

# The Tandem of Full Spin Analysis and qHNMR for the Quality Control of Botanicals Exemplified with *Ginkgo biloba*

José G. Napolitano, Tanja Gödecke, María F. Rodríguez-Brasco, Birgit U. Jaki, Shao-Nong Chen, David C. Lankin, and Guido F. Pauli\*

Institute for Tuberculosis Research and Department of Medicinal Chemistry and Pharmacognosy, College of Pharmacy, University of Illinois at Chicago, Chicago, IL 60612, U.S.A.

## ■ Supporting Information

## TABLE OF CONTENT

	<i>page</i>
<b>S1.</b> The $^1\text{H}$ Fingerprint of Ginkgolide A ( <b>1</b> ) in the PERCH .PMS file format.	S4
<b>S2.</b> $^1\text{H}$ NMR spectrum of Ginkgolide A ( <b>1</b> , DMSO- $d_6$ , 600 MHz).	S5
<b>S3.</b> $^1\text{H}, ^1\text{H}$ -COSY experiment of Ginkgolide A ( <b>1</b> , DMSO- $d_6$ , 600 MHz).	S6
<b>S4.</b> The $^1\text{H}$ Fingerprint of Ginkgolide B ( <b>2</b> ) in the PERCH .PMS file format.	S7
<b>S5.</b> $^1\text{H}$ NMR spectrum of Ginkgolide B ( <b>2</b> , DMSO- $d_6$ , 600 MHz).	S8
<b>S6.</b> $^1\text{H}, ^1\text{H}$ -COSY experiment of Ginkgolide B ( <b>2</b> , DMSO- $d_6$ , 600 MHz).	S9
<b>S7.</b> The $^1\text{H}$ Fingerprint of Ginkgolide C ( <b>3</b> ) in the PERCH .PMS file format.	S10
<b>S8.</b> $^1\text{H}$ NMR spectrum of Ginkgolide C ( <b>3</b> , DMSO- $d_6$ , 600 MHz).	S11
<b>S9.</b> $^1\text{H}, ^1\text{H}$ -COSY experiment of Ginkgolide C ( <b>3</b> , DMSO- $d_6$ , 600 MHz).	S12
<b>S10.</b> The $^1\text{H}$ Fingerprint of Ginkgolide J ( <b>4</b> ) in the PERCH .PMS file format.	S13
<b>S11.</b> $^1\text{H}$ NMR spectrum of Ginkgolide J ( <b>4</b> , DMSO- $d_6$ , 600 MHz).	S14
<b>S12.</b> $^1\text{H}, ^1\text{H}$ -COSY experiment of Ginkgolide J ( <b>4</b> , DMSO- $d_6$ , 600 MHz).	S15
<b>S13.</b> The $^1\text{H}$ Fingerprint of Bilobalide ( <b>5</b> ) in the PERCH .PMS file format.	S16
<b>S14.</b> $^1\text{H}$ NMR spectrum of Bilobalide ( <b>5</b> , DMSO- $d_6$ , 600 MHz).	S17
<b>S15.</b> $^1\text{H}, ^1\text{H}$ -COSY experiment of Bilobalide ( <b>5</b> , DMSO- $d_6$ , 600 MHz).	S18
<b>S16.</b> The $^1\text{H}$ Fingerprint of Quercetin ( <b>6</b> ) in the PERCH .PMS file format.	S19
<b>S17.</b> $^1\text{H}$ NMR spectrum of Quercetin ( <b>6</b> , DMSO- $d_6$ , 600 MHz).	S20
<b>S18.</b> $^1\text{H}, ^1\text{H}$ -COSY experiment of Quercetin ( <b>6</b> , DMSO- $d_6$ , 600 MHz).	S21
<b>S19.</b> The $^1\text{H}$ Fingerprint of Kaempferol ( <b>7</b> ) in the PERCH .PMS file format.	S22
<b>S20.</b> $^1\text{H}$ NMR spectrum of Kaempferol ( <b>7</b> , DMSO- $d_6$ , 600 MHz).	S23
<b>S21.</b> $^1\text{H}, ^1\text{H}$ -COSY experiment of Kaempferol ( <b>7</b> , DMSO- $d_6$ , 600 MHz).	S24
<b>S22.</b> The $^1\text{H}$ Fingerprint of Isorhamnetin ( <b>8</b> ) in the PERCH .PMS file format.	S25
<b>S23.</b> $^1\text{H}$ NMR spectrum of Isorhamnetin ( <b>8</b> , DMSO- $d_6$ , 600 MHz).	S26
<b>S24.</b> $^1\text{H}, ^1\text{H}$ -COSY experiment of Isorhamnetin ( <b>8</b> , DMSO- $d_6$ , 600 MHz).	S27
<b>S25.</b> The $^1\text{H}$ Fingerprint of Rutin ( <b>9</b> ) in the PERCH .PMS file format.	S28
<b>S26.</b> $^1\text{H}$ NMR spectrum of Rutin ( <b>9</b> , DMSO- $d_6$ , 600 MHz).	S29
<b>S27.</b> $^1\text{H}, ^1\text{H}$ -COSY experiment of Rutin ( <b>9</b> , DMSO- $d_6$ , 600 MHz).	S30

**S28.** Combined  $^1\text{H}$  Fingerprints of nine *Ginkgo biloba* constituents (**1-9**) in the PERCH .PMS file format.

S31

**S29.** Calibration of residual solvent signals using primary reference standards of  $\text{DMSO}_2$  and caffeine as internal calibrants. **(A)** qHNMR spectra of stock solutions of caffeine and  $\text{DMSO}_2$  in  $\text{DMSO}-d_6$  (600 MHz). **(B)** Calibration curves for the determination of residual  $\text{DMSO}-d_5$  content in  $\text{DMSO}-d_6$  lot B.

S34

**S30.** The qHNMR analysis of a ginkgolide B reference material ( $\text{DMSO}-d_6$ , 600 MHz).  $\text{DMSO}_2$  was used as Internal Standard (IS) for quantitative calibration. Using qHNMR, ginkgolide A was readily detected as a very minor (0.43 %) impurity, present at an abundance level where intensities/integrals are comparable to those of  $^{13}\text{C}$  satellite signals of the major component (see also reference #17 in the main text).

S35

**S31.** **(A)** Weight percentage of compounds **1-9** in six Ginkgo samples according to qHNMR measurements. **(B)** Variability of selected botanical markers in commercially available *Ginkgo biloba* preparations, expressed as deviation (y-axis, in %) from the *average* percent content (set to 0% deviation, dotted line) of each of the compounds, **1-9**.

S36

**S32.** The effect of random noise on iterative fitting with PERCH in qHNMR analysis. **(A)** Sections of the qHNMR spectra of caffeine ( $\text{DMSO}-d_6$ , 600 MHz), showing the olefinic proton signals ( $\delta_{\text{H}}$  8.01 ppm). The six qHNMR spectra were acquired using a different number of scans to reach the indicated signal-to-noise ratio (S/N) values. **(B)** Fitting of the signals of the olefinic proton of caffeine at the various S/N levels shown in A: comparison of experimental (*Exp*) and calculated (*Calc*) qHNMR spectra. While the root mean square (RMS) deviations for the iterative fits increase with decreasing S/N, the errors always remain <0.1%, even at a S/N level of ~11:1.

S37

## S1. The $^1\text{H}$ Fingerprint of Ginkgolide A (**1**) in the PERCH .PMS file format.

```
* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "...SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: C:\Users\jgnapo_2\Documents\jgnapo\perch\Ginkgolides\GA\ginkgoa
#$@ Date 9. 3.2011; Time 17: 5:18 ginkgoa.5TH

CHEMICAL SHIFTS(PPM): Similarity% = 28.130
PROTON 2*SPIN=1 SPECIES=1H POPULATION(Y)= 1.00000
OH-3 / 1 6.378972 1*1*1 STAT=Y PRED= 4.856 RANGE= 1.805 WIDTH(Y)= 1.273 RESP(Y)= 1.00000
OH-10 / 1 6.814323 1*1*1 STAT=Y PRED= 4.509 RANGE= 1.630 WIDTH(Y)= 1.565 RESP(Y)= 0.9632
H-7beta / 1 2.043863 1*1*1 STAT=Y PRED= 2.070 RANGE= 0.557 WIDTH(Y)= 2.501 RESP(Y)= 0.8989 HSQC= C14
H-1beta / 1 1.805711 1*1*1 STAT=Y PRED= 1.401 RANGE= 0.302 WIDTH(Y)= 1.882 RESP(Y)= 0.9213 HSQC= C8
H-1alpha/ 1 2.750139 1*1*1 STAT=Y PRED= 2.169 RANGE= 0.387 WIDTH(Y)= 1.927 RESP(Y)= 0.9005 HSQC= C8
H-2 / 1 4.833738 1*1*1 STAT=Y PRED= 4.895 RANGE= 0.262 WIDTH(Y)= 1.798 RESP(Y)= 0.9709 HSQC= C9
H-6 / 1 4.945707 1*1*1 STAT=Y PRED= 4.760 RANGE= 0.247 WIDTH(Y)= 1.956 RESP(Y)= 0.9824 HSQC= C13
H-7alpha/ 1 2.027505 1*1*1 STAT=Y PRED= 2.025 RANGE= 0.472 WIDTH(Y)= 2.358 RESP(Y)= 0.8736 HSQC= C14
H-8 / 1 1.710433 1*1*1 STAT=Y PRED= 1.700 RANGE= 0.222 WIDTH(Y)= 2.173 RESP(Y)= 0.9339 HSQC= C15
H-10 / 1 4.941390 1*1*1 STAT=Y PRED= 4.165 RANGE= 0.392 WIDTH(Y)= 1.780 RESP(Y)= 0.9248 HSQC= C17
H-12 / 1 6.023393 1*1*1 STAT=Y PRED= 5.659 RANGE= 0.335 WIDTH(Y)= 1.498 RESP(Y)= 0.9865 HSQC= C19
H-14 / 1 2.937891 1*1*1 STAT=Y PRED= 2.949 RANGE= 0.387 WIDTH(Y)= 1.519 RESP(Y)= 0.9493 HSQC= C21
H-16 / 1 1.117747 1*1*3 STAT=Y PRED= 1.114 RANGE= 0.252 WIDTH(Y)= 1.687 RESP(Y)= 0.9350 HSQC= C23
t-but / 1 1.017131 1*1*9 STAT=Y PRED= 0.928 RANGE= 0.227 WIDTH(Y)= 2.491 RESP(Y)= 0.9025 HSQC= C25-27

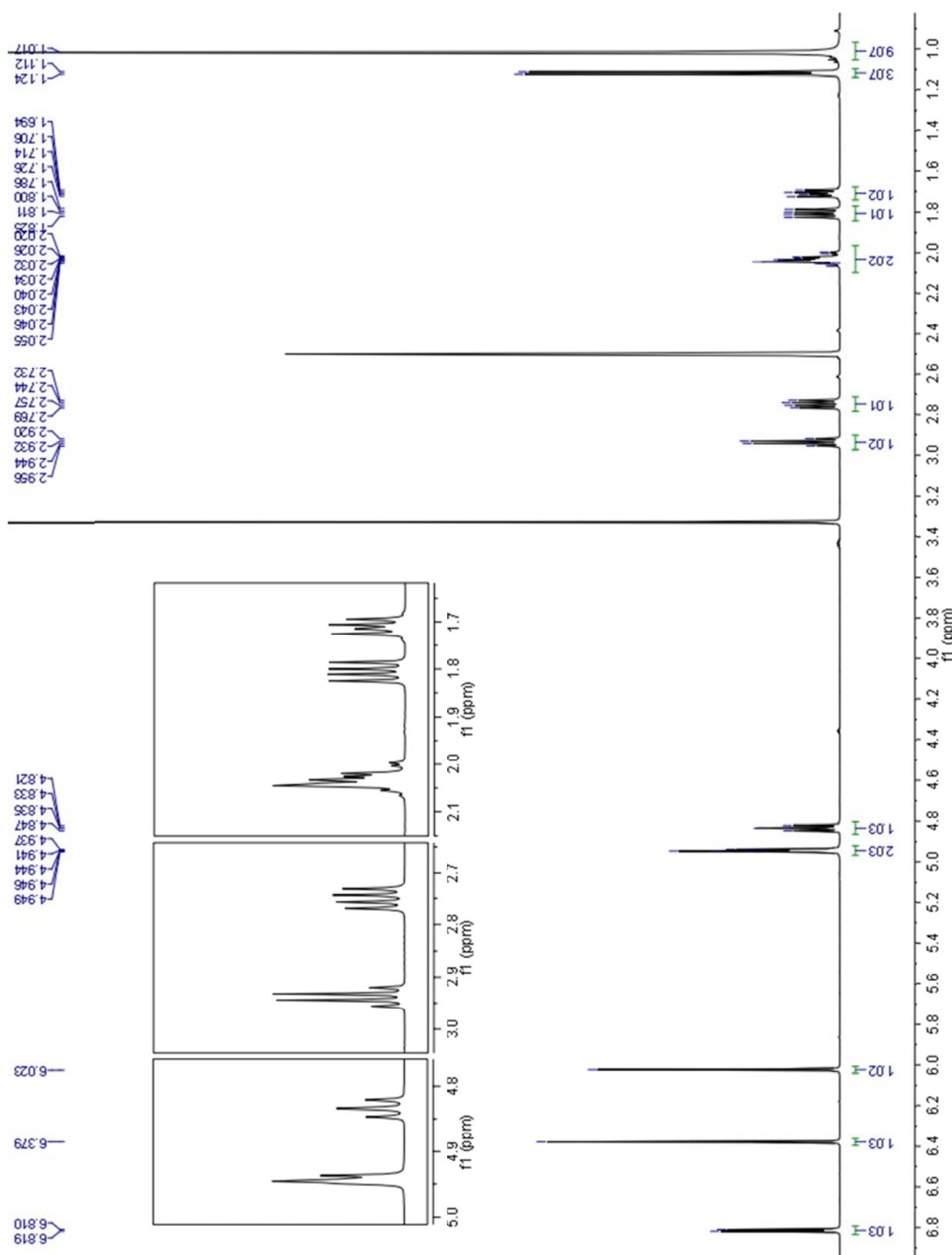
COUPLING CONSTANTS(HZ): Similarity% = 35.913
J6_29      5.1630 J OH-10   H-10    STAT=Y PRED= 5.140 RANGE= 0.750
J12_22     0.6218 J H-7beta H-6    STAT=Y PRED= 1.070 RANGE= 2.000
J12_24    -13.7046 J H-7beta H-7alpha STAT=Y PRED= -12.430 RANGE= 1.000
J12_26     4.5778 J H-7beta H-8    STAT=Y PRED= 4.190 RANGE= 3.000
J14_15    -15.1675 J H-1beta H-1alpha STAT=Y PRED= -12.310 RANGE= 1.000
J14_17     8.2624 J H-1beta H-2    STAT=Y PRED= 8.810 RANGE= 3.000
J15_17     7.2878 J H-1alpha H-2    STAT=Y PRED= 7.110 RANGE= 3.000
J22_24     4.0483 J H-6   H-7alpha STAT=Y PRED= 4.830 RANGE= 3.000
J24_26    14.3434 J H-7alpha H-8    STAT=Y PRED= 13.120 RANGE= 2.200
J35_38     7.1931 J H-14   H-16    STAT=Y PRED= 6.640 RANGE= 0.250

CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
7.50126041 = Left frequency (ppm)
0.00000000 = Right frequency (ppm)
10.000 = Acquisition time (s, for QMTLS)
1.269 = Line-width (for modes D, P & T, 0=use defaults)
0.068609932 = Data-point resolution (Hz)
26.001 = GAUSSIAN (%), 0=use default from INF)
2.174 = Dispersion contribution (%), 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

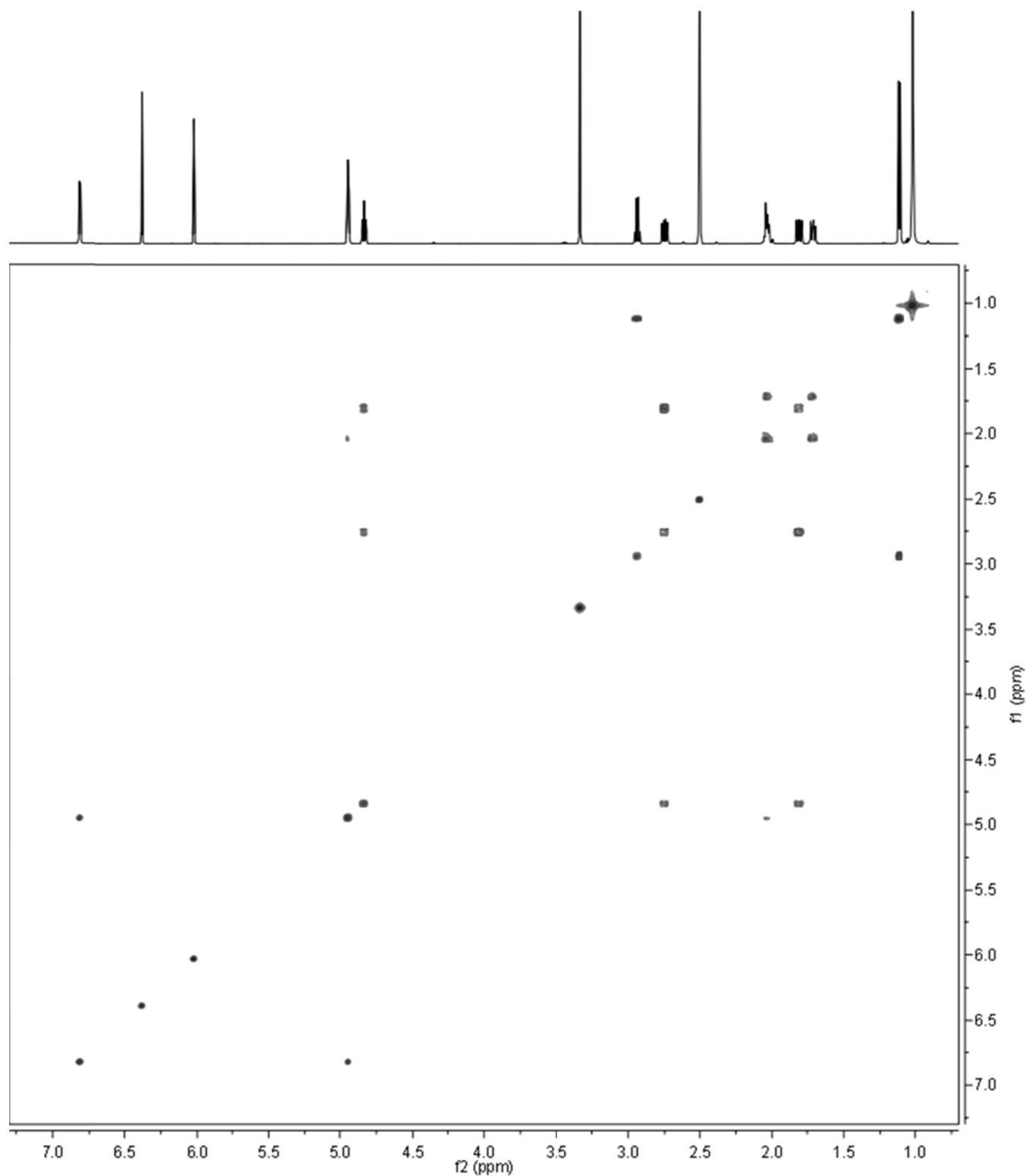
CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

END OF FILE
```

S2.  $^1\text{H}$  NMR spectrum of Ginkgolide A (**1**,  $\text{DMSO}-d_6$ , 600 MHz).



**S3.**  $^1\text{H}$ , $^1\text{H}$ -COSY experiment of Ginkgolide A (**1**, DMSO- $d_6$ , 600 MHz).



#### S4. The $^1\text{H}$ Fingerprint of Ginkgolide B (**2**) in the PERCH .PMS file format.

```
* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: C:\Users\jgnapo_2\Documents\jgnapo\perch\Ginkgolides\GB\ginkgob
#$@ Date 9. 3.2011; Time 17: 8:11 ginkgob.5TH

CHEMICAL SHIFTS(PPM): Similarity% = 11.032
PROTON 2*SPIN=1 SPECIES=LH POPULATION(Y)= 1.00000
OH-3 / 1 6.471908 1*1*1 STAT=Y PRED= 4.913 RANGE= 1.733 WIDTH(Y)= 1.525 RESP(Y)= 1.0000
OH-10 / 1 7.462874 1*1*1 STAT=Y PRED= 4.556 RANGE= 1.695 WIDTH(Y)= 1.929 RESP(Y)= 0.9612
H-7beta / 1 2.137230 1*1*1 STAT=Y PRED= 2.036 RANGE= 0.560 WIDTH(Y)= 2.168 RESP(Y)= 0.9003 HSQC= C14
H-1 / 1 4.042104 1*1*1 STAT=Y PRED= 3.762 RANGE= 0.100 WIDTH(Y)= 1.692 RESP(Y)= 0.9364 HSQC= C8
H-2 / 1 4.642590 1*1*1 STAT=Y PRED= 4.089 RANGE= 0.187 WIDTH(Y)= 1.644 RESP(Y)= 0.9775 HSQC= C9
H-6 / 1 5.303773 1*1*1 STAT=Y PRED= 4.895 RANGE= 0.237 WIDTH(Y)= 1.799 RESP(Y)= 0.9597 HSQC= C13
H-7alpha / 1 1.931786 1*1*1 STAT=Y PRED= 2.041 RANGE= 0.477 WIDTH(Y)= 2.242 RESP(Y)= 0.8904 HSQC= C14
H-8 / 1 1.724094 1*1*1 STAT=Y PRED= 1.670 RANGE= 0.215 WIDTH(Y)= 2.358 RESP(Y)= 0.9326 HSQC= C15
H-10 / 1 5.019748 1*1*1 STAT=Y PRED= 4.165 RANGE= 0.380 WIDTH(Y)= 1.969 RESP(Y)= 0.9334 HSQC= C17
H-12 / 1 6.074154 1*1*1 STAT=Y PRED= 5.659 RANGE= 0.330 WIDTH(Y)= 1.664 RESP(Y)= 0.9898 HSQC= C19
H-14 / 1 2.843287 1*1*1 STAT=Y PRED= 2.949 RANGE= 0.367 WIDTH(Y)= 1.666 RESP(Y)= 0.9457 HSQC= C21
H-16 / 1 1.106901 1*1*3 STAT=Y PRED= 1.102 RANGE= 0.382 WIDTH(Y)= 1.848 RESP(Y)= 0.9294 HSQC= C23
t-butyl / 1 1.028434 1*1*9 STAT=Y PRED= 0.871 RANGE= 0.202 WIDTH(Y)= 2.590 RESP(Y)= 0.8997 HSQC= C25-27
OH-1 / 1 4.918842 1*1*1 STAT=Y PRED= 6.248 RANGE= 2.488 WIDTH(Y)= 1.544 RESP(Y)= 0.9556

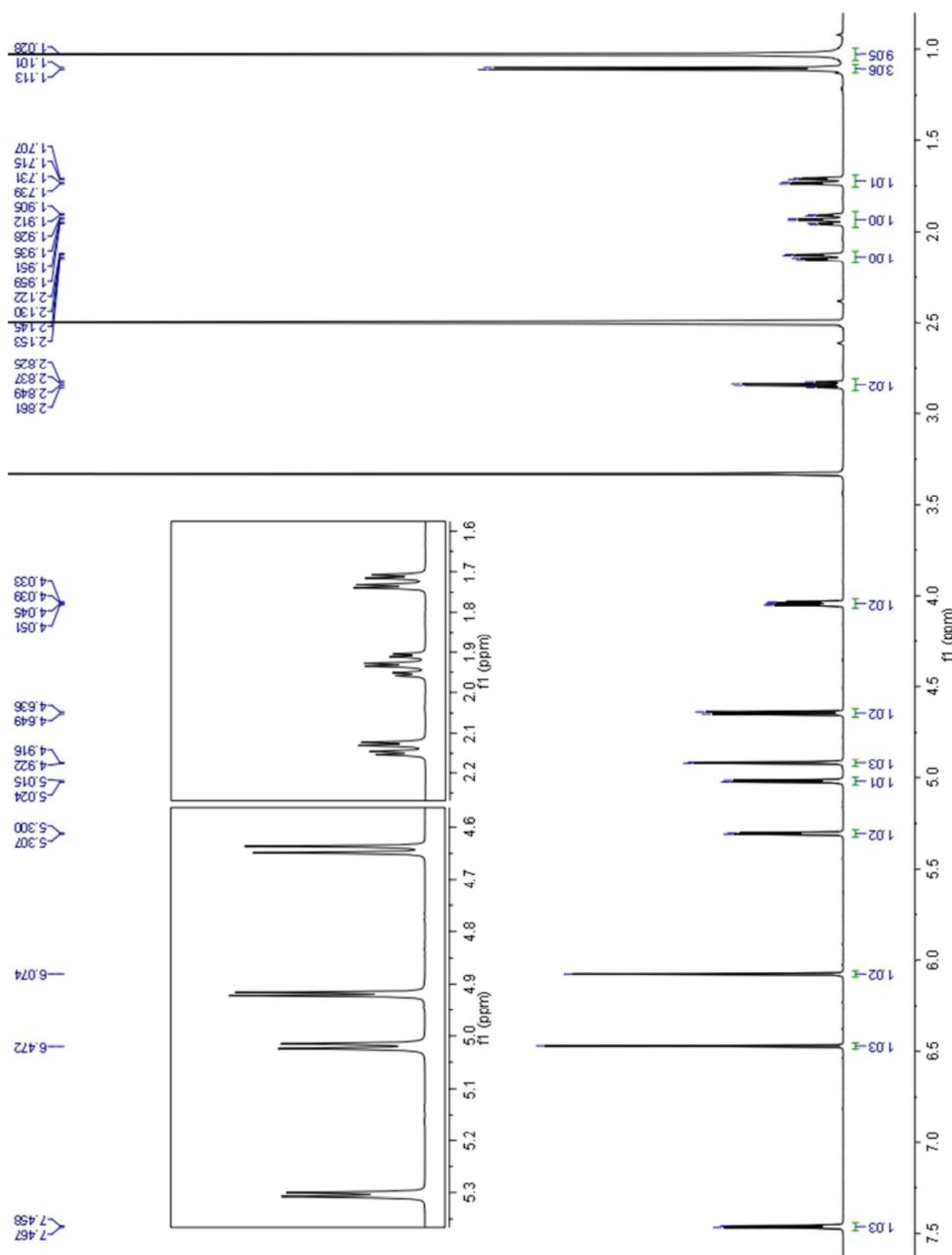
COUPLING CONSTANTS(HZ): Similarity% = 54.436
J6_29 5.5557 J OH-10 H-10 STAT=Y PRED= 5.110 RANGE= 0.750
J12_22 0.8476 J H-7beta H-6 STAT=Y PRED= 1.140 RANGE= 2.200
J12_24 -13.7414 J H-7beta H-7alpha STAT=Y PRED= -12.420 RANGE= 1.000
J12_26 4.6463 J H-7beta H-8 STAT=Y PRED= 4.190 RANGE= 3.000
J14_17 7.4027 J H-1 H-2 STAT=Y PRED= 7.800 RANGE= 3.000
J14_54 3.5631 J H-1 OH-1 STAT=Y PRED= 3.950 RANGE= 0.750
J22_24 4.1428 J H-6 H-7alpha STAT=Y PRED= 4.630 RANGE= 3.000
J24_26 14.3540 J H-7alpha H-8 STAT=Y PRED= 13.130 RANGE= 2.200
J35_38 7.0831 J H-14 H-16 STAT=Y PRED= 6.640 RANGE= 0.250

CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
8.56788059 = Left frequency (ppm)
-0.17374428 = Right frequency (ppm)
10.000 = Acquisition time (s, for QMTLS)
1.426 = Line-width (for modes D, P & T, 0=use defaults)
0.068609932 = Data-point resolution (Hz)
26.703 = GAUSSIAN (%), 0=use default from INF)
3.021 = Dispersion contribution (%), 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

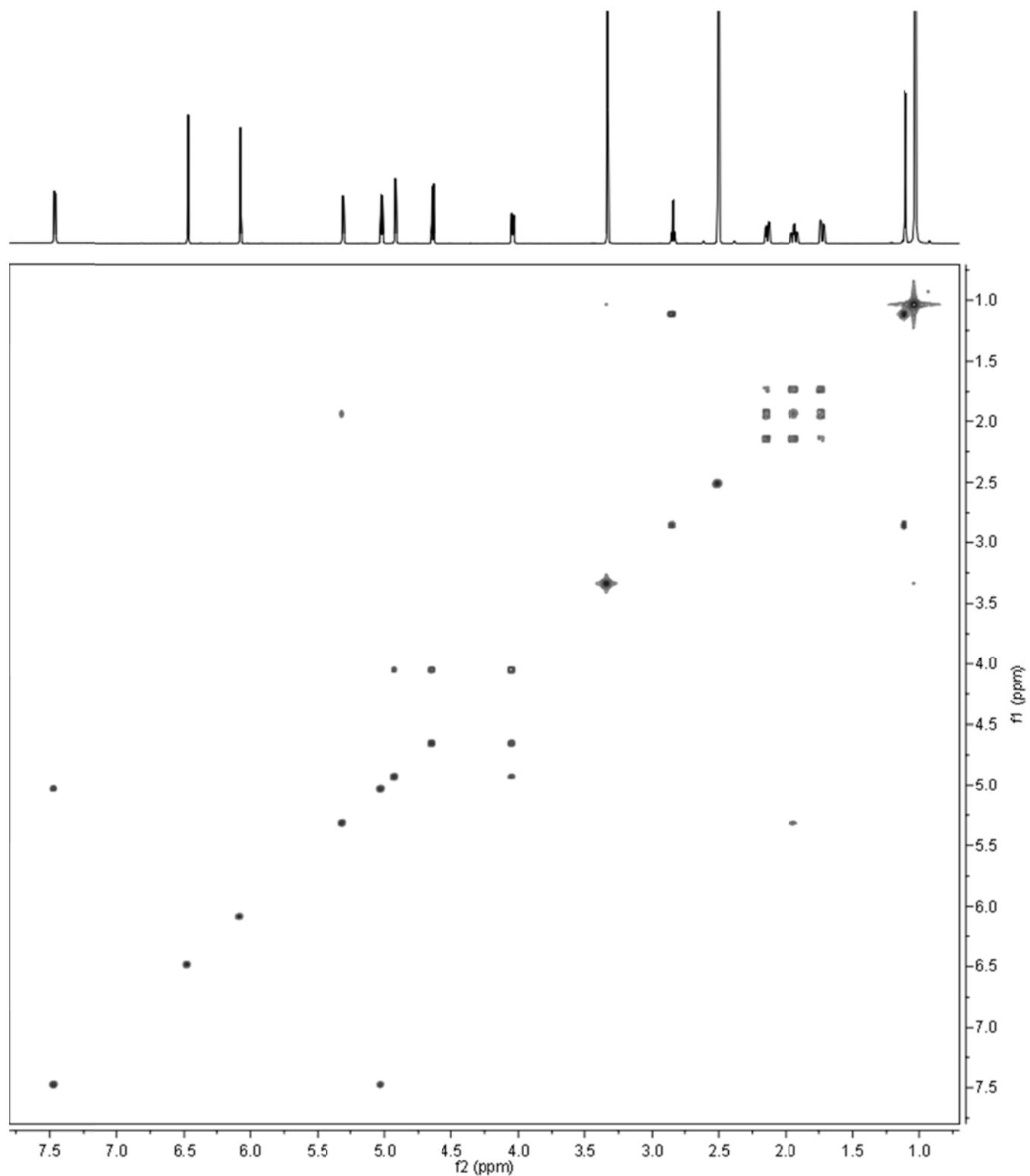
CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

END OF FILE
```

**S5.**  $^1\text{H}$  NMR spectrum of Ginkgolide B (**2**, DMSO- $d_6$ , 600 MHz).



S6.  $^1\text{H}$ , $^1\text{H}$ -COSY experiment of Ginkgolide B (**2**, DMSO-*d*<sub>6</sub>, 600 MHz).



## S7. The $^1\text{H}$ Fingerprint of Ginkgolide C (**3**) in the PERCH .PMS file format.

```

* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: C:\Users\jgnapo_2\Documents\jgnapo\perch\Ginkgolides\GC\ginkgoc
#$@ Date 9. 3.2011; Time 17:10:42 ginkgoc.5TH

CHEMICAL SHIFTS(PPM): Similarity% = 7.110
PROTON 2*SPIN=1H POPULATION(Y)= 1.00000
OH-3 / 1 6.479066 1*1*1 STAT=Y PRED= 5.334 RANGE= 1.700 WIDTH(Y)= 1.736 RESP(Y)= 1.00000
OH-10 / 1 7.538812 1*1*1 STAT=Y PRED= 4.347 RANGE= 1.768 WIDTH(Y)= 2.242 RESP(Y)= 0.9489
OH-7 / 1 5.658435 1*1*1 STAT=Y PRED= 4.985 RANGE= 1.620 WIDTH(Y)= 1.944 RESP(Y)= 0.9704
H-1beta/ 1 3.986275 1*1*1 STAT=Y PRED= 4.316 RANGE= 0.100 WIDTH(Y)= 1.907 RESP(Y)= 0.9449 HSQC= C8
H-2 / 1 4.626842 1*1*1 STAT=Y PRED= 4.089 RANGE= 0.170 WIDTH(Y)= 1.789 RESP(Y)= 0.9761 HSQC= C9
H-6 / 1 4.966384 1*1*1 STAT=Y PRED= 4.852 RANGE= 0.122 WIDTH(Y)= 1.940 RESP(Y)= 0.9965 HSQC= C13
H-7 / 1 4.053494 1*1*1 STAT=Y PRED= 4.559 RANGE= 0.317 WIDTH(Y)= 1.975 RESP(Y)= 0.9273 HSQC= C14
H-8 / 1 1.551161 1*1*1 STAT=Y PRED= 1.808 RANGE= 0.222 WIDTH(Y)= 2.077 RESP(Y)= 0.9481 HSQC= C15
H-10 / 1 4.998365 1*1*1 STAT=Y PRED= 4.165 RANGE= 0.367 WIDTH(Y)= 2.302 RESP(Y)= 0.9280 HSQC= C17
H-12 / 1 6.099451 1*1*1 STAT=Y PRED= 5.659 RANGE= 0.325 WIDTH(Y)= 1.837 RESP(Y)= 0.9848 HSQC= C19
H-14 / 1 2.813690 1*1*1 STAT=Y PRED= 2.949 RANGE= 0.342 WIDTH(Y)= 1.804 RESP(Y)= 0.9418 HSQC= C21
H-16 / 1 1.107483 1*1*3 STAT=Y PRED= 1.103 RANGE= 0.377 WIDTH(Y)= 1.979 RESP(Y)= 0.8915 HSQC= C23
t-butyl/ 1 0.091336 1*1*9 STAT=Y PRED= 0.887 RANGE= 0.225 WIDTH(Y)= 2.733 RESP(Y)= 0.8975 HSQC= C25-27
OH-1 / 1 4.971784 1*1*1 STAT=Y PRED= 5.998 RANGE= 1.965 WIDTH(Y)= 1.840 RESP(Y)= 0.8956

COUPLING CONSTANTS(HZ): Similarity% = 28.377
J6_30      5.6531 J OH-10 H-10      STAT=Y PRED= 4.950 RANGE= 0.750
J13_25     6.2720 J OH-7  H-7       STAT=Y PRED= 4.300 RANGE= 0.500
J15_18     7.2205 J H-1beta H-2     STAT=Y PRED= 7.910 RANGE= 3.000
J15_55     3.6847 J H-1beta OH-1    STAT=Y PRED= 4.300 RANGE= 0.500
J23_25     4.1999 J H-6   H-7       STAT=Y PRED= 4.670 RANGE= 3.000
J25_27     12.3964 J H-7   H-8       STAT=Y PRED= 11.070 RANGE= 2.200
J36_39     7.0914 J H-14 H-16     STAT=Y PRED= 6.640 RANGE= 0.250

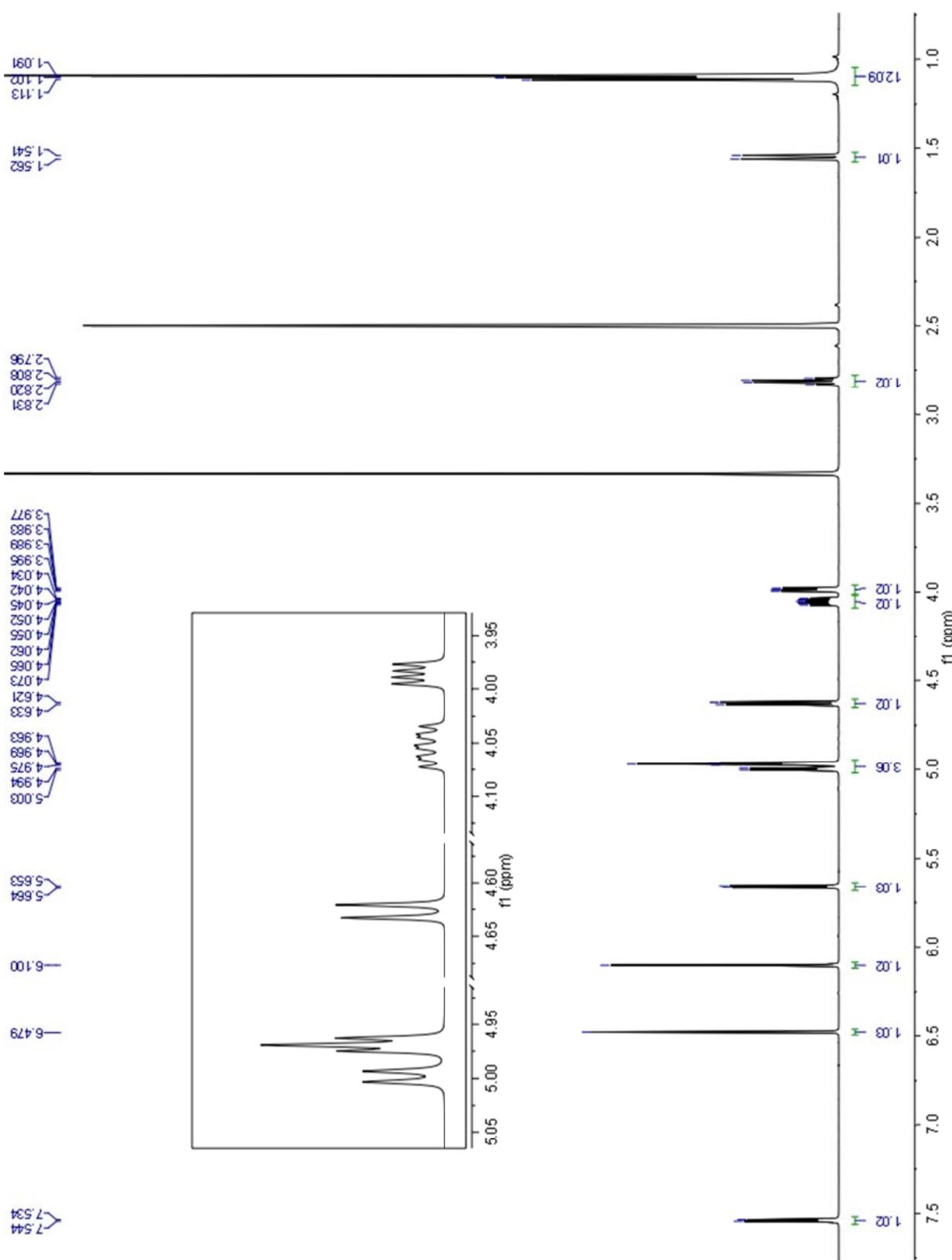
CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
8.89596571 = Left frequency (ppm)
0.00000000 = Right frequency (ppm)
10.000 = Acquisition time (s, for QMTLS)
1.640 = Line-width (for modes D, P & T, 0=use defaults)
0.068609932 = Data-point resolution (Hz)
33.440 = GAUSSIAN (%), 0=use default from INF)
-0.834 = Dispersion contribution (%), 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

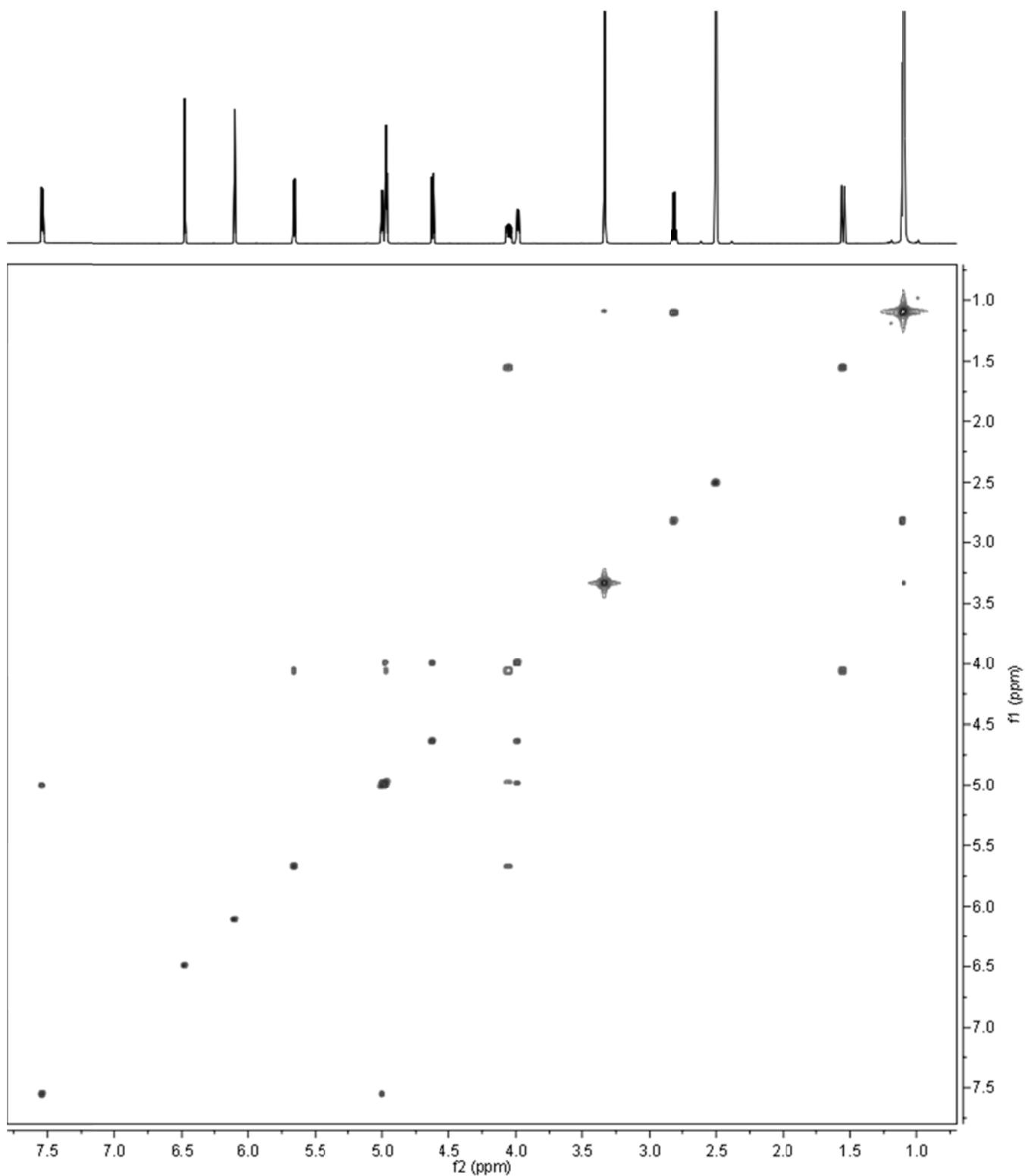
END of FILE

```

**S8.**  $^1\text{H}$  NMR spectrum of Ginkgolide C (**3**,  $\text{DMSO-}d_6$ , 600 MHz).



**S9.**  $^1\text{H}$ , $^1\text{H}$ -COSY experiment of Ginkgolide C (**3**,  $\text{DMSO}-d_6$ , 600 MHz).



## S10. The $^1\text{H}$ Fingerprint of Ginkgolide J (**4**) in the PERCH .PMS file format.

```

* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: C:\Users\jgnapo_2\Documents\jgnapo\perch\Ginkgolides\GJ\ginkgoj
#$æ Date 9. 3.2011; Time 17:14:19 ginkgoj.5TH

CHEMICAL SHIFTS(PPM): Similarity% = 13.626
PROTON 2*SPIN= 1 SPECIES=1H POPULATION(Y)= 1.00000
OH-3 / 1 6.393136 1*1*1 STAT=Y PRED= 4.925 RANGE= 1.775 WIDTH(Y)= 1.498 RESP(Y)= 1.0000
OH-10 / 1 6.908209 1*1*1 STAT=Y PRED= 4.324 RANGE= 1.710 WIDTH(Y)= 1.844 RESP(Y)= 0.9617
OH-7 / 1 5.545649 1*1*1 STAT=Y PRED= 4.759 RANGE= 1.580 WIDTH(Y)= 1.670 RESP(Y)= 0.9709
H-1beta / 1 1.752901 1*1*1 STAT=Y PRED= 1.338 RANGE= 0.287 WIDTH(Y)= 2.106 RESP(Y)= 0.9100 HSQC= C8
H-1alpha/ 1 2.717097 1*1*1 STAT=Y PRED= 2.169 RANGE= 0.377 WIDTH(Y)= 2.200 RESP(Y)= 0.8989 HSQC= C8
H-2 / 1 4.832003 1*1*1 STAT=Y PRED= 4.895 RANGE= 0.232 WIDTH(Y)= 1.942 RESP(Y)= 0.9577 HSQC= C9
H-6 / 1 4.645476 1*1*1 STAT=Y PRED= 4.895 RANGE= 0.110 WIDTH(Y)= 1.872 RESP(Y)= 0.9688 HSQC= C13
H-7 / 1 4.171447 1*1*1 STAT=Y PRED= 4.559 RANGE= 0.325 WIDTH(Y)= 1.862 RESP(Y)= 0.9203 HSQC= C14
H-8 / 1 1.537556 1*1*1 STAT=Y PRED= 1.768 RANGE= 0.212 WIDTH(Y)= 1.979 RESP(Y)= 0.9531 HSQC= C15
H-10 / 1 4.928198 1*1*1 STAT=Y PRED= 4.165 RANGE= 0.390 WIDTH(Y)= 2.064 RESP(Y)= 0.9291 HSQC= C17
H-12 / 1 6.046780 1*1*1 STAT=Y PRED= 5.659 RANGE= 0.332 WIDTH(Y)= 1.684 RESP(Y)= 0.9761 HSQC= C19
H-14 / 1 2.906534 1*1*1 STAT=Y PRED= 2.949 RANGE= 0.362 WIDTH(Y)= 1.711 RESP(Y)= 0.9406 HSQC= C21
H-16 / 1 1.117886 1*1*3 STAT=Y PRED= 1.135 RANGE= 0.260 WIDTH(Y)= 1.906 RESP(Y)= 0.9293 HSQC= C23
t-butyl / 1 1.081520 1*1*9 STAT=Y PRED= 0.894 RANGE= 0.230 WIDTH(Y)= 2.587 RESP(Y)= 0.9000 HSQC= C25-27

COUPLING CONSTANTS(HZ): Similarity% = 17.523
J6_30 5.3487 J OH-10 H-10 STAT=Y PRED= 5.040 RANGE= 0.750
J13_25 6.8614 J OH-7 H-7 STAT=Y PRED= 4.300 RANGE= 0.500
J15_16 -15.1380 J H-1beta H-1alpha STAT=Y PRED= -12.310 RANGE= 1.000
J15_18 8.1701 J H-1beta H-2 STAT=Y PRED= 9.280 RANGE= 3.000
J16_18 7.2493 J H-1alpha H-2 STAT=Y PRED= 7.040 RANGE= 3.000
J23_25 4.1855 J H-6 H-7 STAT=Y PRED= 4.830 RANGE= 3.000
J25_27 12.2977 J H-7 H-8 STAT=Y PRED= 11.060 RANGE= 2.200
J36_39 7.2044 J H-14 H-16 STAT=Y PRED= 6.640 RANGE= 0.250

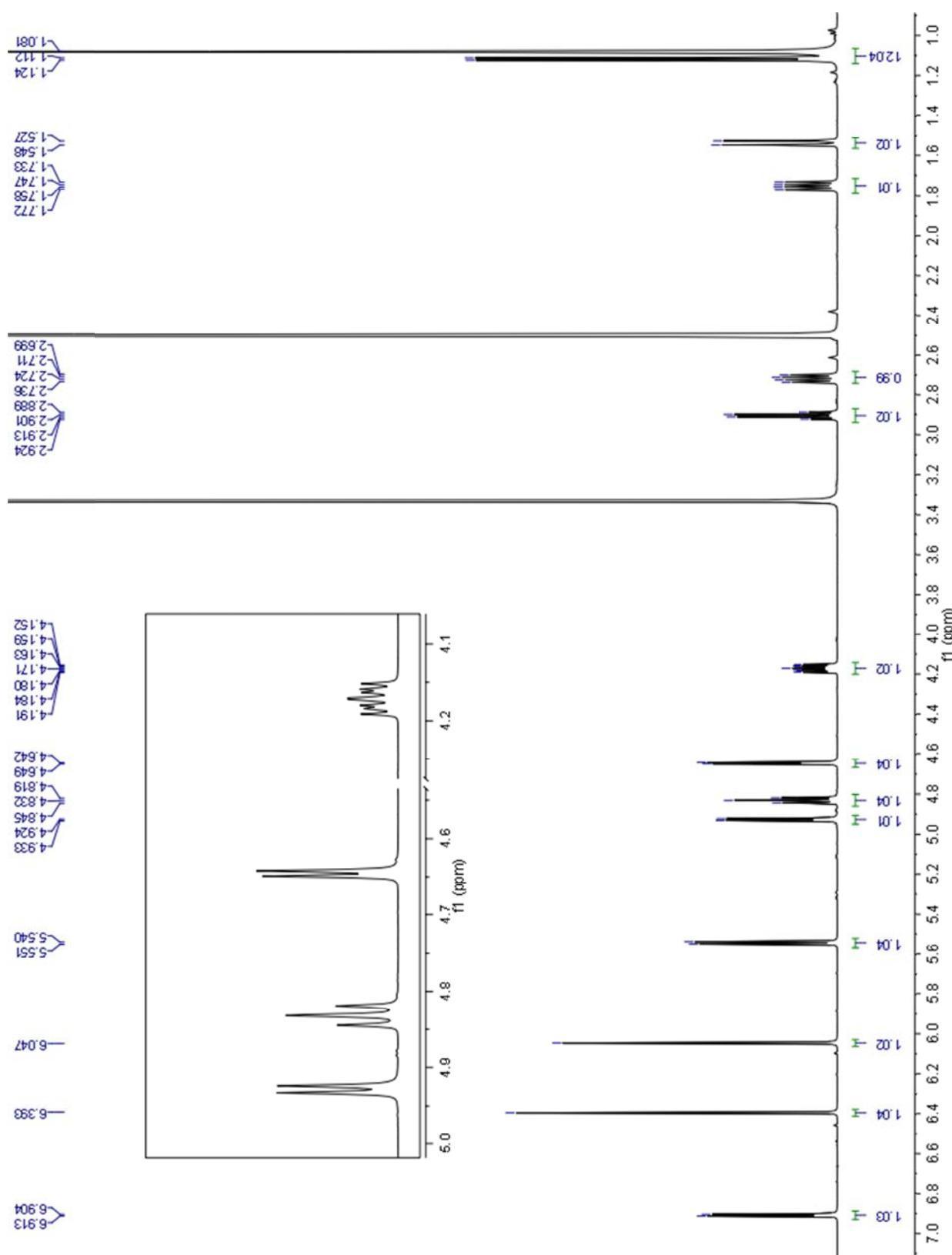
CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
7.83034303 = Left frequency (ppm)
-0.20624264 = Right frequency (ppm)
10.000 = Acquisition time (s, for QMTLS)
1.433 = Line-width (for modes D, P & T, 0=use defaults)
0.068609932 = Data-point resolution (Hz)
25.038 = GAUSSIAN (%), 0=use default from INF)
-2.507 = Dispersion contribution (%), 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

CONSTRAINTS (in equations X0 = 1.0)...use no empty lines
EQUAL H-2 = H-6

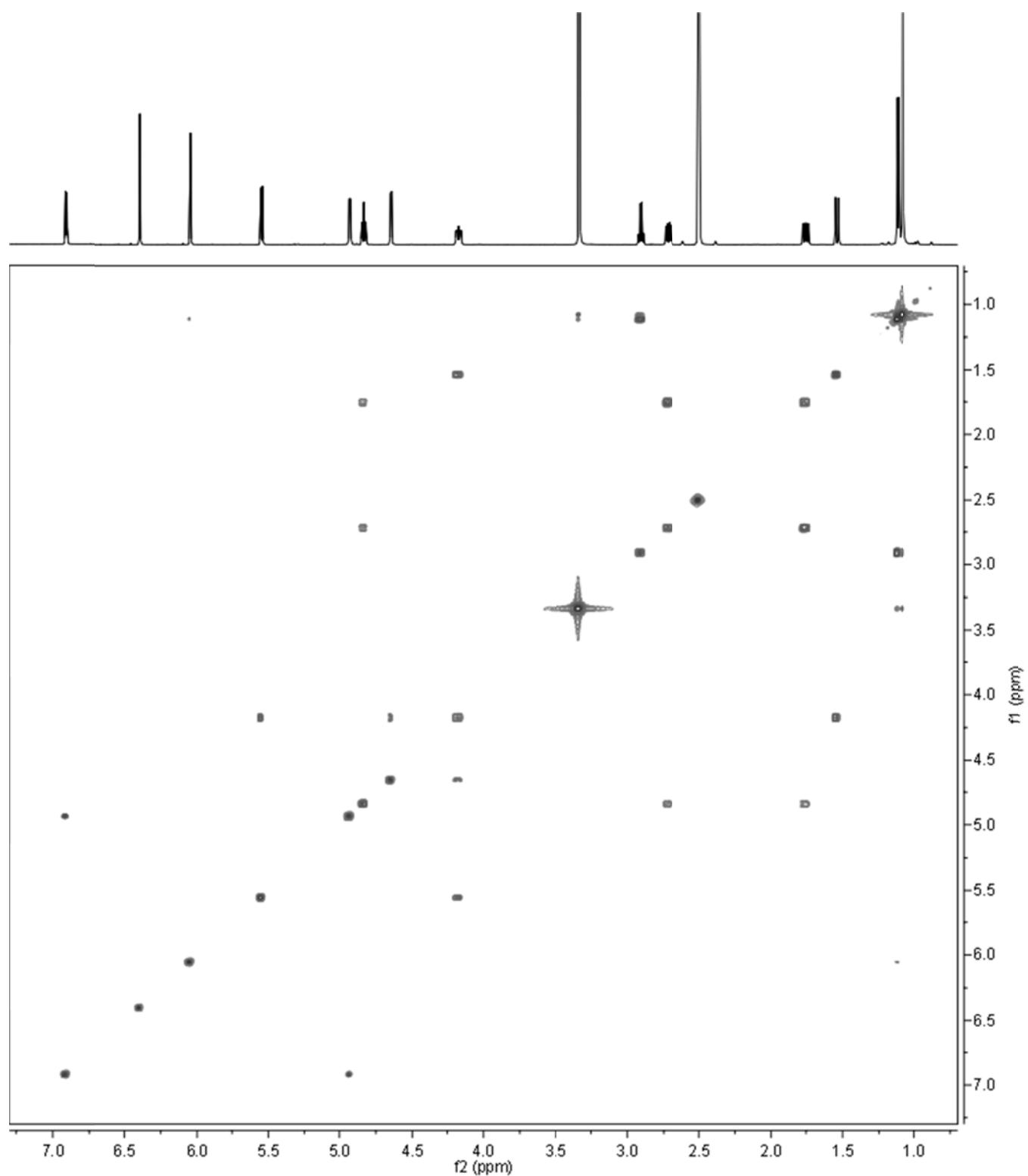
END OF FILE

```

**S11.**  $^1\text{H}$  NMR spectrum of Ginkgolide J (**4**,  $\text{DMSO}-d_6$ , 600 MHz).



**S12.**  $^1\text{H}$ , $^1\text{H}$ -COSY experiment of Ginkgolide J (**4**, DMSO- $d_6$ , 600 MHz).



### S13. The $^1\text{H}$ Fingerprint of Bilobalide (**5**) in the PERCH .PMS file format.

```
* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: C:\Users\jgnapo_2\Documents\jgnapo\perch\Ginkgolides\BB\bb
#$æ Date 9. 3.2011; Time 17: 1:31 bb.5TH

CHEMICAL SHIFTS(PPM): Similarity% = 12.410
PROTON 2*SPIN= 1 SPECIES=1H POPULATION(Y)= 1.00000
OH-10 / 1 7.253250 1*1*1 STAT=Y PRED= 4.828 RANGE= 1.750 WIDTH(Y)= 1.600 RESP(Y)= 0.9729
H-7beta / 1 2.077084 1*1*1 STAT=Y PRED= 1.896 RANGE= 0.357 WIDTH(Y)= 1.891 RESP(Y)= 0.9170 HSQC= C14
H-6 / 1 4.916794 1*1*1 STAT=Y PRED= 4.888 RANGE= 0.282 WIDTH(Y)= 1.768 RESP(Y)= 0.9697 HSQC= C13
H-7alpha/ 1 2.561866 1*1*1 STAT=Y PRED= 2.518 RANGE= 0.282 WIDTH(Y)= 1.976 RESP(Y)= 0.8993 HSQC= C14
H-10 / 1 5.149952 1*1*1 STAT=Y PRED= 4.165 RANGE= 0.370 WIDTH(Y)= 1.774 RESP(Y)= 0.9421 HSQC= C17
H-12 / 1 6.274102 1*1*1 STAT=Y PRED= 5.659 RANGE= 0.102 WIDTH(Y)= 1.500 RESP(Y)= 1.0000 HSQC= C19
t-Butyl / 1 1.034063 1*1*9 STAT=Y PRED= 0.914 RANGE= 0.237 WIDTH(Y)= 2.678 RESP(Y)= 0.9023 HSQC= C25-27
OH-8 / 1 5.403089 1*1*1 STAT=Y PRED= 3.979 RANGE= 1.923 WIDTH(Y)= 1.508 RESP(Y)= 0.9843
H-1beta / 1 2.774028 1*1*1 STAT=Y PRED= 2.359 RANGE= 0.370 WIDTH(Y)= 1.686 RESP(Y)= 0.9603 HSQC= #27
H-1alpha/ 1 2.897044 1*1*1 STAT=Y PRED= 2.405 RANGE= 0.462 WIDTH(Y)= 1.740 RESP(Y)= 0.9580 HSQC= #27

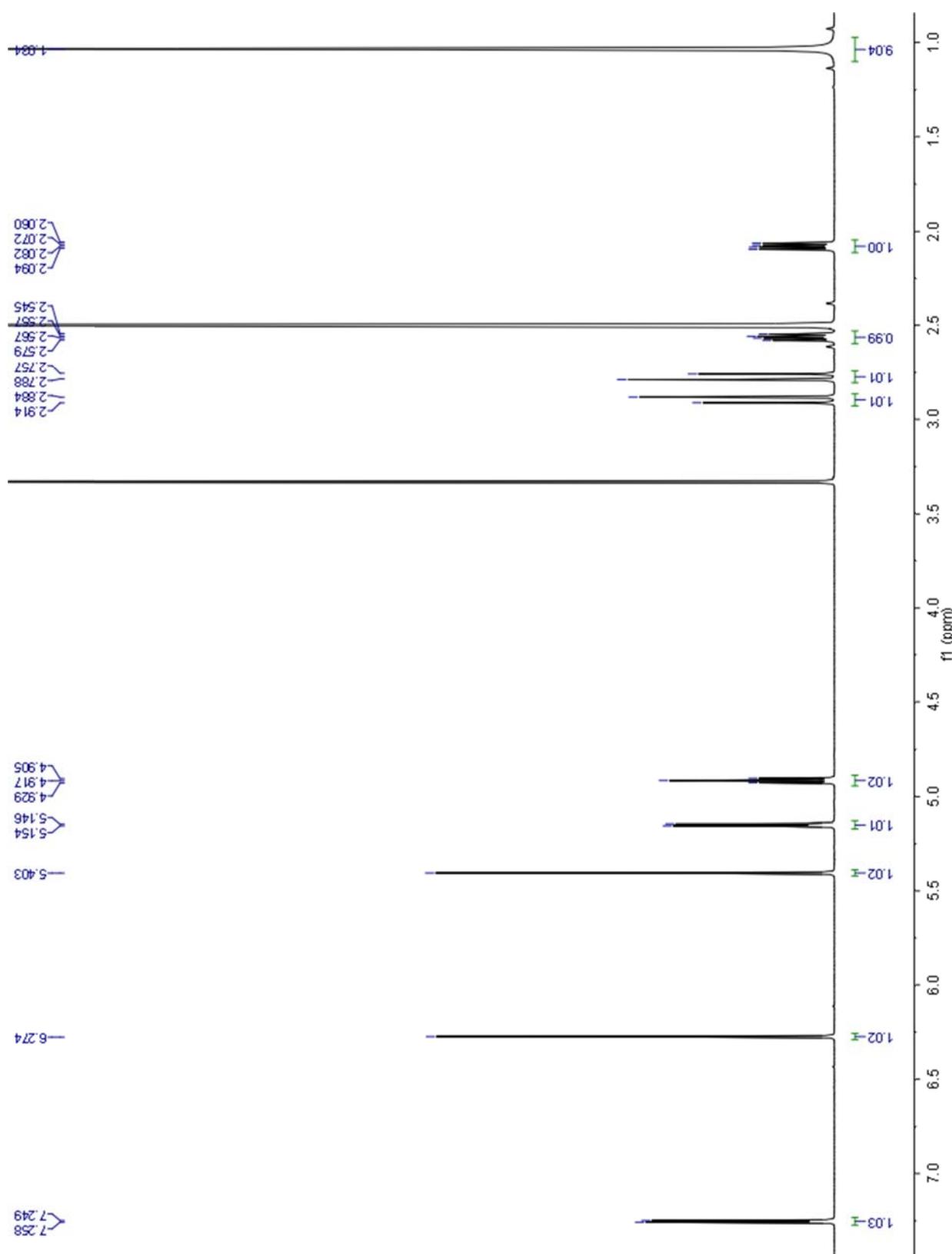
COUPLING CONSTANTS(HZ): Similarity% = 64.488
J4_18 5.1195 J OH-10 H-10 STAT=Y PRED= 4.890 RANGE= 0.750
J7_11 6.8503 J H-7beta H-6 STAT=Y PRED= 9.550 RANGE= 3.000
J7_13 -13.3259 J H-7beta H-7alpha STAT=Y PRED= -12.140 RANGE= 1.000
J11_13 7.0969 J H-6 H-7alpha STAT=Y PRED= 6.600 RANGE= 3.000
J40_41 -18.1406 J H-1beta H-1alpha STAT=Y PRED= -15.860 RANGE= 2.000

CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
8.04426687 = Left frequency (ppm)
0.00000000 = Right frequency (ppm)
10.000 = Acquisition time (s, for QMTLS)
1.505 = Line-width (for modes D, P & T, 0=use defaults)
0.068609932 = Data-point resolution (Hz)
25.060 = GAUSSIAN (%), 0=use default from INF)
2.716 = Dispersion contribution (%), 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

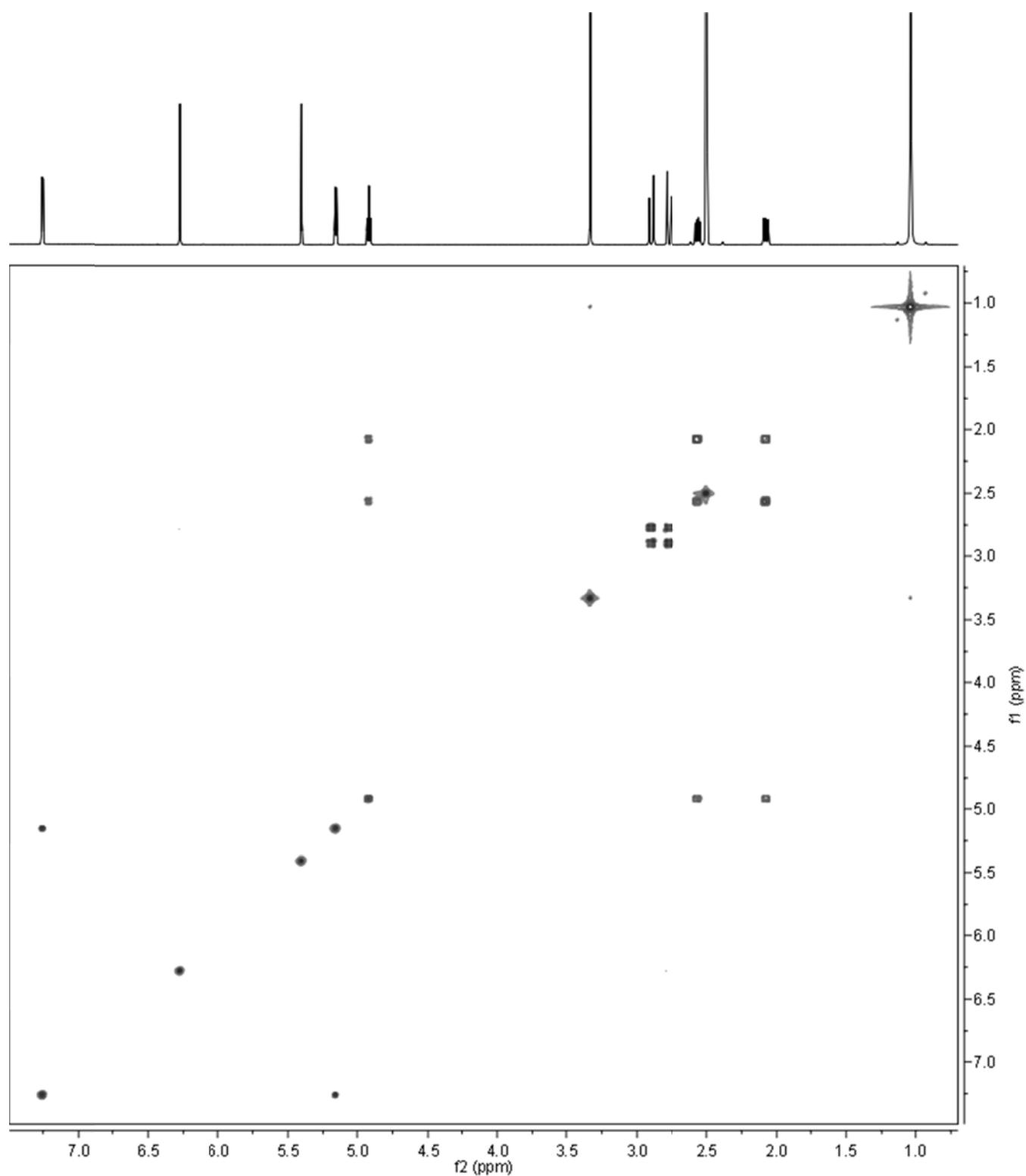
CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

END of FILE
```

**S14.**  $^1\text{H}$  NMR spectrum of Bilobalide (**5**, DMSO- $d_6$ , 600 MHz).



**S15.**  $^1\text{H}$ , $^1\text{H}$ -COSY experiment of Bilobalide (**5**, DMSO- $d_6$ , 600 MHz).



## S16. The $^1\text{H}$ Fingerprint of Quercetin (**6**) in the PERCH .PMS file format.

```
* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: C:\Users\jgnapo_2\Documents\users\jgnapo\perch\Ginkgo\Flavonoids\
#$@ Date 20.10.2011; Time 16: 2:48 perch.3rd

CHEMICAL SHIFTS(PPM) :
PROTON   2*SPIN= 1 SPECIES=1H   POPULATION(Y)=    1.00000
H6 / 1    6.183050  1*1*1   STAT=Y  PRED= 6.298 RANGE= 0.242 WIDTH(Y)= 1.416 RESP(Y)= 1.0000 HSQC= C2
H8 / 1    6.403402  1*1*1   STAT=Y  PRED= 6.461 RANGE= 0.172 WIDTH(Y)= 1.407 RESP(Y)= 0.9861 HSQC= C4
OH7 / 1   10.783504  1*1*1   STAT=Y  PRED= 10.896 RANGE= 1.000 WIDTH(Y)= 1.739 RESP(Y)= 0.9816
OH5 / 1   12.494440  1*1*1   STAT=Y  PRED= 13.379 RANGE= 0.850 WIDTH(Y)= 1.942 RESP(Y)= 0.9532
H6' / 1   7.536549  1*1*1   STAT=Y  PRED= 7.627 RANGE= 0.265 WIDTH(Y)= 1.496 RESP(Y)= 0.9718 HSQC= C14
H5' / 1   6.880921  1*1*1   STAT=Y  PRED= 6.929 RANGE= 0.197 WIDTH(Y)= 1.406 RESP(Y)= 0.9618 HSQC= C15
H2' / 1   7.674662  1*1*1   STAT=Y  PRED= 7.698 RANGE= 0.247 WIDTH(Y)= 1.416 RESP(Y)= 0.9741 HSQC= C18
OH3'/ 1   9.374357  1*1*1   STAT=Y  PRED= 9.494 RANGE= 1.215 WIDTH(Y)= 1.780 RESP(Y)= 0.9566
OH4'/ 1   9.311783  1*1*1   STAT=Y  PRED= 9.682 RANGE= 1.315 WIDTH(Y)= 2.880 RESP(Y)= 0.9305
OH3 / 1   9.597692  1*1*1   STAT=Y  PRED= 9.115 RANGE= 1.115 WIDTH(Y)= 2.732 RESP(Y)= 0.9341

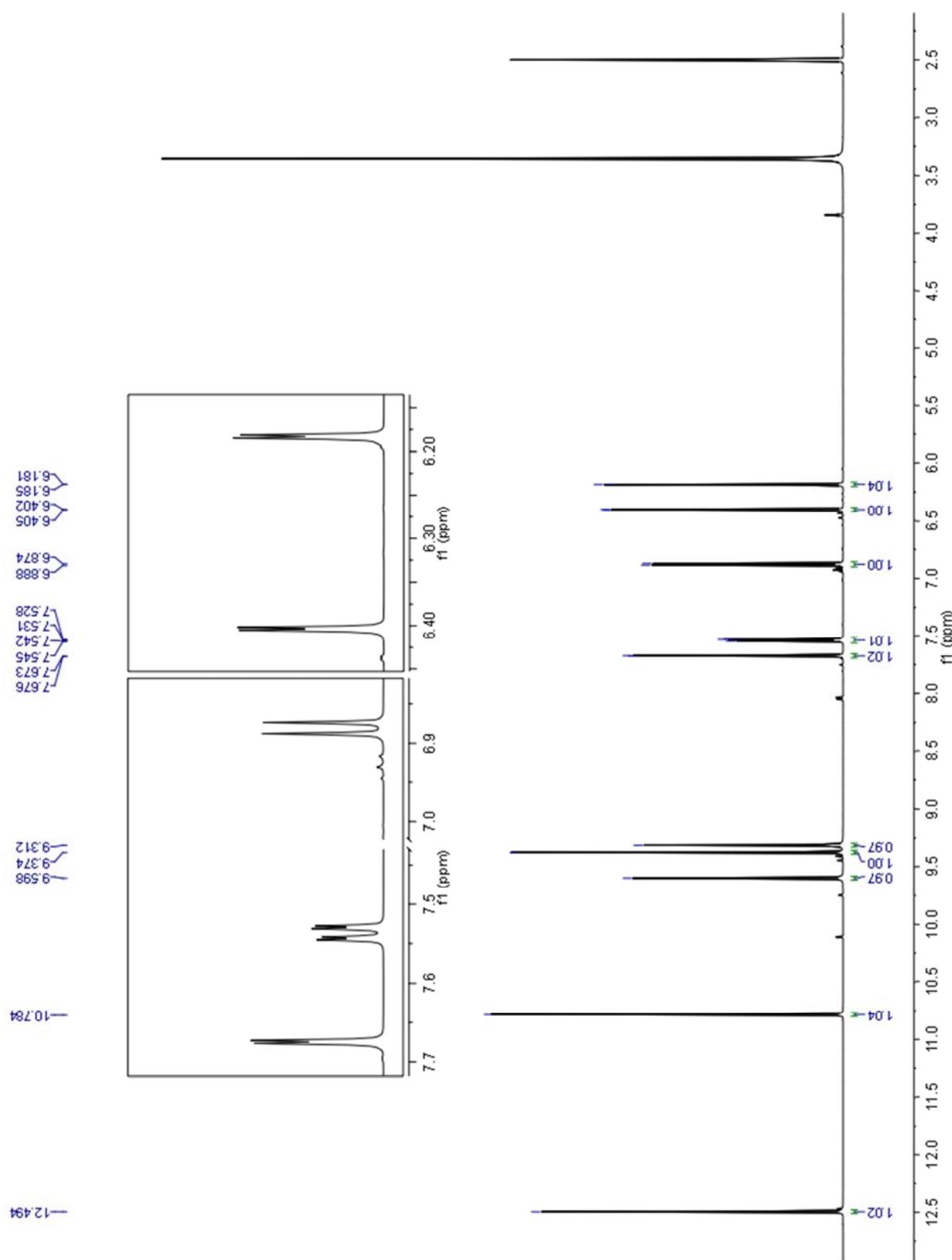
COUPLING CONSTANTS(HZ) :
J22_23     2.0315   J H6    H8      STAT=Y  PRED= 2.110 RANGE= 0.890
J27_28     8.4728   J H6'   H5'     STAT=Y  PRED= 8.380 RANGE= 0.500
J27_29     2.2019   J H6'   H2'     STAT=Y  PRED= 2.080 RANGE= 0.800
J28_29     0.4659   J H5'   H2'     STAT=Y  PRED= 0.460 RANGE= 0.320

CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
14.00343391 = Left frequency (ppm)
0.00000000 = Right frequency (ppm)
0.000 = Acquisition time (s, for QMTLS)
1.402 = Line-width (for modes D, P & T, 0=use defaults)
0.068609932 = Data-point resolution (Hz)
37.540 = GAUSSIAN (%), 0=use default from INF)
5.203 = Dispersion contribution (%), 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

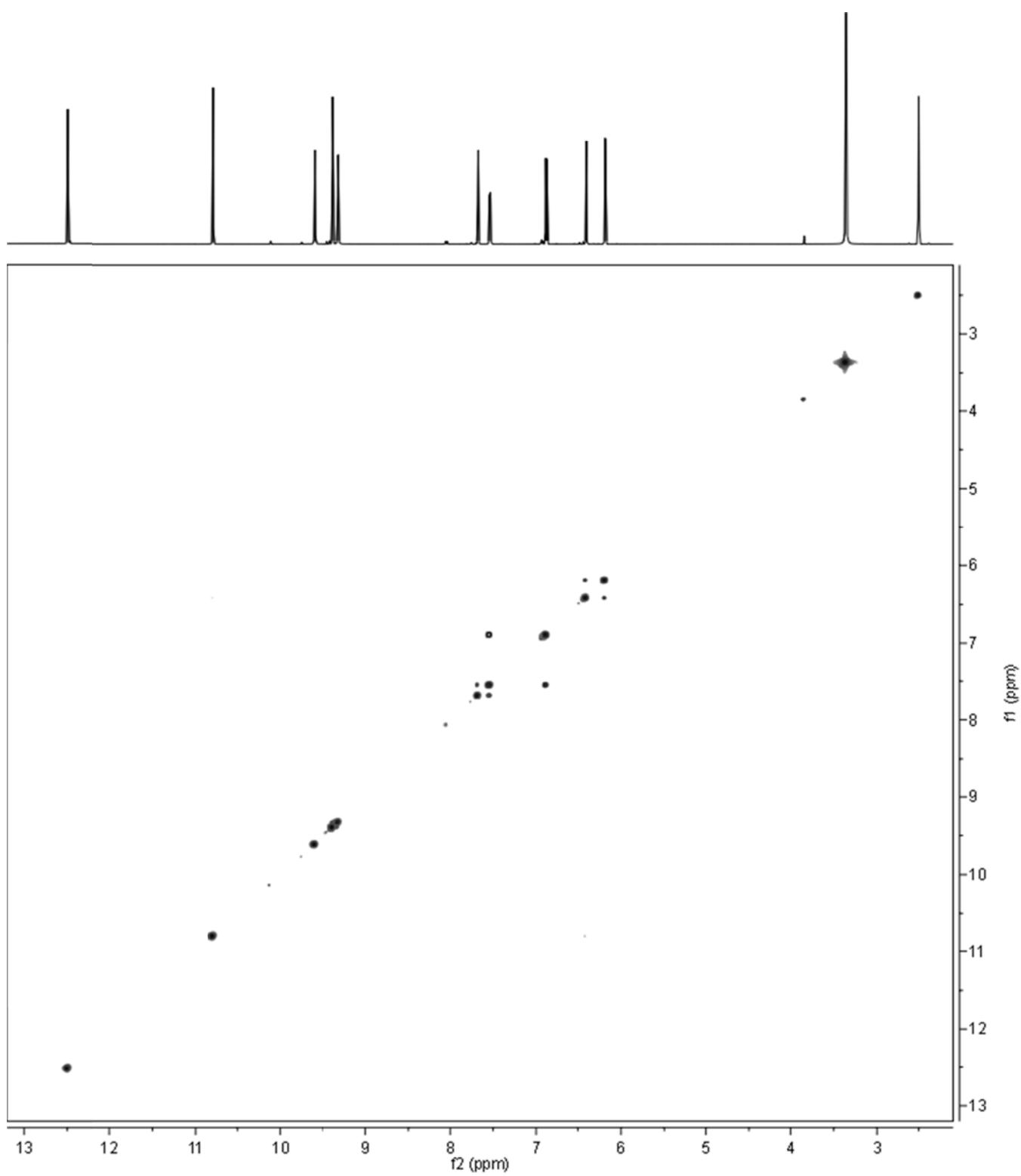
CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

END OF FILE
```

S17.  $^1\text{H}$  NMR spectrum of Quercetin (**6**,  $\text{DMSO}-d_6$ , 600 MHz).



**S18.**  $^1\text{H}$ , $^1\text{H}$ -COSY experiment of Quercetin (**6**, DMSO- $d_6$ , 600 MHz).



## S19. The $^1\text{H}$ Fingerprint of Kaempferol (7) in the PERCH .PMS file format.

```
* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: C:\Users\jgnapo_2\Documents\users\jgnapo\perch\Ginkgo\Flavonoids\
#$æ Date 20.10.2011; Time 17: 5:18 perch.3rd

CHEMICAL SHIFTS(PPM):
PROTON 2*SPIN= 1 SPECIES=1H   POPULATION(Y)= 1.00000
H3'_5' / 1    6.922821 1*2*1  STAT=Y  PRED= 6.925 RANGE= 0.177 WIDTH(Y)= 1.511 RESP(Y)= 0.9770 HSQC= C15_17
H6     / 1    6.186969 1*1*1  STAT=Y  PRED= 6.295 RANGE= 0.252 WIDTH(Y)= 1.406 RESP(Y)= 1.0000 HSQC= C2
H8     / 1    6.435905 1*1*1  STAT=Y  PRED= 6.468 RANGE= 0.172 WIDTH(Y)= 1.404 RESP(Y)= 0.9947 HSQC= C4
OH7    / 1    10.788285 1*1*1  STAT=Y  PRED= 10.903 RANGE= 0.985 WIDTH(Y)= 17.150 RESP(Y)= 0.8071
OH5    / 1    12.484003 1*1*1  STAT=Y  PRED= 13.446 RANGE= 0.850 WIDTH(Y)= 1.739 RESP(Y)= 0.9625
H2'_6' / 1    8.041525 1*2*1  STAT=Y  PRED= 8.080 RANGE= 0.252 WIDTH(Y)= 1.495 RESP(Y)= 0.9722 HSQC= C14_18
OH4'   / 1    9.408526 1*1*1  STAT=Y  PRED= 9.807 RANGE= 1.500 WIDTH(Y)= 11.480 RESP(Y)= 0.8274
OH3    / 1    10.109684 1*1*1  STAT=Y  PRED= 9.155 RANGE= 1.070 WIDTH(Y)= 11.760 RESP(Y)= 0.8250

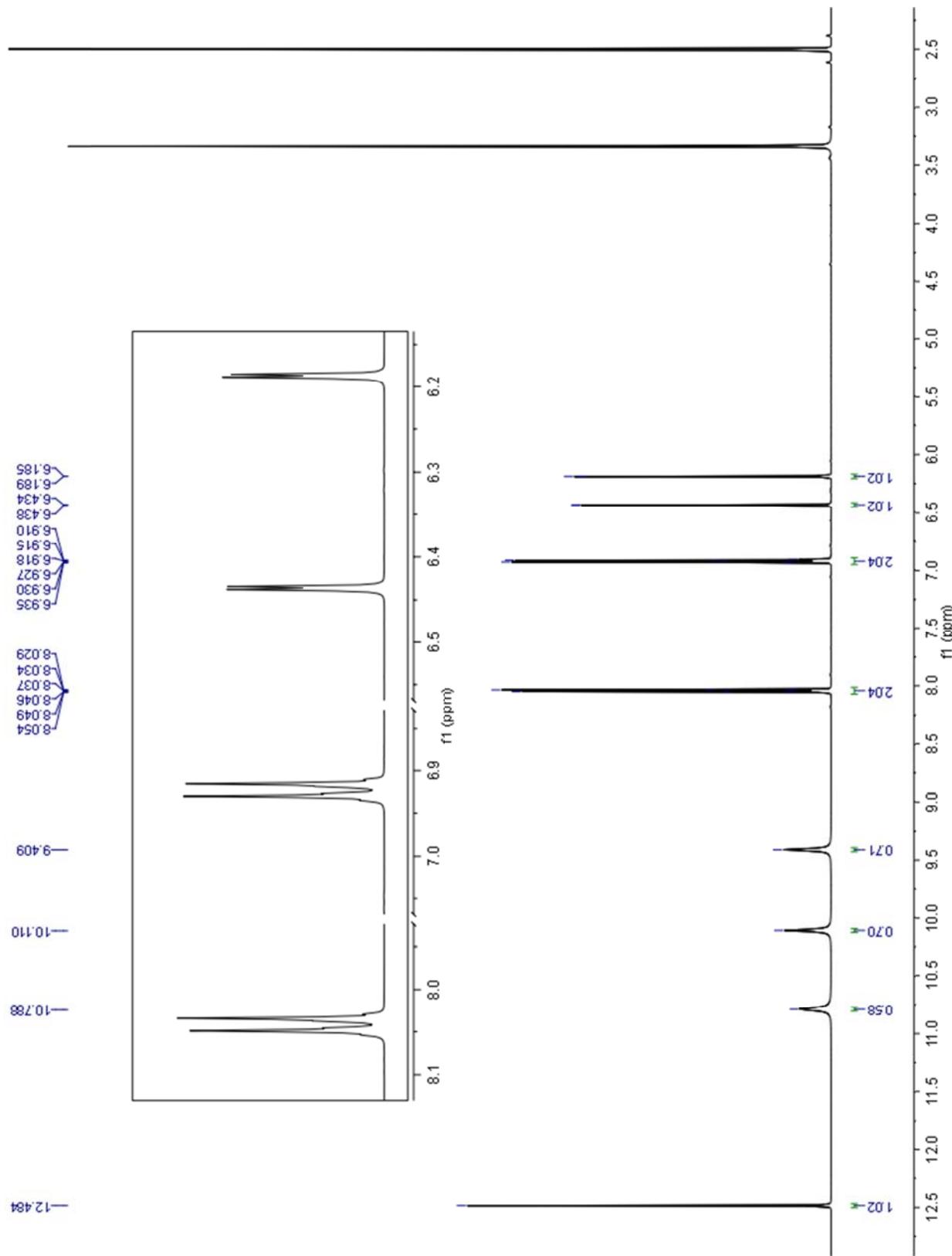
COUPLING CONSTANTS(HZ):
J21_28      2.4778  J H3'_5' H3'_5'  STAT=Y  PRED= 2.710 RANGE= 0.700
J21_27      0.2690  J H2'_6' H3'_5'  STAT=Y  PRED= 0.460 RANGE= 0.320
J21_29      8.7801  J H3'_5' H2'_6'  STAT=Y  PRED= 8.530 RANGE= 0.440
J22_23      2.0325  J H6   H8      STAT=Y  PRED= 2.110 RANGE= 0.890
J29_27      2.4747  J H2'_6' H2'_6'  STAT=Y  PRED= 2.380 RANGE= 0.700

CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
13.07908736 = Left frequency (ppm)
0.00000000 = Right frequency (ppm)
0.000 = Acquisition time (s, for QMTLS)
1.391 = Line-width (for modes D, P & T, 0=use defaults)
0.068609932 = Data-point resolution (Hz)
47.309 = GAUSSIAN (%), 0=use default from INF)
6.174 = Dispersion contribution (%), 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

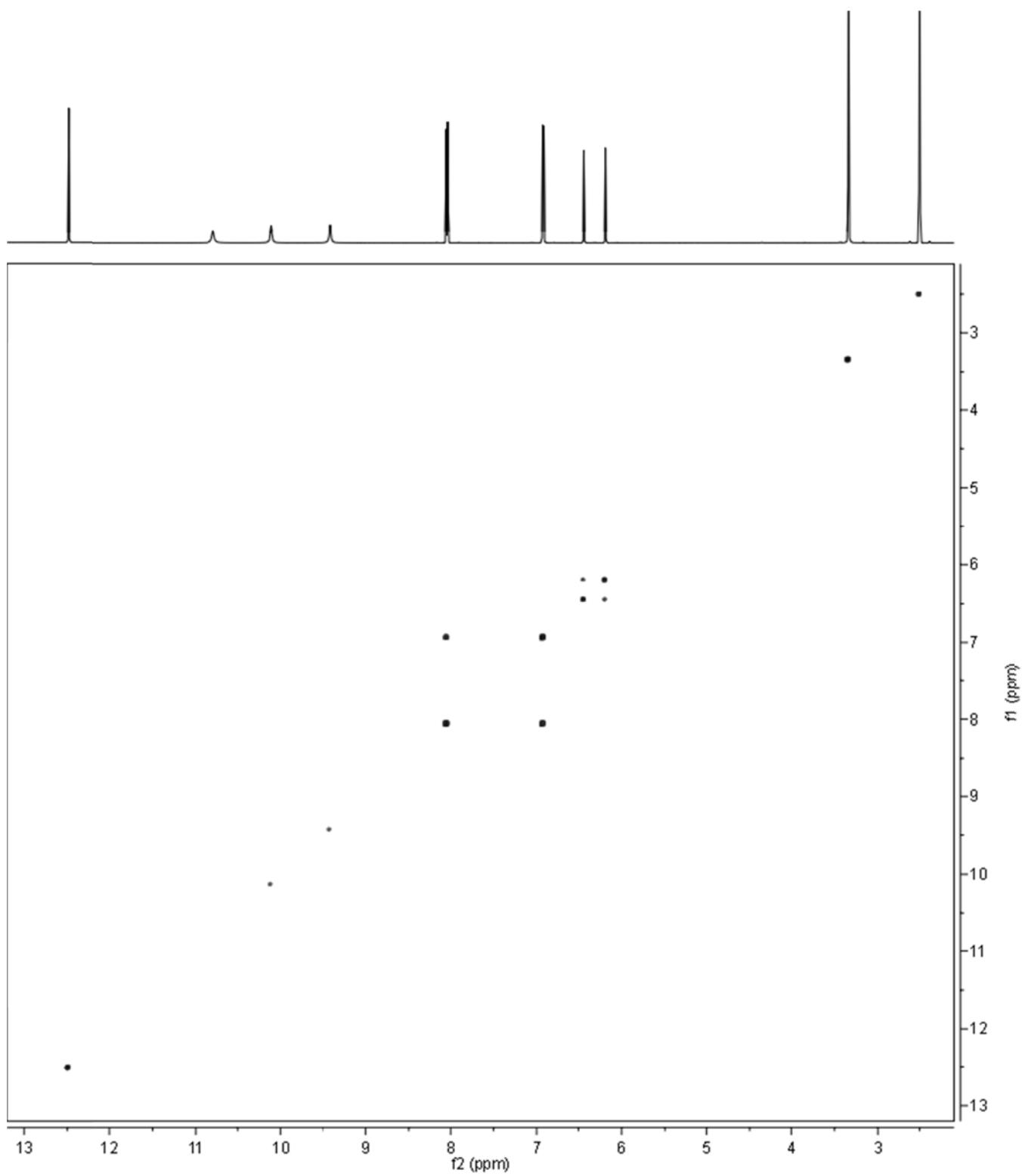
CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

END of FILE
```

S20.  $^1\text{H}$  NMR spectrum of Kaempferol (**7**,  $\text{DMSO}-d_6$ , 600 MHz).



**S21.**  $^1\text{H}$ , $^1\text{H}$ -COSY experiment of Kaempferol (**7**, DMSO- $d_6$ , 600 MHz).



**S22.** The  $^1\text{H}$  Fingerprint of Isorhamnetin (**8**) in the PERCH .PMS file format.

```

* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: c:\users\jgnapo_2\documents\users\jgnapo\perch\ginkgo\flavonoids\
#$@ Date 20.10.2011; Time 16:43:26 perch.3rd

CHEMICAL SHIFTS(PPM) :
PROTON   2*SPIN= 1 SPECIES=1H   POPULATION(Y)=    1.00000
H6 / 1      6.192379  1*1*1  STAT=Y  PRED=  6.306 RANGE=  0.242 WIDTH(Y)=  1.456 RESP(Y)=  0.9979 HSQC= C2
H8 / 1      6.476736  1*1*1  STAT=Y  PRED=  6.462 RANGE=  0.182 WIDTH(Y)=  1.446 RESP(Y)=  0.9882 HSQC= C4
OH7 / 1     10.773796  1*1*1  STAT=Y  PRED= 10.912 RANGE= 1.015 WIDTH(Y)= 2.985 RESP(Y)= 0.9504
OH5 / 1     12.467391  1*1*1  STAT=Y  PRED= 13.376 RANGE= 0.855 WIDTH(Y)= 1.433 RESP(Y)= 1.0000
H6' / 1     7.687450  1*1*1  STAT=Y  PRED= 7.638 RANGE= 0.297 WIDTH(Y)= 1.538 RESP(Y)= 0.9733 HSQC= C14
H5' / 1     6.936966  1*1*1  STAT=Y  PRED= 6.904 RANGE= 0.242 WIDTH(Y)= 1.475 RESP(Y)= 0.9759 HSQC= C15
H2' / 1     7.752072  1*1*1  STAT=Y  PRED= 7.676 RANGE= 0.305 WIDTH(Y)= 1.697 RESP(Y)= 0.9921 HSQC= C18
OH4' / 1    9.445056  1*1*1  STAT=Y  PRED= 9.712 RANGE= 0.752 WIDTH(Y)= 2.159 RESP(Y)= 0.9537
OH3 / 1     9.747292  1*1*1  STAT=Y  PRED= 9.072 RANGE= 1.125 WIDTH(Y)= 2.173 RESP(Y)= 0.9713
OMe / 1     3.840464  1*1*3  STAT=Y  PRED= 3.899 RANGE= 0.132 WIDTH(Y)= 1.727 RESP(Y)= 0.9945 HSQC= H20

COUPLING CONSTANTS(HZ) :
J22_23      2.0139   J H6    H8      STAT=Y  PRED= 2.110 RANGE= 0.890
J27_28      8.4712   J H6'   H5'    STAT=Y  PRED= 8.340 RANGE= 0.500
J27_29      2.0570   J H6'   H2'    STAT=Y  PRED= 2.080 RANGE= 0.800
J28_29      0.2967   J H5'   H2'    STAT=Y  PRED= 0.460 RANGE= 0.320

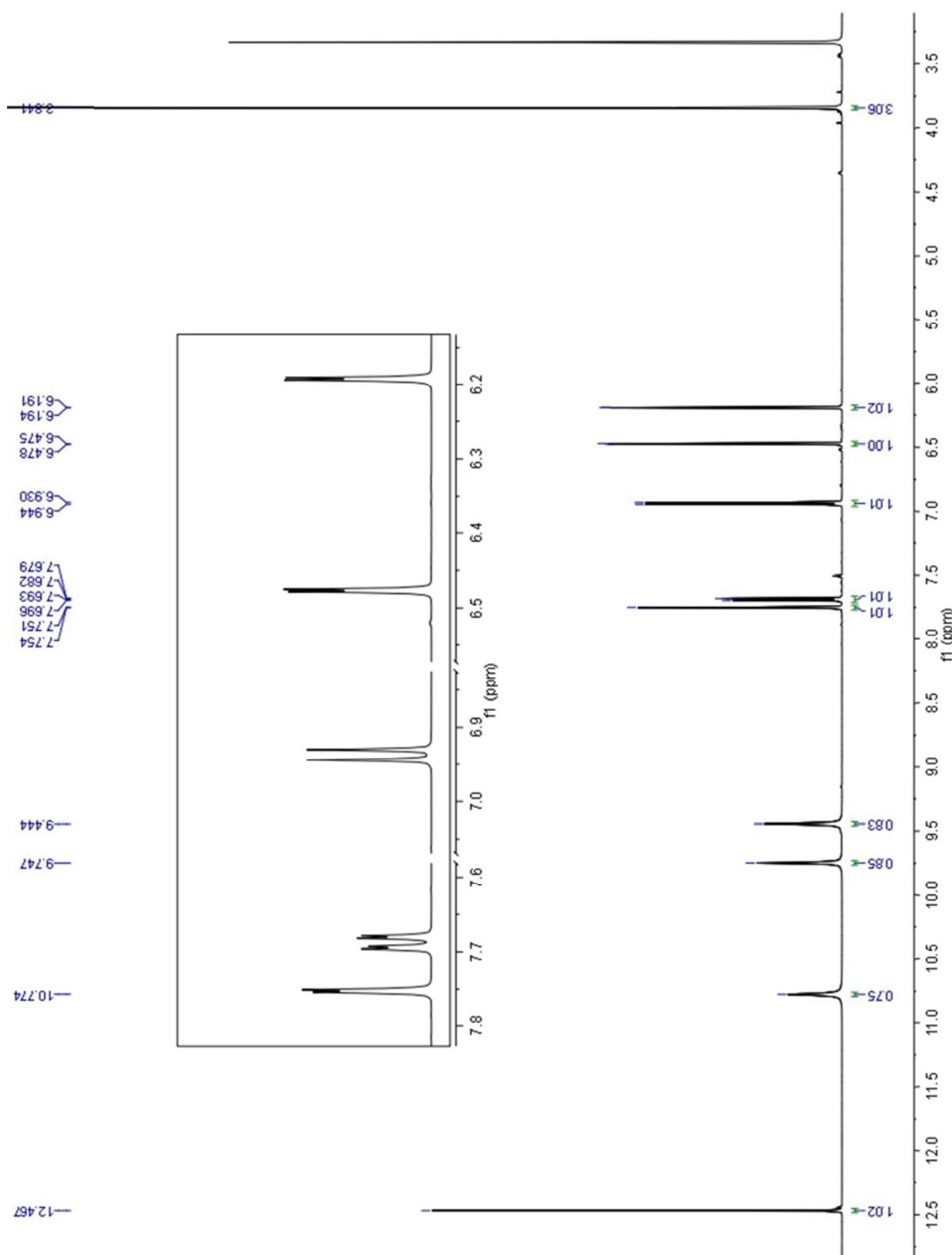
CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
13.09593996 = Left frequency (ppm)
0.00000000 = Right frequency (ppm)
10.000 = Acquisition time (s, for QMTLS)
0.000 = Line-width (for modes D, P & T, 0=use defaults)
0.071649999 = Data-point resolution (Hz)
30.935 = GAUSSIAN (%), 0=use default from INF)
-0.908 = Dispersion contribution (%), 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

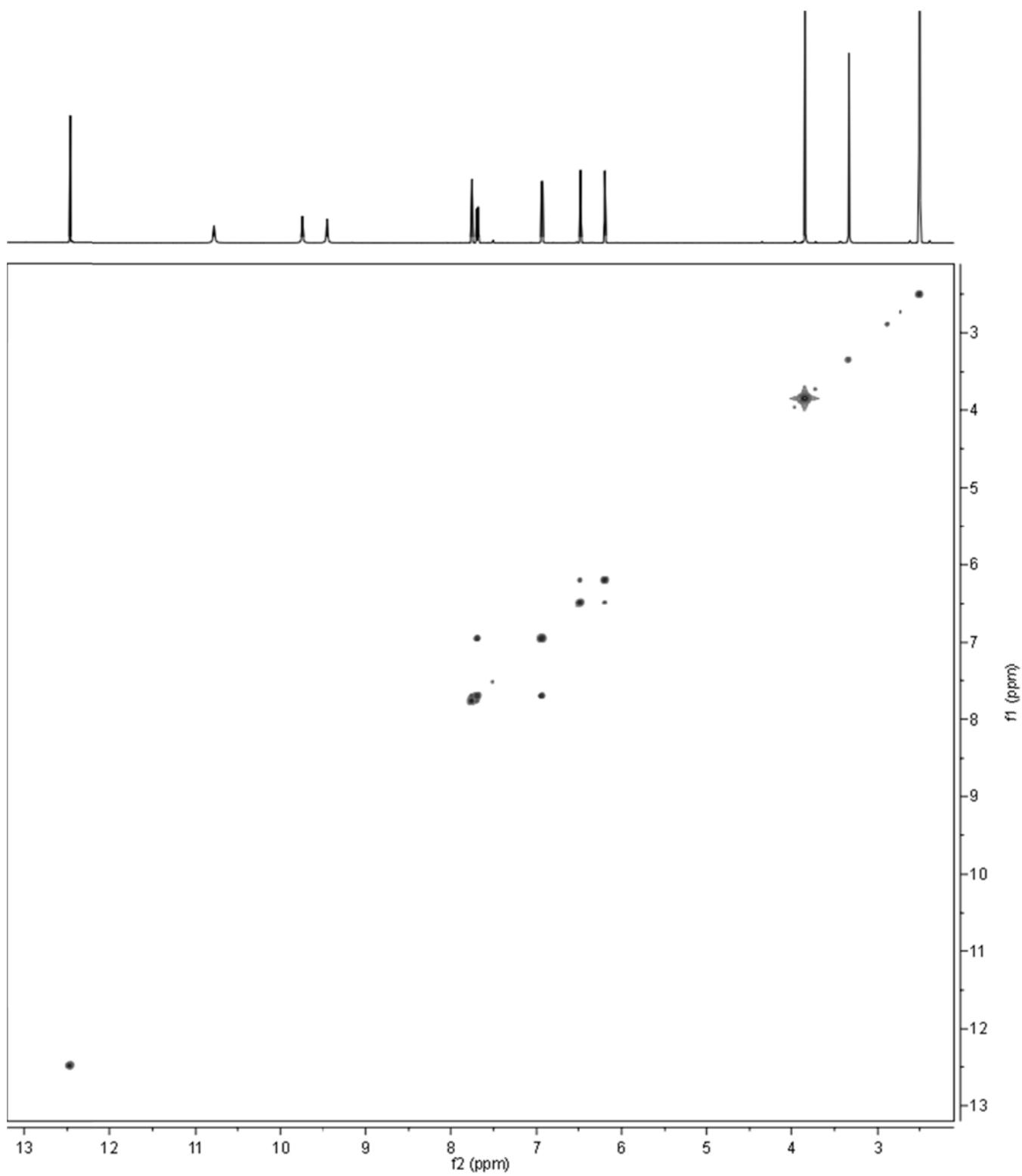
END OF FILE

```

S23.  $^1\text{H}$  NMR spectrum of Isorhamnetin (**8**,  $\text{DMSO}-d_6$ , 600 MHz).



**S24.**  $^1\text{H}$ , $^1\text{H}$ -COSY experiment of Isorhamnetin (**8**, DMSO- $d_6$ , 600 MHz).



## S25. The $^1\text{H}$ Fingerprint of Rutin (**9**) in the PERCH .PMS file format.

```

* NEW: the lines beginning by * are comment lines !
* To keep all the chemical shifts fixed during iteration
* replace "CHEMICAL SHIFTS(HZ):" by "..SHIFTS(HZ): fixed"
* The couplings can be fixed in the same way

NMR-data: c:\users\jgnapo_2\documents\users\jgnapo\perch\ginkgo\flavonoids\
#$@ Date 21.10.2011; Time 0:33:18 perch.5TH

CHEMICAL SHIFTS(PPM): Similarity% = 16.777
PROTON 2*SPIN=1H POPULATION(Y)= 1.00000
H6 / 1 6.190056 1*1*1 STAT=Y PRED= 6.412 RANGE= 0.305 WIDTH(Y)= 1.062 RESP(Y)= 0.9766 HSQC= C2
H8 / 1 6.380863 1*1*1 STAT=Y PRED= 6.581 RANGE= 0.362 WIDTH(Y)= 1.055 RESP(Y)= 0.9738 HSQC= C4
OH7 / 1 10.829194 1*1*1 STAT=Y PRED= 10.802 RANGE= 1.260 WIDTH(Y)= 6.056 RESP(Y)= 0.7582
OH5 / 1 12.599850 1*1*1 STAT=Y PRED= 13.831 RANGE= 0.985 WIDTH(Y)= 1.142 RESP(Y)= 1.0000
H6' / 1 7.537800 1*1*1 STAT=Y PRED= 7.839 RANGE= 0.327 WIDTH(Y)= 1.146 RESP(Y)= 0.9079 HSQC= C14
H5' / 1 6.836057 1*1*1 STAT=Y PRED= 7.028 RANGE= 0.445 WIDTH(Y)= 1.149 RESP(Y)= 0.9452 HSQC= C15
H2' / 1 7.525994 1*1*1 STAT=Y PRED= 7.813 RANGE= 0.392 WIDTH(Y)= 1.146 RESP(Y)= 0.9585 HSQC= C18
OH3' / 1 9.671753 1*1*1 STAT=Y PRED= 9.599 RANGE= 1.830 WIDTH(Y)= 9.146 RESP(Y)= 0.7385
OH4' / 1 9.182772 1*1*1 STAT=Y PRED= 9.443 RANGE= 1.775 WIDTH(Y)= 6.955 RESP(Y)= 0.7480
Glc-1 / 1 5.341488 1*1*1 STAT=Y PRED= 5.565 RANGE= 0.342 WIDTH(Y)= 1.356 RESP(Y)= 0.9078 HSQC= H21
Glc-2 / 1 3.219724 1*1*1 STAT=Y PRED= 3.526 RANGE= 0.201 WIDTH(Y)= 1.897 RESP(Y)= 0.9171 HSQC= C23
Glc-5 / 1 3.238384 1*1*1 STAT=Y PRED= 3.485 RANGE= 0.247 WIDTH(Y)= 2.050 RESP(Y)= 0.8314 HSQC= C25
Glc-3 / 1 3.204949 1*1*1 STAT=Y PRED= 3.317 RANGE= 0.155 WIDTH(Y)= 2.091 RESP(Y)= 0.8548 HSQC= C26
Glc-4 / 1 3.047931 1*1*1 STAT=Y PRED= 3.387 RANGE= 0.287 WIDTH(Y)= 2.008 RESP(Y)= 0.8374 HSQC= C27
OH-Glc-2 / 1 5.286398 1*1*1 STAT=Y PRED= 5.633 RANGE= 1.215 WIDTH(Y)= 1.765 RESP(Y)= 0.8978
Glc-6 / 1 3.281188 1*1*1 STAT=Y PRED= 3.351 RANGE= 0.450 WIDTH(Y)= 2.220 RESP(Y)= 0.8318 HSQC= C29
Glc-6' / 1 3.701113 1*1*1 STAT=Y PRED= 3.509 RANGE= 0.232 WIDTH(Y)= 2.257 RESP(Y)= 0.8536 HSQC= C29
OH-Glc-3 / 1 5.115871 1*1*1 STAT=Y PRED= 4.296 RANGE= 1.420 WIDTH(Y)= 2.048 RESP(Y)= 0.8798
OH-Glc-4 / 1 5.074713 1*1*1 STAT=Y PRED= 4.702 RANGE= 1.568 WIDTH(Y)= 2.000 RESP(Y)= 0.8679
Rh-1 / 1 4.376166 1*1*1 STAT=Y PRED= 4.453 RANGE= 0.300 WIDTH(Y)= 1.672 RESP(Y)= 0.9527 HSQC= C33
Rh-5 / 1 3.265194 1*1*1 STAT=Y PRED= 2.961 RANGE= 0.627 WIDTH(Y)= 1.684 RESP(Y)= 0.9335 HSQC= C36
Rh-Me / 1 0.987386 1*1*3 STAT=Y PRED= 0.586 RANGE= 0.262 WIDTH(Y)= 1.730 RESP(Y)= 0.8538 HSQC= C40
Rh-4 / 1 3.067177 1*1*1 STAT=Y PRED= 2.968 RANGE= 0.287 WIDTH(Y)= 3.477 RESP(Y)= 0.9349 HSQC= C37
Rh-2 / 1 3.383564 1*1*1 STAT=Y PRED= 3.609 RANGE= 0.242 WIDTH(Y)= 2.395 RESP(Y)= 0.9502 HSQC= C38
Rh-3 / 1 3.275197 1*1*1 STAT=Y PRED= 3.082 RANGE= 0.582 WIDTH(Y)= 2.131 RESP(Y)= 0.8779 HSQC= C39
OH-Rh-2 / 1 4.350559 1*1*1 STAT=Y PRED= 4.719 RANGE= 1.395 WIDTH(Y)= 4.049 RESP(Y)= 0.7880
OH-Rh-3 / 1 4.400478 1*1*1 STAT=Y PRED= 4.127 RANGE= 1.613 WIDTH(Y)= 3.183 RESP(Y)= 0.8379
OH-Rh-4 / 1 4.534535 1*1*1 STAT=Y PRED= 4.732 RANGE= 1.673 WIDTH(Y)= 10.065 RESP(Y)= 0.8554

COUPLING CONSTANTS(HZ): Similarity% = 36.900
J22_23 2.0571 J H6 H8 STAT=Y PRED= 2.110 RANGE= 1.790
J27_28 8.4604 J H6' H5' STAT=Y PRED= 8.380 RANGE= 1.000
J27_29 2.2300 J H6' H2' STAT=Y PRED= 2.080 RANGE= 1.600
J28_29 0.2251 J H5' H2' STAT=Y PRED= 0.460 RANGE= 0.640
J38_40 7.7199 J Glc-1 Glc-2 STAT=Y PRED= 7.190 RANGE= 3.580
J40_43 9.0266 J Glc-2 Glc-3 STAT=Y PRED= 8.920 RANGE= 3.580
J40_48 4.7130 J Glc-2 OH-Glc-2 STAT=Y PRED= 4.300 RANGE= 1.000
J41_46 9.8132 J Glc-5 Glc-4 STAT=Y PRED= 10.550 RANGE= 2.560
J41_49 6.9219 J Glc-5 Glc-6 STAT=Y PRED= 2.700 RANGE= 3.000
J41_50 1.7617 J Glc-5 Glc-6' STAT=Y PRED= 1.280 RANGE= 3.000
J43_46 8.7207 J Glc-3 Glc-4 STAT=Y PRED= 9.140 RANGE= 3.580
J43_51 4.8865 J Glc-3 OH-Glc-3 STAT=Y PRED= 4.300 RANGE= 1.000
J46_52 5.8261 J Glc-4 OH-Glc-4 STAT=Y PRED= 4.300 RANGE= 1.000
J49_50 -11.2374 J Glc-6 Glc-6' STAT=Y PRED= -12.300 RANGE= 0.800
J56_68 1.5856 J Rh-1 Rh-2 STAT=Y PRED= 2.330 RANGE= 2.800
J57_59 6.2129 J Rh-5 Rh-Me STAT=Y PRED= 6.250 RANGE= 0.600
J57_66 9.2814 J Rh-5 Rh-4 STAT=Y PRED= 9.940 RANGE= 3.580
J66_69 9.5011 J Rh-4 Rh-3 STAT=Y PRED= 9.590 RANGE= 3.580
J66_73 3.6210 J Rh-4 OH-Rh-4 STAT=Y PRED= 4.300 RANGE= 1.000
J68_69 3.0081 J Rh-2 Rh-3 STAT=Y PRED= 3.420 RANGE= 3.800
J68_71 4.0866 J Rh-2 OH-Rh-2 STAT=Y PRED= 4.440 RANGE= 1.500
J69_72 5.6937 J Rh-3 OH-Rh-3 STAT=Y PRED= 4.300 RANGE= 1.000

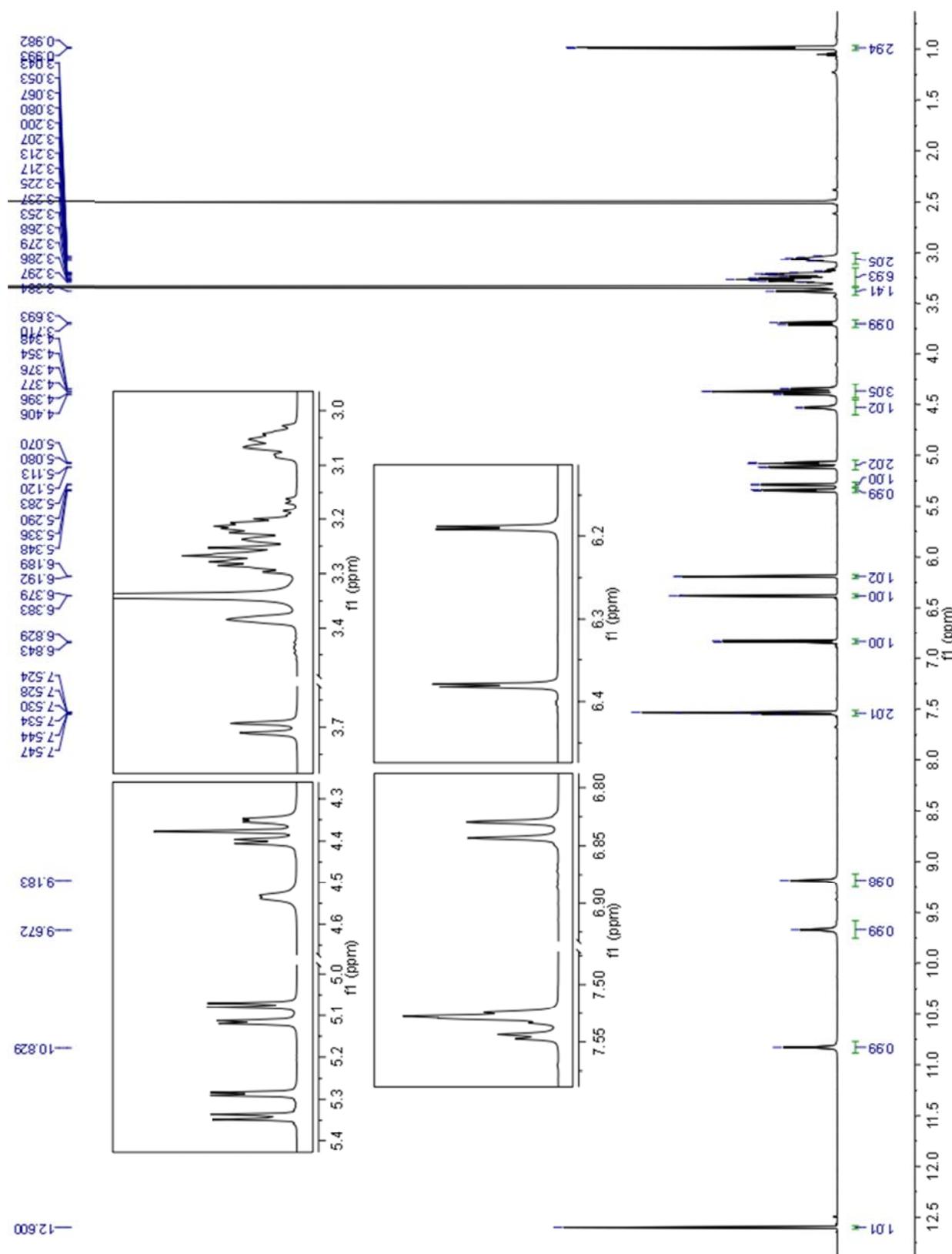
CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
13.25602790 = Left frequency (ppm)
0.00000000 = Right frequency (ppm)
10.000 = Acquisition time (s, for QMTLS)
1.046 = Line-width (for modes D, P & T, 0=use defaults)
0.068609922 = Data-point resolution (Hz)
70.615 = GAUSSIAN (% , 0=use default from INF)
-1.053 = Dispersion contribution (% , 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

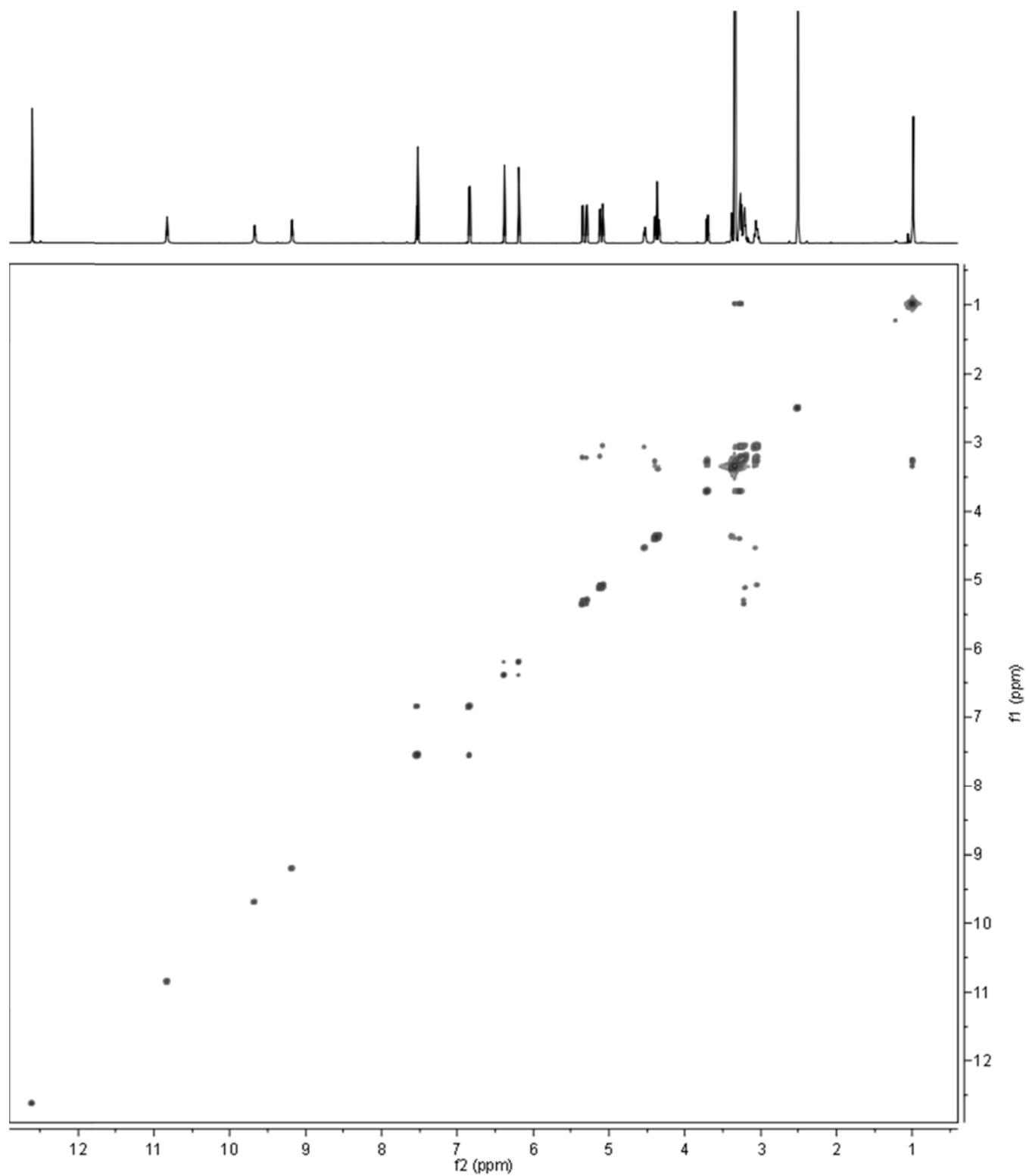
END OF FILE

```

S26.  $^1\text{H}$  NMR spectrum of Rutin (**9**,  $\text{DMSO}-d_6$ , 600 MHz).



**S27.**  $^1\text{H}$ , $^1\text{H}$ -COSY experiment of Rutin (**9**, DMSO- $d_6$ , 600 MHz).



**S28.** Combined  $^1\text{H}$  Fingerprints of nine *Ginkgo biloba* constituents (**1–9**) in the PERCH .PMS file format.

\* NEW: the lines beginning by \* are comment lines !  
 \* To keep all the chemical shifts fixed during iteration  
 \* replace "CHEMICAL SHIFTS(HZ):" by "...SHIFTS(HZ): fixed"  
 \* The couplings can be fixed in the same way

NMR-data: C:\Users\jgnapo\_2\Documents\jgnapo\perch\Ginkgo\  
 #\$\$@ Date 10. 15.2011; Time 15: 5:45 Ginkgo.5TH

CHEMICAL SHIFTS(PPM) :

BILOBALIDE	2*SPIN= 1 SPECIES=1H	POPULATION(Y)= 1.00000
BB-OH-10 / 1	7.253250 1*1*1	STAT=Y PRED= 4.828 RANGE= 1.750 WIDTH(Y)= 1.600 RESP(Y)= 0.9729
BB-H-7beta / 1	2.077084 1*1*1	STAT=Y PRED= 1.896 RANGE= 0.357 WIDTH(Y)= 1.891 RESP(Y)= 0.9170 HSQC= C14
BB-H-6 / 1	4.916794 1*1*1	STAT=Y PRED= 4.888 RANGE= 0.282 WIDTH(Y)= 1.768 RESP(Y)= 0.9697 HSQC= C13
BB-H-7alpha / 1	2.561866 1*1*1	STAT=Y PRED= 2.518 RANGE= 0.282 WIDTH(Y)= 1.976 RESP(Y)= 0.8993 HSQC= C14
BB-H-10 / 1	5.149952 1*1*1	STAT=Y PRED= 4.165 RANGE= 0.370 WIDTH(Y)= 1.774 RESP(Y)= 0.9421 HSQC= C17
BB-H-12 / 1	6.274102 1*1*1	STAT=Y PRED= 5.659 RANGE= 0.102 WIDTH(Y)= 1.500 RESP(Y)= 1.0000 HSQC= C19
BB-t-Butyl / 1	1.034063 1*1*9	STAT=Y PRED= 0.914 RANGE= 0.237 WIDTH(Y)= 2.678 RESP(Y)= 0.9023 HSQC= C25-27
BB-OH-8 / 1	5.403089 1*1*1	STAT=Y PRED= 3.979 RANGE= 1.923 WIDTH(Y)= 1.508 RESP(Y)= 0.9843
BB-H-1beta / 1	2.774028 1*1*1	STAT=Y PRED= 2.359 RANGE= 0.370 WIDTH(Y)= 1.686 RESP(Y)= 0.9603 HSQC= #27
BB-H-lalpha / 1	2.897044 1*1*1	STAT=Y PRED= 2.405 RANGE= 0.462 WIDTH(Y)= 1.740 RESP(Y)= 0.9580 HSQC= #27
GINKGOLIDE_A	2*SPIN= 1 SPECIES=1H	POPULATION(Y)= 1.00000
GA-OH-3 / 2	6.378972 1*1*1	STAT=Y PRED= 4.856 RANGE= 1.805 WIDTH(Y)= 1.273 RESP(Y)= 1.0000
GA-OH-10 / 2	6.814323 1*1*1	STAT=Y PRED= 4.509 RANGE= 1.630 WIDTH(Y)= 1.565 RESP(Y)= 0.9632
GA-H-7beta / 2	2.043863 1*1*1	STAT=Y PRED= 2.070 RANGE= 0.557 WIDTH(Y)= 2.501 RESP(Y)= 0.8989 HSQC= C14
GA-H-1beta / 2	1.805711 1*1*1	STAT=Y PRED= 1.401 RANGE= 0.302 WIDTH(Y)= 1.882 RESP(Y)= 0.9213 HSQC= C8
GA-H-lalpha / 2	2.750139 1*1*1	STAT=Y PRED= 2.169 RANGE= 0.387 WIDTH(Y)= 1.927 RESP(Y)= 0.9005 HSQC= C8
GA-H-2 / 2	4.833738 1*1*1	STAT=Y PRED= 4.895 RANGE= 0.262 WIDTH(Y)= 1.798 RESP(Y)= 0.9709 HSQC= C9
GA-H-6 / 2	4.945707 1*1*1	STAT=Y PRED= 4.760 RANGE= 0.247 WIDTH(Y)= 1.956 RESP(Y)= 0.9824 HSQC= C13
GA-H-7alpha / 2	2.027505 1*1*1	STAT=Y PRED= 2.025 RANGE= 0.472 WIDTH(Y)= 2.358 RESP(Y)= 0.8736 HSQC= C14
GA-H-8 / 2	1.710433 1*1*1	STAT=Y PRED= 1.700 RANGE= 0.222 WIDTH(Y)= 2.173 RESP(Y)= 0.9339 HSQC= C15
GA-H-10 / 2	4.941390 1*1*1	STAT=Y PRED= 4.165 RANGE= 0.392 WIDTH(Y)= 1.780 RESP(Y)= 0.9248 HSQC= C17
GA-H-12 / 2	6.023393 1*1*1	STAT=Y PRED= 5.659 RANGE= 0.335 WIDTH(Y)= 1.498 RESP(Y)= 0.9865 HSQC= C19
GA-H-14 / 2	2.937891 1*1*1	STAT=Y PRED= 2.949 RANGE= 0.387 WIDTH(Y)= 1.519 RESP(Y)= 0.9493 HSQC= C21
GA-H-16 / 2	1.117747 1*1*3	STAT=Y PRED= 1.114 RANGE= 0.252 WIDTH(Y)= 1.687 RESP(Y)= 0.9350 HSQC= C23
GA-t-but / 2	0.017131 1*1*9	STAT=Y PRED= 0.928 RANGE= 0.227 WIDTH(Y)= 2.491 RESP(Y)= 0.9025 HSQC= C25-27
GINKGOLIDE_B	2*SPIN= 1 SPECIES=1H	POPULATION(Y)= 1.00000
GB-OH-3 / 3	6.471908 1*1*1	STAT=Y PRED= 4.913 RANGE= 1.733 WIDTH(Y)= 1.525 RESP(Y)= 1.0000
GB-OH-10 / 3	7.462874 1*1*1	STAT=Y PRED= 4.556 RANGE= 1.695 WIDTH(Y)= 1.929 RESP(Y)= 0.9612
GB-H-7beta / 3	2.137230 1*1*1	STAT=Y PRED= 2.036 RANGE= 0.560 WIDTH(Y)= 2.168 RESP(Y)= 0.9003 HSQC= C14
GB-H-1 / 3	4.042104 1*1*1	STAT=Y PRED= 3.762 RANGE= 0.100 WIDTH(Y)= 1.692 RESP(Y)= 0.9364 HSQC= C8
GB-H-2 / 3	4.642590 1*1*1	STAT=Y PRED= 4.089 RANGE= 0.187 WIDTH(Y)= 1.644 RESP(Y)= 0.9775 HSQC= C9
GB-H-6 / 3	5.303773 1*1*1	STAT=Y PRED= 4.895 RANGE= 0.237 WIDTH(Y)= 1.799 RESP(Y)= 0.9597 HSQC= C13
GB-H-7alpha / 3	1.931786 1*1*1	STAT=Y PRED= 2.041 RANGE= 0.477 WIDTH(Y)= 2.242 RESP(Y)= 0.8904 HSQC= C14
GB-H-8 / 3	1.724094 1*1*1	STAT=Y PRED= 1.670 RANGE= 0.215 WIDTH(Y)= 2.358 RESP(Y)= 0.9326 HSQC= C15
GB-H-10 / 3	5.019748 1*1*1	STAT=Y PRED= 4.165 RANGE= 0.380 WIDTH(Y)= 1.969 RESP(Y)= 0.9334 HSQC= C17
GB-H-12 / 3	6.074154 1*1*1	STAT=Y PRED= 5.659 RANGE= 0.330 WIDTH(Y)= 1.664 RESP(Y)= 0.9898 HSQC= C19
GB-H-14 / 3	2.843287 1*1*1	STAT=Y PRED= 2.949 RANGE= 0.367 WIDTH(Y)= 1.666 RESP(Y)= 0.9457 HSQC= C21
GB-H-16 / 3	1.106901 1*1*3	STAT=Y PRED= 1.102 RANGE= 0.382 WIDTH(Y)= 1.848 RESP(Y)= 0.9294 HSQC= C23
GB-t-butyl / 3	1.028434 1*1*9	STAT=Y PRED= 0.871 RANGE= 0.202 WIDTH(Y)= 2.590 RESP(Y)= 0.8997 HSQC= C25-27
GB-OH-1 / 3	4.918842 1*1*1	STAT=Y PRED= 6.248 RANGE= 2.488 WIDTH(Y)= 1.544 RESP(Y)= 0.9556
GINKGOLIDE_C	2*SPIN= 1 SPECIES=1H	POPULATION(Y)= 1.00000
GC-OH-3 / 4	6.479066 1*1*1	STAT=Y PRED= 5.334 RANGE= 1.700 WIDTH(Y)= 1.736 RESP(Y)= 1.0000
GC-OH-10 / 4	7.538812 1*1*1	STAT=Y PRED= 4.347 RANGE= 1.768 WIDTH(Y)= 2.242 RESP(Y)= 0.9489
GC-OH-7 / 4	5.658435 1*1*1	STAT=Y PRED= 4.985 RANGE= 1.620 WIDTH(Y)= 1.944 RESP(Y)= 0.9704
GC-H-1beta / 4	3.986275 1*1*1	STAT=Y PRED= 4.316 RANGE= 0.100 WIDTH(Y)= 1.907 RESP(Y)= 0.9449 HSQC= C8
GC-H-2 / 4	4.626842 1*1*1	STAT=Y PRED= 4.089 RANGE= 0.170 WIDTH(Y)= 1.789 RESP(Y)= 0.9761 HSQC= C9
GC-H-6 / 4	4.966384 1*1*1	STAT=Y PRED= 4.852 RANGE= 0.122 WIDTH(Y)= 1.940 RESP(Y)= 0.9965 HSQC= C13
GC-H-7 / 4	4.053494 1*1*1	STAT=Y PRED= 4.559 RANGE= 0.317 WIDTH(Y)= 1.975 RESP(Y)= 0.9273 HSQC= C14
GC-H-8 / 4	1.551161 1*1*1	STAT=Y PRED= 1.808 RANGE= 0.222 WIDTH(Y)= 2.077 RESP(Y)= 0.9481 HSQC= C15
GC-H-10 / 4	4.998365 1*1*1	STAT=Y PRED= 4.165 RANGE= 0.367 WIDTH(Y)= 2.302 RESP(Y)= 0.9280 HSQC= C17
GC-H-12 / 4	6.099451 1*1*1	STAT=Y PRED= 5.659 RANGE= 0.325 WIDTH(Y)= 1.837 RESP(Y)= 0.9848 HSQC= C19
GC-H-14 / 4	2.813690 1*1*1	STAT=Y PRED= 2.949 RANGE= 0.342 WIDTH(Y)= 1.804 RESP(Y)= 0.9418 HSQC= C21
GC-H-16 / 4	1.107483 1*1*3	STAT=Y PRED= 1.103 RANGE= 0.377 WIDTH(Y)= 1.979 RESP(Y)= 0.8915 HSQC= C23
GC-t-butyl / 4	1.091336 1*1*9	STAT=Y PRED= 0.887 RANGE= 0.225 WIDTH(Y)= 2.733 RESP(Y)= 0.8975 HSQC= C25-27
GC-OH-1 / 4	4.971784 1*1*1	STAT=Y PRED= 5.998 RANGE= 1.965 WIDTH(Y)= 1.840 RESP(Y)= 0.8956
GINKGOLIDE_J	2*SPIN= 1 SPECIES=1H	POPULATION(Y)= 1.00000
GJ-OH-3 / 5	6.393136 1*1*1	STAT=Y PRED= 4.925 RANGE= 1.775 WIDTH(Y)= 1.498 RESP(Y)= 1.0000
GJ-OH-10 / 5	6.908209 1*1*1	STAT=Y PRED= 4.324 RANGE= 1.710 WIDTH(Y)= 1.844 RESP(Y)= 0.9617
GJ-OH-7 / 5	5.545649 1*1*1	STAT=Y PRED= 4.759 RANGE= 1.580 WIDTH(Y)= 1.670 RESP(Y)= 0.9709
GJ-H-1beta / 5	1.752901 1*1*1	STAT=Y PRED= 1.338 RANGE= 0.287 WIDTH(Y)= 2.106 RESP(Y)= 0.9100 HSQC= C8
GJ-H-lalpha / 5	2.717097 1*1*1	STAT=Y PRED= 2.169 RANGE= 0.377 WIDTH(Y)= 2.200 RESP(Y)= 0.8989 HSQC= C8
GJ-H-2 / 5	4.832003 1*1*1	STAT=Y PRED= 4.895 RANGE= 0.232 WIDTH(Y)= 1.942 RESP(Y)= 0.9577 HSQC= C9
GJ-H-6 / 5	4.645476 1*1*1	STAT=Y PRED= 4.895 RANGE= 0.110 WIDTH(Y)= 1.872 RESP(Y)= 0.9688 HSQC= C13
GJ-H-7 / 5	4.171447 1*1*1	STAT=Y PRED= 4.559 RANGE= 0.325 WIDTH(Y)= 1.862 RESP(Y)= 0.9203 HSQC= C14
GJ-H-8 / 5	1.537556 1*1*1	STAT=Y PRED= 1.768 RANGE= 0.212 WIDTH(Y)= 1.979 RESP(Y)= 0.9531 HSQC= C15
GJ-H-10 / 5	4.928198 1*1*1	STAT=Y PRED= 4.165 RANGE= 0.390 WIDTH(Y)= 2.064 RESP(Y)= 0.9291 HSQC= C17
GJ-H-12 / 5	6.046780 1*1*1	STAT=Y PRED= 5.659 RANGE= 0.332 WIDTH(Y)= 1.684 RESP(Y)= 0.9761 HSQC= C19
GJ-H-14 / 5	2.906534 1*1*1	STAT=Y PRED= 2.949 RANGE= 0.362 WIDTH(Y)= 1.711 RESP(Y)= 0.9406 HSQC= C21
GJ-H-16 / 5	1.117886 1*1*3	STAT=Y PRED= 1.135 RANGE= 0.260 WIDTH(Y)= 1.906 RESP(Y)= 0.9293 HSQC= C23
GJ-t-butyl / 5	1.081520 1*1*9	STAT=Y PRED= 0.894 RANGE= 0.230 WIDTH(Y)= 2.587 RESP(Y)= 0.9000 HSQC= C25-27

QUERCETIN    2\*SPIN= 1 SPECIES=1H    POPULATION(Y)= 1.00000  
 Q-H6 / 6 6.183050 1\*1\*1 STAT=Y PRED= 6.298 RANGE= 0.242 WIDTH(Y)= 1.416 RESP(Y)= 1.0000 HSQC= C2  
 Q-H8 / 6 6.403402 1\*1\*1 STAT=Y PRED= 6.461 RANGE= 0.172 WIDTH(Y)= 1.407 RESP(Y)= 0.9861 HSQC= C4  
 Q-OH7 / 6 10.783504 1\*1\*1 STAT=Y PRED= 10.896 RANGE= 1.000 WIDTH(Y)= 1.739 RESP(Y)= 0.9816  
 Q-OH5 / 6 12.494440 1\*1\*1 STAT=Y PRED= 13.379 RANGE= 0.850 WIDTH(Y)= 1.942 RESP(Y)= 0.9532  
 Q-H6' / 6 7.536549 1\*1\*1 STAT=Y PRED= 7.627 RANGE= 0.265 WIDTH(Y)= 1.496 RESP(Y)= 0.9718 HSQC= C14  
 Q-H5' / 6 6.880921 1\*1\*1 STAT=Y PRED= 6.929 RANGE= 0.197 WIDTH(Y)= 1.406 RESP(Y)= 0.9618 HSQC= C15  
 Q-H2' / 6 7.674662 1\*1\*1 STAT=Y PRED= 7.698 RANGE= 0.247 WIDTH(Y)= 1.416 RESP(Y)= 0.9741 HSQC= C18  
 Q-OH3' / 6 9.374357 1\*1\*1 STAT=Y PRED= 9.494 RANGE= 1.215 WIDTH(Y)= 1.780 RESP(Y)= 0.9566  
 Q-OH4' / 6 9.311783 1\*1\*1 STAT=Y PRED= 9.682 RANGE= 1.315 WIDTH(Y)= 2.880 RESP(Y)= 0.9305  
 Q-OH3 / 6 9.597692 1\*1\*1 STAT=Y PRED= 9.115 RANGE= 1.115 WIDTH(Y)= 2.732 RESP(Y)= 0.9341  
 KAEMPFEROL 2\*SPIN= 1 SPECIES=1H    POPULATION(Y)= 1.00000  
 H3\_5' / 7 6.922821 1\*2\*1 STAT=Y PRED= 6.925 RANGE= 0.177 WIDTH(Y)= 1.511 RESP(Y)= 0.9770 HSQC= C15\_17  
 K-H6 / 7 6.186969 1\*1\*1 STAT=Y PRED= 6.295 RANGE= 0.252 WIDTH(Y)= 1.406 RESP(Y)= 1.0000 HSQC= C2  
 K-H8 / 7 6.435905 1\*1\*1 STAT=Y PRED= 6.468 RANGE= 0.172 WIDTH(Y)= 1.404 RESP(Y)= 0.9947 HSQC= C4  
 K-OH7 / 7 10.788285 1\*1\*1 STAT=Y PRED= 10.903 RANGE= 0.985 WIDTH(Y)= 17.150 RESP(Y)= 0.8071  
 K-OH5 / 7 12.484003 1\*1\*1 STAT=Y PRED= 13.446 RANGE= 0.850 WIDTH(Y)= 1.739 RESP(Y)= 0.9625  
 H2\_6' / 7 8.041525 1\*2\*1 STAT=Y PRED= 8.080 RANGE= 0.252 WIDTH(Y)= 1.495 RESP(Y)= 0.9722 HSQC= C14\_18  
 K-OH4' / 7 9.408526 1\*1\*1 STAT=Y PRED= 9.807 RANGE= 1.500 WIDTH(Y)= 11.480 RESP(Y)= 0.8274  
 K-OH3 / 7 10.109684 1\*1\*1 STAT=Y PRED= 9.155 RANGE= 1.070 WIDTH(Y)= 11.760 RESP(Y)= 0.8250  
 ISORHAMNETIN 2\*SPIN= 1 SPECIES=1H    POPULATION(Y)= 1.00000  
 I-H6 / 8 6.192379 1\*1\*1 STAT=Y PRED= 6.306 RANGE= 0.242 WIDTH(Y)= 1.456 RESP(Y)= 0.9979 HSQC= C2  
 I-H8 / 8 6.476736 1\*1\*1 STAT=Y PRED= 6.462 RANGE= 0.182 WIDTH(Y)= 1.446 RESP(Y)= 0.9882 HSQC= C4  
 I-OH7 / 8 10.773796 1\*1\*1 STAT=Y PRED= 10.912 RANGE= 1.015 WIDTH(Y)= 2.985 RESP(Y)= 0.9504  
 I-OH5 / 8 12.467391 1\*1\*1 STAT=Y PRED= 13.376 RANGE= 0.855 WIDTH(Y)= 1.433 RESP(Y)= 1.0000  
 I-H6' / 8 7.687450 1\*1\*1 STAT=Y PRED= 7.638 RANGE= 0.297 WIDTH(Y)= 1.538 RESP(Y)= 0.9733 HSQC= C14  
 I-H5' / 8 6.936966 1\*1\*1 STAT=Y PRED= 6.904 RANGE= 0.242 WIDTH(Y)= 1.475 RESP(Y)= 0.9759 HSQC= C15  
 I-H2' / 8 7.752072 1\*1\*1 STAT=Y PRED= 7.676 RANGE= 0.305 WIDTH(Y)= 1.697 RESP(Y)= 0.9921 HSQC= C18  
 I-OH4' / 8 9.445056 1\*1\*1 STAT=Y PRED= 9.712 RANGE= 0.752 WIDTH(Y)= 2.159 RESP(Y)= 0.9537  
 I-OH3 / 8 9.747292 1\*1\*1 STAT=Y PRED= 9.072 RANGE= 1.125 WIDTH(Y)= 2.173 RESP(Y)= 0.9713  
 I-OMe / 8 3.840464 1\*1\*3 STAT=Y PRED= 3.899 RANGE= 0.132 WIDTH(Y)= 1.727 RESP(Y)= 0.9945 HSQC= H20  
 RUTIN 2\*SPIN= 1 SPECIES=1H    POPULATION(Y)= 1.00000  
 R-H6 / 9 6.190056 1\*1\*1 STAT=Y PRED= 6.412 RANGE= 0.305 WIDTH(Y)= 1.062 RESP(Y)= 0.9766 HSQC= C2  
 R-H8 / 9 6.380863 1\*1\*1 STAT=Y PRED= 6.581 RANGE= 0.362 WIDTH(Y)= 1.055 RESP(Y)= 0.9738 HSQC= C4  
 R-OH7 / 9 10.829194 1\*1\*1 STAT=Y PRED= 10.802 RANGE= 1.260 WIDTH(Y)= 6.056 RESP(Y)= 0.7582  
 R-OH5 / 9 12.599850 1\*1\*1 STAT=Y PRED= 13.831 RANGE= 0.985 WIDTH(Y)= 1.142 RESP(Y)= 1.0000  
 R-H6' / 9 7.537800 1\*1\*1 STAT=Y PRED= 7.839 RANGE= 0.327 WIDTH(Y)= 1.146 RESP(Y)= 0.9079 HSQC= C14  
 R-H5' / 9 6.836057 1\*1\*1 STAT=Y PRED= 7.028 RANGE= 0.445 WIDTH(Y)= 1.149 RESP(Y)= 0.9452 HSQC= C15  
 R-H2' / 9 7.525994 1\*1\*1 STAT=Y PRED= 7.813 RANGE= 0.392 WIDTH(Y)= 1.146 RESP(Y)= 0.9585 HSQC= C18  
 R-OH3' / 9 6.671753 1\*1\*1 STAT=Y PRED= 9.599 RANGE= 1.830 WIDTH(Y)= 9.146 RESP(Y)= 0.7385  
 R-OH4' / 9 9.182772 1\*1\*1 STAT=Y PRED= 9.443 RANGE= 1.775 WIDTH(Y)= 6.955 RESP(Y)= 0.7480  
 R-Glc-1 / 9 5.341488 1\*1\*1 STAT=Y PRED= 5.565 RANGE= 0.342 WIDTH(Y)= 1.356 RESP(Y)= 0.9078 HSQC= H21  
 R-Glc-2 / 9 3.219724 1\*1\*1 STAT=Y PRED= 3.526 RANGE= 0.200 WIDTH(Y)= 1.897 RESP(Y)= 0.9171 HSQC= C23  
 R-Glc-5 / 9 3.238384 1\*1\*1 STAT=Y PRED= 3.485 RANGE= 0.247 WIDTH(Y)= 2.050 RESP(Y)= 0.8314 HSQC= C25  
 R-Glc-3 / 9 3.204949 1\*1\*1 STAT=Y PRED= 3.317 RANGE= 0.155 WIDTH(Y)= 2.091 RESP(Y)= 0.8548 HSQC= C26  
 R-Glc-4 / 9 3.047931 1\*1\*1 STAT=Y PRED= 3.387 RANGE= 0.287 WIDTH(Y)= 2.008 RESP(Y)= 0.8374 HSQC= C27  
 R-OH-Glc-2 / 9 5.286398 1\*1\*1 STAT=Y PRED= 5.633 RANGE= 1.215 WIDTH(Y)= 1.765 RESP(Y)= 0.8978  
 R-Glc-6 / 9 3.281188 1\*1\*1 STAT=Y PRED= 3.351 RANGE= 0.450 WIDTH(Y)= 2.220 RESP(Y)= 0.8318 HSQC= C29  
 R-Glc-6' / 9 3.701113 1\*1\*1 STAT=Y PRED= 3.509 RANGE= 0.232 WIDTH(Y)= 2.257 RESP(Y)= 0.8536 HSQC= C29  
 R-OH-Glc-3 / 9 5.115871 1\*1\*1 STAT=Y PRED= 4.296 RANGE= 1.420 WIDTH(Y)= 2.048 RESP(Y)= 0.8798  
 R-OH-Glc-4 / 9 5.074713 1\*1\*1 STAT=Y PRED= 4.702 RANGE= 1.568 WIDTH(Y)= 2.000 RESP(Y)= 0.8679  
 R-Rh-1 / 9 4.376166 1\*1\*1 STAT=Y PRED= 4.453 RANGE= 0.300 WIDTH(Y)= 1.672 RESP(Y)= 0.9527 HSQC= C33  
 R-Rh-5 / 9 3.265194 1\*1\*1 STAT=Y PRED= 2.961 RANGE= 0.627 WIDTH(Y)= 1.684 RESP(Y)= 0.9335 HSQC= C36  
 R-Rh-Me / 9 0.987386 1\*1\*3 STAT=Y PRED= 0.586 RANGE= 0.262 WIDTH(Y)= 1.730 RESP(Y)= 0.8538 HSQC= C40  
 R-Rh-4 / 9 3.067177 1\*1\*1 STAT=Y PRED= 2.968 RANGE= 0.287 WIDTH(Y)= 3.477 RESP(Y)= 0.9349 HSQC= C37  
 R-Rh-2 / 9 3.383564 1\*1\*1 STAT=Y PRED= 3.609 RANGE= 0.242 WIDTH(Y)= 2.395 RESP(Y)= 0.9502 HSQC= C38  
 R-Rh-3 / 9 3.275197 1\*1\*1 STAT=Y PRED= 3.082 RANGE= 0.582 WIDTH(Y)= 2.131 RESP(Y)= 0.8779 HSQC= C39  
 R-OH-Rh-2 / 9 4.350559 1\*1\*1 STAT=Y PRED= 4.719 RANGE= 1.395 WIDTH(Y)= 4.049 RESP(Y)= 0.7880  
 R-OH-Rh-3 / 9 4.400478 1\*1\*1 STAT=Y PRED= 4.127 RANGE= 1.613 WIDTH(Y)= 3.183 RESP(Y)= 0.8379  
 R-OH-Rh-4 / 9 4.534535 1\*1\*1 STAT=Y PRED= 4.732 RANGE= 1.673 WIDTH(Y)= 10.065 RESP(Y)= 0.8554

#### COUPLING CONSTANTS (HZ) :

J1_2	5.1195	J BB-OH-10	BB-H-10	STAT=Y	PRED= 4.890	RANGE= 0.750
J3_4	6.8503	J BB-H-7beta	BB-H-6	STAT=Y	PRED= 9.550	RANGE= 3.000
J3_4	-13.3259	J BB-H-7beta	BB-H-7alpha	STAT=Y	PRED= -12.140	RANGE= 1.000
J4_5	7.0969	J BB-H-6	BB-H-7alpha	STAT=Y	PRED= 6.600	RANGE= 3.000
J6_7	-18.1406	J BB-H-1beta	BB-H-1alpha	STAT=Y	PRED= -15.860	RANGE= 2.000
J8_9	5.1630	J GA-OH-10	GA-H-10	STAT=Y	PRED= 5.140	RANGE= 0.750
J10_11	0.6218	J GA-H-7beta	GA-H-6	STAT=Y	PRED= 1.070	RANGE= 2.000
J10_12	-13.7046	J GA-H-7beta	GA-H-7alpha	STAT=Y	PRED= -12.430	RANGE= 1.000
J10_13	4.5778	J GA-H-7beta	GA-H-8	STAT=Y	PRED= 4.190	RANGE= 3.000
J14_15	-15.1675	J GA-H-1beta	GA-H-1alpha	STAT=Y	PRED= -12.310	RANGE= 1.000
J14_16	8.2624	J GA-H-1beta	GA-H-2	STAT=Y	PRED= 8.810	RANGE= 3.000
J15_16	7.2878	J GA-H-1alpha	GA-H-2	STAT=Y	PRED= 7.110	RANGE= 3.000
J11_12	4.0483	J GA-H-6	GA-H-7alpha	STAT=Y	PRED= 4.830	RANGE= 3.000
J12_13	14.3434	J GA-H-7alpha	GA-H-8	STAT=Y	PRED= 13.120	RANGE= 2.200
J17_18	7.1931	J GA-H-14	GA-H-16	STAT=Y	PRED= 6.640	RANGE= 0.250
J19_20	5.5557	J GB-OH-10	GB-H-10	STAT=Y	PRED= 5.110	RANGE= 0.750
J21_22	0.8476	J GB-H-7beta	GB-H-6	STAT=Y	PRED= 1.140	RANGE= 2.200
J21_23	-13.7414	J GB-H-7beta	GB-H-7alpha	STAT=Y	PRED= -12.420	RANGE= 1.000
J21_24	4.6463	J GB-H-7beta	GB-H-8	STAT=Y	PRED= 4.190	RANGE= 3.000
J25_26	7.4027	J GB-H-1	GB-H-2	STAT=Y	PRED= 7.800	RANGE= 3.000
J25_27	3.5631	J GB-H-1	GB-OH-1	STAT=Y	PRED= 3.950	RANGE= 0.750
J22_23	4.1428	J GB-H-6	GB-H-7alpha	STAT=Y	PRED= 4.630	RANGE= 3.000
J23_24	14.3540	J GB-H-7alpha	GB-H-8	STAT=Y	PRED= 13.130	RANGE= 2.200
J28_29	7.0831	J GB-H-14	GB-H-16	STAT=Y	PRED= 6.640	RANGE= 0.250

J30_31	5.6531	J GC-OH-10	GC-H-10	STAT=Y	PRED= 4.950	RANGE= 0.750
J32_33	6.2720	J GC-OH-7	GC-H-7	STAT=Y	PRED= 4.300	RANGE= 0.500
J34_35	7.2205	J GC-H-1beta	GC-H-2	STAT=Y	PRED= 7.910	RANGE= 3.000
J34_36	3.6847	J GC-H-1beta	GC-OH-1	STAT=Y	PRED= 4.300	RANGE= 0.500
J37_33	4.1999	J GC-H-6	GC-H-7	STAT=Y	PRED= 4.670	RANGE= 3.000
J33_38	12.3964	J GC-H-7	GC-H-8	STAT=Y	PRED= 11.070	RANGE= 2.200
J39_40	7.0914	J GC-H-14	GC-H-16	STAT=Y	PRED= 6.640	RANGE= 0.250
J41_42	5.3487	J GJ-OH-10	GJ-H-10	STAT=Y	PRED= 5.040	RANGE= 0.750
J43_44	6.8614	J GU-OH-7	GJ-H-7	STAT=Y	PRED= 4.300	RANGE= 0.500
J45_46	-15.1380	J GJ-H-1beta	GJ-H-1alpha	STAT=Y	PRED= -12.310	RANGE= 1.000
J45_48	8.1701	J GU-H-1beta	GJ-H-2	STAT=Y	PRED= 9.280	RANGE= 3.000
J46_48	7.2493	J GU-H-1alpha	GJ-H-2	STAT=Y	PRED= 7.040	RANGE= 3.000
J47_44	4.1855	J GJ-H-6	GJ-H-7	STAT=Y	PRED= 4.830	RANGE= 3.000
J44_49	12.2977	J GJ-H-7	GJ-H-8	STAT=Y	PRED= 11.060	RANGE= 2.200
J50_51	7.2044	J GJ-H-14	GJ-H-16	STAT=Y	PRED= 6.640	RANGE= 0.250
J52_53	2.0315	J Q-H6	Q-H8	STAT=Y	PRED= 2.110	RANGE= 0.890
J54_55	8.4728	J Q-H6'	Q-H5'	STAT=Y	PRED= 8.380	RANGE= 0.500
J54_56	2.2019	J Q-H6'	Q-H2'	STAT=Y	PRED= 2.080	RANGE= 0.800
J55_56	0.4659	J Q-H5'	Q-H2'	STAT=Y	PRED= 0.460	RANGE= 0.320
J57_58	2.4778	J H3'-5'	H3'-5'	STAT=Y	PRED= 2.710	RANGE= 0.700
J57_59	0.2690	J H2'-6'	H3'-5'	STAT=Y	PRED= 0.460	RANGE= 0.320
J57_60	8.7801	J H3'-5'	H2'-6'	STAT=Y	PRED= 8.530	RANGE= 0.440
J61_62	2.0325	J K-H6	K-H8	STAT=Y	PRED= 2.110	RANGE= 0.890
J60_59	2.4747	J H2'-6'	H2'-6'	STAT=Y	PRED= 2.380	RANGE= 0.700
J64_63	2.0139	J I-H6	I-H8	STAT=Y	PRED= 2.110	RANGE= 0.890
J65_66	8.4712	J I-H6'	I-H5'	STAT=Y	PRED= 8.340	RANGE= 0.500
J65_67	2.0570	J I-H6'	I-H2'	STAT=Y	PRED= 2.080	RANGE= 0.800
J66_67	0.2967	J I-H5'	I-H2'	STAT=Y	PRED= 0.460	RANGE= 0.320
J68_69	2.0571	J R-H6	R-H8	STAT=Y	PRED= 2.110	RANGE= 1.790
J70_71	8.4604	J R-H6'	R-H5'	STAT=Y	PRED= 8.380	RANGE= 1.000
J70_72	2.2300	J R-H6'	R-H2'	STAT=Y	PRED= 2.080	RANGE= 1.600
J71_72	0.2251	J R-H5'	R-H2'	STAT=Y	PRED= 0.460	RANGE= 0.640
J73_74	7.7199	J R-Glc-1	R-Glc-2	STAT=Y	PRED= 7.190	RANGE= 3.580
J74_75	9.0266	J R-Glc-2	R-Glc-3	STAT=Y	PRED= 8.920	RANGE= 3.580
J74_80	4.7130	J R-Glc-2	R-OH-Glc-2	STAT=Y	PRED= 4.300	RANGE= 1.000
J77_76	9.8132	J R-Glc-5	R-Glc-4	STAT=Y	PRED= 10.550	RANGE= 2.560
J77_78	6.9219	J R-Glc-5	R-Glc-6	STAT=Y	PRED= 2.700	RANGE= 3.000
J77_79	1.7617	J R-Glc-5	R-Glc-6'	STAT=Y	PRED= 1.280	RANGE= 3.000
J75_76	8.7207	J R-Glc-3	R-Glc-4	STAT=Y	PRED= 9.140	RANGE= 3.580
J75_81	4.8865	J R-Glc-3	R-OH-Glc-3	STAT=Y	PRED= 4.300	RANGE= 1.000
J76_82	5.8261	J R-Glc-4	R-OH-Glc-4	STAT=Y	PRED= 4.300	RANGE= 1.000
J78_79	-11.2374	J R-Glc-6	R-Glc-6'	STAT=Y	PRED= -12.300	RANGE= 0.800
J83_84	1.5856	J R-Rh-1	R-Rh-2	STAT=Y	PRED= 2.330	RANGE= 2.800
J87_88	6.2129	J R-Rh-5	R-Rh-Me	STAT=Y	PRED= 6.250	RANGE= 0.600
J87_86	9.2814	J R-Rh-5	R-Rh-4	STAT=Y	PRED= 9.940	RANGE= 3.580
J86_85	9.5011	J R-Rh-4	R-Rh-3	STAT=Y	PRED= 9.590	RANGE= 3.580
J86_91	3.6210	J R-Rh-4	R-OH-Rh-4	STAT=Y	PRED= 4.300	RANGE= 1.000
J84_85	3.0081	J R-Rh-2	R-Rh-3	STAT=Y	PRED= 3.420	RANGE= 3.800
J84_89	4.0866	J R-Rh-2	R-OH-Rh-2	STAT=Y	PRED= 4.440	RANGE= 1.500
J85_90	5.6937	J R-Rh-3	R-OH-Rh-3	STAT=Y	PRED= 4.300	RANGE= 1.000

CONTROL PARAMETERS:

```

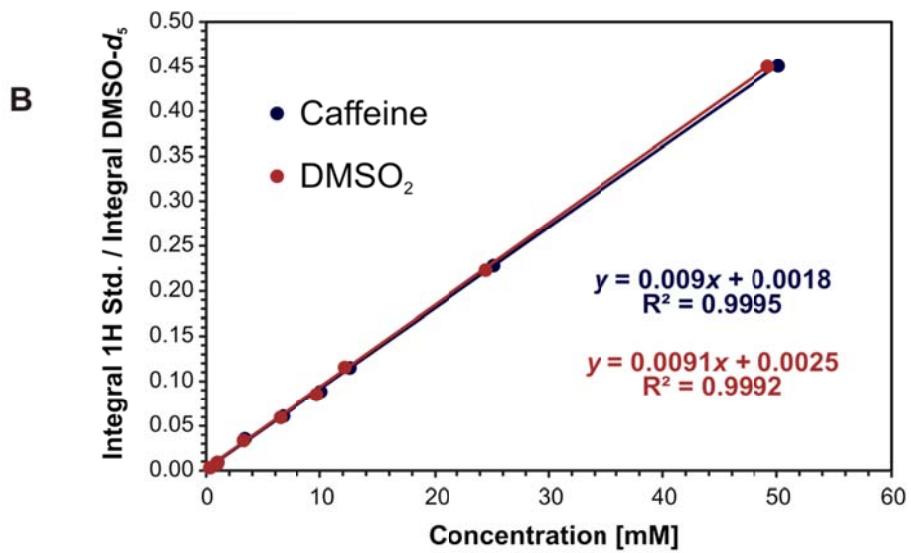
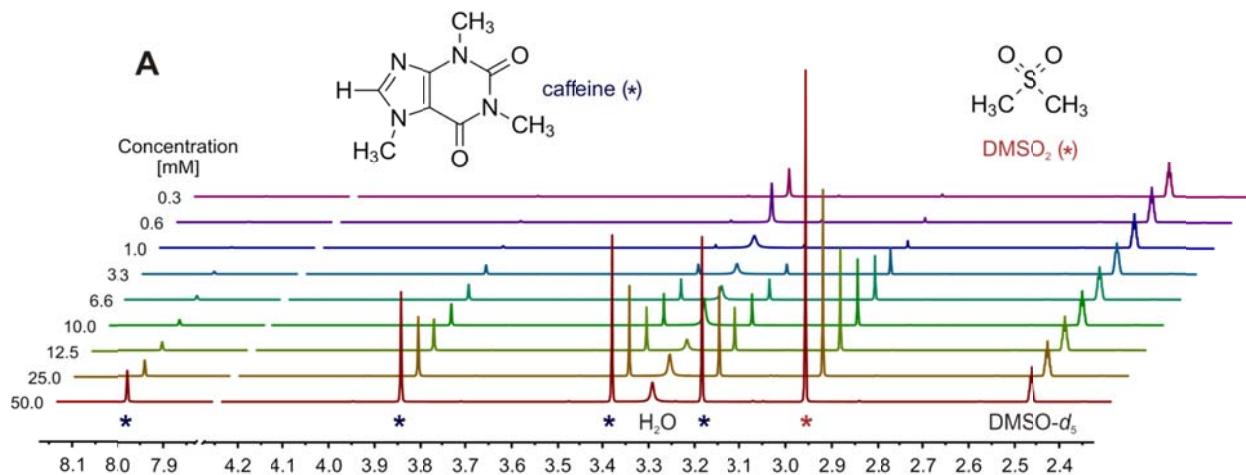
Solvent = none (def. 99% enriched)
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
8.04426687 = Left frequency (ppm)
0.00000000 = Right frequency (ppm)
10.000 = Acquisition time (s, for QMTLS)
1.505 = Line-width (for modes D, P & T, 0=use defaults)
0.068609932 = Data-point resolution (Hz)
25.060 = GAUSSIAN (%), 0=use default from INF)
2.716 = Dispersion contribution (%), 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)

```

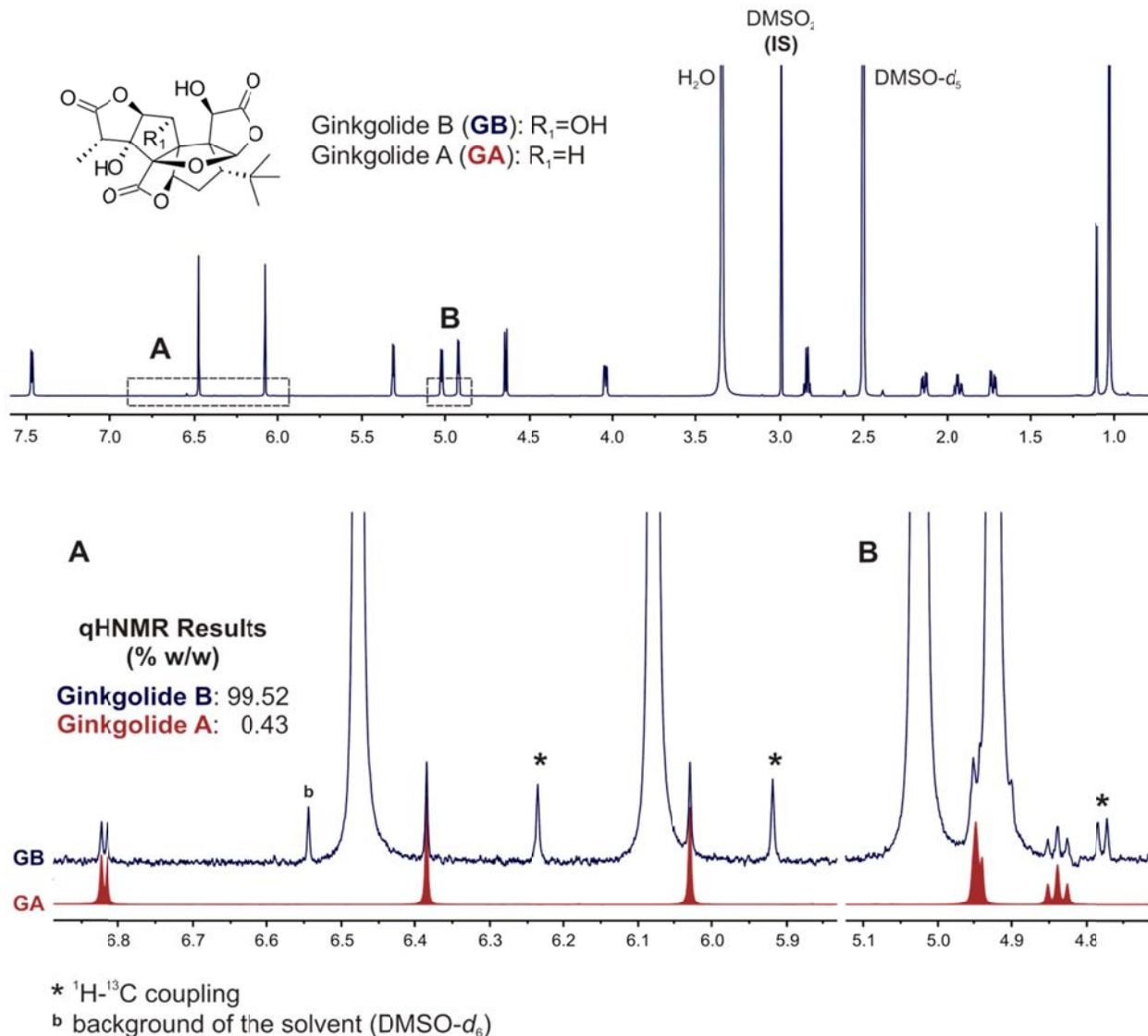
CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

END OF FILE

**S29.** Calibration of residual solvent signals using primary reference standards of  $\text{DMSO}_2$  and caffeine as internal calibrants. **(A)** qHNMR spectra of stock solutions of caffeine and  $\text{DMSO}_2$  in  $\text{DMSO}-d_6$  (600 MHz). **(B)** Calibration curves for the determination of residual  $\text{DMSO}-d_5$  content in  $\text{DMSO}-d_6$  lot B.



**S30.** The qHNMR analysis of a ginkgolide B reference material ( $\text{DMSO}-d_6$ , 600 MHz).  $\text{DMSO}_2$  was used as Internal Standard (IS) for quantitative calibration. Using qHNMR, ginkgolide A was readily detected as a very minor (0.43 %) impurity, present at an abundance level where intensities/integrals are comparable to those of  $^{13}\text{C}$  satellite signals of the major component (see also reference #17 in the main text).



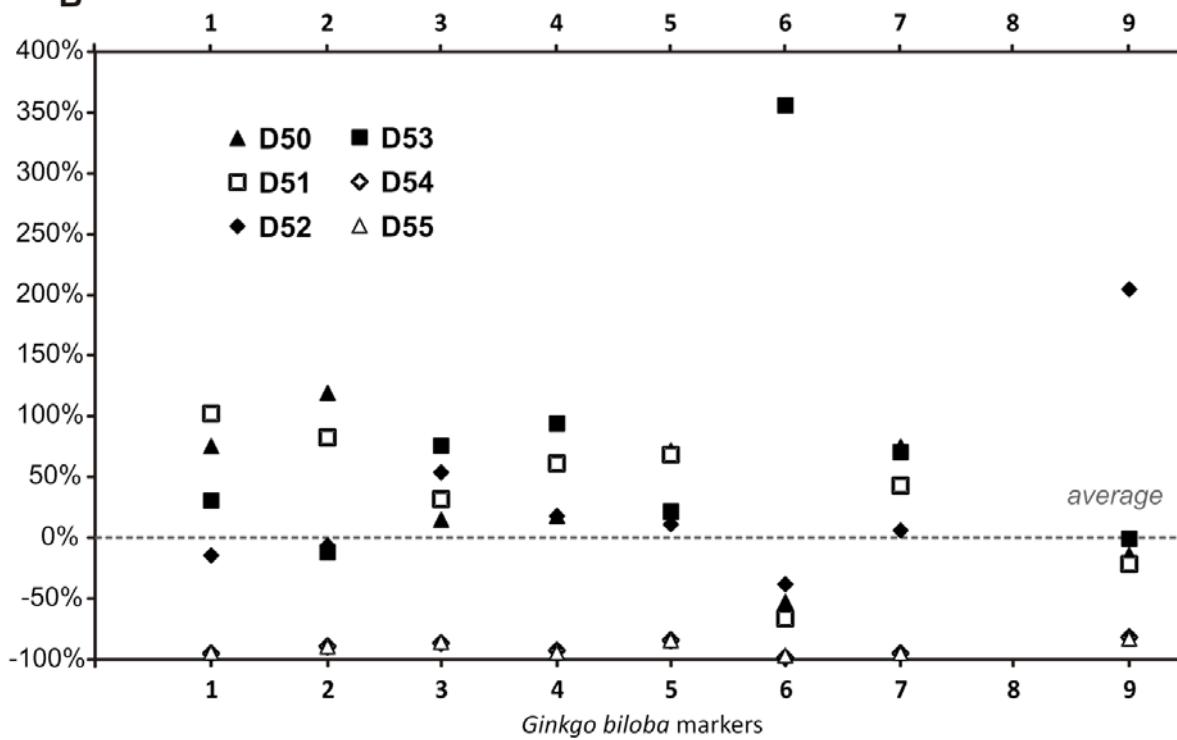
**S31.** (A) Weight percentage of compounds **1-9** in six Ginkgo samples according to qHNMR measurements. (B) Variability of selected botanical markers in commercially available *Ginkgo biloba* preparations, expressed as deviation (y-axis, in %) from the *average* percent content (set to 0% deviation, dotted line) of each of the compounds, **1-9**.

**A**

sample	<i>Ginkgo biloba</i> markers								
	concentration, % (w/w)								
	1	2	3	4	5	6	7	8	9
D50	2.34	1.92	1.15	0.68	4.03	0.73	0.38	Ø <sup>a</sup>	3.33
D51	2.70	1.60	1.32	0.93	3.95	0.52	0.31	Ø	3.05
D52	1.14	0.81	1.54	0.68	2.60	0.96	0.23	Ø	11.91
D53	1.74	0.77	1.76	1.12	2.85	7.14	0.37	Ø	3.86
D54	0.06	0.09	0.13	0.04	0.36	0.02	<0.01	Ø	0.70
D55	0.07	0.09	0.14	0.03	0.36	0.04	<0.01	Ø	0.65

<sup>a</sup>Ø denotes values below the calculated limit of detection (LOD).

**B**



**S32.** The effect of random noise on iterative fitting with PERCH in qHNMR analysis. **(A)** Sections of the qHNMR spectra of caffeine (DMSO-*d*<sub>6</sub>, 600 MHz), showing the olefinic proton signals ( $\delta_H$  8.01 ppm). The six qHNMR spectra were acquired using a different number of scans to reach the indicated signal-to-noise ratio (S/N) values. **(B)** Fitting of the signals of the olefinic proton of caffeine at the various S/N levels shown in **A**: comparison of experimental (*Exp*) and calculated (*Calc*) qHNMR spectra. While the root mean square (RMS) deviations for the iterative fits increase with decreasing S/N, the errors always remain <0.1%, even at a S/N level of ~11:1.

