

Supplemental Figures

Figure S1. Generation of intestinal epithelial-specific STAT5 knock out mice (STAT5^{IEC KO}).

- A.** STAT5^{IEC KO} mice were generated using cre-mediated recombination to delete the *stat5* locus in IECs.
B. Genotypes of *villin*-cre *Stat5* floxed mice were determined by PCR.

Figure S2.

- A.** Mice were treated with 3% DSS for 7 days and then followed by 14 days of water. Weight gain and loss percentage were evaluated. Results were expressed as the mean \pm SEM (n = 5 or 6), * $p < 0.01$ versus STAT5^{IEC WT} mice.
B. The tyrosine phosphorylation of STAT5 (pSTAT5) was determined by IH in the inflamed colonic crypts and proliferating crypt cells in STAT5^{IEC KO} and littermate control mice exposed to DSS for 7 days, n = 7. Original magnification, $\times 100$, bar = 100 μm .

Figure S3. Mice were treated with 3% DSS for 5 days and then followed by 5 days of water.

- A.** Regulatory T cells (CD4⁺CD25⁺Foxp3⁺), activated CD4⁺ T cells (CD4⁺CD44^{hi}CD62L^{lo}), and macrophages (CD4⁻CD11c⁻CD11b⁺F4/80⁺) in MLNs were determined by FACS. Representative scatter graphs were shown.
B, C. Paraffin-embedded colonic sections were labeled with F4/80 and Foxp3. F4/80 and Foxp3 positive cells appeared brown, n = 5. Original magnification, $\times 400$; bar = 100 μm .
D. MLNs were imaged and weighed, open circles represent MLNs.

Figure S4. IEC apoptosis and proliferation in STAT5^{IEC KO} mice.

- A, B.** IEC apoptosis was detected by cleaved Caspase-3 immunohistochemistry staining (A), and IEC proliferation was determined using BrdU incorporation measured by immunohistochemistry staining (B); original magnification, $\times 400$, bar = 100 μm . Results were expressed as the mean \pm SEM (n = 7).

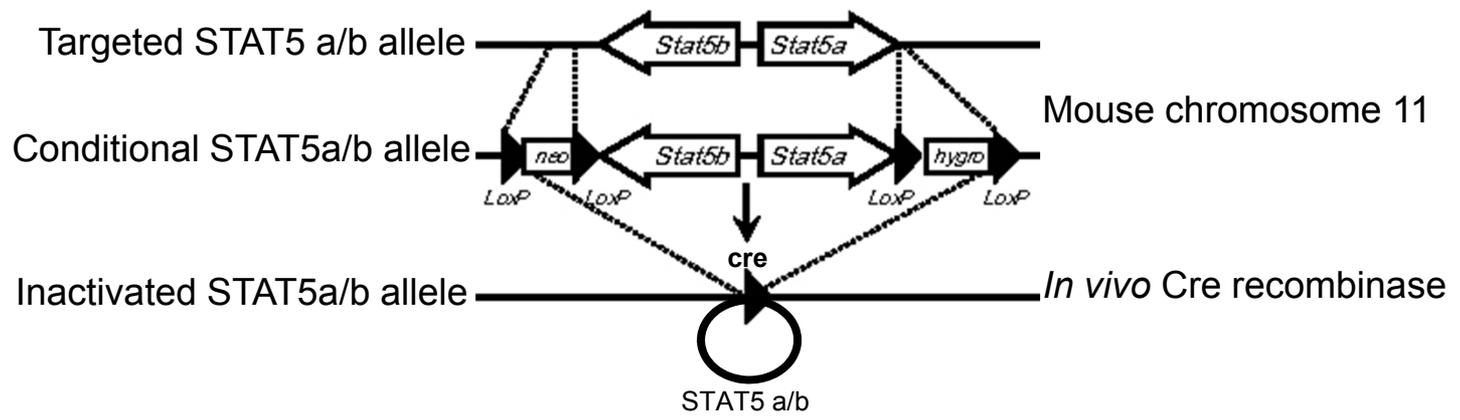
Figure S5.

- A.** Colonic IECs were isolated from STAT5^{IEC WT} and STAT5^{IEC KO} mice, ZO-1, -2 and -3, and occludin were measured with WB. HT-29 IEC monolayers on transwell filters, with and without stat5 RNAi, were exposed to IFN γ (10 ng/ml) induction for 18 hrs and then TNF α (10 ng/ml) stimulation for 12 hrs.
B. HT-29 cell monolayers on the transwell inserts were immunostained with ZO-1 & 3, n = 3. Original magnification is $\times 600$, bar = 100 μm .
C. Nuclear proteins (NE) and cytosolic proteins (CE) were isolated from HT-29 monolayers on the transwell inserts, pSTAT5 and STAT5 abundances in NE and CE were measured by WB, The representative result is shown from three repeated experiments.

Supplemental Table: PCR Primers.

Fig S1

A



B

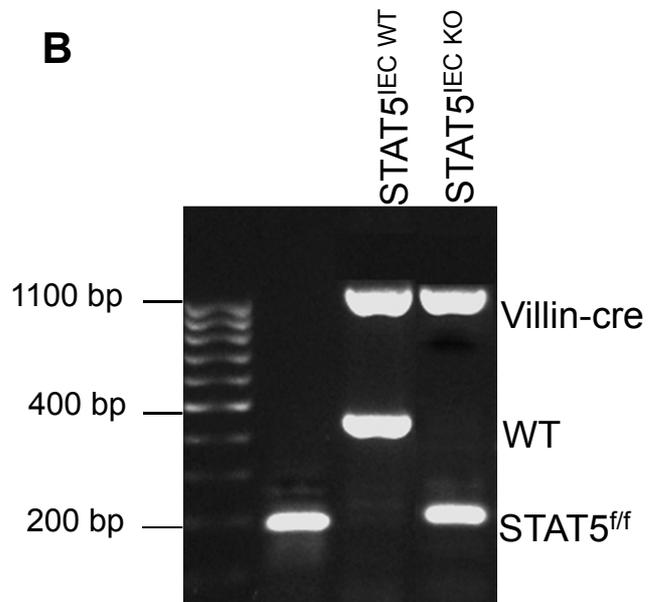


Fig S2

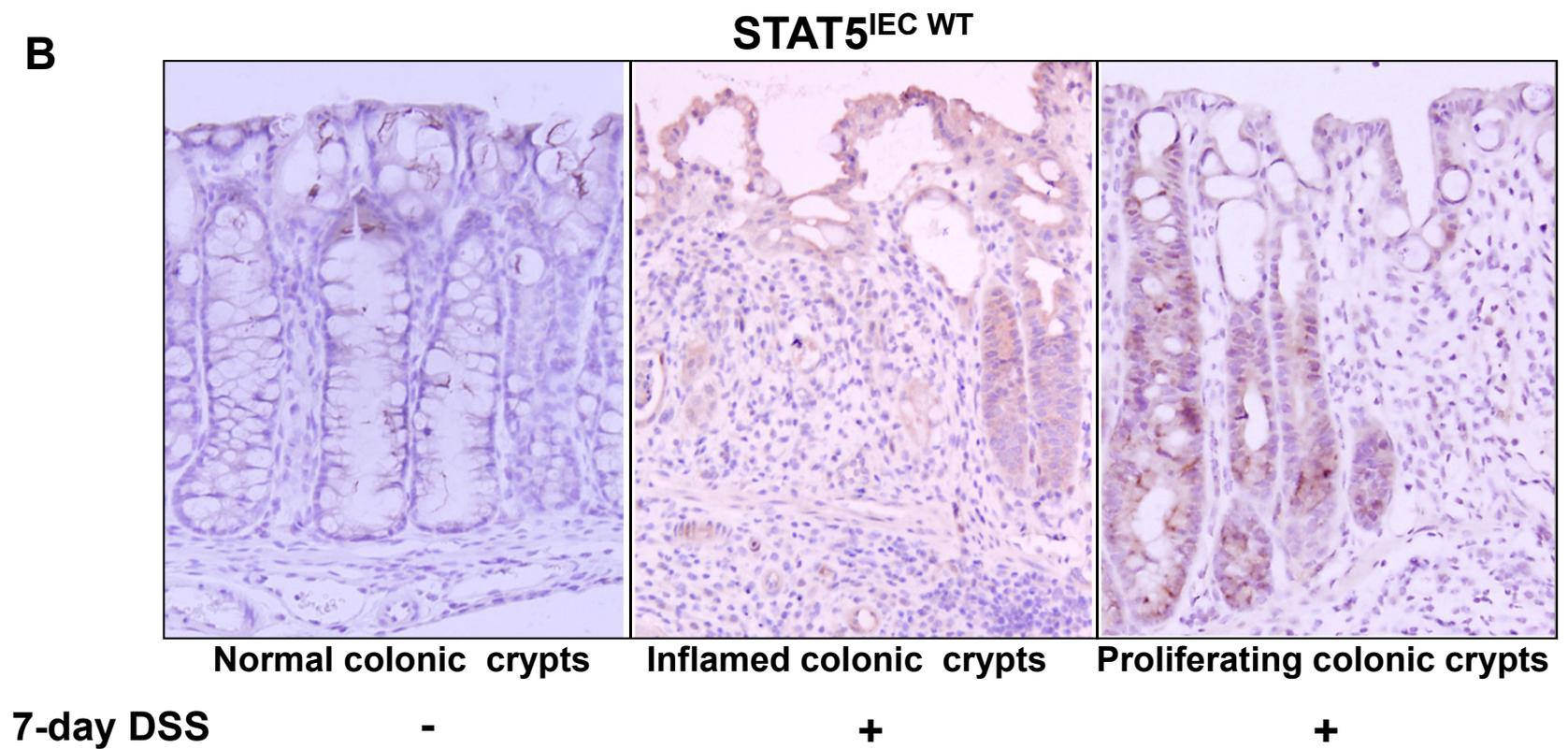
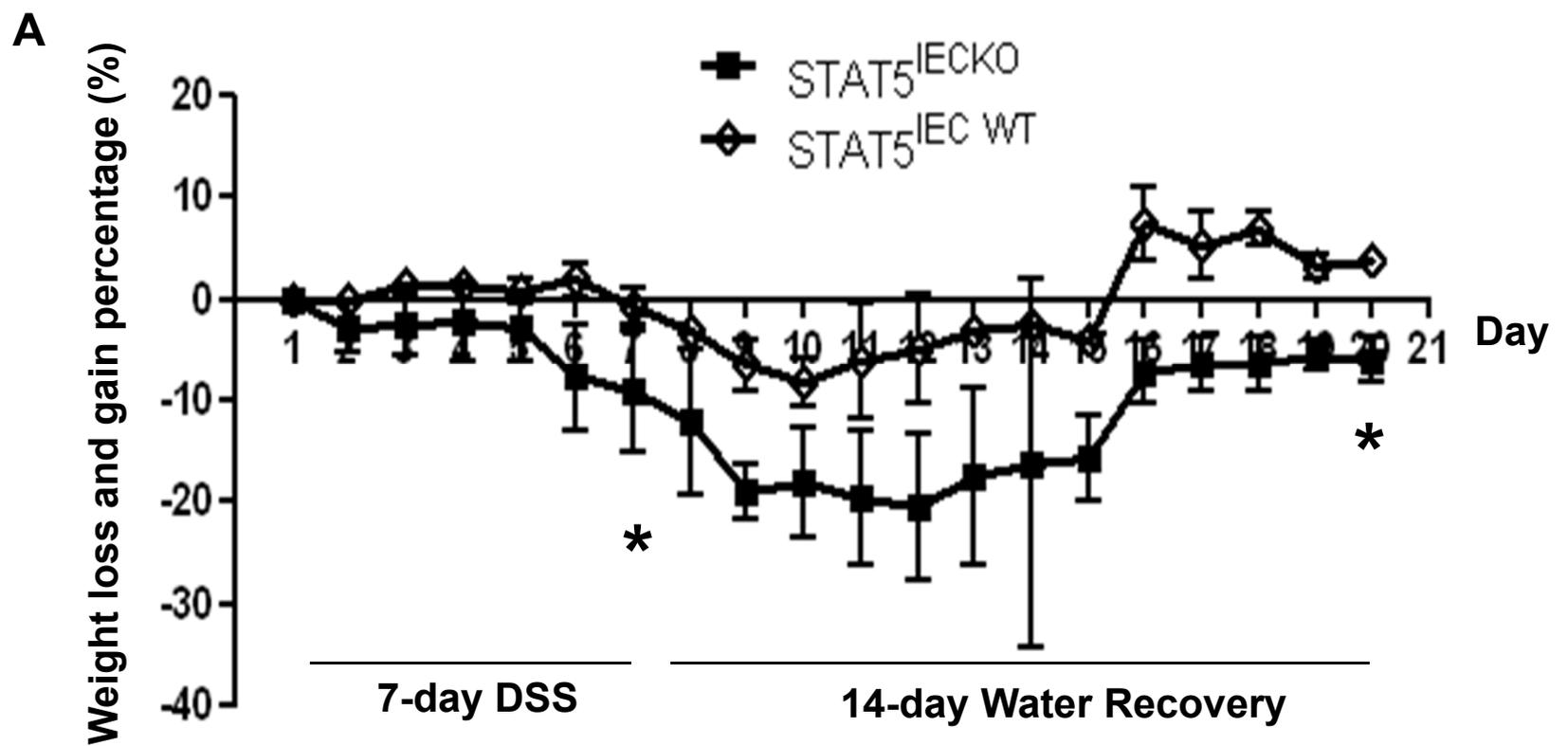
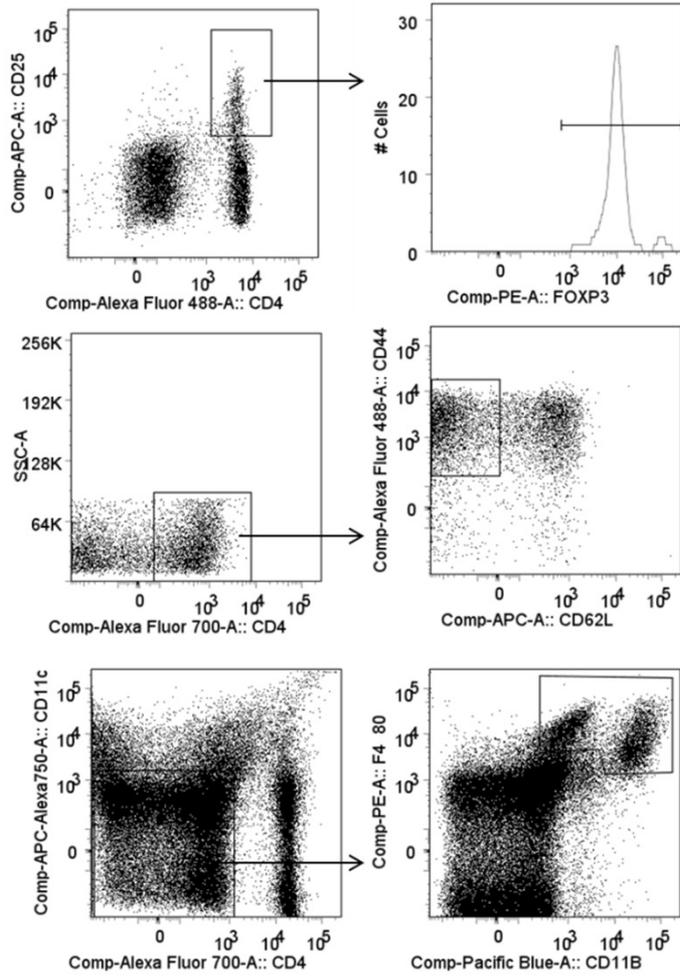
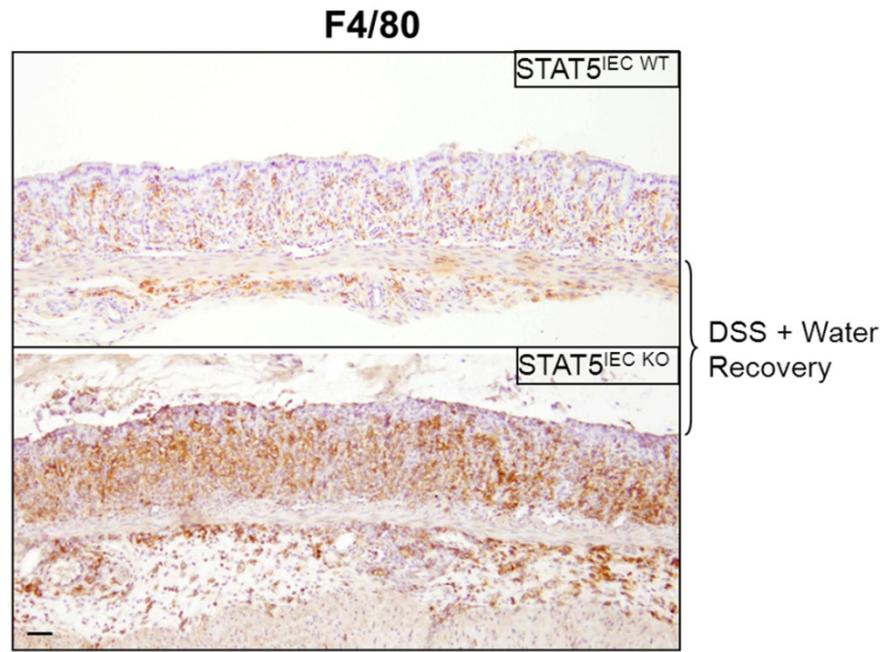


Fig S3

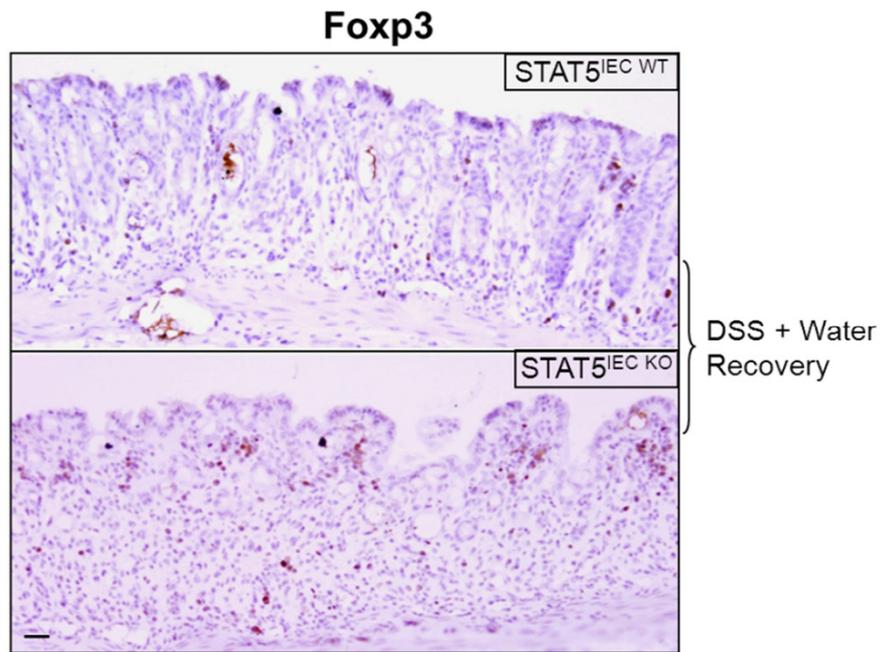
A



B



C



D

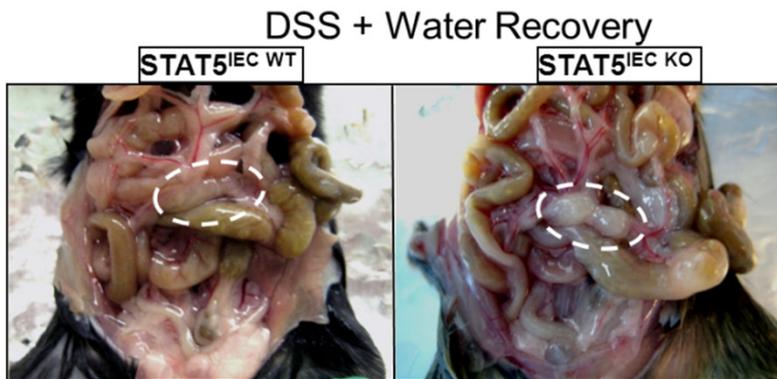
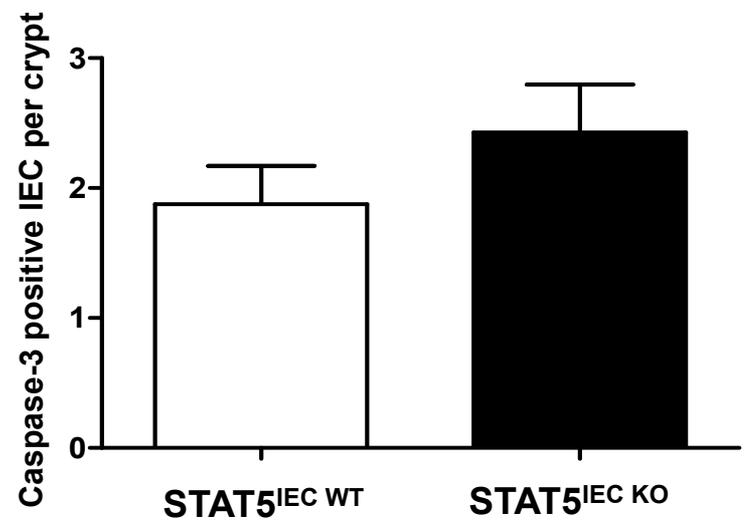
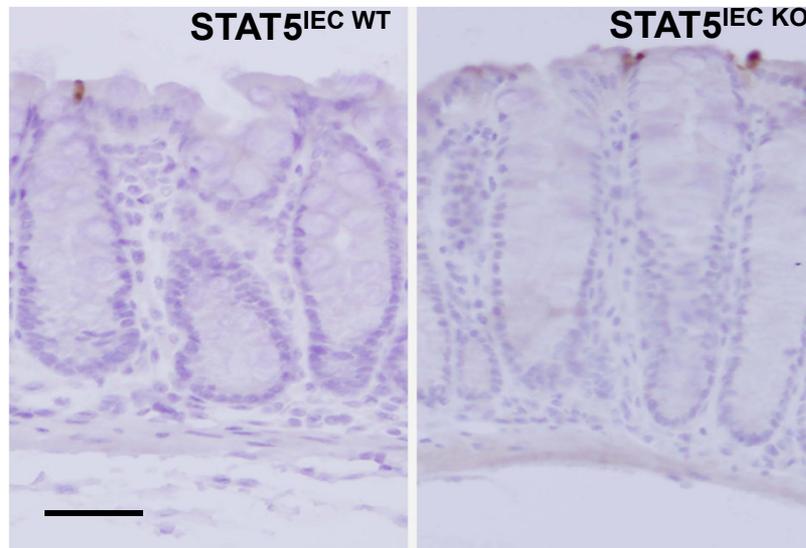


Fig S4

A



B

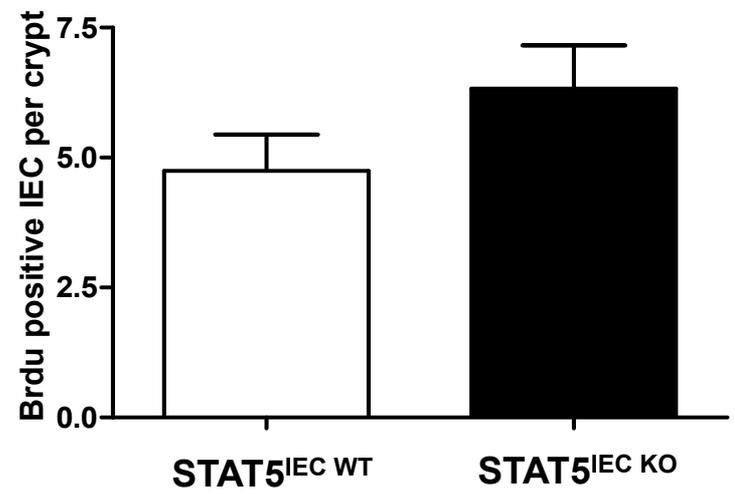
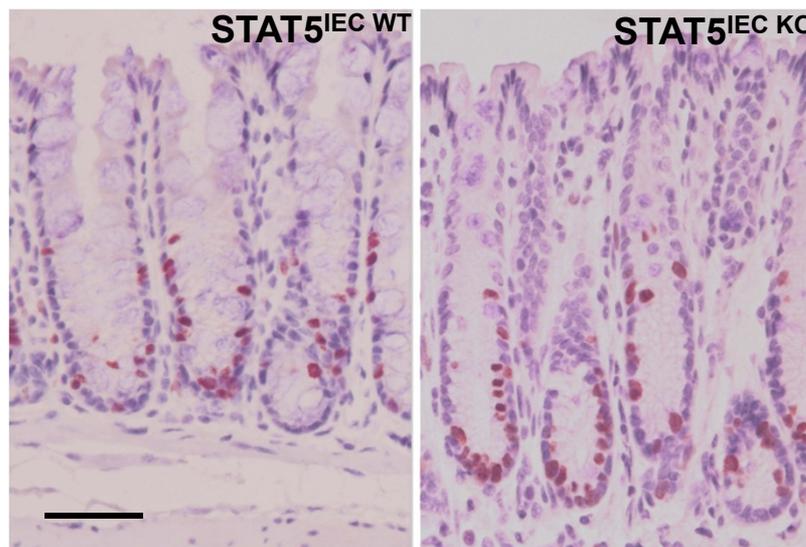
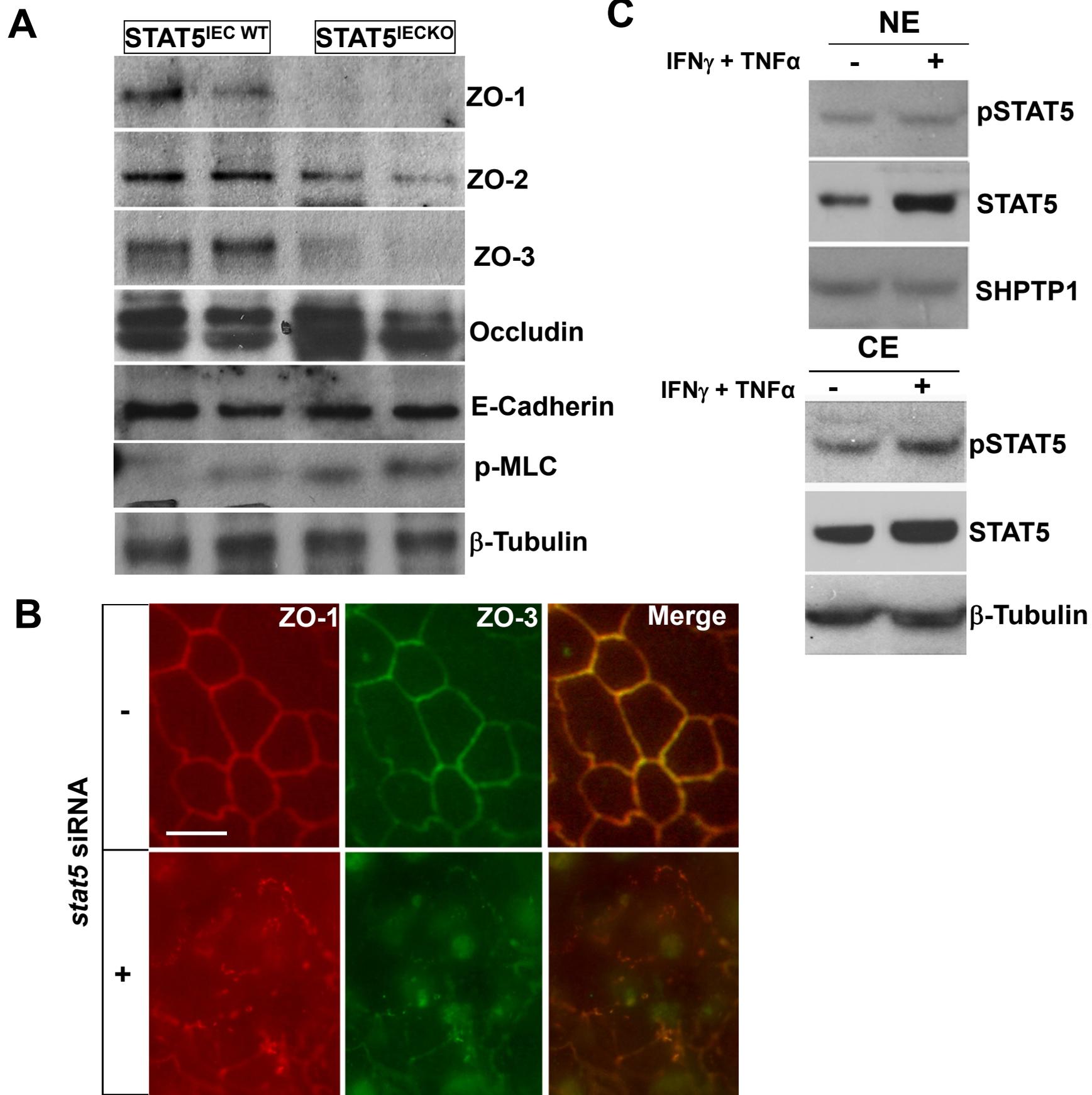


Fig S5



Supplemental Table: PCR Primers and EMSA Oligonucleotide Sequences
(Denning et al, 2007; Han et al, 2011; Han et al, 2009; Lewkowich et al, 2008; Shivakumar et al, 2004; Villarino et al, 2008).

Primers	Sequences
GAPDH	Forward: 5'-GGTGGGTGGTCCAAGGTTTC-3' Reverse: 5'-TGGTTTGACAATGAATACGGCTAC-3'
β actin	Forward: 5'- CCCTAAGGCCAACCGTGAA -3' Reverse: 5'- CAGCCTGGATGGCTACGTACA -3'
TNF α	Forward: 5' - AATGGCCTCCCTCTCATCAGTT - 3' Reverse: 5' - CCACTTGGTGGTTTGCTACGA - 3'
IFN γ	Forward: 5' - GGCTGTCCCTGAAAGAAAGC - 3' Reverse: 5' - GAGCGAGTTATTTGTCATTCGG - 3'
IL-6	Forward: 5' - CAAAGCCAGAGTCCTTCAGAGAGATAC - 3' Reverse: 5' - GGATGGTCTTGGTCCTTAGCCAC - 3'
IL-10	Forward: 5' - TGGTTTCTCTTCCCAAGACC - 3' Reverse: 5' - CCCTTTGCTATGGTGTCCTT - 3'
IL-17	Forward: 5' - TTAACTCCCTTGGCGCAAAA - 3' Reverse: 5' - CTTTCCCTCCGCATTGACAC - 3'
IL-23	Forward: 5' - CATGGGGCTATCAGGGAGTA - 3' Reverse: 5' - GACCCACAAGGACTCAAGGA - 3'
Long MLCK	Forward: 5' - AAGTCATGGATGGAAGCCAGGTCA- 3' Reverse: 5' - AATCTCATTCCTCGTGAAGCCA- 3'
NF- κ B	Sense: 5- AGT TGA GGG GAC TTT CCC AGG C-3 Anti-sense: 3-TCA ACT CCC CTG AAA GGG TCC G-5
κ B binding	Sense: 5-CAA CTT CAG GAG CTT CCC AGC CGA G-3 Anti-sense: 3-GTT GAA GTC CTC GAA GGG TCG GCT C-5

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Supplemental References

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Han X, Ren X, Jurickova I, Groschwitz K, Pasternak BA, Xu H, Wilson TA, Hogan SP, Denson LA (2009) Regulation of intestinal barrier function by signal transducer and activator of transcription 5b. *Gut* 58: 49-58

Lewkowich IP, Lajoie S, Clark JR, Herman NS, Sproles AA, Wills-Karp M (2008) Allergen uptake, activation, and IL-23 production by pulmonary myeloid DCs drives airway hyperresponsiveness in asthma-susceptible mice. *PLoS One* 3: e3879

Shivakumar P, Campbell KM, Sabla GE, Miethke A, Tiao G, McNeal MM, Ward RL, Bezerra JA (2004) Obstruction of extrahepatic bile ducts by lymphocytes is regulated by IFN-gamma in experimental biliary atresia. *J Clin Invest* 114: 322-329

Villarino AV, Artis D, Bezbradica JS, Miller O, Saris CJ, Joyce S, Hunter CA (2008) IL-27R deficiency delays the onset of colitis and protects from helminth-induced pathology in a model of chronic IBD. *Int Immunol* 20: 739-752