

1 Supplementary Table S1: *Mycobacterium tuberculosis complex* isolates used in this study

| Species | Strain | Country of Isolation | Origin ^a | Remark |
|---|----------|----------------------|-----------------------|------------------------------|
| <i>M. tuberculosis</i> | 22 | Mongolia | RIVM | Beijing lineage |
| <i>M. tuberculosis</i> | 53 | Argentina | RIVM | Haarlem lineage |
| <i>M. tuberculosis</i> | 112 | The Netherlands | RIVM | CAS lineage |
| <i>M. tuberculosis</i> | 67 | Comoro Islands | RIVM | EAI lineage |
| <i>M. tuberculosis</i> | 41 | Chile | RIVM | LAM lineage |
| <i>M. tuberculosis</i> | 103 | China | RIVM | T-family lineage |
| <i>M. tuberculosis</i> | 12594_02 | Former Soviet Union | Borstel | Beijing lineage |
| <i>M. tuberculosis</i> | 1500_03 | Former Soviet Union | Borstel | Beijing lineage |
| <i>M. tuberculosis</i> | 1934_03 | Former Soviet Union | Borstel | Beijing lineage |
| <i>M. tuberculosis</i> | 1428_02 | Ghana | Borstel | Cameroon lineage |
| <i>M. tuberculosis</i> | 5390_02 | Ghana | Borstel | Cameroon lineage |
| <i>M. tuberculosis</i> | 5400_02 | Ghana | Borstel | Cameroon lineage |
| <i>M. tuberculosis</i> | 2637_02 | Germany | Borstel | Delhi/CAS lineage |
| <i>M. tuberculosis</i> | 7936_01 | Germany | Borstel | Delhi/CAS lineage |
| <i>M. tuberculosis</i> | 1797_03 | Germany | Borstel | EAI lineage |
| <i>M. tuberculosis</i> | 4850_03 | Germany | Borstel | EAI lineage |
| <i>M. tuberculosis</i> | 947_01 | Germany | Borstel | EAI lineage |
| <i>M. tuberculosis</i> | 2336_02 | Germany | Borstel | Haarlem lineage |
| <i>M. tuberculosis</i> | 9532_03 | Germany | Borstel | Haarlem lineage |
| <i>M. tuberculosis</i> | 7968_03 | Germany | Borstel | LAM lineage |
| <i>M. tuberculosis</i> | 8885_03 | Germany | Borstel | LAM lineage |
| <i>M. tuberculosis</i> | 946_03 | Germany | Borstel | LAM lineage |
| <i>M. tuberculosis</i> | 2151_03 | Germany | Borstel | S-type lineage |
| <i>M. tuberculosis</i> | 2318_06 | Germany | Borstel | S-type lineage |
| <i>M. tuberculosis</i> | 10469_01 | NA ^c | Borstel | Ghana lineage |
| <i>M. tuberculosis</i> | 10493_01 | NA ^c | Borstel | Ghana lineage |
| <i>M. tuberculosis</i> | 2570_02 | NA ^c | Borstel | Ghana lineage |
| <i>M. tuberculosis</i> | 2201_99 | Uganda | Borstel | Uganda I lineage |
| <i>M. tuberculosis</i> | 2333_99 | Uganda | Borstel | Uganda I lineage |
| <i>M. tuberculosis</i> | 2176_99 | Uganda | Borstel | Uganda II lineage |
| <i>M. tuberculosis</i> | 2191_99 | Uganda | Borstel | Uganda II lineage |
| <i>M. tuberculosis</i> | 4412_04 | Germany | Borstel | X-type lineage |
| <i>M. tuberculosis</i> | 9953_04 | Germany | Borstel | X-type lineage |
| <i>M. tuberculosis</i> | 11313_03 | Germany | Borstel | Tur lineage |
| <i>M. tuberculosis</i> | 1657_03 | Germany | Borstel | Ural lineage |
| <i>M. tuberculosis</i> | 10264_03 | Germany | Borstel | Tur lineage |
| <i>M. tuberculosis</i> | 10529_03 | Germany | Borstel | Tur lineage |
| <i>M. tuberculosis</i> | 8431_03 | Germany | Borstel | Ural lineage |
| <i>M. tuberculosis</i> | 3493_07 | | Borstel | Hamburg lineage |
| <i>M. tuberculosis</i> | 10707_07 | | Borstel | Hamburg lineage |
| <i>M. tuberculosis</i> | 9679_00 | NA ^c | Borstel | Laboratory strain ATCC H37Rv |
| <i>M. tuberculosis</i> (19 clinical) | - | NA ^c | Mario Vaneechoutte | Clinical isolates |

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|---------------------|------------------------|-----------------|--------------|---|
| <i>isolates)</i> | | | | |
| <i>M. canettii</i> | 116 | Somalia | RIVM | Smooth growing strain described by van Soolingen <i>et al.</i> 1997 |
| <i>M. canettii</i> | 1997-1549 | Switzerland | RIVM | Swiss isolate described in Pfyffer <i>et al.</i> 1998 |
| <i>M. canettii</i> | NLA000701671 | Somalia | RIVM | Characterised on the basis of their spoligotype, IS6110 RFLP type and smooth growth as |
| <i>M. canettii</i> | NLA000200937 | Eritrea | RIVM | Characterised on the basis of their spoligotype, IS6110 RFLP type and smooth growth |
| <i>M. canettii</i> | 1996-46 | France | RIVM | Canetti strain |
| <i>M. canettii</i> | 3040_99 | The Netherlands | Borstel | |
| <i>M. canettii</i> | 3151_08 | NA ^c | Borstel | |
| <i>M. canettii</i> | 3041_99 | The Netherlands | Borstel | |
| <i>M. bovis</i> | 117 | Argentina | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. bovis</i> | 126 | Argentina | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. bovis</i> | 73 | The Netherlands | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. bovis</i> | 130 | The Netherlands | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. bovis</i> | 24 | Saudi Arabia | RIVM | Isolated from an oryx, Antelope clade, see also Smith <i>et al.</i> 2006 |
| <i>M. bovis</i> | 4258_00 | Germany | Borstel | |
| <i>M. bovis</i> | 751_01 | Germany | Borstel | |
| <i>M. bovis</i> | 7540_01 | Germany | Borstel | |
| <i>M. bovis</i> | - | NA ^c | Mario | Clinical isolates |
| (6 isolates) | | | Vaneechoutte | |
| <i>M. bovis</i> BCG | 48 (2) | The Netherlands | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. bovis</i> BCG | 71 | Japan | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. bovis</i> BCG | 83 | Russia | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. bovis</i> BCG | 2008-714 ^b | NA ^c | RIVM | Identified on basis of characteristic IS6110/IS1081 RFLP patterns according to van Soolingen <i>et al.</i> 1992 |
| <i>M. bovis</i> BCG | 2008-1601 ^b | NA ^c | RIVM | Identified on basis of characteristic IS6110/IS1081 RFLP patterns according to van Soolingen <i>et al.</i> 1992 |
| <i>M. bovis</i> BCG | DSM 43990 | NA ^c | DSMZ | <i>Mycobacterium bovis</i> Karlson and Lessel 1970 BCG, Chicago 1 |
| <i>M. bovis</i> BCG | DSM 45071 | NA ^c | DSMZ | <i>Mycobacterium bovis</i> Karlson and Lessel 1970 |
| <i>M. caprae</i> | 2006-1960 ^b | The Netherlands | RIVM | Characterised using Hain genotype MTBC kit |
| <i>M. caprae</i> | 2007-0039 ^b | The Netherlands | RIVM | Characterised using Hain genotype MTBC kit |
| <i>M. caprae</i> | 1694_00 | Germany | Borstel | |
| <i>M. caprae</i> | 8986_99 | Germany | Borstel | |
| <i>M. caprae</i> | 9577_99 | Germany | Borstel | |
| <i>M. microti</i> | 62 | United Kingdom | RIVM | see van Soolingen <i>et al.</i> 1998 |
| <i>M. microti</i> | 25 | United Kingdom | RIVM | see van Soolingen <i>et al.</i> 1998 |

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|----------------------|------------------------|-----------------|---------|--------------------------------------|
| <i>M. microti</i> | 15274 ^b | United Kingdom | RIVM | see van Soolingen <i>et al.</i> 1998 |
| <i>M. microti</i> | 15912 ^b | Belgium | RIVM | see van Soolingen <i>et al.</i> 1998 |
| <i>M. microti</i> | 15911 ^b | Netherlands | RIVM | see van Soolingen <i>et al.</i> 1998 |
| <i>M. microti</i> | 417/01 | Germany | | Llama lineage |
| <i>M. pinnipedii</i> | 76 | Argentina | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. pinnipedii</i> | 81 | Argentina | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. pinnipedii</i> | 101 | Argentina | RIVM | See Kremer <i>et al.</i> 2005 |
| <i>M. pinnipedii</i> | 7011_02 | Germany | Borstel | |
| <i>M. pinnipedii</i> | 7739_01 | Germany | Borstel | |
| <i>M. africanum</i> | 6 | The Netherlands | RIVM | <i>M. africanum</i> clade 2 |
| <i>M. africanum</i> | 128 (85) | The Netherlands | RIVM | <i>M. africanum</i> clade 2 |
| <i>M. africanum</i> | 2007-1386 ^b | The Netherlands | RIVM | <i>M. africanum</i> clade 2 |
| <i>M. africanum</i> | 2007-1154 ^b | The Netherlands | RIVM | <i>M. africanum</i> clade 2 |
| <i>M. africanum</i> | 2007-1073 ^b | The Netherlands | RIVM | <i>M. africanum</i> clade 2 |
| <i>M. africanum</i> | 1449_02 | Ghana | Borstel | <i>M. africanum</i> clade 1 |
| <i>M. africanum</i> | 1473_02 | Ghana | Borstel | <i>M. africanum</i> clade 1 |
| <i>M. africanum</i> | 10473_01 | Ghana | Borstel | <i>M. africanum</i> clade 1 |
| <i>M. africanum</i> | 10494_01 | Ghana | Borstel | <i>M. africanum</i> clade 1 |
| <i>M. africanum</i> | 1443_02 | Ghana | Borstel | <i>M. africanum</i> clade 1 |
| <i>M. africanum</i> | 10476_01 | Ghana | Borstel | <i>M. africanum</i> clade 2 |
| <i>M. africanum</i> | 10514_01 | Ghana | Borstel | <i>M. africanum</i> clade 2 |
| <i>M. africanum</i> | 5468_02 | Ghana | Borstel | <i>M. africanum</i> clade 2 |
| <i>M. africanum</i> | 9550_99 | Ghana | Borstel | <i>M. africanum</i> clade 2 ATCC |

^a RIVM = National Tuberculosis Reference Laboratory, National Institute for Public Health and the Environment, Bilthoven, The Netherlands; *Borstel = National Reference Center for Mycobacteria, Forschungszentrum Borstel, Germany; *DSM = The German Collection of Microorganisms

^b Represent RIVM strains not previously described in literature, however have been characterised to the species level using techniques outlined in references supplied in the manuscript.

^c This information was not available (NA) for this study.

Supplementary Table S2: Non *tuberculosis* mycobacteria and other strains of bacteria used in this study

| Non <i>tuberculosis</i> mycobacteria | Strain designation ^a | Remark |
|---|---------------------------------|---|
| <i>Mycobacterium aichiense</i> | DSM 44147 | Type strain, isolated from soil |
| <i>Mycobacterium alvei</i> | DSM 44176 | Type strain, isolated from water sample |
| <i>Mycobacterium arupense</i> | DSM 44942 | Type strain, isolated from a tendon |
| <i>Mycobacterium asiaticum</i> | ITG 8182 | See De Baere <i>et al.</i> 2002 |
| <i>Mycobacterium avium</i> | ITG 7886 | See Vaneechoutte <i>et al.</i> 1993 |
| <i>Mycobacterium boenickei</i> | DSM 44677 | Type strain, isolated from a leg wound |
| <i>Mycobacterium branderi</i> | DSM 44624 | Type strain, isolated from human sputum |
| <i>Mycobacterium brisbanense</i> | DSM 44680 | Type strain, isolated from a sinus |
| <i>Mycobacterium brumae</i> | DSM 44177 | Type strain, isolated from water sample |
| <i>Mycobacterium canariense</i> | DSM 44828 | Type strain, isolated from human blood |
| <i>Mycobacterium celatum</i> | ITG 6147 | See De Baere <i>et al.</i> 2002 |
| <i>Mycobacterium chelonae</i> | ITG 4975 | NA ^b |
| <i>Mycobacterium chelonae</i> subsp. <i>abscessus</i> | DSM 44196 | Type strain |
| <i>Mycobacterium confluentis</i> | DSM 44017 | Type strain, isolated from human sputum |

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| <i>Mycobacterium conspicuum</i> | DSM 44136 | Type strain, isolated from patient with disseminated infection |
| <i>Mycobacterium flavescens</i> | VUB A016 | See De Baere <i>et al.</i> 2002 |
| <i>Mycobacterium fortuitum</i> | ITG 8020 | See Vaneechoutte <i>et al.</i> 1993 |
| <i>Mycobacterium genavense</i> | ITG 97-102 | See De Baere <i>et al.</i> 2002 |
| <i>Mycobacterium gilvum</i> | DSM 9487 | Isolated from soil |
| <i>Mycobacterium goodii</i> | DSM 44492 | Type strain |
| <i>Mycobacterium gordonae</i> | ITG 7704 | See Vaneechoutte <i>et al.</i> 1993 |
| <i>Mycobacterium heckeshornense</i> | DSM 44428 | Type strain, isolated from human respiratory tract |
| <i>Mycobacterium houstonense</i> | DSM 44676 | Type strain, isolated from a facial abscess |
| <i>Mycobacterium intracellulare</i> | DSM 43223 | Type strain |
| <i>Mycobacterium kansasii</i> | ITG 7727 | See Vaneechoutte <i>et al.</i> 1993 |
| <i>Mycobacterium kubicaiae</i> | DSM 44627 | Type strain, isolated from human sputum |
| <i>Mycobacterium lacus</i> | DSM 44577 | Type strain, isolated from human elbow |
| <i>Mycobacterium mageritense</i> | DSM 44476 | Type strain, isolated from human sputum |
| <i>Mycobacterium malmoense</i> | ITG 940611 | See De Baere <i>et al.</i> 2002 |
| <i>Mycobacterium marinum</i> | ITG 1727 | See Vaneechoutte <i>et al.</i> 1993 |
| <i>Mycobacterium massiliense</i> | DSM 45103 | Type strain, isolated from human blood |
| <i>Mycobacterium moriokaense</i> | DSM 44221 | Type strain, isolated from soil |
| <i>Mycobacterium mucogenicum</i> | DSM 44625 | Type strain, isolated from human cyst |
| <i>Mycobacterium nebraskense</i> | DSM 44803 | Type strain, isolated from human sputum |
| <i>Mycobacterium neworleansense</i> | DSM 44679 | Type strain, isolated from human scalp |
| <i>Mycobacterium paratuberculosis</i> | ITG 2666 | See De Baere <i>et al.</i> 2002 |
| <i>Mycobacterium scrofulaceum</i> | DSM 43992 | Type strain, isolated from human cervical lymph node |
| <i>Mycobacterium shimoidei</i> | DSM 44152 | Type strain, isolated from sputum of patient with tuberculosis-like disease |
| <i>Mycobacterium simiae</i> | ITG 4485 | See Vaneechoutte <i>et al.</i> 1993 |
| <i>Mycobacterium smegmatis</i> | DSM 43756 | Type strain |
| <i>Mycobacterium szulgai</i> | ITG 4979 | NA ^b |
| <i>Mycobacterium tusciae</i> | DSM 44338 | Type strain, isolated from human cervical lymph node |
| <i>Mycobacterium ulcerans</i> | ITG 96-1439 | NA ^b |
| <i>Mycobacterium xenopi</i> | ITG 4986 | See De Baere <i>et al.</i> 2002 |
| Other bacteria | Strain designation | Remark |
| <i>Staphylococcus aureus</i> | DSM 20231 | Type strain, isolated from human pleural fluid |
| <i>Listeria monocytogenes</i> | DSM 20600 | Type strain, isolated from a rabbit |
| <i>Escherichia coli</i> | DSM 301 | Disinfectant test strain |
| <i>Klebsiella oxytoca</i> | ATCC 43086 | NA ^b |
| <i>Enterococcus faecalis</i> | DSM 20371 | Isolated from pleural fluid |
| <i>Proteus mirabilis</i> | DSM 4479 | Type strain |

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| <i>Bacillus cereus</i> | DSM 31 | Type strain |
| <i>Bordetella pertussis</i> | CCUG 13475 | Isolated from patient suffering from whooping cough |
| <i>Streptococcus agalactiae</i> | DSM 2134 | Type strain |
| <i>Rhodococcus equi</i> | DSM 20307 | Type strain, isolated from lung abscess of foal |
| <i>Streptomyces albidoflavus</i> | DSM 40455 | Type strain |
| <i>Nocardioides sp.</i> | DSM 17401 | Proposed type strain, isolated from marine sediment |
| <i>Nocardia salmonicida</i> | DSM 40472 | Type strain, isolated from blueback salmon |
| <i>Nocardia asiatica</i> | clinical isolate | Isolated from human wound, see Wauters <i>et al.</i> 2005 |
| <i>Nocardia nova</i> | clinical isolate | Isolated from human abscess, see Wauters <i>et al.</i> 2005 |
| <i>Nocardia cyriacigeorgica</i> | clinical isolate | Isolated from human bronchial aspirate, see Wauters <i>et al.</i> 2005 |
| <i>Nocardia farcinica</i> | clinical isolate | Isolated from human abscess, see Wauters <i>et al.</i> 2005 |

^a DSM = The German Collection of Microorganisms; *ITG = Institute of Tropical Medicine, Antwerp; *VUB = Department of Microbiology, Academic Hospital of the Free University of Brussels, Brussels, Belgium Germany; *ATCC = American Type Culture Collection; *CCUG = Culture Collection, University of Göteborg, Sweden.

^bThis information was not available (NA) for this study.

Supplementary Table S3: Oligonucleotide primers and probes used in this study

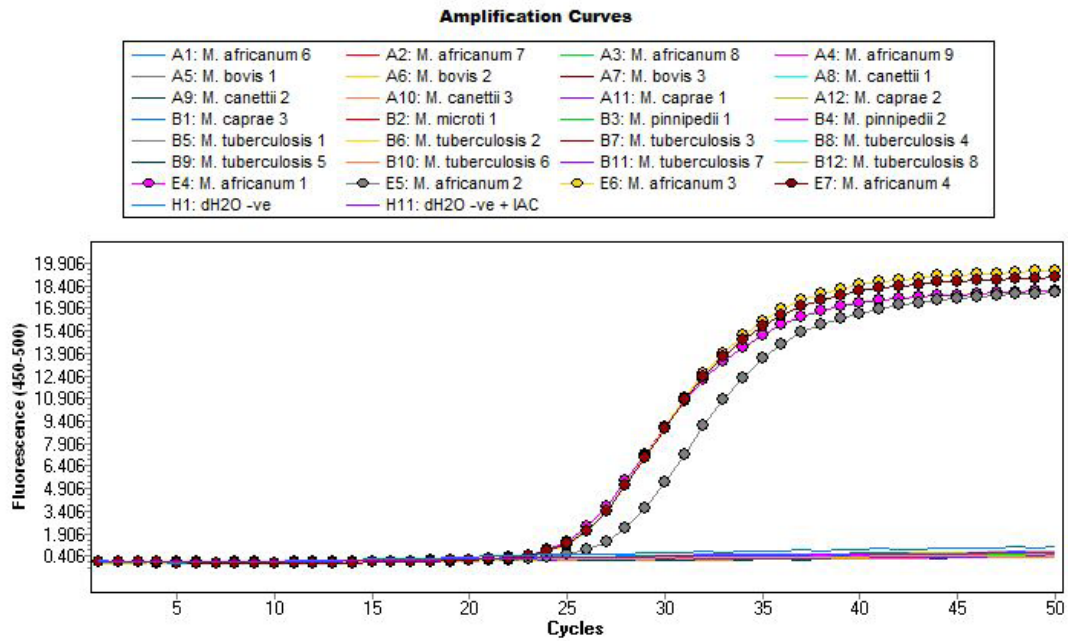
| Name | Function | Sequence 5'→3' |
|-----------------------------|---|-------------------------------------|
| MTC_Fw | Forward Sequencing primer, forward MTC real-time PCR assay primer | AGACCGTGCGGATCTTG |
| MTC_Rv | Reverse Sequencing primer, Reverse MTC real-time PCR assay primer | CATGGAGATCACCCGTGA |
| MTC Probe | MTC probe | HEX-ATTGGTCACCCGGATTTCCGGT-BHQ1 |
| <i>wbb11</i> _Fw | Forward sequencing primer, Forward real-time PCR assay primer | TACCAGCTTCAGTTCCGT |
| <i>wbb11</i> _Rv | Reverse sequencing primer, Reverse real-time PCR assay primer | GCACCTATATCTTCTTAGCCG |
| <i>wbb11</i> probe | <i>wbb11</i> probe | FAM-ATGGTGCGCAGTTCCTGC |
| <i>M. canetti</i> sp Fw | Forward <i>M. canetti</i> specific primer | ATGTGGTTTCAGTACGACTTC |
| <i>M. canetti</i> sp Rv | Reverse <i>M. canetti</i> specific primer | GATGGCAGTGTCTTATCCAA |
| <i>M. canetti</i> sp probe | <i>M. canetti</i> specific probe | ROX-TGAGAGGTGTTGGCACGCAA-BHQ2 |
| <i>lpqT</i> Fw | Forward sequencing primer, Forward real-time PCR assay primer | ACGAATCCGGCGATGATC |
| <i>lpqT</i> _Rv | Reverse sequencing primer, Reverse real-time PCR assay primer | CGACTGCACACCTGGAA |
| <i>lpqT</i> probe | <i>lpqT</i> Probe | FAM-TTGGCCGGCGCCGGTT-BHQ1 |
| RD1_Fw | Forward sequencing primer, Forward real-time PCR assay primer | CATCGCTGATGTGCTTGC |
| RD1_Rv | Reverse sequencing primer, Reverse real-time PCR assay primer | TGCGCCGAGCTGTATTC |
| RD1_probe | RD1 Probe | ROX-ACACTAGCGTCAATGCGGTCA-BHQ2 |
| <i>M. caprae lepA</i> _Fw | Forward sequencing primer, Forward real-time PCR assay primer | AGACCGTGCGGATCTTG |
| <i>M. caprae lepA</i> _Rv | Reverse sequencing primer, Reverse real-time PCR assay primer | CATGGAGATCACCCGTGA |
| <i>M. caprae lepA</i> probe | <i>M. caprae lepA</i> Probe | Cyan 500-TATCGGGTACACAAAGACGA – BBQ |
| RD713_Fw S | RD713 Forward sequencing primer | CCATCTGCGCTTTCGGTGCTTCT |
| RD713_RvS | RD 713 Reverse sequencing primer | CTGCCAGTCGCTTCCCCATTGTG |

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|----------------------|---|-----------------------------------|
| RD713_Fw | Forward real-time PCR assay primer | ACGGAACGGTCAAGAAC |
| RD713_Rv | Reverse real-time PCR assay primer | GCTCAAGAATCGTCGCTA |
| RD713 probe | RD 713 Probe | Cyan 500-ACGTCCTTGTGACCGCGAC- BBQ |
| RD701_FwS | RD 701 Forward sequencing primer | CTGTGCAGGTGGTCGTTTC |
| RD701_RvS | RD 701 Reverse sequencing primer | CGAATTGCTCATCCCGTAAC |
| RD701_Fw | Forward real-time PCR assay primer | AACGGGTCCGATTCTCC |
| RD701_Rv | Reverse real-time PCR assay primer | CCGAAACCCTCGTTGATC |
| RD701 probe | RD 701 Probe | ROX-TCAGCCGCCGGCCAACC-BHQ2 |
| IAC MSMEG_0660_Fw | Forward sequencing primer, Forward real-time PCR assay primer | TCACCGACCATGTCCAG |
| IAC MSMEG_0660_Rv | Reverse sequencing primer, Reverse real-time PCR assay primer | CGTTGCCCAATCCGTATG |
| IAC MSMEG_0660 probe | IAC MSMEG_0660 probe | Cy5-CAGCAGTACCATCGCCATCG-BHQ2 |
| <i>ssrA</i> _Fw | <i>ssrA</i> Forward sequencing primer | TGGGGCTGAAAGGTTTCGA |
| <i>ssrA</i> _Rv | <i>ssrA</i> Reverse sequencing primer | TGGAGCTGCCGGAAT |

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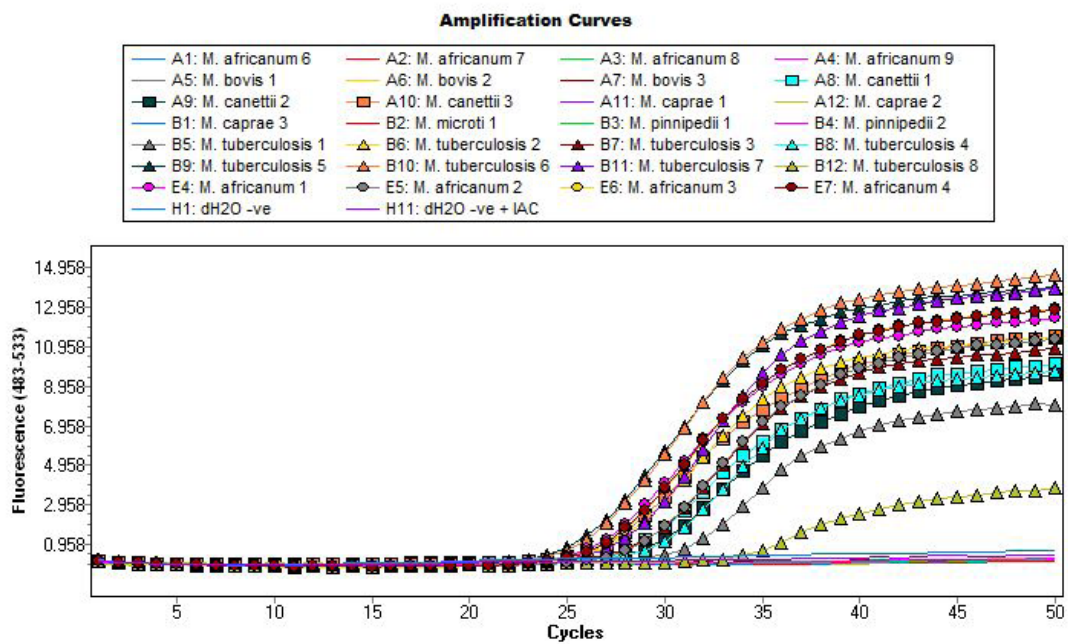
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Fig. S1A



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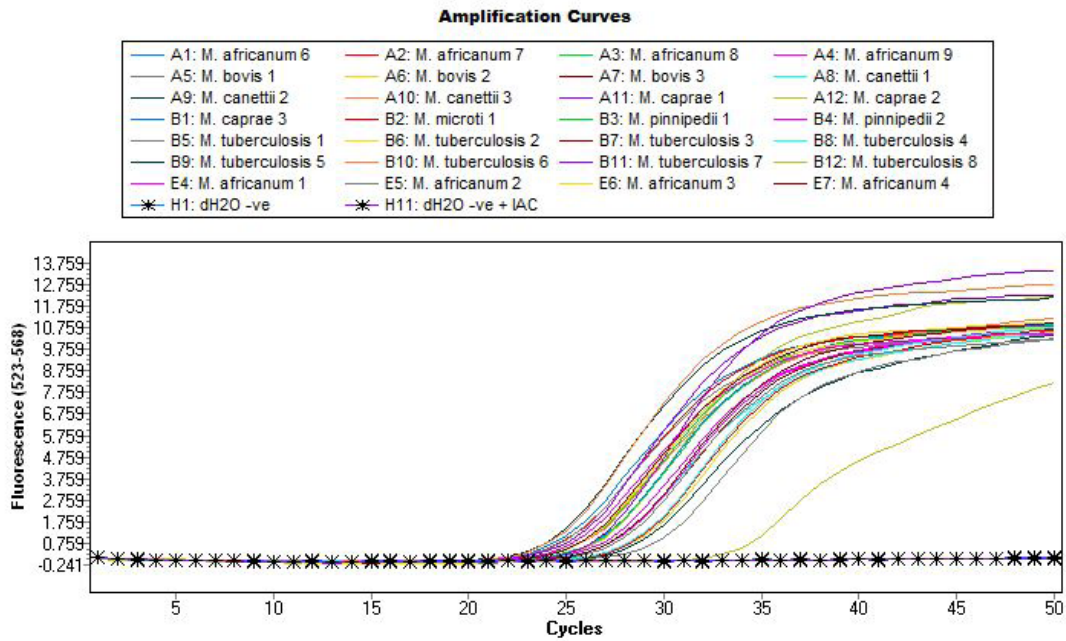
Fig. S1B



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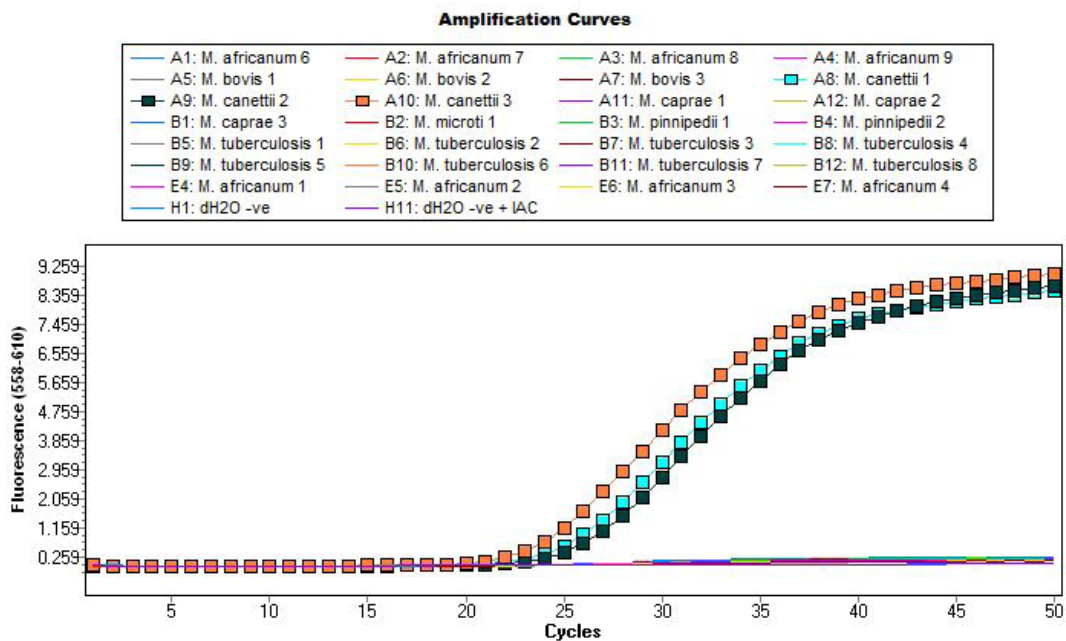
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Fig. S1C



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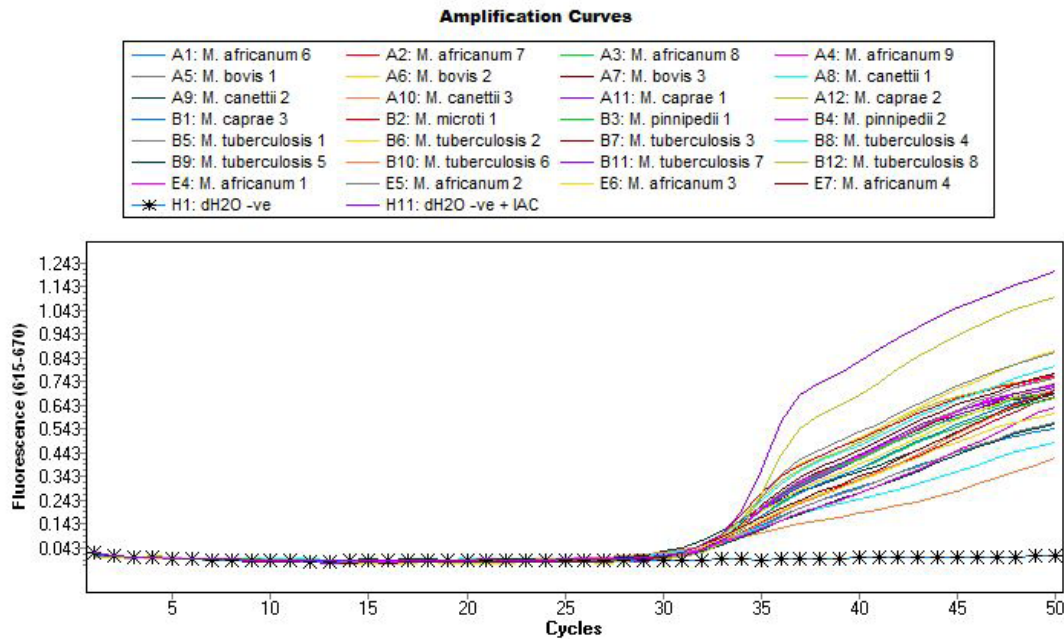
Fig. S1D



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Fig. S1E



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Supplementary Figure Legend

97 **Fig. S1**

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99 **Amplification curves for multiplex 1 real-time PCR assay**

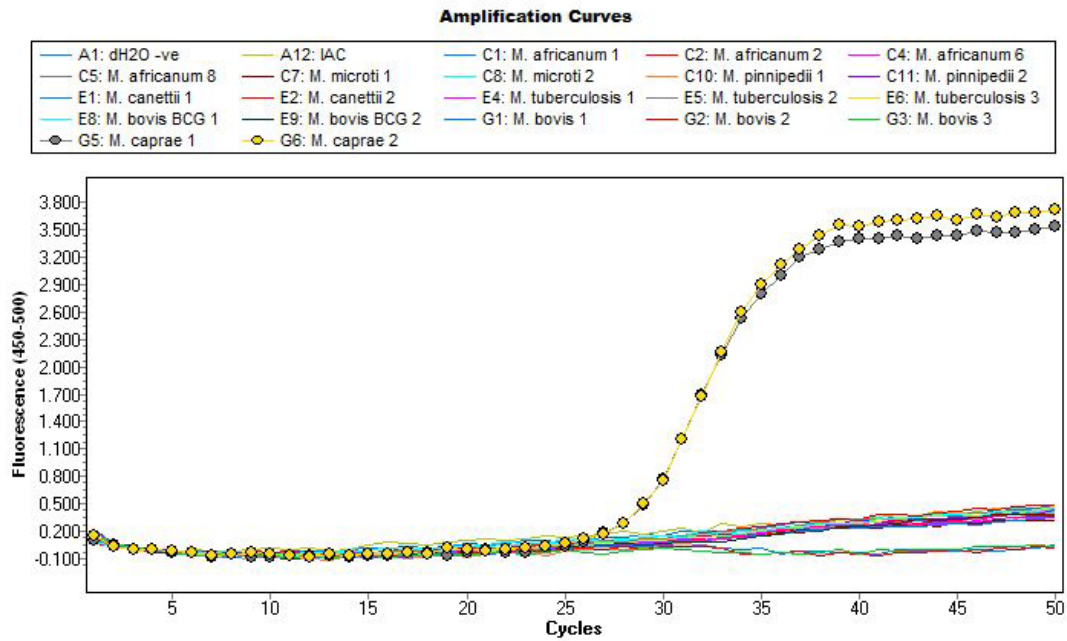
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101 Figure S1A Real-time amplification curves for *M. africanum* West African 1 (circle) using
102 RD713 in Cyan500 channel (450-500). Figure S1B Amplification curves for *M. africanum*
103 West African 1 (circle), *M. tuberculosis* (triangles) and *M. canettii* (rectangles) using the
104 *wbb11* gene in FAM channel (483-533). Figure S1C Amplification curves for all members of
105 the MTC using the *lepA* gene, with the no template control highlighted (stars) in the HEX
106 channel (523-568). Figure S1D Amplification curves of *M. canettii* (rectangles) in the ROX
107 channel (558-610) using RD^{canettii1}. Figure S1E Amplification curves for IAC in Cy5 channel
108 (615-670) with the no template control highlighted with stars through amplification curve.

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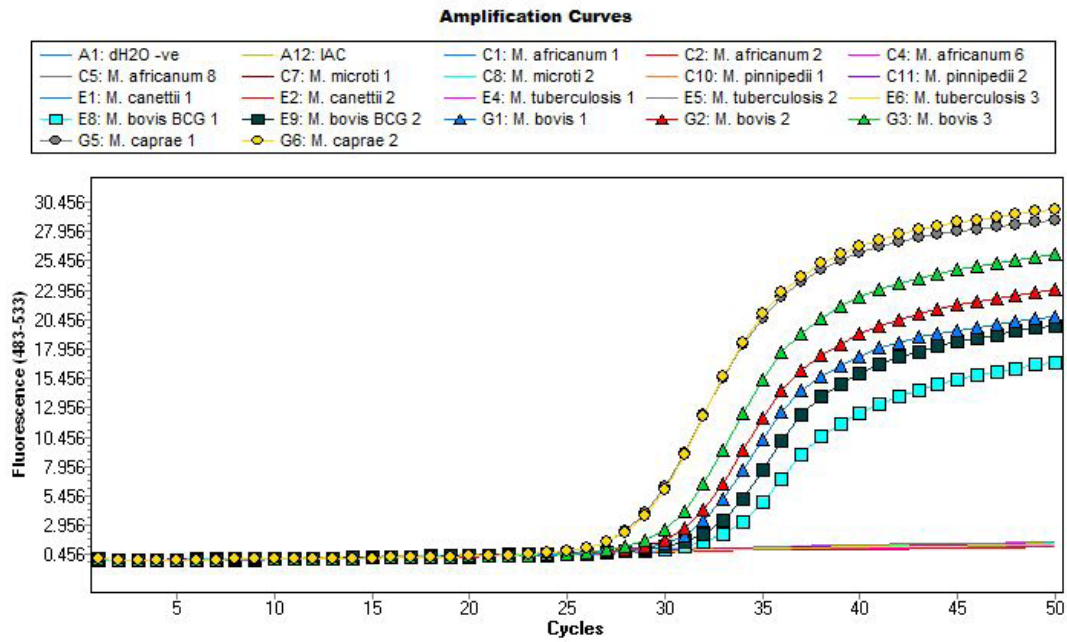
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Fig. S2A



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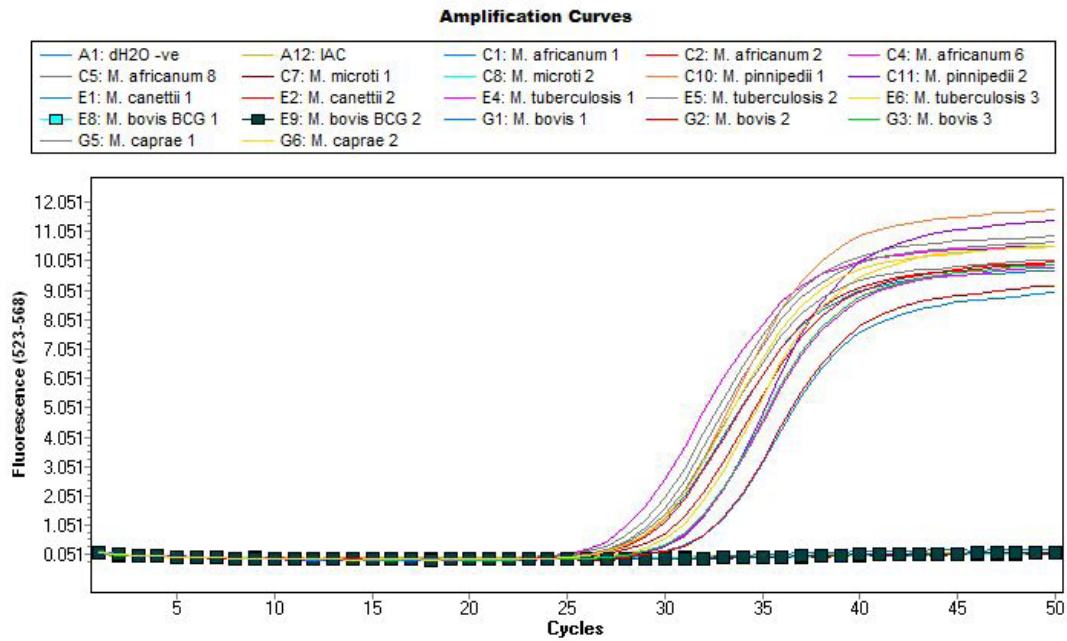
Fig. S2B



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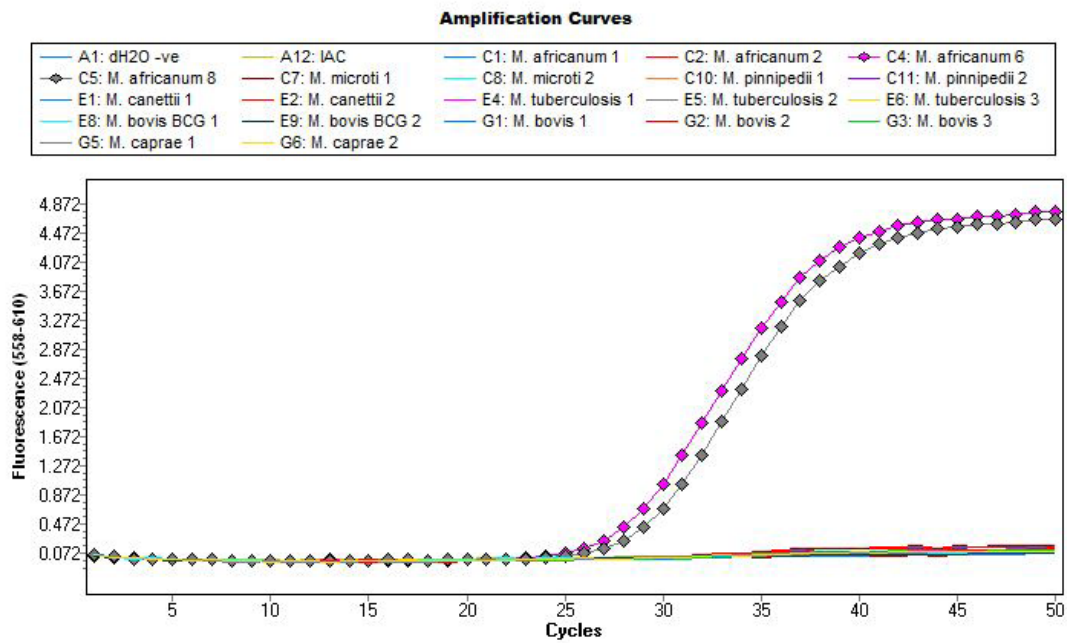
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Fig. S2C



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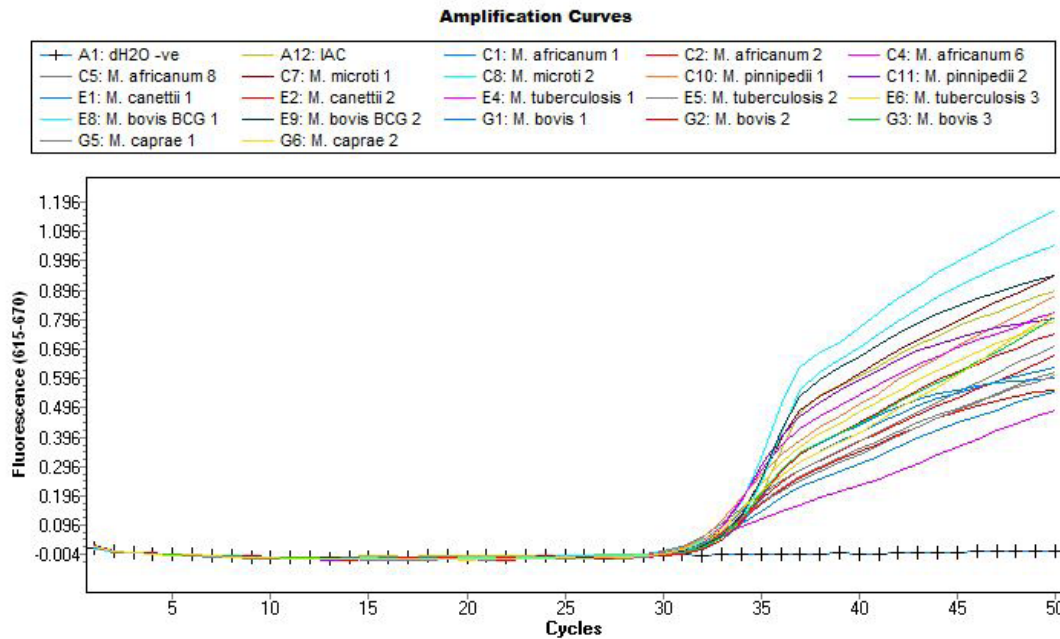
Fig. S2D



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Fig. S2 E



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Fig. S2

Amplification curves for multiplex 2 real-time PCR assay

160 Figure S2A Real-time amplification curves for *M. caprae* (circle) using *lepA* gene in
161 Cyan500 channel (450-500). Figure S2B Amplification curves for *M. caprae* (circle), *M.*
162 *bovis* (triangles) and *M. bovis BCG* (rectangles) using the *lpqT* gene in FAM channel (483-
163 533). Figure S2C Amplification curves for all members of the MTC with the exception of *M.*
164 *bovis BCG* (diamonds) and *M. microti* in the HEX channel (523-568). Figure S2D
165 Amplification curves of *M. africanum* West African 2 (diamonds) in the ROX channel (558-
166 610) using RD701. Figure S2E Amplification curves for IAC in Cy5 channel (615-670) with
167 the no template control highlighted with stars through amplification curve.

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