SUPPLEMENTAL FIGURE LEGENDS

FIGURE S1. **Comparison of growth characteristics of *FTL_0325* mutant and WT *F. tularensis* LVS.** (A) Growth curves of *FTL_0325* mutant (Pink lines) and *F. tularensis* LVS (Blue lines) were generated by growing bacteria in Chamberlain's medium (CDM), Mueller-Hinton broth (MHB) or Brain Heart Infusion (BHI) broth with or without 10% fetal bovine serum (FBS). Left panel: OD_{600} were recorded at 2-hour intervals in a Biotek synergy HT plate reader (BioTek). Right panel: the bacterial numbers were quatitated by plating ten-fold dilutions on MH-chocolate agar plates and expressed as Log_{10} CFU. (B) Disc diffusion assay to determine the sensitivity of *FTL_0325* mutant to polymyxin B (0-200 µg/ml), SDS (0.25-2% W/V), Triton-X100 (0.25-2% V/V) and superoxide generating compound pyrogallol (0.125 mM-1M).

FIGURE S2. ***FTL_0325* mutant of *F. tularensis* LVS does not exhibit enhanced serum sensitivity.** Mid log phase grown cultures of WT *F. tularensis* LVS and *FTL_0325* mutants were exposed to indicated concentrations of human serum. Aliquots were collected at the indicated times, diluted 10 fold and plated on chocolate agar plates to quantitated bacterial numbers. Results are expressed as the percentage of bacteria that survived the treatment. The data are cumulative of two independent experiments.

Table S1: Primers, plasmid vectors, and bacterial strains used and generated in the present study

S. No.	Primer	Sequence	Purpose
1	ompA Apa1 for pcDNA 3.1 (-)	5'AAACGGGCCCATGAAAAAA TTACTGAAACTATG3'	To express FTT0831c in pcDNA 3.1
2	ompA EcoR1 for pcDNA 3.1 (-)	5'CCGGAATTCTTACTTGTCGTG TCGTCCTTGTAGTCTAAAATAG TCGTTGCCA TTGAAT3'	To express FTT0831c in pCDNA
3	ompA upF	5'ACCGCTCGAGAGCCTCTACC ATAACTACTCCTT3'	Amplify region upstream of *FTT0831c* to make ∆*FTT0831c*.
4	ompA upR	5'GATAATACCATTAAAAACTTA ATTATAC3'	Amplify region upstream of *FTT0831c* to make ∆*FTT0831c*.
5	ompA DnF	5'AGTTTTTAATGGTATTATCAAT CAAGTATAAAACCTAAAATAGT ATTTTATATCATT3'	Amplify region downstream of *FTT0831c* to make ∆*FTT0831c*.
6	ompA DnR	5'AGGAGATCTCAGTCTCAACA GCACGACTTAC3'	Amplify region downstream of *FTT0831c* to make ∆*FTT0831c*.
7	Pkk214 F	5, GTA ACT GCA GAT GAA AAA ATT ACT TGA AAC TAT GCT3'	Amplification of *FTT0831c* for cloning in transcomplementation vector pKK214.
8	Pkk214R	TTA TAA AAT AGT CGT TGC CAT TGA AT	Amplification of *FTT0831c* for cloning in. transcomplementation vector pKK214
	Plasmid Vectors		
	pDMK		(26)
	pKK214::GFP		Albany Med. Microbiology core
	pcDNA 3.1		Invitrogen
	Bacterial Strains		

***E. coli* strains**

S17-1 (pDMK::Δ*FTT0831c*)	This study
$DH_5α$ (pKK214::*FTT0831c*)	This study
$DH_5α$ (pcDNA::*FTT0831c*)	This study

***Francisella* strains**

FTL_0325 mutant	This study
Δ*FTT0831c* mutant	This study
Δ*FTT0831c*+p*FTT0831c*	This study

Figure S1

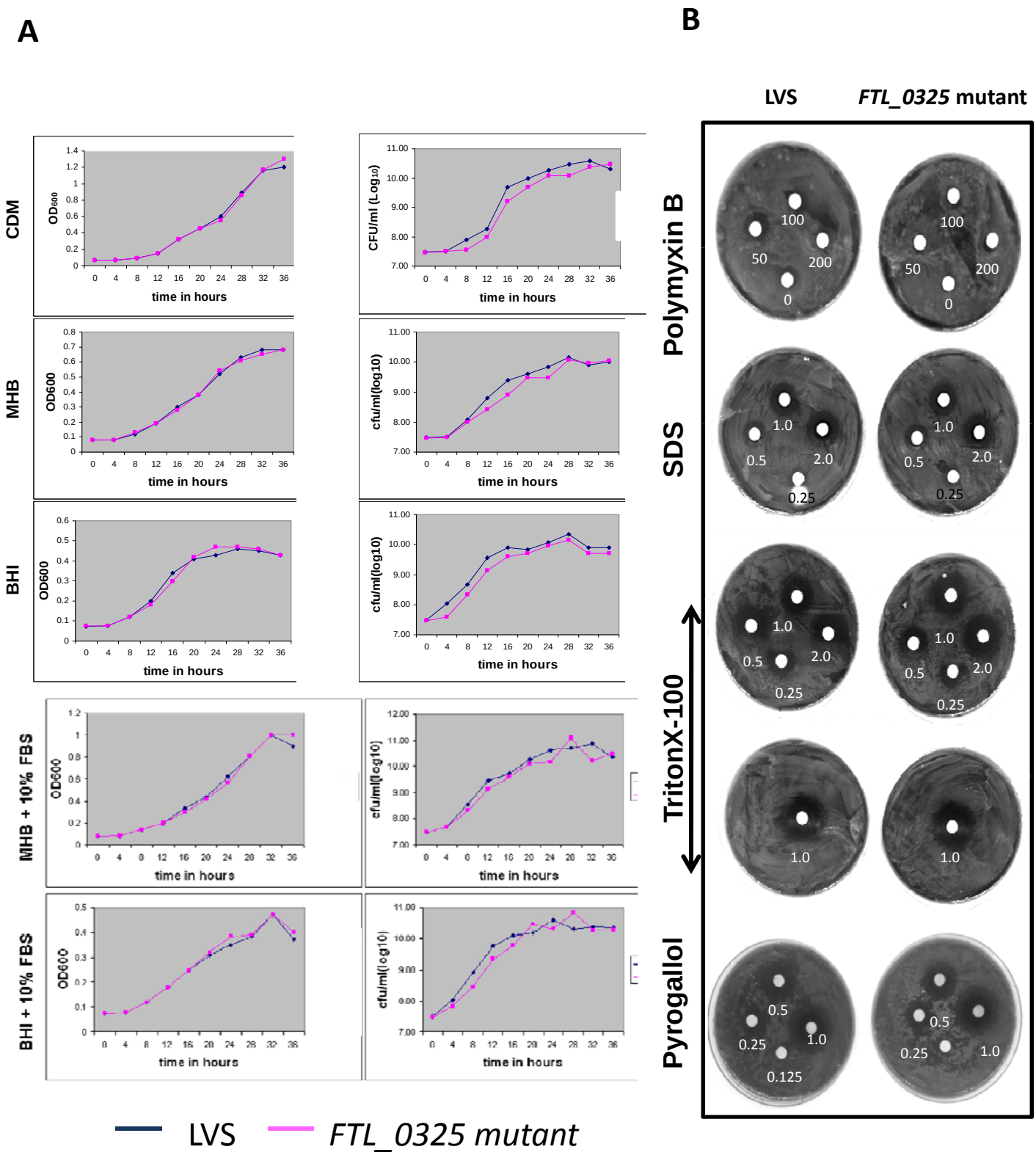
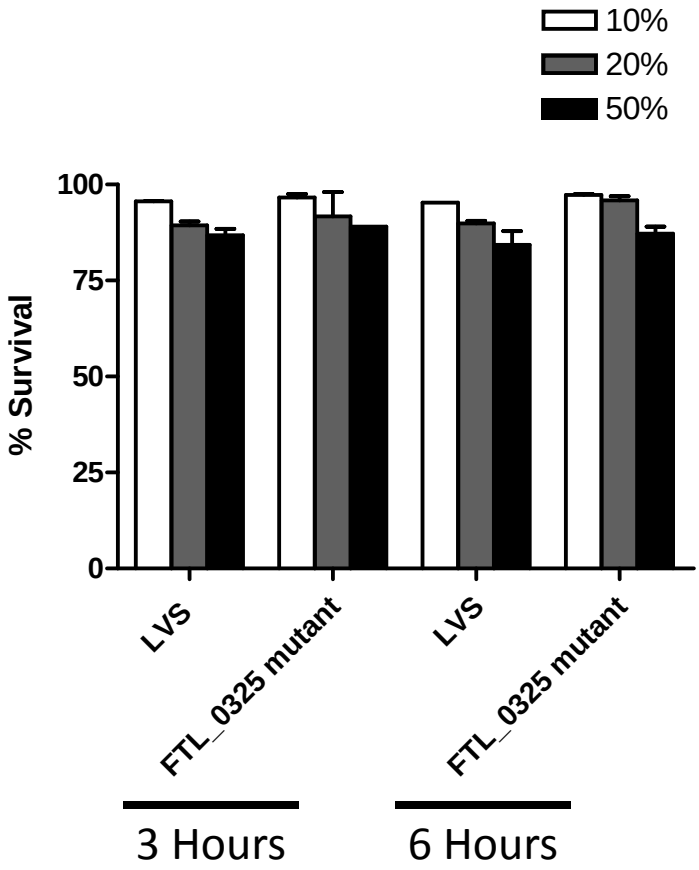

A
B
CDM
MHB
BHI
MHB + 10% FBS
BHI + 10% FBS
OD600
CFU/ml (Log10)
cfu/ml(log10)
time in hours
LVS
FTL_0325 mutant
Polymyxin B
SDS
TritonX-100
Pyrogallol
100
50
200
0
1.0
0.5
2.0
0.25
0.125

Figure S2


10%
20%
50%
100
75
50
25
0
% Survival
LVS
FTL_0325 mutant
LVS
FTL_0325 mutant
3 Hours
6 Hours