Supplemental Materials for

The NOX4 pathway as a source of selective insulin resistance and responsiveness

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Supplemental Background

Prior literature has documented major site-specific phosphorylations and functions in the AKT limb of insulin receptor signaling (see Figure 1). Insulinstimulated phosphorylation of FOXO1 has been reported at Thr24, a key site normally phosphorylated by activated AKT, leading to exclusion of FOXO1 from the nucleus¹ and arresting its activity as a transcription factor for apoC-III and gluconeogenic genes.²⁻⁵

The AKT limb also affects several molecules involved in lipogenesis. The AKT target site on glycogen synthase kinase-3ß (GSK3ß) is Ser9.⁶ Phosphorylation by AKT inactivates GSK3,⁶ and may thereby block this factor from inhibiting ACL, a key enzyme in fatty acid, cholesterol, and new glucose biosynthesis,^{7, 8} as well as glycogen synthase. Insulin induces activation and site-specific serine-phosphorylation of a second enzyme in fatty acid biosynthesis, acetyl-CoA carboxylase-1 (ACC).^{9, 10} The effect of phosphorylation of ACC at Ser79 on its activity in vitro remains an open question,¹¹ whereas in vivo, T2DM increases hepatic ACC activity and Ser79-phosphorylation together.¹² We infer that overphosphorylation at Ser79 in T2DM liver in vivo could be driven by hyperinsulinemia in combination with continued responsiveness of this portion of the insulin receptor-AKT signaling cascade.

Another lipogenic target of AKT is the mammalian target of rapamycin complex-1 (mTORC1), which is activated in the tissues of obese mice.^{13, 14} This complex has two AKT-dependent inputs: PRAS40¹⁵ and TSC2.^{16, 17} Insulin stimulates AKT to phosphorylate PRAS40^{15, 18} at Thr246.¹⁹ Insulin-stimulated phosphorylation of TSC2 occurs at a key site acted on by AKT, Thr1462.²⁰ Insulin-induced mTORC1 activity can be assess by the phosphorylation of one of its substrates – namely, the ribosomal protein S6 kinase 1 (S6K1), an enzyme that desensitizes IRS1 in states of overnutrition and obesity (reference²¹ and Figure 1). Activated mTORC1 induces *Srebp1c* mRNA, which encodes a major insulin-responsive transcription factor in lipogenesis.²²⁻²⁶ Activated mTORC1 might also increase lipogenesis through induction of ER stress¹⁷ and hence

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cleavage and activation of the SREBP1c protein.^{27, 28} All of these effects are displayed schematically in Figure 1.

Pathways of interest that are not in Figure 1 include suppression of Irs2 mRNA by insulin;²⁹ insulin-induced activation of mitogen-activated protein kinases in addition to ERK1,2;³⁰ our sequence analysis of NOX4 and subsequent inference that NOX4 in cholesterol-rich caveolae could contribute to the putative generation of oxysterol ligands for LXR after insulin stimulation (Methods); inhibition of GLUT translocation by activated mTORC1;³¹ insulin-stimulated production of the vasodilator NO via activated AKT and of the vasoconstrictor endothelin-1 via activated ERK:³² direct effects of activated PI3K on ER stress:^{33,} ³⁴ insulin-stimulated phosphorylation and inhibition of FOXA2, a transcriptional factor that otherwise drives the expression of genes encoding enzymes of fatty acid oxidation, ketogenesis, and glycolysis;³⁵ insulin-induced cleavage and activation of SREBP1c protein via PI3K;³⁶ ERK-mediated phosphorylation of SREBP1c at Ser93, which enhances transactivation of its target genes;³⁷⁻³⁹ the ability of activated MEK to bind, inhibit, and provoke the expulsion of PPARy from the nucleus;^{40, 41} and effects of insulin on sympathetic activity,^{42, 43} renal sodium excretion,^{42, 44-46} ERK-mediated phosphorylation and activation of the Na⁺/K⁺ ATPase,⁴⁷ coagulation,⁴⁸⁻⁵⁰ expression of matrix metalloproteinases,³⁰ and secretion of apoB-containing lipoproteins.^{5, 51-54}

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Supplemental figure legends

Supplemental Figure I: Type 2 diabetes renders the liver unable to inactivate PTP1B in response to insulin.

Displayed are PTP1B activities from the same liver samples as in Figure 2, which were obtained just before (*Pre*) and 10min after (*Post*) an intravenous injection of insulin into lean db/m mice (controls) and their hyperphagic, obese T2DM db/db littermates, as indicated. PTP1B activities in liver homogenates were assayed under strictly anaerobic conditions (mean±SEM, n=3). Statistical comparisons by the paired *t*-test are indicated.

Supplemental Figure II: Type 2 diabetes impairs a key insulin-stimulated hypolipidemic and hypoglycemic pathway in liver, yet preserves lipogenic pathways and robust ERK activation.

Displayed are immunoblots from the same liver samples as in Figure 2, which were obtained just before (*Pre*) and 10min after (*Post*) an intravenous injection of insulin into lean *db/m* mice (controls) and their hyperphagic, obese T2DM *db/db* littermates, as indicated. Panel A: Resistance of FOXO1 to insulin-stimulated phosphorylation in T2DM *db/db* livers compared to control *db/m* livers (*TG-rich lipoprotein clearance* and *Gluconeogenesis* pathways from Figure 1), yet continued responsiveness of GSK3ß (*Lipogenesis-I* pathway from Figure 1). Panel B: Continued activation of molecules upstream (PRAS40) and downstream (S6K1) of mTORC1 in T2DM *db/db* livers (*Lipogenesis-D* pathway from Figure 1). The 70-kDa isoform of S6K1 is indicated. Panel C: Continued responsiveness of ERK to insulin-stimulated phosphorylations in T2DM *db/db* livers (*pT202-ERK*, *pY204-ERK*). Immunoblots for total (*t*-, meaning phosphorylated plus unphosphorylated) amounts of each target protein are shown for each sample. Numbers over the lanes refer to individual animals.

Supplemental Figure III: Inhibition of NOX4 in primary rat hepatocytes impairs the ability of insulin to suppress *Irs2* mRNA levels. Displayed are mRNA quantifications from the same set of cultured hepatocytes as in Figure 5A-C. As indicated, primary rat hepatocytes were pretreated with 0 (vehicle) or 1.0 μ M DPI (an inhibitor of NOX4), exposed to 0 or 10nM insulin for 6 h, and then harvested. Displayed are *Irs2* mRNA levels normalized to *B-actin* mRNA levels (Δ Ct) and then expressed relative to the mean value from the cells that had been incubated with neither DPI nor insulin (2^{- $\Delta\Delta$ Ct}; mean±SEM, n=4). P<0.001 by ANOVA; columns labeled with different lowercase letters (a, b, c) are statistically different by the Student-Newman-Keuls test (P<0.01).

Supplemental Table I: Antibodies against target proteins.

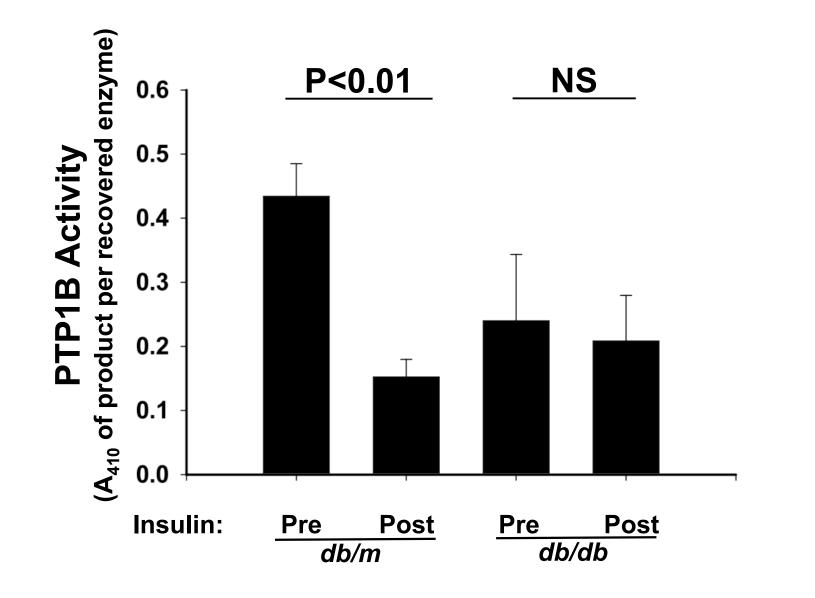
Target Protein	Epitope	Supplier/ Catalog Number	Description	Molecular weight of target protein
ACC	p-ACC	Cell Signaling #3661	Rabbit polyclonal antibody (Ab) against acetyl-CoA carboxylase that is phosphorylated at Ser79	280 kDa
	t-ACC	Cell Signaling #3676	Rabbit monoclonal antibody (mAb) against total acetyl-CoA carboxylase (phosphorylated and non-phosphorylated)	280 kDa
AKT	рТ308-АКТ	Cell Signaling #4056	Rabbit mAb against AKT phosphorylated at Thr308	60 kDa
	pS473-AKT	Cell Signaling #4051	Mouse mAb against AKT phosphorylated at Ser473	60 kDa
	t-AKT	Cell Signaling #2920	Mouse mAb against total AKT (phosphorylated and non- phosphorylated; clone 40D4)	60 kDa
ß-actin	ß-actin	Cell Signaling #3700	Mouse mAb against ß-actin	45 kDa
ERK	pT202-ERK pY204-ERK	Cell Signaling #4376	Rabbit mAb against Erk1/2 that is phosphorylated at Thr202, regardless of the status of Tyr204 Rabbit mAb against Erk1/2 that is	42, 44 kDa
		Cell Signaling #4377	phosphorylated at Tyr204, regardless of the status of Thr202	42, 44 kDa
	t-ERK	Cell Signaling #4695	Rabbit mAb against total ERK1/2, also known as the P44/42 mitogen- activated protein kinase	42, 44 kDa
FOXO1	t-FOXO1	Cell Signaling #9454	Rabbit polyclonal Ab against total FOXO1 (used here for immunoprecipitation)	78 to 82 kDa
	pT24-FOXO1	Cell Signaling #9464	Rabbit polyclonal Ab against FOXO1 phosphorylated at Thr24 or FOXO3A phosphorylated at Thr32	78 to 82, 95 kDa
	t-FOXO1	Cell Signaling #2880	Rabbit mAb against total FOXO1 (used here for immunoblotting)	78 to 82 kDa
GSK3ß	pS9-GSK3ß	Cell Signaling #9322	Rabbit mAb against GSK3ß phosphorylated at Ser9	46 kDa
	t-GSK3ß	Cell Signaling #9315	Rabbit mAb against total GSK3ß	46 kDa
NOX4	NOX4	LifeSpan Biosciences #LS-C33986	Rabbit polyclonal Ab against NOX4	67 kDa (isoform 1) 31.8 kDa (isoform 4)
PRAS40	pT246-PRAS40	Cell Signaling #2997	Rabbit mAb against PRAS40 phosphorylated at Thr246	40 kDa
	t-PRAS40	Cell Signaling #2691	Rabbit mAb against total PRAS40	40 kDa
PTEN	PTEN	Cell Signaling #9188	Rabbit mAb against PTEN (used here for immunoprecipitation)	54KDa
	PTEN	Cell Signaling #9556	Mouse mAb against PTEN (used here for immunoblotting)	54KDa
PTP1B	PTP1B	Santa Cruz sc-1718	Goat polyclonal Ab against PTP1B (used here for immunoprecipitation)	50KDa
	PTP1B	Santa Cruz sc-1718-R	Rabbit polyclonal Ab against PTP1B (used here for immunoblotting)	50KDa
S6K1	pT389-S6K1 (70-kDa isoform)	Cell Signaling #9206	Mouse mAb against the 70-kDa isoform of S6 kinase-1 that is phosphorylated at Thr389	70, 85 kDa
	t-S6K1	Cell Signaling #2708	Rabbit mAb against total S6K1	70, 85 kDa
SULF2	SULF2	Santa Cruz sc-68436 (Lot # C1909)	Rabbit polyclonal Ab against SULF2	72 kDa
TSC2	pT1462-TSC2	Cell Signaling #3617	Rabbit mAb against TSC2 (tuberin) phosphorylated at Thr1462	200 kDa
	t-TSC2	Cell Signaling #4308	Rabbit mAb against total TSC2	200 kDa

Supplemental Table II: Primer and probe sequences for quantitative real-time PCR.

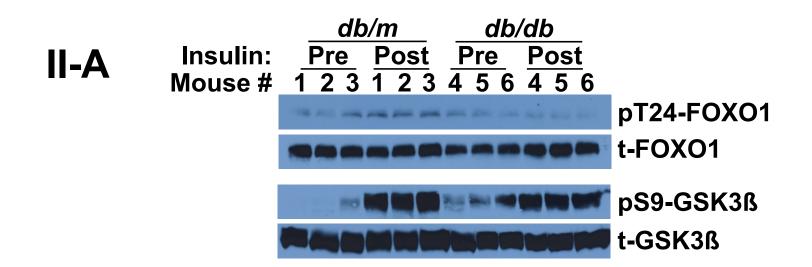
Target mRNA	Forward primer	Reverse primer	Probe
Арос3	5'-atg cag ggc tac atg gaa ca-3'	5'-cac agc tat atc aga ctc ct-3'	F-5'-tcc aag acg gtc cag gat gca ct- 3'-Q
lrs2	5'-atg aac ctg gac ttc agt tct-3'	5'-atc cat gga gcc tac tgt gt-3'	F-5'-tcc ccc aag cct agc acc cgc-3'- Q
Pepck	5'-agt cac cat cac ttc ctg ga-3'	5'-cag aat cgc gag ttg gga tg-3'	F-5'-cg gtt cct cat cct gtg gtc tcc ac- 3'-Q
Srebp1c	5'-gga gcc atg gat tgc aca tt-3'	5'-cat caa ata ggc cag gga ag-3'	F-5'-tg ctt cag ctc atc aac aac caa gac a-3'-Q
ß-actin	5'-tgc ctg acg gtc agg tca-3'	5'-cag gaa gga agg ctg gaa g-3'	F-5'-ca cta tcg gca atg agc ggt tcc g- 3'-Q

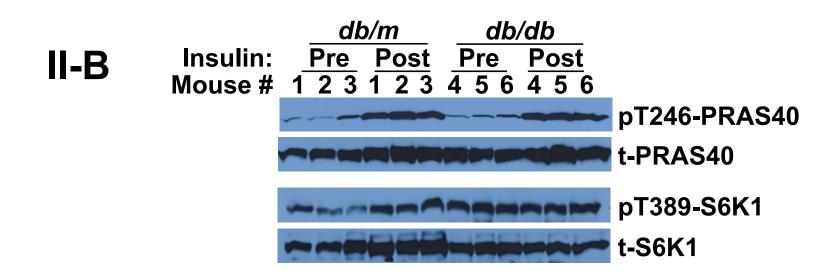
In the probe sequences, F and Q denote the positions of the FAM fluorophore and TAMRA quencher, respectively.

Supplemental Figure I

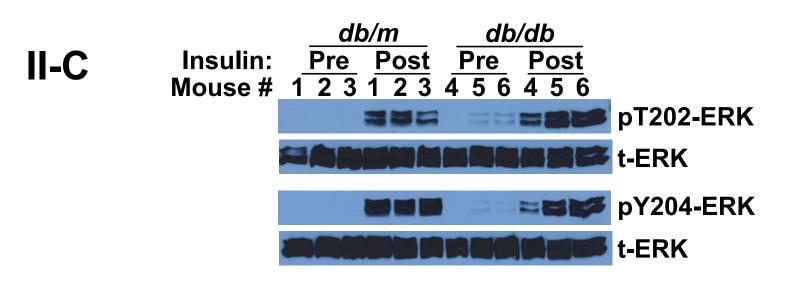


Supplemental Figure II-A,B





Supplemental Figure II-C



Supplemental Figure III

