

Supplemental Figure 1 – IFN- $\gamma$  plasma levels and lymphocyte apoptosis in mice treated with metalloporphyrins. (A) Effects of treatment with CoPP in vivo on the mean plasma levels of IFN $\gamma$  at 8 dpi. Samples were taken from 3-8 individual mice and evaluated by ELISA. (B) Effects of in vivo treatment with CoPP on the lymphocyte apoptosis, as detected by flow cytometry of splenic lymphocytes for annexin V and CD4 / CD8 at 8 dpi. The percentages inside the contour plots represent annexin V+ cells among the respective T cell subset (CD4 or CD8). Graphs are representative of at least 3 independent samples; each sample was derived from spleens of 3-5 pooled mice. NI= non-infected; - = infected untreated controls.



Supplemental Figure 2. CoPP does not directly kill trypomastigotes, interfere with intracellular amastigogenesis or induce secretion of a soluble killing factor. (A) Effects of treatment with CoPP and SnPP on the mean trypomastigote number in thioglycollate-elicited macrophages. Trypomastigotes were counted in supernatants after 5 days of cell culture. (B) Effects of CoPP and SnPP on the expression of HO-1, as shown by immunoblot. (C) Effects of treatment with CoPP, SnPP, biliverdin (Blv), or bilirubin (Bb) on mean trypomastigote survival, as assessed by motility. Three independent wells were counted for each point. (D) Effects of treatment with CoPP or benzonidazole (100µM) on the viability of trypomastigotes, assessed by iodide propidium (IP) exclusion (flow cytometry). (E) Effects of treatment with supernatants (SN) from derived macrophages as in (A) on trypomastigote survival. Three independent wells were counted for each point. (F) Effects of treatment with CoPP and SnPP or left untreated, at 5 dpi or (G) 9 dpi. (H) Effects of treatment with CoPP and SnPP on the mean amastigogenesis period). (I) Effects of treatment in vivo with CoPP starting 2 days after infection on the mean parasitemia (n=8). Controls treated with CoPP starting 2h after infection (usual regimen) are also shown. NI= non-infected; -=infected non-treated controls. Errors bars represent SEM. All experiments were repeated at least twice with similar results, except for F.



Supplemental Figure 3. TNF is not involved in CoPP-induced reduction of parasitism. (**A**) Effects of treatment in vivo with CoPP or SnPP on the mean TNF production by LPS-stimulated peritoneal macrophages at 8 dpi. Cells were pooled from 3-5 mice / group and results represent 3-4 such pools (ELISA). (**B**) Effects of treatment with CoPP or SnPP on the mean TNF production by LPS-stimulated thioglycollate-elicited macrophages. Results represent 3 independent wells (ELISA). (**C**) Effects of treatment with CoPP or SnPP on mean parasite burden of thioglycollate-elicited peritoneal macrophages derived from wild-types or  $Tnfr1^{-/-}$  mice. Cells were stained with Giemsa and amastigotes were counted in each cell of a sample of 100 infected cells. (**D**) Effects of treatment in vivo with CoPP or SnPP on the mean parasitemia of wild-types and  $Tnfr1^{-/-}$  (n=4-6 mice) and (**E**) mortality. NI = non-infected; - = infected untreated controls. Error bars represent SEM.



Supplemental Figure 4. Macrophage parasite burden does not change 6 h after treatment with CoPPi. Effects of treatment with CoPP on mean parasite burden of macrophages. Thioglycollate-elicited macrophages were infected with 10:1 trypomastigotes in vitro for 1 h and treated with CoPP for 6 h. Cells were stained with Giemsa and amastigotes were counted in each cell of a sample of 100 infected cells.



Supplemental Figure 5. Labile iron pool in infected versus non-infected cells. We used the quenching of calcein fluorescence as an indicator of labile iron presence. Thioglycollate-elicited macrophages were infected with 3:1 trypomastigotes in vitro for 12 h or left untreated, then washed and left untreated for additional 6 h. Cells were then scrapped off the plate and loaded with calcein as described in Methods.