

# Supramolecular hydrogels formed by the conjugates of nucleobases,

## Arg-Gly-Asp (RGD) peptides, and glycosamine

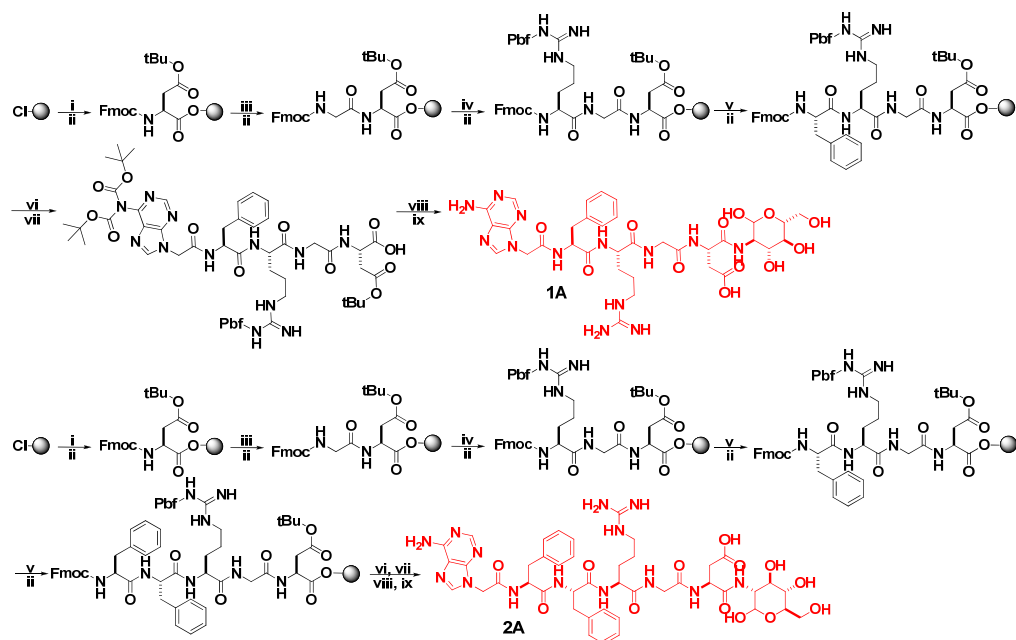
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### Supporting information

#### 1) Materials and methods

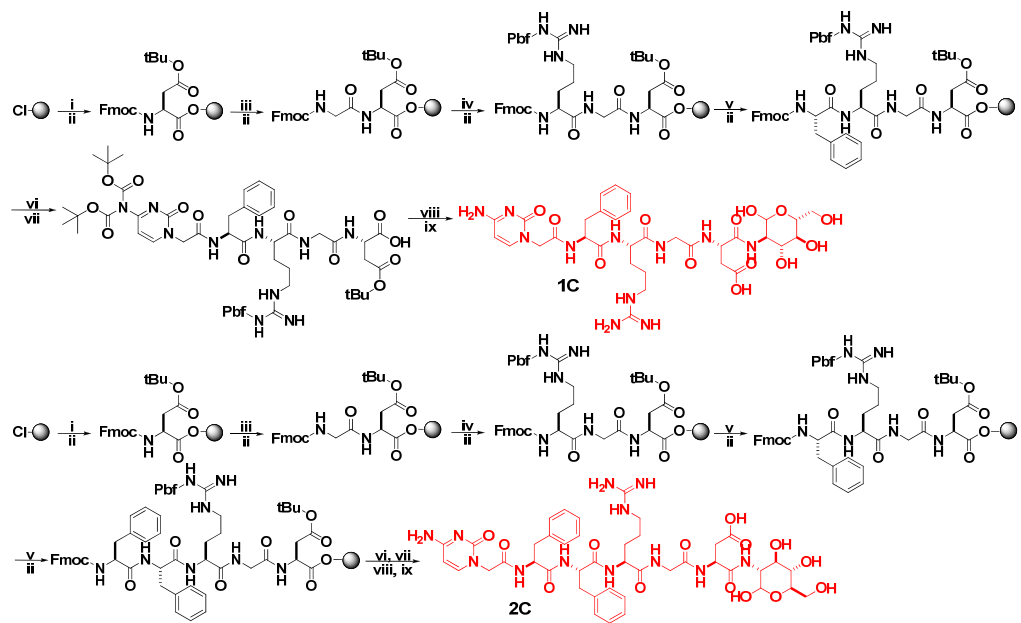
Chemical reagents and solvents were used as received from commercial sources unless otherwise stated.  $^1\text{H}$  and  $^{13}\text{C}$  spectra were obtained on a Varian Unity Inova 400 spectrometer, CD on a JASCO J-810 spectrometer, LC-MS on a Waters Acquity ultra Performance LC with Waters MICROMASS detector, and TEM on a Morgagni 268 transmission electron microscope.

#### 2) Synthesis of hydrogelators of 1A, 2A, 1C, 2C, 1G and 2G.



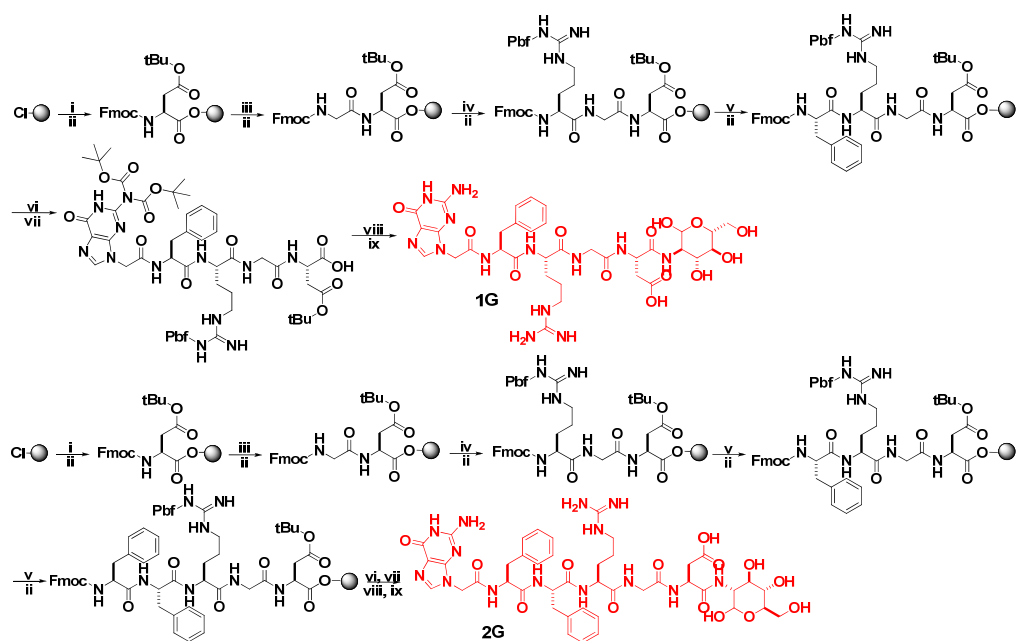
i) L-Asp(OtBu), DIEA; ii) 20 % piperidine; iii) L-Gly, HBTU, DIEA; iv) L-Arg(Pbf), HBTU, DIEA; v) L-Phe, HBTU, DIEA; vi) Bis-boc adenine acetic acid, HBTU, DIEA; vii) 20 % TFE in DCM; viii) D-glucosamine, HBTU, DIEA; ix) TFA:TIS:water (95:2.5:2.5)

**Figure S1.** Molecular structures and the typical synthetic routes of hydrogelators 1A and 2A.



i) L-Asp(OtBu), DIEA; ii) 20 % piperidine; iii) L-Gly, HBTU, DIEA; iv) L-Arg(Pbf), HBTU, DIEA; v) L-Phe, HBTU, DIEA; vi) Bis-boc cytosine acetic acid, HBTU, DIEA; vii) 20 % TFE in DCM; viii) D-glucosamine, HBTU, DIEA; ix) TFA:TIS:water (95:2.5:2.5)

**Figure S2.** Molecular structures and the typical synthetic routes of hydrogelators **1C** and **2C**.

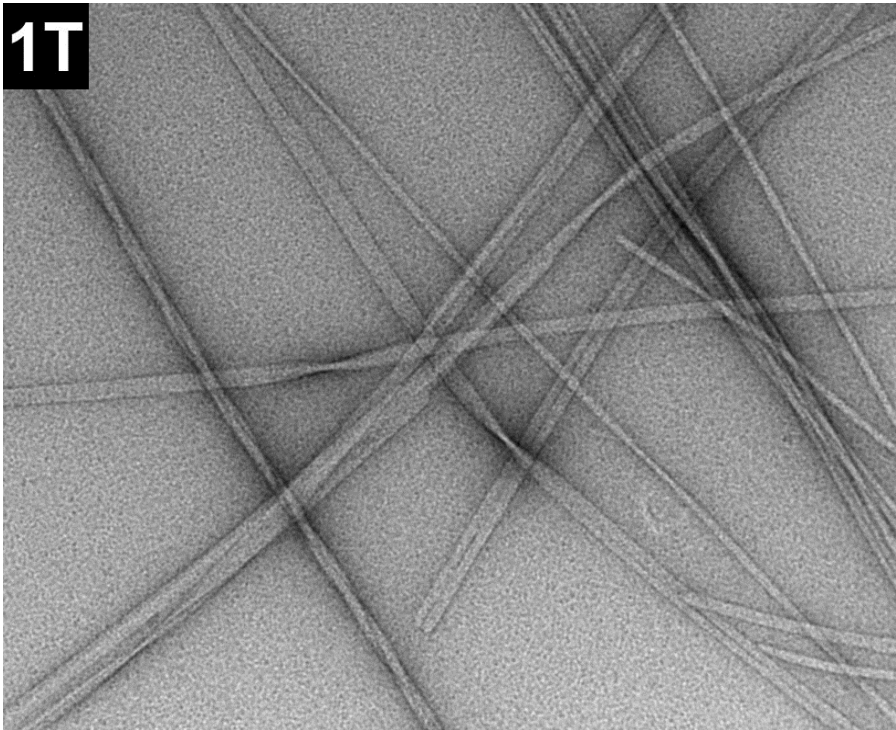


i) L-Asp(OtBu), DIEA; ii) 20 % piperidine; iii) L-Gly, HBTU, DIEA; iv) L-Arg(Pbf), HBTU, DIEA; v) L-Phe, HBTU, DIEA; vi) Bis-boc guanine acetic acid, HBTU, DIEA; vii) 20 % TFE in DCM; viii) D-glucosamine, HBTU, DIEA; ix) TFA:TIS:water (95:2.5:2.5)

**Figure S3.** Molecular structures and the typical synthetic routes of hydrogelators **1G** and **2G**.

**3) Transmission electron micrograph (TEM) of hydrogels of 1T, 1A, 1C, 1G, 2A and 2C, and solution of 2G.**

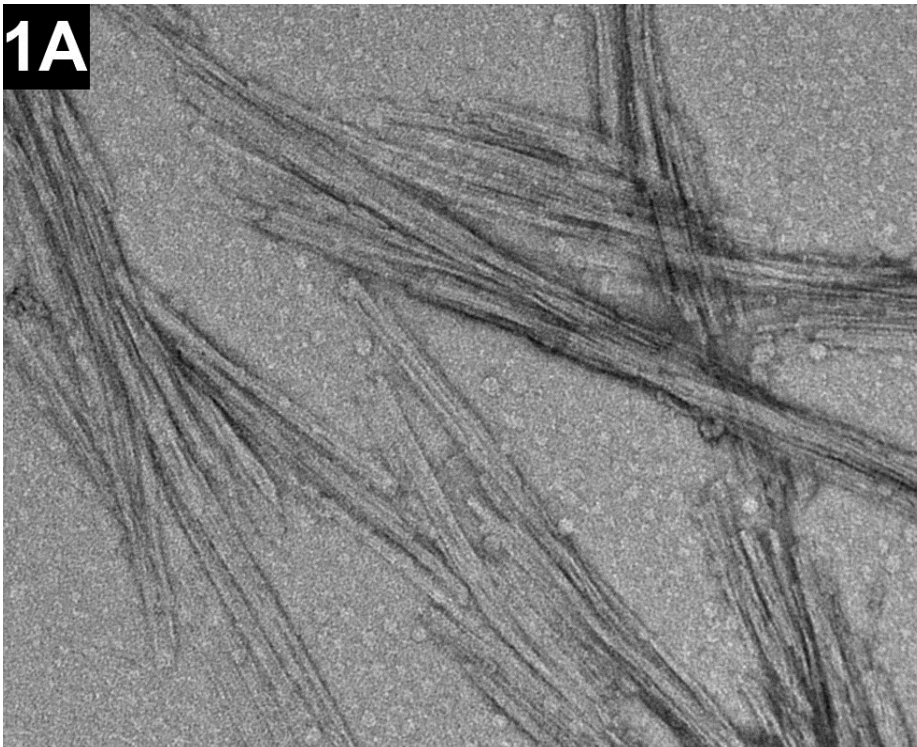
**1T**



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Thy-FRGD-glucos  
Cal: 1.026pix/nm

100 nm  
HV=100kV  
Direct Mag: 18000x  
AMT Camera System

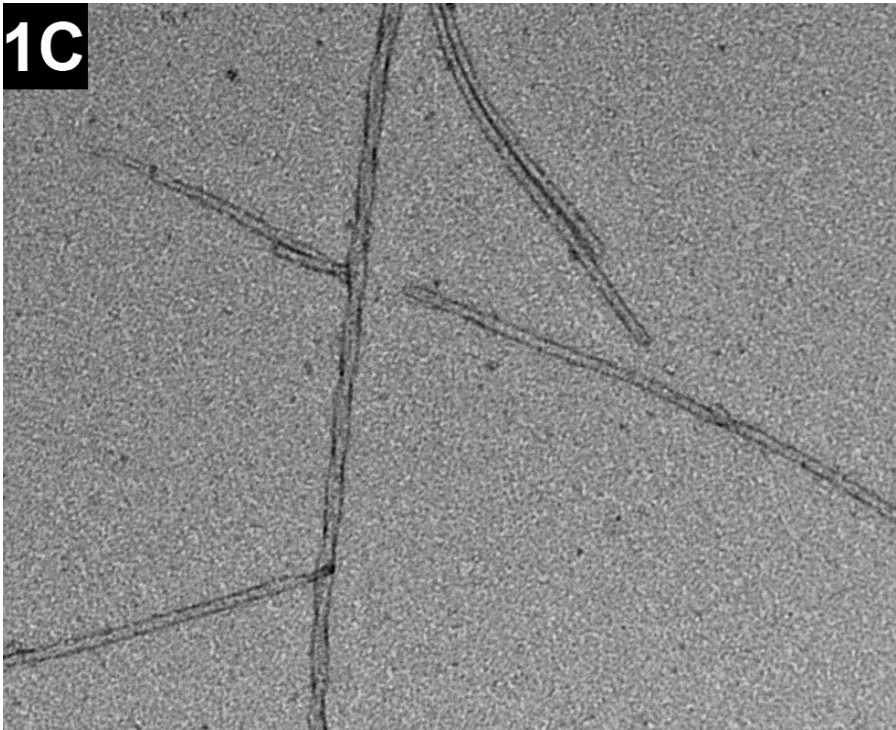
**1A**



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Ade-FRGD-glucosamine  
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20:13 12/14/11

100 nm  
HV=80kV  
Direct Mag: 28000x  
AMT Camera System

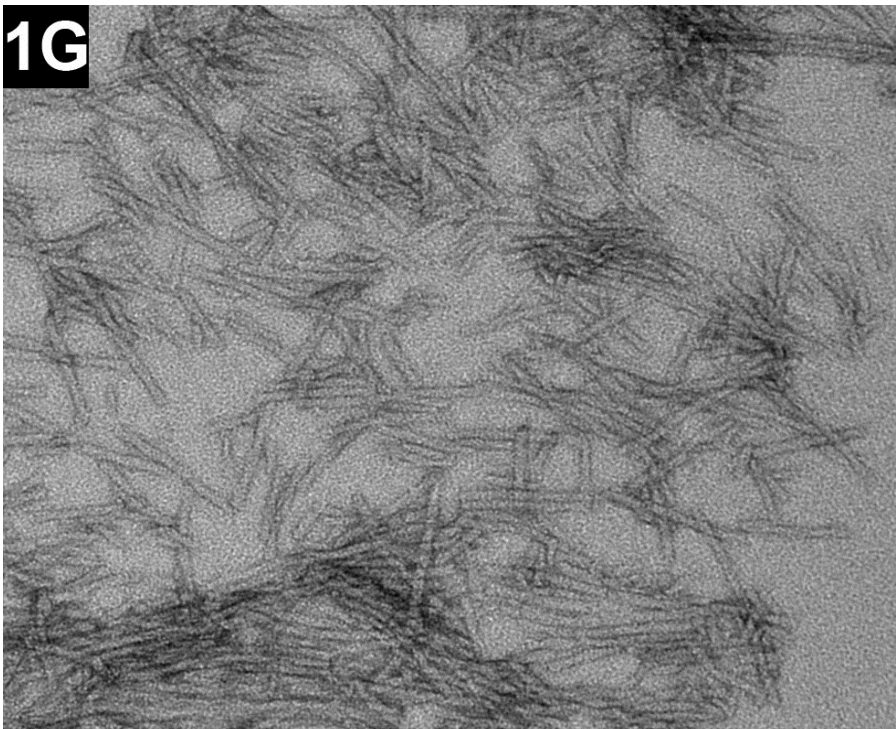
**1C**



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Cyt-FRGD-glucosamine  
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20:44 12/14/11

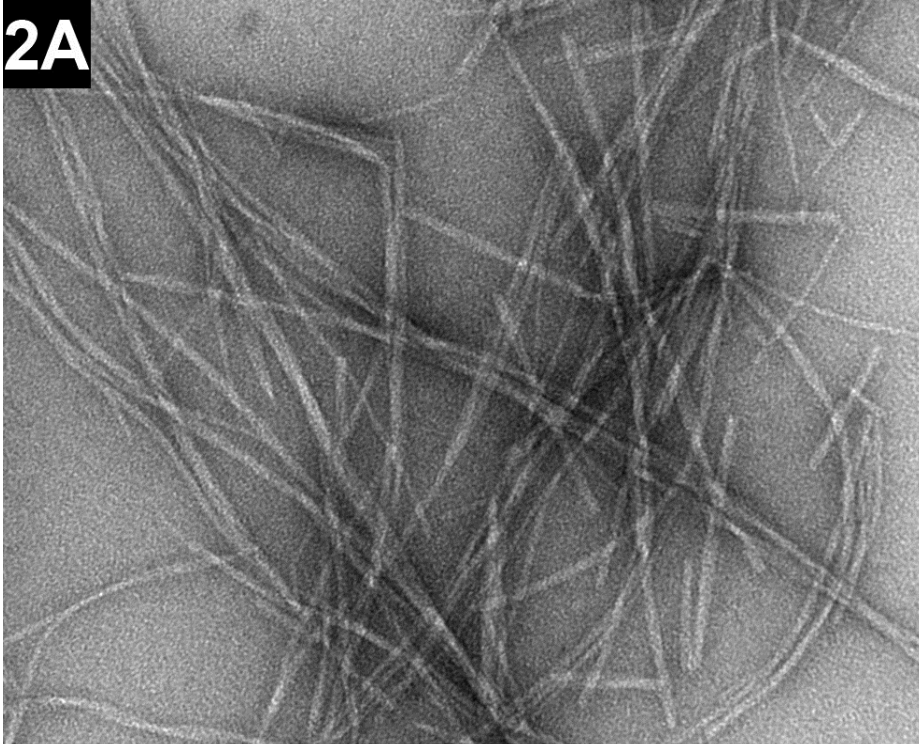
100 nm  
HV=80kV  
Direct Mag: 28000x  
AMT Camera System

**1G**



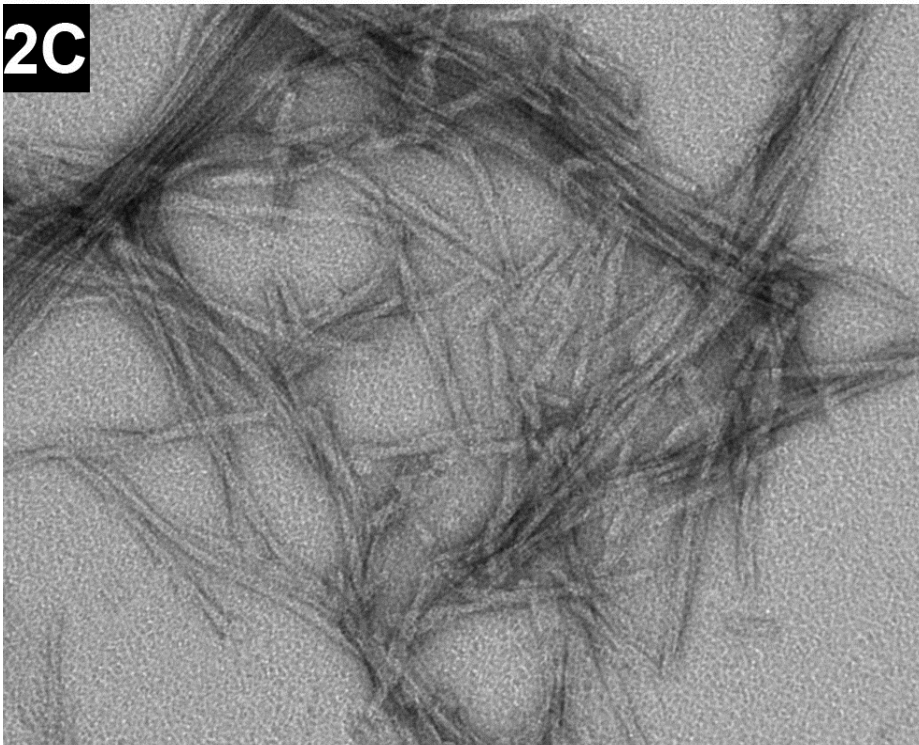
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G-FRGD-Glucosamine  
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21:42 12/28/11

100 nm  
HV=80kV  
Direct Mag: 28000x  
AMT Camera System



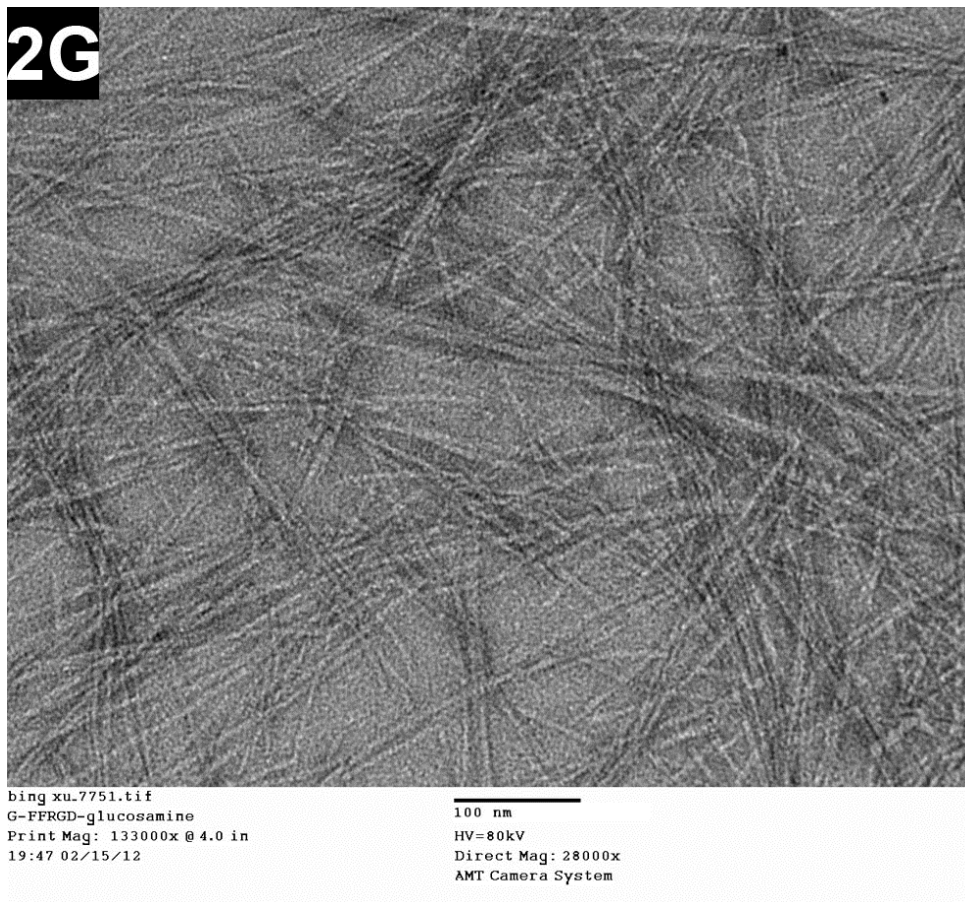
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Ade-FFRGD-glucosamine  
Print Mag: 133000x @ 4.0 in  
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100 nm  
HV=80kV  
Direct Mag: 28000x  
AMT Camera System



bing xu.7432.tif  
CytoFFRGD-Glucosamine  
Print Mag: 133000x @ 4.0 in  
21:03 01/16/12

100 nm  
HV=80kV  
Direct Mag: 28000x  
AMT Camera System



**Figure S4.** High magnification TEM images of hydrogels 1T, 1A, 1C, 1G, 2A and 2C, and solution of 2G.

#### 4) Rheological measurement

Rheological tests were conducted on a TA ARES G2 rheometer (with TA Orchestrator Software). 25 mm cone-plates were used during the experiment. 0.3 mL of hydrogel sample was placed on the cone-plate.

##### i) Dynamic Strain Sweep Test

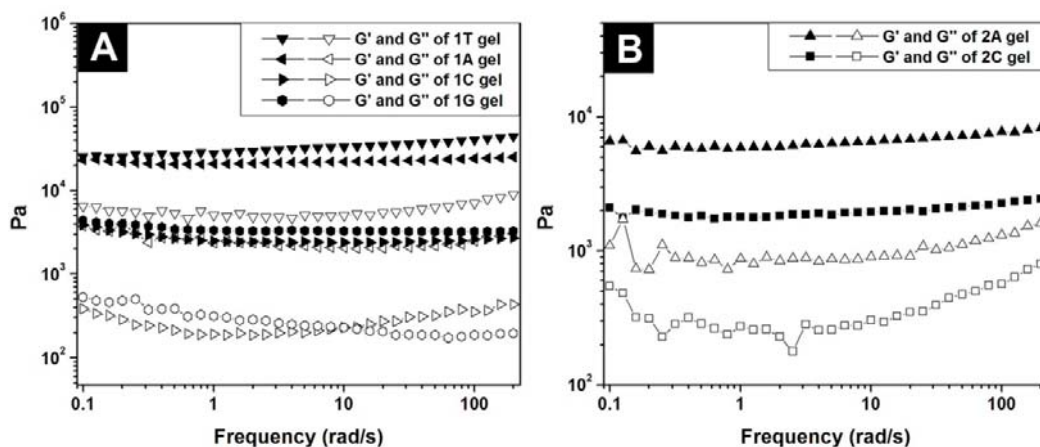
Test range (0.1 to 10 % strain, frequency =10  $\text{rads}^{-1}$ ), 10 points per decade. Sweep mode is “log” and temperature was carried out at 25 °C.

##### ii) Critical strain determination

The critical strain ( $\gamma_c$ ) value was determined from the storage-strain profiles of the hydrogel sample. The strain applied to the hydrogel sample increased from 0.1 to 10 % (10 rad/s and 25 °C). Over a certain strain, a drop in the elastic modulus was observed, and the strain amplitude at the onset of decrease to 5 % decrease from its maximum value was determined and taken as a measure of the critical strain of the hydrogels, which correspond to the breakdown of the cross-linked network in the hydrogel sample.

i) Dynamic Frequency Sweep Test

Test range (0.1 to 200 rad/s, strain = 0.4%), 10 points per decade. Sweep mode is “log” and was carried at 25 °C.

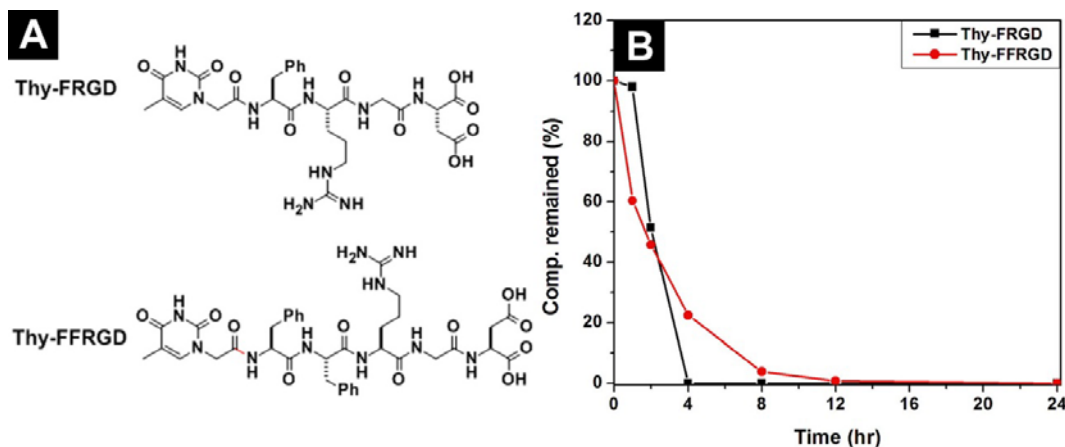


**Figure S5.** Frequency dependence of dynamic storage moduli ( $G'$ ) and loss moduli ( $G''$ ) of (A) the hydrogels of 1T, 1C, 1A, and 1G, (B) the hydrogels of 2A, and 2C shown in Fig. 1.

**5) Biostability test with proteinase K**

1 mg of each compound was dissolved in 5 mL of HEPES buffer at pH 7.5. Then 3.2 units/mL of proteinase K were added and incubated at 37 °C for 24 hr, then 100  $\mu$ L of sample were taken out at 2, 4, 8, 12, and 24h and analyzed by HPLC.

For the control experiment, 1 mg of Thymine-FRGD and 1 mg of thymine-FFRGD (nucleopeptides without glucosamine in conjugation) were dissolved in 5 mL of HEPES buffer at pH 7.5 respectively. Then 3.2 units/mL of proteinase K were added and incubated at 37 °C for 24 hr, then 100  $\mu$ L of sample were taken out each time and analyzed by HPLC.



**Figure S6.** (A) The molecular structures of Thymine-FRGD and thymine-FFRGD, and (B) their time-dependent course of the digestions by proteinase K as control experiment, in which

Thymine-FRGD and thymine-FFRGD are the nucleopeptides without D-glucosamine in conjugation.