

# Supplemental Figure S1

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AtSnRK1.1 MFKRVDDEFNLVSSTIDHRIFKSRMDGSG-TGSRSGVESILPNYKLGRTLIGIGSFGRVKIA 59
S1SnRK1 -----MDGTA-VQGTSSVDSFLRNYKLGKTLGIGSFQKVKIA 36
AtSnRK1.2 -----MDHSSNRFNGNGVESILPNYKLGKTLGIGSFQKVKIA 37
AtSnRK1.3 -----MDGSS-EKTTNKLVSILPNYRIGKTLGHGSAKVKLA 36
AtSnRK2.1 -----MDKYDVVKDLGAGNFGVARLL 21
AtSnRK2.2 -----MDPATNSPIMPIDLPIMHSDRYDFVKDIGSGNFGVARLM 40
AtSnRK2.3 -----MDRAP-VTTGPLDMPIMHSDRYDFVKDIGSGNFGVARLM 39
AtSnRK3.1 -----MEK-----KGSVLMRLRYEVGKFLGQGTFAKVYHA 29
AtSnRK3.2 -----MEN-----KPSVLTERYEVGRLGQGTFAKVYFG 29
AtSnRK3.3 -----MESPYPKSPEKITGTVLLGKYELGRRLGSGSFAKVHVA 38
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AtSnRK1.1 EHALTGHKVAIKILNRRKIKNMEMEKKVRRREIKILR-LFMHPHIIRLYEVIETPTDIYLV 118
S1SnRK1 EHTLTGHKVAVKILNRRKIRNMDMEKKVRRREIKILR-LFMHPHIIRLYEVIETPSDIYVV 95
AtSnRK1.2 EHVVTGHKVAIKILNRRKIKNMEMEKKVRRREIKILR-LFMHPHIIRQYEVIEETSDIYVV 96
AtSnRK1.3 LHVATGHKVAIKILNRSKIKNMGIEIKVQREIKILR-FLMHPHIIRQYEVIEETPNDIYVV 95
AtSnRK2.1 RHKDTKELVAMKYIER----GRKIDENVAREIINHR-SLKHPNIIRFKEVILTPHLAIV 76
AtSnRK2.2 TDRVTKELVAVKYIER----GEKIDENVQREIINHR-SLRHPNIVRFKEVILTPSHLAIV 95
AtSnRK2.3 RDKLTKELVAVKYIER----GDKIDENVQREIINHR-SLRHPNIVRFKEVILTPHLAII 94
AtSnRK3.1 RHLKTGDSVAIKVIDKERILKVGMTQIKREISAMR-LLRHPNIVELHEVMATKSKIYFV 88
AtSnRK3.2 RSNHTNESVAIKMIDKDKVMRVGLSQQIKREISVMR-IAKHPNVVELYEVMATKSRIFV 88
AtSnRK3.3 RSISTGELVAIKIIDKQKTIDSGMEPRIIREIEAMRRLHNHPNVLKIEVMATKSKIYLV 98
* . **:* :: : . : *** * **::: **:* . : . :

AtSnRK1.1 MEYVNSGELFDYIVEKGRLOEDEARNFFQOIISGVEYCHRMVVRDLKPENLLLDKSKCN 178
S1SnRK1 MEYVKSSELFDYIVEKGRLOEDEARNFFQOIISGVEYCHRMVVRDLKPENLLLDKSKWN 155
AtSnRK1.2 MEYVKSSELFDYIVEKGRLOEDEARNFFQOIISGVEYCHRMVVRDLKPENLLLDKSRCN 156
AtSnRK1.3 MEYVKSSELFDYIVEKGRLOEDEARNFFQOIISGVEYCHRMVVRDLKPENLLLDKQCN 155
AtSnRK2.1 MEYASGGELFDRICTAGRFSEAEARYFFQQLICGVYCHSLQICHRDLKLENTLLDGSPA 136
AtSnRK2.2 MEYASGGELYERICNAGRFSEAEARYFFQQLISGVSYCHAMQICHRDLKLENTLLDGSPA 155
AtSnRK2.3 MEYASGGELYERICNAGRFSEAEARYFFQQLISGVSYCHSMQICHRDLKLENTLLDGSPA 154
AtSnRK3.1 MEHVKGSELFNKVST-GKLRDVARVYFQQLVRAVDVFCVSRGVCHRDLKPENLLLDDEHGN 147
AtSnRK3.2 IEYCKGSELFNKVAK-GKLRDVARVYFQQLISAVDVFCHSRGVYHRDIKPENLLLDNDN 147
AtSnRK3.3 VEYASGGELFTKLIRFGRLNESAAARYFQQLASALSFCVHRDGIHRDVKPQNLKQGN 158
*: .***: : **: * * * *: .:*** : **:* : * **

AtSnRK1.1 --VKIADFGLSNIM---RDGHFLKTSVCGSPNYAAPEVISGKLYAGPEVDVWSCGVILYAL 233
S1SnRK1 --VKIADFGLSNIM---RDGHFLKTSVCGSPNYAAPEVISGKLYAGPEVDVWSCGVILYAL 210
AtSnRK1.2 --IKIADFGLSNVM---RDGHFLKTSVCGSPNYAAPEVISGKLYAGPEVDVWSCGVILYAL 211
AtSnRK1.3 --IKIVDFGLSNVM---HDGHFLKTSVCGSPNYAAPEVISGKPYG-PDVDIWSGVILYAL 209
AtSnRK2.1 PLLKICDFGYSKSS---ILHSRPKSTVGTTPAYIAPEVLSRREYDGGKADVWSCGVTLVYM 193
AtSnRK2.2 PRLKICDFGYSKSS---VLHSQPKSTVGTTPAYIAPEILLRQYDGGKADVWSCGVTLVYM 212
AtSnRK2.3 PRLKICDFGYSKSS---VLHSQPKSTVGTTPAYIAPEVLLRQYDGGKADVWSCGVTLVYM 211
AtSnRK3.1 --LKISDFGLSALSRRQDGLLHTTCGTPAYVAPEVISRNGYDGGKADVWSCGVILFVL 205
AtSnRK3.2 --LKVSDFGLSALADCKRQDGLLHTTCGTPAYVAPEVINRKGYEGTKADIWSCGVVLFVL 205
AtSnRK3.3 --LKVSDFGLSALPEHRSNGLLHTTACGTPAYVAPEVIAQRGYDGGKADAWSCGVVLFVL 216
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AtSnRK1.1 LCGTLPFDDENIPNLFKKIKGGIYTLPS-----HLSPGARDLIPRMLVVDPMKRVTIPE 287
S1SnRK1 LCGTLPFDDENIPNLFKKIKGGIYTLPS-----HLSAGARDLIPRMLIVDPMKRMTIPE 264
AtSnRK1.2 LCGTLPFDDENIPNLFKKIKGGIYTLPS-----HLSSEARDLIPRMLIVDPVKRITPE 265
AtSnRK1.3 LCGTLPFDDENIPNVFEKIKRGMYYTLPN-----HLSHFARDLIPRMLMVDPTMRISITE 263
AtSnRK2.1 LVGAYPFEDPNDPKNFRKTIQRIMAVQYKIPDYVHISQECKHLLSRIFVTNSAKRITLKE 253
AtSnRK2.2 LVGAYPFEDPQEPDRYKTIQRILSVTYSIPEDLHLSPECRHLISRIFVADPATRITPE 272
AtSnRK2.3 LVGAYPFEDPEPRDYKTIQRILSVKYSIPDDIRISPECCHLISRIFVADPATRISIPE 271
AtSnRK3.1 LAGYLPFRDNLMELYKKGKAEVKFPN-----WLAGAKRLLKRILDPNPTRVSTEK 259
AtSnRK3.2 LAGYLPFHDTNLMEMRYKIKGADFKCPS-----WFAPEVKRLLCKMLDPNHETRITIAK 259
AtSnRK3.3 LAGYVPFDDANIVAMYRKIHKRDYRFPS-----WISKPARSIYKLLDPNPETRMSIEA 270
* * * * : : * : : : : * :

AtSnRK1.1 IRQHPWFQAHLPYLAVPPDTPVQQAkki-----DEEILQEVINMGFDRNHL 334
S1SnRK1 IRLHPWFQAHLPYLAVPPDTPVQQAkki-----DEEILQEVVKGDFDRNHL 311
AtSnRK1.2 IRQHRWFQTHLPYLAVSPDTPVEQAkki-----NEEIVQEVVNMGFDRNQV 312
AtSnRK1.3 IRQHPWFNNHLPYLSIPPLDTIDQAkki-----EEEIIONVVNIGFDRNHV 310
AtSnRK2.1 IKNHPWYLKLNLPKELLESA-QAAYYKRDT-----SFSLOSVEDIMKIVGEAR 299
AtSnRK2.2 ITSDKWFLKLNLPGLMDEN-RMGSQFQEP-----EQPMQSLDTIMQIISEAT 318
AtSnRK2.3 IKTHSWFLKLNLPADLMNES-NTGSQFQEP-----EQPMQSLDTIMQIISEAT 317
AtSnRK3.1 IMKSSWFRKGLQEEVK--ESVEETEVD-----EAEGNASAEK----EKKRCINLNAFEI 309
AtSnRK3.2 IKESSWFRKGLHLKQKKMEKMEKQOVREATNPMEAGGSGQENGENHEPPRLATLNAFDI 319
AtSnRK3.3 VMGTVWFQKSLEISEFQSSVFEldrFLEK-----EAKSSNAITAFDL 312
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# Supplemental Figure S1 contd.

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AtSnRK1.1	IESLRNRTQNDGTVTYYLILDNRFR--ASSGYLGAEFQETMEG--TPRMHPAESVASPVSH	391
<b>SISnRK1</b>	<b>TESLRNRVQNEGTVAAYLLLDNRHR--VSTGYLGAEFQESMEYGYNRINSNETAASPVGQ</b>	<b>369</b>
AtSnRK1.2	LESLRNRTQNDATVTYYLILDNRFR--VPSGYLESEFQETTWF-----	353
AtSnRK1.3	VDSLARIQNEATVAYHLILDNRNQNVSVPNDPFQSKFKEISDGIFNSTLPVQNITSHVGH	370
AtSnRK2.1	NPAPSTSAVKSSGSGADEEEEEEDVEAEVEEEEEDEDEYEKHKVKEAQSCQESDKA-----	353
AtSnRK2.2	IPTVRNRCLDDFMADN-LDLDDDMDDFDSESEIDVDSSGEIVYAL-----	362
AtSnRK2.3	IPAVRNRCLDDFMTDN-LDLDDDMDDFDSESEIDIDSSGEIVYAL-----	361
AtSnRK3.1	ISLSTGFDSLGLFEKGEKEEMRFTSNREASEITEKLVEIGKDLKMKVRKK-EHEWRVKM	368
AtSnRK3.2	IALSTGFGLAGLFGDVYDKRESRFASQKPASEIISKLVKAKCLKLRKQAGLFLKLER	379
AtSnRK3.3	ISLSSGLDLGLFER-RKRKEKRFTARVSAERVVEKAGMIGEKLGRFVEKK--EETKVVG	369
:		
AtSnRK1.1	RLPGLMEYQGVGLRSQYPVERKWALGLQSRAPREIMTEVLKALQDLNVCWKKIGHYNMK	451
<b>SISnRK1</b>	<b>RFPGIMDYQQAGAR-QFPIERKWALGLQSRAPREIMTEVLKALQELNVCWKKIGQYNMK</b>	<b>428</b>
AtSnRK1.2	-----QSYAHT-----	359
AtSnRK1.3	SFSALYGLKSNVKD-----DKTWTGLGLQSQGSPYDIMTEIFKALQNLKICWKKIGLYNIK	425
AtSnRK2.1	-----	
AtSnRK2.2	-----	
AtSnRK2.3	-----	
AtSnRK3.1	SAEAT----VVEAEVFEIAPSYHMVVLKKSQGDTAEYKRVMK--ESIRPALIDFVLAWH-	421
AtSnRK3.2	VKEGKNGILTMDAEIFQVTPTFHLVEVKKCNQDTEYQKLV--EDLRPALADIVVWVQG	437
AtSnRK3.3	LGKGR---TAVVVEVVEFAEGLVVADVKKVVVEGEEEEEEVESHWSELIVELEEIVLSWHN	426
:		
AtSnRK1.1	CRWVPNSS--ADGMLSNSMHDNNYFGDESSIIENEAAVKSPNVVKFEIQLYKTRDDKYLL	509
<b>SISnRK1</b>	<b>CRWVPSLPGHHEGMVNSMHGNQFFGDDSSIIENDGATKLTNVVKFEVQLYKTREEKYLL</b>	<b>488</b>
AtSnRK1.2	-----	
AtSnRK1.3	CRWVRSFAYYKN-----HTIEDECAIILPTVIKFEIQLYKVREGKYLL	468
AtSnRK2.1	-----	
AtSnRK2.2	-----	
AtSnRK2.3	-----	
AtSnRK3.1	-----	
AtSnRK3.2	EKEKEEQLLQDEQGEQEPS-----	456
AtSnRK3.3	-----	
:		
AtSnRK1.1	DLQRVQGPQFLFLDLCAAFLAQLRVL	535
<b>SISnRK1</b>	<b>DLQRVQGPQFLFLDLCAAFLAQLRVL</b>	<b>514</b>
AtSnRK1.2	-----	
AtSnRK1.3	DILRIDGPQFIFDLCAVFLRELGVL	494
AtSnRK2.1	-----	
AtSnRK2.2	-----	
AtSnRK2.3	-----	
AtSnRK3.1	-----	
AtSnRK3.2	-----	
AtSnRK3.3	-----	

**Supplemental Figure S1.** Alignment of SnRK proteins from tomato and *Arabidopsis*. The following SnRK sequences were aligned using clustalW: SISnRK1 (shown in bold; AF143743), AtSnRK1.1 (NP850488), AtSnRK1.2 (NP974375), AtSnRK1.3 (NP198760), AtSnRK2.1 (P43292), AtSnRK2.2 (Q39192), AtSnRK2.3 (Q39193), AtSnRK3.1 (P92937), AtSnRK3.2 (Q9LYQ8), AtSnRK3.3 (Q9SUL7). The invariant lysine responsible for ATP binding is shown in yellow outlined in black; K48 in SISnRK1. The activation phosphorylation site, T175 in SISnRK1, is shown in red outlined in black.

# Supplemental Figure S2 contd.

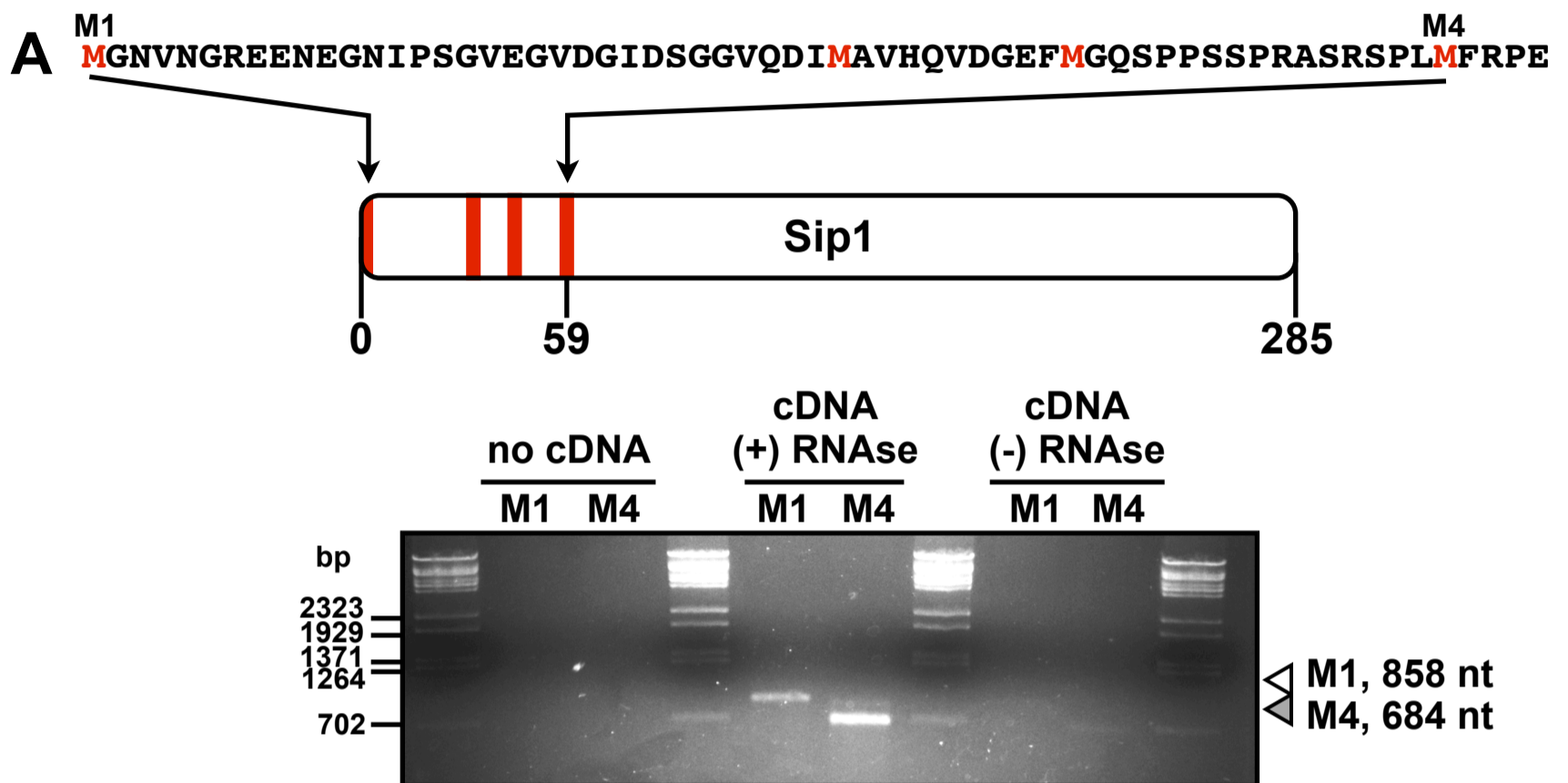
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AtAKINβ1	MGNANGKDEDAAGSGGADVTSSSAR	<b>S</b> NGGDPSARSRH----	RRPSSDSMSSSPPGSPAR	56		
<b>SlGal83</b>	<b>MGNANAR-EDGAAGDGDG</b>	<b>D</b> GQVSGRR <b>S</b> NVE <b>S</b> GIVEDHHALNSRVPSADLMVNSPPQSPHR		59		
<b>SlSip1</b>	<b>MGNVNGREENEGNI</b>	<b>P</b> SGVEGVG--IDSGGVQDIMAVHQ----	VDGEFMGQSPSSPRA	53		
<b>SlTau2</b>	<b>MGNVNGREE</b>	-----ID-- <b>Q</b> <b>S</b> SVGIQETMDAR-----	DGEFMGQSPSSPRA	39		
AtAKINβ2	MGNVNAREEANSNNASAVEDED----	AEICSRREAMSAASDGNHVAPPELMGQSPPHSPRA		56		
<b>SlTau1</b>	<b>MGNVSGKKKEGESAE</b>	<b>S</b> SGIKNQ----- <b>E</b> HGEEYMEYG-----	LFPDSMVQSPPHSPKA	49		
HsAMPKβ1	MGNTSSERAALERHGGHKTPRR-----		<b>D</b> <b>S</b> GGTKDGRPK	35		
HsAMPKβ2	MGNTTSDRVSGERHG-AKAARS-----		EGAGGHAPGKEHK	34		
	***...	:	:			
AtAKINβ1	-SPSPFLFAPQVPVAPLQORANAPPNNIQW-NQS-QRVFDNPP-EQGIPTIITWNQGGND			112		
<b>SlGal83</b>	<b>-SASPLLF</b>	<b>G</b> QVPVPLQGGDGNPVSNQMWGNEC-EDASDHSL-EGGIPTLITWSYGGNN		116		
<b>SlSip1</b>	<b>-SRSPL</b>	<b>M</b> FRPEMPVPLQRPDEGHGPSISWSQT--TSGYEPCDEQGVPTLISWTLDGKE		110		
<b>SlTau2</b>	<b>-SHSPLM</b>	<b>F</b> RQMPVPLQRPPEELHISNPSWMQN--TSGYEDLNEEKGVPTLISWTYEGKD		96		
AtAKINβ2	-TQSPLMFAPQVPVPLQRPDEIHIPNPSWMQSP-SSLYEEASNEQGIPTMITWCHGGKE			114		
<b>SlTau1</b>	<b>YHHSPLD</b>	<b>F</b> TPQVPIFPLQRPDEILMQNQSGNIVQKTMEYGDMPCENGIPTMITWSHGGHE		109		
HsAMPKβ1	-----ILMDSPEADLDFHSEEIKAPEKEEF	FLAWQHDLEVNDKAPAQARPTVFRWTGGGKE		90		
HsAMPKβ2	-----IMVGSTDDPSVFS	LPDSKLPGDKEFVSWQQDLEDSVKPTQQARPTVIRWSEGGKE		89		
	: . . . . .	:	:			
AtAKINβ1	VAVEGSWDNWRSRKKLQK	<b>S</b> GKDHSILFVLPSGIYHYKVI	VDGESKYIPDLPFVADEVGNV	172		
<b>SlGal83</b>	<b>VAIQGS</b>	<b>W</b> DNWTSRKILQK <b>S</b> GKDYTVLLVLP	SGIYHYKFIVDGEVRYIPELPCVADETVV	176		
<b>SlSip1</b>	<b>VAVEGS</b>	<b>W</b> DNWKSRLMPLQK <b>S</b> GKDFITLKVLP	SGVYQYRFIVDQWRCSPDLPCVQDEAGNT	170		
<b>SlTau2</b>	<b>IAVEGS</b>	<b>W</b> DNWKSRLMPLQK <b>S</b> GKDFITLKVLP	SGVYQYRFIVDQWRCSPDLPCVQDEAGNT	156		
AtAKINβ2	IAVEGSWDNWKTRSRQLQK	<b>S</b> GKDFITMKNVLP	SGVYQYRFIVDQWRHAPPELPLARDDAGNT	174		
<b>SlTau1</b>	<b>VAIEGS</b>	<b>W</b> DGKTKDFLQK <b>S</b> GKDFITVMKVF	PSGVYHYRFIVDQWRYAPDYPYERDDTGNV	169		
HsAMPKβ1	VYLSGSFNNWS-KLPLTR	<b>S</b> HNNFVAI	LDLPEGEHQYKFFVDGQWTHDPSEPIVTSQLGTV	149		
HsAMPKβ2	VFISGSFNNWSTKIPLIK	<b>S</b> HNDFVAI	LDLPEGEHQYKFFVDGQWVHDPSEPVVTSQLGTI	149		
	: . : . : .	:	:			
AtAKINβ1	CNILDVHNFVNPENPESIVEFEA-----	PP	SPDHSYGQTLPAA--EDYAKEPLAVP	220		
<b>SlGal83</b>	<b>FNLLDV</b>	<b>N</b> DPENLESVAEFEA-----	PP	SPDSSYAQALMGE--EDFEKEPVAVP	224	
<b>SlSip1</b>	<b>YNLLDM</b>	<b>K</b> DYVPEDIESISGFEP-----	PQ	SPDSSYNNLHLVS--EDYAKEPPVVP	218	
<b>SlTau2</b>	<b>YNILDV</b>	<b>K</b> DYVPEDIESISGFEP-----	PL	SPDSSYNLELGA--EDYAKEPPLVP	204	
AtAKINβ2	FNILDLDQDYVPEDIQISISGFEP-----	PQ	SPENSYSNLLLGA--EDYSKEPPVVP	222		
<b>SlTau1</b>	<b>FNVLDL</b>	<b>Q</b> DI IPEVLNNTNWSDA-----	PP	SPESSYSNAPFSS--EDFSEKLPDLP	217	
HsAMPKβ1	NNIIQVKKTD	FEVFDALMVDSQKCS	DVS--ELSS	PPGPYHQEPYVCKPEERFRAPPILP	207	
HsAMPKβ2	NNLIHVKKSD	FEVFDALKLDSMESSE	TS	CRDLSS	PPGPYQEMYAFRSEERFKSPILP	209
	* : : : .	:	:			
AtAKINβ1	PQLHLTLLG--TTEETA---	IATKPQHVVNLNHVFIEQGWTPQSIVALGLTHR	FESKYITV	275		
<b>SlGal83</b>	<b>PQLHLT</b>	<b>V</b> LGSENSEAP---SSPKQHVVNLNHLFIEK	GWASQSIVALGLTHR	FQSKYVTV	281	
<b>SlSip1</b>	<b>PHLQMT</b>	<b>L</b> LNVPSPHMEI-PPPLSRPQHVVNLNHL	YMQKDRSTPSVVALGSTNRFLSKYVTV	277		
<b>SlTau2</b>	<b>PHLQMT</b>	<b>L</b> LNVPSSPMEILPPPLSRPQHVVNLNHL	YMQKGSNPSLVALSSTNRFLFKYVTV	264		
AtAKINβ2	PHLQMTLLNLPANPDI-PSPLPRPQH	VVNLNHL	YMQKGSVVALGSTRFLAKYVTV	281		
<b>SlTau1</b>	<b>PLLQOT</b>	<b>P</b> LDQSSSAGS-VETFRKPLPAVLNHL	YIQKTRSSQSMVLSSTHRFRTKYVTA	276		
HsAMPKβ1	PHLLQVILNKDTGISCD-PALLPEPNH	VMLNHL	YALS--IKDGMVLSATHRYKKKYVTT	264		
HsAMPKβ2	PHLLQVILNKDTNISCD-PALLPEPNH	VMLNHL	YALS--IKDSVMVLSATHRYKKKYVTT	266		
	* * . *	:	:			
AtAKINβ1	VLYKPLTR			283		
<b>SlGal83</b>	<b>VLYKPLKR</b>			289		
<b>SlSip1</b>	<b>VLYKSIQS</b>			285		
<b>SlTau2</b>	<b>VLYKSIQR</b>			272		
AtAKINβ2	VLYKSLQR			289		
<b>SlTau1</b>	<b>VLFKSLKK</b>			284		
HsAMPKβ1	LLYKPI--			270		
HsAMPKβ2	LLYKPI--			272		
	: * : * . :	:	:			

**Supplemental Figure S2.** Alignment of SnRK complex β-subunits. The following β-subunit protein sequences were aligned using clustalW: SlGal83 (JF895513), SlSip1 (JF8955212), SlTau1 (XXXX), SlTau2 (XXXX), AtAKINβ1 (AAM6584), AtAKINβ2 (CAB64719), HsAMPKβ1 (NP\_006244), and HsAMPKβ2 (NP\_005390). The N-terminal end of the original SlGal83 (D18) is shown in purple and outlined in black. The Adi3 phosphorylation site on SlGal83 (S26) and the MS identified phosphorylation site S30 are shown in red outlined in black. The originally identified SlSip1 start site (M33) is shown in orange outlined in black. The HsAMPKβ1 phosphorylation sites are shown in pink outlined in black.

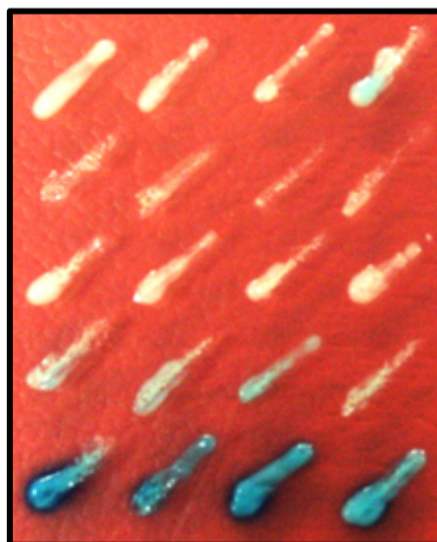
# Supplemental Figure S3

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## B

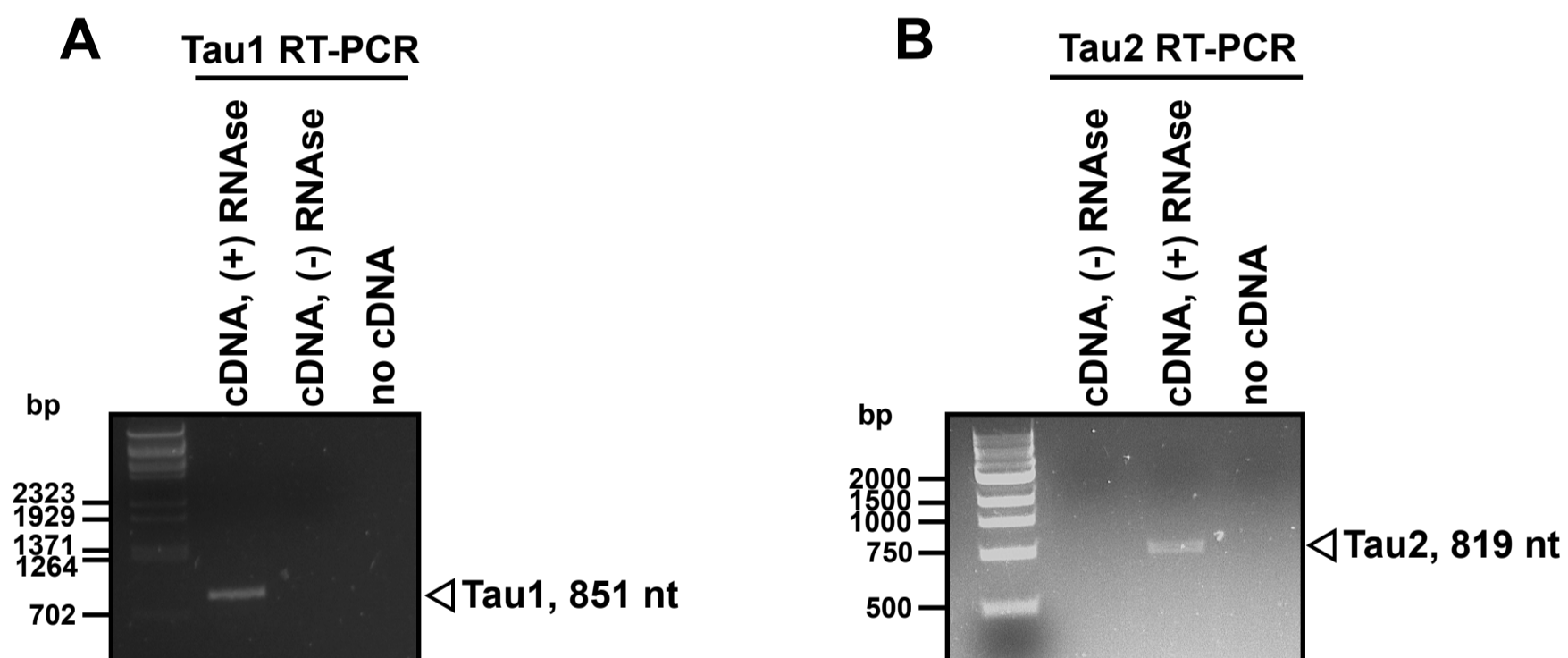
Bait	Prey
Adi3	pJG4-5
pEG202	GAL83
Adi3	BICOID
Dorsal	GAL83
Adi3	GAL83



**Supplemental Figure S3.** RT-PCR amplification of S1Sip1 and Adi3/S1Gal83 yeast two-hybrid interaction. A, Schematic of S1Sip1 showing originally identified start site (M4) and newly identified start site (M1). Also shown below is the RT-PCR gel showing amplification of the larger S1Sip1 from tomato leaf total RNA. B, Yeast two-hybrid interaction between Adi3 and Gal83. The indicated bait and prey constructs were tested in a standard yeast two-hybrid assay for expression of the *lacZ* gene on X-Gal plates.

# Supplemental Figure S4

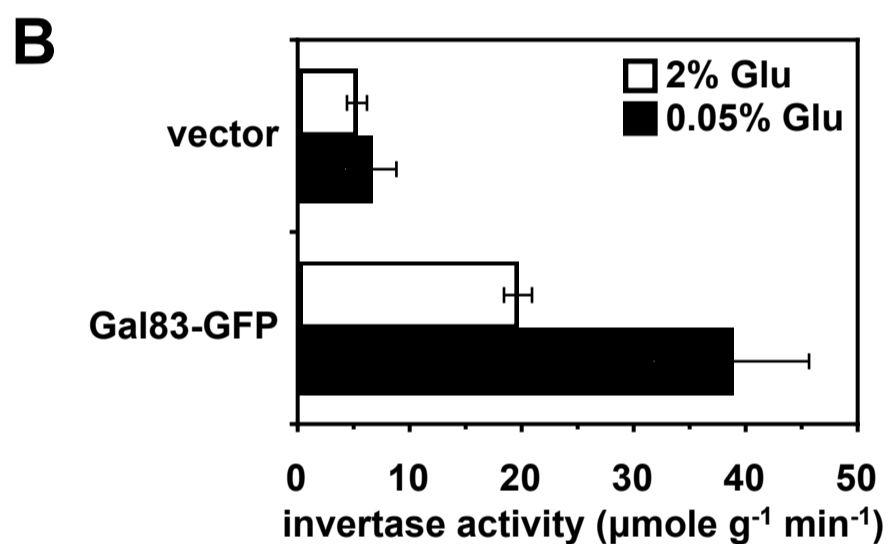
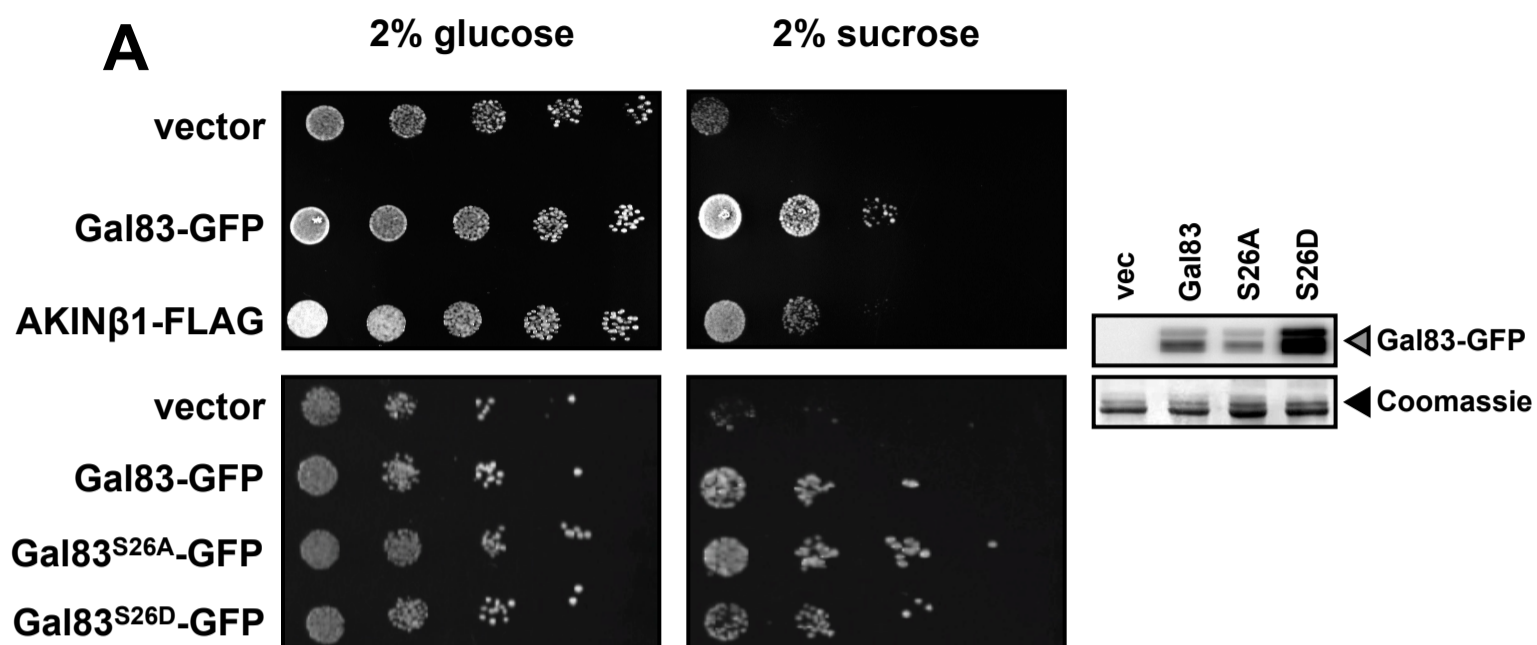
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**Supplemental Figure S4.** RT-PCR amplification of S1Tau1 and S1Tau2. A, RT-PCR gel showing amplification of the S1Tau1 cDNA from tomato leaf total RNA. B, RT-PCR gel showing amplification of the S1Tau2 cDNA from tomato leaf total RNA.

# Supplemental Figure S5

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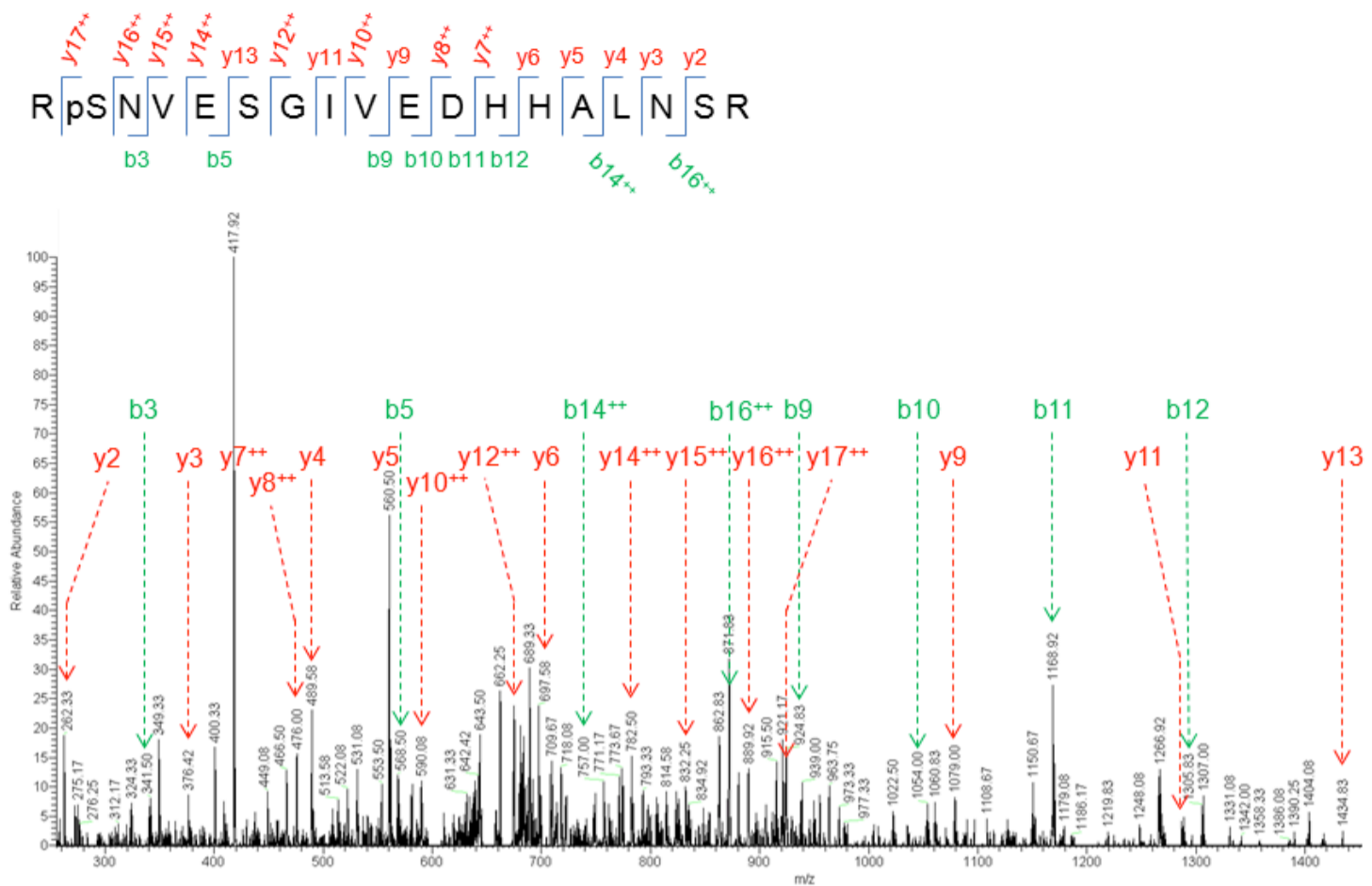


**Supplemental Figure S5.** SIGal83 complementation of *sip1 $\Delta$ sip2 $\Delta$ gal83 $\Delta$*  yeast. A, Tomato Gal83 can complement the yeast  $\beta$ -subunit triple knockout. *sip1 $\Delta$ sip2 $\Delta$ gal83 $\Delta$*  yeast cells were transformed with empty vector, the indicated SIGal83 constructs, or AKIN $\beta$ 1 and plated on 2% glucose or sucrose CM plates at 5-fold dilutions. SIGal83-GFP protein expression detected by  $\alpha$ -GFP western blot is shown on the right. B, SIGal83 complementation of yeast invertase activity. The indicated constructs were transformed into *sip1 $\Delta$ sip2 $\Delta$ gal83 $\Delta$*  yeast and extracts from the yeast tested for invertase activity in the presence of high (2%) and low (0.05%) glucose. Values are averages of three independent experiments and error bars are standard error.

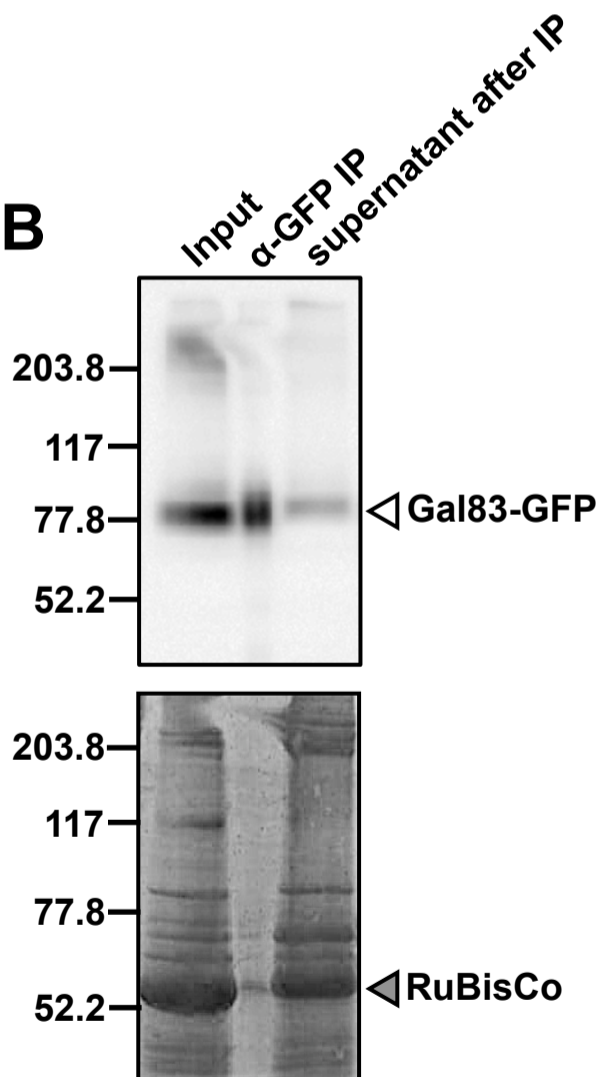
# Supplemental Figure S6

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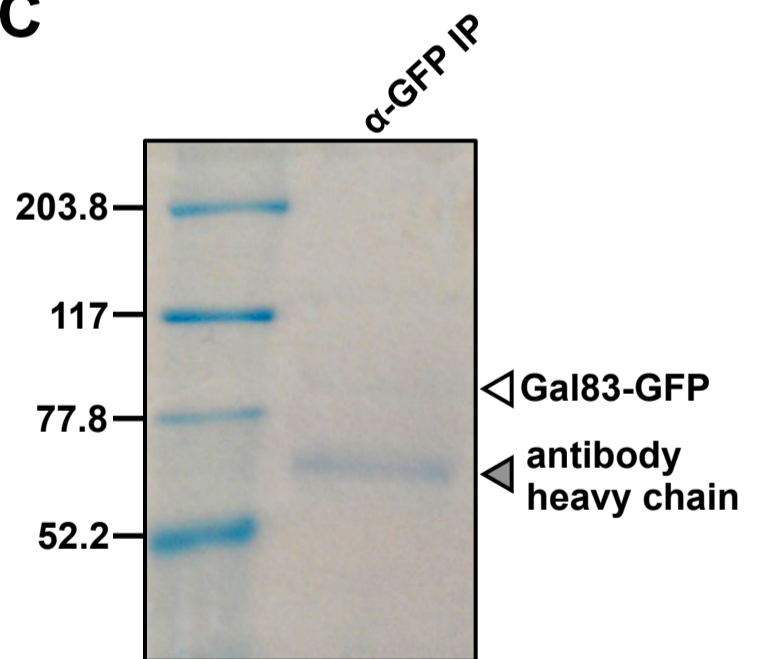
**A**



**B**



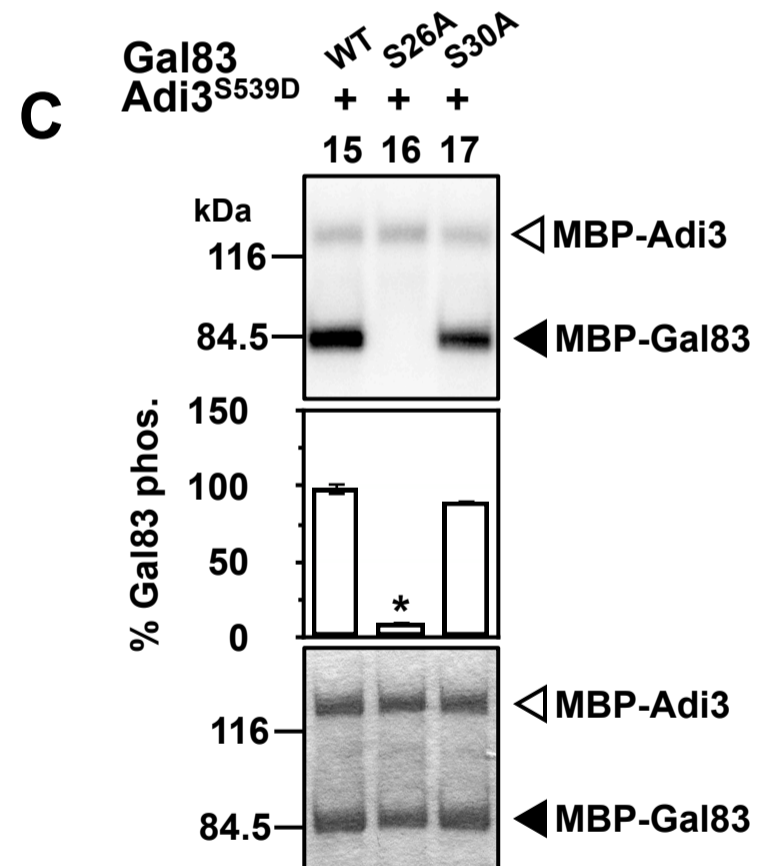
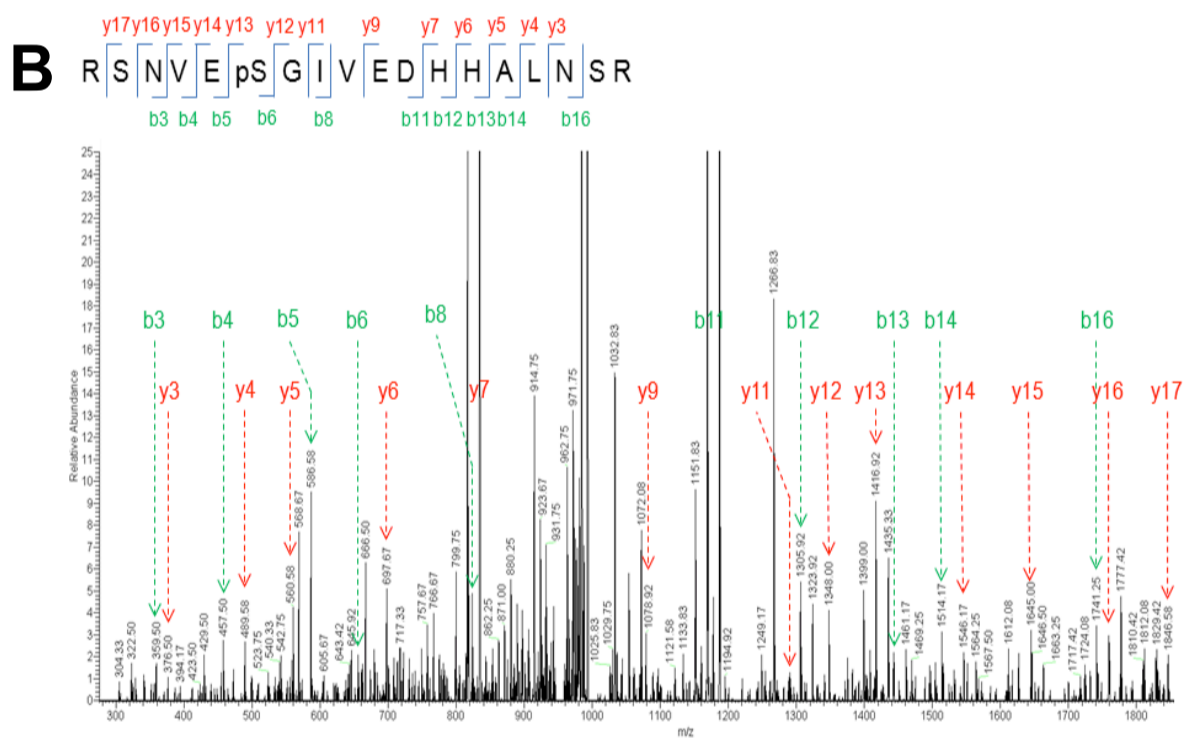
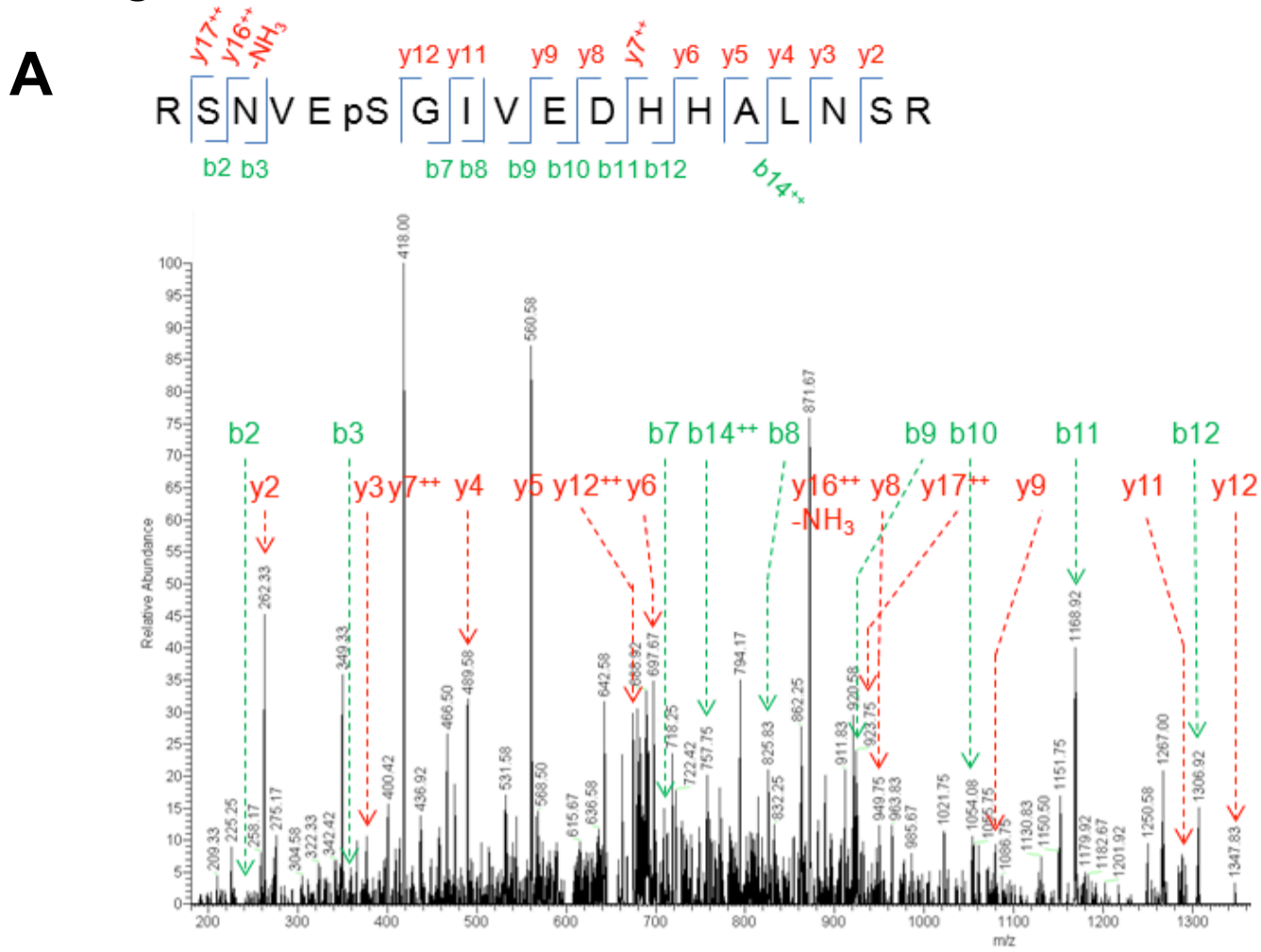
**C**



**Supplemental Figure S6.** MS identification of SIGal83 S26 phosphorylation and  $\alpha$ -GFP immunoprecipitation of SIGal83-GFP. A, Identification of SIGal83 S26 phosphorylation *in vitro* in peptide R p S N V E S G I V E D H H A L N S R. B, SIGal83-GFP can be pulled down with an  $\alpha$ -GFP antibody. Protoplasts expressing SIGal83-GFP for 16 hrs were lysed, immunoprecipitated with  $\alpha$ -GFP antibody, and analyzed by  $\alpha$ -GFP western blot. C, Immunoprecipitated SIGal83-GFP used for MS analysis. SIGal83-GFP was expressed and immunoprecipitated as in (A) and the sample separated by SDS-PAGE. The band corresponding to SIGal83-GFP was cut from the gel, trypsin digested and analyzed by MS.

# Supplemental Figure S7

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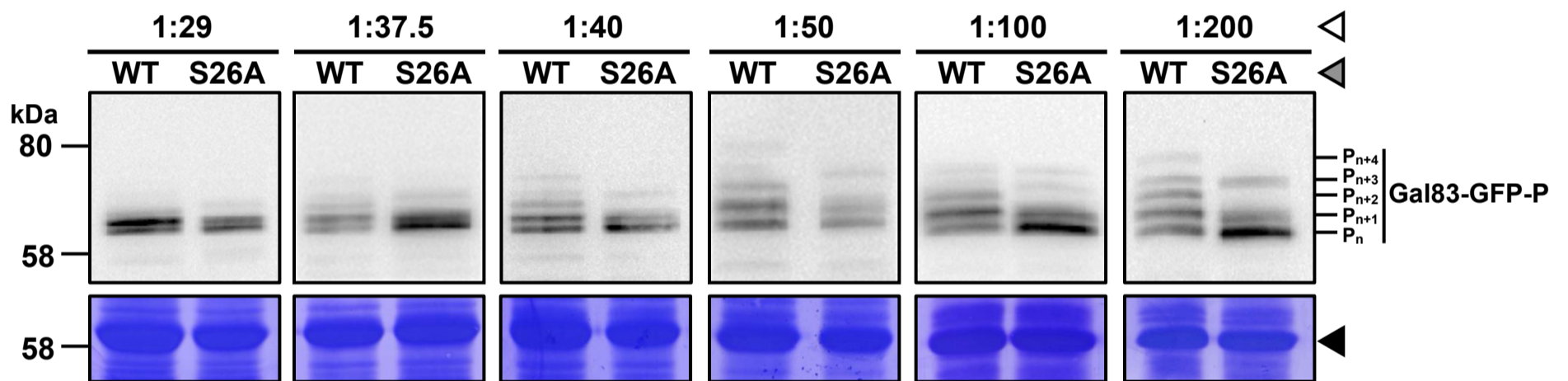


**Supplemental Figure S7.** MS identification of S1Gal83 S30 phosphorylation. A, Identification of S1Gal83 S30 phosphorylation *in vitro* in peptide RSNVEpSGIVEDHHALNSR. B, Identification of S1Gal83 S30 phosphorylation *in vivo* in peptide RSNVEpSGIVEDHHALNSR. C, Adi3 does not phosphorylate S1Gal83 S30 *in vitro*. One asterisk indicates significant decrease in phosphorylation of S1Gal83 Ser to Ala mutants compared to wild-type S1Gal83 phosphorylation (Student's *t* test,  $p < 0.05$ ).



# Supplemental Figure S8

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**Supplemental Figure S8.** Separation of SI Gal83-GFP phosphoproteins by SDS-PAGE with varying bis-acrylamide:acrylamide ratios. SI Gal83-GFP was stably transformed into *Arabidopsis*, protein extracts made from leaf tissue, and the extracts analyzed by SDS-PAGE. Open triangle, bis-acrylamide:acrylamide ratio; grey triangle, SI Gal83-GFP protein; black triangle, RuBisCo

# Supplemental Table S1

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Gene	Primer Name	Puopose	Direction	Restriction site	Sequence
Adi3	Adi3 BamHI-F	Cloning into pGEX	Forward	BamHI	CACGGATCCATGGAAAGGATACCTGAAGTT
Adi3	Adi3 EcoRI-R	Cloning into pGEX	Reverse	EcoRI	CACGAATTCCTAAAAGAAGTCAAAGTCAAG
SnRK1	SnRK EcoRI-F	ORF Amplification Cloning into pEG202/pJG4-5	Forward	EcoRI	CACGAATTCATGGACGGAACAGCAGTG
SnRK1	SnRK BamHI-R	ORF Amplification Cloning into pEG202	Reverse	BamHI	CACGGATCCCTTAAAGTACTCGAAGCTG
SnRK1	SnRK PstIR	Cloning into pMAL	Reverse	PstI	CACCTGCAGTTAAAGTACTCGAAGCTG
SnRK1	SnRK T175D-F	Mutagenesis	Forward		GGTCATTTTCTGAAGGATAGTTGCGGAAGCCCA
SnRK1	SnRK T175D-R	Mutagenesis	Reverse		TGGGCTTCCGCAACTATCCTTCAGAAAATGACC
SnRK1	SnRK K48Q-F	Mutagenesis	Forward		CACAAAGTTGCTGTCCAGATTCTTAATCGTCGA
SnRK1	SnRK K48Q-R	Mutagenesis	Reverse		TCGACGATTAAGAATCTGGACAGCAACTTTGTG
Snf4	Snf4 EcoRI	ORF Amplification Cloning into	Forward	EcoRI	CACGAATTCATGCAGGCAACAGCGGAG
Snf4	Snf4 Sall	ORF Amplification Cloning into	Reverse	Sall	CACGTCGACTCACTGCAAAAACTCAG
Sip1	Sip1 EcoRI	ORF Amplification Cloning into pMAL	Forward	EcoRI	CACGAATTCATGTTTAGACCTGAGATG
Sip1	Sip1 BamHI	ORF Amplification Cloning into pMAL	Reverse	BamHI	CACCTCGAGTCACCTCTGTATTGACTTG
Gal83	Gal83 EcoRI	ORF Amplification Cloning into pMAL,	Forward	EcoRI	CACGAATTCATGGGGAATGCGAACGCC
Gal83	Gal83 SallR	ORF Amplification Cloning into	Reverse	Sall	CACGTCGACTCACCTCTTCAGTGGCTTG
Gal83	Gal83 BamHI	Cloning into MBB263, pTEX cGFP	Forward	BamHI	CACGGATCCATGGGGAATGCGAACGCC
GFP	GFP EcoRI	Cloning of Gal83-GFP into MBB263	Reverse	EcoRI	CACGAATTCCTATTGTATAGTTCATCC
Gal83	Gal83 SallR NS	Cloning Gal83 without stop codon into pTEX cGFP	Reverse	Sall	CACGTCGACCCTCTTCAGTGGCTTGGTAG
Gal83	S22A	Mutagenesis	Forward		GGCGACGGT CAGGTAGCGGGAAGAAGA TCTAATG
Gal83	S22A	Mutagenesis	Reverse		CATTAGATCTTCTCCCGCTACCTGACCGTCGCC
Gal83	S26A	Mutagenesis	Forward		CGGGAAGAAGAGCTAATGTTGAATCTGG
Gal83	S26A	Mutagenesis	Reverse		CCAGATTCAACATTAGCTCTTCTTCCG
Gal83	S45A	Mutagenesis	Forward		CGCGAGTGCCTGCGGCTGACTTGATGG
Gal83	S45A	Mutagenesis	Reverse		CCATCAAGTCAGCCGCGAGGCACTCGCG
Gal83	S60/62A	Mutagenesis	Forward		GCAGAGTCCACATCGTGCAGCTGCACCTCTCTTGTTCGG
Gal83	S60/62A	Mutagenesis	Reverse		CCGAACAAGAGAGGTTGCAGCTGCACAGTGTGGACTCTGC
Gal83	S122A	Mutagenesis	Forward		GTTGCTATCCAAGGAGCTTGGGACAACCTGGAC
Gal83	S122A	Mutagenesis	Reverse		GTCCAGTTGTCCCAAGCTCCTTGGATAGCAAC
Gal83	S135A F	Mutagenesis	Forward		GGAAAATTCTCCAAAGAGCAGGCAAGGACTATACCG
Gal83	S135A R	Mutagenesis	Reverse		CGGTATAGTCTTGCCAGCTCTTTGGAGAATTTCC
Gal83	S147A	Mutagenesis	Forward		CTTTTGGTCTTCCAGCGGGTATATATCATTAC
Gal83	S147A	Mutagenesis	Reverse		GTAATGATATATACCCTGCTGGAAGGACCAAAAAG
Gal83	S192A	Mutagenesis	Forward		CCAGAGAACCTCGAAGCTGTTGCAGAGTTTGAG
Gal83	S192A	Mutagenesis	Reverse		CTCAAACCTGCAACAGCTTCGAGGTTCTCTGG
Gal83	S204/205A	Mutagenesis	Forward		CCACCATCACCTGACGCTGCCTATGCGCAAGCTTTG
Gal83	S204/205A	Mutagenesis	Reverse		CAAAGCTTGCGCATAGGCAAGCTCAGGTGATGGTGG
Gal83	S234A	Mutagenesis	Forward		CTAACTGTTCTTGGTGTGAAAACCTCAGAAGAAGC
Gal83	S234A	Mutagenesis	Reverse		GCTTCTTCTGAGTTTGTAGCACCAGAAGACAGTTAG
Gal83	S237A	Mutagenesis	Forward		GGTCTGAAAACGCAAGAAGAA GCACCTTC
Gal83	S237A	Mutagenesis	Reverse		GAAGGTGCTTC TTCTGCGTTTTC AGAACC
Gal83	S242/243A	Mutagenesis	Forward		CAGAAGAAGCACCTGCTGCT CCAAAACCCAGCACG
Gal83	S242/243A	Mutagenesis	Reverse		GTGCTGGGGTTTTGGAGCAGCAGGTGCTTCTCTG
Gal83	S262A/S264R	Mutagenesis	Forward		GAGAAAGGATGGGCTGCTCAAGCCATTGTTGCTCTTG
Gal83	S262A/S264R	Mutagenesis	Reverse		CCAAGAGCAACAATCCGTTGAGCAGCCCATCCTTTCTC
Tau1	Tau1-F	ORF amplification	Forward		ATGGGGAATGTGAGTGGG
Tau1	Tau1-R	ORF amplification	Reverse		TCACTTTTTCAAGGACTTAAAAAG
Tau1	Tau1 EcoRI	Cloning into pMAL	Forward	EcoRI	CACGAATTCATGGGGAATGTGAGTGGG
Tau1	Tau1 Sall	Cloning into pMAL	Reverse	Sall	CACGTCGACTCACTTTTCAAGGACTTAAAAAG
Tau2	Tau2-F	ORF amplification	Forward		ATGGGGAATGTTAATGGAAGAG
Tau2	Tau2-R	ORF amplification	Reverse		TCACTCTGTATGGACTTGTAAG
Tau2	Tau2 EcoRI	Cloning into pMAL	Forward	EcoRI	CACGAATTCATGGGGAATGTTAATGGA
Tau2	Tau2 Sall	Cloning into pMAL	Reverse	Sall	CACGTCGACTCACCTCTGTATGGACTTGTA

**Supplemental Table S1.** Primers used in this study are listed by gene name and purpose. In the primer sequences restriction sites are underlined, start and stop codons are in bold, and mutation sites are in bold and underlined.