1.20x10 <sup>-4</sup>	B	4x10 <sup>-5</sup>	22	7x10 <sup>-14</sup>	Q	4x10 <sup>-15</sup>
8	C	Stromule	GO:0010319	5x10 <sup>-5</sup>	22/33	12	1x10 <sup>-9</sup>	B	2x10 <sup>-13</sup>	21	7x10 <sup>-9</sup>	Q	2x10 <sup>-6</sup>
9	C	NAD(P)H dehydro-genase complex	GO:0010598	8x10 <sup>-5</sup>	9/9	12	2x10 <sup>-7</sup>	Y	1x10 <sup>-9</sup>	22	1x10 <sup>-10</sup>	Q	2x10 <sup>-11</sup>
10	P	Tetrapyrrole metabolic process	GO:0033013	6x10 <sup>-6</sup>	38/61	12	5x10 <sup>-6</sup>	B	2x10 <sup>-10</sup>	23	1x10 <sup>-6</sup>	P	3x10 <sup>-6</sup>
11	P	Plastid organization	GO:0009657	5x10 <sup>-6</sup>	49/84	30	2x10 <sup>-7</sup>	Z	7x10 <sup>-10</sup>	21	1.66x10 <sup>-4</sup>	N	7x10 <sup>-9</sup>
12	P	Carotenoid metabolic process	GO:0016116	7.41x10 <sup>-4</sup>	16/24	N/A	N/A	B	1x10 <sup>-7</sup>	N/A	N/A	N	9x10 <sup>-5</sup>
13	C	Plastid ribosome	GO:0009547	3.55x10 <sup>-4</sup>	16/24	N/A	N/A	EE	2x10 <sup>-7</sup>	N/A	N/A	J	4x10 <sup>-5</sup>
14	C	Plastid chromosome	GO:0009508	1.95x10 <sup>-4</sup>	13/16	10	1x10 <sup>-6</sup>	Y	1x10 <sup>-5</sup>	25	1.26x10 <sup>-2</sup>	N	1x10 <sup>-5</sup>
15	C	Plastid membrane	GO:0042170	2.82x10 <sup>-3</sup>	57/104	N/A	N/A	Z	2x10 <sup>-6</sup>	N/A	N/A	P	2x10 <sup>-6</sup>
<b>Translation</b>													
1	P	Translation	GO:0006412	2x10 <sup>-6</sup>	32/59	32	5x10 <sup>-24</sup>	E	1x10 <sup>-19</sup>	23	7.08x10 <sup>-3</sup>	N	9x10 <sup>-23</sup>
2	C	Cytosolic ribosome	GO:0022626	1.91x10 <sup>-1</sup>	70/181	32	1x10 <sup>-21</sup>	E	6x10 <sup>-18</sup>	24	4.57x10 <sup>-1</sup>	N	3x10 <sup>-12</sup>
3	P	Ribosome biogenesis	GO:0042254	2x10 <sup>-5</sup>	8/10	32	1x10 <sup>-12</sup>	E	8x10 <sup>-8</sup>	23	3.72x10 <sup>-4</sup>	N	2x10 <sup>-8</sup>
<b>Growth and development</b>													
1	P	Cell cycle	GO:0007049	7.41x10 <sup>-1</sup>	39/123	N/A	N/A	EE	9x10 <sup>-7</sup>	N/A	N/A	K	4.79x10 <sup>-3</sup>
2	P	Embryonic development	GO:0009790	1.62x10 <sup>-1</sup>	123/326	N/A	N/A	Z	1x10 <sup>-5</sup>	N/A	N/A	N	2.88x10 <sup>-1</sup>
3	C	Cell wall	GO:0005618	1.05x10 <sup>-1</sup>	64/123	31	2.57x10 <sup>-1</sup>	N/A	N/A	18	2x10 <sup>-5</sup>	N/A	N/A
4	P	Cell wall modification	GO:0042545	1.23x10 <sup>-1</sup>	27/61	N/A	N/A	CC	1.66x10 <sup>-3</sup>	N/A	N/A	K	7x10 <sup>-6</sup>
5	P	DNA replication	GO:0006260	6.31x10 <sup>-2</sup>	21/43	27	4x10 <sup>-6</sup>	DD	9x10 <sup>-5</sup>	28	7x10 <sup>-7</sup>	K	4x10 <sup>-6</sup>
6	P	Regulation of post-embryonic development	GO:0048580	1.58x10 <sup>-2</sup>	57/135	N/A	N/A	GG	1x10 <sup>-5</sup>	N/A	N/A	M	3.31x10 <sup>-4</sup>
<b>Regulation of gene expression</b>													
1	P	Regulation of transcription	GO:0045449	9.33x10 <sup>-1</sup>	206/558	4	9x10 <sup>-7</sup>	R	4x10 <sup>-10</sup>	21	3.16x10 <sup>-1</sup>	M	2x10 <sup>-5</sup>
2	P	Cellular protein catabolic process	GO:0044257	3.02x10 <sup>-1</sup>	56/134	33	4x10 <sup>-7</sup>	GG	3x10 <sup>-7</sup>	20	7.94x10 <sup>-4</sup>	L	1.86x10 <sup>-4</sup>
2	C	Ubiquitin ligase complex	GO:0000151	1.66x10 <sup>-2</sup>	19/42	16	2x10 <sup>-5</sup>	GG	1.12x10 <sup>-2</sup>	20	1x10 <sup>-7</sup>	L	3x10 <sup>-6</sup>
3	P	RNA processing	GO:0006396	7x10 <sup>-6</sup>	47/96	6	7x10 <sup>-6</sup>	GG	6x10 <sup>-7</sup>	21	5.25x10 <sup>-1</sup>	O	3x10 <sup>-5</sup>
<b>Metabolism</b>													
1	P	Carbon utilization	GO:0015976	3.31x10 <sup>-4</sup>	11/13	12	1.05x10 <sup>-3</sup>	Y	6.31x10 <sup>-4</sup>	21	8x10 <sup>-8</sup>	Q	3x10 <sup>-6</sup>
3	P	Cellular amino acid metabolism	GO:0006520	3x10 <sup>-7</sup>	59/113	34	1x10 <sup>-6</sup>	E	3x10 <sup>-6</sup>	22	2.10x10 <sup>-3</sup>	N	4x10 <sup>-5</sup>
4	P	Lipid transport	GO:0006869	4.79x10 <sup>-4</sup>	9/16	N/A	N/A	F	3x10 <sup>-7</sup>	N/A	N/A	J	9x10 <sup>-7</sup>
5	P	Starch metabolic process	GO:005982	1.23x10 <sup>-3</sup>	21/33	32	2.45x10 <sup>-4</sup>	N/A	N/A	23	3x10 <sup>-6</sup>	N/A	N/A
6	P	Monosaccharide metabolic process	GO:0005996	6.61x10 <sup>-4</sup>	22/45	30	4x10 <sup>-6</sup>	B	8x10 <sup>-6</sup>	21	3.72x10 <sup>-4</sup>	P	1.82x10 <sup>-3</sup>
<b>Other cellular components</b>													
1	C	Endoplasmic reticulum	GO:0005783	2.00x10 <sup>-1</sup>	103/267	N/A	N/A	H	1x10 <sup>-5</sup>	N/A	N/A	J	2.88x10 <sup>-2</sup>
2	C	Mitochondrion	GO:0005739	8x10 <sup>-5</sup>	232/551	32	2x10 <sup>-9</sup>	Z	6x10 <sup>-16</sup>	29	4.72x10 <sup>-2</sup>	J	2x10 <sup>-5</sup>

Testing for the enrichment of GO terms was described in Materials and Methods, Fig. 2, Supplemental Figs. S2, S3, S4, and S5. Enrichment in all genes refers to the significance of enrichment of a particular GO term in the entire dataset of 7104 genes. Ontologizer 2.0 was used to calculate the indicated p-values. The ratio of genes in our dataset that are associated with a particular GO term to the total number of genes that are associated with that same GO term is presented for each GO term. The most significantly enriched light- and plastid-regulated expression pattern and cluster is indicated for each GO term. If a particular pattern or cluster was not significantly enriched (p<0.01), a p-value is not presented (N/A).

**Supplemental Table S2.** Significance of enriched biological process and response to stimulus GO terms in particular expression patterns

GO Term	GO ID	Enrichment in all genes	Expressed genes / Total genes in GO term	Light pattern of expression with the highest enrichment		Light cluster of expression with the highest enrichment		Plastid pattern of expression with the highest enrichment		Plastid cluster of expression with the highest enrichment			
				p-value	Pattern number	p-value	Cluster letter	p-value	Pattern number	p-value	Cluster letter		
<b>Oxidative Stress and ROS scavenging</b>													
1	R	Oxidative stress	GO:0006979	6x10 <sup>-10</sup>	102/173	34	3.16x10 <sup>-4</sup>	I	1x10 <sup>-5</sup>	21	8x10 <sup>-6</sup>	O	5.13x10 <sup>-4</sup>
2	P	Phenyl propanoid Metabolic process	GO:0009698	4x10 <sup>-5</sup>	43/82	N/A	N/A	AA	1x10 <sup>-9</sup>	N/A	N/A	J	2.45x10 <sup>-6</sup>
3	P	Glycoside metabolic process	GO:0016137	3x10 <sup>-4</sup>	30/48	16	1x10 <sup>-6</sup>	AA	1x10 <sup>-6</sup>	18	1x10 <sup>-7</sup>	K	3x10 <sup>-6</sup>
4	P	Cell death	GO:0008219	1.86x10 <sup>-3</sup>	44/80	N/A	N/A	FF	4x10 <sup>-7</sup>	N/A	N/A	M	2.09x10 <sup>-2</sup>
<b>Biotic and Abiotic stress</b>													
1	R	Heat	GO:0009408	4x10 <sup>-8</sup>	57/99	6	1x10 <sup>-6</sup>	C	8x10 <sup>-25</sup>	20	3.16x10 <sup>-3</sup>	M	1.82x10 <sup>-3</sup>
2	R	Cold	GO:0009409	1x10 <sup>-13</sup>	128/216	34	1x10 <sup>-7</sup>	S	4x10 <sup>-5</sup>	22	5x10 <sup>-5</sup>	J	2x10 <sup>-6</sup>
3	R	Water deprivation	GO:0009414	2x10 <sup>-13</sup>	97/145	34	6x10 <sup>-9</sup>	H	2x10 <sup>-8</sup>	20	6x10 <sup>-10</sup>	J	2x10 <sup>-8</sup>
4	R	Salt stress	GO:0009651	6x10 <sup>-12</sup>	117/202	33	5x10 <sup>-6</sup>	H	3x10 <sup>-8</sup>	20	8x10 <sup>-5</sup>	M	2x10 <sup>-7</sup>
5	R	Wounding	GO:0009611	1x10 <sup>-7</sup>	67/114	15	6x10 <sup>-5</sup>	CC	3.24x10 <sup>-3</sup>	22	2.69x10 <sup>-3</sup>	J	6.76x10 <sup>-4</sup>
6	R	Carbohydrate stimulus	GO:0009743	4x10 <sup>-7</sup>	93/174	15	2x10 <sup>-6</sup>	D	1x10 <sup>-5</sup>	20	3.02x10 <sup>-2</sup>	M	7x10 <sup>-6</sup>
7	R	Metal ion	GO:0010038	9x10 <sup>-6</sup>	44/70	11	2.24x10 <sup>-3</sup>	D	6x10 <sup>-5</sup>	25	2.14x10 <sup>-2</sup>	M	1.26x10 <sup>-3</sup>
<b>Light</b>													
1	R	UV light	GO:0009411	5x10 <sup>-7</sup>	38/55	4	2x10 <sup>-6</sup>	AA	4x10 <sup>-16</sup>	20	4x10 <sup>-5</sup>	K	3.02x10 <sup>-2</sup>
2	R	High light intensity	GO:0009644	3x10 <sup>-10</sup>	34/41	4	8x10 <sup>-5</sup>	C	1x10 <sup>-9</sup>	21	9x10 <sup>-6</sup>	P	1x10 <sup>-7</sup>
3	R	Red and far-red light	GO:0009639	6x10 <sup>-6</sup>	77/129	4	2x10 <sup>-6</sup>	R	2x10 <sup>-7</sup>	21	3x10 <sup>-8</sup>	M	9x10 <sup>-6</sup>
4	R	Blue light	GO:0009637	1x10 <sup>-6</sup>	30/44	4	8.71x10 <sup>-4</sup>	U	1.38x10 <sup>-3</sup>	21	3.98x10 <sup>-4</sup>	Q	8x10 <sup>-5</sup>
5	P	Circadian rhythm	GO:0007623	6x10 <sup>-6</sup>	30/43	33	1.66x10 <sup>-4</sup>	BB	1.51x10 <sup>-4</sup>	21	8x10 <sup>-5</sup>	L	4.37x10 <sup>-4</sup>
<b>Hormones</b>													
1	R	Abscisic acid stimulus	GO:0009737	4x10 <sup>-12</sup>	137/245	17	2x10 <sup>-6</sup>	H	9x10 <sup>-9</sup>	20	6x10 <sup>-8</sup>	J	9x10 <sup>-9</sup>
2	R	Auxin stimulus	GO:0009733	3.02x10 <sup>-3</sup>	76/169	3	2x10 <sup>-6</sup>	X	7x10 <sup>-11</sup>	21	1.95x10 <sup>-3</sup>	M	2x10 <sup>-9</sup>
3	R	Jasmonic acid stimulus	GO:0009753	1x10 <sup>-8</sup>	80/140	2	1.10x10 <sup>-4</sup>	T	1x10 <sup>-5</sup>	23	1.86x10 <sup>-3</sup>	J	1.51x10 <sup>-4</sup>
5	R	Gibberellin stimulus	GO:0009739	3x10 <sup>-8</sup>	59/99	11	9x10 <sup>-6</sup>	H	4x10 <sup>-7</sup>	20	2.35x10 <sup>-3</sup>	L	2.00x10 <sup>-3</sup>

Testing for the enrichment of GO terms was described in Materials and Methods, Fig. 2, Supplemental Figs. S2, S3, S4, and S5. Enrichment in all genes refers to the significance of enrichment of a particular GO term in the entire dataset of 7104 genes. Ontologizer 2.0 was used to calculate the indicated p-values. The ratio of genes in our dataset that are associated with a particular GO term to the total number of genes that are associated with that same GO term is presented for each GO term. The most significantly enriched light- and plastid-regulated expression pattern and cluster is presented for each GO term. If a particular pattern or cluster was not significantly enriched ( $p < 0.01$ ), a p-value is not presented (N/A).

**Supplemental Table S3.** Genes that exhibit enhanced light-induced expression in lincomycin-treated seedlings and that lack publicly available T-DNA alleles

Number	Arabidopsis Genome Initiative (AGI) Identifier	Light Induction	Plastid regulation	Name/Description	Biological function, process, and location
4	At1g14345	2.45	-5.92	Aldo/keto reductase domain	Oxidation reduction <sup>IEA</sup> , Chloroplast thylakoid membrane <sup>a</sup>
9	At1g11380	2.06	-1.80	Cys-rich domain	Unknown
12	At3g49580	1.98	-1.67	LSU1, RESPONSE TO LOW SULFUR 1	Unknown
18	At2g30520	1.87	-1.21	RPT2, ROOT PHOTOTROPISM 2 <sup>b</sup>	Phototropism <sup>b</sup> , Nucleus <sup>ISS</sup>
24	At5g47610	1.76	-3.56	Zinc finger (C3HC4-type RING finger)	Protein binding <sup>ISS</sup>
26	At2g42540	1.73	5.80	COR15A, COLD-REGULATED 15A	Cold acclimation <sup>c</sup> , Chloroplast <sup>d</sup>
27	At1g12250	1.73	-2.48	Expressed protein	Chloroplast thylakoid membrane <sup>a</sup>
30	At1g16720	1.68	-2.19	HCF173, HIGH CHLOROPHYLL FLUORESCENCE 173 <sup>e</sup>	Translational initiation <sup>e</sup> , Photosystem II assembly <sup>e</sup> , Chloroplast <sup>f</sup>
31	At1g42550	1.65	-3.08	PMI1, PLASTID MOVEMENT IMPAIRED1 <sup>g</sup>	Chloroplast relocation <sup>g</sup> , Plasma membrane <sup>h</sup>
32	At1g79270	1.65	-1.68	ECT8, EVOLUTIONARILY CONSERVED C-TERMINAL REGION 8	Unknown
33	At1g32080	1.64	-6.56	LrgB-like domain protein	Chloroplast inner membrane <sup>i</sup>
34	At5g49330	1.57	1.38	ATMYB111, MYB DOMAIN PROTEIN 111	Transcription factor activity <sup>ISS</sup>
37	At2g02950	1.52	1.17	PKS1, PHYTOCHROME KINASE SUBSTRATE 1	Phototropism <sup>l</sup> , Red-far red light signaling <sup>k</sup> , Cytoplasm <sup>k</sup> , Plasma membrane <sup>j</sup>

Thirteen genes are ranked by their light induction. Light induction is defined as the ratio of light-induced expression in lincomycin-treated seedlings to light-induced expression in untreated seedlings at 1 h following the BR-fluence-rate shift. Plastid regulation is represented as the ratio of induced or repressed (-) expression in lincomycin-treated seedlings to expression in untreated seedlings at 0 h relative to the BR-fluence-rate shift. Gene names and descriptions are based on available literature or on TIGR gene annotation records. Biological function, process, and location are based on the current literature or the gene ontology with evidence codes as described in Materials and Methods. References: a (Peltier et al., 2004); b (Sakai et al., 2000); c (Lin and Thomashow, 1992); d (Kleffmann et al., 2004); e (Schult et al., 2007); f (Zybailov et al., 2008); g (DeBlasio et al., 2005); h (Nühse et al., 2003); i (Ferro et al., 2003); j (Lariguet et al., 2006); k (Fankhauser et al., 1999).



**Supplemental Table S4.** Alleles of genes that exhibit similar light-induced expression in lincomycin-treated and untreated seedlings that cause *end* phenotypes

No.	AGI code	Light Induction	Plastid regulation	Name/Description	Biological function, process, and location	T-DNA line(s)	Transcript phenotype in the homozygote
67	At5g59220	1.20	-0.10	HAI1, HIGHLY ABA-INDUCED PP2C GENE 1	Chloroplast <sup>EA</sup> , protein serine/threonine phosphatase activity <sup>SS</sup> , ABA signaling <sup>a-c</sup>	Salk_142672 SAIL_520_H12	Null Upregulated
70	At5g23730	0.83	-0.64	RUP2/EFO2, WD-40 repeat family protein	UV-B signaling <sup>d</sup> , vegetative development and flowering <sup>e</sup> , CUL4 RING ubiquitin ligase complex <sup>SS</sup>	Salk_015765	Knockdown

The light induction and the plastid regulation were calculated as described in Supplemental Table S3. Gene names and descriptions are based on available literature or on TIGR gene annotation records. Biological function, process, and location are based on the current literature or the gene ontology with evidence codes as described in Materials and Methods. References: a(Fujita et al., 2009); b(Guo et al., 2010); c(Antoni et al., 2012); d(Gruber et al., 2010); e(Wang et al., 2011).

**Supplemental Table S5.** The diverse regulators of genes that exhibit enhanced light-induced expression in lincomycin-treated seedlings

Number	AGI Code	Regulators of gene expression
1	At5g24120	Far-red light <sup>E,1</sup> , Red light <sup>E,1</sup> , UV-B <sup>M,2</sup> , Plastid signals <sup>SS,3</sup> , Blue light <sup>E,4</sup>
2	At3g56290	UV-B <sup>M,2</sup>
3	At2g30040	Nitrogen <sup>E,5</sup> , Far-red light <sup>E,1</sup> , Red light <sup>E,1</sup> , Brassinosteroid <sup>E,6</sup> , Auxin <sup>E,6</sup> , Cold <sup>E,7</sup>
5	At5g08050	NA
6	At5g24660	Sulfur <sup>M,L,8</sup>
7	At3g17040	Red light <sup>E,9</sup>
8	At1g44000	NA
10	At2g24540	Salt <sup>M,10</sup> , Osmotic <sup>M,10</sup> , Cold <sup>M,10</sup> , Circadian clock <sup>11</sup> , UV-B <sup>E,12</sup>
11	At5g35970	High light <sup>M,13</sup> , UV-B <sup>E,12</sup>
13	At4g11360	Phenylglycosides <sup>E,14</sup> , Sucrose starvation <sup>L,15</sup> , Immune response to flg22E <sup>16</sup> , Sucrose addition <sup>E,17</sup> , Chitin <sup>E,18</sup>
14	At5g14970	NA
15	At5g58650	NA
16	At2g41660	UV-B <sup>E,12</sup>
17	At5g13770	NA
19	At2g16365	NA
20	At5g52780	NA
21	At3g54990	Photoperiod <sup>19</sup> , Nitrogen <sup>E,5</sup>
22	At5g62430	Circadian clock <sup>20</sup>
23	At3g02380	Reactive oxygen <sup>SS,21</sup> , Far-red light <sup>E,1</sup> , Red light <sup>E,1</sup>
25	At1g43160	Wounding <sup>E,22</sup> , Intense light <sup>E,23</sup> , Brassinosteroid <sup>E,6</sup> , Auxin <sup>E,6</sup> , Drought <sup>L,24</sup> , Cytokinin <sup>E,25</sup> , Cold <sup>M,26</sup> , Blue light <sup>E,27</sup> , Far-red light <sup>E,28</sup> , Red light <sup>E,28</sup>
28	At5g52250	Far-red light <sup>E,1</sup> , Red light <sup>E,1</sup> , UV-B <sup>E,29</sup> , High light <sup>M,13</sup>
29	At1g04770	Sulfur <sup>M,L,8</sup>
35	At2g33250	NA
36	At2g46340	Far-red light <sup>E,1</sup> , Red light <sup>E,1</sup> , Blue light <sup>E,30</sup> , Green light <sup>E,31</sup>
38	At4g28740	NA

Only signals that both regulate the expression of these genes and are indicated in the text (including embedded tables) of the indicated references were considered. Large datasets provided in the supplemental material online were not considered. The kinetics of regulated expression is also provided: E, Early, 0-2 h; M, Mid, 2-8 h; L, Late, 8-24 h; SS, Steady State. References: 1 (Khanna et al., 2006); 2 (Brown et al., 2005); 3 (Ankele et al., 2007); 4 (Onda et al., 2008); 5 (Scheible et al., 2004); 6 (Goda et al., 2004); 7 (Lee et al., 2005); 8 (Maruyama-Nakashita et al., 2005); 9 (Monte et al., 2004); 10 (Kreps et al., 2002); 11 (Harmon and Kay, 2003); 12 (Oravecz et al., 2006); 13 (Kleine et al., 2007); 14 (Guan and Nothnagel, 2004); 15 (Contento et al., 2004); 16 (Navarro et al., 2004); 17 (Osuna et al., 2007); 18 (Libault et al., 2007); 19 (Schmid et al., 2003); 20 (Imaizumi et al., 2005); 21 (Charron et al., 2008); 22 (Yan et al., 2007); 23 (Rossel et al., 2007); 24 (Catala et al., 2007); 25 (Rashotte et al., 2003); 26 (Fowler and Thomashow, 2002); 27 (Folta et al., 2003); 28 (Tepperman et al., 2004); 29 (Ulm et al., 2004); 30 (Fittinghoff et al., 2006); 31 (Dhingra et al., 2006).

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