SUPPLEMENTARY DATA

Myosin Regulatory Light Chain Diphosphorylation Slows Relaxation of Arterial Smooth Muscle

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FIGURE LEGENDS

Supplementary FIGURE S1. Effect of exogenous calmodulin and MLCK on LC_{20} phosphorylation in Triton-skinned rat caudal arterial smooth muscle. Triton-skinned rat caudal arterial smooth muscle strips were incubated for 15 min in pCa 9 solution in the absence or presence of exogenous calmodulin (CaM) (1.8 µM) and/or MLCK (10 or 50 nM). [Ca²⁺] was then increased to pCa 4.5 under otherwise identical conditions and, after steady-state force was achieved (10 and 20 min), tissues were harvested for Phos-tag SDS-PAGE and western blotting with anti-pan LC_{20} . A control experiment in which tissue was treated with microcystin at pCa 9 was included to indicate the positions of unphosphorylated (0P), mono- (1P) and diphosphorylated LC_{20} (2P) bands. Phosphorylation levels were quantified and the results are shown in Supplementary Table 1.

Supplementary FIGURE S2. Identification of thiophosphorylated forms of LC_{20} using phosphospecific antibodies. Triton-skinned rat caudal arterial smooth muscle strips were treated as shown in Fig. 8A and harvested at selected times for Phos-tag SDS-PAGE and western blotting with anti-pan LC_{20} , anti-pS19- LC_{20} , anti-pT18- LC_{20} and anti-pT18,pS19- LC_{20} . A control experiment (lane 1) in which tissue was treated with microcystin at pCa 9 was included to indicate the positions of unphosphorylated (0P), mono- (1P) and diphosphorylated LC_{20} (2P) bands. Tissue incubated at pCa 9 (lane 2) is also included as a control. Numbers 3 - 6 indicate tissues harvested at the times indicated in Fig. 8A.

Supplementary FIGURE S3. ILK does not use ATP γ S as a substrate. Triton-skinned rat caudal arterial smooth muscle strips were treated as shown in *A*. Tissues were harvested at the indicated times for Phostag SDS-PAGE and western blotting with anti-pan LC₂₀ (*B*). Numbers 1 (pCa 9) and 2 (pCa 4.5 + ATP γ S) correspond to the times indicated in Fig. 8*A*, and numbers 3 and 4 to the times indicated in panel *A* of this figure (i.e. tissue treated with ATP γ S + microcystin at pCa 9 and following washout at pCa 9, respectively).

Supplementary FIGURE S4. Contraction of Triton-skinned rat caudal arterial smooth muscle induced by Ca²⁺ or okadaic acid in the absence of Ca²⁺. Following control Ca²⁺-induced contraction-relaxation cycles, Triton-skinned rat caudal arterial smooth muscle strips were treated with okadaic acid (OA; 20 μ M) at pCa 9 (*A*) or with Ca²⁺ (pCa 4.5) (*B*). Relaxation followed washout of OA or removal of Ca²⁺, respectively.

Supplementary FIGURE S5. Dephosphorylation of MYPT1 following washout of okadaic acid. Tissues were treated as in Fig. S4A and harvested at the indicated times as follows: under relaxed conditions at pCa 9 (pCa 9); at the peak of pCa 4.5-induced contraction (pCa 4.5); at the plateau of okadaic acid-induced contraction at pCa 9 (100% Force) and during the relaxation following washout of okadaic acid corresponding to force levels of 90, 80, 70, 60, 50 and 25% and at complete relaxation (0% Force). Tissue samples were subjected to SDS-PAGE and western blotting with anti-pT697-MYPT1 or anti-pT855-MYPT1. Blots were also probed with anti-actin (shown beneath the corresponding phosphospecific antibody blots). Identical results were obtained in 5 independent experiments.

Conditions	% total LC ₂₀		
	0P	1P	2P
	o 4 -	(7 0	0
pCa 4.5, 10 min	34.7	65.3	0
pCa 4.5, 20 min	22.6	76.0	1.4
pCa 4.5, CaM, 10 min	16.7	80.1	3.2
pCa 4.5, CaM, 20 min	14.7	81.3	4.0
nC_{2} 4.5 $C_{2}M$ MICK (10 nM) 10 min	22.3	73 5	4.2
pCa 4.5, CaN, MLCK (10 nM), 10 min	17.8	77.7	4.5
a Co 4.5, Co M, MI CV (50 a M), 10 aria	20.7	75 1	4.2
pCa 4.5, Calvi, MILCK (50 mM), 10 min	20.7	/3.1	4.2
pCa 4.5, CaM, MLCK (50 nM), 20 min	24.0	70.3	5.7
MC, pCa 9	16.8	26.4	56.8

Supplementary Table 1. Effect of exogenous calmodulin and MLCK on LC₂₀ phosphorylation in Triton-skinned rat caudal arterial smooth muscle¹

¹These data were calculated from the western blot in Supplementary Fig. S1.













