

Supporting Information

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SI Discussion

Although carbachol activated the M1 receptor/Gq/PLC pathway, leading to the hydrolysis of PI(4,5)P₂ and activation of TRPC6, it failed to stimulate TRPML1 activity. In contrast, the breakdown of PI(4,5)P₂ by direct activation of 5-phosphatase was sufficient to activate TRPML1. One potential explanation for this discrepancy is that the affinity of ion channels determines their sensitivity to PI(4,5)P₂ (1). The insensitivity of TRPML1 to M1 receptor activation might reflect the channel's high affinity to PI(4,5)P₂ (IC₅₀ = 0.2 μM). Although we observed a rapid carbachol-induced translocation of GFP-PLCδ1-PH, a commonly used PI(4,5)P₂ probe with a K_d of 1.7 μM (2),

it remains unknown whether M1 receptor activation could sufficiently deplete the PI(4,5)P₂ level to modulate TRPML1. Moreover, activation of TRPML1 by GTP-γ-S, but not M1 receptor activation, could potentially be explained by a "receptor-specific phosphoinositide signaling" theory that highlights the importance of a local PI(4,5)P₂ pool in regulating the channels only adjacent to the receptors (3). Finally, the insensitivity of TRPML1 to M1 receptor/PLC activation could also be explained by the fact that receptor stimulation might activate other signaling pathways such as diacylglycerol, inositol triphosphate, Ca²⁺, and PKC, concurrently to nullify the effect of PI(4,5)P₂ depletion on TRPML1 activation.

1. Suh BC, Hille B (2008) PIP₂ is a necessary cofactor for ion channel function: How and why? *Annu Rev Biophys* 37:175–195.
2. Lemmon MA, Ferguson KM, O'Brien R, Sigler PB, Schlessinger J (1995) Specific and high-affinity binding of inositol phosphates to an isolated pleckstrin homology domain. *Proc Natl Acad Sci USA* 92:10472–10476.

3. Gamper N, Shapiro MS (2007) Regulation of ion transport proteins by membrane phosphoinositides. *Nat Rev Neurosci* 8:921–934.

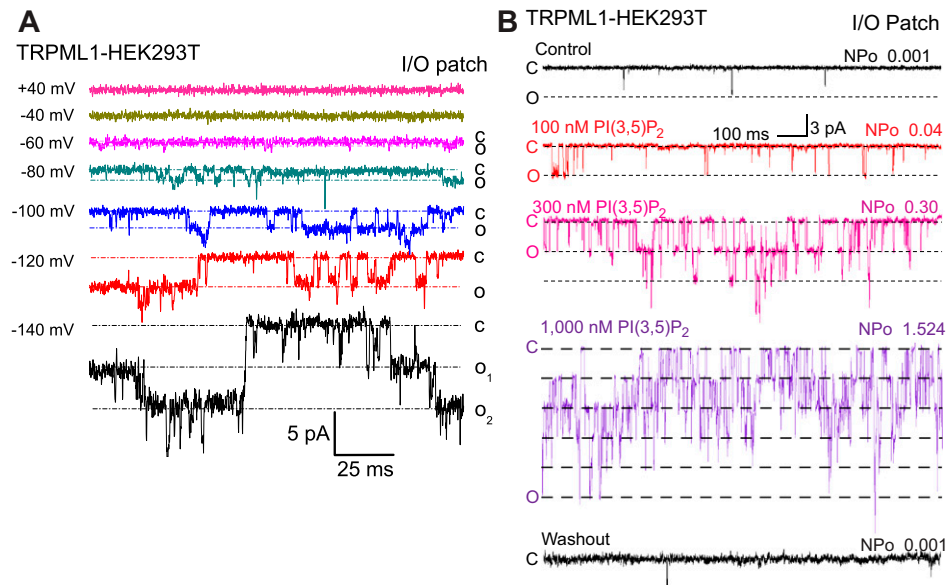


Fig. S1. Single-channel currents in an I/O patch excised from a TRPML1-expressing HEK293T cell. (A) Single-channel openings were elicited using a voltage step protocol from -140 mV to $+80$ mV (HP = 0 mV). At negative, but not positive potentials, channel openings were frequently observed. C, closed state; O, open state. (B) PI(3,5)P₂ activates single channel TRPML1 currents at the plasma membrane (PM) in a dose-dependent manner. The PI(3,5)P₂ effect was examined in an I/O patch excised from TRPML1-GFP-expressing HEK293T cells.

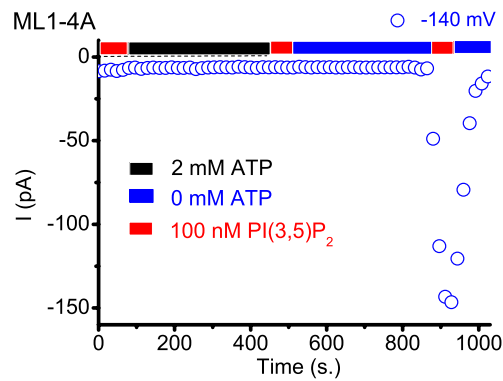


Fig. 52. Mg-ATP prevents run-up of TRPML1 currents in I/O patches. In an I/O patch, bath application of 2 mM Mg-ATP for 6 min prevented run-up of ML1-4A. Upon removal of Mg-ATP for 6 min, I_{ML1-4A} was activated by 100 nM PI(3,5)P₂.

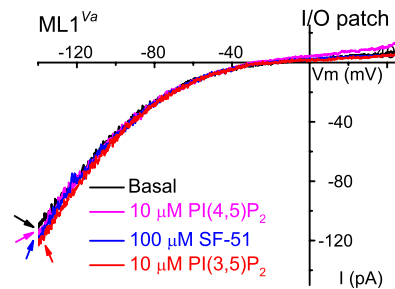


Fig. 53. Insensitivity of TRPML1^{Va} to PI(4,5)P₂. Large basal I_{ML1-Va} was detected in an I/O patch excised from ML1^{Va}-expressing cells. PI(4,5)P₂ (10 μM) failed to inhibit I_{ML1-Va} .

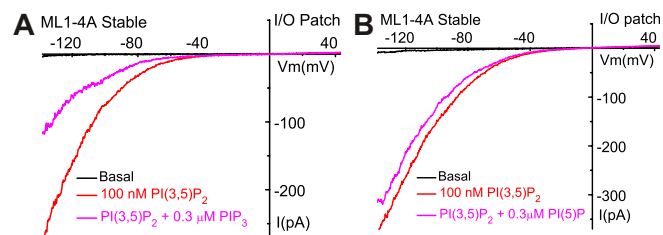


Fig. 54. Effects of PI(3,4,5)P₃ and PI(5)P on I_{ML1-4A} . (A) Inhibition PI(3,5)P₂-activated I_{ML1-4A} by 300 nM PI(3,4,5)P₃ in an I/O patch excised from ML1-4A stable cells. (B) Lack of an effect of PI(5)P (300 nM) on PI(3,5)P₂-activated I_{ML1-4A} .

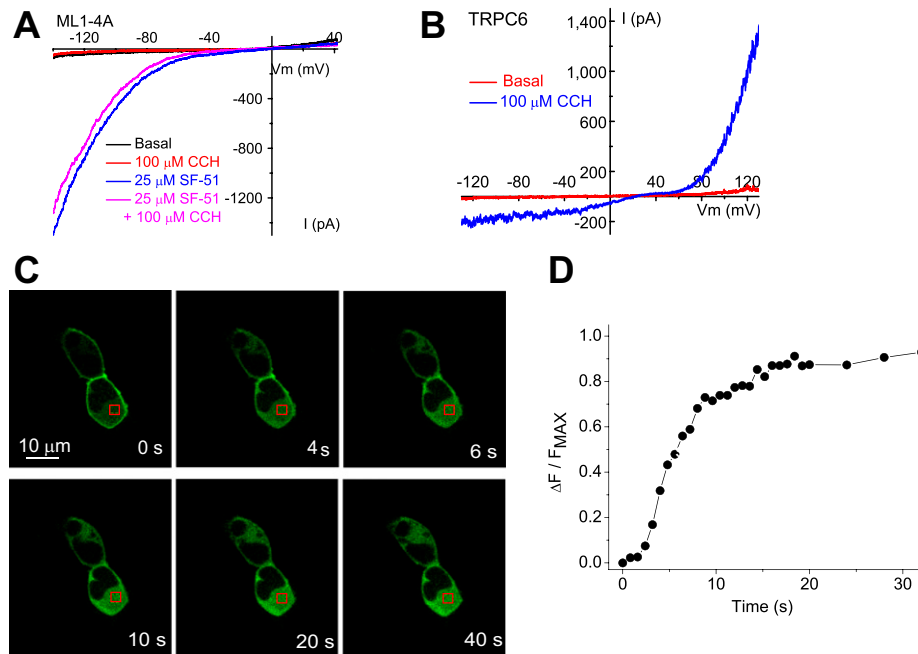


Fig. 55. Receptor-mediated activation of PLC induces PI(4,5)P₂ hydrolysis but fails to activate TRPML1. (A) Activation of PLC-coupled M1 receptors had no effect on I_{ML1-4A} . Carbachol (CCH; 100 μ M) was used to activate the Gq/PLC pathway in HM1 cells. (B) Whole-cell TRPC6 currents were activated by CCH (100 μ M). (C and D) CCH-induced hydrolysis of PI(4,5)P₂ detected using a PI(4,5)P₂ probe. (C) CCH (100 μ M) induced a translocation of GFP-PLC δ 1-PH from the plasma membrane to the cytosol. Images shown were taken 0, 4, 6, 10, 20, and 40 s after CCH application. Red boxes indicate cytosolic regions where fluorescence intensity (F) was measured. (D) Time course of GFP-PLC δ 1-PH translocation. Changes in average fluorescence intensity ($\Delta F = F - F_{\text{basal}}$) are normalized to maximum change of fluorescence intensity (F_{max}) and plotted with time. Data were analyzed with ImageJ software (National Institutes of Health).

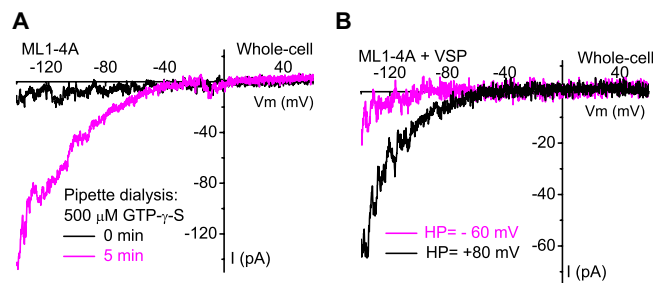


Fig. 56. GTP- γ -S and depolarization-activated Ci-VSP evoke whole-cell TRPML1 currents. (A) In an HM1 cell overexpressing TRPML1-4A, small whole-cell I_{ML1-4A} developed gradually upon pipette dialysis of GTP- γ -S (500 μ M) for 5 min. (B) Depolarization-activated Ci-VSP increases basal I_{ML1-4A} . Representative traces of whole-cell currents in Ci-VSP-transfected ML1-4A stable cells before (HP = -60 mV) and 8 min after depolarization (HP = +80 mV). I_{ML1-4A} was elicited by voltage ramps (from -140 mV to +100 mV; 100-ms duration).

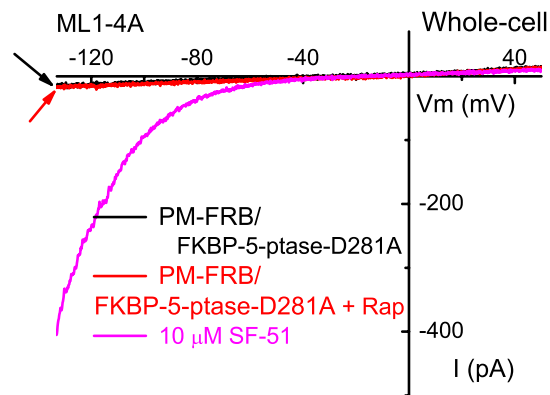


Fig. S7. PM-translocation of inactive mutant FKBP-5-phosphatase fails to activate whole-cell TRPML1 currents. No detectable I_{ML1-4A} was observed with or without Rapamycin ($0.5 \mu\text{M}$) application in ML1-4A-expressing HEK293 stable cells transfected with inactive mutant (D281A) of FKBP-5-phosphatase (FKBP-5-ptase-D281A) together with PM-FRB, which contains a signal for the PM localization. The dimerization of FRB and FKBP induced by rapamycin ($0.5 \mu\text{M}$ for 10 min) led to translocation of 5-phosphatase from the cytosol to the PM. I_{ML1-4A} was readily activated by SF-51 in the same cell.

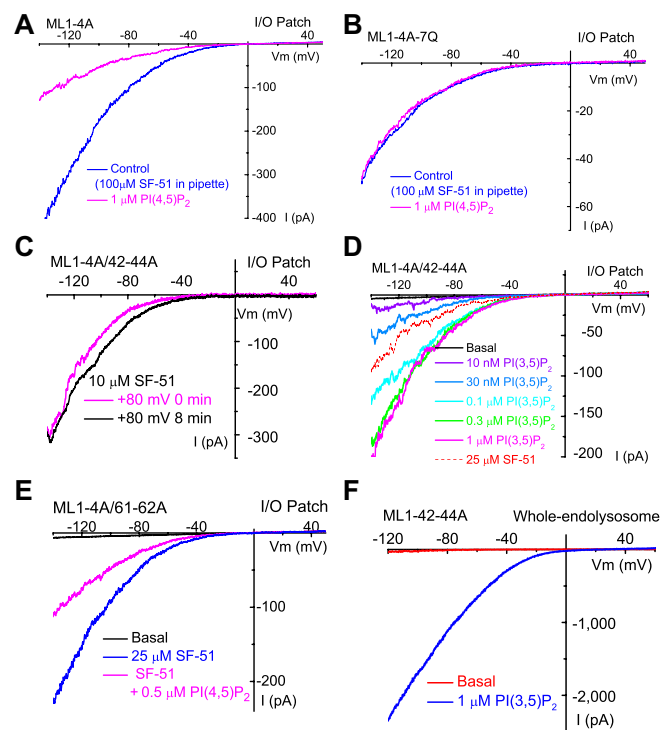


Fig. S8. Mutational analysis of $\text{PI}(4,5)\text{P}_2$ inhibition and $\text{PI}(3,5)\text{P}_2$ activation of TRPML1. (A and B) $\text{PI}(4,5)\text{P}_2$ inhibition is abolished by 7Q mutations in I/O patches. (A) Activation of I_{ML1-4A} by SF-51 ($100 \mu\text{M}$; included in pipette solution) was significantly inhibited by bath application of $\text{PI}(4,5)\text{P}_2$ ($1 \mu\text{M}$). (B) Activation of $I_{ML1-4A-7Q}$ by SF-51 ($100 \mu\text{M}$; included in pipette solution) was insensitive to $\text{PI}(4,5)\text{P}_2$ ($1 \mu\text{M}$). (C) Triple mutations in the PIP_2 -interacting domain abolish the activation effect of Ci-VSP. Activation of Ci-VSP by depolarization (HP = $+80 \text{ mV}$) failed to potentiate the SF-51 ($10 \mu\text{M}$)-activated $I_{ML1-4A/42-44A}$. (D) Dose-dependent activation of ML1-4A/42-44A by $\text{PI}(3,5)\text{P}_2$. $I_{ML1-4A/42-44A}$ was activated by various concentrations of $\text{PI}(3,5)\text{P}_2$ (10 nM , 30 nM , $0.1 \mu\text{M}$, $0.3 \mu\text{M}$, and $1 \mu\text{M}$). Effect of SF-51 ($25 \mu\text{M}$) is shown for comparison. (E) $\text{PI}(4,5)\text{P}_2$ sensitivity of R61A/K62A double mutations. Activation of $I_{ML1-4A/61-62A}$ by SF-51 ($25 \mu\text{M}$) was inhibited by $\text{PI}(4,5)\text{P}_2$ ($0.5 \mu\text{M}$). (F) Activation of ML1-42-44A by $\text{PI}(3,5)\text{P}_2$ in the lysosome. Whole-endolysosome I_{42-44A} was robustly activated by $\text{PI}(3,5)\text{P}_2$ ($1 \mu\text{M}$).