# **Supporting Information:**

- **4** Table S1. Antibodies used in flow cytometry and definition of T cell subsets.
- **4** Table S2. Geometric mean and range of multiple cytokine-producing  $CD4^+$  cells.
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### Figure S1. Gating strategies.

Whole blood cells stimulated with PMA/ionomycin were fixed with 4% paraformaldehyde after lysing the RBCs, and cryopreserved until used. At the time of analysis the cells were thawed and stained with all antibodies in the cocktail (Table S1) except the one that registers in the channel of particular interest [Fluorescence Minus one (FMO) Staining]. The gate drawn for each channel was used to determine the frequency cytokine producing cells in response to antigen stimulation in that channel.

# Downloaded from http://jimmunol.org/ at NIH Library, National Institutes of Health on July 3, 2012

# Table S1

List of antibodies used in flow cytometry and definition of T helper cell subsets.

Marker	Species	Clone	Fluorochrome	Company	
CD3	Mouse	UCHT1	Alexa Fluor 700	eBioscience	
CD4	Mouse	RPA-T4	Alexa Fluor 750/PE-Cy5	eBioscience	
CD8	Mouse	3B5	Qdot605	Invitrogen	
CD25	Mouse	M-A251	APC-Cy7	BD Bioscience	
CD127	Mouse	hIL-7R-M21	PE	BD Bioscience	
CD152	Mouse	BNI3	APC	BD Bioscience	
Foxp3	Rat	PCH101	Alexa Fluor 488	eBioscience	
IFN-γ	Mouse	4S.B3	PE-Cy7	eBioscience	
IL-4	Rat	MP4-25D2	FITC	eBioscience	
IL-5	Rat	TRFK5	РЕ	BD Bioscience	
IL-10	Rat	JES3-9D7	APC/Pacific Blue	eBioscience	
IL-17A	Mouse	eBio64DEC17	PerCp-Cy5.5	eBioscience	
TNF-α	Mouse	MAb11	Pacific Blue	eBioscience	
Th1	$CD3^{+}CD4^{+}IFN-\gamma^{+}IL-10^{+}TNF-\alpha^{+}IL-17A^{-}IL-5^{-}IL-4^{-}, CD3^{+}CD4^{+}IFN-\gamma^{+}IL-10^{+}TNF-\alpha^{-}IL-10^{+}TNF-\alpha^{-}IL-10^{+}ID-10^{+$				
	IL-17A <sup>-</sup> IL-5 <sup>-</sup> IL-4 <sup>-</sup> , and CD3 <sup>+</sup> CD4 <sup>+</sup> IFN- $\gamma^+$ IL-10 <sup>-</sup> TNF- $\alpha^-$ IL-17A <sup>-</sup> IL-5 <sup>-</sup> IL-4 <sup>-</sup> ;				
Th2	CD3 <sup>+</sup> CD4 <sup>+</sup> IL-4 <sup>+</sup> IL-5 <sup>+</sup> IL-10 <sup>+</sup> IFN-γ <sup>-</sup> TNF-α <sup>-</sup> IL-17A <sup>-</sup> , CD3 <sup>+</sup> CD4 <sup>+</sup> IL-4 <sup>+</sup> IL-5 <sup>+</sup> IL-10 <sup>-</sup>				
	IFN- $\gamma^{-}$ TNF- $\alpha^{-}$ IL-17A <sup>-</sup> , CD3 <sup>+</sup> CD4 <sup>+</sup> IL-4 <sup>+</sup> IL-5 <sup>-</sup> IL-10 <sup>-</sup> IFN- $\gamma^{-}$ TNF- $\alpha^{-}$ IL-17A <sup>-</sup> , and				
	$CD3^{+}CD4^{+}IL-5^{+}IL-4^{-}IL-10^{-}IFN-\gamma^{-}TNF-\alpha^{-}IL-17A^{-}$				
Th17	$CD3^{+}CD4^{+}IL-17A^{+}TNF-\alpha^{+}IL-10^{+}IFN-\gamma^{-}IL-5^{-}IL-4^{-}, CD3^{+}CD4^{+}IL-17A^{+}TNF-\alpha^{+}IL-10^{$				
	10 <sup>-</sup> IFN- $\gamma$ <sup>-</sup> IL-5 <sup>-</sup> IL-4 <sup>-</sup> , CD3 <sup>+</sup> CD4 <sup>+</sup> IL-17A <sup>+</sup> IL-10 <sup>+</sup> TNF- $\alpha$ <sup>-</sup> IFN- $\gamma$ <sup>-</sup> IL-5 <sup>-</sup> IL-4 <sup>-</sup> , and				
	$CD3^{+}CD4^{+}IL-17A^{+}TNF-\alpha^{-}IL-10^{-}IFN-\gamma^{-}IL-5^{-}IL-4^{-}$				
aTreg/Tr1	aTreg/Tr1 consisted of CD3 <sup>+</sup> CD4 <sup>+</sup> IL-10 <sup>+</sup> IL-4 <sup>-</sup> IL-5 <sup>-</sup> IFN-γ <sup>-</sup> TNF-α <sup>-</sup> IL-17A <sup>-</sup> Foxp3 <sup>-</sup> ;				
nTreg	CD3 <sup>+</sup> CD4 <sup>+</sup> CD25 <sup>+</sup> Foxp3 <sup>+</sup> CD127 <sup>-</sup>				
"TNF- α"	$CD3^{+}CD4^{+}TNF - \alpha^{+}IL - 10^{+}IFN - \gamma^{-}IL - 17A^{-}IL - 5^{-}IL - 4^{-} and CD3^{+}CD4^{+}TNF - \alpha^{+}IL - 10^{-}IL - 10^{-$				
	IFN-γ <sup>-</sup> IL-17A <sup>-</sup> IL-5 <sup>-</sup> IL-4 <sup>-</sup>				

Flow Cytometry

Cryopreserved fixed whole blood cells were thawed at room temperature and washed twice with 1X PBS and transferred to 96-wells V-bottom costar plates (Corning). The cells were then incubated with 1X Permeabilization buffer (eBioscience) for 15 minutes at room temperature and washed again with 1X Permeabilization buffer. The cells were then blocked for 15 minutes at room temperature then washed again with 1X Permeabilization buffer. The cells were then blocked for 15 minutes at room temperature then washed again with 1X Permeabilization buffer. The cells were then blocked for 15 minutes at room temperature then washed again with 1X Permeabilization buffer. The cells were then incubated with antibodies for the effector and regulatory panels for 30 minutes then washed twice with PBS. After washing, the cells were resuspended in 0.2 ml of PBS and transferred to 96-plug micro tubes (Bio-Rad) and acquired on an LSRII (BD Bioscience).

## Table S2

Median and range of multiple cytokine-producing  $\text{CD4}^+$  cells

	Fil-	Fil+	
	GM (Range)	GM (Range)	P value
Lymphocyte count	1.9 x 106 (0.85x106 – 2.5x106)	1.9 x106 (1.2 x 106 – 3.8 106)	>0.05
CD4+ count	0.51 x106 (0.7 x105 – 9.2 x 105)	0.46 x105 (0.09 x105 - 1.16 x106)	> 0.05
Malaria antigen			Do
$CD4^{+}IFN\text{-}\gamma^{+}$	1767.0 (322.0 – 5023.0)	481.4 (0.0 – 1631.0)	0.0013 ond
CD4 <sup>+</sup> IL-4 <sup>+</sup>	715.4 (78.5 – 1794.0)	567.3 (19.49 - 5020.0)	>0.05 for
CD4 <sup>+</sup> IL-5 <sup>+</sup>	5213.0 (113.7 - 72892.0)	3455.0 (53.82 - 21093.0)	>0.05 lig
$CD4^{+}IL-10^{+}$	884.3 (92.38 - 2929.0)	226.0 (37.12 - 11239.0)	0.04
$CD4^{+}IL-17A^{+}$	1533.0 (437.0 - 6080.0)	949.1 (12.06 - 4474.0)	>0.05
$CD4^{+}TNF-\alpha^{+}$	2064.0 (234.5 - 7458.0)	1055.0 (48.26 – 4078.0)	0.03 at Z
Staphylococcus aureus enteroto	oxin B		IH Lib
$CD4^{+}IFN\text{-}\gamma^{+}$	7241 (629.3 – 21462.0)	9187 (17.63 - 14916)	>0.05 y, N
CD4 <sup>+</sup> IL-4 <sup>+</sup>	2816.0 (227.4 - 8511.0)	2725.0 (194.0 - 30680.0)	>0.05 affional
CD4 <sup>+</sup> IL-5 <sup>+</sup>	6817.0 (199.0 - 79534.0)	4810.0 (202.4 - 30371.0)	>0.05 Institu
CD4 <sup>+</sup> IL-10 <sup>+</sup>	3229.0 (142.1 - 8900.0)	2185.0 (41.76 - 16259.0)	>0.05 of
$CD4^{+}IL-17A^{+}$	3229.0 (270.0 - 10376.0)	4079.0 (41.76 - 11130.0)	>0.05 Health
$CD4^{+}TNF-\alpha^{+}$	13763.0 (682.2 – 26837.0)	15185.0 (595.6 – 52628.0)	>0.05 <sup>on</sup> July

\* P value adjusted for multiple comparisons.

Note: Bold indicates statistically significant

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Figure S1