Supplemental materials for:

Pt(II)-Catalyzed Synthesis of 1,2-Dihydropyridines from Aziridinyl Propargylic Esters

Massoud Motamed, Eric M. Bunnelle, Surendra W. Singaram and Richmond Sarpong* Department of Chemistry, University of California, Berkeley, California 94720

Email: rsarpong@berkeley.edu

Table of contents:

Materials and Methods	2
Representative Procedure for the Formation of Aziridines	3
Representative Procedure for the Formation of Azyridinyl Aldehydes	5
Representative Procedure for the Formation of Azyridinyl Propargylic Esters	7
Representative Procedure for the Formation of Dihydropyridines	17
Representative Procedure for the Formation of Hydroxypyridines	26
References	27
HPLC Traces	28
ORTEP Structure and Refinement Data	31
Selected NMR spectra	33

Materials and Methods. All air or moisture sensitive reactions were conducted in flame-dried glassware under an atmosphere of nitrogen using dry, deoxygenated solvents. Toluene, methylene chloride, and acetonitrile were distilled under nitrogen from calcium hydride immediately prior to use and tetrahydrofuran (THF) was distilled under nitrogen from sodium benzophenone ketyl. Alumina was washed with water (3 mL per 100 g) prior to use. Dess-Martin periodinane was prepared according to literature precedent.¹ PtCl₂ was purchased from Strem or obtained by donation from Johnson Matthey. All other reagents were purchased from Aldrich, Acros, or Lancaster and used without further purification. Reaction temperatures were controlled by an IKAmag[®] temperature modulator. Thin-layer chromatography (TLC) was performed using E. Merck silica gel 60 F254 precoated plates (0.25 mm) and visualized by UV indication and anisaldehyde stain. Fisher silica gel 240-400 mesh (particle size 0.032-0.063) and Aldrich neutral alumina oxide 150 mesh were used for flash chromatography. ¹H and ¹³C NMR spectra were recorded on a Bruker AV-500 (at 500 MHz (¹H) and 125 MHz (¹³C)), on a Bruker DRX-500 (at 500 MHz (¹H) and 125 MHz (¹³C)), or on a Bruker AVB-400 (at 400 MHz (¹H) and 100 MHz (¹³C)) in chloroform-d, benzene- d_6 or methanol- d_4 at 23 °C. Chemical shifts were referenced to the residual chloroform-H peak, which was set at 7.26 ppm for ¹H and 77.0 ppm (center peak) for ¹³C spectra; to the residual methanol-H peak, which was set at 3.34 ppm for ¹H and 49.9 ppm (center peak) for ¹³C spectra; or to the residual benzene-H peak, which was set at 7.15 ppm for ¹H and 128.6 ppm (center peak) for ¹³C spectra. Data for ¹H NMR are reported as follows: chemical shifts (δ ppm), multiplicity, (s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, dt = doublet of triplets, dq = doublet of quartets, qd = quartet of doublets, m = multiplet, br = broad resonance), coupling constants (Hz) and integration. Data for ¹³C NMR are reported in terms of chemical shift. IR spectra were recorded on a Nicolet MAGNA-IR 850 spectrometer and are reported in frequency of absorption (cm⁻¹). Low and high resolution mass spectral data were obtained from the University of California, Berkeley Mass Spectral Facility, on a VG 70-Se Micromass spectrometer for FAB (120 kV), and a VG Prospec Micromass spectrometer for EI (70 eV). Enantiomeric excesses were obtained utilizing a Shimatzu 10 A VP series chiral HPLC. Polarization data was obtained on a Perkin-Elmer 241 Polarimeter with a 59 nm sodium lamp.

Procedure for Synthesis of Propargylic Esters:



S1

Representative Procedure A for the Formation of Aziridines:²

(7-tosyl-7-aza-bicyclo[4.1.0]heptan-1-yl)methanol (S1). Cyclohexenylmethanol was synthesized by a known method.³ A flame-dried 50 mL round-bottom flask equipped with a stir bar was charged with cyclohexenylmethanol (1 g, 9.3 mmol, 1 equiv) and acetonitrile (30 mL). To the flask, anhydrous Chloramine-T (2.2 g, 9.7 mmol, 1.05 equiv) was added. To the stirring cloudy white mixture, *N*-bromosuccinimide (329 mg, 1.85 mmol, 0.2 equiv) was added. The resulting cloudy yellow mixture was stirred overnight at room temperature, and the solids were removed by filtration. The resulting yellow solution was concentrated under reduced pressure (rotary evaporation) and then

purified by column chromatography (100 mL silica gel, 4:1 hexanes:ethyl acetate) to yield 1.86 g (58%) of **S1** as a white semi-solid. **R**_f = 0.23 (4:1 hexanes:ethyl acetate); ¹**H NMR** (500 MHz, CDCl₃) δ ppm 7.82 (d, *J* = 8.3 Hz, 2H), 7.32 (d, *J* = 8.1 Hz, 2H), 4.05 (m, 2H), 3.24 (dd, *J* = 5.0, 0.7 Hz, 1H), 3.02 (s, 1H), 2.44 (s, 3H), 2.26 (td, *J* = 12.0, 5.8, 5.8 Hz, 1H), 1.79 (m, 2H), 1.65 (m, 2H), 1.35 (m, 3H), 1.23-1.14 (m, 1H); ¹³**C NMR** (125 MHz, CDCl₃) δ ppm 143.8, 138.1, 129.5, 126.8, 65.8, 56.4, 45.4, 27.4, 22.7, 21.5, 19.6, 19.3; **IR** (film) 3424, 2940, 1644 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₁₄H₁₉NO₃S]⁺: *m/z* 282.1164, found 282.1161.



S2

(2-methyl-3-phenyl-1-tosylaziridin-2-yl)methanol (S2). 2-methyl-3-phenylprop-2-en-1-ol, the aziridine precursor, was purchased from Aldrich. Following procedure A, a yellow gel was obtained in 82% yield following purification by column chromatography (silica gel, 6:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.18$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, CDCl₃) δ ppm 7.87 (d, J = 8.3 Hz, 2H), 7.33 (d, J = 8.1 Hz, 2H), 7.25 (dd, J= 6.4, 2.9 Hz, 3H), 7.03 (dd, J = 6.5, 3.0 Hz, 2H), 4.24 (s, 1H), 4.18 (m, 2H), 3.20 (m, 1H), 2.45 (s, 3H), 1.18 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ ppm 144.2, 137.4, 133.0, 129.6, 128.3, 127.8, 127.1, 126.9, 65.5, 59.2, 51.3, 21.6, 16.0; **IR** (film) 3441, 1644 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₁₇H₁₉NO₃S]⁺: *m/z* 318.1164, found 318.1162.



(3-isobutyl-2-isopropyl-1-tosylaziridin-2-yl)methanol (S3). 2-isopropyl-5-methyl-2hexenol, the aziridine precursor, was obtained by reduction of the corresponding aldehyde using a known procedure⁴ and was utilized crude. Following procedure A, a yellow gel was obtained in 48% yield following purification by column chromatography (silica gel, 6:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.36$ (4:1 hexanes:ethyl acetate); ¹H NMR (400 MHz, CDCl₃) δ ppm 7.88 (d, J = 8.3 Hz, 2H), 7.36 (d, J = 8.0 Hz, 2H), 4.41 (dd, J= 14.0, 9.0 Hz, 1H), 3.93 (dd, J = 14.0, 6.1 Hz, 1H), 3.25 (m, 2H), 2.49 (s, 3H), 1.69 (td, J = 14.1, 7.1, 7.1 Hz, 1H), 1.63 (s, 1H), 1.49-1.35 (m, 2H), 1.24 (d, J = 7.1 Hz, 3H), 1.08 (d, J = 7.1 Hz, 3H), 0.95 (d, J = 6.2 Hz, 3H), 0.84 (d, J = 6.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ ppm 143.9, 137.6, 129.4, 127.2, 63.4, 61.1, 49.4, 35.5, 31.4, 26.9, 22.9, 21.9, 21.5, 19.7, 18.0; IR (film) 3519, 2961, 1599 cm⁻¹; HRMS (FAB⁺) calc'd for [C₁₇H₂₇NO₃S]⁺: *m/z* 326.1802, found 326.1800.

Representative Procedure B for the Formation of Aziridinyl Aldehydes:



7-tosyl-7-aza-bicyclo[4.1.0]heptane-1-carbaldehyde (S4). A flame-dried 50 mL roundbottom flask equipped with a stir bar was charged with **S1** (1.5 g, 4.2 mmol, 1 equiv) and

methylene chloride (15 mL). The resulting clear solution was cooled to 0 °C and then stirred for an additional 10 min. Sodium bicarbonate (689 mg, 8.4 mmol, 2 equiv) and Dess-Martin periodinane (1.9 g, 4.6 mmol, 1.1 equiv) were then added. The resulting cloudy white mixture was then warmed to room temperature and until all the starting material was consumed as judged by TLC analysis (1 h). The white mixture was washed with sat. aq. sodium bicarbonate and extracted with methylene chloride (2 x 15 mL). The combined organic extracts were dried over MgSO₄ and concentrated under reduced pressure (rotary evaporation). The crude white oil was purified by column chromatography (150 mL silica gel, 4:1 hexanes:ethyl acetate) to yield 1.2 g (80%) of S4 as a white semi-solid. $\mathbf{R}_{\mathbf{f}} = 0.35$ (4:1 hexanes:ethyl acetate); ¹H NMR (400 MHz, CDCl₃) δ ppm 9.53 (s, 1H), 7.83 (d, J = 8.3 Hz, 2H), 7.34 (d, J = 8.0 Hz, 2H), 3.70 (dd, J= 3.9, 2.1 Hz, 1H), 2.51-2.41 (m, 4H), 1.87-1.73 (m, 3H), 1.48-1.38 (m, 1H), 1.38-1.29 (m, 1H), 1.28-1.14 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ ppm 194.7, 144.5, 136.9, 129.8, 127.2, 56.9, 46.2, 23.0, 21.6, 21.15, 19.07, 19.05; **IR** (film) 3423, 1643 cm⁻¹; **HRMS** (FAB⁺) calc'd for $[C_{14}H_{17}NO_3S]^+$: *m/z* 280.1007, found 280.1007.



2-methyl-3-phenyl-1-tosylaziridine-2-carbaldehyde (S5). Obtained following Procedure B, a white semi-solid was obtained in quantitative yield following purification by column chromatography (silica gel, 6:1 hexanes:ethyl acetate). $\mathbf{R}_{f} = 0.38$ (4:1 hexanes:ethyl acetate); ¹H NMR (400 MHz, CDCl₃) δ ppm 9.76 (s, 1H), 7.93 (d, J = 8.2Hz, 2H), 7.40 (d, J = 8.1 Hz, 2H), 7.37-7.27 (m, 3H), 7.16-7.09 (m, 2H), 4.73 (s, 1H), 2.51 (s, 3H), 1.21 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ ppm 194.1, 144.8, 136.2, 131.1, 129.8, 128.6, 128.6, 127.4, 127.1, 59.8, 52.1, 21.6, 11.3; **IR** (film) 3064, 1721, 1598 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₁₇H₁₇NO₃S]⁺: *m/z* 316.1007, found 316.1004.



3-isobutyl-2-isopropyl-1-tosylaziridine-2-carbaldehyde (S6). Following procedure B, the product was obtained and the crude material was taken on directly.

Representative Procedure C for the Formation of Aziridinyl Propargylic Esters:



1a

1-(2-methyl-3-phenyl-1-tosylaziridin-2-yl)prop-2-ynyl acetate (1a). A flame-dried 50 mL round-bottom flask equipped with a stir bar was charged with **S5** (500 mg, 1.6 mmol, 1 equiv) and THF (7 mL). To the stirred clear solution, cooled to -78 °C was added ethynylmagnesium bromide (0.5M in THF, 9.5 mL, 4.7 mmol, 3 equiv). After stirring at room temperature for 1 h, acetyl chloride (0.41 mL, 5.3 mmol, 3.3 equiv) was added dropwise over 10 min to the orange solution. The resulting red solution was stirred until all the starting material was consumed as judged by TLC analysis (20 min) and

subsequently quenched with sat. aqueous NH₄Cl (15 mL) and extracted with ethyl acetate (2 x 7mL). The combined organic extracts were dried over MgSO₄ and concentrated under reduced pressure (rotary evaporation). The crude yellow oil was purified by column chromatography (50 mL silica gel, 4:1 hexanes:ethyl acetate) to yield 498 mg (81% yield) of **1a** as a white gel. **R**_f = 0.41 (4:1 hexanes:ethyl acetate); ¹**H NMR** (500 MHz, CDCl₃) δ ppm 7.86 (d, *J* = 8.2 Hz, 2H), 7.31 (d, *J* = 8.1 Hz, 2H), 7.28-7.22 (m, 3H), 7.04 (dd, *J* = 6.5, 2.6 Hz, 2H), 6.07 (d, *J* = 2.0 Hz, 1H), 4.16 (s, 1H), 2.62 (d, *J* = 2.1 Hz, 1H), 2.43 (s, 3H), 2.13 (s, 3H), 1.28 (s, 3H); ¹³**C NMR** (125 MHz, CDCl₃) δ ppm 169.2, 144.4, 137.0, 132.1, 129.6, 128.4, 128.1, 127.5, 127.0, 78.7, 75.1, 64.9, 56.5, 50.4, 21.6, 20.7, 14.0; **IR** (film) 3279, 2983, 1752 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₂₁H₂₁NO₄S]⁺: *m/z* 384.1270, found 384.1279.

Enriched sample: $[\alpha]_D^{23} = -12.7^\circ$ (c = 0.7 in CHCl₃). A racemic sample of 1a was resolved using a HPLC system with Rainin SD-1 pumps, Sonntek UV detector and a 2 X 25 cm Chiraplak OD-H column (using a solvent system of 20% ethanol in hexane at 3mL/min). Analysis of enantiomers by chiral HPLC (Chiraplak OD-H, 20% ethanol in hexane at 3mL/min, T_r major 3.68, minor 2.52 at > 99.9% ee. Enantiomeric excess was determined using HPLC analysis, which was conducted by Ms. Christina Kraml (AccelaPure Corp., Princeton, NJ, USA). HPLC traces are provided on pages 27-28.



1b

1-(2-methyl-3-phenyl-1-tosylaziridin-2-yl)prop-2-ynyl 4-chlorobenzoate (1b). Following procedure C, a clear semi-solid was obtained in 75% yield following purification by column chromatography (silica gel, 4:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.47$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, CDCl₃) δ ppm 7.94 (d, J = 8.4 Hz, 2H), 7.82 (d, J = 8.1 Hz, 2H), 7.42 (d, J = 8.4 Hz, 2H), 7.34-7.30 (m, 3H), 7.22-7.15 (m, 4H), 6.30 (d, J = 1.4 Hz, 1H), 4.28 (s, 1H), 2.68 (d, J = 1.7 Hz, 1H), 2.34 (s, 3H), 1.39 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ ppm 164.2, 144.9, 140.2, 137.1, 132.537, 131.8, 130.0, 129.1, 128.9, 128.7, 128.4, 127.9, 127.6, 78.8, 76.1, 65.8, 56.6, 50.2, 22.0, 14.6; **IR** (film) 3284, 2980, 1733 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₂₆H₂₂ClNO₄S]⁺: *m/z* 480.1045, found 480.1043.



1c

1-(2-methyl-3-phenyl-1-tosylaziridin-2-yl)prop-2-ynyl pivalate (1c). Following procedure C, a clear oil was obtained in 81% yield following purification by column chromatography (silica gel, 6:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.54$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, CDCl₃) δ ppm 7.87 (d, J = 8.3 Hz, 2H), 7.30 (d, J = 8.0 Hz, 2H), 7.27-7.24 (m, 3H), 7.03 (dd, J = 6.4, 2.6 Hz, 2H), 6.08 (d, J = 2.0 Hz, 1H), 4.15 (s, 1H), 2.60 (d, J = 2.1 Hz, 1H), 2.43 (s, 3H), 1.30 (s, 12H); ¹³C NMR (125 MHz,

CDCl₃) δ ppm 176.3, 144.3, 137.1, 132.2, 129.6, 128.3, 128.0, 127.5, 126.9, 78.9, 74.6, 64.8, 56.8, 50.3, 38.9, 27.0, 21.6, 14.0; **IR** (film) 3279, 2978, 1742 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₂₄H₂₇NO₄S]⁺: *m/z* 426.1751, found 426.1747.



1d

1-(2-methyl-3-phenyl-1-tosylaziridin-2-yl)prop-2-ynyl benzoate (1d). Following procedure C, a clear viscous oil was obtained in 79% yield following purification by column chromatography (silica gel, 4:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.49$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, CDCl₃) δ ppm 7.98 (d, J = 7.3 Hz, 2H), 7.81 (d, J = 8.2 Hz, 2H), 7.59 (dd, J = 7.4, 7.4 Hz, 1H), 7.44 (dd, J = 7.8, 7.8 Hz, 2H), 7.35-7.30 (m, 3H), 7.25-7.21 (m, 2H), 7.12 (d, J = 8.1 Hz, 2H), 6.31 (d, J = 1.9 Hz, 1H), 4.30 (s, 1H), 2.66 (d, J = 2.0 Hz, 1H), 2.28 (s, 3H), 1.39 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ ppm 164.5, 144.3, 136.5, 133.2, 132.1, 129.9, 129.5, 129.4, 128.4, 128.2, 128.1, 127.4, 127.2, 78.6, 75.3, 64.9, 56.1, 49.4, 21.5, 14.1; **IR** (film) 3284, 1731 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₂₆H₂₃NO₄S]⁺: *m/z* 446.1426, found 446.1430.



1-(3-isobutyl-2-isopropyl-1-tosylaziridin-2-yl)prop-2-ynyl acetate (12c). Following procedure C, a white foam was obtained in 75% yield after purification by column chromatography (silica gel, 6:1 hexanes:ethyl acetate). **R**_f = 0.62 (4:1 hexanes:ethyl acetate); ¹**H NMR** (500 MHz, C₆D₆) δ ppm 7.85 (d, J = 8.2 Hz, 2H), 6.66 (d, J = 8.1 Hz, 2H), 6.56 (d, J = 2.3 Hz, 1H), 3.15 (dd, J = 7.0, 5.8 Hz, 1H), 2.02 (d, J = 2.4 Hz, 1H), 1.88 (td, J = 14.4, 7.1, 7.1 Hz, 1H), 1.77 (s, 3H), 1.76 (s, 3H), 1.47 (d, J = 7.1 Hz, 3H), 1.39-1.31 (m, 4H), 1.24-1.19 (m, 2H), 0.75 (d, J = 6.6 Hz, 3H), 0.66 (d, J = 6.6 Hz, 3H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 167.8, 143.2, 138.4, 129.0, 127.6, 127.6, 78.0, 75.4, 65.2, 59.9, 50.0, 35.5, 32.6, 26.7, 22.5, 21.8, 21.2, 20.7, 19.5; **IR** (film) 3268, 2963, 1752 cm⁻¹; **HRMS** (EI) calc'd for [C₂₁H₂₉NO₄S]⁺: *m/z* 392.1851, found 392.1852.



1-(7-tosyl-7-aza-bicyclo[4.1.0]heptan-1-yl)prop-2-ynyl acetate (14). Following

procedure C, a yellow gel was obtained in 69% yield after purification by column chromatography (silica gel, 4:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.20$ (4:1 hexanes:ethyl acetate); ¹H NMR (400 MHz, CDCl₃) δ ppm 7.80 (d, J = 8.3 Hz, 2H), 7.30 (d, J = 8.0 Hz, 2H), 5.87 (d, J = 2.2 Hz, 1H), 3.16 (dd, J = 4.8, 1.0 Hz, 1H), 2.54 (d, J = 2.2 Hz, 1H), 2.43 (s, 3H), 2.32-2.22 (m, 1H), 2.12-2.06 (m, 1H), 2.04 (s, 3H), 1.87-1.76 (m, 1H), 1.68-1.59 (m, 2H), 1.54-1.41 (m, 1H), 1.40-1.20 (m, 3H); ¹³C NMR (125 MHz, CDCl₃) δ ppm 169.2, 144.0, 137.8, 129.6, 127.2, 78.9, 74.9, 65.1, 54.0, 44.0, 25.0, 22.6, 21.6, 20.7, 19.8, 19.3; IR (film) 3424, 2942, 1750,1644 cm⁻¹; HRMS (FAB⁺) calc'd for [C₁₈H₂₁NO₄S]⁺: *m/z* 348.1262, found 348.1261.

Representative Procedure D for the Formation of Aziridinyl Propargylic Esters:



12a

3-cyclopropyl-1-(3-isobutyl-2-isopropyl-1-tosylaziridin-2-yl)prop-2-ynyl acetate (12a). A flame-dried 50 mL round-bottom flask equipped with a stir bar was charged with ethynylcyclopropane (197 mg, 3 mmol, 1.3 equiv) and THF (20 mL). To the stirred clear solution, cooled to -78 °C, was added butyllithium (2.5M in hexanes, 1 mL, 2.5 mmol, 1.1 equiv) dropwise over 15 min, and the resulting mixture was stirred for an additional 10 min. To the faint yellow solution, **S6** (750 mg, 2.3 mmol, 1 equiv) in THF

(1 mL) was added dropwise over 10 min. The yellow solution was stirred at 23 °C for 2 h, then acetyl chloride (0.30 mL, 4.6 mmol, 2 equiv) was added dropwise over 10 min and then stirred until all the starting material was consumed as judged by TLC analysis (20 min). The reaction was then quenched with sat. aqueous NH₄Cl (15 mL) and extracted with ethyl acetate (2 x 7mL). The combined organic extracts were dried over MgSO₄ and concentrated under reduced pressure (rotary evaporation). The crude red oil was purified by column chromatography (50 mL silica gel, 6:1 hexanes:ethyl acetate) to yield 725 mg (73 % yield) of a yellow gel. $\mathbf{R}_{f} = 0.43$ (4:1 hexanes:ethyl acetate); ¹H **NMR** (500 MHz, C_6D_6) δ ppm 7.87 (d, J = 8.3 Hz, 2H), 6.68 (d, J = 8.0 Hz, 2H), 6.55 (d, J = 1.5 Hz, 1H), 3.18 (dd, J = 6.5, 6.5 Hz, 1H), 1.91 (td, J = 14.3, 7.2, 7.2 Hz, 1H),1.80 (s, 3H), 1.78 (s, 3H), 1.50 (d, J = 7.1 Hz, 3H), 1.42-1.38 (m, 1H), 1.36 (d, J = 7.3Hz, 3H), 1.27-1.23 (m, 2H), 0.90 (dqd, *J* = 8.2, 5.0, 5.0, 5.0, 2.0 Hz, 1H), 0.76 (d, *J* = 6.6 Hz, 3H), 0.68 (d, J = 6.6 Hz, 3H), 0.50 (m, 2H), 0.26 (m, 2H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 168.1, 143.2, 138.7, 129.1, 127.6, 90.7, 72.4, 66.0, 60.8, 50.1, 35.6, 32.6, 26.8, 22.5, 21.9, 21.3, 20.8, 20.3, 19.7, 7.7, 7.7, -0.4; **IR** (film) 2962, 1754 cm⁻¹; HRMS (FAB^+) calc'd for $[C_{24}H_{33}NO_4S]^+$: m/z 432.2209, found 432.2208.



1-(3-isobutyl-2-isopropyl-1-tosylaziridin-2-yl)-3-phenylprop-2-ynyl acetate (12b).

Following procedure D, a clear semi-solid was obtained in 70% yield after purification by column chromatography (silica gel, 8:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.55$ (4:1 hexanes:ethyl acetate); ¹**H NMR** (400 MHz, C₆D₆) δ ppm 7.92 (d, J = 8.3 Hz, 2H), 7.40-7.33 (m, 2H), 6.93-6.89 (m, 3H), 6.86 (s, 1H), 6.72 (d, J = 8.0 Hz, 2H), 3.30 (dd, J = 6.5, 6.5 Hz, 1H), 2.00 (td, J = 14.2, 7.1, 7.1 Hz, 1H), 1.86 (s, 3H), 1.81 (s, 3H), 1.59 (d, J = 7.0 Hz, 3H), 1.49 (d, J = 7.2 Hz, 3H), 1.42 (td, J = 13.1, 6.6, 6.6 Hz, 1H), 1.33-1.26 (m, 2H), 0.77 (d, J = 6.5 Hz, 3H), 0.70 (d, J = 6.6 Hz, 3H); ¹³**C NMR** (100 MHz, C₆D₆) δ ppm 168.0, 143.3, 138.5, 131.6, 129.1, 128.5, 128.2, 127.6, 122.3, 86.9, 86.0, 66.0, 60.5, 50.1, 35.5, 32.6, 26.8, 22.4, 21.8, 21.3; 20.734, 20.201, 19.615; **IR** (film) 2962, 1755, 1598 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₂₇H₃₃NO₄S]⁺: *m/z* 468.2209, found 468.2209.

Representative Procedure E for the Formation of Aziridinyl Propargylic Esters:



10

1-(2-methyl-3-phenyl-1-tosylaziridin-2-yl)prop-2-ynyl 2,2,2-trichloroacetate (10). A flame-dried 50 mL round-bottom flask equipped with a stir bar was charged with **S5** (500 mg, 1.6 mmol, 1 equiv) and THF (8 mL). To the stirred clear solution, cooled to -78 °C, was added ethynylmagnesium bromide (0.5M in THF, 9.6 mL, 4.7 mmol, 3 equiv). The resulting red solution was warmed to 23 °C and stirred for 1 h. To the solution, 2,2,2-trichloroacetonitrile (0.52 mL, 5.2 mmol, 3.3 equiv) was added dropwise over 10 min.

The black solution was stirred until all the starting material was consumed as judged by TLC analysis (6 h), and then quenched with water (15 mL) and extracted with ethyl acetate (2 x 7mL). The combined organic extracts were dried over MgSO₄ and concentrated under reduced pressure (rotary evaporation). The crude red product was used without further purification. $\mathbf{R}_{\mathbf{f}} = 0.33$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, C₆D₆) δ ppm 8.67 (s, 1H), 7.85 (d, J = 8.3 Hz, 2H), 7.06 (m, 2H), 6.97-6.89 (m, 3H), 6.80 (d, J = 2.0 Hz, 1H), 6.61 (d, J = 8.1 Hz, 2H), 4.23 (s, 1H), 2.01 (d, J = 2.1 Hz, 1H), 1.73 (s, 3H), 1.45 (s, 3H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 160.0, 143.6, 137.9, 132.5, 129.3, 128.2, 127.9, 127.0, 91.0, 78.2, 75.2, 69.1, 56.7, 49.8, 20.7, 13.67 (16 of 17 observed).

Representative Procedure F for the Formation of Aziridinyl Propargylic Esters:





1-(1-acetyl-3-isobutyl-2-isopropylaziridin-2-yl)prop-2-ynyl acetate (16a). A 20 mL oven-dried vial was charged with 12c (500 mg, 1.3 mmol, 1 equiv) and methanol (5 mL). Freshly ground magnesium (153 mg, 6.4 mmol, 5 equiv) was then added to the vial. The resulting clear mixture with the magnesium settled to the bottom was sonicated for 90 min at power 9 using a VWR model 75D sonicator to yield a cloudy green mixture without observable magnesium. This mixture was quenched with sat. aqueous NH₄Cl (20

mL) and extracted with ethyl acetate (3 x 20 mL). The combined organic extracts were dried over MgSO₄ and concentrated under reduced pressure (rotary evaporation). A 20 mL oven-dried vial, equipped with a stir bar, was charged with the crude amino alcohol and methylene chloride (5 mL). To the resulting solution was added acetic anhydride (0.29 mL, 3.1 mmol, 2.4 equiv), triethylamine (0.46 mL, 3.4 mmol, 2.6 equiv) and dimethylaminopyridine (DMAP) (5 mg). The pale red solution was stirred overnight, then guenched with sat. aqueous NH₄Cl (5 mL) and extracted with methylene chloride (2 x 5 mL). The combined organic extracts were dried over MgSO₄ and concentrated under reduced pressure (rotary evaporation). The crude clear oil was purified by column chromatography (50 mL silica gel, 8:1 hexanes:ethyl acetate) to yield 289 mg (81%) yield) of 16a as a clear oil. $R_f = 0.27$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, C_6D_6) δ ppm 6.00 (d, J = 2.3 Hz, 1H), 2.83 (dd, J = 7.9, 4.8 Hz, 1H), 1.98 (d, J = 2.3 Hz, 1H), 1.93 (s, 3H), 1.73 (m, 2H), 1.60 (s, 3H), 1.37-1.24 (m, 2H), 1.20 (dd, J = 7.8, 7.8Hz, 3H), 0.99-0.92 (m, 10H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 177.3, 168.0, 78.5, 76.6, 64.1, 50.9, 42.1, 36.4, 31.3, 27.3, 24.1, 22.5, 22.2, 19.8, 19.4, 18.7; **IR** (film) 3249, 2961, 2122,1751,1686 cm⁻¹; **HRMS** (EI) calc'd for $[C_{16}H_{25}NO_3]^+$: m/z 281.1933, found 281.1938.



16b

1-(1-benzoyl-3-isobutyl-2-isopropylaziridin-2-yl)prop-2-ynyl benzoate (16b). Following procedure F, a white foam was obtained in 78% yield following purification by column chromatography (silica gel, 9:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.84$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, C₆D₆) δ ppm 8.02 (dd, J = 7.9, 1.6 Hz, 2H), 7.92 (m, 2H), 7.01-6.96 (m, 1H), 6.94-6.84 (m, 5H), 5.98 (d, J = 2.3 Hz, 1H), 3.08 (dd, J = 7.2, 5.5 Hz, 1H), 2.14 (td, J = 14.3, 7.2, 7.2 Hz, 1H), 1.84-1.74 (m, 2H), 1.56-1.44 (m, 2H), 1.40 (d, J = 7.0 Hz, 3H), 1.20 (d, J = 7.3 Hz, 3H), 0.95 (dd, J = 6.6, 4.2 Hz, 6H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 176.8, 164.2, 135.6, 132.7, 131.5, 129.7, 129.4, 129.0, 128.0, 127.7, 78.8, 76.4, 65.8, 54.0, 43.2, 36.6, 31.7, 27.4, 22.5, 22.3, 20.0, 19.9; IR (film) 2959, 1727,1667 cm⁻¹; HRMS (EI) calc'd for [C₂₆H₂₉NO₃]⁺: *m/z* 404.2252, found 404.2252.

Representative Procedure G for the Formation of Dihydropyridines:



5-methyl-6-phenyl-1-tosyl-1,6-dihydropyridin-3-yl acetate (5a). A 4 mL oven-dried vial equipped with a stir bar was charged with PtCl₂ (6.2 mg, 0.023 mmol, 0.1 equiv). To the vial, **1a** (90 mg, 0.23 mmol, 1 equiv) and toluene (1.1 mL) were added. The vial was purged with nitrogen and then tightly sealed with a Teflon[®]-coated cap. The resulting pale yellow mixture was stirred at 100 °C until all the starting material was consumed as judged by TLC analysis (3 h). The resulting black mixture was concentrated under reduced pressure (rotary evaporation). Purification of the crude oil by column chromatography (20 mL neutral deactivated alumina, 16:1 hexanes:ethyl acetate) yielded

68 mg (76%) of a clear gel. **R**_f = 0.39 (4:1 hexanes:ethyl acetate); ¹**H NMR** (500 MHz, CDCl₃) δ ppm 7.72 (d, J = 7.5 Hz, 2H), 7.70-7.67 (m, 2H), 7.17-7.14 (m, 2H), 7.03 (dd, J = 7.3, 7.3 Hz, 1H), 6.69 (d, J = 7.9 Hz, 2H), 6.42 (s, 1H), 5.53-5.51 (m, 1H), 5.33 (s, 1H), 1.76 (s, 3H), 1.51 (s, 3H), 1.16 (s, 3H); ¹³**C NMR** (125 MHz, CDCl₃) δ ppm169.2, 143.7, 139.9, 136.7, 136.0 132.0, 129.4, 128.5, 128.5, 127.9, 126.6, 117.5, 112.1, 60.7, 21.6, 20.7, 20.71; **IR** (film) 3396, 2451, 1764 cm⁻¹; **HRMS** (FAB⁺) calc'd for $[C_{21}H_{21}NO_4S]^+$: *m/z* 384.1270, found 384.1279.

Enriched sample: $[\alpha]_D{}^{26} = -467.9^\circ$ (c = 0.7 in CHCl₃). Analysis of enantiomers by chiral HPLC (Chiraplak OD-H, 10% isopropanol in hexane at 1mL/30 min, T_r major 17.34, minor 8.67 at 99% ee. HPLC traces are provided on pages 28-29.



5b

5-methyl-6-phenyl-1-tosyl-1,6-dihydropyridin-3-yl 4-chlorobenzoate (5b). Following procedure G, a clear semi-solid was obtained in 72% yield following purification by column chromatography (20 mL neutral deactivated alumina, 16:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.56$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, C₆D₆) δ ppm 7.82 (d, J = 7.3 Hz, 2H), 7.77 (d, J = 8.3 Hz, 2H), 7.68 (m, 2H), 7.22 (dd, J = 7.7, 7.7 Hz, 2H), 7.11 (s, 1H), 6.94 (m, 2H), 6.76 (d, J = 8.0 Hz, 2H), 6.56 (s, 1H), 5.61 (s, 1H), 5.42 (m, 1H), 1.81 (s, 3H), 1.21 (s, 3H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 163.6, 143.1, 139.8, 139.7, 137.2, 136.9, 131.7, 131.2, 129.2, 128.6, 128.6, 128.5, 128.1, 126.7, 117.7, 113.0, 60.8, 20.7, 20.2 (19 of 20 observed); **IR** (film) 3399, 1736, 1730 cm⁻¹; **HRMS** (FAB⁺) calc'd for $[C_{26}H_{22}CINO_4S]^+$: *m/z* 480.1036, found 480.1038.



5-methyl-6-phenyl-1-tosyl-1,6-dihydropyridin-3-yl pivalate (5c). Following procedure G, a yellow gel was obtained in 76% yield after purification by column chromatography (20 mL neutral deactivated alumina, 25:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.67$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, CDCl₃) δ ppm 7.68 (d, J = 8.3 Hz, 2H), 7.53 (d, J = 6.7 Hz, 2H), 7.36-7.29 (m, 3H), 7.25 (s, 2H), 6.27 (s, 1H), 5.56 (s, 1H), 5.36 (s, 1H), 2.41 (s, 3H), 1.55 (s, 3H), 1.26 (s, 9H); \mathbf{C}^{13} NMR (125 MHz, CDCl₃) 177.1, 143.7, 139.8, 136.8, 136.1, 131.9, 129.5, 128.6, 128.5, 127.9, 126.7, 117.6, 111.9, 60.7, 38.9, 27.0, 21.6, 20.7; IR (film) 3419, 2974, 1751 cm⁻¹; HRMS (FAB⁺) calc'd for [C₂₄H₂₇NO₄S]⁺: *m/z* 425.1661, found 425.1669.



5d

5-methyl-6-phenyl-1-tosyl-1,6-dihydropyridin-3-yl benzoate (5d). Following procedure G, a clear viscous oil was obtained in 76% yield after purification by column chromatography (20 mL neutral deactivated alumina, 16:1 hexanes:ethyl acetate). $\mathbf{R_f} = 0.58$ (4:1 hexanes:ethyl acetate); ¹H NMR (400 MHz, C₆D₆) δ ppm 8.01 (dd, J = 8.3, 1.3

Hz, 2H), 7.84 (d, J = 7.4 Hz, 2H), 7.77 (d, J = 8.3 Hz, 2H), 7.21 (dd, J = 7.7, 7.7 Hz, 2H), 7.11-7.05 (m, 2H), 7.00-6.94 (m, 2H), 6.74 (d, J = 8.0 Hz, 2H), 6.58 (s, 1H), 5.61 (s, 1H), 5.44 (m 1H), 1.80 (s, 3H), 1.20 (s, 3H); ¹³C NMR (400 MHz, C₆D₆) δ ppm 164.5, 143.0, 140.0, 137.2, 136.9, 133.1, 131.6, 129.9, 129.3, 129.2, 128.6, 128.5, 128.2, 128.1, 126.8, 118.0, 112.9, 60.8, 20.7, 20.1; **IR** (film) 3415, 1737 cm⁻¹; **HRMS** (FAB⁺) calc'd for $[C_{26}H_{23}NO_4S]^+$: m/z 445.1348, found 445.1352.



11

2,2,2-trichloro-N-(5-methyl-6-phenyl-1-tosyl-1,6-dihydropyridin-3-yl)acetamide

(11). Following procedure G, a red gel was obtained in 56% yield after purification by column chromatography (20 mL neutral deactivated alumina, 16:1 hexanes:ethyl acetate). $\mathbf{R_f} = 0.33$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, C₆D₆) δ ppm 8.00 (d, J = 8.3 Hz, 2H), 7.10 (m, 2H), 7.02-6.94 (m, 3H), 6.77 (d, J = 8.0 Hz, 2H), 6.23 (dd, J = 2.6, 2.6 Hz, 1H), 5.28 (dd, J = 2.9, 1.4 Hz, 1H), 4.56 (dd, J = 2.3, 1.5 Hz, 1H), 4.17 (s, 1H), 1.83 (s, 3H), 0.84 (s, 3H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 167.1, 155.5, 144.0, 137.0, 132.3, 129.4, 128.3, 128.1, 127.9, 127.5, 104.0, 86.6, 85.8, 56.5, 48.0, 20.8, 11.6; **IR** (film) 3442, 1607 cm⁻¹; **HRMS** (FAB⁺) calc'd for $[C_{21}H_{19}Cl_3N_2O_3S]^+$: *m/z* 485.0244, found 485.0243.



13a

2-cyclopropyl-6-isobutyl-5-isopropyl-1-tosyl-1,6-dihydropyridin-3-yl acetate (13a). Following procedure G, a yellow oil was obtained in 62% yield after purification by column chromatography (20 mL neutral deactivated alumina, 16:1 hexanes:ethyl acetate). $\mathbf{R}_{f} = 0.51$ (4:1 hexanes:ethyl acetate); ¹H NMR (400 MHz, C₆D₆) δ ppm 8.05 (d, J = 8.2 Hz, 2H), 6.91 (d, J = 8.2 Hz, 2H), 5.17 (s, 1H), 4.70 (dd, J = 11.4, 2.1 Hz, 1H), 2.30-2.04 (m, 2H), 1.93 (s, 3H), 1.80 (s, 3H), 1.23 (d, J = 6.4 Hz, 3H), 1.09-0.97 (m, 4H), 0.94 (d, J = 6.8 Hz, 4H), 0.81 (d, J = 6.8 Hz, 4H), 0.65 (d, J = 6.9 Hz, 4H); ¹³C NMR (100 MHz, C₆D₆) δ ppm 167.4, 146.9, 143.5, 142.5, 137.4, 128.7, 128.6, 124.4, 113.7, 55.7, 38.9, 31.2, 23.4, 22.8, 21.0, 20.7, 20.7, 20.4, 19.8, 12.3, 7.6, 6.2; IR (film) 2959, 1764, 1348 cm⁻¹; HRMS (FAB⁺) calc'd for [C₂₄H₃₃NO₄S]⁺: *m/z* 431.2144, found 431.2138.



13b

6-isobutyl-5-isopropyl-2-phenyl-1-tosyl-1,6-dihydropyridin-3-yl acetate (13b). Following procedure G, a clear oil was obtained in 69% yield after purification by column chromatography (20 mL neutral deactivated alumina, 18:1 hexanes:ethyl acetate). $\mathbf{R}_{f} = 0.62$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, C₆D₆) δ ppm 7.76 (d, J = 7.8

Hz, 2H), 7.73 (d, J = 7.9 Hz, 2H), 7.25 (dd, J = 7.5, 7.5 Hz, 2H), 7.13-7.09 (m, 1H), 6.76 (d, J = 7.9 Hz, 2H), 5.35 (s, 1H), 4.97 (d, J = 10.4 Hz, 1H), 2.46-2.37 (m, 1H), 2.28 (m, 1H), 1.96 (m, 1H), 1.85 (s, 3H), 1.48 (s, 3H), 1.21 (d, J = 6.31 Hz, 3H), 1.10 (m, 1H), 0.96 (d, J = 6.7 Hz, 3H), 0.85 (d, J = 6.8 Hz, 3H), 0.77 (d, J = 6.8 Hz, 3H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 167.8, 148.5, 142.7, 141.3, 137.1, 135.3, 129.2, 128.7, 123.3, 114.3, 56.4, 39.4, 31.5, 23.6, 23.5, 21.2, 21.0, 20.7, 20.4, 19.8 (20 of 23 peaks observed); **IR** (film) 2960, 1765, 1350 cm⁻¹; **HRMS** (FAB⁺) calc'd for $[C_{27}H_{33}NO4S]^+$: *m/z* 467.2130, found 467.2137.



13c

6-isobutyl-5-isopropyl-1-tosyl-1,6-dihydropyridin-3-yl acetate (13c). Following procedure G, a white gel was obtained in 70% yield after purification by column chromatography (20 mL neutral deactivated alumina, 16:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.43$ (4:1 hexanes:ethyl acetate); ¹**H NMR** (500 MHz, C₆D₆) δ ppm 7.78 (d, J = 8.3 Hz, 2H), 6.70 (d, J = 8.0 Hz, 2H), 6.64 (dd, J = 1.1, 1.1 Hz, 1H), 5.26 (dd, J = 1.3, 1.3 Hz, 1H), 4.58 (m, 1H), 2.20 (dtdd, J = 13.2, 9.8, 6.6, 6.6, 3.1 Hz, 1H), 2.03 (ddd, J = 14.2, 11.1, 3.1 Hz, 1H), 1.83-1.77 (m, 1H), 1.76 (s, 3H), 1.57 (s, 3H), 1.14 (d, J = 6.4 Hz, 3H), 0.97-0.92 (m, 1H), 0.89 (d, J = 6.8 Hz, 3H), 0.66 (d, J = 6.8 Hz, 3H), 0.51 (d, J = 6.9 Hz, 3H); ¹³**C NMR** (125 MHz, C₆D₆) δ ppm 168.1, 144.5, 142.8, 141.1, 137.5, 129.1, 127.0, 113.1, 112.4, 54.7, 40.1, 31.3, 23.4, 23.1, 21.2, 21.1, 20.7, 20.2, 19.8; **IR** (film) 2961,

2870, 1766, 1598 cm⁻¹; **HRMS** (EI) calc'd for $[C_{21}H_{29}NO_4S]^+$: *m/z* 391.1739, found 391.1816.



15

1-tosyl-1,5,6,7,8,8a-hexahydroquinolin-3-yl acetate (15). Following procedure G, a white foam was obtained in 76% yield after purification by column chromatography (20 mL neutral deactivated alumina, 16:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.40$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, C₆D₆) δ ppm 7.89 (d, J = 8.0 Hz, 2H), 6.83 (s, 1H), 6.77 (d, J = 8.0 Hz, 2H), 5.11 (s, 1H), 4.42 (dd, J = 11.9, 3.9 Hz, 1H), 2.05 (m, 1H), 1.93 (ddd, J = 24.4, 12.2, 3.5 Hz, 1H), 1.76 (s, 3H), 1.72-1.66 (m, 1H), 1.59 (s, 3H), 1.46 (d, J = 15.4 Hz, 1H), 1.42-1.31 (m, 2H), 1.24 (m, 1H), 1.08-0.98 (m, 1H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 168.2, 143.2, 140.8, 136.6, 135.7, 129.6, 127.0, 113.6, 112.7, 59.9, 37.4, 35.0, 29.9, 25.8, 20.8, 19.9; **IR** (film) 3214, 2885, 1721 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₁₈H₂₁NO₄S]⁺: *m/z* 347.1185, found 347.1187.



1-acetyl-6-isobutyl-5-isopropyl-1,6-dihydropyridin-3-yl acetate (17a).

Following procedure G, a white foam was obtained in 71% yield after purification by column chromatography (20 mL neutral deactivated alumina, 16:1 hexanes:ethyl acetate).

 $\mathbf{R}_{f} = 0.73$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, C₆D₆) δ ppm 6.07 (s, 1H), 5.68 (s, 1H), 5.55 (dd, J = 10.9, 1.7 Hz, 1H), 2.11 (m, 1H), 2.03 (ddd, J = 14.1, 10.9, 3.4Hz, 1H), 1.71 (s, 3H), 1.63-1.61 (m, 4H), 1.16 (d, J = 6.4 Hz, 3H), 1.06 (ddd, J = 13.3, 10.1, 2.9 Hz, 1H), 0.97-0.89 (m, 9H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 168.654, 168.282, 147.154, 137.673, 112.966, 112.653, 49.199, 39.847, 31.730, 24.386, 23.516, 21.950, 21.739, 20.380, 20.001, 19.939; **IR** (film) 2962, 1754, 1681 cm⁻¹; **HRMS** (EI) calc'd for [C₁₆H₂₅NO₃]⁺: *m/z* 280.1854, found 280.1859.



17b

1-benzoyl-6-isobutyl-5-isopropyl-1,6-dihydropyridin-3-yl benzoate (17b). Following procedure G, an oil was obtained in 74% yield after purification by column chromatography (20 mL neutral deactivated alumina, 25:1 hexanes:ethyl acetate). **R**_f = 0.80 (4:1 hexanes:ethyl acetate); ¹**H NMR** (500 MHz, C₆D₆) δ ppm 8.02 (d, *J* = 7.5 Hz, 2H), 7.55 (d, *J* = 3.4 Hz, 2H), 7.04 (dd, *J* = 7.4, 7.4 Hz, 1H), 6.97-6.94 (m, 5H), 6.22 (s, 1H), 5.78 (s, 1H), 5.68 (d, *J* = 9.6 Hz, 1H), 2.27-2.20 (m, 1H), 2.15 (m, 1H), 1.91-1.82 (m, 1H), 1.29-1.21 (m, 4H), 0.98 (dd, *J* = 6.4, 6.4 Hz, 6H), 0.92 (d, *J* = 6.7 Hz, 3H); ¹³**C NMR** (125 MHz, C₆D₆) δ ppm 169.0, 164.7, 146.8, 137.1, 135.2, 133.0, 130.0, 129.9, 129.5, 128.6, 128.3, 128.0, 114.8, 113.4, 50.7, 39.9, 31.9, 24.8, 23.6, 22.3, 21.9, 20.1; **IR** (film) 2961, 1735, 1669 cm⁻¹; **HRMS** (EI) calc'd for [C₂₆H₂₉NO₃]⁺: *m/z* 404.2185, found 404.2187.



17c

6-isobutyl-5-isopropyl-1-(4-methylbenzoyl)-1,6-dihydropyridin-3-yl

4-methylbenzoate (17c). Following procedure G, clear crystals were obtained in 69% yield after purification by column chromatography (20 mL neutral deactivated alumina, 25:1 hexanes:ethyl acetate). $\mathbf{R_f} = 0.85$ (4:1 hexanes:ethyl acetate); **MP** 67 °C- 69 °C; ¹**H NMR** (500 MHz, C₆D₆) δ ppm 8.02 (d, J = 7.9 Hz, 2H), 7.54 (d, J = 7.2 Hz, 2H), 6.81 (dd, J = 8.1, 8.1 Hz, 4H), 6.33 (s, 1H), 5.81 (s, 1H), 5.66 (d, J = 9.5 Hz, 1H), 2.23 (m, 1H), 2.17 (m, 1H), 1.91 (d, J = 3.9 Hz, 6H), 1.29-1.20 (m, 4H), 0.97 (d, J = 6.3 Hz, 6H), 0.92 (d, J = 6.7 Hz, 4H); ¹³**C NMR** (125 MHz, C₆D₆) δ ppm 169.0, 164.8, 146.7, 143.9, 140.2, 137.0, 132.4, 130.0, 129.1, 128.8, 128.7, 126.9, 115.1, 113.6, 50.7, 39.9, 31.9, 24.8, 23.6, 22.4, 22.0, 21.0, 20.8, 20.1; **IR** (film) 3425, 1734, 1658 cm⁻¹; **HRMS** (FAB⁺) calc'd for [C₂₈H₃₃NO₃]⁺: *m/z* 432.2539, found 432.2539.



17d

1-(4-chlorobenzoyl)-6-isobutyl-5-isopropyl-1,6-dihydropyridin-3-yl 4chlorobenzoate (17d). Following procedure G, a yellow dense oil was obtained in 65% yield after purification by column chromatography (20 mL neutral deactivated alumina, 25:1 hexanes:ethyl acetate). $\mathbf{R}_{\mathbf{f}} = 0.85$ (4:1 hexanes:ethyl acetate); ¹H NMR (500 MHz, C₆D₆) δ ppm 7.74 (d, J = 8.5 Hz, 2H), 7.27 (d, J = 8.2 Hz, 2H), 6.90 (d, J = 8.4 Hz, 4H), 6.06 (s, 1H), 5.74 (s, 1H), 5.60 (d, J = 10.1 Hz, 1H), 2.21-2.08 (m, 2H), 1.85-1.76 (m, 1H), 1.25 (d, J = 6.2 Hz, 3H), 1.22-1.17 (m, 1H), 0.97 (dd, J = 7.1, 7.1 Hz, 6H), 0.92 (d, J = 6.7 Hz, 3H); ¹³C NMR (125 MHz, C₆D₆) δ ppm 167.8, 163.8, 147.4, 139.8, 137.2, 136.3, 133.2, 131.2, 130.1, 129.4, 128.7, 128.3, 114.5, 113.1, 50.8, 39.9, 34.6, 31.9, 24.8, 23.6, 21.9, 20.1; IR (film) 2960, 1738, 1666 cm⁻¹; HRMS (FAB⁺) calc'd for [C₂₆H₂₇Cl₂NO₃]⁺: *m/z* 427.1433, found 427.1433.

Representative Procedure H for the Formation of Hydroxypyridines:



5-methyl-6-phenylpyridin-3-ol (20). A 10 mL flame-dried round-bottom flask was charged with **5b** (100 mg, 0.2 mmol, 1 equiv) and THF (2 mL). Potassium trimethylsilanolate (27 mg, 0.2 mmol, 1 equiv) was then added to the flask in one portion. The resulting cloudy brown mixture was stirred under a nitrogen atmosphere until all the starting material was consumed as judged by TLC analysis (3 h) and subsequently quenched with acetic acid (12.5 mg, 0.2 mmol, 1 equiv). The brown solution was concentrated under reduced pressure (rotary evaporation). The crude cloudy brown oil was purified by column chromatography (50 mL deactivated neutral alumina, 4:1 hexanes:ethyl acetate then ethyl acetate) to yield 39 mg (79%) of **20** as a white foam. **R**_f = 0.0 (4:1 hexanes:ethyl acetate); ¹**H NMR** (500 MHz, CD₃OD) δ ppm 153.3, 149.7, 139.9, 133.7, 132.4, 128.7, 127.8, 127.4, 125.1, 18.6; **IR** (film) 3423, 2528, 1647 cm⁻¹; **HRMS** (EI) calc'd for [C₁₂H₁₁NO]⁺: *m/z* 185.0827, found 185.0833.

References:

- (1) K. W. Henderson, J. K. William, M. H. Jennifer, Tetrahedron 2002, 58, 4573-4587.
- (2) J. U. Jeong, I. Sagasser, K. B. Sharpless, J. Am. Chem. Soc. 1998, 120, 6844-6845.
- (3) M. Kimura, M. Shimizo, S. Tanaka, Y. Tamaru, Tetrahedron 2005, 61, 3709-3718.
- (4) A. G. Kallianos, A. H. Warfield, M. I. Simpson, U.S. Patent 3, 704, 714, December 5, 1972.

HPLC Traces:

Propargylic Ester (1a) racemic and enriched HPLC traces:





					and the second se	
	[Min]			[mAU]	[mAU*min]	[%]
1	3.66	0.00	0.00	121.24	14.46	100.000
Total					14.46	100.000

Dihydropyridine (5a) racemic and enriched HPLC traces:





X-ray Structure of 17c:



17c

Table 1.	Crystal	data and	structure	refinement	for	$C_{28}H_3$	₃₃ NO ₃	(17c).
----------	---------	----------	-----------	------------	-----	-------------	-------------------------------	--------

Identification code	$C_{28}H_{33}NO_3$	
Empirical formula	C ₂₈ H ₃₃ NO ₃	
Formula weight	431.55	
Temperature	155(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P2(1)/c	
Unit cell dimensions	a = 20.270(3) Å	a= 90°.
	b = 8.1071(12) Å	b=96.360(3)°.
	c = 14.964(2) Å	$g = 90^{\circ}$.
Volume	2443.8(6) Å ³	
Z	4	

Density (calculated)	1.173 Mg/m ³
Absorption coefficient	0.075 mm ⁻¹
F(000)	928
Crystal size	0.27 x 0.20 x 0.12 mm ³
Theta range for data collection	1 to 23.5°.
Index ranges	-21<=h<=25, -9<=k<=9, -18<=l<=10
Reflections collected	4932
Independent reflections	3410 [R(int) = 0.0341]
Completeness to theta = 26.41°	98.9 %
Absorption correction	Empirical
Max. and min. transmission	1 and 0.8311
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	4932 / 0 / 421
Goodness-of-fit on F ²	0.886
Final R indices [I>2sigma(I)]	R1 = 0.0450, wR2 = 0.1203
R indices (all data)	R1 = 0.0733, wR2 = 0.1373
Largest diff. peak and hole	0.232 and -0.158 e.Å ⁻³

















































