

Baibakov et al., http://www.jcb.org/cgi/content/full/jcb.201203062/DC1

Figure S1. **Transgenic mice expressing human ZP1.** (A) Schematic of a 11.9-kbp human ZP1 transgene composed of a 2.2-kbp promoter, 8.2-kbp coding region, and 1.5-kbp 3' of the last exon. Exons are indicated by numbers. Arrowheads represent exon-spanning forward (F) and reverse (R) PCR primers used for genotyping. (B) In situ hybridization of human ZP1 transgenic mouse ovaries. Specific  $^{35}$ S-labeled sense and antisense human ZP1 cRNA probes were hybridized to Tissue-Tek OCT-embedded ovarian sections from 15-d-old human ZP1 transgenic females. Sections were viewed with brightand dark-field microscopy. Insets are magnifications. (C) Plastic-embedded ovarian sections (5 µm) from 3–4-wk-old normal, human ZP1 transgenic, mouse Zp1-null, and human ZP1 rescue female mice were stained with periodic acid Schiff's reagent to highlight the zona pellucida (arrows) and counterstained with hematoxylin. (D) DIC microscopy of human ZP1 rescue and normal ovulated eggs. (E) Immunoblot of human and mouse eggs. Lane 1, noninseminated immature human oocytes (n = 6); lane 2, huZP4 transgenic eggs (n = 30); lane 3, huZP1 transgenic eggs (n = 30). The blot was probed with a monoclonal antibody specific to human ZP1, which was detected with HRP-conjugated secondary antibodies and chemiluminescence. Molecular masses are indicated to the left.

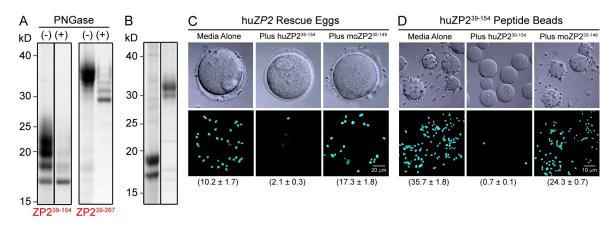


Figure S2. Human and mouse ZP2 peptides. (A) Coomassie blue-stained SDS-PAGE of recombinant huZP2<sup>39-154</sup> and huZP2<sup>39-267</sup> peptides expressed in High Five insect cells before (-) and after (+) deglycosylation with PNGase F to release three (N<sup>89</sup>, N<sup>105</sup>, and N<sup>122</sup>) or four (N<sup>89</sup>, N<sup>105</sup>, N<sup>122</sup>, and N<sup>223</sup>) Nglycans, respectively. (B) Coomassie blue-stained SDS-PAGE of recombinant moZP2<sup>35-149</sup> (left) and moZP2<sup>35-262</sup> (right) peptides after purification on IMAC beads. (C) DIC (top) and confocal (bottom) images after staining with Hoechst. Capacitated human sperm binding to huZP2 rescue eggs (left) was inhibited by coincubation with huZP2<sup>39-154</sup> (middle), but not moZP2<sup>35-149</sup> (right), peptides. Number (mean  $\pm$  SEM) of bound sperm from three independent experiments, each with seven to eight eggs. (D) Same as C but with 18–27 huZP2<sup>39-154</sup> peptide beads.

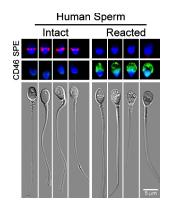


Figure S3. Acrosome status of human sperm. Acrosome-intact and -reacted human sperm bound antibodies to SPESP1 (SPE; top) and CD46 (middle), respectively. Sperm, stained with Hoechst and antibody, were imaged by confocal microscopy (top and middle) and DIC (bottom). Bar, 5 µm.

Table S1.	Fertility of human ZP1 transgenic and rescue mi	ce
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0	Litter size
Ovulated eggs	
29.0 ± 7.5	7.8 ± 0.7
34.3 ± 2.8	6.3 ± 1.5
26.5 ± 3.7	6.7 ± 1.0
	29.0 ± 7.5 34.3 ± 2.8

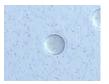
Values are given as means ± SEM.



Video 1. **Human sperm interacting with huZP1 rescue eggs.** Human sperm were incubated with huZP1 rescue eggs for 4 h at 37°C in HTF and imaged for 1 min at 30 frames/s with a charge-coupled device (CCD) camera (KP-D20A/B) mounted on a stereomicroscope (SMZ-U; Nikon) using Studio II Plus software (Pinnacle) and exported as MPEG files.



Video 2. Human sperm interacting with huZP2 rescue eggs. Human sperm were incubated with huZP2 rescue eggs for 4 h at 37°C in HTF and imaged for 1 min at 30 frames/s with a CCD camera (KP-D20A/B) mounted on a stereomicroscope (SMZ-U) using Studio II Plus software and exported as MPEG files.



Video 3. **Human sperm interacting with huZP3 rescue eggs.** Human sperm were incubated with huZP3 rescue eggs for 4 h at 37°C in HTF and imaged for 1 min at 30 frames/s with a CCD camera (KP-D20A/B) mounted on a stereomicroscope (SMZ-U) using Studio II Plus software and exported as MPEG files.



Video 4. **Human sperm interacting with huZP4 transgenic eggs.** Human sperm were incubated with huZP4 transgenic eggs for 4 h at 37°C in HTF and imaged for 1 min at 30 frames/s with a CCD camera (KP-D20A/B) mounted on a stereomicroscope (SMZ-U) using Studio II Plus software and exported as MPEG files.