

Supplemental Figure 1. Copper or ascorbate alone do not elicit plasma membrane currents; copper and ascorbate together elicit a barium-permeable inward conductance. Whole cell patch clamp recordings from epidermal protoplast plasma membrane. Recording conditions as in Figure 1 unless otherwise stated. (A) Col-0. Left, control currents (*I*) elicited by step voltage (*V*) changes, from a representative protoplast. Centre, *I* after exposure to extracellular 1 mM CuCl₂. Right, mean ± S.E.M. *I-V* relationships for control (open circles) and test exposure (n = 3). (B) Col-0 response to 1 mM ascorbate (n = 3). (C) ann1/ANN1 response to 1 mM CuCl₂ (n = 3). (D) WS response to 1 mM CuCl₂ (n = 3). (E) Col-0 response to extracellular OH' generated by 1 mM copper and 1 mM ascorbic acid with BaCl₂ replacing CaCl₂ in the bath solution (n = 3). Negative current to the left of the equilibrium potential for K⁺ (E_{κ}) can be carried by Ba²⁺ (as a proxy for Ca²⁺) with a possible K⁺ component. Negative current to the right of E_{κ} is carried by Ba²⁺.

Figure S2



Supplemental Figure 2. Expanded planar lipid bilayer traces. Current traces are those shown in Figure 7C for control (**A**) and in the presence of hydroxyl radicals (**B**). There are measurable increases in current magnitude evoked by radicals even at the lowest voltages applied (±20 mV).

Supplemental Data. Laohavisit et al. (2012). Plant Cell 10.1105/tpc.112.097881 Figure S3



Supplemental Figure 3. Immunoprecipitated ANN1 does not support a conductance in planar lipid bilayers. The ANN1 preparation was incubated for 30 minutes with anti-ANN1 peptide antibody in a 2:1 ratio. Antibody (Ab) alone was used as a control. (A). Schematic representation of the voltage pulse protocol used in planar lipid bilayer studies. (B). Representative current traces and mean IV relationships. The cis chamber contained 1 mM Ca²⁺, the trans contained 200 mM Ca²⁺, both pH 6. Left panel, OH[•] were generated in the *trans* chamber by Cu-Asc. Centre panel, OH were generated in the trans chamber by Cu-Asc, the cis chamber contained Ab. Right panel, *IV* relationship for the PLB with Ab and OH[•]. (C). Ionic conditions as (B) but in the centre panel, immunoprecipitated ANN1 (ANN1/Ab) was in the cis chamber and OH[•] were generated in the trans chamber. Right panel, IV relationship for ANN1/Ab. (D). Cis contained 200 mM KCl; trans 50 mM KCl; both pH 6. Left panel, OH[•] were generated in the trans chamber. Centre panel, OH[•] were generated in the *trans* chamber, ANN1/Ab was in the *cis* chamber. Right panel, IV relationship for ANN1/Ab. (E). Cis chamber held 200 mM K⁺, 1 mM CaCl₂ and the trans held 1 mM K⁺, 200 mM CaCl₂, both pH 6. Left panel, OH[•] were generated in the trans chamber. Centre panel, OH were generated in the trans chamber, ANN1/Ab was in the cis chamber. Right panel, *IV* relationship for ANN1/Ab. Observations were made for 2 hours. For all data sets, n = 43 except (**E**), where n = 3.

Supplemental Table 1. Root plasma membrane currents. Mean \pm SEM currents (in pA) are reported for epidermal protoplasts (n = 6) and root hair apical spheroplasts (n = 6) of the genotypes used in this study, determined using whole cell mode of patch clamp electrophysiology. *ann1/ANN1* denotes the complemented *ann1* mutant. Values were determined under control conditions and at maximal response to the generation of OH[•] by 1 mM Cu-Asc addition. Current recorded at a membrane potential of -200 mV (inside negative) was inward and denoted by a negative value. That at +200 mV was outward and denoted by a positive value.

	-200 mV (Ca _{in})		200 mV (K _{out})	
Epidermis	Control	OH	Control	OH
WT (Col-0)	-181 ± 56	-487 ± 123	280 ± 102	829 ± 156
ann1	-142 ± 58	-160 ± 43	263 ± 108	249 ± 106
ann1/ANN1	-231 ± 78	-569 ± 133	298 ± 128	847 ± 148
WT (WS)	-191 ± 88	-562 ± 94	246 ± 122	840 ± 164
gork	-222 ± 99	-527 ± 128	265 ± 84	811 ± 144
Root hair apex	Control	OH•	Control	OH
WT (Col-0)	-53 ± 38	-280 ± 73	185 ± 68	511 ± 106
ann1	-17 ± 4	-10 ± 3	231 ± 66	249 ± 56