Supplementary Methods

Primers used for sAC mt-sAC amplification:

Forward: 5'-3' CGC<u>ACCGGT</u>ATGAGTGCCCGAAGGCAG (*AgeI* restriction site underlined);

Reverse: 5'-3'CGCGCGGCCGCCTAAGCGTAGTCTGGGACG

TCGTATGGGTAGACTTTCTCATTGAGGC (*NotI* restriction site underlined, HA sequence in bold).

Primers used in RT-PCR for mtDNA quantification: COXI Forward: 5'-3'CTAACAGACCGCAACCT COXI Reverse: 5'-3'GTGGCGAGTCAGCTAA 28sRNA Forward: 5'-3' ATCCTTCGATGTCGGC 28sRNA Reverse: 5'-3' AGCACATACACCAAATGTCT

Primers used in RT-PCR to quantify mRNA expression:

PGC1α Forward: 5'-3' GGCACGCAATCCTATT

PGC1a Reverse: 5'-3' GCTGTAGGGCGATCTT

NRF-1 Forward: 5'-3'AGGTAACCGAAGTATTAGGG

NRF-1 Reverse: 5'-3' CCTTCCGCGTACTGAC

GAPDH Forward: 5'-3'AGAGCTGAACGGGAAG

GAPDH Reverse: 5'-3' GTTGAAGTCGCAGGAG

Supplementary Figures



Supplementary Figure 1. Cyt c transcription regulation by PKA and ROS depends on NRF1-CREB

Luciferase activity in WT and CA75 cells transfected with Cyt c-LUC 66, whose promoter lacks the NRF-1 and CREB binding sites, after 48 hrs treatment with 8BrcAMP, H89, NAC, or H89 plus NAC. Luminescence was normalized by the number of cells (RLU/cell ratio). Values are shown as a percentage of untreated controls for each cell line and represent the mean of three independent transfections, each measured in triplicates.



Supplementary Figure 2. Cytoplasmic cAMP does not affect ROS-dependent OXPHOS biogenesis

(A) ROS production (n=12), (B) mtDNA content (n=9), and (C) cytrate synthase (CS) activity (n=6) were significantly changed in cells treated with membrane permeant 8Br-cAMP for 48 hrs, but not in cells treated with forskolin-IBMX (fsk-IBMX). *, p<0.01; **, p<0.001. (D) Western blot analyses of Cyt c and COXIV in homogenates from cells treated with fsk-IBMX for 48 hrs. Re-probing the membrane with GAPDH showed that comparable amount of protein was loaded in the gel.



Supplementary Figure 3. Effects of sAC transient expression on OXPHOS biogenesis

(A) Expression of sAC detected with the R21 antibody in transiently transfected 293T HEK cells. Steady state levels of mitochondrial proteins COXIV and Cyt c were reduced in sAC expressing cells in comparison to untransfected (Unt) or mock transfected cells. GAPDH is used as protein loading control. (B) MtDNA content (n=6) and (C) cytrate synthase (CS) activity (n=9) in 293T HEK cells transiently expressing sAC and mock transfected cells. Values are shown as a percentage of Unt cells. **, p<0.001; ***, p<0.0001.



Supplementary Figure 4. Effect of 8Br-cAMP on DT glu/gal in WT and COX deficient cells

Treatment with 8Br-cAMP impairs growth in galactose of WT cells, whereas CA75 cells, which do not grow in galactose, were unaffected by 8Br-cAMP (n=3). ***, p<0.0001.



Supplementary Figure 5. Adenylyl cyclase activity and sensitivity to KH7 in COXI mutant cells stably expressing mt-sAC

(A) cAMP levels measured in total cell homogenate (T), and the pellet fraction containing mitochondrial (P) in mt-sAC CA75 expressing cells and mock transfected cells (n=3). (B) KH7 suppresses the increase of COX enzymatic activity in CA75 cells induced by mt-sAC expression (n=5). *, p<0.01; ***, p<0.0001.