

FIGURE S1: $\text{NO}_2^-/\text{NO}_3^-$ concentration and $\log(\text{NO}_2^-/\text{NO}_3^- \text{ concentration})$ in an enrichment culture over time. The growth rate of the culture is calculated as the linear increase of the log transformed the $\text{NO}_2^-/\text{NO}_3^-$ concentration over time. The lag phase was determined as the time before the culture started to grow logarithmic.

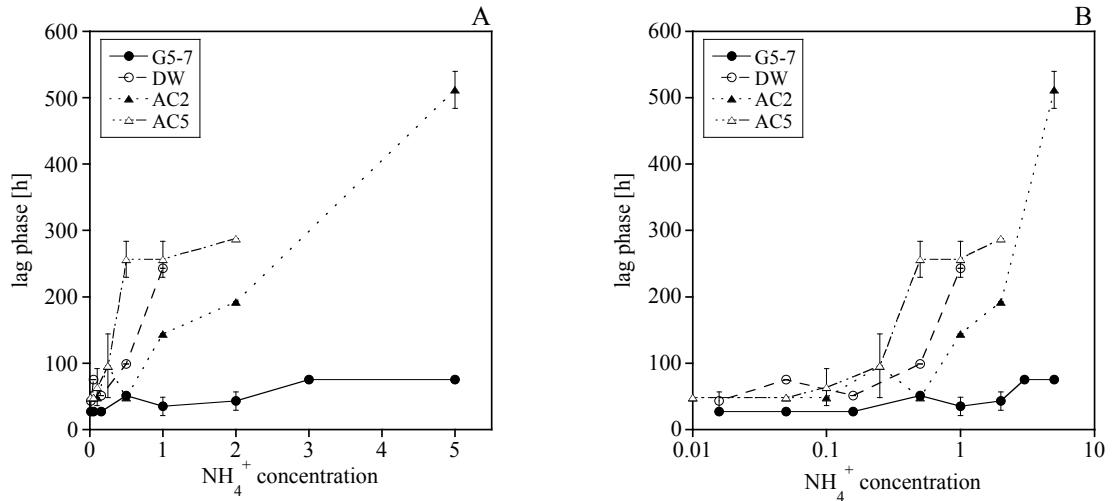


FIGURE S2: Influence of NH_4^+ concentration on the lag phase before onset of logarithmic growth in the enrichment cultures AOA-AC2; AOA-AC5, AOA-DW, and AOB-G5-7 (mean \pm SD; n=3). A: NH_4^+ concentration linear scale; B: NH_4^+ concentration logarithmic scale.

TABLE S1: Primers and PCR conditions used in this study.

	Primer	Anneal. temp	# cycles
AOA-amoA (5)	Arch amoA F: 5'-STA ATG GTC TGG CTT AGA CG-3' Arch amoA R: 5'-GCG GCC ATC CAT CTG TAT GT-3'	53	35
Archaeal 16S rRNA (4, 6)	Arch 109F: 5'-ACK GCT CAG TAA CAC GT-3' Arch 915R: 5'-YCC GGC GTT GAM TCC AAT T-3'	46	30
AOB-amoA (10)	amoA-1F: 5'-GGG GTT TCT ACT GGT GGT-3' amoA-2R KS: 5'-CCC CTC KGS AAA GCC TTC TTC-3'	55	35
AOB-16S rRNA (7)	CTO 189F-A: 5'-GGA GAA AAG CAG GGG ATC G-3' CTO 189F-B: 5'-GGA GGA AAG CAG GGG ATC G-3' CTO 189F-C: 5'-GGA GGA AAG TAG GGG ATC G-3' CTO 654R: 5'-CTA GCY TTG TAG TTT CAA ACG C-3'	57.5	35
M13-cloning	M13-F: 5'-GTA AAA CGA CGG CCA G-3' M13-R: 5'-CAG GAA ACA GCT ATG AC-3'	50	30

TABLE S2: Oligonucleotide probes used for CARD-FISH

Probe	Sequence (5'-3')	Reference
Eub338I (Bacteria)	GCTGCCTCCCGTAGGAGT	(1)
Eub338II (Bacteria)	GCAGCCACCCGTAGGTGT	(2)
Eub338III (Bacteria)	GCTGCCACCCGTAGGTGT	(2)
Cren 554 (Crenarchaeota)	TTAGGCCAATAATCMTCCT	(8)
Ntspa712 (Nitrospira)	CGCCTTCGCCACCGGCCTTCC Competitor: CGCCTTCGCCACCGGTGTTCC	(3)
AOB NSO1225	CGCCATTGTATTACGTGTGA	(9)
AOB NSO156	TATTAGCACATCTTCGAT	(9)

TABLE S3: Influence of the NH_4^+ concentration on the growth rates [h^{-1}] of the enrichment culture AOA-AC2, AOA-AC5, AOA-DW and AOB-G5-7 (data are similar to data in Figure 2) (mean \pm SD, n=3; different letters behind values indicate significant differences between values determined by one-way ANOVA followed by Tukey test; P < 0.05).

NH_4^+ [mM]	AOA-AC2	AOA-AC5	AOA-DW	AOB-G5-7
0.01	$0.020 \pm 0.002^{\text{a}}$	$0.023 \pm 0.003^{\text{a}}$		
0.0158			$0.023 \pm 0.003^{\text{a}}$	$0.030 \pm 0.002^{\text{a}}$
0.05	$0.019 \pm 0.001^{\text{ab}}$	$0.021 \pm 0.000^{\text{ab}}$	$0.017 \pm 0.000^{\text{b}}$	$0.042 \pm 0.000^{\text{b}}$
0.1	$0.017 \pm 0.000^{\text{ab}}$	$0.020 \pm 0.001^{\text{bcd}}$		
0.158			$0.017 \pm 0.000^{\text{b}}$	$0.051 \pm 0.004^{\text{bc}}$
0.25	$0.017 \pm 0.001^{\text{ab}}$	$0.020 \pm 0.001^{\text{abc}}$		
0.5	$0.017 \pm 0.002^{\text{ab}}$	$0.018 \pm 0.001^{\text{bcd}}$	$0.016 \pm 0.000^{\text{b}}$	$0.057 \pm 0.005^{\text{cd}}$
1	$0.017 \pm 0.001^{\text{ab}}$	$0.020 \pm 0.001^{\text{ab}}$	$0.016 \pm 0.000^{\text{b}}$	$0.059 \pm 0.003^{\text{cd}}$
2	$0.015 \pm 0.002^{\text{b}}$	$0.016 \pm 0.001^{\text{cd}}$		$0.060 \pm 0.005^{\text{cd}}$
3				$0.065 \pm 0.004^{\text{d}}$
5		$0.016 \pm 0.000^{\text{d}}$		$0.061 \pm 0.003^{\text{d}}$

TABLE S4: Influence of NH₄⁺ concentration on the lag phase [h] before onset of logarithmic growth in the enrichment cultures AOA-AC2; AOA-AC5, AOA-DW, and AOB-G5-7 (data are similar to data in Figure S3; mean \pm SD, n=3; different letters behind values indicate significant differences between values determined by one-way ANOVA followed by Tukey test; P < 0.05).

NH ₄ ⁺ [mM]	AOA-AC2	AOA-AC5	AOA-DW	AOB-G5-7
0.01	48.0 \pm 0.0 ^a	48.0 \pm 0.0 ^a		
0.0158			43.0 \pm 13.9 ^a	27.0 \pm 0.0 ^a
0.05	48.0 \pm 0.0 ^a	48.0 \pm 0.0 ^a	75.5 \pm 0.0 ^a	27.0 \pm 0.0 ^a
0.1	64.0 \pm 27.7 ^a	48.0 \pm 0.0 ^a		
0.158			51.0 \pm 0.0 ^b	27.0 \pm 0.0 ^a
0.25	96.0 \pm 0.0 ^a	96.0 \pm 0.0 ^{ab}		
0.5	256.5 \pm 27.3 ^b	48.0 \pm 0.0 ^a	99.0 \pm 0.0 ^c	51.0 \pm 0.0 ^b
1	256.5 \pm 27.3 ^b	144.0 \pm 0.0 ^{bc}	243.0 \pm 0.0 ^d	35.0 \pm 13.9 ^{ab}
2	288.0 \pm 0.0 ^b	192.0 \pm 0.0 ^c		43.0 \pm 13.9 ^{ab}
3				75.5 \pm 0.0 ^c
5		512.0 \pm 27.7 ^d		75.5 \pm 0.0 ^c

TABLE S5: Influence of the calculated O₂ concentrations in the headspace of the bottle on the growth rates [h⁻¹] of the enrichment cultures AOA-AC2, AOA-AC5, AOA-DW, and AOB-G5-7 (data are similar to data in Figure 3; mean ± SD, n=3; different letters behind values indicate significant differences between values determined by one-way ANOVA followed by Tukey test; P < 0.05).

O ₂ [%]	AC2	AC5	DW	G5-7
0.5	0.000 ± 0.000 ^a	0.005 ± 0.000 ^a	0.014 ± 0.004 ^a	0.010 ± 0.000 ^a
1	0.008 ± 0.001 ^b	0.008 ± 0.000 ^b	0.016 ± 0.000 ^a	0.018 ± 0.001 ^b
2	0.011 ± 0.001 ^{bc}	0.009 ± 0.001 ^c	0.015 ± 0.001 ^a	0.031 ± 0.003 ^c
21	0.013 ± 0.001 ^c	0.008 ± 0.000 ^{bc}	0.019 ± 0.003 ^a	0.045 ± 0.002 ^d

TABLE S6: Influence of the pH value on the growth rates [h^{-1}] of the enrichment cultures AOA-AC2, AOA-AC5, AOA-DW, and AOB-G5-7 (data are similar to data in Figure 4; mean \pm SD, n=3; different letters behind values indicate significant differences between values determined by one-way ANOVA followed by Tukey test; P < 0.05).

pH	AC2	AC5	DW	G5-7
6	0.000 \pm 0.000 ^a	0.010 \pm 0.000 ^{ab}	0.017 \pm 0.001 ^a	0.034 \pm 0.003 ^a
6.5	0.011 \pm 0.001 ^{bc}	0.011 \pm 0.001 ^{abc}	0.018 \pm 0.001 ^{ad}	0.046 \pm 0.004 ^b
7	0.013 \pm 0.000 ^d	0.013 \pm 0.001 ^c	0.024 \pm 0.000 ^b	0.049 \pm 0.000 ^b
7.5	0.012 \pm 0.000 ^{cd}	0.011 \pm 0.001 ^{abc}	0.022 \pm 0.001 ^{bc}	0.051 \pm 0.004 ^b
8	0.010 \pm 0.000 ^b	0.012 \pm 0.000 ^{bc}	0.020 \pm 0.002 ^{cd}	0.047 \pm 0.005 ^b
8.5	0.005 \pm 0.000 ^e	0.008 \pm 0.000 ^a	0.020 \pm 0.001 ^{cd}	0.025 \pm 0.001 ^c
9	0.006 \pm 0.000 ^e	0.010 \pm 0.002 ^{ab}	0.020 \pm 0.001 ^{cd}	0.018 \pm 0.001 ^c

TABLE S7: Influence of white, red, and blue light with the intensity of 30 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ and blue light with the intensity of 3 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ on the growth rates [h^{-1}] of the enrichment cultures AOB-G5-7 (data are similar to data in Figure 5; mean \pm SD, n=3; different letters behind values indicate significant differences between values determined by one-way ANOVA followed by Tukey test; P < 0.05).

G5-7	white	red	blue	blue low intensity
light	0.056 \pm 0.001 ^a	0.051 \pm 0.005 ^a	0.000 \pm 0.000 ^a	0.057 \pm 0.004 ^a
light -> dark	0.056 \pm 0.004 ^a	0.047 \pm 0.002 ^a	0.051 \pm 0.002 ^b	0.068 \pm 0.004 ^b
dark -> light	0.058 \pm 0.001 ^a	0.049 \pm 0.002 ^a	0.064 \pm 0.004 ^c	0.055 \pm 0.002 ^a
dark	0.055 \pm 0.001 ^a	0.045 \pm 0.002 ^a	0.066 \pm 0.004 ^c	0.063 \pm 0.003 ^{ab}

TABLE S8: Influence of white, red, and blue light with the intensity of 30 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ and blue light with the intensity of 3 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ on the growth rates [h^{-1}] of the enrichment cultures AOA-DW (data are similar to data in Figure 5; mean \pm SD, n=3; different letters behind values indicate significant differences between values determined by one-way ANOVA followed by Tukey test; P < 0.05).

	white	red	blue	blue low intensity
light	0.000 \pm 0.000 ^a	0.012 \pm 0.000 ^a	0.000 \pm 0.000 ^a	0.011 \pm 0.001 ^a
light -> dark	0.000 \pm 0.000 ^a	0.012 \pm 0.001 ^a	0.000 \pm 0.000 ^a	0.013 \pm 0.001 ^{bc}
dark -> light	0.018 \pm 0.000 ^b	0.018 \pm 0.001 ^b	0.018 \pm 0.001 ^b	0.012 \pm 0.001 ^{ab}
dark	0.017 \pm 0.001 ^b	0.016 \pm 0.000 ^c	0.020 \pm 0.000 ^c	0.015 \pm 0.001 ^c

References:

1. **Amann, R. I., B. J. Binder, R. J. Olson, S. W. Chisholm, R. Devereux and D. A. Stahl.** 1990. Combination of 16S ribosomal-RNA targeted oligonucleotide probes with flow-cytometry for analyzing mixed microbial populations. *Appl. Environ. Microbiol.* **56**:1919-1925.
2. **Daims, H., A. Bruhl, R. Amann, K.-H. Schleifer and M. Wagner.** 1999. The domain-specific probe EUB338 is insufficient for the detection of all bacteria: Development and evaluation of a more comprehensive probe set. *Syst. Appl. Microbiol.* **22**:434-444.
3. **Daims, H., J. L. Nielsen, P. H. Nielsen, K.-H. Schleifer and M. Wagner.** 2001. In situ characterization of *Nitrospira*-like nitrite-oxidizing bacteria active in wastewater treatment plants. *Appl. Environ. Microbiol.* **67**:5273-5284.
4. **DeLong, E. F.** 1992. Archaea in costal marine environments. *Proc. Natl. Acad. Sci.* **89**:5685-5689.
5. **Francis, C. A., K. J. Roberts, J. M. Beman, A. E. Santoro and B. B. Oakley.** 2005. Ubiquity and diversity of ammonia-oxidizing archaea in water columns and sediments of the ocean. *Proc. Natl. Acad. Sci.* **102**:14683-14688.
6. **Grosskopf, R. P. H. Janssen, and W. Liesack.** 1998. Diversity and structure of the methanogenic community in anoxic rice paddy soil microcosms as examined by cultivation and direct 16S rRNA gene sequence retrieval. *Appl. Environ. Microbiol.* **64**: 960-969.
7. **Kowalchuk, G. A., J. R. Stephen, W. DeBoer, J. I. Prosser, T. M. Embley and J. W. Woldendorp.** 1997. Analysis of ammonia-oxidizing bacteria of the beta subdivision of the class *Proteobacteria* in coastal sand dunes by denaturing gradient gel electrophoresis

- and sequencing of PCR-amplified 16S ribosomal DNA fragments. *Appl. Environ. Microbiol.* **63**:1489-1497.
8. **Massana, R., A. E. Murray, C. M. Preston and E. F. DeLong.** 1997. Vertical distribution and phylogenetic characterization of marine planktonic Archaea in the Santa Barbara Channel. *Appl. Environ. Microbiol.* **63**:50-56.
 9. **Mobarry, B. K., M. Wagner, V. Urbain, B. E. Rittmann and D. A. Stahl.** 1996. Phylogenetic probes for analyzing abundance and spatial organization of nitrifying bacteria. *Appl. Environ. Microbiol.* **62**:2156-2162.
 10. **Rotthauwe, J. H., K. P. Witzel and W. Liesack.** 1997. The ammonia monooxygenase structural gene *amoA* as a functional marker: Molecular fine-scale analysis of natural ammonia-oxidizing populations. *Appl. Environ. Microbiol.* **63**:4704-4712.