Supplementary material

Supplementary figure 1:

An-LxrA	1	MSRSLEGKFAIITGGSRGIGEAIAHNL
An-XhrA	1	MGLKGKVAIVTGGARGIGAGIVRSL
Tr - LXR4	1	MARBYEGKLAIVTGASRGIGAAVARRL
Tr-LXR3	1	MTQMKNGAFPHDNAAVPNVERVLPLFSLKGRTAIVSGAGAGIGLAVAQAF
An-LxrA	28	ASKGCSLLLNYTSDSSRTRTESLCNTLSTTHKITCIPVQADLSDPAPAVN
An-XhrA	26	SEOGAKVAFNYVSSSSRKAADALIESLRON - NNEATAVOADITDPN - APK
Tr-LXR4	28	AAKGSNVLITFTSDSSRDLTRGLVEELSSKHGVHVOSVOTDLAKASTAAP
Tr-LXR3	51	AFAGANVATWYNSNKOA VT - SAFDIAKTYGVKCKAYOVNVTSAE - AVD
An-LxrA	78	TIISAAKTHETSPT TNTLTIDILINNAGVSKDR - ELNDPSSGPIDP
An-XhrA	74	MITOAALEAFOT DRIDILVNNAGAGDNR - PLEEV TMDS
Tr-LXR4	78	TIVEAARTLEDSYSPPSGGKKFOVDILINNAGVSSNO-ELNDPEKGATDE
Tr-LXR3	97	KATTEIIKEENGRLDVFVANSGITWTEGAFIDGSVES
An-LxrA	123	AYFNWHYTINVLAPILLTOACAEYLPRKPA
An-XhrA	111	- YMRLMDVNVRAVIFMTOAILPYIPRGGRIIN
Tr-LXR4	127	AEFTRVYAINVLAPLLLTOAVAPHLPADRSGRIVN
Tr-LXR3	134	ARNVMSVNVDGVMWCAKSAGAHFRROKEEGTTIDGKPLDNFIAGSFTA
11 1	101	
An-LxrA	160	ISSISSIGF TGOSVYGGTKAALEAMTRTWARELADV ATVNAVNPG
An-XhrA	142	LSSISSRGGY ATOSVYAASKAAVEGLTRVWATELGHKYGVTVNAANPG
Tr-LXR4	162	VSSVSASIGY LGOSVYAGSKGALEVMTRTWARELAER ATVNSVNPG
Tr-LXR3	182	TASMSGSIVNVPOLOAVYNSSKAAVIHFCKSLAVEWTGF ARVNTVSPG
An-LxrA	206	PVVGDMYFATGEEFWKOMOGFODNTPLSKLVDGEEAVEELLSEEOKR
An-XhrA	190	PVD TDMYOAA GEVHLKRMEEONK
Tr-LXR4	208	PAWGDMYA EAGPTFWRRNOPYVDAAPLMA - YDGEEDVLRRAGGEADKFDR
Tr-LXR3	230	YIITEISNFVPPETKT
	(1997) (1	
An-LxrA	253	LIREKMGGRRPAFTREIAGVVGMLCTEDGAWCTGSVVCANGGLK FT
An-XhrA	213	KVPAGORCGTVODIGDIVSFLAEERSRWVTGDVICANGGMLYI
Tr-LXR4	257	LVREOMGGRRPGFADEIAGTVDMLCTEESGWTTGSVVCANGGMRMSIA
Tr-LXR3	246	LWKDKIVMGREGRVGELKGAYLYLASDASSYTTGLDMIVDGGYSLP

Legend figure S1:

Protein sequence alignment of LxrA, XhrA, LXR4, and LXR3. In *Aspergillus niger*, two close homologues, LxrA and XhrA, function in distinct catabolic pathways for L-arabinose and for D-galactose, respectively. In *Trichoderma reesei* (*Hypocrea jecorina*), the close homologue of these two proteins, LXR4, is responsible for L-xylo-3-hexulose conversion to D-sorbitol in the oxido-reductive D-galactose pathway; while the more distant homologue, LXR3, carries out the L-xylulose reduction in the L-arabinose catabolic pathway.

Supplementary figure 2:



The Vista track (comparative analysis) of the *xhrA* and *galX* in selected Aspergilli

Legend figure S2:

Comparison of the genomic regions covering the *galX* and *xhrA* genes in three *Aspergillus* species with *A. niger*. While all of the species contain the *galX* gene, only *A. oryzae* and *A. fumigatus*, but not *A. nidulans*, seem to have the *xhrA* gene (the figure is a modification of a vista visualization obtained from the JGI Aspergillus niger v3.0 web database - http://genome.jgi-psf.org/Aspni5/).

Supplementary figure 3:

	spo	ores	mycelium	
	wт	∆ladB	wт	∆ladB
without CS	•	•	Ø	8
2% D-galactose	٠	•	۲	
2% D-galactose 0.025% D-glucose	۲	•	•	Ø
2% D-galactose 0.025% D-xylose	0	0		
2% galactitol	-	•		٠
2% D-glucose				6

Legend figure S3:

Growth of the *A. niger* strain ATCC 1015 (WT) and the ATCC 1015 with a deletion in *ladB* ($\Delta ladB$) on different carbon sources (CS). Comparison of the growth originated from either conidiospores (left panel) or pre-grown mycelium (right panel). The conidiospores were collected from cultures grown on PDA (potato-dextrose agar) plates and kept at -80°C. The mycelia were produced by inoculating the spores into YESG (2% yeast extract; 4% sucrose; 3% gelatin) medium and cultivating at 28°C for 18 hours. The mycelium was collected by filtration, washed with sterile water, and cut into identical pieces

prior to the inoculation on the agar plates. The test plates contained YNB (yeast nitrogen base), 2% agar, and a carbon source as indicated in the figure. The cultivation was carried out at 28°C for 4 days in the case of spore-inoculation or 3 days in the case of mycelium-inoculation.

Here we summarise the observations supporting our claims:

- The mycelium pre-grown in the presence of sucrose and peptides is not able to grow after transfer to D-galactose.
- The addition of a small amount of D-xylose into the D-galactose-containing medium leads to considerable enhancement of growth of the wild type stain regardless of the form of inoculum. This is caused by induction of XyrA-dependent D-galactose reductase activity and not by the induction of the Leloir pathway (1).
- On the other hand, addition of a small amount D-glucose into the D-galactose-containing medium leads to only minimal growth probably caused by the D-glucose itself rather than by D-galactose. This is expected as the genes encoding the D-galactose oxido-reductive metabolism are repressed (Fig. 2, (1)). However, Fekete et al. showed that in their *A. niger* strain the genes encoding the Leloir pathway are expressed even in presence of D-glucose (2).
- The *A. niger* strain we used seemed to have no difficulty to sporulate in the presence of D-galactose when supplemented by a small amount of D-xylose (Fig. S3).
- 1. Mojzita, D., Koivistoinen, O. M., Maaheimo, H., Penttilä, M., Ruohonen, L., and Richard, P. (2012) *Fungal Genet. Biol.* **49**, 152-159
- 2. Fekete, E., de Vries, R. P., Seiboth, B., Vankuyk, P. A., Sándor, E., Fekete, E., Metz, B., Kubicek, C. P., and Karaffa, L. (2012) *FEMS Microbiol Lett.* **329**, 198-203