Supplemental information for

IL-6 TRIGGERS IL-21 PRODUCTION BY HUMAN CD4$^+$ T CELLS TO DRIVE STAT3-DEPENDENT PLASMA CELL DIFFERENTIATION IN B CELLS

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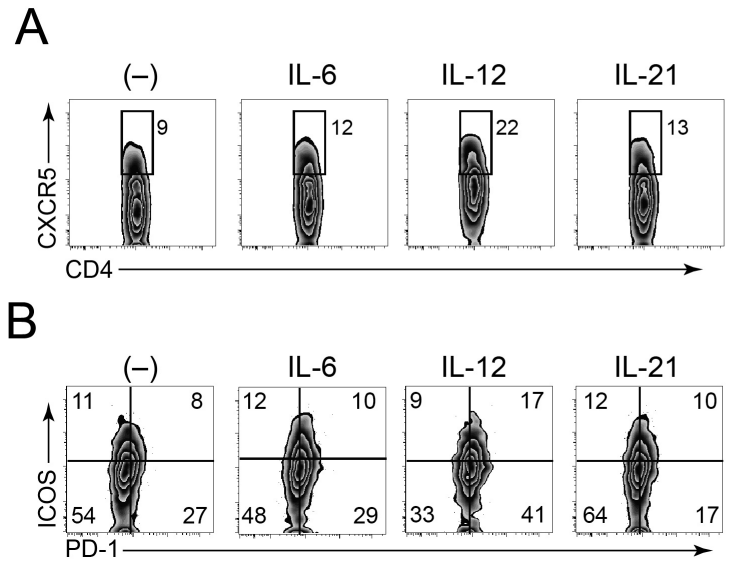

Figure S1: Representative surface phenotype plots for data presented in Fig 4. (**A**) CXCR5 expression on $CD4^+CD3^+$ T cells following *in vitro* stimulation with anti-CD3/CD28 in the presence or absence (–) of the indicated cytokines. (**B**) ICOS and PD-1 expression on $CXCR5^+CD4^+$ T cells. Gates were determined based on isotype control staining.

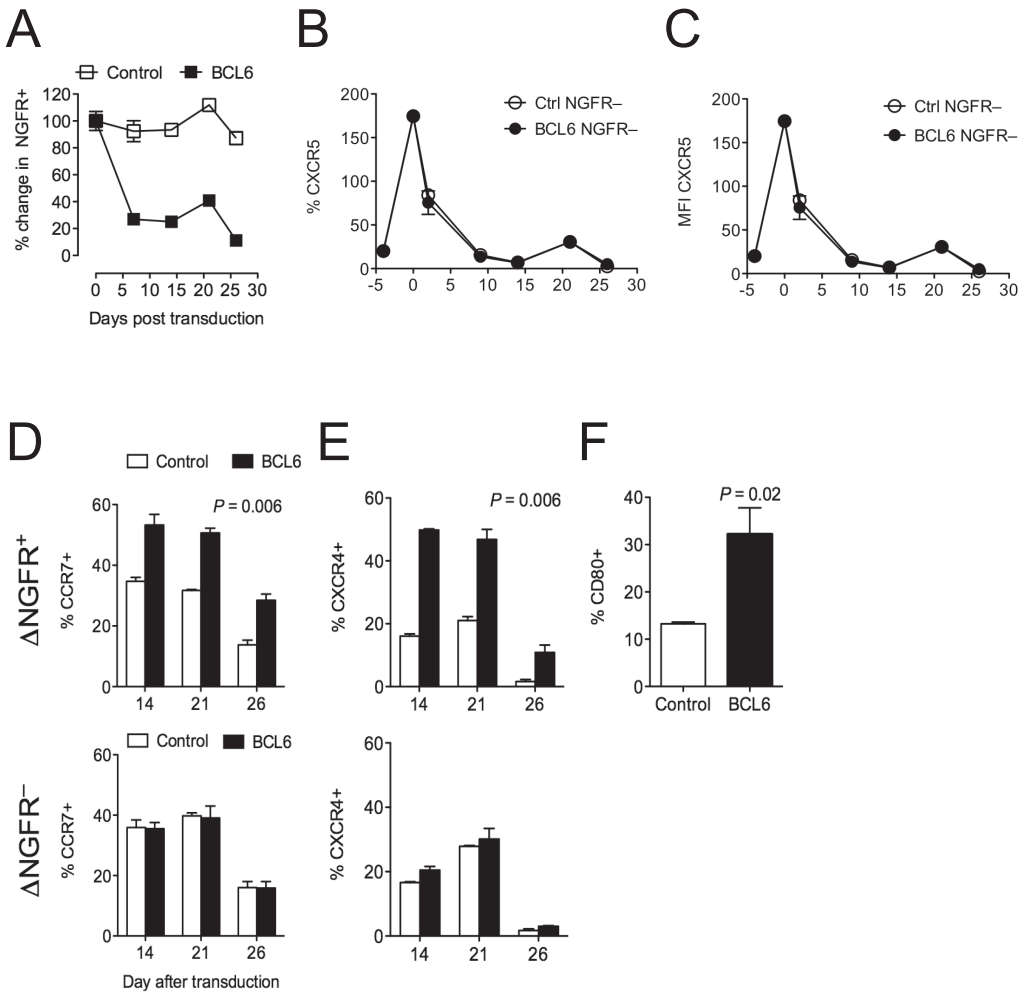

Figure S2: BCL6 overexpression in primary human CD4$^+$ T cells.

Total CD4$^+$ T cells from peripheral blood were transduced with retroviral vectors encoding either LZRS-IRES-ΔNGFR (Control, open symbols) or LZRS-BCL6-IRES-ΔNGFR (BCL6, closed symbols). Transduced cells were cultured on irradiated allogeneic PBMC feeders with PHA (2 μg/ml) and IL-2 (20 U/ml) (methods). (**A**). Frequency of NGFR$^+$ cells over time relative to initial transduction efficiency as determined by flow cytometry. Gated on CD4$^+$NGFR$^–$ cells, (**B**) Percent CXCR5$^+$ cells and (**C**) CXCR5 surface mean fluorescence intensity (MFI) were determined. Quantitation of (**D**) CCR7 and (**E**) CXCR4 expression on from CD4$^+$NGFR$^+$ (upper panels) CD4$^+$NGFR$^–$ gated cells (lower panels). See Figure 6 for representative FACS plots. (**F**) Quantitation of CD80 expression in CD4$^+$NGFR$^+$ cells at day 21 after

transduction. Results are means ± SD of two independent experiments each containing two donors. Two-way ANOVA was used to calculate *P*-values for data in panels D, E and student's t-test for data in panel F.