## **Supplementary Material**

## Function of the cytochrome P450 enzymes MycCI and MycG in *Micromonospora griseorubida*, a producer of the macrolide antibiotic mycinamicin

Yojiro Anzai, Shu-ichi Tsukada, Ayami Sakai, Ryohei Masuda, Chie Harada, Ayaka Domeki, Shengying Li, Kenji Kinoshita, David H. Sherman, and Fumio Kato

Primer	Sequence (5'-3')	Restricton site
DmycCIF	CTGTCTGACGATGGGCAGAAGATGGTGGTCTGGCCCATGgggtgccagggcgtgccctt	-
DmycCIR	CCAGGACTATCCGCATACCGCACCCCCATTCGTCCGTCAtgtaggctggagctgcttc	-
DmycGF	TAGATATGAGGAGGGCTTTCTGGCAGCCTCGGTCGGGTGgggtgccagggcgtgccctt	-
DmycGR1	ACGTCACATCCTCCACCGCGTAGCGCGGGAACGCCGTTCtgtaggctggagctgcttc	-
DmycGR2	GCGGTCGGGTCGATCCGGCCCGACCACCGGCGCCTGTCAtgtaggctggagctgcttc	-
RecognitionDmycGF	CGGAGGAGAGTCGGGAGGTC	-
RecognitionDmycGR1	CGAACCCGAGATGCTGATTG	-
RecognitionDmycGR2	ACGAGATCGTCGAGATCGAC	-
mycCIproF	GCT <u>GCATGC</u> CAGAAATACCTCCTTGCCTGGTG	Sph I
mycCIIR	TGAC <u>AAGCTT</u> ACTCCTGTTGGCCCACCTGTCCCGTG	Hin dIII
myrBpF	GT <u>AAGCTT</u> AACGCTCGGTGATCGGGGTG	Hin dIII
myrBpR	GC <u>AAGCTT</u> GATGA <u>CATATG</u> TTTCCTTGGGTTC	Hin dIII, Nde I
myrBpF3	TGAAGCAGCAGGCCCGCCAGTTCCTTC	-
myrBpartialF	ACATGCTCCGGTACGACTTC	-
mycBpR1	CGACCGAGGCTGCCAGAAAG	-
myrBmycGR1	GGCTCGGTCTCCTGCAACTC	-
RecognitionDmycCIF	ACTTCCCGTCAGAATTCCTG	-
RecognitionDmycCIR	CAGCAGTTCCGGTGTCACTG	-
mycGpartialF	GTGCTCGGTGACGGACGCTTC	-
mycGpartialR	CCAGTGCCACCTGGAGTTCCAC	-
mycCIpartialF	TCAGCAGATACGAGCACGTC	-
mycCIpartialR	CGAGGACAGCAGAATTATCAG	-
152attPF	AATCGCTCTTCGTTCGTCTG	-
152intR	AATGCCCGACGAACCTGAAC	-
MGneo860F	TCTCCTGTCATCTCACCTTG	-
MGneo630R	TCATAGCCGAATAGCCTCTC	-

Table S1. PCR primers used in this study

The unique priming sites of disruption cassettes are shown in lower case. The relevant restriction sites for genetic manipulation are underlined.

	M-I <sup>a</sup>	M-III <sup>b</sup>	M-V <sup>a</sup>	M-IX <sup>a</sup>	F-1 (14-hydroxy-M-III, M-IX)				
Position	<sup>13</sup> C (ppm)	<sup>13</sup> C (ppm)	<sup>13</sup> C (ppm)	<sup>13</sup> C (ppm)		<sup>13</sup> C (ppm)	<sup>1</sup> H (ppm)	Multiplicity	Coupling (Hz)
1	166.7	166.1	166.3	166.4	С	166.4	-	-	-
2	120.1	120.9	120.7	121.1	СН	120.8	5.83	d	15.4
3	151.5	151.7	151.8	151.9	СН	152.0	6.60	dd	9.8,15.5
4	41.9	41.3	41.3	41.3	СН	41.3	2.71	m	-
5	87.5	87.8	87.7	87.9	СН	87.9	3.26 <sup>c</sup>	-	-
6	34.2	34.1	34.1	34.4	СН	34.1	1.22 <sup>c</sup>	-	-
7	32.1	32.6	32.7	33.2	$CH_2$	32.7	1.52	dd	3.4,14.0
							1.57 <sup>c</sup>	-	-
8	44.7	44.9	44.8	44.8	СН	44.9	2.54 <sup>c</sup>	-	-
9	200.8	203.7	203.8	203.4	С	203.9	-	-	-
10	125.6	123.2	123.8	124.6	СН	124.0	6.27	d	14.9
11	143.7	141.8	141.4	141.3	СН	141.4	7.14	dd	11.2,15.1
12	59.6	133.0	130.5	130.6	СН	130.8	6.47	dd	11.1,15.0
13	59.0	141.3	143.5	143.3	СН	143.4	6.02	d	15.1
14	47.5	49.2	77.4	77.6	СН	77.6	-	-	-
15	72.4	73.7	75.8	76.1	СН	75.7	4.81	dd	2.2,11.0
16	24.7	25.2	21.4	21.6	$CH_2$	21.5	1.56 <sup>c</sup>	-	-
							1.84	m	-
17	8.9	9.7	10.4	10.4	$CH_3$	10.4	0.91	t	7.5
18	18.9	19.4	19.5	19.5	$CH_3$	19.6	1.23	d	6.6
19	17.1	17.4	17.4	17.4	$CH_3$	17.4	0.99	d	6.6
20	17.5	17.7	17.6	17.6	$CH_3$	17.6	1.14	d	6.9
21	67.1	68.7	75.2	74.7	CH <sub>2</sub>	75.6	3.77	d	11.0
							3.88	d	10.0
1'	105.5	104.9	104.8	105.2	СН	104.9	4.24	d	7.3
2'	70.3	70.4	70.4	70.3	СН	69.3	3.49 <sup>c</sup>	-	-
3'	65.8	65.8	65.8	66.2	СН	66.0	2.55 <sup>c</sup>	-	-
4'	28.5	28.4	28.4	28.9	$CH_2$	28.7	1.26 <sup>c</sup>	-	-
							1.69	dd	1.7,12.7
5'	69.4	69.5	69.4	69.5	СН	69.1	4.21	t	3.1
6'	21.2	21.2	21.2	21.2	CH <sub>3</sub>	21.1	1.24	d	6.0
NMe <sub>2</sub>	40.1	40.2	40.2	40.4	CH <sub>3</sub>	40.3	2.34	S	-
1"	100.9	100.8	101.6	101.4	СН	101.5	4.57	d	7.8
2"	81.9	80.1	81.7	80.3	СН	79.9	3.02	dd	3.0,7.8
3"	79.3	69.8	79.2	70.9	СН	70.4	3.28 <sup>c</sup>		-
4"	72.7	72.8	72.5	72.9	СН	72.5	3.21	dd	3.1,9.5
5"	70.6	69.8	70.7	70.9	СН	70.2	3.69	m	-
6"	17.8	17.7	17.6	17.6	$CH_3$	17.5	1.30	d	6.2
2"-OCH <sub>3</sub>	59.0	59.4	59.1	58.9	$CH_3$	58.9	3.50	S	-
3"-OCH <sub>3</sub>	61.6	-	61.7	-	-	-	-	-	-

Table S2. <sup>13</sup>C- and <sup>1</sup>H-NMR data of F-1 (14-hydroxy-M-III, M-IX), and comparison with <sup>13</sup>C-NMR data of mycinamicins.

<sup>a</sup> Kinoshita, K., S. Takenaka, H. Suzuki, T. Morohoshi, and M. Hayashi. 1992. Mycinamicins, new macrolide antibiotics. XIII. Isolation and structures of novel fermentation products from *Micromonospora griseorubida* (FERM BP-705). J. Antibiot. 45:1-9.

<sup>b</sup> Hayashi, M., M. Ohno, K. Kinoshita, S. Satoi, M. Suzuki, and K. Harada. 1981. Mycinamicins, new macrolide antibiotics. III Isolation and structures of mycinamicin aglycones, mycinolide IV and V. J. Antibiot. 34:346-349.

<sup>c</sup> Overlapping

	M-I <sup>a</sup>	M-III <sup>a</sup>	F-2 (12,13-epoxy-M-III)				
	<sup>13</sup> C (ppm)	<sup>13</sup> C (ppm)		<sup>13</sup> C (ppm)	<sup>1</sup> H (ppm)	Multiplicity	Coupling (Hz)
1	166.7	166.1	С	165.7	-	-	-
2	120.1	120.9	СН	120.5	5.83	d	
3	151.5	151.7	СН	151.0	6.59	dd	
4	41.9	41.3	СН	41.7	2.72	m	
5	87.5	87.8	СН	87.6	3.32 <sup>b</sup>	-	-
6	34.2	34.1	СН	34.2	1.18 <sup>b</sup>	-	
7	21.1	32.6	$CH_2$	32.0	1.47 <sup>b</sup>	-	-
					1.62 <sup>b</sup>	-	-
8	44.7	44.9	СН	44.6	2.56	m	
9	200.8	203.7	С	200.7	-	-	-
10	125.6	123.2	СН	125.6	6.56	d	
11	143.7	141.8	СН	144.0	6.40	dd	
12	59.6	133.0	СН	59.0	3.25 <sup>b</sup>	-	-
13	59.0	141.3	СН	59.0	3.09	dd	
14	47.5	49.2	СН	47.4	1.40	m	
15	72.4	73.7	СН	72.5	5.32	m	
16	24.7	25.2	$CH_2$	24.8	1.54	m	
					1.86	qd	
17	8.9	9.7	$CH_3$	8.9	0.88	dd	
18	18.9	19.4	CH <sub>3</sub>	19.0	1.21	d	
19	17.1	17.4	$CH_3$	17.0	0.96	d	
20	17.5	17.7	CH <sub>3</sub>	17.5	1.15	d	
21	67.1	68.7	$CH_2$	67.4	3.60 <sup>b</sup>	-	
					4.18	dd	
1'	105.5	104.9	CH	103.6	4.35	d	
2'	70.3	70.4	CH	69.6	3.52	m	
3'	65.8	65.8	СН	66.7	3.30	m	
4'	28.5	28.4	$CH_2$	31.2	1.47	m	
					1.99	m	
5'	69.4	69.5	СН	67.9	3.57 <sup>b</sup>	-	-
6'	21.2	21.2	CH <sub>3</sub>	20.8	1.30	d	
7'	40.1	40.2	$CH_3$	39.0	2.85	br s	
8'	40.1	40.2	CH <sub>3</sub>	41.5	2.85	br s	
1"	100.9	100.8	СН	100.7	4.55	d	
2"	81.9	80.1	СН	80.1	3.06	dd	
3"	79.3	69.8	СН	69.5	4.20	dd	
4"	72.7	72.8	СН	72.9	3.24 <sup>b</sup>	-	-
5"	70.6	69.8	CH	70.0	3.65	dd	
6"	17.8	17.7	$CH_3$	17.6	1.30	d	-
2"-OCH <sub>3</sub>	59.0	59.4	$CH_3$	59.3	3.57	S	
3"-OCH <sub>3</sub>	61.6	-	-	-	-	-	-

Table S3. <sup>13</sup>C- and <sup>1</sup>H-NMR data of F-2 (12,13-epoxy-M-III), and comparison with <sup>13</sup>C-NMR data of mycinamicins.

<sup>a</sup> Kinoshita, K., S. Takenaka, H. Suzuki, T. Morohoshi, and M. Hayashi. 1992. Mycinamicins, new macrolide antibiotics. XIII. Isolation and structures of novel fermentation products from *Micromonospora griseorubida* (FERM BP-705). J. Antibiot. 45:1-9.

<sup>b</sup> Overlapping

	MIC (µg/ml)						
Microorganism	14-hydroxy-M-III (M-IX, F-1)	12,13-epoxyl-M-III (F-2)	M-III	M-IV			
S. aureus ATCC 25923	0.78	0.20	0.10	0.10			
M. luteus ATCC 9341	0.10	0.05	0.02	0.02			
E. coli ATCC 25922	100	100	>100	100			

Table S4. Antibacterial activities of mycinamicins



Figure S1. HPLC chromatograms of the EtOAc extract obtained from the culture broth of *M. griseorubida* A11725 (wild), TPMA0023 ( $\Delta mycCI$ ), and TPMA0034 (mycCI complemented). The EtOAc extract from the culture broth of TPMA0023 was not treated with 0.1% TFA. HPLC conditions: column, ODS-80T<sub>M</sub> (Tosoh); mobile phase, a 20-min. linear gradient from 35% MeCN to 70% MeCN containing 0.1% TFA; flow rate, 0.8 mL/min; UV wavelength, 200–300 nm. Blue line, at 240 nm; red line, at 280 nm.



Figure S2. HPLC chromatograms of the EtOAc extract obtained from the culture broth of wild strain *M. griseorubida* A11725, *mycG* disruption mutant TPMA0025, and *mycG* complementation strains TPMA0034, TPMA0047, and TPMA0062. HPLC conditions: column, ODS-80T<sub>M</sub> (Tosoh); mobile phase, MeCN, 0.06% TFA (35:65); flow rate, 0.8 mL/min; UV wavelength, 200–300 nm. Blue line, at 240 nm; Red line, at 280 nm.